Installation of biologically active pyrimidine moiety into pyridopyrimidine framework and evaluation of their antibacterial activities

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Pyrido[2,3-d]pyrimidines **3** have been synthesized starting from 5-benzylidene barbiturate and maloninitrile in presence of ammonium acetate. Installation of pharmacologically active pyrimidine moiety into pyridopyrimidine framework **3** has been accomplished by condensation of **3** with formic acid, acetic acid, urea, thiourea, formamide and hydrazine hydrate. The antibacterial activities of the synthesized compounds have been evaluated. Good to moderate activities of the tested compounds have been observed.

Keywords: Pyrimidine, pyrido[2,3-d]pyrimidine, 5-arylidene-barbituric acids, maloninitrile

Pyridopyrimidines are annelated uracils that have attracted considerable interest in recent years. The pyrido[2,3-d]pyrimidine ring system is present in a number of biologically active organic compounds which antitumour¹, antibacterial², antiviral³. includes antioxidant⁴. antifungal⁵, hepatoprotective⁶ and anticonvulsant agents². It also has antipyretic⁷, analgesic⁸ and CNS depressant activity⁹. More specifically, pyrido [2,3-d] pyrimidines were considered as inhibitors of Pneumocystis carinii (pc), Toxoplasma gondii (tg), of tumour cell lines in culture¹⁰ and some have anticytokinin activity¹¹. The activity is mainly due to inhibition of dihydrofolate reductase (DHFR)^{12,13}.

In view of significant biological contribution of pyridopyrimidine heterocycles and in continuation of our work on the synthesis of biologically active heterocycles¹⁴, herein is reported the installation of another pyrimidine moiety into pyridopyrimidine framework possessing antibacterial activity with expectations of an enhancement of biological properties of target molecules using barbiturates as synthetic precursors.

Results and Discussion

5-Arylidene-barbituric acids 2 was synthesized by condensation of appropriate barbiturates 3 with equimolar amount of various arylaldehydes in acetic acid as previously reported Scheme I (Ref 15). The one-pot reaction of 5-arylidene-1,3-diphenyl thiobarbituric acids **2a-f**, malononitrile and excess of ammonium acetate in dioxane produced pyridopyrimidine-6-carbonitrile derivatives **3a-f**.

The effect of substituents of aryl system in arylidene derivatives **2a-f** in the formation of **3a-f** has also been studied. The reaction of **2b** (X=S, R=Ph, Ar = p-OMeC₆H₄), and **2c** (R=Ph; p-N(Me)₂C₆H₄) with malononitrile and ammonium acetate afforded the mixture of products **3b,c** isolated from water and the corresponding **4a,b** formed in the reaction set in 51%, 42% and 20%, 35% yield, respectively.

The possible explanation of the reaction mechanism for the product **3a-f** is supposed to occur through non-isolable Michael adduct formed by refluxing the mixture of arylidene **2a-f** and malononitrile under the above said reaction conditions and subsequent *in situ* oxidation of the intermediate. The regioselective reaction of arylidene derivative with malononitrile is monitored by *p*-substituted benzaldehyde to form the products **3a-f** and **4a,b**. In fact, *p*-NO₂, *p*-Cl and *p*-Br substituents at the phenyl ring increases the reactivity at β -carbon atom of α,β -unsaturated carbonyl scaffold of arylidene derivatives due to -M effect. On the contrary, the

CN

selectivity of reaction of malononitrile increases at carbonyl carbon of the arylidene derivatives 2b,c caused by +M effect due to p-OMe and p-Me₂N substituents at phenyl ring (Scheme I). Subsequently, another set of 5-arylidene barbituric acid 2g-j (R = H, X = O) was examined as the replacement of 5arylidene 1,3-diphenyl thiobarbituric acid 2a-f acid in the reaction with malononitrile and ammonium acetate Scheme I. Surprisingly, under the reaction conditions described above, a mixture of 4c-f as major product and **3g-j** being the minor were generated. Here, in case of 5-arylidene barbituric acid (R = H, X = O) the comparative increase in the reactivity of carbonyl function at 4 and 6 positions of the arylidene intermediate 2 (X = O, R = H) renders significant contribution in the formation of the major product 4 due to the presence of uriedo scaffold. This is still a puzzle to us, and further efforts are underway to clarify this problem. The IR spectrum of 2a (R = Ph; X = S) shows bathochromic shift of two carbonyl stretching frequencies from 1728 and 1709 cm^{-1} in 1 to 1712 and 1683 cm⁻¹ suggesting the formation of exocyclic double bond at position-5 of thiobarbiturate scaffold. The characteristic peaks in the region 3176-3446 cm⁻¹ and 2140-2194 cm⁻¹ corresponding to -NH and $-C \equiv N$ stretching band, respectively in the IR spectrum of **3a-f**, clearly demonstrate the formation of pyridopyrimidine derivative 3a-f. Disappearance of one of carbonyl band at 1712 cm^{-1} in 2 and leaving the other carbonyl group unaffected suggests



Scheme I — Synthesis of pyridopyrimidine-6-carbonitrile derivative **3** and 2-[6-dicyanomethylene-5-arylidene-pyrimidin-4-ylidene]-malononitrile **4**

unambiguously the participation of more active carbonyl function in the process of cyclocondensation followed by *in situ* oxidation. The appearance of two carbonyl stretching frequencies at 1688-1691 and 1646-1649 cm⁻¹ along with the presence of nitrile peak at 2129-2136 cm⁻¹ in the IR spectrum, suggests the formation of the pyridopyrimidine derivatives **3g-j**. In the ¹H NMR spectrum of compounds **3a-j**, a broad singlet at δ 4.43-4.96 exchangeable with D₂O and a complex multiplet resonating at δ 6.57-7.91 corresponding to NH₂ and phenyl protons, respectively confirms the formation of **3a-j**. ¹³C NMR spectrum of compound **3a-f** and **3g-j** showed signals at δ 173.15-174.66 and 155.21-157.32, 159.10-159.23 due to carbonyl groups, respectively. In addition, elemental analyses are consistent with the structures proposed for the compounds 3a-j. The formation of dicyano pyrimido derivatives 4a,b was ensured by the disappearance of both carbonyl bands of 2b,c and appearance of nitrile peak at 2221-2222 cm⁻¹ in the IR spectrum while the dicyano pyrimido derivatives 4c-f displayed absorption bands at 2222-2224 cm⁻¹ and 1672-1676 cm⁻¹ corresponding to nitrile and carbonyl groups, respectively. In the ¹H NMR spectrum of **4a-f** a sharp singlet emerged at δ 8.03-8.39 for benzvlidene proton.

In continuation of our work on the synthesis¹⁴ of new fused heterocyclic systems having potential biological activites, we have installed another pyrimidine moiety into pyridopyrimidine framework **3a,b,d-f**. The pyrido [2,3-*d*]pyrimidine-6-carbonitrile **3a,b,d-f** being a β -enaminonitrile derivative, pyrimidine ring was annulated with **3a,b,d-f** in presence of formic acid/acetic acid and catalytic amount of conc. HCl via partial hydrolysis of nitrile group in 3a,b,d-f to amide group followed by dehydrative cyclisation to form pyrimido pyrido pyrimidine derivative 5 in good vield (Scheme II). Disappearance of CN stretching, and appearance of second carbonyl stretching in the lower frequency region at 1645-1647 cm⁻¹ assignable to cyclic amide function in the IR spectrum of **5a-h** are in favour of the proposed structure **5a-h**. In the 1 H NMR spectrum of 5a-d and 5e-h, the methine and methyl protons of pyrimidin-4(3H)-one ring appeared as singlet at δ 8.53-8.61 and δ 1.80-1.89, respectively. The ¹H NMR spectrum of **5a-h** also showed a broad singlet at δ 12.05-12.29 exchangeable with D₂O for NH proton. ¹³C NMR spectrum of compound 5a-h showed signals at δ 160.80-160.98 and δ 159.65-160.21 due to carbonyl group of PhNC=O and -NHC=O,



Scheme II — Synthesis of pyrimido pyrido pyrimidine derivative 5-7

respectively. These signals clearly demonstrate the formation of pyrimido pyrido pyrimidine derivative **5a-h**. The analytical data also supports the formation of the product **5a-h**.

Pyrimido pyrido pyrimidine 6 was constructed by condensation of equimolar pyridopyrimidine derivative 3a, b, d-f and urea/ or thiourea on a sand bath at 170°C in good yield via deaminative cyclisation. The appearance of absorption bands at 3285-3301 cm⁻¹ and 3185-3198 cm⁻¹ corresponding to NH₂ and NH stretchings in the IR spectrum of 6a-h along with absorption bands for carbonyl groups at 1664-1671 for **6a-h** and 1645-1646 cm^{-1} for **6e-h** ensures the formation of pyrimido pyrido pyrimidine derivative **6a-h**. The ¹H NMR spectrum of **6a-h** exhibited two distinct singlets at δ 7.85-8.02 and δ 8.55-8.78 corresponding to two protons of NH₂ and one proton of NH groups, respectively. Moreover, in the ¹³C NMR spectra of compounds 6a-h, the -C=S and -C=O signals were observed at δ 180.07-181.09 and δ 152.95-153.64, respectively. Furthermore, the elemental analyses are compatible with the structure **6a-h**. The reaction of formamide with **3a,b,d-f** under reflux generated pyrimido pyrido pyrimidine derivatives 7. The IR spectrum of compounds 7a-d displayed absorption bands in the region 1668-1670 cm⁻¹ and 3188-3371 cm⁻¹ corresponding to carbonyl and NH₂ groups, respectively. The ¹H NMR spectrum of **7a-d** showed a singlet at δ 9.87-9.98 for CH proton of pyrimidine ring. A multiplet for aromatic protons



Scheme III — Synthesis of pyrimido pyrido pyrimidine derivative 9

appeared at δ 6.60-7.56 and signals due to NH₂ protons were found to be merged with aromatic protons suggesting the formation of **7a-d**.

The condensation of equimolar mixture of 3a, b, **d-f** and triethylorthoformate on an oil bath produced formimidic acid ethyl ester derivative 8 which on subsequent treatment with hydrazine hydrate in refluxing ethanol in presence of NaOEt as catalyst underwent cyclisation to form pyrimido pyrido pyrimidine derivatives 9 (Scheme III). The formation of 8 is ensured on the basis of analytical and IR spectral data. Disappearance of NH stretching in the IR spectrum of **8a-d**, and appearance of a characteristic sharp absorption band at 1612-1617 cm⁻¹ indicate the formation of azomethine group in the product. The retention of nitrile band at 2214-2222 cm⁻¹ in the IR region suggests the non-interfering of nitrile function during the reaction while that of **9a-d** revealed the absence of the nitrile group. The ¹H NMR spectrum of **8a-d** displayed a quartet at δ 4.26-4.30 for CH₂ protons and a triplet at δ 1.29-1.33 corresponding to methyl protons along with a singlet at δ 7.67-7.78 for azomethine proton suggesting the formation of ester derivative **8a-d**. The signals for the NH_2 and NHprotons appearing as distinct broad singlets at δ 12.39-12.60 and δ 12.89-12.97 along with a singlet at δ 8.56-8.78 corresponding to pyrimidine proton confirm the formation of pyrimido pyrido pyrimidine derivative 9a-d. Further, C=NH in ¹³C NMR spectrum of 9a-d showed peak at δ 161.92-162.89.

Antibacterial activity

The antibacterial study of the synthesized compounds was carried out using disc diffusion and

broth dilution method^{16,17}. The synthesized compounds were screened for their antibacterial activity in vitro against Gram negative bacterium (GNB) Escherichia coli isolated from local pond water of Sambalpur University Campus, Odisha, India and Gram positive bacterium (GPB) Staphylococcus aureus isolated from drug-vial contaminants, were identified in Medical Microbiology Laboratory, School of Life Science, Sambalpur University by following routine diagnostic microbiological tests. Activity of each compound was compared with Penicilin-G. Erythromycin, Ampicilin, Cephalothin, Clindamycin, Co-Trimoxazole, and Kanamycin as standards against Gram positive and Gram negative strains respectively. The minimum inhibitory concentrations (MIC) were determined by broth dilution technique. To the nutrient broth, the overnight bacterial cultures diluted at a ratio of 1:100 counting 10^3 CFU/mL (colony forming unit) was added and graded concentrations (0.2-500 µg/mL) of the synthesized compounds in dimethyl sulfoxide were added. The cultures were incubated for 12-14 h at 37°C. Visible turbidity in the MIC tubes was taken as the parameter for determination of minimum inhibitory concentration (MIC). The lowest concentration (highest dilution) of the drug required to arrest the growth of bacteria was determined as MIC (µg/mL).A number of antimicrobial discs are placed on the agar for the sole purpose of producing zones of inhibition in the bacterial lawn. Agar media were poured into each Petri dish. Excess of suspension was decanted and placed in incubator at 37°C for 1 h to dry the plates¹⁷. The disc measuring 5 mm in diameter were prepared from Whatman no.1 filter paper and sterilized by dry heat at 140°C for 1 h. The sterile discs were placed in nutrient agar medium and 50 µL of minimum inhibitory concentration of the test compounds in DMSO were delivered into each labeled disc. Running control with pure DMSO as solvent, has shown no effect. The discs were prepared in triplicate in each Petri dish and incubated for 48 h at 37°C. Antibacterial activity was determined by measuring the diameter of inhibition zone. Activity of each compound was compared with Penicilin-G (10 µg), Erythromycin (10 µg), Ampicilin (25 µg), Cephalothin (30 μ g), Clindamycin (2 μ g), Co-Trimoxazole (25 µg), and Kanamycin (5 µg) as standard. The antibacterial activity was classified on the basis of diameter of inhibition zone such as highly active (>21 mm), moderately active (15-21 mm), least active (12-15 mm) and less than 12 mm was

taken as inactive. The zones of inhibition of the standard drugs and the synthesized compounds is summarized in **Table** I. The investigation of preliminary antibacterial screening data (Table I) revealed that most of the tested compounds showed moderate to good bacterial inhibition. Compounds 5c, 5f, 5g, 6f, 7b, 8b, and 9b exhibited good antibacterial activity against Staphylococcus spp. and E. coli strains. Compound 5f, 7b, 8a, 8b, and 9b showed good activity against both S. aureus and E. coli bacterial strains whereas compounds 5c, and 5g illustrated good activity against E. coli bacterial strain. It is interesting to note that installation of another pyrimidine moiety into pyridopyrimidine framework 3 increases the antibacterial activity. Enhanced activity has been observed in cases of compounds having *p*-OMe and *p*-Cl substituents in the phenyl rings.

Experimental Section

Melting points were taken in open capillaries using sulfuric acid bath and are uncorrected. IR spectra were recorded with a Shimadzu FT-IR Prestige-21 spectrophotometer in potassium bromide (KBr) using diffuse reflectance system (DRS) technique. NMR spectra were recorded on a VARIAN 400 MHz (400 MHz for ¹H, 100 MHz for ¹³C) NMR spectrometer in dimethyl sulfoxide- d_6 (DMSO), unless otherwise stated. Chemical shifts are expressed in parts per million downfield from tetramethylsilane as an internal standard. Data are given in the following order: δ value, multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad), number of protons, coupling constants J is given in Hertz. Elemental analyses were performed by Flash 2000 CHN elemental analyzer, expressed in % and were in agreement with the calculated values within $\pm 0.4\%$. All the reagents and solvents used were of the best grade available and were used without further purification.

General procedure for the synthesis of pyrido[2,3*d*]pyrimidine-6-carbonitrile derivatives 3a-j and (pyrimidin-4-ylidene)-malononitrile derivatives, 4a-f

A mixture of the 5-benzylidenepyrimidine 2 (10 mmol), malononitrile (10 mmol) and ammonium acetate (90 mmol) in dioxane (60 mL) was refluxed with stirring for 6 h. The reaction mixture was allowed to cool to RT and the solution was poured onto crushed ice. The precipitated product thus formed was filtered, dried and recrystallised from ethanol to afford the desired product.

				Zone of Inhibition in mm (MIC in µg/mL)	
Compd	Ar	R	Х	Gram Positive Bacteria ^a	Gram Negative Bacteria ^a
-				Sa	Ec
3a	Ph	_	S	16(26.5)	18(30.1)
3b	<i>p</i> -OMeC ₆ H ₄	_	S	19(28.5)	20(29.5)
3c	$p-N(Me)_2C_6H_4$	_	S	17(25.4)	20(26.4)
3d	$p-ClC_6H_4$	_	S	19(26.4)	20(25.4)
3e	p-BrC ₆ H ₄	_	S	14(27)	16(27.4)
3f	$p-NO_2C_6H_4$	-	S	16(27.1)	18(27.7)
3g	Ph	-	0	14(41)	13(32)
3h	<i>p</i> -OMeC ₆ H ₄	-	0	18(32.2)	16(33.1)
3i	p-ClC ₆ H ₄	-	0	16(34.2)	16(36.4)
3ј	p-BrC ₆ H ₄	-	0	13(35.6)	14(34.2)
5a	Ph	Н	-	18(23.3)	19(25.2)
5b	p-OMeC ₆ H ₄	Н	-	19(24.5)	20(25.4)
5c	p-ClC ₆ H ₄	Н	-	20(23)	22(24.1)
5d	p-BrC ₆ H ₄	Н	-	17(26.5)	16(26.2)
5e	Ph	Me	-	19(24)	20(26.3)
5f	<i>p</i> -OMeC ₆ H ₄	Me	-	21(22.9)	21(22.1)
5g	p-ClC ₆ H ₄	Me	-	18(24.6)	22(25.4)
5h	p-BrC ₆ H ₄	Me	-	14(27)	16(27.4)
6a	Ph	-	0	17(24.2)	18(25.1)
6b	p-OMeC ₆ H ₄	-	0	18(25.3)	19(27.2)
6c	p-ClC ₆ H ₄	-	0	17(24.3)	19(23)
6d	p-BrC ₆ H ₄	-	0	16(26.1)	18(26.3)
6e	Ph	-	S	19(23.9)	19(25.3)
6f	<i>p</i> -OMeC ₆ H ₄	-	S	19(25.1)	21(26.2)
6g	p-ClC ₆ H ₄	-	S	18(24)	20(23.2)
6h	p-BrC ₆ H ₄	-	S	17(24.6)	19(25.2)
7a	Ph	-	-	18(23)	19(25)
7b	p-OMeC ₆ H ₄	-	-	20 (25.6)	21(26.5)
7c	p-ClC ₆ H ₄	-	-	16(24.2)	20(23.5)
7d	p-BrC ₆ H ₄	-	-	15(24.8)	19(25.9)
8a	Ph	-	-	20(22.1)	19(24.9)
8b	p-OMeC ₆ H ₄	-	-	21(25)	20(25.9)
8c	p-ClC ₆ H ₄	-	-	19(24.2)	18(23.6)
8d	p-BrC ₆ H ₄	-	-	16(24.2)	17(25.6)
9a	Ph	-	-	19(23.7)	17(25.1)
9b	p-OMeC ₆ H ₄	-	-	21(24.5)	20(25.1)
9c	p-ClC ₆ H ₄	-	-	17(22.7)	19(23)
9d	p-BrC ₆ H ₄	-	-	18(24.9)	17(26.1)
Penicilin-G ^o	_	-	-	0	0
Erythromycin	_	-	-	22	20
Ampicilin	-	_	_	0	0
Cephalothin ^o	_	-	-	0	0
Clindamycin [°]	-	_	_	26	12
Co-Trimoxazole"	-	_	_	0	21
Kanamycin	—	-	-	12	15.5

In case when X, R, Ar = S, Ph, *p*-OMeC₆H₄; S, Ph, *p*-N(Me)₂C₆H₄; O, H, Ph; O, H, *p*-OMeC₆H₄; O, H, *p*-ClC₆H₄; O, H, *p*-BrC₆H₄, the solid formed in the reaction set during reflux was filtered, dried and recrystallised from DMF to give the respective (pyrimidin-4-ylidene)-malononitrile derivatives **4a-f**. Thereafter, the filtrate was poured onto crushed ice. The solid thus formed was filtered, dried and recrystallised from ethanol to yield the respective pyrido[2,3-*d*]pyrimidine derivatives **3b,c** and **3g-j**.

7-Amino-4-oxo-1,3,5-triphenyl-2-thioxo-1,2,3,4-

tetrahydropyrido [2,3-d]pyrimidine-6-carbonitrile, 3a: Yield 80%. m.p.150°C. IR (KBr): 3429, 3331, 3176 (-NH₂), 2196 (C=N), 1683 cm⁻¹ (C=O); ¹H NMR (DMSO-*d*₆, 400 MHz): $\delta_{\rm H}$ 4.43 (br, 2H, -NH₂, D₂O exchangeable), 6.96-7.61 (m, 15H, PhH); ¹³C NMR (DMSO-*d*₆, 400 MHz,): $\delta_{\rm C}$ 79.79, 110.36, 116.39, 124.82, 125.21, 125.9, 126.54, 127.92, 128.18, 128.49, 129.32, 129.79, 133.76, 134.43, 135.02, 158.95, 159.69, 160.95, 167.29, 173.15. Anal. Calcd for C₂₆H₁₇N₅OS: C, 69.80; H, 3.80; N, 15.66. Found: C, 69.54; H, 3.51; N, 15.36%.

2-[6-Dicyanomethylene-5-(4-methoxy-benzylidene)-1,3-diphenyl-2-thioxo-tetrahydro-pyrimidin-4-ylidene]malononitrile, 4a: Yield 20%. m.p.150°C. IR (KBr): 2222 cm⁻¹ (C≡N); ¹H NMR (DMSO-*d*₆, 400 MHz): $\delta_{\rm H}$ 3.88 (s, 3H, Me), 6.90-7.10 (m, 4H, PhH), 7.17-7.20 (m, 4H, PhH), 7.41-7.44 (m, 4H, PhH), 7.95-7.98 (m, 2H, PhH), 8.39 (s, 1H, C=CH); ¹³C NMR (DMSO-*d*₆, 400 MHz,): $\delta_{\rm C}$ 55.19, 55.85, 76.78, 113.85, 114.74, 115.14, 124.05, 126.45, 127.72, 129.87, 133.30, 160.39, 161, 164.28, 173. Anal. Calcd for C₃₀H₁₈N₆OS: C, 70.59; H, 3.53; N, 16.47. Found: C, 70.27; H, 3.25; N, 16.16%.

1,3,5-Triphenyl-2-thioxo-1,2,3,4-tetrahydropyrimido [4",5":2',3']pyrido[5,6-b]pyrimidine-4,6(7*H*)dione, 5a

A mixture of compound **3a** (1 mmol), formic acid (10 mL) and a catalytic amount of concentrated hydrochloric acid was refluxed at 160°C for 6 h. The reaction mixture was allowed to cool to RT and poured into ice cold water. The solid thus formed was filtered, dried and recrystallised from DMF to give the respective pyrimido pyrido pyrimidine derivative **5a**. Yield 75%. m.p.220°C. IR (KBr): 3196 (NH), 1689, 1645 cm⁻¹ (C=O); ¹H NMR (DMSO-*d*₆, 400 MHz): $\delta_{\rm H}$ 6.79-7.90 (m, 15H, PhH), 8.53 (s, 1H, C8-H), 12.25 (br, 1H, -NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 400 MHz,): $\delta_{\rm C}$ 99.21, 107.42, 121.32, 123.99, 124.64, 126.55, 127.36, 128.95, 129.12, 129.38, 132.81, 136.85, 137.96, 145.45, 149.59, 151.21, 160.03, 160.98, 162.22, 177.03. Anal. Calcd for $C_{27}H_{17}N_5O_2S$: C, 68.21; H, 3.58; N, 14.74. Found: C, 67.96; H, 3.32; N, 14.42.

6-Amino-1,3,5-triphenyl-2-thioxo-1,2,3,4-tetrahydropyrimido[4",5":2',3']pyrido[5,6-b]pyrimidine-4,8(9*H*)dione, 6a

A mixture of compound **3a** (1 mmol), and urea (1 mmol) was heated at 170°C in a test tube on a sand-bath for 4 h. The mixture was allowed to cool to RT and to it DMF (20 mL) was added. The solution was poured into ice cold water and the solid thus formed was filtered, dried and recrystallised from DMF to give the respective pyrimido pyrido pyrimidine derivative 6a. Yield 72%. m.p.245°C. IR (KBr): 3298 (-NH₂), 3194 (NH), 1664, 1645 cm⁻¹ (C=O); ¹H NMR (DMSO- d_6 , 400 MHz): $\delta_{\rm H}$ 6.94-7.62 (m, 15H, PhH), 7.95 (s, 2H, -NH₂, D₂O exchangeable), 8.67 (br, 1H, -NH, D_2O exchangeable); ¹³C NMR (DMSO-*d*₆, 400 MHz,): δ_C 90.12, 98.56, 122.16, 124.32, 124.75, 126.51, 127.33, 128.29, 129.04, 129.52, 129.86, 133.02, 136.58, 138.05, 143.12, 151.34, 153.52, 156.22, 160.87, 163.33, 177. Anal. Calcd for C₂₇H₁₈N₆O₂S: C, 66.12; H, 3.67; N, 17.14. Found: C, 65.84; H, 3.39; N, 16.85%.

6-Amino-1,3,5-triphenyl-2-thioxo-1,2,3,4-tetrahydropyrimido[4",5":2',3']pyrido[5,6-b]pyrimidine-4-one, 7a

A mixture of compound **3a** (1 mmol), formamide (5 mL), and formic acid (2 mL) was refluxed in DMF (50 mL) for 6 h. The reaction mixture was allowed to cool to RT, poured into water and neutralized with ammonia solution. The solid thus formed was filtered, dried and recrystallised from DMF to give the respective pyrimido pyrido pyrimidine derivative 7a. Yield 70%. m.p.185°C. IR (KBr): 3361, 3309, 3188 $(-NH_2)$, 1668 cm⁻¹ (C=O); ¹H NMR (DMSO- d_6 , 400 MHz): δ_H 7.00-7.56 (m, 17H, PhH and -NH₂), 9.96 (s, 1H, C8-H); ¹³C NMR (DMSO-*d*₆, 400 MHz,): δ_C 98.05, 102.34, 121.30, 124.06, 124.60, 126.58, 127.34, 128.91, 129.09, 129.27, 132.75, 136.78, 138, 151.20, 155.23, 156.06, 156.99, 157.34, 160.98, 162.76, 177. Anal. Calcd for C₂₇H₁₈N₆OS: C, 68.35; H, 3.80; N, 17.72. Found: C, 68.07; H, 3.53; N, 17.43%.

N-(6-Cyano-4-oxo-1,3,5-triphenyl-2-thioxo-1,2,3,4tetrahydro-pyrido[2,3-d]pyrimidin-7-yl)-formimidic acid ethyl ester, 8a

A mixture of compound 3a (2 mmol) and triethyl orthoformate (16 mmol) was stirred under reflux in

acetic anhydride (20 mL) for 6 h. The reaction mixture was allowed to cool to RT, poured into water and neutralized with ammonia solution. The solid thus formed was filtered, dried and recrystallised from ethanol to give the respective formimidic acid ethyl ester derivative 8a. Yield 85%. m.p.145°C. IR (KBr): 2214 (C=N), 1683 (C=O), 1612 cm⁻¹ (C=N); ¹H NMR (DMSO- d_6 , 400 MHz): δ_H 1.29 (t, 3H, J = 7.2 Hz, Me), 4.30 (q, 2H, J = 7.2 Hz, CH₂), 7.07-7.64 (m, 15H, PhH), 7.78 (s, 1H, HC=N); 13 C NMR (DMSO- d_6 , 400 MHz,): δ_C 21.03, 62.10, 88.32, 112.54, 117, 124.84, 125.13, 125.89, 126.55, 127.25, 27.97, 128.52, 129.11, 129.80, 132.15, 135.21, 136.29, 156.04, 157.12, 159.45, 160.85, 169.76, 175.21. Anal. Calcd for C₂₉H₂₁N₅O₂S: C, 69.18; H, 4.17; N, 13.92. Found: C, 68.88; H, 3.91; N, 13.60%.

7-Amino-6-imino-1,3,5-triphenyl-2-thioxo-1,2,3,4tetrahydro-pyrimido[4",5":2',3]pyrido-[5,6-b]pyrimidine-4-one, 9a

A mixture of compound **3a** (1 mmol) and hydrazine hydrate (99%, 10 mmol) was refluxed for 3 h in ethanol (20 mL) in presence of sodium ethoxide [prepared by dissolving sodium metal (10 mmol, 0.23) g) in 20 mL absolute ethanol]. The reaction mixture was allowed to cool to RT, poured into water and neutralized with 2N HCl. The solid thus formed was filtered, dried and recrystallised from ethanol to give the respective pyrimido pyrido pyrimidine derivative 9a. Yield 73%. m.p.160°C. IR (KBr): 3435, 3317 (-NH₂), 3197 (-NH), 1689 cm⁻¹ (C=O); ¹H NMR (DMSO-*d*₆, 400 MHz): δ_H 7.22-8.04 (m, 15H, PhH), 8.78 (s, 1H, C8-H), 12.60 (br, 2H, -NH₂, D₂O exchangeable), 12.89 (br, 1H, -NH, D₂O exchangeable); ¹³C NMR (DMSO-*d*₆, 400 MHz,): δ_C 96.7, 103.12, 121.09, 123.99, 124.38, 124.87, 126.76, 127.55, 128.23, 129.12, 129.34, 129.56, 131.77, 136.07, 137.98, 146.15, 150.03, 160.12, 162.43, 164.01, 177.12. Anal. Calcd for C₂₇H₁₉N₇OS: C, 66.26; H, 3.88; N, 20.04. Found: C, 66.02; H, 3.60; N, 19.73%.

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References

- (a) Piper J R, Montgomery J A & Sirotnak F M, US Patent 4725687 (1989); Chem Abstr, 109 (1989) 129064x; (b) Piper J R, Montgomery J A, Sirotnak F J & DeGraw I, US Patent 5077404 (1991); Chem Abstr, 116 (1992) 128965z.
- 2 Ram V J, Berghe D A V & Vlietinck A J, J Heterocycl Chem, 28 (1988) 217.
- 3 Tenser R B, Gaydos A & Hay K A, Antimicrob Agents Chemother, 45 (2001) 3657.
- 4 de la Cruz J P, Carrasco T, Ortega G & de la Cuesta F S, Lipids, 27 (1992) 192.
- 5 Nizamuddin, Mishra M, Srivastava M K & Khan M H, Indian J Chem, 40B (2001) 49.
- 6 Ram V J, Goel A, Sarkhel S & Maulik P R, *Bioorg Med Chem*, 10 (2002) 1275.
- 7 Piper J R, McCalab G S, Montgomery J A, Kisliuk R L, Gaumont Y & Sirotnak F M, *J Med Chem*, 29 (1986) 1080.
- 8 Robins R K & Hitchings G H, *J Am Chem Soc*, 80 (1958) 3449.
 9 Madkour H M F, Salem M A I, Abdel-Rahman T M &
- Azab M E, *Heterocycles*, 38 (1994) 57.
 Lowe J A, *Austrian Patent* 388378 (1989); *Chem Abstr*, 112 (1990) 21008.
- 11 Kouichiro G, Kurimoto Y & Kitamura N, *Eur Patent Appl* 243311 (1987); *Chem Abstr*, 108 (1988) 75422.
- 12 Gangjee A, Adair O O & Queener S F, J Med Chem, 46 (2003) 5074.
- 13 Iwamura H, Murakami S, Koshimizu K & Matsubara S, J Med Chem, 28 (1985) 577.
- (a) Behera A K, Behera R K, Pradhan R, Pati A & Patra M, Indian J Heterocycl Chem, 16 (2006) 167; (b) Behera R K, Behera A K, Pradhan R & Patra M, Indian J Chem, 45B (2006) 933; (c) Behera R K, Behera A K, Pradhan R, Pati A & Patra M, Synth Commun, 36 (2006) 3729; (d) Behera R K, Behera A K, Pradhan R, Pati A & Patra M, J Chem Environ, 10 (2006) 18; (e) Behera R K, Behera A K, Pradhan R, Pati A & Patra M, Phosphorus, Sulfur and Silicon, 184 (2009) 753; (f) Behera R K, Pati A, Patra M & Behera A K, Phosphorus, Sulfur and Silicon, 184 (2009) 2827.
- 15 Das P K, Behera G B, Sahay A K & Mishra B K, Indian J Chem, 24B (1985) 437.
- 16 Pattnaik S D, Padhan K & Jana G K, *J Pharm Res*, 3 (2010) 414.
- 17 Gillespie S H, *Medical Microbiology-Illustrated* (Butterworth Heinemann Ltd., Oxford, UK), 234 (1994).