

## Association of Polymorphisms Calpastatin Gene with Body Weight of Local Sheep in Jonggol, Indonesia

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### ABSTRACT

Calpastatin (CAST) gene is located on the fifth chromosome of sheep and plays important roles in formation of muscles and meat tenderness after slaughtering. Association of genetic polymorphism in the CAST gene locus *MspI* and *NcoI* with body weight was examined in local sheep from Jonggol Animal Science Teaching and Research Unit (JASTRU), Faculty of Animal Science, Bogor Agricultural University. The genotypes for CAST were determined by the PCR-RLFP method. Blood samples were collected from 264 local sheep belonging to JASTRU located in Singosari Village, Bogor District, West Java Province. Extraction of genomic DNA was based on the phenol chloroform method. CAST locus *MspI* had three genotypes including in MM, MN and NN with frequencies of 0.75, 0.23, and 0.02 respectively. CAST locus *NcoI* had two genotypes including in MM and MN with frequencies of 0.92, 0.08 respectively. Chi-square test confirmed Hardy-Weinberg equilibrium for the CAST locus *MspI* and *NcoI*. There was no significant effects ( $P > 0.05$ ) of CAST locus *MspI* and *NcoI* genotypes on body weight of local sheep in JASTRU.

**Key words:** CAST locus *MspI*, CAST locus *NcoI*, body weight, local sheep

### INTRODUCTION

Marker assisted selection is one of new DNA based methods that improves accuracy and progress of selection in animal programmes. Selection using genetic markers are commonly performed to improve productivity in the livestock industry. In general the most documented gene for meat tenderness is calpastatin gene. Calpastatin (CAST) is a specific inhibitor of calpains, making the CAST gene becomes an excellent candidate for controlling meat traits in livestock.

CAST gene is located on the fifth chromosome of sheep and plays important roles in formation of muscles, degradation and meat tenderness after slaughtering. Associations between variation in CAST and carcass and meat quality traits in cattle have been reported previously (Casas *et al.*, 2006; Schenkel *et al.*, 2006) and in sheep, there was also a genetic variation in the CAST gene (Palmer *et al.*, 2000; Zhou *et al.*, 2007; Sumantri *et al.*, 2008; Gabor *et al.*, 2009).

A high degree of polymorphism at the CAST locus had also been reported in studies with Angus bulls

(Chung *et al.*, 2001), crossbred steer and pigs (Kurly *et al.*, 2003). Chung *et al.* (2001) and Tahmoorespour (2005) have described that there were three allele systems of polymorphic variants (CAST a, b, and c) analysed by PCR-SSCP in a region of the ovine and cattle CAST. Kurly *et al.* (2003) observed that pigs with the genotype DD at locus CAST/*MspI* and FF at locus CAST/*RsaI* had less fatty acid, thinner back fat and a lower weight of back fat with skin.

The Jonggol Animal Science Teaching and Research Unit (JASTRU) Faculty of Animal Science-Bogor Agricultural University is a Breeding Farm Station located in Singosari Village, Bogor District, West Java Province. Local sheep in JASTRU were produced from crosses and selection programmes since 1980. Sheep keeping under extensive management in JASTRU can adapt on hot tropical climate and dry environment. Genetic variation of sheep from JASTRU was higher compared to another local sheep such microsatellite DNA variation (Sumantri *et al.*, 2008) and gene *Pit1* polymorphism (Sumantri *et al.*, 2009). Body weight of local sheep in JASTRU still varied, increased body weight of local sheep can be done through selection and crossing in a sustainable system. These observations suggest that CAST may be considered as a candidate gene for body weight and lean meat in cattle and sheep. In this study

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we have chosen CAST as a candidate gene for body weight in local sheep because of the evidence in cattle (Chung *et al.*, 2001), pig (Kurly *et al.*, 2003) and sheep (Nassiry *et al.*, 2006) implicating a role for the CAST gene in skeletal muscle (Edyta *et al.*, 2002). The aim of this study was to determine existence of any association of polymorphism at the CAST locus *MspI* and *NcoI* with body weight characteristics of the local sheep in the JASTRU.

## MATERIALS AND METHODS

### Samples

Blood samples were collected from 264 local sheep belonging to the JASTRU Breeding Station located in Singosari Village, Bogor District, West Java Province, Indonesia.

### Procedure

**DNA extraction.** DNA was extracted from 200  $\mu$ l of blood as described by Sambrook *et al.* (1989) by using proteinase K digestion. Blood samples previously stored in absolute alcohol were washed with deionized water and vortexed for 5 min, and then centrifuged at 8000 rpm for 5 min. Pellet was added 400  $\mu$ l lysis buffer containing 10% SDS 40  $\mu$ l, proteinase K 10  $\mu$ l, and 1x STE 350  $\mu$ l incubated at 55 °C for 2 hr. The mixture was added phenol solution 400  $\mu$ l, CIAA (chloroform: isoamyl alcohol = 24:1) 400  $\mu$ l, and 5 M NaCl 40  $\mu$ l and then shaken slowly at room temperature for 1 hr.

The mixture was centrifuged gently at 12,000 rpm for 5 min. Supernatant was collected in fresh sterilized tubes 1.5 ml. The DNA was added with absolute alcohol 800  $\mu$ l and 5 M NaCl 40  $\mu$ l and then freeze at -20 °C overnight. After being stored overnight, it was centrifuged at 12,000 rpm for 5 min, and then the supernatant was discarded. DNA was washed with 70% ethanol and dried. Finally the DNA was dissolved with elution buffer and then stored at -20 °C.

**PCR condition.** Reaction was carried out in a total volume of 25  $\mu$ l which consisted of 50-100 ng of template DNA, 2.5  $\mu$ l PCR buffer 10x (200 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.1 mM tween 20, 750 mM tris-HCl, pH 8.8), 2.5 mM MgCl<sub>2</sub>, 200  $\mu$ M dNTPs, and 10 pM of each forward and reverse primers and 1 U of taq DNA polymerase. Thermal conditions started with a primary denaturation at 95 °C (5 min) followed by 35 cycles at 95 °C (45 sec), 62 °C (45 sec), and 72 °C (45 sec) and then at 72 °C (5 min) for final extension.

Exon 1C/1D from domain 1 region including the intron of the ovine CAST gene were amplified to a 622 bp fragment by using primers based on Palmer *et al.* (1998). Primer sequences were:

Ovine 1C: 5'-TGGGGCCCAATGACGCCATCGATG-3'  
Ovine 1D: 5'-GGTGGAGCAGCACTTCTGATCACC-3'

PCR products were visualized after electrophoresis on 1.5% agarose gel stained with ethidium bromide.

### RFLP (restriction fragment length polymorphisms).

The amplified fragment of CAST was digested with *MspI* for CAST locus *MspI* and *NcoI* for CAST locus *NcoI*. 5  $\mu$ l of PCR product was digested with 0.7  $\mu$ l buffer, 5 U (0.3) of *MspI* or *NcoI* and 1  $\mu$ l H<sub>2</sub>O up to a total volume of 2  $\mu$ l then were incubated at 37 °C for 12-16 h. The digestion products were electrophoresed on 2% agarose gel in 0.5 x TBE and visualized by ethidium bromide staining for 40 min at 100 V.

### Statistical Analysis

**Frequencies of genotypes and alleles.** The relative frequency of particular allele in a population is called the allele frequency (Nei & Kumar, 2000). Considering a locus with two allele A<sub>1</sub> and A<sub>2</sub>, the allele frequency of A<sub>1</sub> is calculated by

$$X_1 = (2N_{11} + N_{12})/2N$$

Description:

X<sub>1</sub> = allele frequency of A<sub>1</sub>

N<sub>11</sub> = number of sample in genotype A<sub>11</sub>

N<sub>12</sub> = number of sample in genotype A<sub>12</sub>

N = total sample of population

The genotype frequencies can be detected by calculating the ratio of specific genotypes in the population, A<sub>11</sub> genotype frequency can be calculated by following formula (Nei & Kumar, 2000).

$$X_{11} = N_{11}/N$$

Description:

X<sub>11</sub> = genotype frequency of A<sub>11</sub>

N<sub>11</sub> = number of sample in genotype A<sub>11</sub>

N = total sample of population.

**Hardy-Weinberg equilibrium.** Hardy-Weinberg equilibrium was tested by the X<sup>2</sup> statistic (Nei & Kumar, 2000; Noor, 2008).

$$x^2 = \sum \frac{(O - E)^2}{E}$$

Description:

X<sup>2</sup> = Hardy-Weinberg equilibrium test

O = observed number of genotype A<sub>11</sub>

E = expected number of genotype A<sub>11</sub>

### Association of CAST genotypes with body weight.

Association of CAST genotypes locus *MspI* and *NcoI* with body weight were analyzed with the following statistical model (Mattjik & Sumertajaya, 2002).

$$Y_{ij} = \mu + \alpha_i + \epsilon_{ij}$$

Description:

Y<sub>ij</sub> = mean value of body weight

μ = general mean

α<sub>i</sub> = effect of genotypes CAST gen locus *MspI* or *NcoI* (i= MM, MN, and NN)

ε = random error

Data of body weight were corrected by formula below before being used for statistical analysis.

$$X_{i\text{ corrected}} = \frac{\bar{x}_{\text{ standard}}}{\bar{x}_{\text{ observed}}} \times X_{i\text{ observed}}$$

Description:

- X<sub>i</sub> corrected= value of body weight after being corrected by sex and age
- X standard = mean of body weight of standard population
- X observed = mean of body weight of observed population
- X<sub>i</sub> observed= value of body weight before being corrected by sex and age

### RESULTS AND DISCUSSION

CAST gene fragment was successfully amplified by using a thermal cycler machine (Bio AB System) with annealing temperature at 62 °C (Figure 1). The structure of amplified CAST gen was 622 bp including 61 bp of exon 1C, 473 bp in intron 1 and 88 bp in exon 1D (Figure 2) with GenBank accession no AF16006 and AF16007 (Palmer *et al.*, 1998). Annealing temperature used for amplification of this segment was the same with annealing temperature being used by Palmer *et al.* (1998), Nassiry *et al.* (2006), and Shahroudi *et al.* (2006).

#### Genetic Polymorphism of CAST Gen Locus *MspI*

The digestion of 622 bp PCR product for CAST gene with restriction endonucleases *MspI* differentiated

alleles M and N. The *MspI* digestion of the PCR products produced digestion fragments of 336 bp and 286 bp for allele M but the allele N was not digested. The size of fragment for allele N was 622 bp after restriction digestion (Figure 3).

In population of local sheep from JASTRU it was detected three genotypes. The homozygous genotype MM (336 bp, 286 bp) was detected in 197 sheep. The heterozygous genotype MN (622 bp, 336 bp, 286 bp) was detected in 62 sheep. The homozygous genotype NN (622 bp) was detected in 5 sheep.

This result differs with Sumantri *et al.* (2008) work that detected two genotypes only. There had heterozygous genotype MN and homozygous genotype NN. A similar result was also obtained by Shahroudi *et al.* (2006) in Karakul sheep in Iran. It was detected that there were three genotypes MM, MN, and NN. A similar study conducted by Gabor *et al.* (2009) in local sheep in Slovakia and only found two genotypes MM and MN.

#### Genetic Polymorphism of CAST Gen Locus *NcoI*

The digestion of 622 bp PCR product for CAST gene with restriction endonucleases *NcoI* differentiated alleles M and N. The *NcoI* digestion of the PCR products produced digestion fragments of 374 bp and 248 bp for allele N but the allele M was not digested. The size of fragment for allele M was 622 bp after restriction digestion (Figure 4).

In population of local sheep in JASTRU there were detected two genotypes only. The homozygous genotype MM (622 bp) was detected in 242 sheep while the

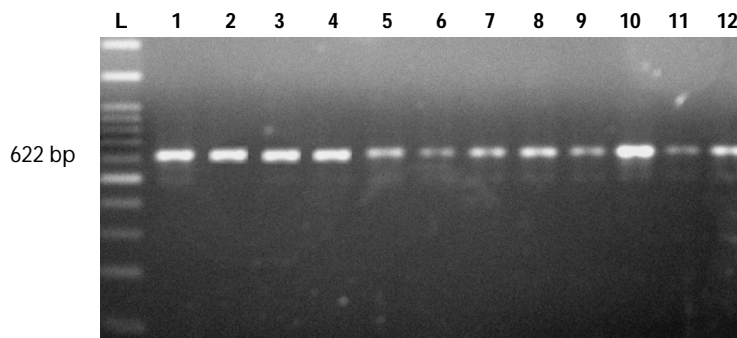


Figure 1. PCR product of CAST gen analyzed by electrophoresis (622 bp). L= ladder 100 bp (Fermentas), and 1-12= PCR product of CAST gen.

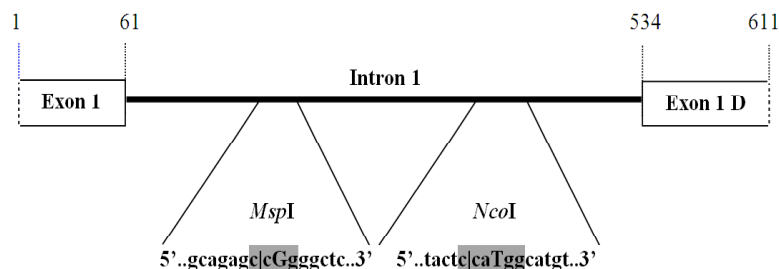


Figure 2. Position of mutation of CAST gene *MspI* and *NcoI* loci. Restriction site of enzymes *MspI* and *NcoI* (dark shading), point of mutations (bold capital letters and dark shading).

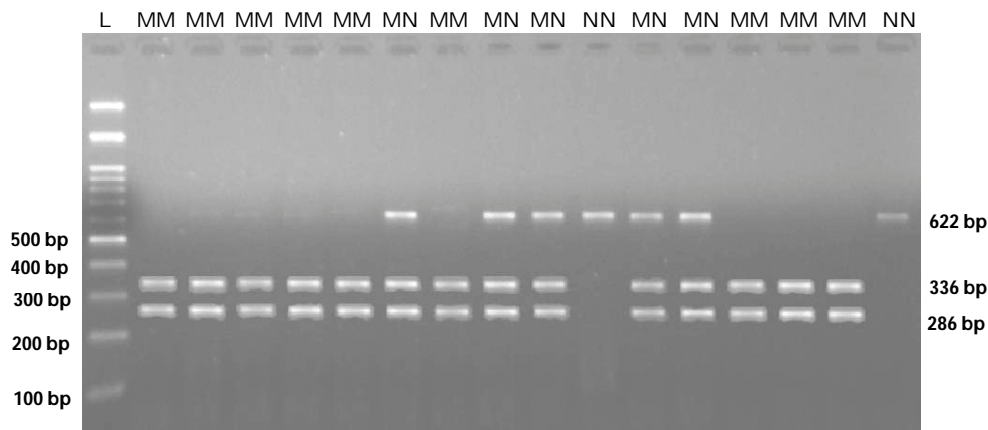


Figure 3. Representatively results of analysis PCR-RFLP for CAST gene by restriction enzyme *MspI* on 2 % agarose gel. L= ladder 100 bp (Fermentas), MM genotype (336 bp, 287 bp), MN genotype (622 bp, 336 bp, 287 bp), NN genotype (622 bp).

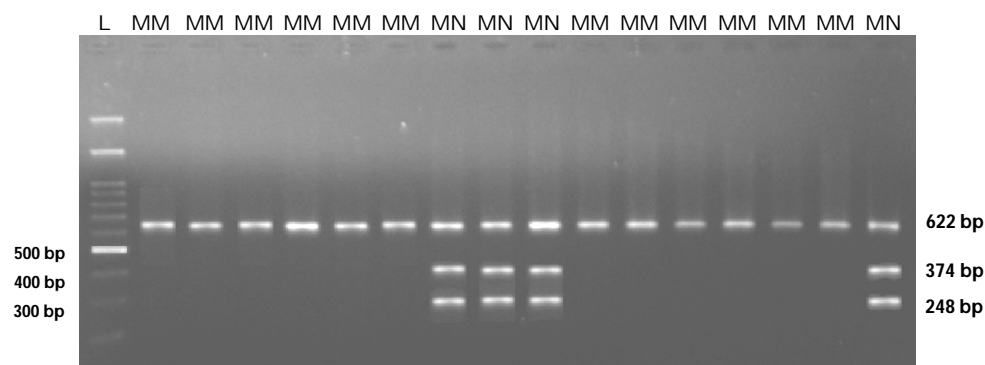


Figure 4. Representatively results of analysis PCR-RFLP for CAST gene by restriction enzyme *NcoI* on 2 % agarose gel. L= ladder 100 bp (Fermentas), MM genotype (622 bp), MN genotype (622 bp, 374 bp, 248 bp).

heterozygous genotype MN (622 bp, 374 bp, 248 bp) was detected in 22 sheep. The homozygous genotype NN (374 bp, 248 bp) was not determined in the population. This result differed with Palmer *et al.* (1998) who finding that there were detected three genotypes MM, MN and NN in Dorset Down, Dorset Down × Coopworth, and Corriedale sheep.

**Frequencies of Genotypes and Alleles**

The frequencies of the CAST genotypes, alleles and  $\chi^2$  test are shown in Table 1. From the total population of local sheep in JASTRU in CAST locus *MspI*, it was

determined that the frequency of homozygous genotype MM was 0.75; heterozygous genotype MN had frequency of 0.23, while homozygous genotype NN had frequency of 0.02. This suggests that a superiority of M allele 0.86 then N allele 0.14 in population 264 local sheep could be kept in JASTRU. In the total population of local sheep in JASTRU of CAST locus *NcoI* was determined that the frequency of homozygous genotype MM had 0.92, while heterozygous genotype MN had frequency of 0.08. This suggests a superiority of M allele 0.96 then N allele 0.04 in population 264 local sheep could be kept in JASTRU.

Polymorphism of CAST gen has also been reported in a variety of other sheep in the world such as the

Table 1. Observed allele and genotypic frequencies and  $\chi^2$  test for CAST locus *MspI* and *NcoI*

CAST	n	Alleles		Genotypes			$\chi^2$
		M	N	MM	MN	NN	
Locus <i>MspI</i>	264	0.86	0.14	0.75	0.23	0.02	0.060 <sup>ns</sup>
Locus <i>NcoI</i>	264	0.96	0.04	0.92	0.08	0.00	0.576 <sup>ns</sup>

n= total of sample;  $\chi^2_{0.05;1} = 3.84$ ; ns= non-significant; MM= homozygous genotype (336 bp, 287 bp); MN= heterozygous genotype (622 bp, 336 bp, 287 bp); NN= homozygous genotype (622 bp).



Table 2. Average of body weight local sheep in The Jonggol Animal Science Teaching and Research Unit (JASTRU), Faculty of Animal Science-Bogor Agricultural University, by CAST genotype locus *Mspl* and *Ncol* (kg)

CAST	N	Genotypes		
		MM	MN	NN
Lokus <i>Mspl</i>	264	15.46±2.65 N= 197	15.03±2.09 N= 62	14.90±3.28 N= 5
Lokus <i>Ncol</i>	264	15.36±2.58 N= 242	15.24±2.0 N= 22	0 N= 0

N = total of sample; ns = non-significant ( $P>0.05$ ); MM= homozygous genotype (336 bp, 287 bp); MN= heterozygous genotype (622 bp, 336 bp, 287 bp); NN= homozygous genotype (622 bp).

Dorset sheep (Palmer *et al.*, 1998), Kurdi sheep in Iran (Nassiry *et al.*, 2006), Karakul sheep in Iran (Shahroudi *et al.*, 2006), Merino, Corriedale, Romney, Poll Dorset, and crossbred NZ sheep in New Zealand (Zhou *et al.*, 2007), and Tsigai sheep, Valachian sheep, East Friesian sheep, Lacaune sheep, and crossbred sheep Lacaune with Tsigai (Gabor *et al.*, 2009).

There was a tendency of local sheep in JASTRU might have higher frequencies M allele than the N allele (Table 1). This result differs from that reported by Sumantri *et al.* (2008) who found that the genotype frequency of MN and NN was 0.32 and 0.68 respectively of local sheep in JASTRU with 22 samples. Some results of other studies also showed similar high frequency of allele M, and low frequency of allele N. Shahroudi *et al.* (2006) reported that the results of gene CAST locus *Mspl* in Karakul sheep frequencies M allele was 0.79 and N was 0.21. Gabor *et al.* (2009) also reported that the results of CAST gene locus *Mspl* in Tsigai sheep, Improved Valachian, Lacaune, East Friesian, and Tsigai sheep × Lacaune sheep had allele frequencies of M 0.94 and N 0.06.

Genetic equilibrium of population was evaluated by using  $\chi^2$ -test. The CAST locus *Mspl* had  $\chi^2$  value of 0.06. For CAST locus *Ncol* the  $\chi^2$  was 0.576. A Large population in Hardy-Weinberg equilibrium is defined, if the frequency of genotype ( $P^2$ ,  $2pq$ ,  $q^2$ ) and frequencies of allele ( $p$  and  $q$ ) are constant from generation to generation, because mating occurs roughly at random in a large population (Vasconcellos *et al.*, 2003; Noor, 2008). Selection is one factor that can alter the balance in the population rapidly (Noor, 2008). Hardy-Weinberg equilibrium can be affected by inbreeding, assortative mating, natural selection and population subdivision (Nei & Kumar, 2000). Current results show that locus *Mspl* and *Ncol* of CAST gene of local sheep in JASTRU were in Hardy-Weinberg equilibrium.

#### Association Analysis CAST Genotypes with Body Weight

The average of body weight of local sheep in JASTRU at each CAST genotype locus *Mspl* and *Ncol* was summarized in Table 2. The genotype effect for body weight of local sheep in JASTRU was not significant. The present data did not show any influences of CAST genotypes on the body weight of local sheep in JASTRU.

Similarly, Nassiry *et al.* (2006) reported it did not show any influence of CAST genotype on sheep at weaning to six months old (GWS), six to nine months (GSN), and nine to yearling (GNY).

The rate and extent of skeletal muscle growth ultimately depends mainly on three factors: rate of muscle protein synthesis, rate of muscle protein degradation, and the number and size of skeletal muscle cells. The calpain activity is required for myoblast fusion and cell proliferation in addition to cell growth (Kurly *et al.*, 2003). The calpain system may also affect the number of skeletal muscle cell (fibres) in domestic animals by altering rate of myoblast proliferation and modulation of myoblast fusion. The calpain system is also important in normal skeletal muscle growth. Increased rate of skeletal muscle growth can result from a decreased rate of muscle protein degradation, and this is associated with a decrease in activity of the calpain system. This could be due principally to a large increase in calpastatin activity (Kurly *et al.*, 2003). body weight traits was controlled by many genes (polygenes) and was mostly influenced by environmental factor (Noor, 2008).

CAST is a specific inhibitor of the calcium-dependent calpain protease family and plays a regulatory role in muscle growth and wastage and in meat tenderization following slaughter (Edyta *et al.*, 2002). This result did not show any influence to body weight of local sheep in JASTRU but may have influence for meat quality. Associations have been reported between variation in CAST and carcass and meat quality traits in cattle (Casas *et al.*, 2006; Schenkel *et al.*, 2006).

Polymorphisms result could be applied for marker in the selection of local sheep with meat quality in future. Currently the consumers preference tend to focus on both quantity and quality traits of sheep meat. This research is a beginning stage to find candidate genes for selection of quantity and quality traits such as double muscle and meat tenderness.

#### CONCLUSION

Gene of CAST locus *Mspl* and *Ncol* were polymorphic in local sheep from JASTRU. The CAST locus *Mspl* had three genotypes MM, MN and NN with frequencies of 0.75, 0.23, and 0.02 respectively. The gene of CAST locus *Ncol* had two genotypes MM and MN with frequencies of 0.92 and 0.08 respectively. The genotypes

of CAST gene locus *MspI* and *NcoI* did not significantly affect on body weight of local sheep in JASTRU.

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