

Supplementary data for article:

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Electronic Supplementary Information

Biliverdin-copper complex at physiological pH

Milena Dimitrijević, Jelena Bogdanović Pristov, Milan Žižić, Dalibor Stanković, Danica Bajuk-Bogdanović, Marina Stanić, Snežana Spasić, Wilfred Hagen, Ivan Spasojević*

*E-mail: : redoxsci@gmail.com

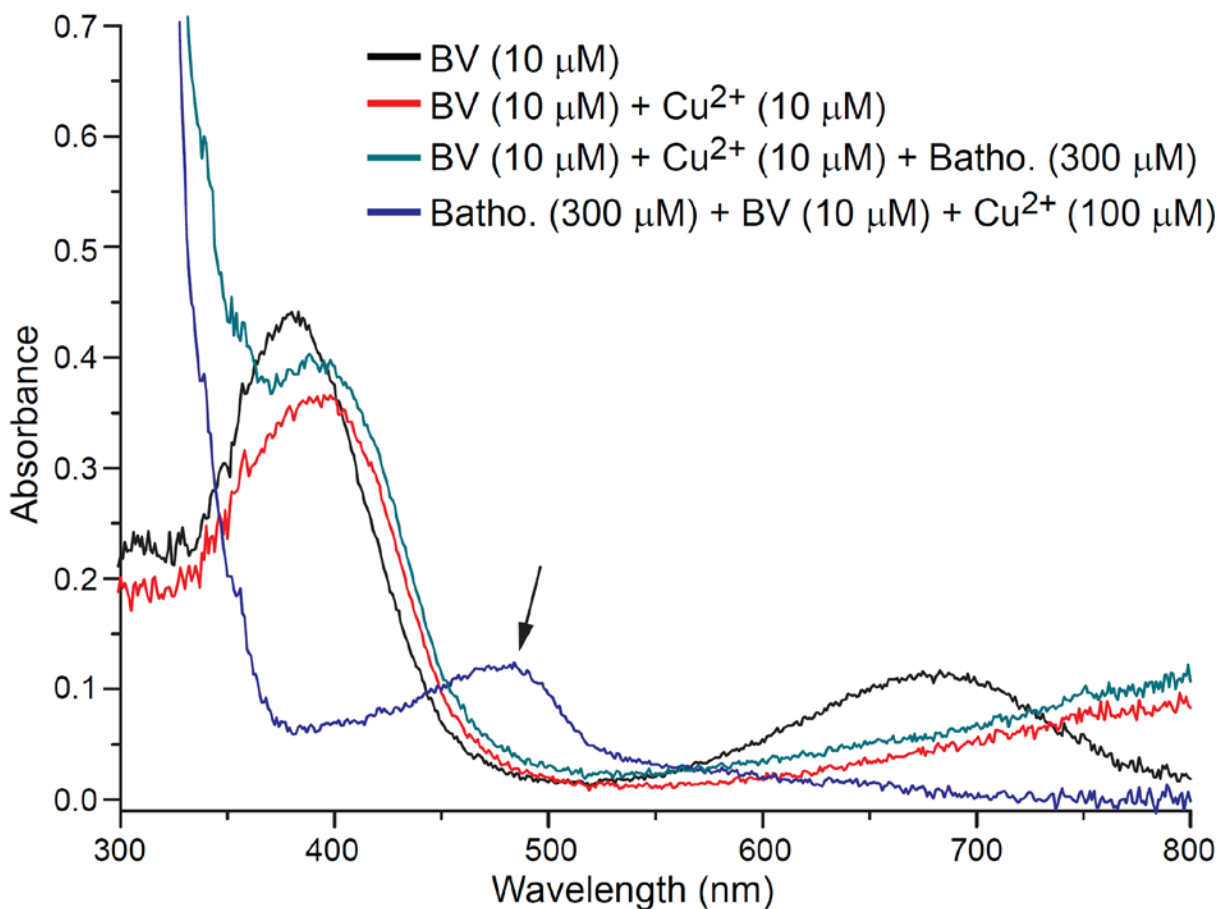


Figure S1. The stability of $[BV]/[Cu^{2+}] = 1$ system in the presence of copper chelating agent bathocuproine in phosphate buffer (50 mM; pH 7.4). Green line - BV and Cu^{2+} were incubated for 5 min before the addition of bathocuproine. Blue line - Cu^{2+} was added to the buffer after BV and bathocuproine (arrow – absorbance line of bathocuproine complex with copper). It is important to note that bathocuproine is a non-innocent copper chelator. In the presence of bathocuproine, the reduction potential for the Cu^{2+}/Cu^{1+} couple is raised by approximately 500 mV, making Cu^{2+} a powerful oxidant.¹ In the process, Cu^{2+} oxidizes BV-Cu complex, resulting in BV degradation (note that BV-related absorbance is completely lost (blue line)) and Cu release.

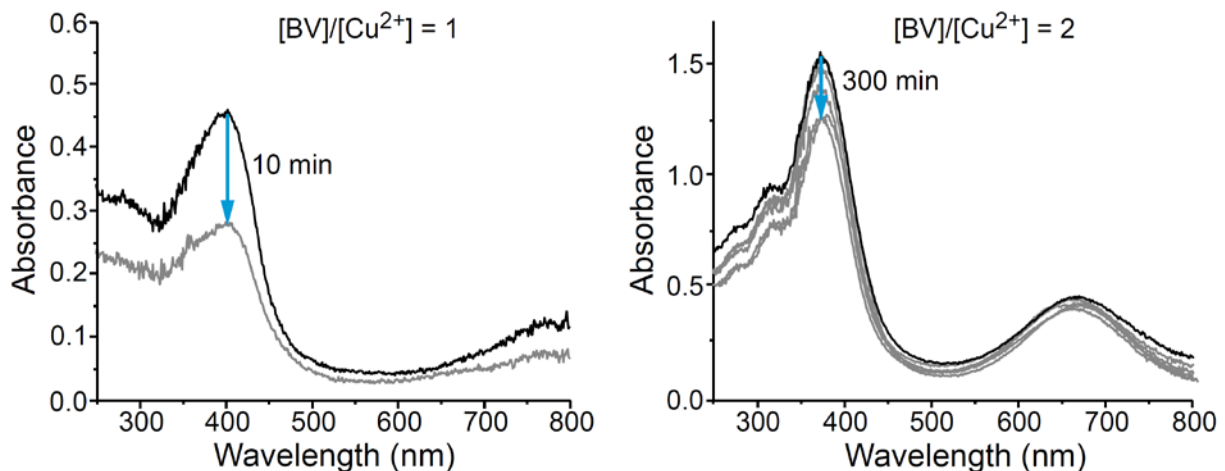


Figure S2. Changes in UV-Vis spectra of BV-Cu²⁺ systems prepared at high concentrations in 50 mM phosphate buffer, pH 7.4. Left: [BV]/[Cu²⁺] = 1; [BV] = [Cu²⁺] = 0.3 mM. Right: [BV]/[Cu²⁺] = 2; [BV] = 2mM; [Cu²⁺] = 1 mM. Aliquots were taken from each system and diluted to lower final concentrations (10 or 40 μ M, respectively), to allow spectra acquisition. It can be observed that the [BV]/[Cu²⁺] = 1 system underwent degradation within 10 min, whereas the [BV]/[Cu²⁺] = 2 system was relatively stable for 5 h.

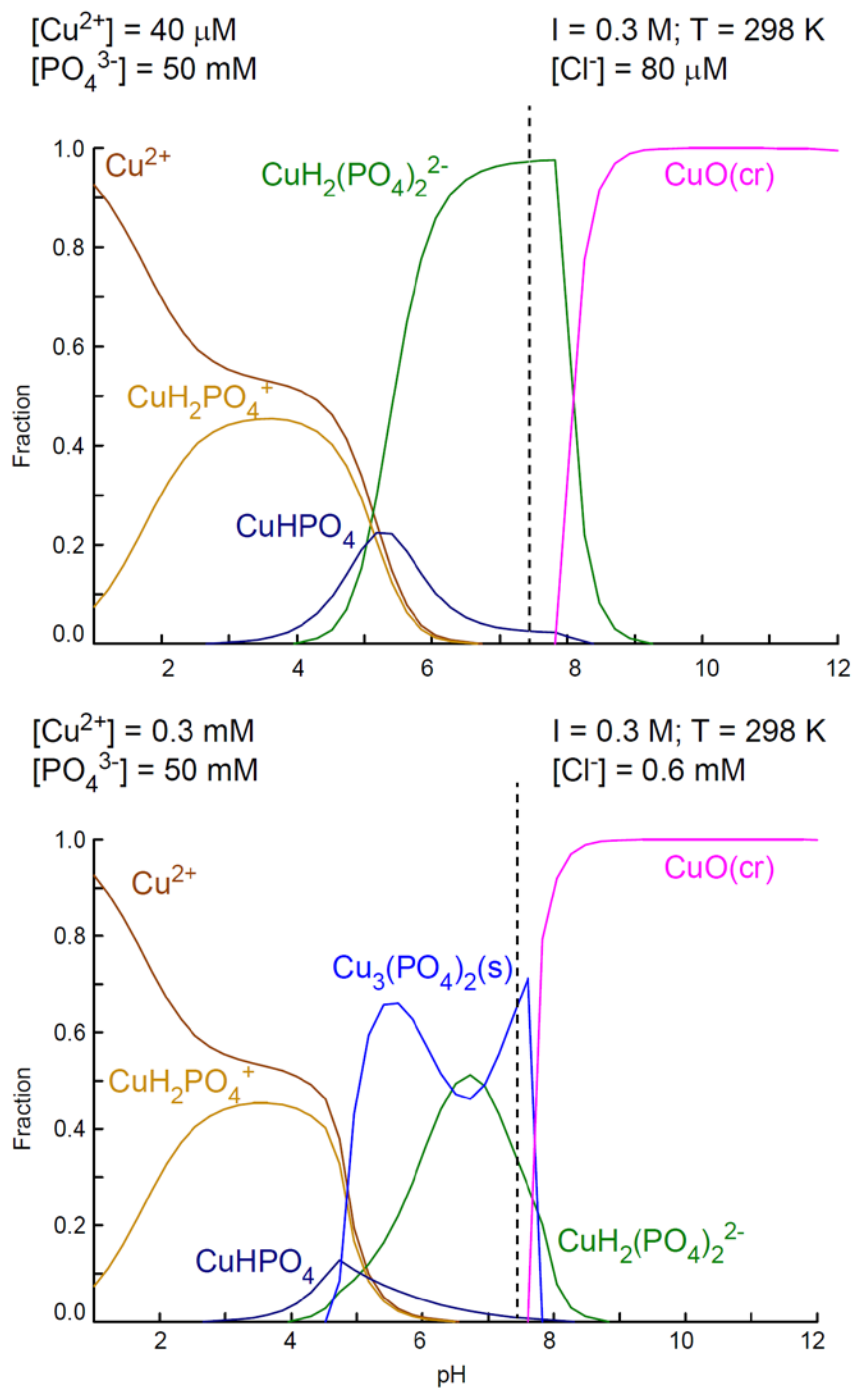


Figure S3. Speciation diagrams of Cu^{2+} in phosphate buffer (50 mM) at two concentrations – 40 μM (top) and 300 μM (bottom). Diagrams were prepared in Hydra-Medusa Software, using the presented parameters.

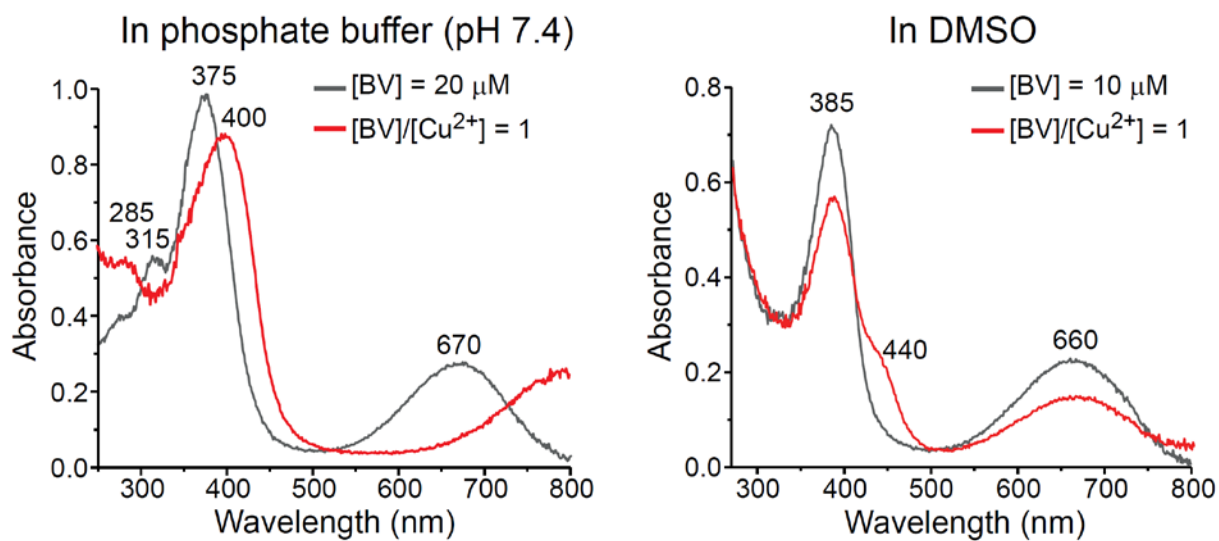


Figure S4. Comparison of UV-Vis spectra of biliverdin (BV) in the absence and the presence of Cu^{2+} in phosphate buffer (50 mM; pH 7.4) and in DMSO. Spectra were recorded after 5 min incubation period.

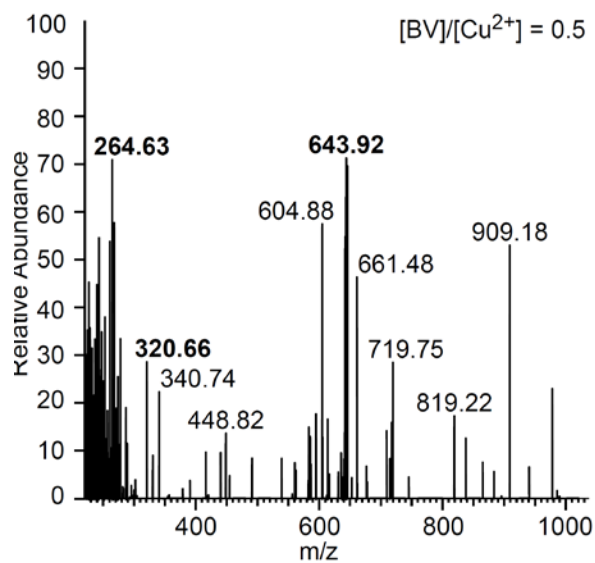


Figure S5. HESI-MS spectrum (full scan mode) of the system with [BV] = 20 μ M and [Cu] = 40 μ M ([BV]/[Cu] = 0.5). Assignment: m/z 643, BV-Cu complex; m/z 264, propentdyopent; m/z 320, propentdyopent complex with copper.²

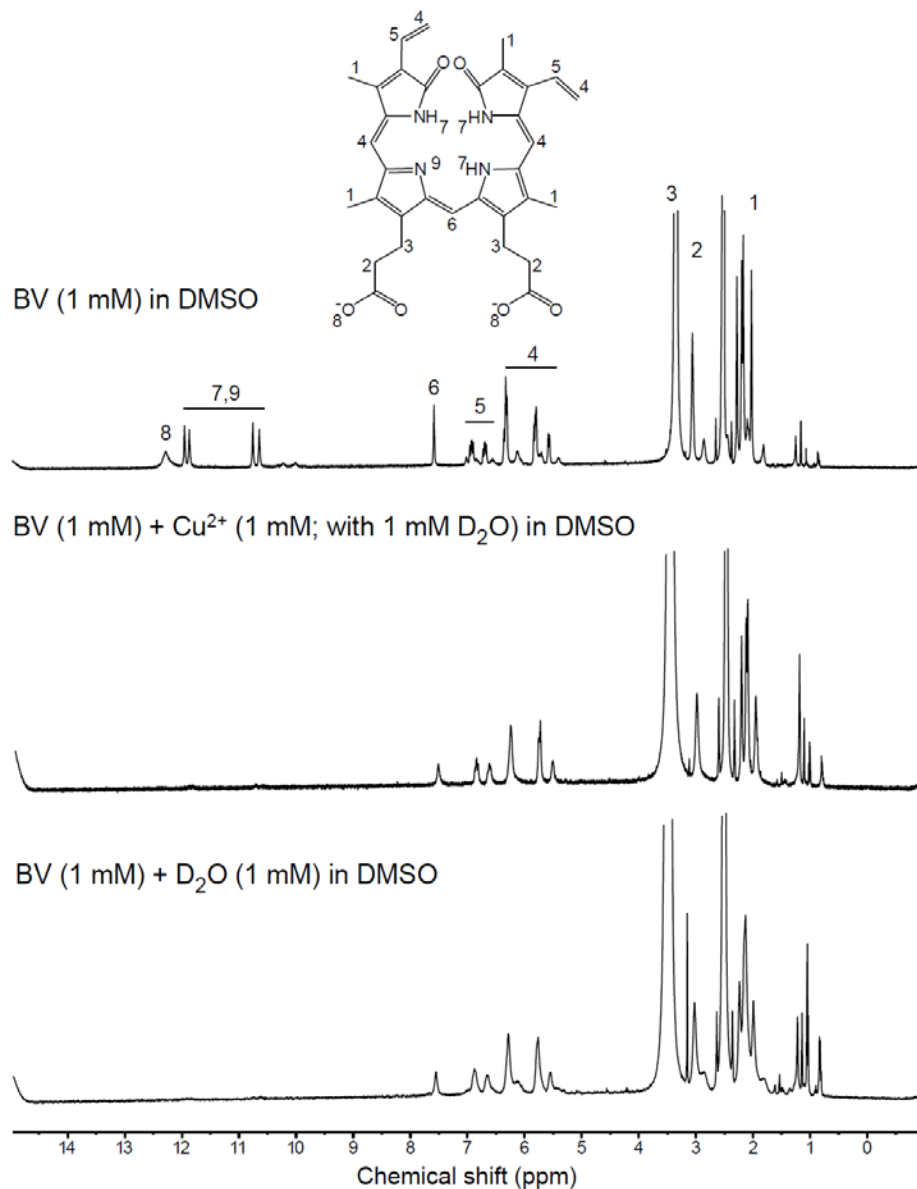


Figure S6. ¹H NMR spectra of biliverdin (0.3 mM) in DMSO-d₆ in the absence or the presence of Cu²⁺ at equimolar concentration. The bottom spectrum was recorded in a copper-free system, with the equimolar amount of D₂O as in experiments with copper. It can be observed that Cu²⁺ and D₂O induced similar (but not identical) changes. Changes in NH signals could not be observed in the presence of D₂O because of the chemical exchange. The peaks were assigned in accordance to previous reports.³ The signals labeled with 4 come from two types of protons (-CH= and =CH₂). The spectra were collected within 20 min after sample preparation.

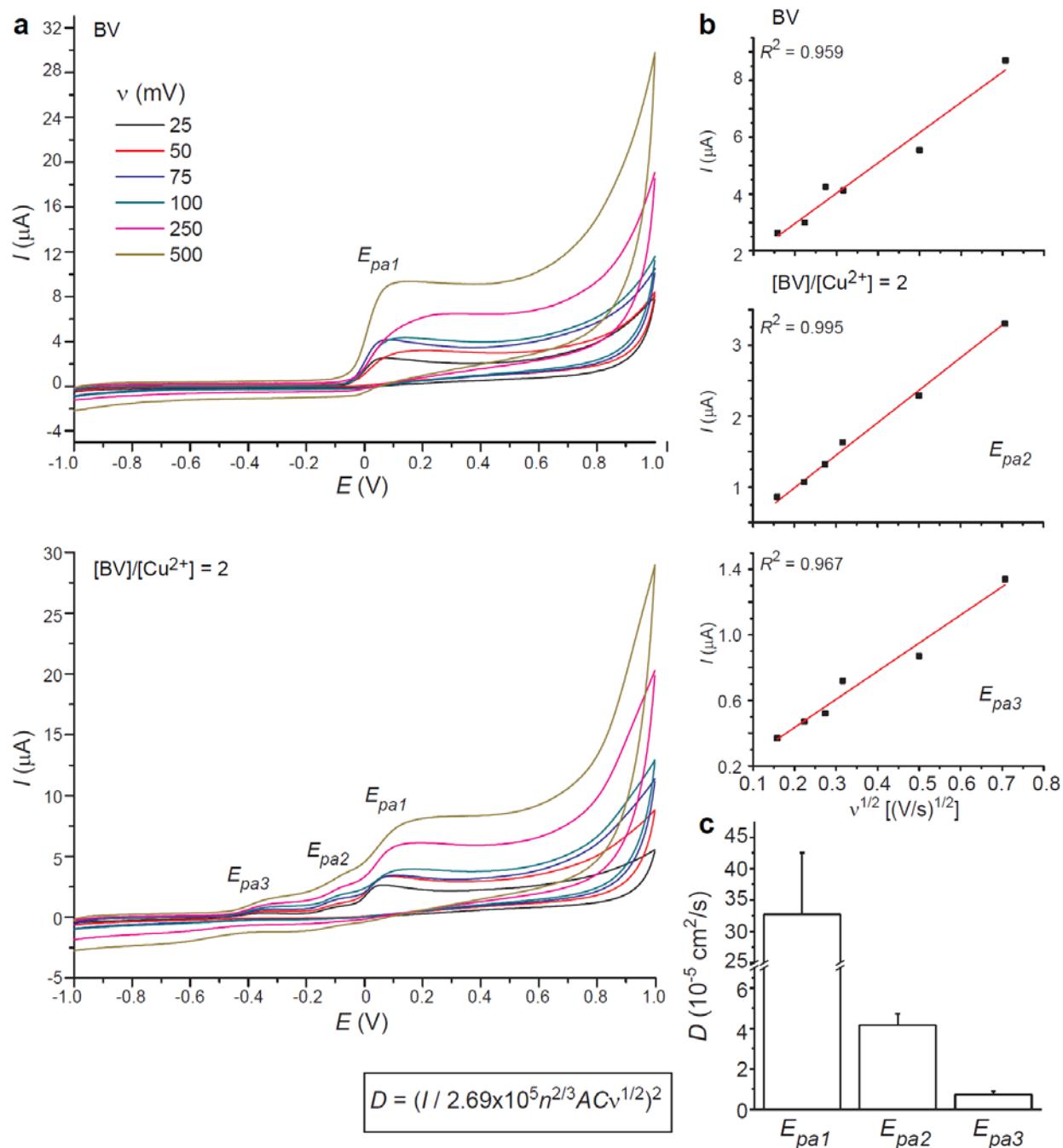


Figure S7. Scan rate analysis of BV and BV-Cu complex in phosphate buffer (50 mM; pH 7.4). (a) Cyclic voltammograms of BV (0.4 mM) in absence or presence of Cu^{2+} (0.2 mM) at the boron doped diamond electrode obtained at different scan rates ($v = 0.025\text{--}0.5$ V/s). (b) The dependence between anodic peak currents I at potentials E_{pa1} (oxidation of BV), and E_{pa2} and E_{pa3} (oxidation of BV-Cu complex(es)) and $v^{1/2}$. Linear fit and R^2 values are presented. (c) D for BV and BV-Cu complex(es). Randles–Sevick equation (in the box): n , number of transferred e^-

($1e^-$ for all peak currents), A , area of the working electrode (0.0707 cm^2); C , concentration of redox species in solution ($[\text{BV}] = 0.4 \text{ mM}$; $[\text{BV-Cu}] = 0.2 \text{ mM}$). Results are presented as means (\pm standard deviation) of measurements made at various v . All three D values were statistically different ($p < 0.001$; ANOVA with *post hoc* Duncan's test).

Table S1. Raman spectral lines that were observed for BV (1 mM), using the $\lambda = 532$ nm laser excitation line.

Line [cm⁻¹]	Assignment	References
1619	Lactam stretching	4
1470	C–C deformation, likely between rings	4,5
1443	Stretching CC, stretching CN	6
1393	CH ₃ asymmetric deformation	6
1362	CH ₃ deformation	6
1331	In plane bending CH(CH ₃)	6
1303	CH wagging	6
1254	Lactam ring	6
1179	C–H twisting	4
1101	Stretching C–C, stretching C–N	6
1003	Asymmetric CH ₃ deformation	6
971	C–C stretching mixed with C–H rocking	4
954	Stretching C–C–O	7
844	Stretching ring	6
767	In plane ring deformation	6
717	Out of plane ring deformation	6
684	Out of plane bending C=O	6

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