Exposing the human nude phenotype

The recent discovery of the human coun-terpart of the hairlessmousephenotype1has helped our understanding of the molecular genetics of hair growth.But there are no reports of a defect in thehuman homologue of the best known of the 'bald' mouse phenotypes, the nudemouse2. This may be because affected individualsare so gravely ill from the accompanyingimmunodeficiency that their baldness goesunnoticed. We have carried out a geneticanalysis that reveals a human homologue of the nudemouse. The nudemouse is characterized by acongenital absence of hair and a severeimmunodeficiency2, resulting from muta-tions in the whn(winged-helix–nude;Hfh11nu) gene, which encodes a member of the forkhead/winged-helix transcription factor family with restricted expression inthymus and skin3. The simultaneous occur-rence of severe functional T-cell immunodeficiency, congenital alopecia and nail dys-trophy (MIM database no. 601705) in twoaffected sisters led to the recognition that he clinical phenotype was reminiscent of the nudemouse4. We therefore investigated whether this syndrome represents the human counterpart of the nudemousephenotype. We obtained DNA samples from mem-bers of the sisters' family in a small villagein southern Italy. The affected sisters wereborn with a complete absence of scalp hair(Fig. 1a), eyebrows and eyelashes and haddystrophic nails, and no thymic shadow wasevident upon X-ray examination. The firstaffected child revealed a striking impair-ment of T-cell function shortly after birth, and died at the age of 12 months. Her sisterhad similar immunological abnormalities, but bonemarrow transplantation at fivemonths of age led to full immunological reconstitution, although the alopecia andnail dystrophy are still present4. We performed linkage analysis usingmicrosatellite markers near the humanWHNlocus on chromosome 17, and founda lod score of 1.32, suggestive of linkage. Wethen sequenced the human WHNgene5andfound a homozygous C-to-T transition atnucleotide position 792 of the WHNcDNA(GenBank accession no. Y11739) (Fig. 1b). This leads to a nonsense mutation atresidue 255 (R255X) in exon 5, and predicts the complete absence of functional proteinas a result of nonsense-mediated decay of messenger RNA. Because the proband's bonemarrowtransplant was from her brother, we exam-ined her leukocyte DNA both before and after the graft for the presence of chi-maerism. Genotyping the proband before the transplant showed that her leukocyteDNA was homozygous only for the mutantallele (Fig. 1c). Four years after the transplant, we detected the haplotype specific for the wild-type paternal WHNallele received from the brother, as well as the mutantallele, indicative of chimaerism. Genderdetermination revealed that the proband'sleukocyte DNA was genotypically XXbefore the transplant, and the brother'sDNA was XY. Afterwards, the proband'sleukocyte DNA was found to be XY (Fig.1c), providing evidence of longtermengraftment and expansion of the bone-marrow graft. The WHNgene encodes a transcriptionfactor, which is developmentally regulated and directs cell-fate decisions6.In mam-mals, whnis expressed specifically in theepithelial cells of the skin and thymus, where it helps to maintain the balancebetween growth and differentiation7,8.Recent evidence9has highlighted theimportance of the thymic microenviron ment in determining the T-cell repertoire, as both positive and negative selection of developing T cells depends on cell-cellinteractions with the thymic epithelium. Inwhnknockout mice, the defect has beenlocalized to the differentiating thymicmicroenvironment rather than to a defectin the developing T cells7. The proband wasfree of infections for four years after thebone-marrow transplant, indicating that T-cell function was at least partly restored. This is probably due to mature T cells ofdonor origin, although we cannot exclude the possibility that

positive selection of Tlymphocytes occurs in the peripherydespite the mutated whngene.Our findings provide evidence of ahuman immunodeficiency caused by a geneexpressed not in haematopoietic cells10, butin specific epithelial cells. In the human hairfollicle, expression of WHNis sharplydemarcated in defined cell populations (Fig.1d). Although nudemice appear to be com-pletely naked, the dermis contains a normalnumber of hair follicles, but they areincompletely developed. The fact that onlyshort, bent hairs occasionally emerge from the epidermis is thought to result from impaired keratinization11. Together with the hairlessgene1, our finding extends the evi-dence implicating cell-type-specific tran-scription factors in hair-follicle cycling andmorphogenesis, and indicates that baldnessis an extremely complex phenotype.



Figure 1. Molecular analysis of the human nudephenotype. a,A five-year-old child with congenital alopeciaand T-cell immunodeficiency. b,Sequence analysis of a nonsense mutation in exon 5 of the WHNgene. Top,homozygous wild-type sequence from an unrelated, unaffected control individual. Middle, sequence from aheterozygous carrier of the mutation R255X; arrow indicates a double T&C peak. Bottom, homozygousmutant R255X sequence from the affected individual; arrow indicates mutant T only, leading to a C-to-T transi-tion (CGA to TGA) and a substitution of an arginine residue

by a nonsense mutation. c,Restriction-enzymedigestion confirms the mutation. The mutation introduced a restriction site for BsrI and, after digestion of the184-base-pair (bp) polymerase chain reaction (PCR) product containing exon 5, the product generated from the mutant allele should cleave into two bands of 120 and 64 bp. Top, the unaffected parents and brother hadthree bands of 184, 120 and 64 bp (lanes 1, 2 and 6), indicating that they were heterozygous carriers of themutation R255X. Both patients had only the two digested bands of 120 and 64 bp (lanes 3 and 4), consistent with the presence of the mutation in the homozygous state. Bottom, evidence for longterm engraftment of the bone-marrow transplant. Gender determination of family members revealed a genotypically XX pattern of an undigested 300-bp band in the mother (lane 1) and affected patients (lanes 3 and 4), and a genotypicallyXY pattern consisting of the 300-bp band and two additional bands of 216 and 84 bp, indicative of the Y chro-mosome, in the brother (lane 2) and father (lane 6). Lane 5, peripheral blood leukocytes from the patient afterthe transplant, demonstrating an XY genotype and the presence of the normal WHNallele, providing evi-dence for fraternal chimaerism and persistence of the graft. M, size markers. d,WHNmRNA expression innormal human scalp skin. In the hair bulb, WHNmRNA is localized to the differentiating cells of the hair follicleprecortex (pc) and the innermost cell layer of the outer root sheath (arrowheads); the dermal papilla (dp)fibroblasts and hair matrix below the level of Auber (small arrows) remain negative for WHNmRNA.

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References: 1.Ahmad, W.et al.Science279,720–724 (1998).2.Flanagan, S. P. Genet. Res.8,295–309 (1966).3.Nehls, M., Pfeifer, D., Schorpp, M., Hedrich, H. & Boehm, T.Nature372,103–107 (1994).4.Pignata, C. et al.Am. J. Med. Genet.65,167–170 (1996).5.Schorpp, M., Hoffmann, M., Dear, T. N. & Boehm, T.Immunogenetics46,509–515 (1997).6.Kaufmann, E. & Knöchel, W. Mech. Dev.57,3–20 (1996).7.Nehls, M. et al.Science272,886–889 (1996).8.Brissette, J. L., Li, J., Kamimura, J., Lee, D. & Dotto, G. P. GenesDev.10,2212–2221 (1996).9.Muller-Hermelink, H. K., Wilisch, A., Schultz, A. & Marx, A.Arch. Histol. Cytol.60,9–28 (1997).10.Fischer, A.et al.Annu. Rev. Immunol.15,93–124 (1997).11.Köpf-Maier, P. et al.Acta Anat.139, 178–190 (1990).