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Supplementary data:

Fulleropyrrolidines Derived from Dioxa- and Trioxaalkyl-tethered Diglycines

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1. General experimental details and procedures

General: IR spectra were recorded with a *Perkin-Elmer FTIR 1725X* spectrophotometer. UV spectra were recorded with a *GBC-Cintra 40* UV-vis spectrophotometer. ¹H and ¹³C NMR spectra were recorded with *Varian Gemini 200* (¹H at 200 MHz, ¹³C at 50 MHz) and *Bruker Avance* spectrometers (¹H at 500 MHz, ¹³C at 125 MHz). Chemical shifts are measured in ppm, *J* in Hz. Sample was dissolved in the indicated solvent system, and TMS was used as an internal reference. The high-resolution MS spectra were obtained with an *Agilent Technologies 6210* TOF LC-MS spectrometer. Dry-column flash chromatography (DCFC) was carried out with Merck silica gel 60 (15-40μm). Thin layer chromatography (TLC) was carried out on precoated silica gel 60 F254 plates.

HPTLC. All substances were chromatographed on HPTLC silica gel 60 aluminium sheets (Merck, 4.0 × 6.0 cm) as a stationary phase. Samples were applied by CAMAG Linomat 5 "linomat5_130827" S/N 130827 (1.00.12) device. Spots were detected by CAMAG TLC Scanner 3 "Scanner3_131003" S/N 131003 (1.14.26), at 340 nm. Software application winCATS Planar Chromatography Manager SN 1311W038, V1.4.2 was used for data processing. In addition, UV spectra of the spots were obtained by scanning from 200-700 nm.

System toluene/ethyl-acetate 7:3 was used for HPTLC of bisadducts **7-9**, while toluene/ethyl-acetate 1:1 was used for adducts **10-14** as a mobile phase.

Morphology investigations: Investigations of sample morphology were carried out with SEM, using a *JEOL JSM-840A* instrument, at an acceleration voltage of 30 kV. The samples for investigation of morphology of self-organized stuctures of **7-14** were prepared by dissolving in differente solvents (ODCB, PhMe, CHCl₃, PhMe/*i*-PrOH (1:1, v/v), PhMe/dioxane (1:1, v/v) at room temperature. A drop of 0.5 mM solution of fullerene derivative was deposited on the surface of a glass substrate (10x10 mm) and left during 24 h to slowly evaporate in a glass Petri dish (diameter 10 cm) under PhMe atmosphere at room temperature (the exceptions were made in case of ODCB solutions, in which 2-3 days were necessary for total evaporation of the solvent). The investigated samples were gold sputtered in a JFC 1100 ion sputter device and then subjected to SEM observations.

Electrochemical Measurements: The electrochemical behavior of C_{60} bis-adducts was investigated using 1mM solutions of bisadducts **7-13** and **15-19** and difullerene **14** in dry and degased mixture ODCB/DMF 2:1, and in DCM (only bisadducts **7-13**), both containing 0.1 M TBAP as a supporting electrolyte. In order to remove oxigen from the electrolyte, the system was bubbled with argon prior to each experiment and argon atmosphere above the liquid surface was maintained during the scans. The electrochemical mesurements were carried out on *CH1760b Electrochemical workstation potentiostat (CH Instruments, Austin, TX)* using conventional three-electrode cell (5 mL) equipped with GCE (glassy carbon electrode), as a working, Ag/Ag⁺ (a silver wire in contact with 0.01 M AgNO₃ and 0.10 M TBAP in acetonitrile), as a reference and the platinum wire as a auxiliary electrodes, calibrated with a ferrocene/ferrocenyl couple (Fc/Fc⁺) as an internal standard. All experiments were performed at room temperature in the potential range of -2.5 to 0.5 V vs Ag/Ag⁺ (i.e. -3.0 to 0.0 V vs Fc/Fc⁺), at sweep rates between 0.01 and 1 V/s. All half-wave reduction potentials are presented in V vs Fc/Fc⁺ (measured $E_{1/2}$ of Fc/Fc⁺: 0.552 and 0.674 V vs Ag/Ag⁺in DCM and ODCB/DMF 2:1, respectively).

Antioxidant Activity in vitro: The antioxidant capacity was determined according to a published procedure with minor changes. Preparation of liposomal gel of fullerene C_{60} and fullerene derivatives 7-14. Liposomes were composed of tested compounds and soybean lecithin in 1:4 mass ratio. Measured fullerene or fullerene bisadduct (0.1-1 mg) and fourfold mass of lecithin are solubilized in minimal volume of PhMe under the ultrasound for 1 minute. Solvent was evaporated and film of lipid-fullerene complex carefully diluted on vortex with deionized water to the concentration of the fullerenic component of 0.02 mg/mL. The final concentration of the pure compound of 0.002 mg/mL was obtained prior to use mixing the solution with water in 1:9 ratio.

¹ M. B. Lens, E. De Marni, R. Gullo, U. Citernesi and R. Crippa, WO 043074 A1, 2007.

FOX reagent preparation². Working FOX reagent was preared by adding 10 mL of Reagent 2 (98 mg of $(NH_4)_2Fe(SO_4)_2x6H_2O$ (FAS) in 100 mL of 250mM H_2SO_4) to 900 mL of Reagent 1 (95 mg of xylenol orange sodium salt (XO) and 880 mg of 2,6-ditbutyl-4-methylphenol (BHT) in 900 mL of MeOH) giving the final concentrations of 250 μ M FAS, 125 μ M of XO, 25 mM H_2SO_4 , and 4 mM BHT. The reagent was consumed within 24 h. The apsorbance was measured at 560 nm by UV-vis spectrophotometer *GBC-Cintra 40* with 90% MeOH as a zero probe.

The applicability of the method in used range of peroxide concentration was confirmed by preparing standard calibration curve using increasing concentrations of peroxide (TBHP or H_2O_2 ; 0-200 μ M) incubated with FOX reagent at room temperature for 30 min. Absorbances measured at 560 nm at the different concentrations confirmed linear correlation.

Sample preparation. The fullerenesomes and vitamin C solutions (0.02 mg/mL) were diluted by nine-fold volume of water to gain 0.002 mg/mL concentration prior to use (0.050 mL : 0.450 mL of water). The same volume of 200 μ M peroxide (obtained by diluting 0.050 mL of 2 mM peroxide with 0.450 mL of water) was added to the sample and vortexed for 1 min. After 10 min of incubation at room temperature, to an aliquot of 0.050 mL of the sample 0.950 mL of FOX reagent was added. Absorbance at 560 mm was determined for each sample after 80 min. of incubation at room temperature.

Standard probe preparation. The standard probe of peroxides were prepared by mixing the same volume of 200 μM peroxide (obtained by diluting 0.050 mL of 2 mM peroxide with 0.450 mL of water) and water. To a 0.050mL of mixture 0.950 mL of FOX reagent was added. The absorbance of the standard probe, determined after 80 min., reffers to the starting (maximum) concentration of the peroxide, prior to incubation. Difference of absorbances of the standard probe (As) and sample (A) is proportional to the quantity of the consumed peroxide by the sample compound.

Blank probe preparation. The blank probe contained 0.950 mL of FOX reagent and 0.050 mL of water. Apsorbance of the blank probe measured at 560 nm (A_0) reffers to the color of the reagent itself in the absence of the peroxide, and all absorbances of the samples and standards are diminished by the value of A_0 for the calculations of the peroxide concentration.

All experiments were performed in triplicates, and the average values were taken.

Antioxidative capacities were calculated according to formula (1):

$$\Delta (\%) = 100 \times (A - A_s) / (A_s - A_0), \tag{1}$$

where A_0 , As and A are apsorbance values determined at the same conditions for blank probe, standard peroxide solution and probe, respectively.

The antioxidant activities relative to the equimolar concentration of vitamin C were calculated using the equation (2):

$$AOA_{mol} \text{ vs vit } C = (\Delta/\Delta_{vit C})/(M/M_{vit C})$$
(2)

where Δ and Δ vit C represent the direct antioxidant capacity of the tested compound and vitamin C, respectively and M and $M_{vit C}$ their molecular weights.

The antioxidant activities relative to the equimolar concentration of the fullerene C_{60} were calculated using the equation (3):

$$AOA_{mol} \text{ vs } C_{60} = (\Delta/\Delta_{C60})/(M/M_{C60})$$
 (3)

where Δ and ΔC_{60} represent the direct antioxidant capacity of the tested compound and the C_{60} , respectively and M and M_{C60} their molecular weights.

² S. Gao, M. Miller, X. Q. Han, EP 1593685 A1, 2005.

2. Synthesis of the compounds $3-6^3$

Dibenzyl-*N*,*N*′-(3,6-dioxaoctane-1,8-diyl)diglycinate (3). To an ice-cooled solution of diamine 1 (4.06 g, 4.00 mL, 0.027 mol, 1 mol equiv) and TEA (5.53 g, 7.80 mL, 0.055 mol, 2 mol equiv) in DCM (160 mL), solution of BBA (12.6 g, 8.60 mL, 0.055 mol, 2 mol equiv) in DCM (80 mL), was added dropwise, during 5h. After additional stirring for 24h, mixture was washed with H₂O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. Na₂SO₄. The solvent was removed in vacuo and the remaining material was purified on a SiO₂ column by dry-flash chromatography. Dibenzyl-*N*,*N*′-(3,6-dioxaoctane-1,8-diyl)diglycinate (3) was isolated as a colourless oil (4.94 g, 41%) using EtOAc/MeOH 9:1 as an eluent. IR (ATR): \tilde{v} /cm⁻¹ 3379, 3031, 2872, 1745, 1665, 1456, 1353, 1198, 1117, 1023, 746, 702. NMR: δH (500 MHz, CDCl₃, Me₄Si): 7.34 (5H, *s*, CH^{Δr}); 5.15 (2H, *s*, CH₂^{Bn}); 3.61 (2H, *s*, CH₂²); 3.51 (2H, *s*, CH₂^{Gly}); 3.31 (1H, *br s*, N*H*); 2.83 ppm (2H, *t*, *J*=5.0 Hz, CH₂¹). δC (125 MHz, CDCl₃, Me₄Si): 171.88 (C=O); 135.49 (C_q^{Ar}); 128.49; 128.28; and 128.25 (CH^{Ar}); 70.22 and 70.04 (CH₂^{2,4}); 66.48 (CH₂^{Bn}); 50.62 (CH₂^{Gly}); 48.63 ppm (CH₂¹). HR-MS: *m/z* calc. for [C₂₄H₃₃N₂O₆+H]⁺: 445.23331, measured 445.23183; calc. for [C₂₄H₃₂N₂O₆+Na]⁺: 467.21526, measured 467.21342.

Dibenzyl-*N*,*N*'-(4,7,10-trioxatridecane-1,13-diyl)diglycinate (4) To an ice-cooled solution of diamine **7** (4.02 g; 4.00 mL; 0.018 mol; 1 mol equiv) and TEA (3.68 g; 5.04 mL; 0.036 mol; 2 mol equiv) in DCM (112 mL), solution of BBA (8.34 g; 5.72 ml; 0.036 mol; 2 mol equiv) in DCM (56 mL), was added dropwise, during 6h. After additional stirring for 20h, mixture was washed with H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL) and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL), and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL), and then with brine (2 x100 mL), and dried over anh. H_2O (3 x100 mL), and then with brine (2 x100 mL), and dried over anh. H_2O (4,7,10-trioxatridecane-1,13-diyl) diglycinate (4) was isolated as a colourless oil (1.54 g, 33%) using EtOAc/MeOH 4:1 as an eluent. IR (ATR): \tilde{w} /cm⁻¹ 3340, 3063, 3033, 2941, 2868, 1742, 1458, 1350, 1212, 1184, 1150, 968, 750, 701. NMR: \tilde{w} (4,7,10-trioxatridecane-1,13-diyl) diglycinate (4) was isolated as a colourless oil (1.54 g, 33%) using EtOAc/MeOH 4:1 as an eluent. IR (ATR): \tilde{w} /cm⁻¹ 3340, 3063, 3033, 2941, 2868, 1742, 1458, 1350, 1212, 1184, 1150, 968, 750, 701. NMR: \tilde{w} (4,7,10-trioxatridecane-1,13-diyl) diglycinate (4) was isolated as a colourless oil (1.54 g, 33%) using EtOAc/MeOH 4:1 as an eluent. IR (ATR): \tilde{w} /cm⁻¹ 3340, 3063, 3033, 2941, 2868, 1742, 1458, 1350, 1212, 1184, 1150, 968, 750, 701. NMR: \tilde{w} (4,00 MHz, CDCl₃, Me₄Si) 7.35 ($\tilde{$

N,N'-(3,6-dioxaoctane-1,8-diyl)diglycine (5). To a solution of dibenzyl ester 3 (1.61 g; 3.622 mmol, MeOH 100 mL) 5% Pd/C was added (161 mg) and suspension was bubbled with argon. Mixture was hydrogenated at 40 psi for 20 h. After filtering the catalyst and evaporating the solvent, crude diacid **5** was isolated as colorless oil (940 mg; 98%). It was characterized spectroscopicaly and used for cycloaddition reaction without further purification. IR (ATR): \tilde{v} /cm⁻¹ 3093, 2955, 2890, 1626, 1573, 1462, 1417, 1371, 1310, 1242, 1211, 1118, 1085, 868, 600, 563. NMR: δH (500 MHz, CD₃OD, Me₄Si) 3.82 (t, t=5.0 Hz, 2H, CH₂²); 3.73 (t=5, 2H, CH₂⁴); 3.61 (t=5, 2H, CH₂^{Gly}); 3.29 (t=5, 0Hz, 2H, CH₂¹) ppm. δC (125 MHz, CD₃OD, Me₄Si) 171.54 (C=O); 71.13 (CH₂⁴); 66.95 (CH₂²); 50.43 (CH₂^{Gly}); 48.16 (CH₂¹) ppm. HR-MS: t=6.15 calc. for t=6.95 (CH₂²); 50.43 (CH₂^{Gly}); 48.16 (CH₂¹) ppm. HR-MS: t=6.11965.

N,N'-(4,7,10-trioxatridecane-1,13-diyl)diglycine (6). To a solution of dibenzyl ester 4 (840 mg; 1.626 mmol , MeOH 100 mL) 5% Pd/C was added (85 mg) and suspension was bubbled with argon. Mixture was hydrogenated at 40 psi for 20 h. After filtering the catalyst and evaporating the solvent, crude diacid 6 was isolated as a colorless oil (530 mg; 97 %). IR (ATR): \tilde{v} /cm⁻¹ 3315, 3064, 2926, 2874, 1740, 1620, 1600, 1454, 1395, 1324, 1243, 1208, 1134, 733, 697. NMR: δH (500 MHz, CD₃OD, Me₄Si) 3.69-3.61 (m, 6H,CH₂^{3,5,6}); 3.52 (s, 2H, CH₂^{Gly}); 3.18 (t, J=5.0 Hz, 2H, CH₂¹); 1.98 ppm (quint, J=6.0, 2H, CH₂²). δC (125 MHz, CD₃OD, Me₄Si) 171.28 (C=O); 71.50 (CH₂^{5,6}); 70.73 (CH₂³); 50.98 (CH₂^{Gly}); 48.32 (CH₂¹); 27.23 ppm (CH₂²). HR-MS: m/z calc. for [C₁₄H₃₀N₂O₇+2H]²⁺: 169.10210, measured 169.10168; calc. for [C₁₄H₂₉N₂O₇+H]⁺: 337.19693, measured 337.19538; calc. for [C₁₄H₂₈N₂O₇+Na]⁺: 359.17887, measured 359.17724.

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³ T. Kop, M. Bjelaković and D. Milić, *Tetrahedron*, 2015, **71**, 4801-4809.

3. Synthesis of the fulleropyrrolidines 7-14³

Bisadducts 7-9. A suspension of C₆₀ (545 mg; 0.757 mmol; 1 mol equiv), diglycine **5** (200 mg; 0.757 mmol; 1 mol equiv) and HCHO (230 mg; 7.570 mmol; 10 mol equiv) in ODCB (150 mL) was maintained at 160°C during 4 h. The obtained reaction mixture was cooled to room temperature, mixed with the same volume of hexane, deposited directly on the top of the SiO₂ column (to remove solvent without further heating) and separated by dry-flash column chromatography. Elution with toluene yielded unreacted C₆₀ (220 mg; 40.4%). Bisadducts (187.7 mg; 27.0 %) were eluted by listed eluents: bisadduct **7** (*cis*-1, 17.3 mg; 2.5%) was eluted with PhMe/EtOAc 7:3, bisadduct **8** (*cis*-2; 124.6 mg; 17.9%) with PhMe/EtOAc 6:4 and bisadduct **9** (*cis*-3; 45.8 mg; 6.6 %) with PhMe/EtOAc 1:1. All products were purified by precipitation with MeOH from highly concentrated CS₂/DCM solutions.

Bisadduct 7 (*cis*-1): R_i =0.52 (PhMe/EtOAc 1:1); UV/Vis: λ_{max} (PhMe)/nm 330 (ε / mol⁻¹dm³cm⁻¹ 30000), 402 (7000), 427 (6000), 622 (210), 654 (200), 684 (160), 722 (160). IR (ATR) \tilde{v} /cm⁻¹ 2923, 2851, 2785, 2334, 2024, 1505, 1453, 1427, 1334, 1304, 1203, 1150, 1115, 964, 757. NMR: δH (500 MHz, CDCl₃, Me₄Si) 4.80 (d, J=10.0 Hz, 2H, CH^{pyrr}); 4.47 (d, J=8.5 Hz 2H, CH^{pyrr}); 4.01 (d, J=8.0 Hz, 2H, CH^{pyrr}); 4.00-3.92 (m, 4H, CH₂^{2,7}), 3.88-3.82 (m, 2H, CH₂^{4,5}); 3.82-3.76 (m, 2H, CH₂^{4,5}); 3.52 (d, J=10.0 Hz, 2H, CH^{pyrr}); 3.38 (ddd, J=3.0; 7.0; 13.5 Hz; 2H, CH^{1,8}); 3.11 (ddd, J=3.0; 5.5; 14.0 Hz; 2H, CH^{1,8}) ppm. δC (125 MHz, CDCl₃, Me₄Si) 151.96 (2C); 151.15 (2C); 150.58 (2C); 144.89 (2C); 147.87 (2C); 147. 08 (2C); 146.82 (2C); 146.20 (2C); 145.96 (2C); 145.92 (1C); 145.37 (2C); 145.18 (2C); 144.89 (2C); 144.84 (2C); 144.42 (2C); 144.11(2C); 143.88 (2C); 143.79 (2C); 143.59 (2C); 142.87 (2C); 142.55 (1C); 142.30 (2C); 142.24 (1C); 142.17 (2C); 141.82 (2C); 141.58 (2C); 140.61 (2C); 137.85 (1C); 135.08 (2C); 134.89 (2C); 69.46 (C^{4,5}); 68.63 (C^{2,7}); 67.59 (2sp3-C^{full}); 66.42 (2CH^{pyrr}); 66.16 (2sp3-C^{full}); 66.06 (2CH^{pyrr}); 52.91 ppm (C^{1,8}). HR-MS: m/z calc. for [C₇₀H₂₀N₂O₂+H]⁺: 921.15975, measured 921.15538.

Bisadduct 8 (*cis-2*): R_i =0.43 (PhMe/EtOAc 1:1); UV/Vis: λ_{max} (PhMe)/nm 310 (ε/ mol⁻¹dm³cm⁻¹ 40000); 374 (4900); 448 (4800); 487 (3000); 572 (910); 647 (430); 680 (280). IR(ATR): \tilde{v} /cm⁻¹ 2930, 2880, 2852, 2808, 2771, 1509, 1457, 1425, 1347, 1314, 1179, 1129, 1110, 1081, 973, 733, 526. NMR: δH (500 MHz, CDCl₃, Me₄Si) 5.34 (*dd*, *J*=10.0; 2.0 Hz, 2H, CH^{pyπ}), 4.27 (*dd*, *J*=9.0; 2.0 Hz, 2H, CH^{pyπ}); 4.02 (*ddd*, *J*=2.5; 8.0; 9.5 Hz; 2H, CH^{2.7}); 3.90-3.80 (*m*, 6H, CH₂^{2⁺,7⁺,4.5}); 3.64 (*d*, *J*=9.0 Hz; 2H, CH^{pyπ}); 3.52 (*ddd*, *J*=2.5; 5.5; 13.0 Hz; 2H, CH^{1,8}); 3.32 (*d*, *J*=10.0 Hz, 2H, CH^{pyπ}); 2.83 ppm (*ddd*, *J*=2.0, 8.0; 13.0 Hz; 2H, CH^{1,8}). δC (125 MHz, CDCl₃, Me₄Si) 159.83 (2C); 155.61 (2C); 149.12 (2C); 148.80 (1C); 148.72 (1C); 148.56 (1C); 147.49 (2C); 147.18 (2C); 147.04 (2C); 146.72 (2C); 146.58 (2C); 146.23 (2C); 146.18 (2C); 146.00 (2C); 145.74 (2C); 145.37 (2C); 145.18 (2C); 145.08 (2C); 144.60 (2C); 144.57 (2C); 144.26 (2C); 143.94 (2C); 143.81 (2C); 143.01 (2C); 141.51 (2C); 140.71 (2C); 139.05 (2C); 133.53 (2C); 132.89 (2C); 129.49 (1C); 71.88 (C^{2,7}); 70.38 (C^{4,5}); 68.50 (2CH^{pyπ}); 68.48 (2CH^{pyπ}); 67.78 (2*sp3*-C^{full}); 67.74 (2*sp3*-C^{full}); 52.55 ppm (C^{1,8}). HR-MS: *m/z* calc. for [C₇₀H₂₀N₂O₂+H]⁺: 921.15975; measured 921.16010.

Bisadduct 9 (*cis*-3): R_i =0.17 (PhMe/EtOAc 1:1); R_i =0.48 (PhMe/MeOH 4:1); UV/Vis: λ_{max} (PhMe)/nm 300 (ε / mol⁻¹dm³cm⁻¹ 47000); 331 (31000); 391 (11000); 431 (2900); 467 (2000); 548 (800); 657 (360); 732 (280). IR (ATR): \tilde{v} /cm⁻¹ 2916, 2859, 2772, 1676, 1451, 1427, 1341, 1305, 1274, 1112, 965, 759, 521. NMR: δH (500 MHz, CDCl₃, Me₄Si) 4.68 (dd, J=9.5; 2.0 Hz, 2H, CH^{pytr}); 4.47 (dd, J=9.0; J=2.0 Hz, 2H, CH^{pytr}); 4.08 (d, J=9.0 Hz, 2H, CH^{pytr}); 3.93-3.90 (m, 2H, CH^{4,5}); 3.90-3.83 (m, 4H, CH₂^{2,7}); 3.83-3.78 (m, 2H, CH^{4',5'}); 3.71 (ddd, 12.5; 9.5; 5.0 Hz; 2H, CH^{1,8}); 3.49 (d, J=9.5 Hz, 2H, CH^{pytr}); 2.85 ppm (dt, J=12.5; 4.0 Hz, 2H, CH^{1',8'}). δC (125 MHz, CDCl₃, Me₄Si) 153.95 (2C); 149.64 (2C); 149.05 (2C); 148.55 (2C); 148.23 (2C); 148.22 (2C); 147.77 (2C); 146.90 (2C); 146.59 (2C); 146.23 (2C); 146.05 (2C); 145.93 (2C); 145.70 (2C); 145.66 (2C); 145.10 (2C); 144.91 (2C); 144.71 (2C); 142.16 (2C); 142.12 (4C); 142.06 (2C); 141.70 (2C); 139.74 (2C); 138.21 (2C); 137.07 (2C); 135.36 (2C); 134.64 (2C); 130.26 (2C); 70.87 (C^{4,5}); 70.36 (2sp3-C^{full}); 69.41 (C^{2,7}); 69.09 (2CH₂^{pytr}); 66.44 (2sp3-C^{full}); 65.72 (2CH₂^{pytr}); 52.28 ppm (C^{1,8}). HR-MS: m/z calc. for [C₇₀H₂₀N₂O₂+H]⁺: 921.15975, measured 921.15997.

Compounds 10-14. A suspension of diacid 6 (255 mg; 0.757 mmol; 1 mol equiv), C₆₀ (545 mg; 0.757 mmol; 1 mol equiv) and HCHO (230 mg; 7.57 mmol; 10 mol equiv) in ODCB (150 ml) was maintained at 160°C during 4 h. The obtained reaction mixture was cooled to room temperature, mixed with the same volume of hexane, deposited directly on the top of the SiO₂ column (to remove solvent without further heating) and separated by dry-flash column chromatography (DFC). DFC yielded: C₆₀ (300 mg; 54.5%; eluent: toluene) difullerene 14 (20.5 mg; 3.2%; eluent: PhMe/EtOAc 8:2) and bisadducts (total yield 194.6 mg; 24.8 %): bisadduct 10 (*cis*-1; 18.9 mg; 2.5%), eluted with PhMe/EtOAc 7:3, bisadduct 13 (*eq*; 24.7 mg; 3.3%), eluted with PhMe/EtOAc 6:4, bisadduct 12 (*cis*-3, 60.5 mg; 7.0 %), eluted with PhMe/EtOAc 1:1 and bisadduct 11 (*cis*-2; 90.5 mg; 12.0 %) eluted also with PhMe/EtOAc 1:1. All products were purified by precipitation with MeOH from highly concentrated DCM solutions.

Bisadduct 10 (*cis*-1): R_i =0.46 (PhMe/EtOAc 1:1). UV/Vis: λ_{max} (PhMe)/nm 328 (ε/ mol⁻¹dm³cm⁻¹ 32000); 406 (7000); 430 (5900); 623 (200); 651 (170); 676 (140); 710 (140). IR (ATR): \tilde{v} /cm⁻¹ 2944, 2870, 2810, 1466, 1426, 1355, 1169, 1124, 735. NMR: δH (500 MHz, CDCl₃, Me₄Si) 4.28 (d, J=9.5 Hz, 2H, CH^{pyrr}); 4.26 (d, J=8.5 Hz, 2H, CH^{pyrr}); 4.04 (d, J=8.5 Hz, 2H, CH^{pyrr}); 3.98 (ddd, J=5.0; 8.0; 10.0 Hz; 2H, CH^{3,11}); 3.86-3.76 (m, 10H, CH^{3,11}, 4CH₂-O); 3.66 (d, J=9.5 Hz, 2H, CH^{pyrr}); 3.23 (ddd, J=7.0; 8.0; 11.5 Hz; 2H, CH^{1,13}); 2.98 (ddd, J=5.5; 8.5; 12.0 Hz; 2H, CH^{1,13}); 2.23-2.07 ppm (m, 4H, CH₂^{2,12}). δC (125 MHz, CDCl₃, Me₄Si) 151.90 (2C); 151.06 (2C); 150.19 (2C); 148.93 (2C); 148.05 (2C); 147. 24 (2C); 146.99 (2C); 146.34 (2C); 146.09 (2C); 145.91 (1C); 145.43 (2C); 145.34 (2C); 145.00 (4C); 144.57 (2C); 144.22 (2C); 144.04 (2C); 143.96 (2C); 143.72 (2C); 143.00 (2C); 142.69 (1C); 142.43 (2C); 142.34 (1C); 142.30 (2C); 141.91 (2C); 141.56 (2C); 140.77 (2C); 137.94 (1C); 135.24 (2C); 135.06 (2C), 70.91 (C-O); 70.22 (C-O); 69.34 (C^{3,11}); 68.46 (2CH₂^{pyrr}); 68.10 (2sp3-C^{full}); 66.59 (2CH₂^{pyrr}); 65.73 (2sp3-C^{full}); 52.47 (C^{1,13}); 28.87 ppm (C^{2,12}). HR-MS: m/z calc. for [C₇₄H₂₉N₂O₃+H]⁺: 993.21727, measured 993.21561.

Bisadduct 11 (*cis-2*): R_i =0.16 (PhMe/EtOAc 1:1); R_i =0.46 (PhMe/MeOH 4:1). UV/Vis: λ_{max} (PhMe)/nm 310 (ε/mol⁻¹dm³cm⁻¹ 43000); 375 (5300); 448 (5100); 483 (3000); 578 (900); 643 (420); 679 (280). IR(ATR): \tilde{v} /cm⁻¹ 2879, 2775, 1452, 1345, 1244, 1120, 1093, 914, 724, 526. NMR: δH (500 MHz, CDCl₃, Me₄Si) 4.01 (d, J=9.0 Hz; 2H, CH^{pytr}); 3.96 (d, J=9.0 Hz; 2H, CH^{pytr}); 3.83-3.75 (m, 4H, CH₂^{3,11}); 3.81 (d, J=9.5 Hz; 2H, CH^{pytr}); 3.81-3.70 (m, 4H, 2CH₂-O); 3.72-3.67 (m, 4H, 2CH₂-O) 3.16 (dt, J=12.0; 7.0 Hz; 2H, CH^{1,13}); 2.90 (dt, J=12.0; 6.0 Hz; 2H, CH^{1',13'}); 2.07 ppm (quint, J=5.5 Hz; 4H, CH₂^{2,12}). δC (125 MHz, CDCl₃, Me₄Si) 159.01 (2C); 156.41 (2C); 149.27 (2C); 148.86 (1C); 148.78 (1C); 148.27 (1C), 147.93 (2C); 147.64 (2C); 147.51 (2C); 147.05 (2C); 146.89 (2C); 146.46 (2C); 146.11 (2C), 145.71 (2C); 145.70 (2C); 145.65 (2C); 145.34 (2C); 145.15 (2C); 144.83 (2C); 144.457 (2C); 144.43 (2C); 144.18 (2C); 144.01 (2C); 142.98 (2C); 141.63 (2C); 140.94 (2C); 138.86 (2C); 133.79 (2C); 133.05 (2C); 129.18 (1C); 70.69 (C-O); 70.58 (C-O); 68.76 (C^{3,11}); 68.22 (2CH^{pytr}); 67.43 (2CH^{pytr}); 67.35 (2sp3-C^{full}); 67.07 (2sp3-C^{full}); 50.96 (C^{1,13}); 28.69 ppm (C^{2,12}). HR-MS: m/z calc. for [C_{74} H₂₉N₂O₃+H]⁺: 993.21727, measured 993.21682.

Bisadduct 12 (*cis-3*): R_i =0.34 (PhMe/EtOAc 1:1). UV/Vis: λ_{max} (PhMe)/nm 299 (ε/ mol⁻¹dm³cm⁻¹ 49000); 330 (34000); 398 (9600), 435 (3900); 464 (2600); 551 (1400); 640 (470); 729 (320). IR(ATR): \tilde{v} /cm⁻¹ 2944, 2864, 2801, 2778, 1455, 1343, 1120, 767, 526. NMR: δH (500 MHz, CDCl₃, Me₄Si) 4.36 (*dd*, *J*=9.0; 1.5 Hz, 2H, CH^{pytr}); 4.28 (*dd*, *J*=9.5; 1.5 Hz, 2H, CH^{pytr}); 3.90-3.84 (*m*, 2H, CH^{3,11}); 3.85-3.75 (*m*, 8H, 4CH₂-O); 3.78 (*d*, *J*=9.0 Hz, 2H, CH^{pytr}); 3.70 (*dt*, *J*=10.0; 5.5 Hz; 2H, CH^{3',11'}); 3.61 (*d*, *J*=9.5 Hz, 2H, CH^{pytr}); 3.25 (*dt*, *J*=12.0; 7.0 Hz; 2H, CH^{1,13}); 2.81 (*dt*, *J*=12.0; 6.0 Hz; 2H, CH^{1',13'}); 2.11-1.99 ppm (*m*, 4H, CH₂^{2,12}). δC (125 MHz, CDCl₃, Me₄Si) 153.49(2C); 149.79(2C); 149.09(2C); 148.61(2C); 148.29(2C); 148.08(2C); 147.78(2C); 147.03(2C); 146.26(2C); 146.15(2C); 146.12(2C); 145.98(2C); 145.76(2C); 145.62(2C); 145.18(2C); 144.98(2C); 144.85(2C); 142.16 (4C); 142.13 (2C); 142.07 (2C); 141.65(2C); 139.79(2C); 138.37(2C); 137.11 (2C); 135.10(2C); 134.05(2C); 130.49(2C); 71.09 (C-O); 70.58 (C-O); 69.69 (2*sp*3-C^{full}); 69.11 (2CH₂^{pytr}); 68.29 (C^{3,11}); 67.33 (2CH₂^{pytr}); 65.66 (2*sp*3-C^{full}); 50.06 (C^{1,13}); 28.40 ppm (C^{2,12}). HR-MS: *m/z* calc. for [C₇₄H₂₉N₂O₃+H]⁺: 993.21727; measured 993.21515.

Bisadduct 13 (*eq*): R_i =0.36 (PhMe/EtOAc 1:1); UV/Vis: λ_{max} (PhMe)/nm 319 (ε/ mol⁻¹dm³cm⁻¹ 42000); 399 (6700); 423 (6100); 456 (5900); 553 (1300); 584 (980); 627 (410); 710 (100). IR(ATR): \tilde{v} /cm⁻¹ 3048, 2947, 2870, 2804, 1677, 1474, 1345, 1235, 1175, 1126, 771, 738, 529. NMR: δH (500 MHz, CDCl₃, Me₄Si) 4.42 (*dd*, *J*=9.0; 1.0 Hz, 2H, CH^{pytr-2}); 4.11 (*s*, 2H, CH₂^{pytr-1}); 4.04 (*s*, 2H, CH₂^{pytr-1}); 3.80 (*dd*, *J*=9.0; 1.0 Hz, 2H, CH^{pytr-2}); 3.81-3.77 (*m*, 2H, CH₂³); 3.78 (*t*, *J*=6.0; 2H, CH₂¹¹); 3.68 (*dd*, *J*=7.0;

6.5 Hz; 2H, CH₂-O); 3.64-3.61 (m, 2H, CH₂-O); 3.53 (dd, J=7.5; 6.0, 2H, CH₂-O); 3.51-3.48 (m, 2H, CH₂-O); 3.10 (t, J=6.0 Hz; 2H, CH₂¹³); 3.08 (t, J=6.0 Hz; 2H, CH₂¹); 2.04 (quint, J=6.0 Hz; 2H, CH₂¹²), 1.98 (quint, J=6.0 Hz; 2H, CH₂²) ppm. δ C (125 MHz, CDCl₃, Me₄Si) 159.39 (2C); 154.59 (2C); 153.72 (2C); 152.72 (2C); 149.67 (1C); 148.84 (2C); 148.10 (2C); 147.71 (1C); 147.54 (2C); 147.44 (2C); 147.37 (2C); 147.17 (2C); 146.02 (2C); 145.77 (2C); 145.15 (2C); 145.03 (2C); 144.69 (2C); 144.62 (2C); 144.40 (2C); 143.54 (2C); 143.33 (2C); 142.30 (2C); 141.59 (2C); 141.42 (2C); 141.26 (2C); 140.89 (2C); 138.96 (2C); 136.73 (2C); 135.54 (2C), 70.75 (2sp3-C^{full(pyrr-2)}), 70.54 (C-O),70.32 (sp3-C^{full(pyrr-1)}), 70.27 (C-O), 69.98 (sp3-C^{full(pyrr-1)}), 69.83 (C-O), 69.63 (C-O), 68.97 (C³), 68.56 (C¹¹), 68.24 (CH₂^{pyrr-1}); 67.27 (2CH₂^{pyrr-2}); 66.70 (CH₂^{pyrr-1}); 50.75 (C¹³); 50.22 (C¹); 29.22 (C¹²); 28.47 ppm (C²). HR-MS: m/z: calc. for [C₇₄H₂₉N₂O₃+H]⁺: 993.21727, measured 993.21577.

1,13-Bis(*N*-fulleropyrrolidino)-4,7,10-trioxatridecane (14): R_1 =0.73 (PhMe/EtOAc 1:1); UV/Vis: λ_{max} (PhMe)/nm 330 (ε/ mol⁻¹dm³cm⁻¹ 39000); 431 (3800); 546 (2100); 610 (1300); 698 (560). IR(KBr): \tilde{v} /cm⁻¹ 2854, 2771, 1731, 1638, 1456, 1423, 1340, 1301, 1232, 1108, 1039, 877, 766, 730, 524. NMR: δH (500 MHz, CDCl₃+CS₂, Me₄Si) 4.41 (s, 8H, CH₂^{pyrr}); 3.86 (t, J=6.5 Hz, 4H, CH₂^{3,11}); 3.77 (br s, 8H, 4CH₂-O); 3.21 (t, J=7.0 Hz, 4H, CH₂^{1,13}); 2.23 ppm (quint, J=6.5 Hz; 4H, CH₂^{2,12}). δC (125 MHz, CDCl₃+CS₂, Me₄Si): 154.91 (8C); 147.18 (4C); 146.14 (8C); 145.95 (16C); 145.57 (4C); 145.33 (8C); 145.18 (8C); 144.46 (8C); 143.01 (4C); 142.54 (8C); 142.14 (8C); 141.97 (8C); 141.79 (8C); 140.08 (8C); 136.15 (8C); 70.80 (2C-O); 70.58 (4sp3-C^{full}); 70.50 (2C-O); 69.31 (C^{3,11}); 67.89 (4CH₂^{pyrr}); 51.64 (C^{1,13}); 29.12 ppm (C^{2,12}). HR-MS: m/z calc. for [C₁₃₄H₂₉N₂O₃+H]⁺: 1713.21727, measured 1713.25252.

4. Table S1. Visible region absorption bands 400–800 nm of bisadduct isomers.

Regioizomer	$\lambda_1/arepsilon$	$\lambda_2/arepsilon$	λ 3 /ε	$\lambda_4/arepsilon$	λ 5 /ε
7 cis-1	402 /7000	427 /6000	654 /200	684 /160	722 /160
10 cis-1	406 /7000	430 /5900	651 /170	676 /140	710 /140
8 cis-2	448 /4800	487 /3000	572 /910	647 /430	680 /280
11 <i>cis</i> -2	448 /5100	483 /3000	578 /900	643 /420	679 /280
9 cis-3	431 /2900	467 /2000	548 /800	657 /360	732 /280
12 <i>cis</i> -3	435 /3900	464 /2600	551 /1400	640 /470	729 /320
13 eq	423 /6100	456 /5900	553 /1300	627 /410	710 /100

5. Table S2 ¹³C NMR chemical shifts of the fullerene moiety of the bridged bisadducts.

cis-1		cis-2		cis-3		eq
-C ₆ O ₂ -	-C ₁₀ O ₃ -	-C ₆ O ₂ -	-C ₁₀ O ₃ -	-C ₆ O ₂ -	-C ₁₀ O ₃ -	-C ₁₀ O ₃ -
151.96	151.90	159.83	159.01	153.95	153.49	159.39
151.15	151.06	155.61	156.41	149.64	149.79	154.59
150.58	150.19	149.12	149.27	149.05	149.09	153.72
148.79	148.93	148.80*	148.86*	148.55	148.61	152.72
147.87	148.05	148.72*	148.78*	148.23	148.29	149.67*
147.08	147.24	148.56*	148.27*	148.22	148.08	148.84
146.82	146.99	147.49	147.93	147.77	147.78	148.10
146.20	146.34	147.18	147.64	146.90	147.03	147.71*
145.96	146.09	147.04	147.51	146.59	146.26	147.54
145.92*	145.91*	146.72	147.05	146.23	146.15	147.44
145.37	145.43	146.58	146.89	146.05	146.12	147.37
145.18	145.34	146.23	146.46	145.93	145.98	147.17
144.89	145.00**	146.18	146.11	145.70	145.76	146.02
144.84	143.00	146.00	145.71	145.66	145.62	145.77
144.42	144.57	145.74	145.70	145.10	145.18	145.15
144.11	144.22	145.37	145.65	144.91	144.98	145.03
143.88	144.04	145.18	145.34	144.71	144.85	144.69
143.79	143.96	145.08	145.15	142.16	142.16**	144.62
143.59	143.72	144.60	144.83	142.12**	142.10	144.40
142.87	143.00	144.57	144.57	142.12	142.13	143.54
142.55*	142.69*	144.26	144.43	142.06	142.07	143.33
142.30	142.43	143.94	144.18	141.70	141.65	142.30
142.24*	142.34*	143.81	144.01	139.74	139.79	141.59
142.17	142.30	143.01	142.98	138.21	138.37	141.42
141.82	141.91	141.51	141.63	137.07	137.11	141.26
141.58	141.56	140.71	140.94	135.36	135.10	140.89
140.61	140.77	139.05	138.86	134.64	134.05	138.96
137.85*	137.94*	133.53	133.79	130.26	130.49	136.73
135.08	135.24	132.89	133.05			135.54
134.89	135.06	129.49*	129.18*			

^{*} Carbon peaks of relative intensity 1.

** Carbon peaks of relative intensity 4 (the others of relative intensity 2).

6. Table S3 ¹H NMR and ¹³C NMR chemical shifts of the non-fullerene moiety of the bridged bisadducts.

	$\begin{array}{c} \delta(sp^3 - \\ C^{\text{full}}) \end{array}$	$\delta(\mathrm{CH_2}^\mathrm{pyrr})$	$\delta \left(\mathrm{CH_2}^{\mathrm{tether}} \right)$
cis-1 (7)	67.59 66.16	4.80d(10.0); 3.52d(10.0); 66.06 4.47d(8.5); 4.01d(8.0); 66.42	CH ₂ (1,8) - 3.11ddd(3.0; 5.5; 14.0); 3.38ddd(3.0; 7.0; 13.5); 52.91 CH ₂ (2,7) - 4.00-3.92m; 68.63 CH ₂ (4,5) - 3.88-3.82m; 3.82-3.76m; 69.46
cis-2 (8)	67.78 67.74	5.34dd(10.0; 2.0); 3.32d(10.0); 68.48 4.27dd(9.0; 2.0); 3.64d(9.0); 68.50	CH ₂ (1,8) - 2.83ddd(2.0; 8.0; 13.0); 3.52ddd(2.5; 5.5; 13.0); 52.55 CH ₂ (2,7) - 4.02ddd(2.5; 8.0; 9.5); 3.80-3.90m; 71.88 CH ₂ (4,5) - 3.80-3.90m; 70.38
cis-3 (9)	70.36 66.44	4.68dd(9.5; 2.0); 3.49d(9.5); 65.72; 4.47dd(9.0; 2.0); 4.08d(9.0); 69.09	CH ₂ (1,8) - 2.85dt(12.5; 4.0); 3.71ddd(12.5; 9.5; 5.0); 52.28 CH ₂ (2,7) - 3.83-3.90m; 69.41 CH ₂ (4,5) - 3.93-3.90m; 3.78-3.83m; 70.87
cis-1 (10)	68.10 65.73	4.28d(9.5); 3.66d(9.5); 68.46; 4.26d(8.5); 4.04d(8.5); 66.59	CH ₂ (1,13) - 3.23ddd(7.0; 8.0; 11.5); 2.98ddd(5.5; 8.5; 12.0); 52.47 CH ₂ (2,12) - 2.07-2.23m; 28.87 CH ₂ (3,11) - 3.98ddd(5.0; 8.0; 10.0); 3.76-3.86m; 69.34 CH ₂ -O - 3.76-3.86m; 70.91 CH ₂ -O - 3.76-3.86m; 70.22
cis-2 (11)	67.35 67.07	4.01d(9.0); 3.96d(9.0); 67.43; 3.93d(9.5); 3.81d(9.5); 68.22	CH ₂ (1,13) - 2.90dt(12.0; 6.0); 3.16dt(12.0; 7.0); 50.96 CH ₂ (2,12) - 2.07quint(5.5); 28.69 CH ₂ (3,11) - 3.75-3.83m; 68.76 CH ₂ -O - 3.70-3.81m; 70.58 CH ₂ -O - 3.67-3.72m; 70.69
cis-3 (12)	69.69 65.66	4.36dd(9.0; 1.5); 3.78d(9.0); 67.33 4.28dd(9.5; 1.5); 3.61d(9.5); 69.11	CH ₂ (1,13) - 2.81dt(12.0; 6.0); 3.25dt(12.0; 7.0); 50.06 CH ₂ (2,12) - 1.99-2.11m; 28.40 CH ₂ (3,11) - 3.84-3.90m; 3.70dt(10; 5.5); 68.29 CH ₂ -O - 3.77-3.85m; 70.58 CH ₂ -O - 3.75-3.81m; 71.09
eq (13)	70.75(2C) 70.32(1C) 69.98(1C)	4.42dd(9.0; 1.0); 3.80dd(9.0; 1.0); 67.27 4.11s; 68.24; 4.04s; 66.70	CH ₂ (1) - 3.08t(6.0),50.22; CH ₂ (13)- 3.10t (6.0), 50.75 CH ₂ (2) - 1.98quint(6.0), 28.47; CH ₂ (12) -2.04quint (6.0), 29.22 CH ₂ (3) - 3.81-3.77m; 68.97; CH ₂ (11) – 3.78t(6.0), 68.56 CH ₂ -O - 3.64-3.61m; 70.54; CH ₂ -O - 3.51-3.48m; 70.27 CH ₂ -O - 3.53dd(7.5; 6.0); 69.83;CH ₂ -O - 3.68dd(7.0; 6.5); 69.63
Bis-C ₆₀ (14)	70.58	4.41s; 67.89	CH ₂ (1,13) - 3.21t(7.0); 51.64 CH ₂ (2,12) - 2.23quint(6.5); 29.12 CH ₂ (3,11) - 3.86t(6.5); 69.31 CH ₂ -O - 3.77br s; 70.80, 70.50
δ range	65-71	3.30-5.40 65-69	Dioxa-C10: $CH_2(1) \rightarrow 52.28-52.91$ $CH_2(2) \rightarrow 68.63-71.88$ $CH_2(4) \rightarrow 69.46-70.71$ Trioxa-C13: $CH_2(1) \rightarrow 50.06-52.47$ $CH_2(2) \rightarrow 28.40-29.22$ $CH_2(3) \rightarrow 68.29-69.34$ $CH_2(5,6) \rightarrow 69-71$

7. Figures S1-S51 NMR and HR-M spectra of compounds 3-14

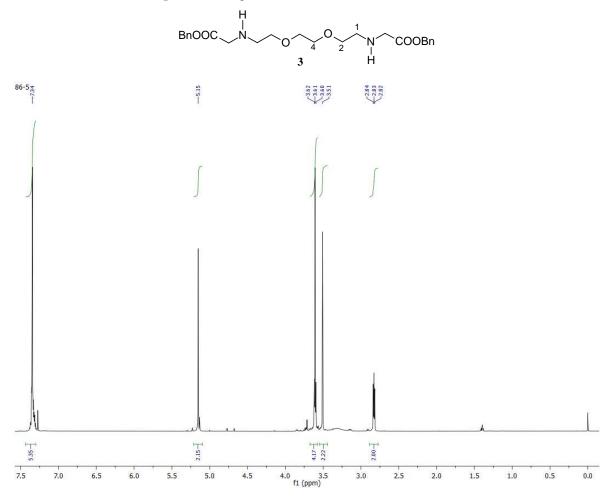


Fig.S1 1 H NMR spectrum of compound 3

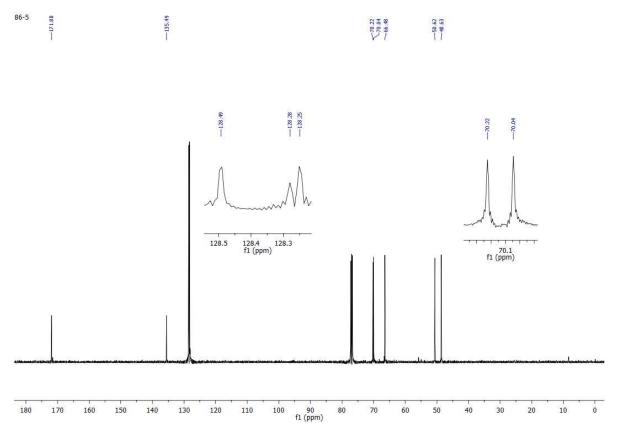
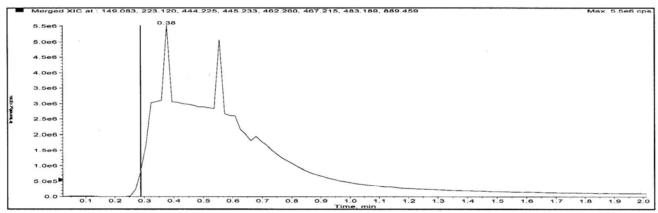
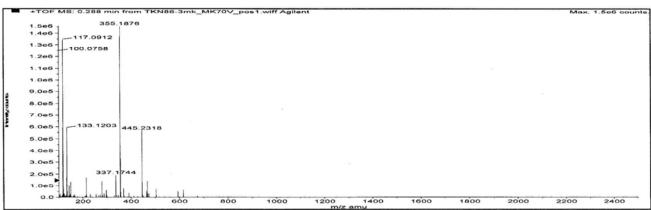
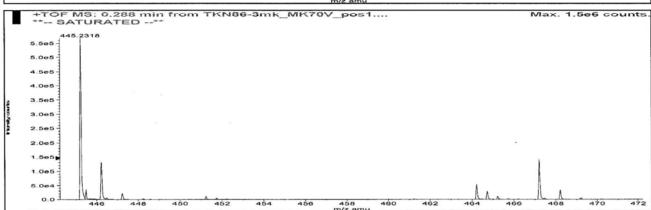


Fig.S2 ¹³C NMR spectrum of compound 3



Merged XIC, Period#:1 Experiment#:1

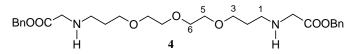




Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C24H32N2O6	-	444.22604	0.38	2.79088 E7	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+H]+	569759.42	445.23331	445.23183	-1.48025	-3.32	-
[M+Na]+	139967.85	467.21526	467.21342	-1.84058	-3.94	-

Fig. S3 HR-MS spectrum of compound 3



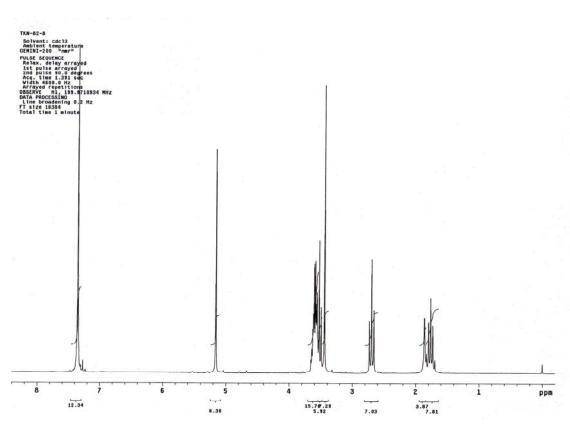


Fig.S4 ¹H NMR spectrum of compound 4

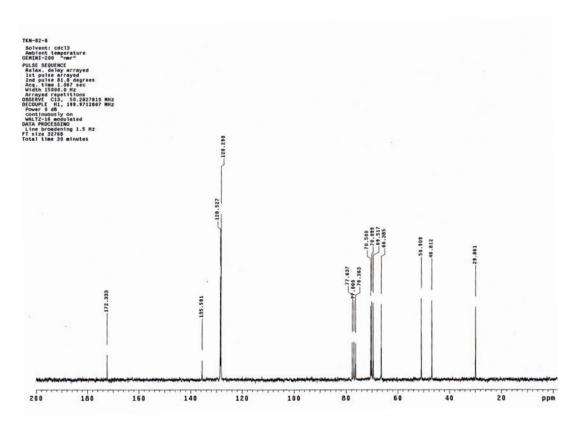
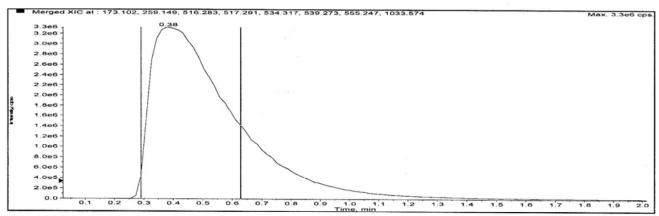


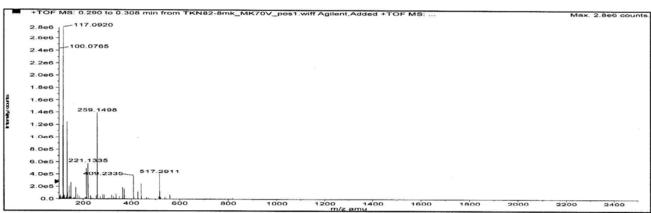
Fig.S5 ¹³C NMR spectrum of compound 4

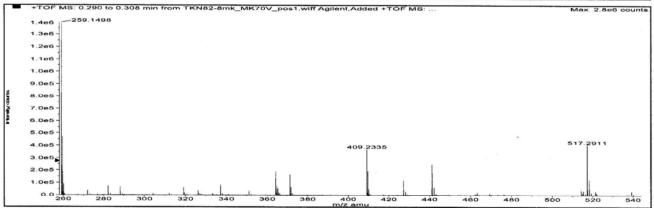
Sample Name: TKN82-8 Sample Location: P1-B5 Sample Id: Operator: Milka

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Merged XIC, Period#:1 Experiment#:1

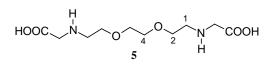




Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C28H40N2O7	-	516.28355	0.38	6.32908 E7	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+	1428936.78	259.14905	259.14980	0.75157	2.90	-
[M+H]+	430308.58	517.29083	517.29112	0.28791	0.56	
[M+Na]+	33890.98	539.27277	539.27333	0.55252	1.02	-

Fig. S6 HR-MS spectrum of compound 4



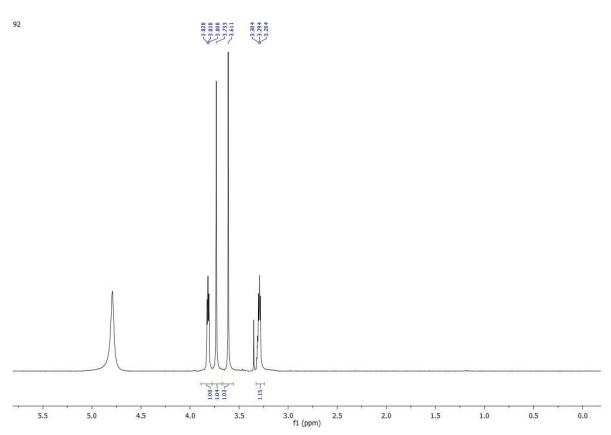


Fig.S7 1 H NMR spectrum of compound 5

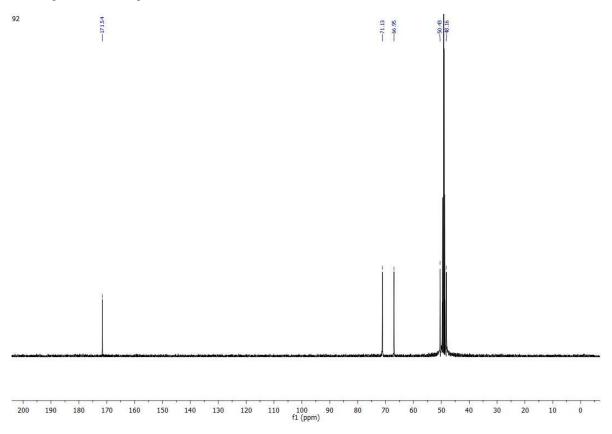
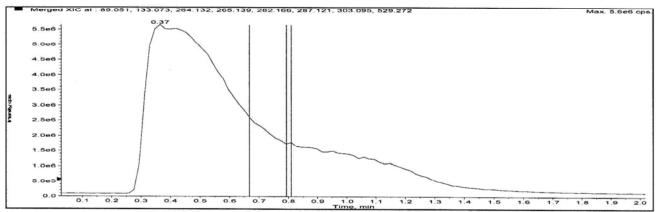


Fig.S8 ¹³C NMR spectrum of compound 5

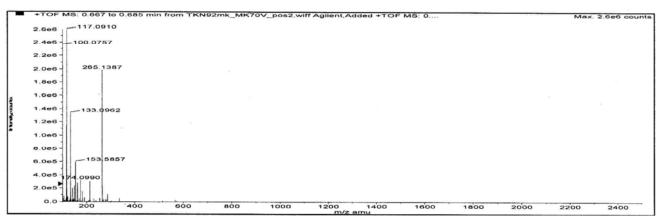
Sample Name: TKN92 Sample Location: P1-B9 Sample Id: Operator: Milka

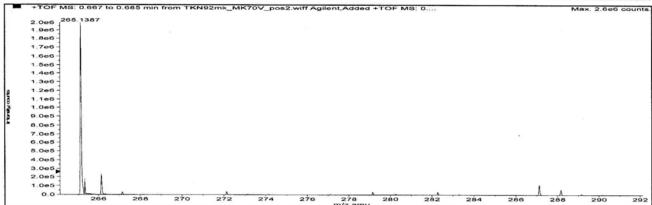
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Merged XIC, Period#:1 Experiment#:1





Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C10H20N2O6		264.13214	0.37	1.46081 E8	-

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+H]+	2082262.46	265.13941	265.13866	-0.75030	-2.83	
[M+Na]+	114103.32	287.12136	287.11965	-1.70249	-5.93	

Fig. S9 HR-MS spectrum of compound 5

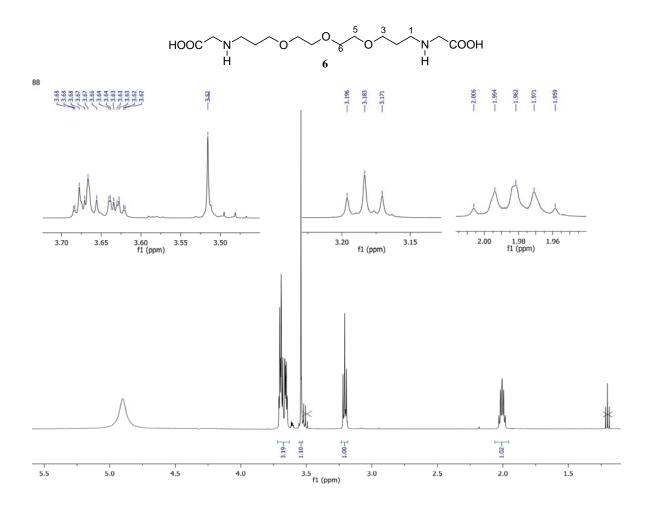


Fig.S10 ¹H NMR spectrum of compound 6

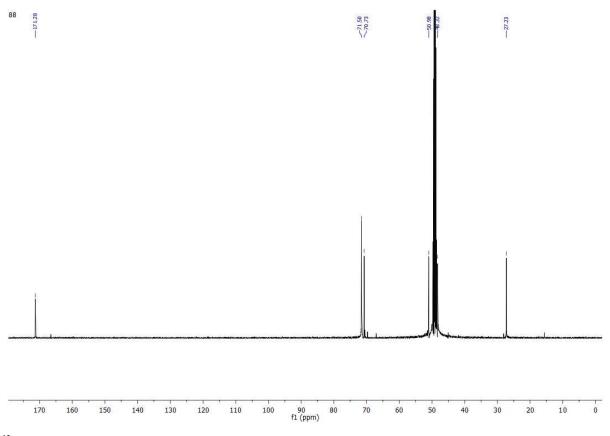
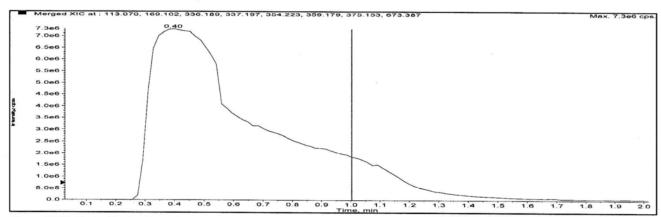


Fig.S11 13 C NMR spectrum of compound 6

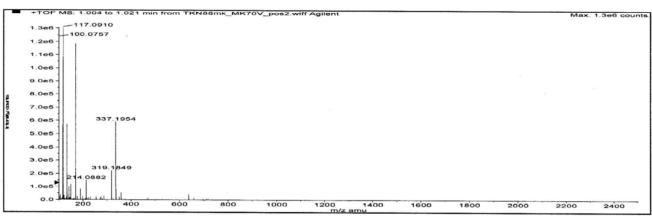
Sample Name: TKN88 Sample Location: P1-B8 Sample Id: Operator: Milka

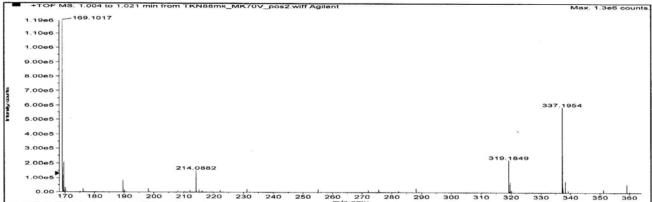
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Merged XIC, Period#:1 Experiment#:1

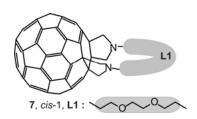




Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C14H28N2O7		336.18965	0.40	1.91919 E8	-

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+					-2.49	
[M+H]+	591544.29	337.19693	337.19538	-1.54941	-4.59	
[M+Na]+	57682.29	359.17887	359.17724	-1.63500	-4.55	-

Fig S12 HR-MS spectrum of compound 6



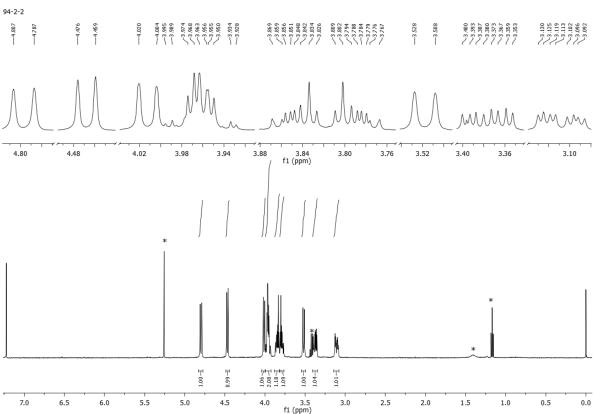


Fig.S13 ¹H NMR spectrum of compound 7

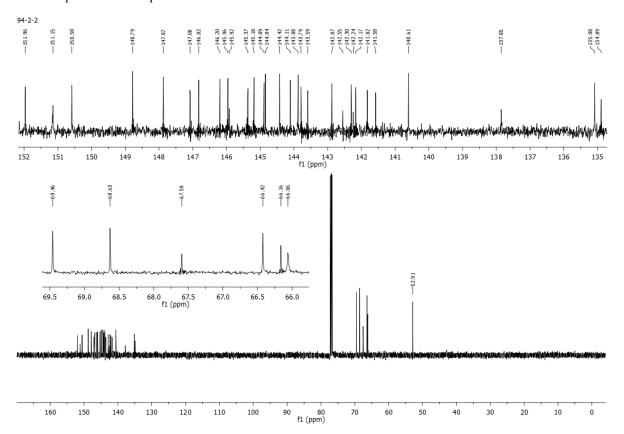


Fig.S14 ¹³C NMR spectrum of compound 7

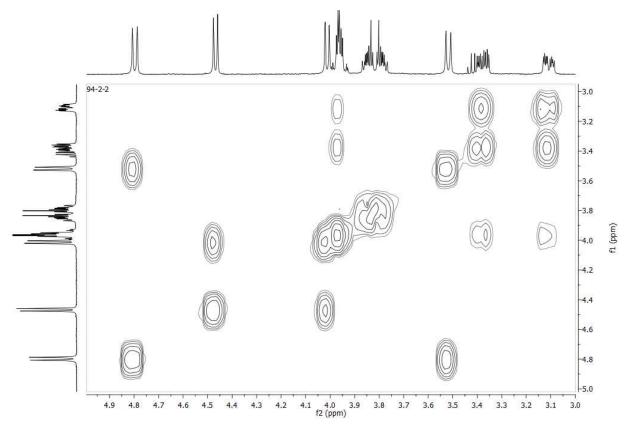


Fig S15 COSY spectrum of compound 7

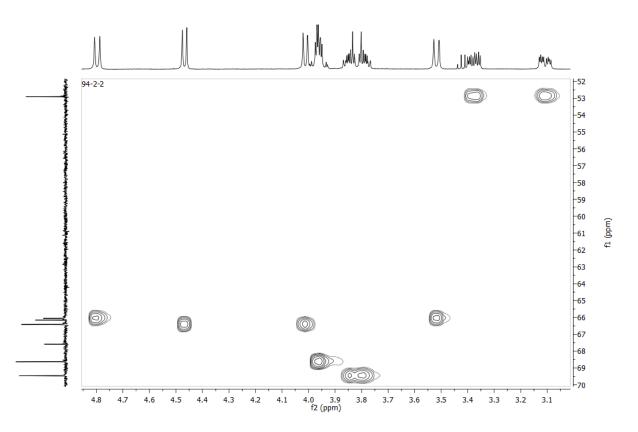
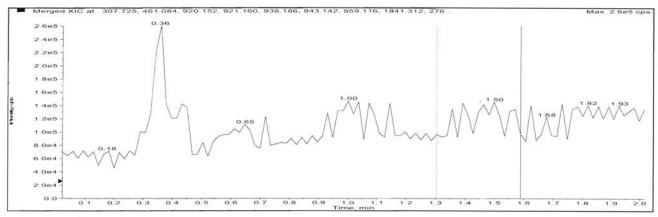


Fig S16 HSQC spectrum of compound 7

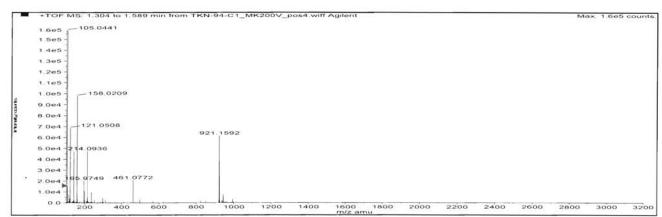
Sample Name: TKN-94-C1 Sample Location: P1-C1 Sample Id: Operator: Milka

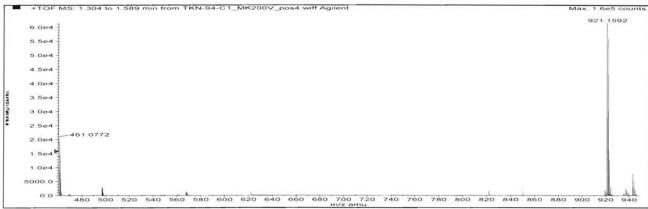
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Merged XIC, Period#:1 Experiment#:1

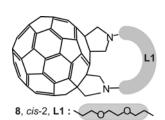




Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C70H20N2O2	-	920.15248	1.50	5.70343 E5	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+	21142.04	461.08352	461.08192	-1.59089	-3.45	_
M+	1640.21	920.15193	920.14423	-7.70308	-8.37	
[M+H]+	61804.07	921.15975	921.15538	-4.36965	-4.74	
[M+NH4]+	1902.36	938.18630	938.15406	-32.23965	-34.36	-
[M+Na]+	7683.76	943.14170	943.13855	-3.14719	-3.34	155

Fig. S17 HR-MS spectrum of compound 7



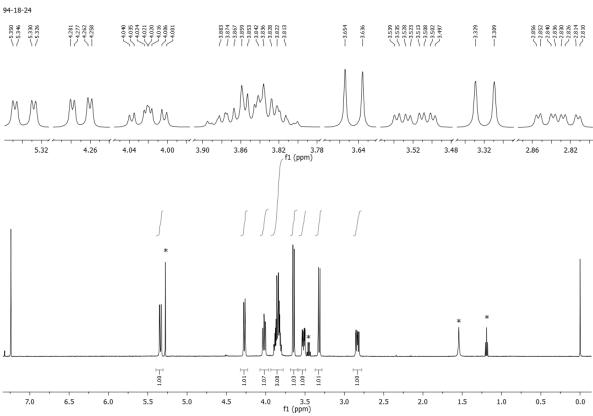


Fig.S18 ¹H NMR spectrum of compound 8

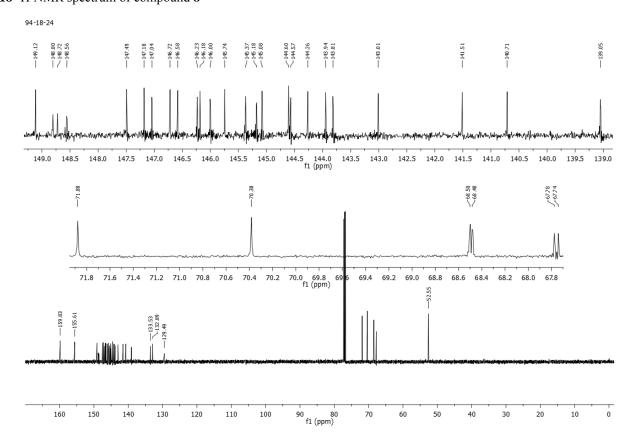


Fig.S19 13 C NMR spectrum of compound 8

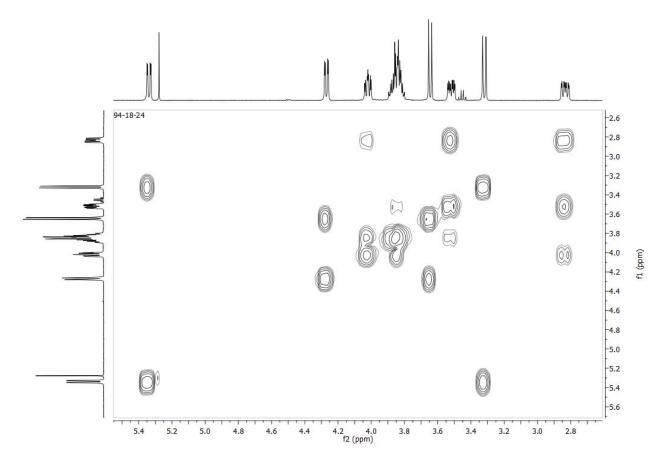


Fig. S20 COSY spectrum of compound 8

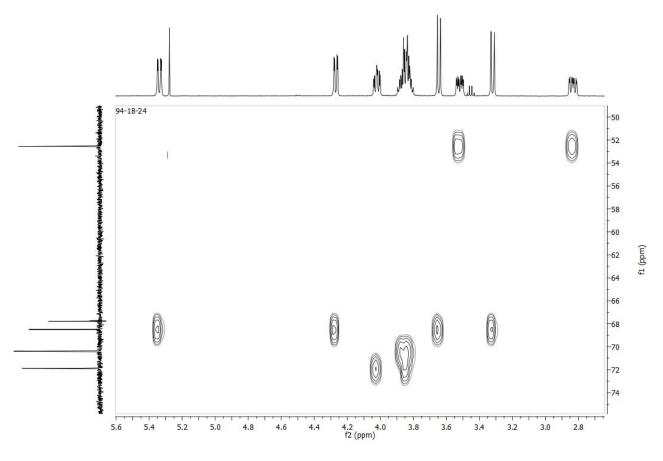
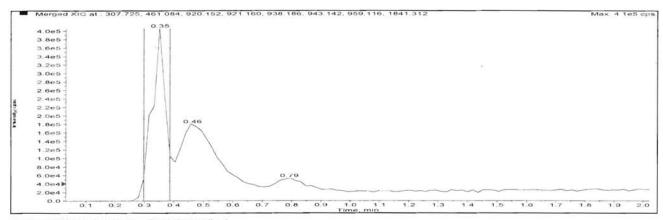


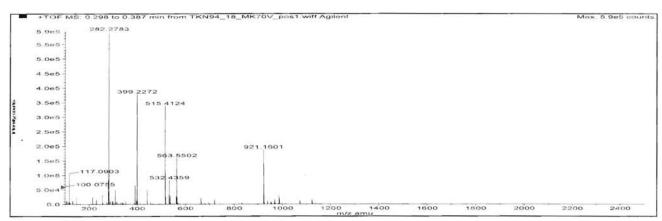
Fig. S21 HSQC spectrum of compound 8

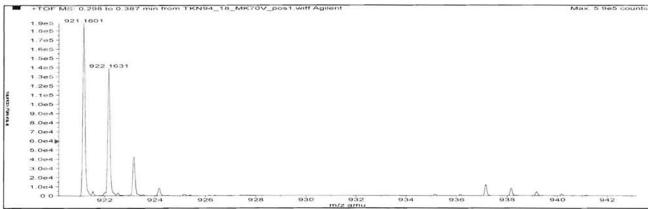
Sample Name: TKN94-18 Sample Location: P1-C3 Sample Id: Operator: Milka

Data File Name: D:\PE Sciex Data\Projects\D_Milic\Data\TKN94_18 MK70V_pos1.wiff Acq Time: April 27 2012, 11:11:04 AM Method: D:\TOF_Data\damethods\Night_Seq_Comp_ident1.anm\efc.xml



Merged XIC, Period#:1 Experiment#:1





Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C70H20N2O2		920.15248	0.35	1.00097 E6	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+H]+	188342.12	921.15975	921.16010	0.34509	0.37	
[M+NH4]+	8444.71	938.18630	938.15891	-27.39731	-29.20	-

Fig. S22 HR-MS spectrum of compound 8

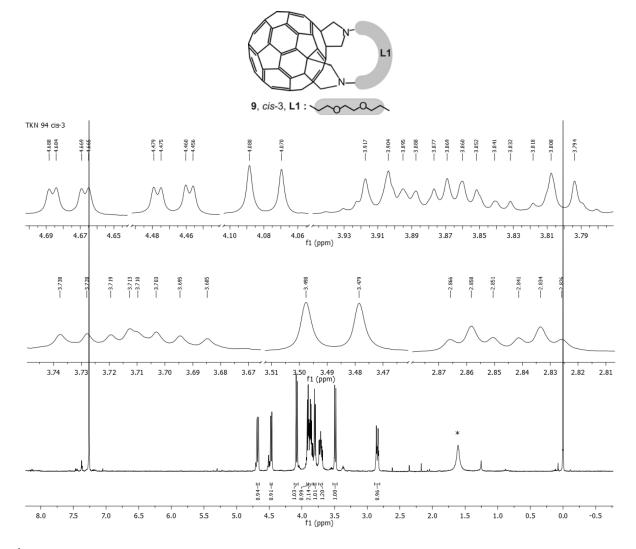


Fig.S23 ¹H NMR spectrum of compound 9

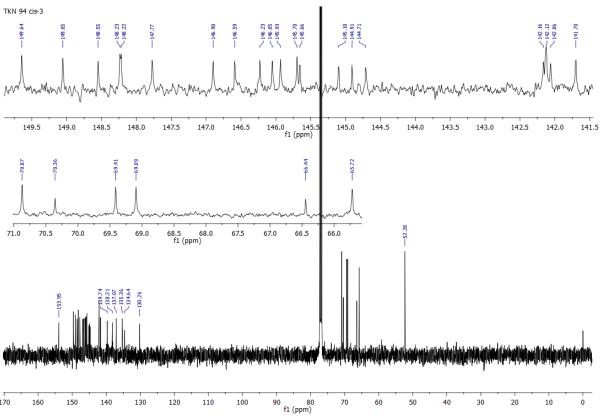


Fig.S24 ¹³C NMR spectrum of compound 9

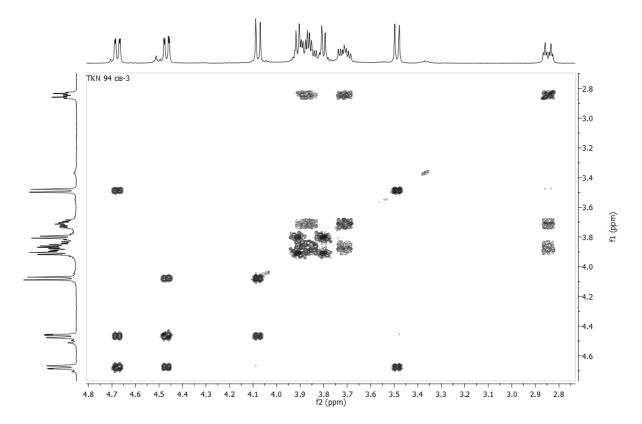


Fig. S25 COSY spectrum of compound 9

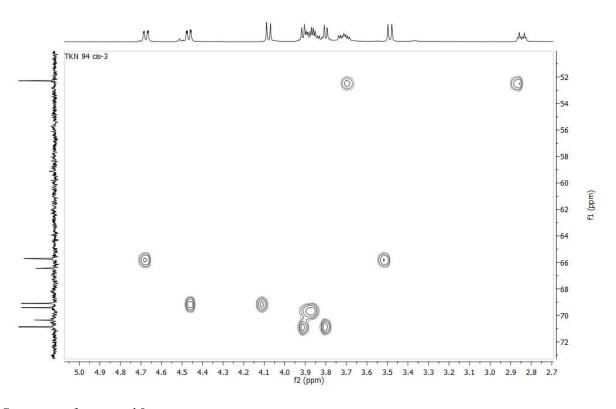
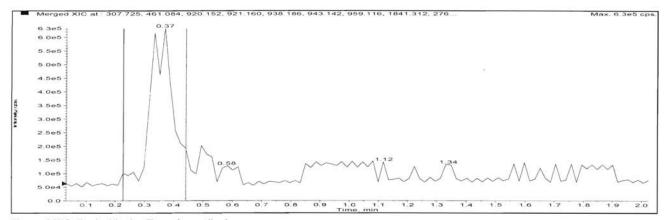
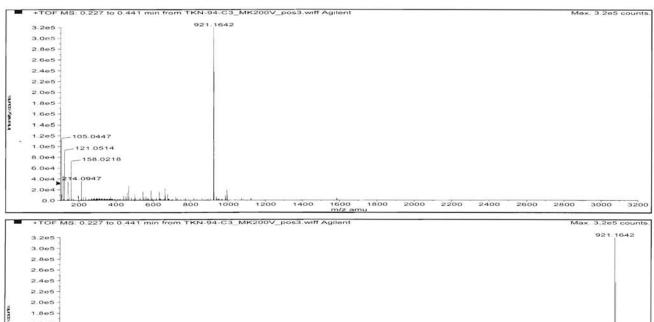


Fig. S26 HSQC spectrum of compound 9



Merged XIC, Period#:1 Experiment#:1



												r	m/z ar	nu											
	0.0	480	500	520	540	560	580	600	620	640	660	680	700	720	740	760	780	800	820	840	860	880	900	920	940
		1	- Li							1					1207100					a-v-				- 1	16
	2.0e4	01									9													- 1	
	4.064																							- 1	
	6.0e4																								
	8.064																							- 1	
	1.0e5																								
	1.2e5 -																								
3	1.465																								
4	1.6e5																								
à	1.8e5																								
8	2.0e5																								
	2.2e5																								
	2.4e5																							1	

Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C70H20N2O2	TT.	920.15248	0.37	2.86071 E6	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+	14217.34	461.08352	461.08479	1.27093	2.76	-
[M+H]+	321713.99	921.15975	921.15997	0.22049	0.24	
[M+NH4]+	5088.87	938.18630	938.16024	-26.06655	-27.78	

Fig. S27 HR-MS spectrum of compound 9

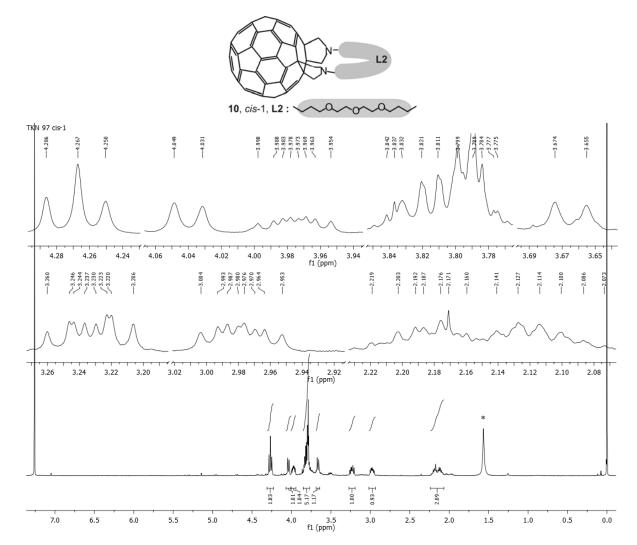


Fig.S28 ¹H NMR spectrum of compound 10

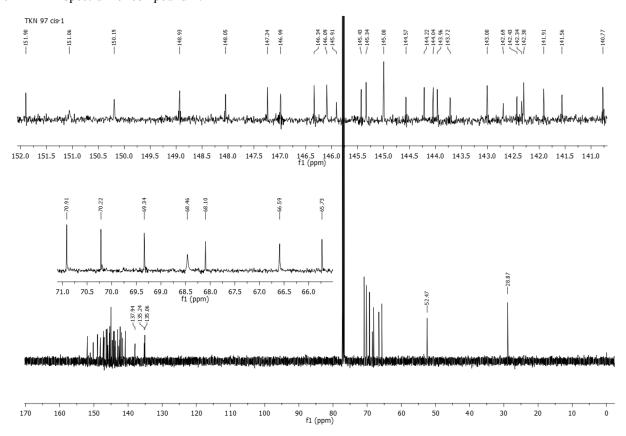


Fig.S29 ¹³C NMR spectrum of compound 10

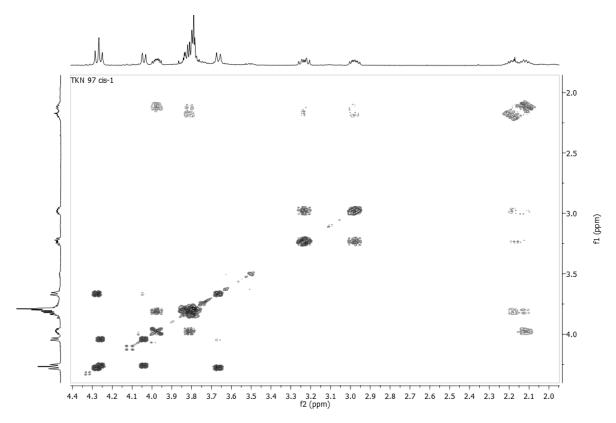


Fig. S30 COSY spectrum of compound 10

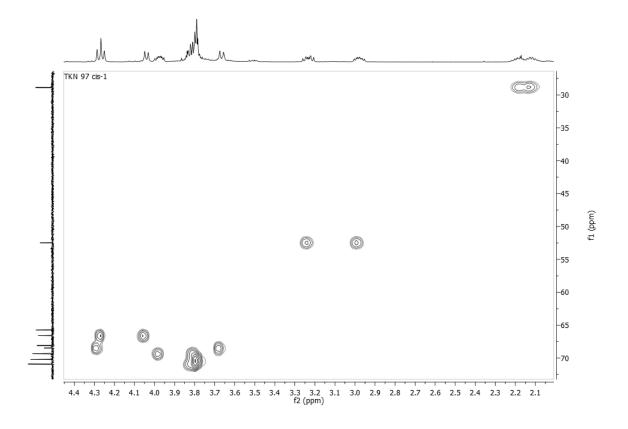
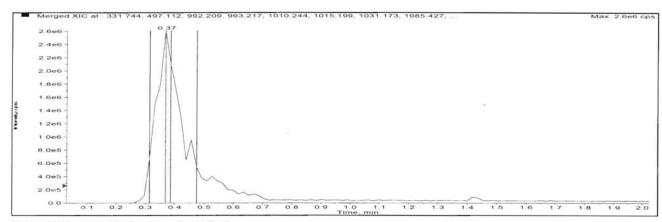


Fig. S31 HSQC spectrum of compound 10

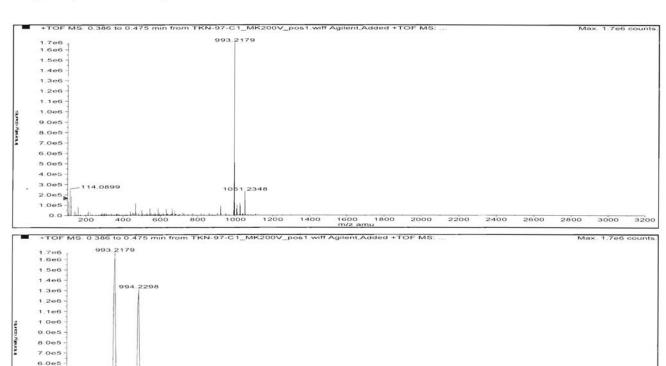
Sample Name: TKN-97-C1 Sample Location: P1-C3 Sample Id: Operator: Milka

Data File Name: D:\PE Sciex Data\Projects\D Milic\Data\TKN-97-C1 MK200V pos1.wiff Acq Time: July 28 2015, 10:19:53 AM

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Merged XIC, Period#:1 Experiment#:1

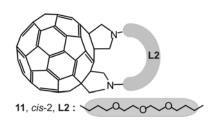


Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C74H28N2O3	-	992.20999	0.37	1.34865 E7	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
M+	36030.60	992.20944	992.20436	-5.08845	-5.13	
[M+H]+	1671238.55	993.21727	993.21561	-1.66295	-1.67	
[M+NH4]+	63146.49	1010.24382	1010.21360	-30.22208	-29.92	1 1

Fig. S32 HR-MS spectrum of compound 10

5.0e5 -4.0e5 -3.0e5 -2.0e5 -1.0e5 -



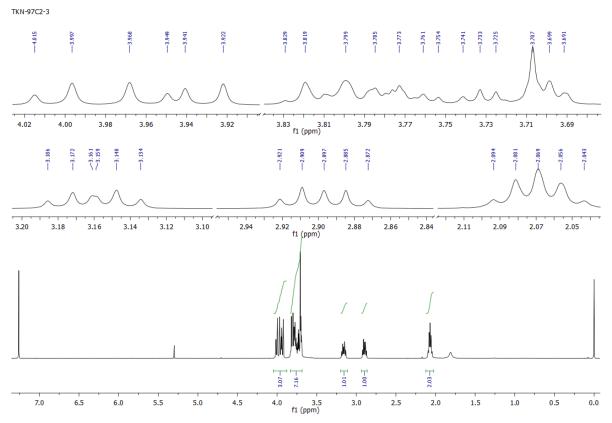


Fig.S33 ¹H NMR spectrum of compound 11

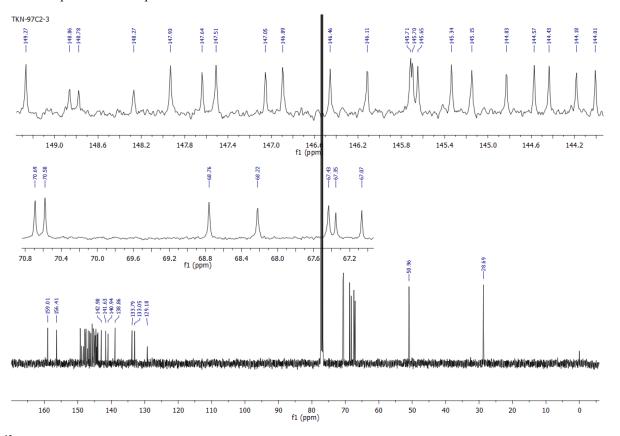


Fig.S34 ¹³C NMR spectrum of compound 11

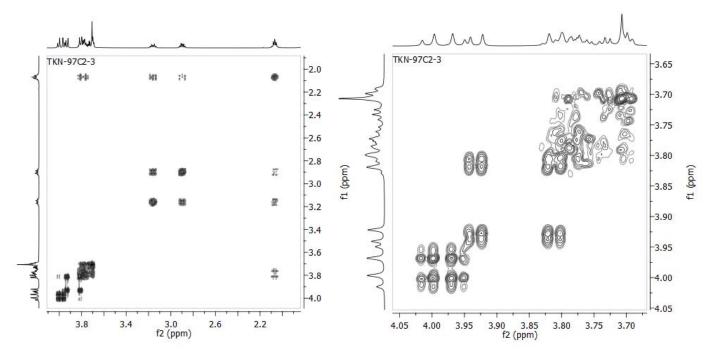


Fig. S35 COSY spectrum of compound 11

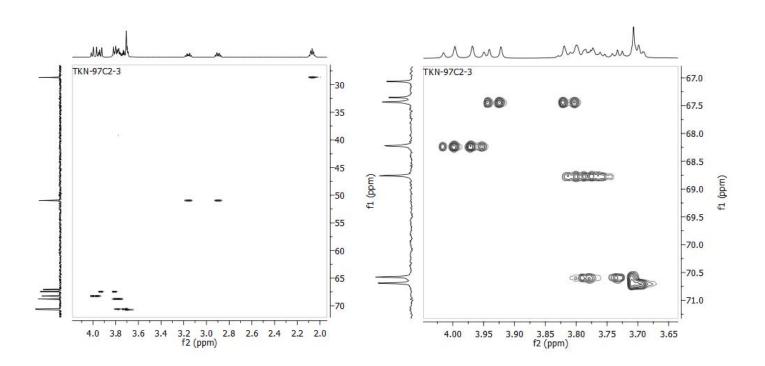
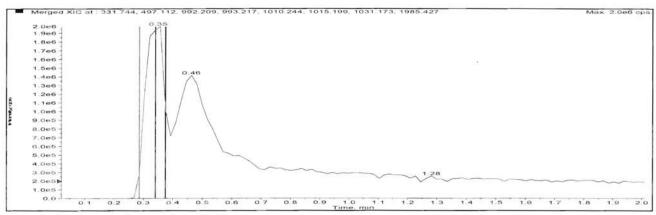
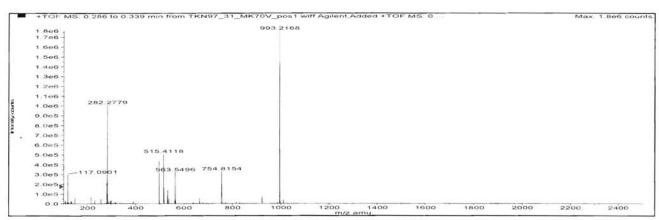


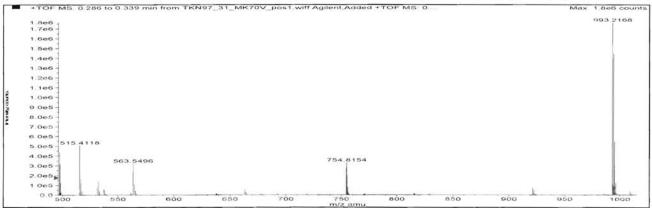
Fig. S36 HSQC spectrum of compound 11

Sample Name: TKN97-31 Sample Location: P1-C4 Sample Id: Operator: Milka Data File Name: D:\PE Sciex Data\Projects\D_Milic\Data\TKN97_31_MK70V_pos1.wiff Acq Time: April 27 2012, 11:14:17 AM Method: D:\TOF_Data\damethods\Night_Seq_Comp_ident1.anm\efc.xml



Merged XIC, Period#:1 Experiment#:1

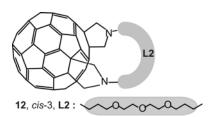




Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C74H28N2O3	্বা	992.20999	0.35	6.50638 E6	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+	437569.35	497.11227	497.11101	-1.26153	-2.54	
[M+H]+	1770072.97	993.21727	993.21682	-0.44446	-0.45	_
[M+NH4]+	30115.51	1010.24382	1010.21427	-29.55150	-29.25	

Fig. S37 HR-MS spectrum of compound 11



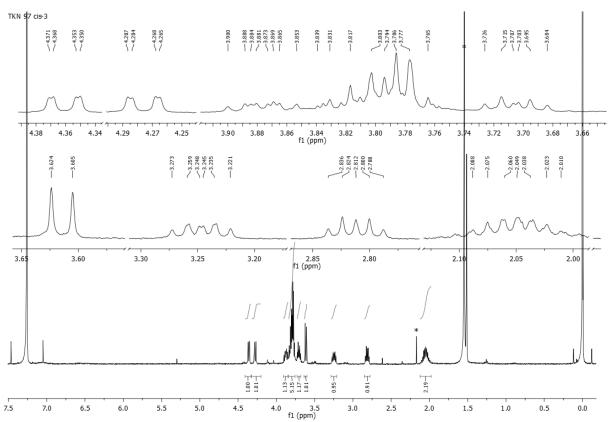


Fig.S38 ¹H NMR spectrum of compound 12

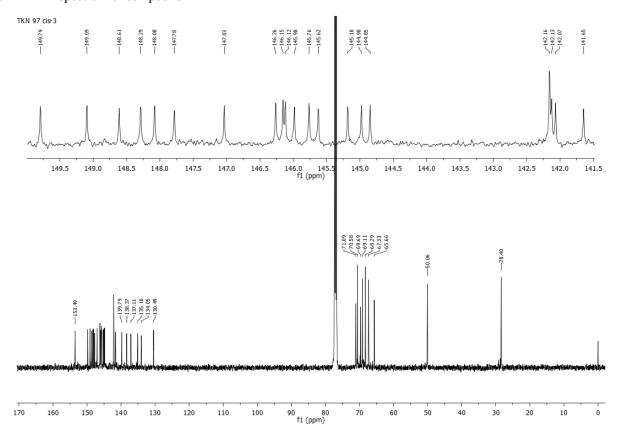


Fig.S39 ¹³C NMR spectrum of compound 12

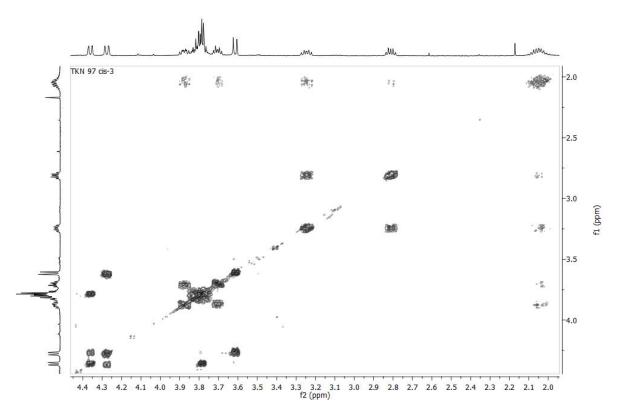


Fig. S40 COSY spectrum of compound 12

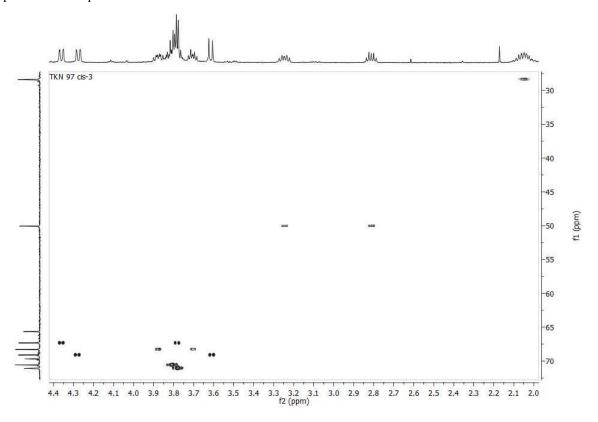
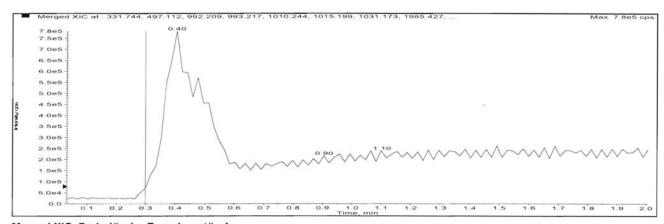
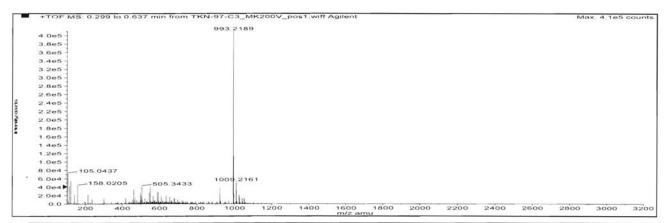
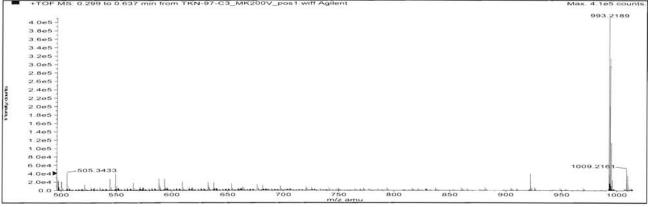


Fig. S41 HSQC spectrum of compound 12



Merged XIC, Period#:1 Experiment#:1





Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C74H28N2O3		992.20999	0.40	6.01842 E6	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+	27465.42	497.11227	497.11162	-0.64813	-1.30	
[M+H]+	411826.29	993.21727	993.21515	-2.12087	-2.14	
[M+NH4]+	37755.98	1010.24382	1010.21458	-29.23997	-28.94	

Fig. S42 HR-MS spectrum of compound 12

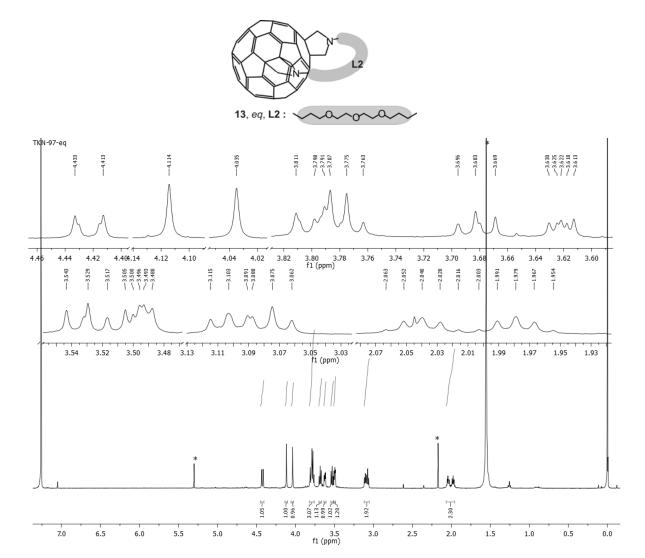


Fig.S43 ¹H NMR spectrum of compound 13

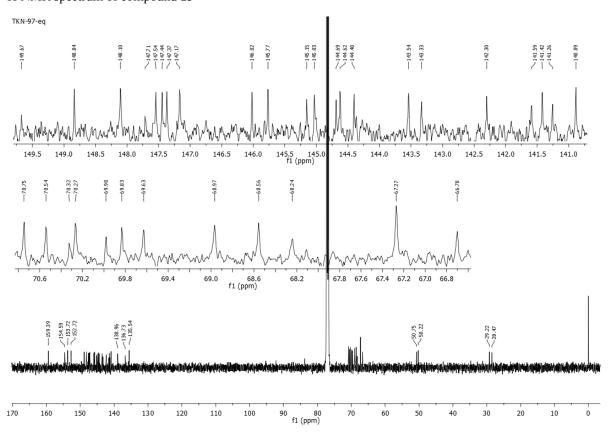


Fig.S44 ¹³C NMR spectrum of compound 13

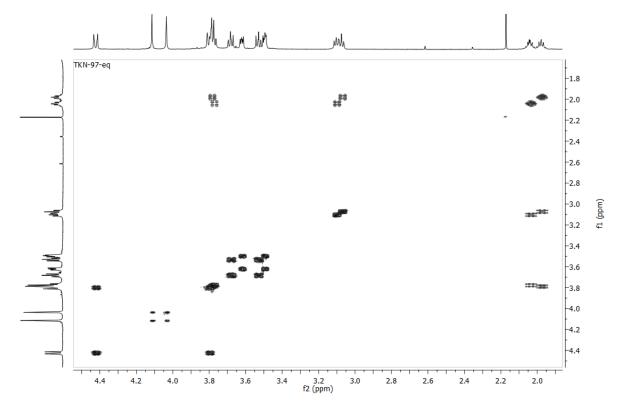


Fig. S44 COSY spectrum of compound 13

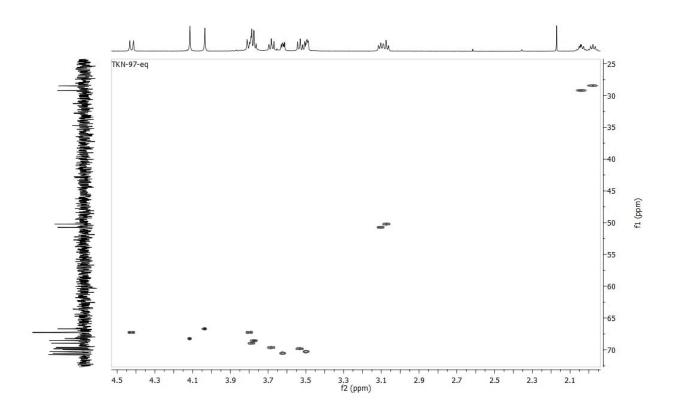
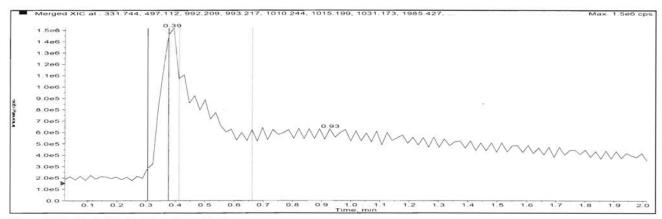
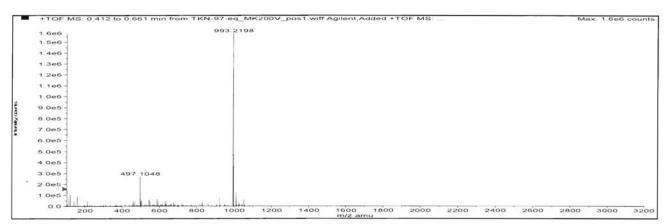
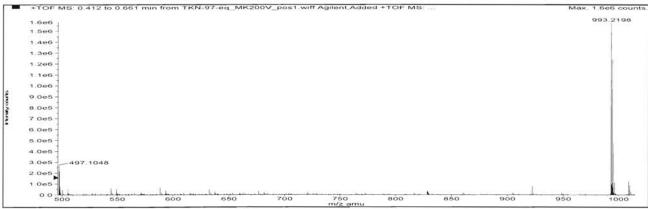


Fig. S45 HSQC spectrum of compound 13



Merged XIC, Period#:1 Experiment#:1





Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C74H28N2O3		992.20999	0.39	9.92219 E6	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+2H]2+	277699.62	497.11227	497.11116	-1.11351	-2.24	-
[M+H]+	1587182.96	993.21727	993.21577	-1.49781	-1.51	
[M+NH4]+_	92095.94	1010.24382	1010.21498	-28.83545	-28.54	

Fig. S46 HR-MS spectrum of compound 13

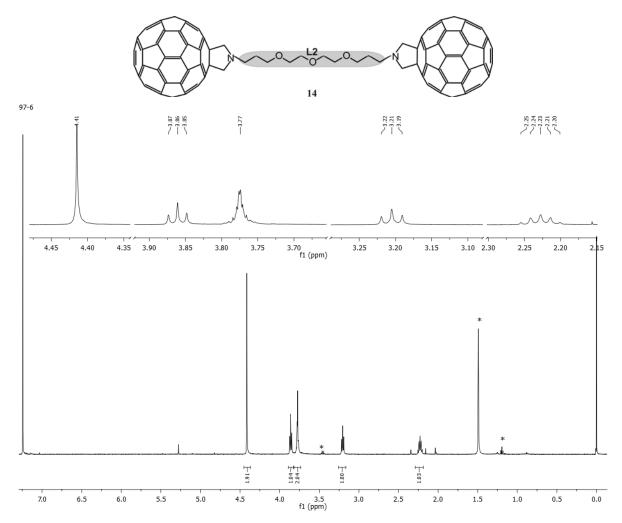


Fig.S47 ¹H NMR spectrum of compound 14

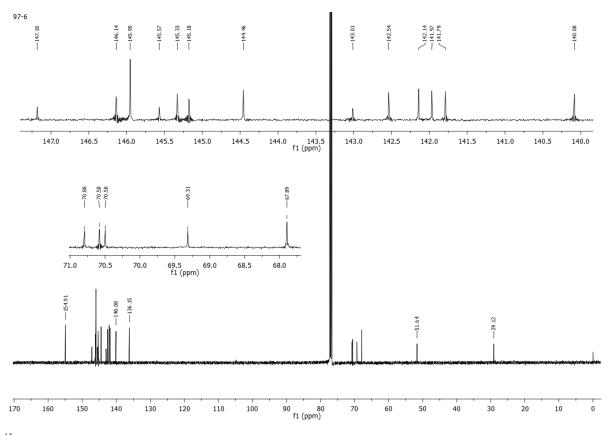


Fig.S48 ¹³C NMR spectrum of compound 14

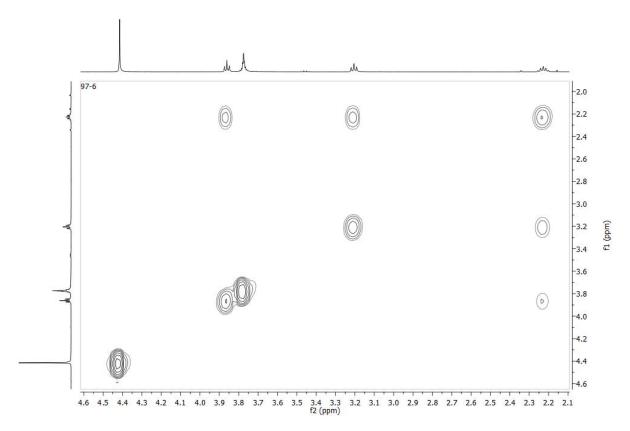


Fig. S49 COSY spectrum of compound 14

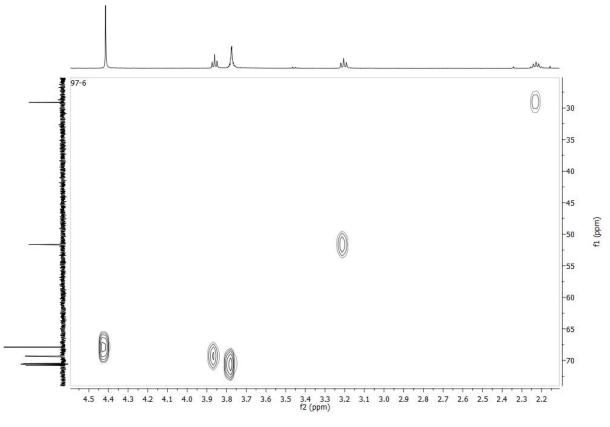


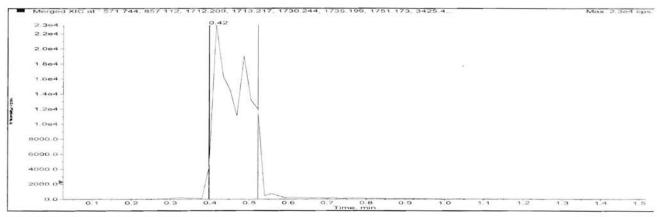
Fig. S50 HSQC spectrum of compound 14

Sample Name: TKN97-6 Sample Location: Vial 4 Sample Id: Operator: Milka

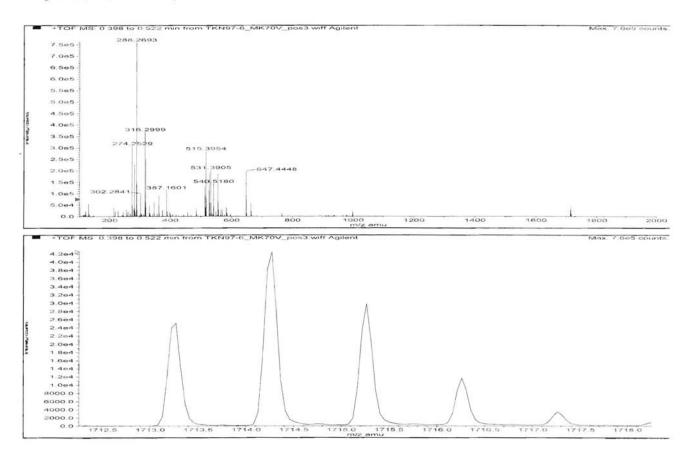
Data File Name: D:\PE Sciex Data\Projects\D_Milic\Data\TKN97-6_MK70V_pos3.wiff Acq Time: July 19 2012, 02:04:41 PM

Method: d:\TOF Software\damethods\Night_Seq_Comp_ident1.anm\efc.xml

One or more scans have failed IRM. Review the data file for details.



Merged XIC, Period#:1 Experiment#:1



Formula	Compound name	Mass	Peak RT (min)	Peak area	Description
C134H28N2O3		1712.20999	0.42	1.22436 E5	

Species	Abundance (counts)	Ion Mass	Measured Mass	Error (mDa)	Error (ppm)	Ret. Time Error (min)
[M+H]+	26096.15	1713.21727	1713.25252	35.24829	20.57	-

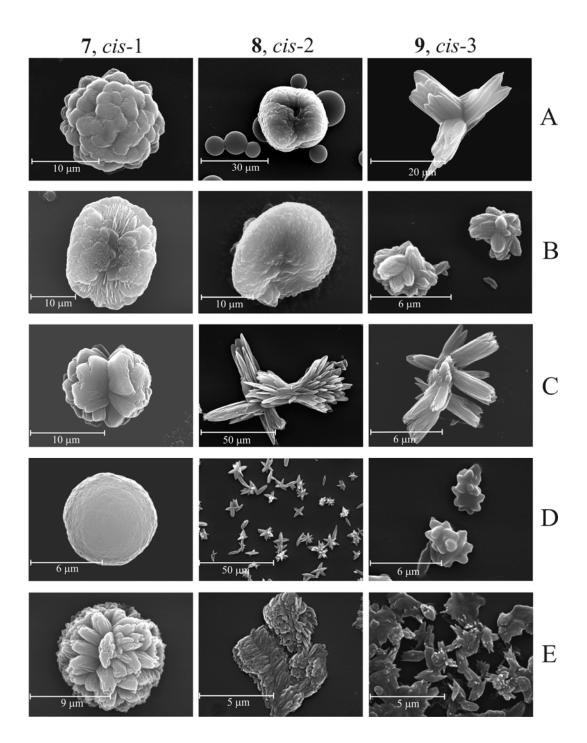


Fig. S52. Representative SEM images of samples prepared from 0.5 mM solution of three regioisomeric bisadducts with the dioxaoctane bridge in **A**) ODCB; **B**) PhMe; **C**) PhMe/iPrOH 1:1; **D**) PhMe/dioksan 1:1 and **E**) CHCl₃, on glass substrate at room temperature.

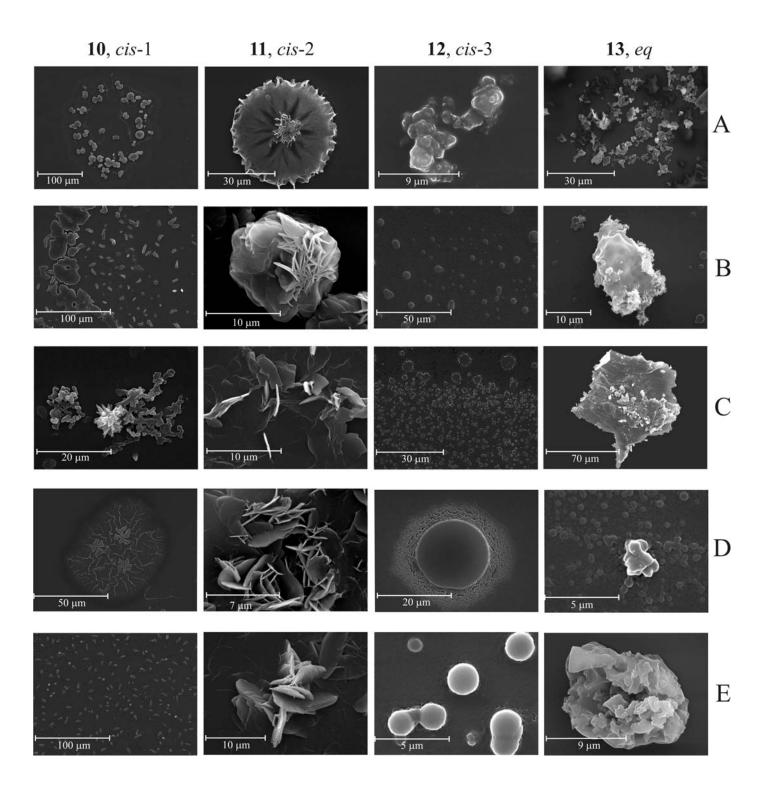


Fig. S53 Representative SEM images of samples prepared from 0.5 mM solutions of regioisomeric bisadducts with the trioxatridecane bridge in **A**) ODCB; **B**) PhMe; **C**) PhMe/*i*-PrOH 1:1; **D**) PhMe/dioxane 1:1, and **E**) CHCl₃ on glass substrate at room temperature.

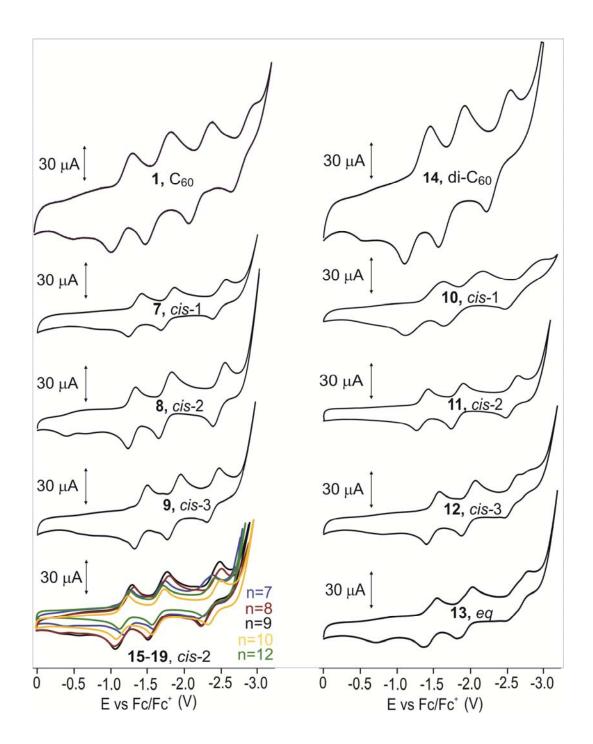


Fig. S54: CVs of compounds **7-19** in ODCB/DMF 2:1, with 0,1 M TBAP as a supporting electrolyte, recorded at the scanning rate of 0.7 V/s, at the room temperature, under the argon atmosphere.

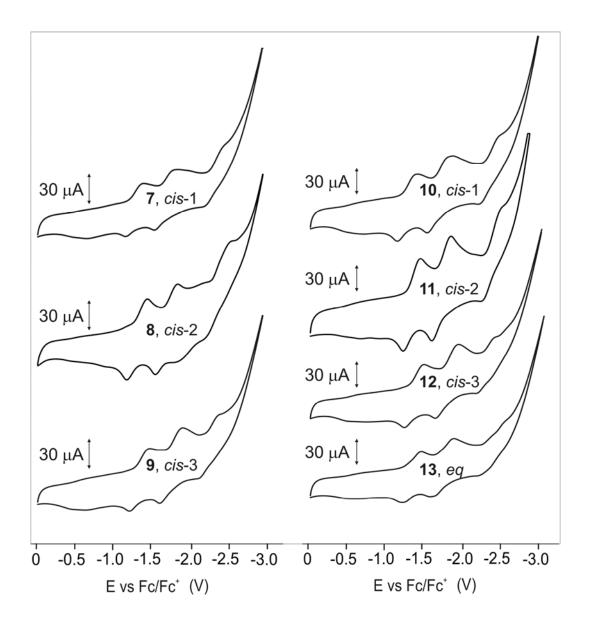


Fig. S55: CVs of compounds **7-13** in DCM, with 0,1 M TBAP as a supporting electrolyte, recorded at the scanning rate of 0.7 V/s, at the room temperature under the argon atmosphere.

500 [AU] 400 350 300 250 200 150 100 40 50 0 30 0.00 0.10 25 20 0.50 0.60 10 0.70 1.00 0 Track 1, ID: 450 94 C1 @ 340 nm 400-350 300-250-200 150 AutoGenerated2 @ 340 nm AutoGenerated3 @ 340 nm 100-50 0.64 0.84 1.04 0.24 0.44 Start Start Max Max Max End End Area Peak Rf Height Rf Height % Rf Height Area % Assigned substance 0.20 0.21 0.20 166.3 1.77 AutoGenerated3 1m 17.8 27.1 6.84 22.1 2m 0.44 32.1 0.45 35.4 8.95 0.46 31.4 270.3 2.88 AutoGenerated2 3m 0.53 4.5 0.64 333.2 84.20 0.90 1.3 8950.2 95.35 94 C1 Spectrum scan Thursday, October 15, 2015 1:13:28 PM 94 C1 on all Tracks Executed by Zivoslav Tesic 100.0 100.0 All detected peaks Mode Slit dimensions 6.00 x 0.30 mm, Micro [AU] [AU] Optimize optical system Resolution 80.0 80.0 Scanning speed 100 nm/s 70.0 70.0 Data resolution 10 nm/step 60.0 60.0 Reference spectrum, pos X 10.0 mm 50.0 50.0 Reference spectrum, pos Y 10.0 mm 40.0 40.0 30.0 30.0 20.0 20.0 10.0 10.0 0.0 200.0 250.0 300.0 350.0 400.0 450.0 500.0 550.0 600.0 [nm] 700.0

Rf

0.64 Rf 94 C1

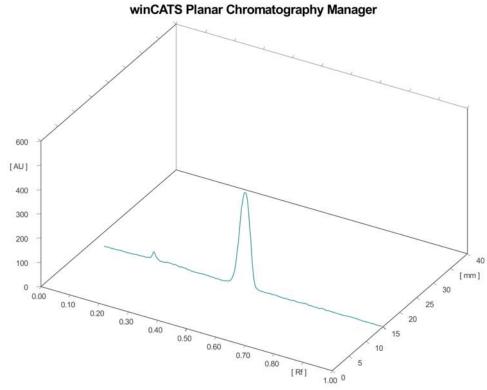
Substance

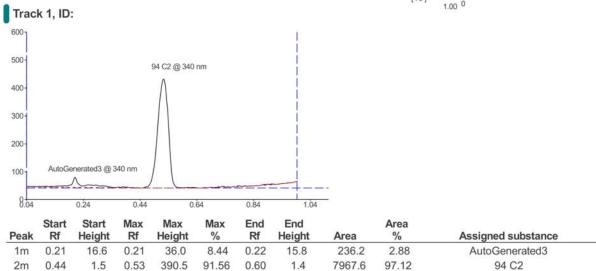
winCATS Planar Chromatography Manager

Fig. S56. HPTLC of bisadduct 7.

Max. @

261 nm



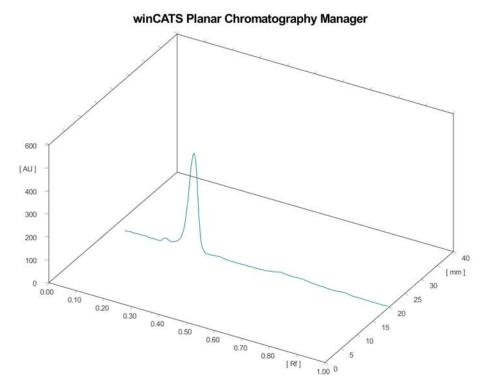


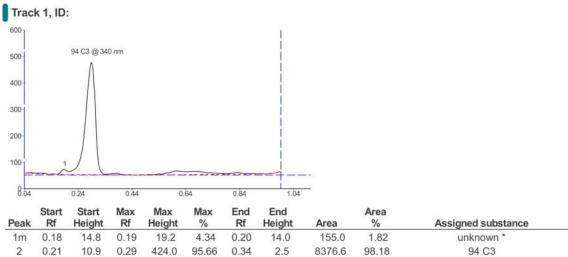
1.4

4 44	1 ·		
recuted by	Zivoslav Tesic	100.0	100
ode	All detected peaks		42530
it dimensions	6.00 x 0.30 mm, Micro	[AU] -	- [AU
ptimize optical system	Resolution	80.0	- 80.
canning speed	100 nm/s	70.0	- 70.
ata resolution	10 nm/step	60.0	- 60.
eference spectrum, pos X	10.0 mm		7455
eference spectrum, pos Y	10.0 mm	50.0	- 50.
1 5 2 11 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1		40.0 -	40.
		30.0 -	- 30.
		20.0 -	- 20.
		10.0 -	- 10.
		200.0 250.0 300.0 350.0 400.0 450.0 500.0 550.0 600.0 [nm] 700.0

Fig. S57. HPTLC of bisadduct 8.

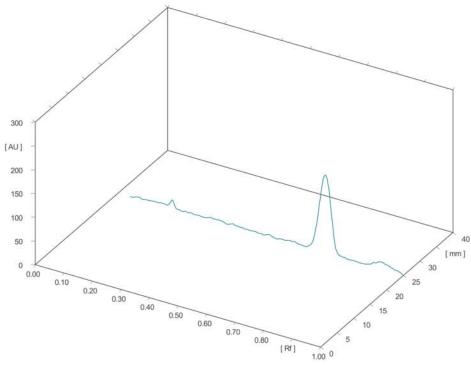
1.5

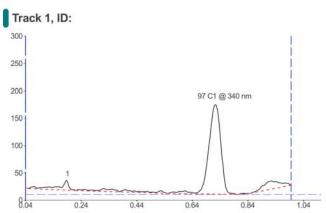




Executed by Mode Slit dimensions Optimize optical system Scanning speed Data resolution Reference spectrum, pos X Reference spectrum, pos Y	Zivoslav Tesic All detected peaks 6.00 x 0.30 mm, Micro Resolution 100 nm/s 10 nm/step 10.0 mm 10.0 mm	T Rf Substance 100.0 100.0 100.0 100.0 100.0 100.0 200.0 250.0 300.0 350.0 400.0 450 T Substance 1 0.29 Rf 94 C3	100.0 - [AU] - 80.0 - 70.0 - 60.0 - 50.0 - 40.0 - 30.0 - 20.0 - 10.0 - 10.0 - Max. @ 266 nm
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Fig. S58. HPTLC of bisadduct 9.





	Start	Start	IVIAX	IVIAX	IVIAX	Ena	Ena		Area		
Peak	Rf	Height	Rf	Height	%	Rf	Height	Area	%	Assigned substance	
1m	0.18	11.2	0.19	18.1	9.92	0.20	12.1	104.6	2.68	unknown *	
2m	0.64	2.6	0.73	164.1	90.08	0.82	0.6	3801.4	97.32	97 C1	

Spectrum scan	Friday, October 16, 2015 10:48:33 AM
Executed by	Zivoslav Tesic
Mode	All detected peaks
Slit dimensions	6.00 x 0.30 mm, Micro
Optimize optical system	Resolution
Scanning speed	100 nm/s
Data resolution	10 nm/step
Reference spectrum, pos X	10.0 mm
Reference spectrum, pos Y	10.0 mm

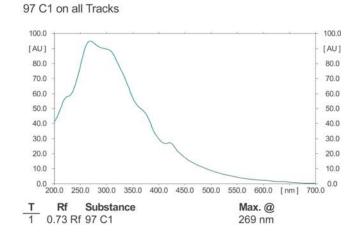
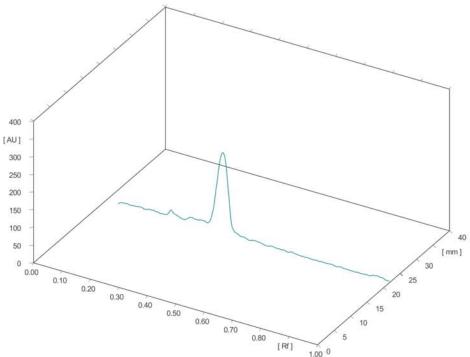
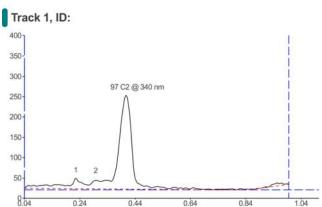


Fig. S59. HPTLC of bisadduct 10.





	Start	Start	Max	Max	Max	End	End		Area	
Peak	Rf	Height	Rf	Height	% R	Rf	Rf Height	Area	%	Assigned substance
1m	0.22	19.5	0.23	25.2	9.29	0.24	18.9	156.7	2.44	unknown *
2m	0.25	16.4	0.26	16.4	6.03	0.25	13.3	107.1	1.67	unknown *
3m	0.28	9.6	0.41	230.1	84 67	0.57	19	6158 4	95.89	97 C2

Spectrum scan	Friday, October 16, 2015 12:36:01 PM			
Executed by	Zivoslav Tesic			

Mode
Slit dimensions
Optimize optical system
Scanning speed
Data resolution
Reference spectrum, pos X
Reference spectrum, pos Y

Zivoslav Tesic
All detected peaks
6.00 x 0.30 mm, Micro
Resolution
100 nm/s
10 nm/step
10.0 mm
10.0 mm

97 C2 on all Tracks

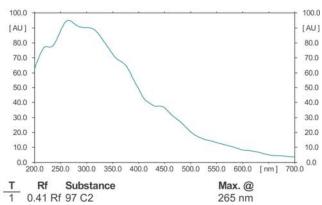
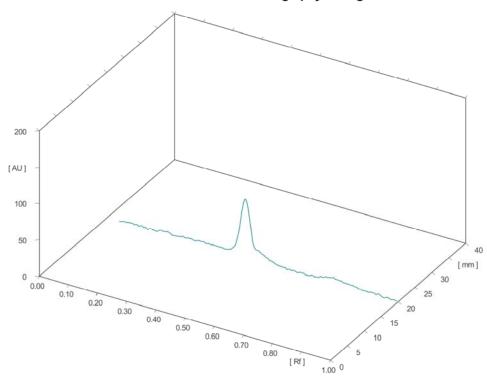
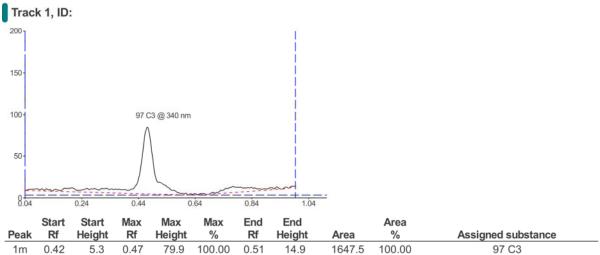


Fig. S60. HPTLC of bisadduct 11.



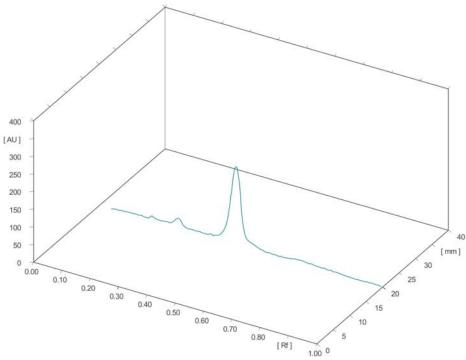


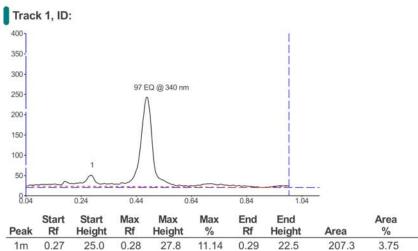
Spectrum scan	Thursday, October 15, 2015 2:15:26 PM	97 C3 on all Tracks	
Executed by Mode Slit dimensions Optimize optical system Scanning speed Data resolution Reference spectrum, pos seference spec		100.0 [AU] 80.0 70.0 60.0 40.0 30.0 20.0 10.0 200.0 250.0 300.0 350.0 400.0 450.0 T Rf Substance	100.0 [AU] 80.0 70.0 60.0 50.0 40.0 30.0 20.0 10.0 Max. @

0.47 Rf 97 C3

Fig. S61. HPTLC of bisadduct 12.

265 nm





88.86

0.59

3.3

5318.4

96.25

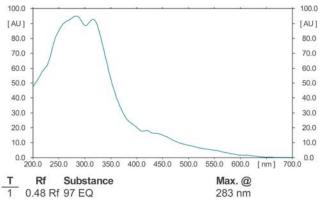
Spectrum scan	Thursday, October 15, 2015 1:53:33 PM
Executed by	Zivoslav Tesic
Mode	All detected peaks
Slit dimensions	6.00 x 0.30 mm, Micro
Optimize optical system	Resolution
Scanning speed	100 nm/s
Data resolution	10 nm/step
Reference spectrum, pos X	10.0 mm
Reference spectrum, pos Y	

221.7

0.48

6.1

97 EQ on all Tracks



Assigned substance

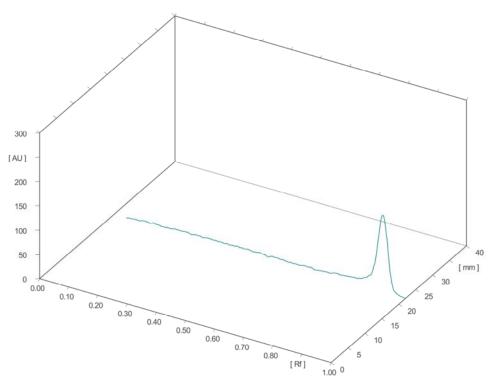
unknown *

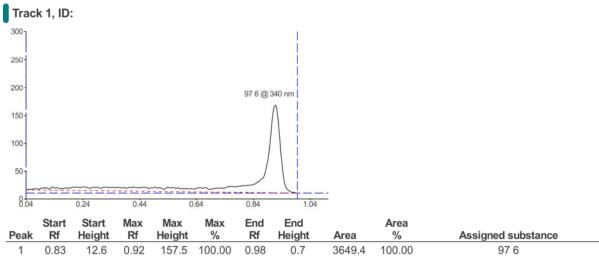
97 EQ

Fig. S62. HPTLC of bisadduct 13.

2m

0.38





Spectrum scan	Friday, October 16, 2015 1:16:29 PM	97 6 on all Tracks	
Executed by Mode Slit dimensions Optimize optical system Scanning speed Data resolution Reference spectrum, pos X Reference spectrum, pos Y	Zivoslav Tesic All detected peaks 6.00 x 0.30 mm, Micro Resolution 100 nm/s 10 nm/step 10.0 mm 10.0 mm	100.0 [AU] - 80.0 - 70.0 - 60.0 - 50.0 - 40.0 - 450 T Rf Substance	
T1 G(2 HDT) G (1)		1 0.92 Rf 97 6	Max. @ 271 nm

Fig. S63. HPTLC of bisadduct 14.