

AUTOMISED POTENTIOMETRIC END POINT TITRATION FOR CARBON AND ORGANIC MATTER DETERMINATIONS

K. Lambert; A.S. Rahardjono; Suryanto S.¹⁾ J. Vanderdeelen

Intisari

Pengukuran bahan organik tanah dengan metode "Dichromate Wet Combustion" meliputi proses titrasi senyawa dikromat yang tersisa dengan menggunakan ion ferro. Titrasi redoks potensiometrik diduga mampu berperan sebagai pengganti metode indikator warna diphenilamina. Modifikasi ini memungkinkan pengukuran akhir yang cepat dan teliti, dan dapat diandalkan bagi laboratoria yang analisis rutinnnya mencakup pengukuran bahan organik. Metode usulan ini diperbandingkan dengan metode konvensional Walkley dan Black, untuk tanah-tanah mineral yang berbeda dari Jawa Barat dan Jawa Tengah, untuk gambut untuk kompos dan limbah organik. Metode usulan terbukti sangat teliti dan dapat diulang serta dapat direkomendasikan bagi pengukuran bahan organik tanah di laboratoria pengujian tanah.

Abstract

The dichromate wet combustion determination of soil organic matter involves a titration of the excess of dichromate with ferrous ions. An automised potentiometric redox titration is proposed as a substitute for the diphenylamine colour indicator method. This modification enables a fast and accurate end point determination, and is very reliable for laboratories where organic matter determinations are part of the routine analysis. The proposed method is compared with the traditional Walkley and Black method for different mineral soils from West and Central Java, for peat, for compost and organic waste. The proposed method is highly accurate and reproducible and can be strongly recommended for soil organic matter determinations in soil testing laboratories.

1. Introduction

The term *soil organic matter* embraces the whole non-mineral fraction of soil and any vegetable or animal matter forming part of the sample analysed, will be included in the determination. Thus analytical results depend in the first place upon the size of the sieve used in the preparation of the sample (*Hesse, 1971*).

Soil organic matter has a structure which is too complex to be determined as such, but its composition is remarkably constant. Carbon is the main building-block of soil organic matter, comprising from 48% to 58% of the total weight (*Nelson and Summers, 1982*).

¹⁾Jurusan Ilmu Tanah, Fakultas Pertanian Universitas Gadjah Mada, Sekip Unit I, Yogyakarta, Indonesia.

Therefore, total organic matter is estimated on a routine basis from the determination of oxidizable carbon in the soil sample.

The principle of total carbon analysis is to convert the element completely into carbon dioxide which is then determined gravimetrically or volumetrically. Oxidation is achieved by either wet or dry combustion.

The quantitative determination of soil organic matter by the dichromate-wet combustion method (*Nelson and Summers, 1982*) is a commonly used procedure, especially for routine analysis in soil testing laboratories. The method is based on the oxidation of organic carbon by dichromate in acid medium and is well-known as the Walkley and Black procedure (*Walkley and Black, 1934*). The main disadvantage of the method is that the oxidation of organic carbon is incomplete and this necessitates the use of a correction factor, which means that the results obtained cannot be considered as quantitative. The main advantage of the procedure is its simplicity and rapidity.

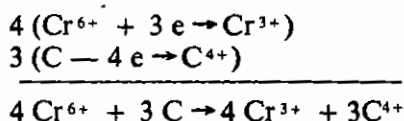
The excess dichromate is back-titrated with a standard ferrous reducing solution. The main difficulty in the titration is the detection of the end point. Several minor modifications of the method exist, using different end point colour indicators: diphenylamine in phosphoric acid medium (*Schollenberger, 1945*), barium diphenylamine sulphonate (*Peech et al., 1947*), ortho-phenantroline (*Jackson, 1958*), N-phenylanthranilic acid (*Simakov, 1957; Mebius, 1960*).

However, instead of relying entirely upon a colour change, the excess dichromate can be titrated potentiometrically (*Raveh and Avnimelech, 1973*), in which case no indicator is required.

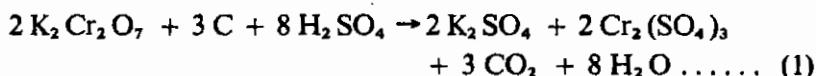
The objective of this paper is to describe a potentiometric method, using an automatic titration system. The proposed method is based on the use of redox potential measurements for the determination of the end point in the titration. A comparative study is done for the diphenylamine and the automatised potentiometric titration, using mineral soils from West and Central Java, peat, compost and organic waste materials. Saccharose ($C_{12}H_{22}O_{11}$), is used to check accuracy and precision.

II. Background and Theory

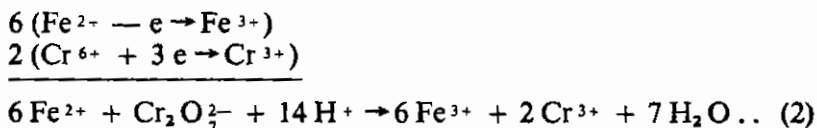
The oxidation of organic C by dichromate in acidic medium proceeds according to the following reactions:



In the Walkley & Black method the reaction is as such:



The soil sample is treated with an excess dichromate. After oxidation, the excess dichromate is back-titrated with a standard ferrous reducing solution, giving the reactions (acidic medium):



Reaction (2) expressed in terms of redox potential (of the half cell relative to the normal hydrogen electrode) consists of two Nernst's equations:

$$E(V) = E^\circ_{\text{Fe}^{2+}} - \frac{0.059}{1} \times \log \frac{[\text{Fe}^{2+}]}{[\text{Fe}^{3+}]} \dots \dots \dots (3)$$

$$E(V) = E^\circ_{\text{Cr}_2\text{O}_7} - \frac{0.059}{6} \times \log \frac{[\text{Cr}^{3+}]^2}{[\text{Cr}_2\text{O}_7^{2-}] \times [\text{H}^+]^{14}} \dots \dots (4)$$

where $E^\circ_{\text{Fe}^{2+}} = 0.77 \text{ V}$ and $E^\circ_{\text{Cr}_2\text{O}_7} = 1.33 \text{ V}$.

Table 1 describes the potentiometric titration in terms of redox-potential and concentration of the different ions involved in the reaction, under the experimental conditions of:

- 10 ml $\text{K}_2\text{Cr}_2\text{O}_7$ 1N = $1.67 \times 10^{-3} \text{ mol Cr}_2\text{O}_7^{2-}$ in 10 ml
- 20 ml H_2SO_4 36N = 0.72 mol H^+ in 20 ml
- a total volume at the start of (10 + 20 + 150 =) 180 ml.

There is a marked voltage shift (drop) at the end point of this reaction in the order of 400 mV for 0.02 ml of titrant. The change in potential during the potentiometric titration of $\text{Cr}_2\text{O}_7^{2-}$ by Fe^{2+} is given in figure 1 for 3 replicates of the blank titration (10 ml $\text{K}_2\text{Cr}_2\text{O}_7$ + 20 ml H_2SO_4 + 150 ml H_2O).

with depth in the profile. Accurate organic matter content estimates require a knowledge of the factor for the particular soil studied. However, if an estimate of organic matter content of surface soils must be made based on organic C data and no information on the exact factor is available, a factor of 2 appears to be most universally acceptable (Nelson and Summer, 1982).

Organic materials

The samples are treated in a slightly different way for peat, compost, sludge and other organic material because of the higher organic C content: 0.1 or 0.5 g material is weighed and 20 ml of 1N $K_2Cr_2O_7$ solution is added. The amount of $K_2Cr_2O_7$ will influence the calculations of C, with a factor 2 (20 meq $K_2Cr_2O_7$ oxidizes 20 meq C or 60 mg organic C).

Walkley and Black method

The organic C determinations with the procedure described above are compared with the results using the conventional Walkley and Black titrimetric determinations: 1 g of soil is weighed, 10 ml of 1N $K_2Cr_2O_7$ is added and immediately afterwards 20 ml concentrated H_2SO_4 (36N). Soil and solution are mixed gently and stored for 30 minutes. Then, 150 ml demineralised water is added and the suspension is filtered through a Whatman 42 filterpaper. Concentrated H_3PO_4 (10 ml) and 10 drops of diphenylamine indicator solution are added. The solution is titrated with a 1N solution of $Fe(NH_4)_2(SO_4)_2 \cdot 6H_2O$, using a microburette.

Initially the solution will be yellowish brown. Progressively the colour will change over blue to violet, which is a sign that the equivalence point is near. Upon further slow addition of $Fe(NH_4)_2(SO_4)_2$ -solution, the colour abruptly changes to green.

A blank titration, containing only dichromate, is carried out following the entire oxidation and titration procedure in order to standardise the $Fe(NH_4)_2(SO_4)_2$ -solution.

IV. Results and Discussion

Accuracy and Precision

The carbon content of a sugar (saccharose) was determined in order to check the accuracy of both methods. The Molecular Weight of saccharose $C_{12}H_{22}O_{11}$ equals 342.30 g, thus 40 mg saccharose con-

tains 16.827 mg C. Using the automised method the amount of C found was 16.820 mg, 16.788 mg, 16.784 mg respectively, or a recovery percentage (relative accuracy) of 99.96%, 99.77% and 99.74% (99.82% on average).

Using the diphenylamine method the amount of C found was 17.144 mg, 17.184 mg, 17.184 mg respectively, or a relative accuracy of 101.88%, 102.12% and 102.12% (102.12% on average).

It shows there is a slight overestimation when using the conventional Walkley and Black method. This may be due to the unexperience of the analyst, but it shows at least that the automatic titration is not influenced by this kind of problems and is thus more accurate and reliable.

During the carbon determinations of all the material listed in table 2a, "compost" (nr.21 in table 2a) was regularly repeated in the sequence of analysis in order to check more thoroughly the statistical meaning of the results obtained by both methods. The results given in table 4 show that the Standard Deviation is higher using the diphenylamine titrimetric determination. The Coefficient of Variation amounts 4.47% for the diphenylamine method and is 2.33% for the automatic redox titration. The precision of the latter is thus slightly better.

Organic Matter Determinations

The results of the organic matter (O.M.) determinations for different material listed in table 2a are shown in table 5. Comparing the results of both methods, one can conclude that:

(1) Nearly all values are slightly lower when using the proposed automised method. Based on the results of the recovery test, one can say there is an overestimation of the O.M. content when using the conventional Walkley & Black procedure. This overestimation is small and thus not of big concern, but it shows at least the higher reliability and accuracy of the automised method.

(2) The Standard Deviation for 3 replicates is usually higher when using the proposed automised method. Comparing this with the Standard Deviation for both methods in the recovery and accuracy tests with saccharose and compost, the lower Standard Deviation here (i.e. for 3 replicates in a row, instead of 10 replicates at random through the whole experiment) can be explained by the presence of the analyst. Again, this is not of big concern, but it favours the automised titration method once more: although giving higher Standard Deviations, the results are more reliable.

(3) The determined value of organic matter for peat is higher than 100% and this is of course not possible. It can be explained by the use of the factor 2 (common for mineral soils) in the calculations when converting % C, to % O.M.

Based on the ash content only, the O.M. content of the organic materials (see table 2a) can be estimated as given in table 6. This results in conversion factors C, /O.M. that differ from the 0.50 used. These factors are not of greater value than the others, any constant factor used remains an approximation at best.

More detailed experiments concerning the total carbon determinations in different organic soils using the proposed automatised method, in combination with the determinations of ash content and degree of decomposition, are necessary to find out what conversion factor(s) could be more suitable for organic soils. The lower the degree of decomposition the lower the recovery. It is recommended to use the Mebius procedure (*Mebius, 1962*) with external heating.

VI. Conclusion

The potentiometric redox titration is highly accurate and precise, and is not giving overestimates as in the case of the titrimetric determinations.

The subjectivity and the prescience of the analyst is excluded using an automatised method. The carbon determinations become thus more reliable.

Unexperienced analysts can confuse the dirty green colour at the beginning of a titration with the green colour of the end point, but this is merely due to lack of practice.

The lesser accuracy for the titrimetric determinations is compensated by the rapidity and simplicity of the method, but analysts confirm the much more pleasant determinations using the automatic titrator.

By using the automatised procedure, the carbon determinations become less expensive in terms of chemicals and glassware.

Resuming all this one can strongly recommend the automatised potentiometric end point titration for routine carbon determinations in soil testing laboratories.

Table 1. Description of the potentiometric redox titration of $\text{Cr}_2\text{O}_7^{2-}$ by Fe^{2+}

Titration Point	ml Fe^{2+} Added	Total Volume (ml)	$[\text{Cr}_2\text{O}_7^{2-}]$ (mol/l)	$[\text{Cr}^{3+}]$ (mol/l)	$[\text{H}^+]$ (mol/l)	E (V)
start	0	180	9.28×10^{-3}	0.00	4.00	—
1% neutralisation	0.1	180.1	9.18×10^{-3}	1.85×10^{-4}	4.00	1.466 ¹
10% neutralisation	1	181	8.30×10^{-3}	1.85×10^{-3}	3.98	1.445 ¹
90% neutralisation	9	189	8.84×10^{-4}	1.59×10^{-2}	3.81	1.415 ¹
equivalent point	10	190	0.00	1.76×10^{-2}	3.79	1.331 ²
0.2% over equiv.pt.	10.02	190.02	0.00	1.76×10^{-2}	3.79	0.929 ³
1% over equiv.pt.	10.1	190.1	0.00	1.76×10^{-2}	3.79	0.888 ³

(¹): equation (4) is used.

(²): equation (3) & (4) are used (at equivalent point, the concentrations are: $[\text{Fe}^{2+}] = 6 \times [\text{Cr}_2\text{O}_7^{2-}]$)

(³): equation (3) is used.

Table 2a. Type and origin of the material used for C-determinations

nr	Type of Material	Origin of Material
1	Vertic Rhodudalf	Mulo, Gunung Kidul, Central Java
2	Typic Rhodudalf	Jasinga, Bogor, West Java
3	Entic Pelludert	Gading, Gunung Kidul, Central Java
4	Entic Udipsamment	Kalitirto, Yogya, Central Java
5	Dystric Troposaprist	Pontianak, West Kalimantan
6	Eutric Troposaprist	Rawapening, Salatiga, Central Java
7	Rhodic Paleudalf	Ungaran, Semarang, Central Java
8	Typic Tropofluvent.	Samas Beach, Central Java
9	Udand	Boyolali, Central Java
10	Compost sugarcane factory	Sugarcane factory, Klaten, Central Java
11	Industrial Waste	Rungkut, Surabaya, East Java
12	Typic Rhodudalf	Pathuk, Gunung Kidul, Central Java
13	Typic Rhodudult	Jasinga, Bogor, West Java
14	Agricultural Waste	Fakultas Peternakan, UGM, Yogyakarta
15	Typic Udipsamment	Balecatuur, Sleman, Central Java
16	Lithic Udorthent	Gunung Cekel, Kulon Progo, Central Java
17	Entic Chromodert	Sedayu, Kulon Progo, Central Java
18	Lithic Udorthent	Sedayu, Kulon Progo, Central Java
19	Entic Pelludert	Sedayu, Kulon Progo, Central Java
20	Waste mushroom factory	Jerami Padi, Kulon Progo, Central Java
21	Compost	Mushroom factory, Dieng, Central Java

Table 2b. Some chemica; properties of the material used

nr.	Moisture Content (% w/w)	pH(H ₂ O)	pH(KCl)	R exch. Ca + K + Na (meq/100 g dry)
1	7.17	6.7	5.5	8.17
2	7.53	4.3	3.4	1.29
3	10.01	6.9	5.8	26.16
4	1.83	6.1	4.9	3.97
5	68.52	3.1	2.0	3.68
6	257.14	5.4	4.6	25.21
7	6.95	5.7	4.8	3.36
8	5.04	7.6	6.9	15.67
9	10.01	5.5	4.7	1.19
10	10.38	6.0	5.9	7.69
11	4.17	6.2	6.1	9.06
12	4.38	5.3	4.4	4.12
13	10.25	4.3	3.3	0.21
14	8.58	7.1	6.8	15.13
15	2.15	6.1	4.9	4.42
16	3.09	6.4	3.7	10.45
17	7.30	6.8	5.4	16.05
18	6.61	7.6	6.9	20.81
19	11.73	7.5	6.3	23.17
20	8.70	8.7	8.0	49.18
21	9.89	8.7	7.8	43.98

Table 3. Settings of the programme used for the titration of dichromate with ferrous ions (end point titration with Titrator Radiometer TTT85)

Parameter	Value
1 st end point	700 mV
dynamic range	1000 mV
2 nd end point	---
delay	5 s
pre dose	0.00 ml
sample volume	1.00 ml
titrant concentration	1.000
unit	M
blank volume	0.00 ml
maximum volume	20.00 ml

Table 4. Carbon determinations (C_v) for compost (10 replicates)

Titration With Diphenylamine	Automatic Redox Titration
21.12	20.35
23.60	20.98
21.48	20.96
21.48	20.91
21.43	19.48
21.07	20.45
23.15	20.47
21.43	20.68
21.12	21.03
23.15	20.99
(21.90 = 0.98)	(20.63 = 0.48)

Table 5. Organic matter determinations with the two different procedures

nr.	Titrimetric				Potentiometric			
	% O.M.		(Mean ± s)		% O.M.		(Mean ± s)	
1	4.06	4.06	4.14	4.09 ± 0.05	3.54	3.49	3.58	3.54 ± 0.05
2	5.09	4.93	5.09	5.04 ± 0.09	4.67	4.76	4.96	4.80 ± 0.15
3	3.42	3.34	3.34	3.37 ± 0.05	2.85	3.07	3.08	3.00 ± 0.13
4	1.99	1.99	1.92	1.97 ± 0.04	1.43	1.50	1.43	1.45 ± 0.04
5	125.14	129.22	127.86	127.41 ± 2.08	116.08	120.02	116.96	117.69 ± 2.07
6	99.58	96.56	99.58	98.57 ± 1.74	81.26	83.82	80.32	81.80 ± 1.81
7	2.83	2.83	2.83	2.83 ± 0.00	1.84	2.15	2.14	2.04 ± 0.18
8	2.66	2.66	2.83	2.72 ± 0.10	2.20	2.20	2.13	2.18 ± 0.04
9	22.84	22.84	22.54	22.74 ± 0.17	18.22	17.78	18.54	18.18 ± 0.38
10	105.46	106.36	106.36	106.06 ± 0.52	92.44	97.18	94.76	94.79 ± 2.05
11	25.55	25.96	26.37	25.96 ± 0.41	23.77	25.94	24.95	24.89 ± 1.09
12	3.18	3.09	3.26	3.18 ± 0.09	2.25	2.21	2.52	2.33 ± 0.17
13	3.32	3.23	3.41	3.32 ± 0.09	2.57	2.53	2.68	2.59 ± 0.08
14	66.76	65.02	67.64	66.47 ± 1.33	73.52	69.42	71.24	71.39 ± 2.05
15	2.99	2.99	2.99	2.99 ± 0.00	1.63	1.67	1.58	1.63 ± 0.05
16	1.68	1.68	1.51	1.62 ± 0.10	1.06	0.86	0.99	0.97 ± 0.10
17	3.66	3.66	3.75	3.69 ± 0.05	3.16	3.15	3.13	3.15 ± 0.02
18	4.57	4.66	4.48	4.57 ± 0.09	4.64	4.45	4.25	4.45 ± 0.20
19	2.29	2.29	2.11	2.23 ± 0.10	2.63	2.63	2.70	2.65 ± 0.04
20	56.60	57.48	59.26	57.78 ± 1.36	56.88	54.78	54.86	55.51 ± 1.19
21	42.14	42.86	42.86	42.62 ± 0.25	40.89	40.93	41.35	41.06 ± 0.42

Table 6. Conversion factors for organic material

nr.	Ash Content (% w/w)	% O.M. (1)	% C _i (2)	C _i /O.M.
5	0.11	99.89	58.85	0.59
6	9.89	90.11	40.90	0.45
10	24.56	75.44	47.40	0.63
14	43.00	57.00	35.70	0.63
20	47.09	52.91	27.76	0.52
21	54.34	45.66	20.53	0.45

(1) % O.M. = (100 - ash content) %

(2) % C_i = (0.5 × O.M. [table 5, potentiometric, mean]) %.

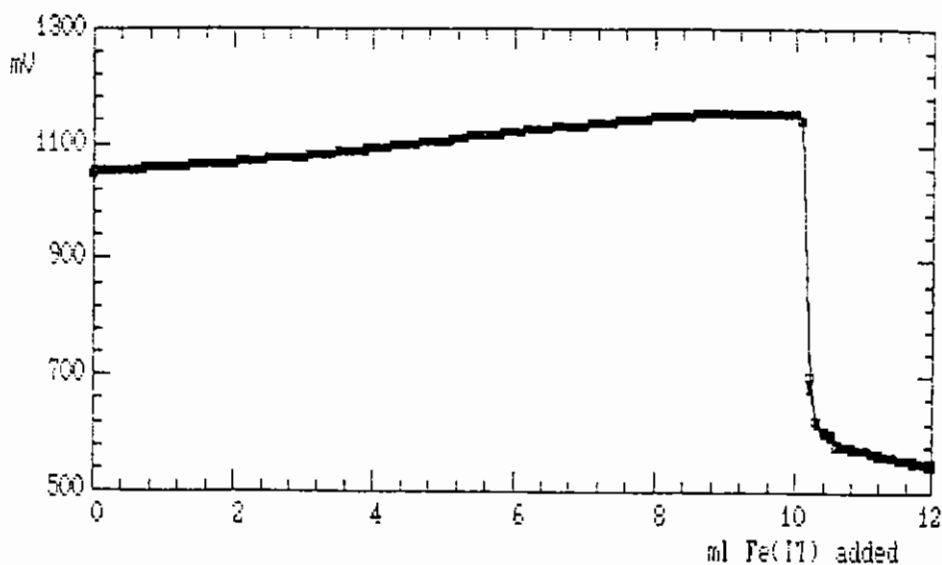


Figure 1. Change in potential during the potentiometric titration of the excess of dichromate with ferrous ions

References

- Christian, G. 1986. *Analytical Chemistry*. 4th edition. Wiley, New York. p. 273.
- Dynoodt, R. 1991. *Organic Carbon*. Practical work notes & personal communication.
- Hesse, P.R. 1971. Carbon and Organic Matter. In: *A Textbook of Soil Chemical Analysis*. Chemical Publishing, New York, 204—254.
- Jackson, M.L. 1958. *Soil Chemical Analysis*. Prentice Hall Englewood Cliffs. New York.
- Mebius, L.J. 1960. A rapid method for the determination of organic carbon in soil. *Analytical Chimia Acta*, 22: 120—124.
- Nelson, D.W. and Sommers, L.E. 1982. Total Carbon, Organic Carbon and Organic Matter. In: Page A.L. (ed.) *Methods of Soil Analysis*, Part 2: Chemical and Microbiological Properties. 2nd edition. Madison, Wisconsin, *Agronomy* 9: 539—579.
- Peech, M., Dean, L.A. and Reed, J. 1947. Methods of soil analysis for soil fertility investigation. U.S. Department of Agriculture Circular 757. (Cited by Hesse, 1971.)
- Ravech, A. and Avnimelech, Y. 1973. Potentiometric determination of soil organic matter. *Soil Science Society of America Proceedings*, 36: 967.
- Schollenberger, C.J. 1945. Determination of soil organic matter. *Soil Science*: 59, 53—56.
- Simankov, V.N. 1957. The use of phenylanthranilic acid in the determination of humus by Tyurin's method. *Pachvovedenie*, 8, 72—73. (Cited by Nelson and Summers (1982)).
- Stumm, W. and Morgan, J. 1981. *Aquatic Chemistry*. 2nd edition. Wiley-Interscience, New York, p. 478.
- Walkley, A. and Black, I.A. (1934). An examination of the Degtjareff method for determining soil organic matter and a proposed modification of the chromic acid titration method. *Soil Science*, 37: 29—38.