Metal Ion Catalysed Adenosine-5'-triphosphate Hydrolysis: Part I-Cu(II) Catalysed Hydrolysis

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Received 26 March 1976; accepted 10 May 1976

The kinetics of the cupric ion catalysed hydrolysis of adenosine-5'-triphosphate (ATP) in aqueous solutions has been studied at 46.0° , 56.0° and 64.0° , over the broad pH-range 3.0-8.0, and $\mu = 0.1$ (KNO₃). pH-rate profiles have been analysed and the overall rate constants have been resolved into specific rate constants relating to the various Cu: ATP chelate species in solution. Activation parameters ΔH^{\ddagger} , ΔS^{\ddagger} and ΔF^{\ddagger} , for the specific rate constants of the various chelate species are reported. The possible mechanisms for the hydrolysis of the various ATP ionic species in the absence and the presence of Cu(II) ions are discussed.

'N a previous publication¹ the kinetics of the nonenzymic hydrolysis of adenosine-5'-triphoshas been reported. However, the presence of bivalent metal ions are found to be essential in the in vivo enzymic hydrolysis reactions of ATP^{2,3}. An understanding of the mechanism of metal ion catalysis in simple nonenzymic systems will be of much help in interpreting the function and role of metal ions in enzymic ATP reactions. It was therefore considered important to carry out a detailed physico-chemical study of the effect of a few bivalent metal ions on the nonenzymic hydrolysis of ATP in aqueous solutions, as a function of pH and temperature. Earlier investigators in this field4-11 have investigated the effect of Mg(II), Ca(II) and Ba(II) ions on the rate of ATP and ADP hydrolysis. The study was extended to metal ions less basic than the alkaline earths by Liebecq12, Lowenstien13 and Westheimer¹⁴. However, these investigations were only qualitative or semiquantitative in nature and various buffers known to complex with metal ions¹⁵ were used to maintain a constant ρ H. Schneider and Brintzinger¹⁶ have also reported the effect of Cu(II) ions on the ATP hydrolysis. In this paper are reported the results of a detailed, physico-chemical study of the effect of Cu(II) ions on the nonenzymic hydrolysis of ATP at 46°, 56° and 64° over the broad pH range of 3-8 and $\mu = 0.1$ (KNO₂).

Materials and Methods

A chromatographically pure crystalline adenosine-5'-triphosphate dipotassium salt (from Mann Research Laboratories, USA) was used. The aqueous solution $Cu(NO_3)_2$ (AR) was standardized using EDTA (disodium salt) by the procedure outlined by Schwarzenbach¹⁷.

The experimental procedure used was essentially the same as that used for the uncatalysed study¹ To maintain a constant temperature of 56°, 64° and 78°, acetone, methanol and ethanol respectively were used along with the experimental set-up described¹. For work at 46°, a double-walled glass cell serviced by a constant temperature water-bath at 46° \pm 0·1° was used. The pH of the solution was initially adjusted to the required pH, by the addition of enough acid or alkali. Any change in pH during the course of the kinetic runs was also adjusted by adding acid or alkali. Buffers were not used to maintain a constant pH, since the basic component of most buffers are known to complex with Cu(II) ions¹⁵.

Kinetic runs were carried out with equimolar amounts of Cu(II) and ATP ($\sim 2.0 \times 10^{-3}M$), over the pH range 3-8 at 46°, 56° and 64°. First order rate constants were evaluated in the initial stages of the hydrolysis reaction from plots of log a (a-x) against t (where a = initial concentration of ATP and x = amount reacted at time t). By restricting the kinetic study to the initial stages of the reaction, complications resulting from further hydrolysis of products was eliminated. All calculations were carried out with an IBM 370/155 computer.

Results

In the previous publication¹, kinetic data for the uncatalysed ATP hydrolysis was reported at 56° and 64°. This work has now been extended by studying the kinetics of the uncatalysed ATP hydrolysis at 78° also. Separate values for the specific rate constants of the monoprotonated species, HATP³⁻ were reported¹, one for the pH region 3-7 and the other for the pH region 7-9. As the pH region 6.0-7.5 is very sensitive to slight changes in pH of the solution, the kinetic runs in this pH region were repeated, and the first order rate constants obtained for the uncatalysed ATP hydrolyses are given in Table 1. Using this data recalculation of the specific rate constants in the

TABLE 1 - EXPERIMENTAL AND CALCULATED	RATE
CONSTANTS FOR THE UNCATALYSED ATP HYDR	OLYSIS
$[u = 0.1 (KNO_{*})]$	

¢Η		Rate c	onstant >	< 10 ⁴ (mi	in-1) at	
	5	6°	6	4°	7	8°
	Expl	Calc.	Expl	Calc.	Expl	Calc.
3.0	1.89	1.84	5.69	5.54	33.37	34.86
3.5	1.80	1.78	5.26	5.22	30.07	31.51
4.0	1.70	1.72	4.87	4.91	26.24	27.66
4.5	1.67	1.66	4.76	4.63	23.66	25.01
5.5	1.52	1.51	4.14	3-98	20.58	21.83
6.0	1.44	1.33	3.80	3.59	19.30	18.32
6·Š	1.12	0.99	2.85	2.60	13.33	12.78
7.0	0.76	0.66	1.87	1.64	7.95	7.43
7.5	0.55	0.47	1.25	1.09	4.77	4.42
8.0	0.37	0.39	0.92	0.87	3.53	3.24
9.0	0.36	0.36	0.74	0.77	2.87	2.70

TABLE	2	SPI	ECIFI	C	RATE	E Co	NST	ANTS	(mi	n^{-1})	FOR	THE
UNCATA	LYSE	D.	AND	Cτ	I(II)	CAT	ALY	SED	ATF	' HY	DROI	LYSIS

	[µ =	= 0.1 (KNO	3)]	
ATP species	46°	56°	6 4 °	78°
H ₂ ATP ²⁻		1.87×10^{-4}	5.75×10^{-4}	3.76×10^{-3}
HATP ³⁻		(± 0.10) 1.63×10 ⁻⁴	(± 0.24) 4.50×10 ⁻⁴	(± 0.13) 2.37 × 10 ⁻³
ATP ⁴⁻		3.56×10^{-5}	(± 0.13) 7.55 × 10 ⁻⁵	2.63×10^{-4}
Cu: ATPH-	6.89×10	(± 0.23) 1.90×10 ⁻³ (±0.05)	(± 0.04) 4.20×10 ⁻³ (±0.07)	(±0.2.5)
Cu: ATP ²⁻	7.53×10^{-4}	2.15×10^{-3} (+0.10)	4.80×10^{-3}	
TCu(OH): ATP*	3.94†	10.96†	23.99†	

*TCu(OH): ATP = $[Cu(OH): ATP^{-3}] + [Cu(OH)_2: ATP^{4-}]$ $+[(Cu(OH): ATP)^{\bullet}].$

†Overall pH independent rate constants.

pH region 3-9 gave a single value for each of the three ionic species of ATP. These values and the corresponding values of the activation parameters are listed in Tables 2 and 3 respectively.

First order rate constants for the kinetic runs carried out with equimolar amounts of Cu(II) and ATP are given in Table 4. These constants have been corrected for the spontaneous hydrolysis of the uncomplexed ATP at a particular pH. The rate of the Cu(II) catalysed ATP hydrolysis was found to increase with pH and an optimum value was found around $\rho H 5.50$. The rate then decreases with increasing $p\dot{H}$. The data in Tables 1 and 2 show that in the presence of Cu(II) ions, the rate of ATP hydrolysis is higher than the uncatalysed rate, over the entire pH range of 3-8.

The thermodynamic data of Taqui Khan and Martell¹⁸ have been used to calculate the concentrations of the various Cu:ATP chelate species in solution at 46°, 56° and 64°. From the concentrations of these various species, the corresponding mole fractions have been calculated. The values obtained at 46° are listed in Table 5. The overall

TABLE 3 - ACTIVATION PARAMETERS* FOR THE UNCATALYSED AND Cu(II) CATALYSED ATP HYDROLYSIS

7	04	177370 17
u ===	0.1	(KNO ₀))
L.		1

ATP species	ΔH^{\ddagger} (kcal/mole)	ΔS‡ (e.u.)	ΔF^{\ddagger} (kcal/mole)
H.ATP ³⁻	31.2	10.9	27.6
HATP ⁸⁻	27.8	0.3	27.7
ATP ⁴⁻	20.9	-24.0	28.7
Cu: ATPH-	21.6	-13.7	26.1
Cu: ATP2-	22.0	-12.2	26.0
TCu(OH): ATP†	21.6	3.6	20.4

*Reference temperature = 56°. †TCu(OH): ATP = [Cu(OH): ATP³⁻] + [Cu(OH)₂: ATP⁴⁻] + [(Cu(OH): ATP)₂⁶⁻].

TABLE 4 - EXPERIMENTAL AND CALCULATED RATE CONSTANTS FOR THE Cu(II) CATALYSED ATP Hydrolysis

		0.4	177370 17
μ	==	0.1	(INO3)

¢H	46°	(a)	56°	(b)	64	° (b)
	Expl	Calc.	Expl	Calc.	Expl	Calc.
3.0	1.34	1.36	0.37	0.38	0.84	0.85
3.5	2.80	2.85	0.73	0.76	1.70	1.73
4.0	4.55	4.50	1.29	1.24	2.76	2.71
4.5	5.71	5.65	1.63	1.56	3.46	3.40
5.5	60.10	5.42	19.95	1.40	41.71	2.81
6.0	39.80	3.49	10.36	0.81	26.61	1.48
6.5	21.10	1.39	7.57	0.28	11.06	0.47
7.0	14.13	0.33	4.27	0.06	6.49	0.09
7.5	7.94	0.05	1.76	0.01	4.41	0.01
8.0	4.17	0.01	1.13	0.001	2.15	0.001
(a) F min ⁻¹ $>$	Rate cons < 10 ³ .	tant, m	$in^{-1} imes 10^4$	and (t) rate	constant,

TABLE 5 - MOLE FRACTIONS OF THE VARIOUS Cu:ATP CHELATE SPECIES IN SOLUTION

$$[\text{Temp.} = 46^\circ; \mu = 0.1 \text{ (KNO_3)}]$$

pН	Cu: ATPH-	Cu: ATP ²⁻	Cu: (OH) ATP ³⁻	Cu: (OH) <u></u> ATP ⁴⁻	(Cu : (OH) ATP) ⁶⁻
3.0	0.153	0.041			
3.5	0.214	0.446			
4.0	0.166	0.446			
4.5	0.080	0.677	0.018		
5.0	0.029	0.776	0.064		
5.5	0.008	0.712	0.187	0.009	0.014
6.0		0.464	0.386	0.056	0.058
6.5		0.185	0.486	0.222	0.092
7.0		0.044	0.367	0.531	0.052
7.5		0.007	0.176	0.803	0.012
8.0		0.001	0.064	0.932	0.002

experimentally evaluated first order rate constants in the pH region 3-5 (Table 4), have been resolved into the specific rate constants relating to the catalytically active monoprotonated (Cu:ATPH⁻) and normal (Cu:ATP²⁻) chelate species, and are listed in Table 2. Correlation between the experimental rate constants and the calculated rate constants in the pH region 3-5 (obtained from the specific rate constants of the monoprotonated and normal chelate species) is good, as seen from Table 4



Fig. 1—pH-rate profile (3-5) for the Cu(II) catalysed ATP hydrolysis [Temp. = 56.0°, $\mu = 0.1$ (KNO₃). Solid line calculated rates; O—measured rates. Similar pH-rate profile was obtained at 46° and 64°]



Fig. 2 — Evaluation of the order (dependence with respect to hydrogen ion concentration) in the Cu(II) catalysed ATP hydrolysis [Temp. = 56°, $\mu = 0.1$ (KNO₃). Similar plots were obtained at 46° and 64°]

and Fig. 1. At $\rho H > 5$ the hydrolysis reaction is found to have a 1/2 order dependence with respect to [H+] as shown in Fig. 2. An overall ρH independent hydrolysis rate constant pertaining to the Cu:ATP hydroxospecies has been evaluated and is listed in Table 2.

Activation parameters ΔH^{\ddagger} , ΔS^{\ddagger} ard ΔF^{\ddagger} related to the specific rate constants of the catalytically active Cu:ATP species were determined by studying the kinetics of the Cu(II) catalysed ATP hydrolysis at 46°, 56° and 64°. These activation parameters are listed in Table 3.

Discussion

Uncatalysed ATP hydrolysis — The kinetics of the spontaneous hydrolysis of ATP is given by the overall rate law

$$\frac{-dL}{dt} = k_{\rm obs} \left[T_L \right]$$

where

$$T_L = [H_2ATP^{2-}] + [HATP^{3-}] + [ATP^{4-}]$$

and

$$k_{\rm obs} = k_1 M_1 + k_2 M_2 + k_3 M_3$$

 k_1 , k_2 and k_3 are the specific rate constants and M_1 , M_2 and M_3 are the corresponding mole fractions of the H₂ATP²⁻, HATP³⁻ and ATP⁴⁻ species, respectively.

The three ionic forms of ATP present in the solution in the pH range 3-9 are shown in Fig. 3. The specific rate constants for these three ionic forms of ATP, listed in Table 2, show that the diprotonated form, H₂ATP²⁻, is the most kinetically active, followed by the moroprotonated species, HATP³⁻ The fully dissociated form, ATP⁴⁻ is kinetically the slowest of the three species.

It is difficult to specify precisely, the mechanisms by which the various forms of ATP undergo hydrolysis, because for the conditions under which the reaction occurs, there is no appropriate criterion



Fig. 3 - Dissociation steps for adenosine-5'-triphosphate

of molecularity with respect to water. The hydrolysis of a particular species of ATP may take place either by a S_N1 mechanism, involving the elimination of a metaphosphate intermediate, or by a S_N2 mechanism involving nucleophilic attack by a polar water molecule on the said ionic species of ATP in solution.

The entropies of activation, ΔS^{\ddagger} , for the hydrolysis reactions of esters, are of much importance in distinguishing between the $S_N 1$ and $S_N 2$ mechanisms. From the study of the ΔS^{\ddagger} of a large number of ester hydrolysis reactions, Schaleger and Long¹⁹ have postulated that S_N1 mechanisms are accompanied by small entropies of either sign, while the $S_N 2$, proceed with large negative entropies of activation. The ΔS^{\ddagger} values for the hydrolysis of the three ionic species of ATP, listed in Table 3, show that the diprotonated and the monoprotonated species of ATP have positive values of 10.9 e.u. and 0.3 e.u. respectively, while the fully dissociated species has a ΔS^{\dagger} value of -24.0 e.u. In the light of the above observations, it may be postulated that the di- and mono-protonated species of ATP hydrolyse by $S_N 1$ mechanism (Fig. 4A) while the fully dissociated form of ATP hydrolyses by a S_N^2 mechanism (Fig. 4B). Further support for the above postulation is obtained from the work of Miller and Westheimer²⁰ on the hydrolysis of γ -phenylpropyl di-(ADP)- and tri-(ATP)-phosphates. The study was carried out at 95° in the pH range 0.3-10, various buffers being used to maintain a constant pH. These authors have shown that in the pH region 3-6, wherein the H_2ATP^{2-} and $HATP^{3-}$ species predominate, the hydrolysis reaction proceeds by the elimination of a monomeric metaphosphate anion. Further confirmation of the mechanism of hydrolysis can be obtained by a detailed investigation of the effect of ionic strength, effect of D₂O, effect of a nucleophile like pyridine, or by carrying out the reaction in a non-pelar solvent like acetonitrile.

Cu(II) catalysed ATP hydrolysis — A detailed potentiometric study²¹ has shown that the 1:1 metal-ATP complexes [where metal = Cu(II), Zp(II), Mn(II), Mg(II) and Ca(II)] have no affinity to further complex with another ATP molecule or a sulphosalicylic acid (SSA) molecule, to form a 1:2 metal-



Fig. 4 — Mechanisms for the uncatalysed hydrolysis of adenosine-5'-triphosphate

ATP or a 1:1:1 metal-ATP-SSA complexes, respectively. This lack of interaction is attributed to the electrostatic repulsion between the negatively charged 1:1 complexes and a second anionic ATP⁴⁻ or SSA molecule. Since 1:2 Cu-ATP complexes are not formed, the kinetics of the Cu(II)-catalysed ATP hydrolysis has been carried out with a 1:1 mole ratio of Cu(II) and ATP.

Potentiometric study of 1:1 Cu-ATP systems¹⁸ has shown that Cu(II) interacts strongly with ATP to give rise to a series of complexes, namely a protonated Cu:ATPH-, a normal Cu:ATP2-, a monohydroxo Cu:(OH)ATP³⁻, a dihydroxo Cu:(OH)₂ATP⁴⁻ and a dimeric species [Cu:(OH)ATP]⁶⁻. The mole fractions of these various species at 46° (listed in Table 5) show that up to pH 5, the Cu:ATP species ir solution predominently consist of Cu:ATPH- and Cu:ATP²⁻ species on ly, the hydroxo species being \angle^2 10% of the total metal chelate species. Concentration of the hydroxo species increases rapidly with ρ H, and at ρ H 8 their concentration approaches 99% of the total Cu:ATP chelate species in solution. In the acidic pH range 3-5 the observed rate can be assigned to the predominant Cu:ATPH- and Cu:ATP²⁻ species. In this pH range, the rate law for the hydrolysis of the Cu:ATP system is given by the relationship

where

and

$$T_L = [Cu:ATPH^-] - [Cu:ATP^{2\bullet}]$$

$$k_{\text{obs}} = k_1 M_1 + k_2 M_2$$

 $\frac{-dL}{dt} = k_{\rm obs}[T_L]$

 k_1 and k_2 are the specific rate constants and M_1 and M_2 are the corresponding mole fractions of the Cu: ATPH- and Cu:ATP²⁻ species respectively. Comparison of the specific rate constants of the Cu:ATPH- and Cu:ATP²⁻ species with those of the corresponding free ligand species ATPH³⁻ and ATP⁴⁻ respectively (Table 2) shows that Cu(II) ions catalyse the rate of ATP hydrolysis considerably. The ratio of the rates for the catalysed and uncatalysed reaction of the above-mentioned species are about 12 and 60 respectively at 56°. Thus the metal ion catalysis is most effective for the fully dissociated form of ATP, i.e. ATP⁴⁻.

The catalytic activity of the Cu(II) ions on the rate of ATP hydrolysis can be directly attributed to the changes brought about in the charge of the ATP molecule and the polarization of the P-O bonds. On complexation with the ATP molecule, Cu(II) ions reduce the negative charge on the molecule by two units. Due to this screening of the negative charge on the phosphorous oxygen atoms, the approach of a nucleophile to the site of attack, will be greatly facilitated leading to enhanced rates of hydrolysis. Further, complexation of the Cu(II) ions with the oxygen atoms of the phosphate chain will result in the withdrawal of electrons from the oxygen atoms towards the sites of metal binding. This leads to polarization of the P-O bonds with an enhancement of positive charge dersity on the phosphorous atom, and a facile nucleophilic attack on the polarized phosphorous atom.

The data in Table 2 show that the normal complex species Cu:ATP²⁻ reacts at a slightly faster rate than the monoprotonated species Cu:ATPH-. The latter species has the advantage of a lower, negative charge and the gamma phosphorous atom at the site of attack is polarized by both the metal ion and the proton. However, the much greater interaction of Cu(II) ions in the Cu:ATP²⁻ species (log $K_{\rm ML} = 5.83$ at 56°) leading to a greater polarization of the P-O bond, is more than enough to compensate for the lower negative charge and the combined polarizing effect in the Cu: ATPH species $(\log K_{\rm MHL} = 2.91 \text{ at } 56^{\circ}).$

The entropies of activation, for the hydrolysis of the monoprotonated and the normal chelate species are -13.7 e.u. and -12.2 e.u. respectively (Table 3). Entropies of this order of magnitude are associated with hydrolysis reactions that proceed by a $S_N 2$ mechanism¹⁹. The monoprotonated uncomplexed ATPH3- species has been suggested to undergo hydrolysis by a S_N1 mechanism, involving the transfer of a proton to the leaving group, and elimination of the metaphosphate anion (Fig. 4A). In the corresponding monoprotonated metal complex, Cu:ATPH-, the negative charge on the gamma phosphorous atom, which is the driving force behind the S_N1 mechanism for the ATPH³⁻ species, is absent. Further complexation of the metal ion with the oxygens of the beta and gamma phosphates^{22,23} hinders the free rotation of the, terminal phosphate moiety, and the transfer of a proton to the leaving group, required for the S_N mechanism, is thus rendered difficult. Due to these reasons the Cu:ATPH- species seems to undergo hydrolysis via a $S_N 2$ mechanism.

For the normal complex species, Cu:ATP²⁻, the entropy of activation for hydrolysis (-12.2 e.u.) is much more positive than the value (-24 e.u.) observed for the fully dissociated species, ATP4-. If the Cu:ATP²⁻ species is also assumed to react with the nucleophile: OH_2 by a $S_N 2$ mechanism, the more positive entropy of activation for the metal chelate species as compared to the corresponding uncomplexed ATP4- species, may be readily explained on the basis of the decreased repulsion between the Cu:ATP²⁻ species and the attacking polar water molecule. In the light of the above discussion, the hydrolysis of ATP in the presence of Cu(II) ions can be visualized as a S_N^2 nucleophilic attack of a polar water molecule on the terminal phosphate moiety of ATP, as evidenced by the work of Moll et al.24. These proposed mechanisms are diagramatically shown in Figs. 5A and B.

Above pH 5, the hydrolysis reaction mechanism is complicated by the formation of hydroxo species. After correction for the contribution of the Cu: ATPH- and Cu:ATP²⁻ species, the overall rate constants should be the sum of the contributions of the monohydroxo, dihydroxo and dimeric species of ATP. Equations assuming a simple first order dependence on the substrate species did not give any constant values for the specific rate constants of the three hydroxo species. Unlike the acidic region, the mechanism of hydrolysis must involve H⁺ or OH- ions in the pH region >5.5. The plot of log k against pH (Fig. 2), is linear with a slope of 1/2,



Fig. 5 - Mechanisms of hydrolysis for the protonated and normal Cu: ATP chelate species

showing a 1/2 order dependence on the [H⁺]. The intercept of the above plot gives an overall (pHindependent) hydrolysis rate constant. The rate law for the hydrolysis of ATP, via the hydroxo species in the pH range 5.5-.80 may then be represented by

$$\frac{-dL}{dt} = k[T_L][\mathrm{H}^*]_{1/2}$$

where

$$T_L = [Cu:(OH)ATP^{3\bullet}] + [Cu:(OH)_2ATP^{4\bullet}] + [(Cu:(OH)ATP^{4\bullet}]]$$

and

k = overall p H independent hydrolysis constant.

The rate constants obtained at 46°, 56° and 64°, and the corresponding activation parameters are listed in Tables 2 and 3. In view of the complex dependence on H⁺ further resolution of the overall rate constants into individual rate constants for the various hydroxo species was not possible with the data on hand. The overall entropy and enthalpy of activation for these species (Table 3) do not lead to any precise mechanism for any one individual hydroxo species.

Acknowledgement

One of the authors, M. Srinivas Mohan, is indebted to the CSIR, New Delhi, for the award of a senior research fellowship.

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