

UNIVERSIDADE ESTADUAL DE CAMPINAS FACULDADE DE ODONTOLOGIA DE PIRACICABA

CIRO DANTAS SOARES

ESTUDO COMPARATIVO DA EXPRESSÃO DE p-AKT1, COX-2 E MARCADORES MITOCONDRIAIS EM MELANOMAS CUTÂNEOS E MUCOSOS

COMPARATIVE STUDY OF THE EXPRESSION OF p-AKT1, COX-2 AND MITOCHONDRIAL MARKERS IN CUTANEOUS AND MUCOSAL MELANOMAS

Piracicaba 2020

CIRO DANTAS SOARES

ESTUDO COMPARATIVO DA EXPRESSÃO DE p-AKT1, COX-2 E MARCADORES MITOCONDRIAIS EM MELANOMAS CUTÂNEOS E MUCOSOS

COMPARATIVE STUDY OF THE EXPRESSION OF p-AKT1, COX-2 AND MITOCHONDRIAL MARKERS IN CUTANEOUS AND MUCOSAL MELANOMAS

Tese apresentada à Faculdade de Odontologia de Piracicaba da Universidade Estadual de Campinas como parte dos requisitos exigidos para a obtenção do título de Doutor em Estomatopatologia, na Área de Patologia.

Thesis presented to the Piracicaba Dental School of the University of Campinas in partial fulfillment of the requirements for the degree of Doctor in Stomatopathology, in Pathology area.

Orientador: Prof. Dr. Jacks Jorge Junior

Este exemplar corresponde a versão final da tese defendida pelo aluno Ciro Dantas Soares e orientada pelo Prof. Jacks Jorge Júnior.

> Piracicaba 2020

Ficha catalográfica Universidade Estadual de Campinas Biblioteca da Faculdade de Odontologia de Piracicaba Marilene Girello - CRB 8/6159

Soares, Ciro Dantas, 1990-

So11e Estudo comparativo da expressão de p-Akt1, COX-2 e marcadores mitocondriais em melanomas cutâneos e mucosos / Ciro Dantas Soares. – Piracicaba, SP : [s.n.], 2020.

Orientador: Jacks Jorge Junior. Tese (doutorado) – Universidade Estadual de Campinas, Faculdade de Odontologia de Piracicaba.

1. Melanoma. 2. Ciclooxigenase 2. 3. Prognóstico. I. Jorge Junior, Jacks, 1962-. II. Universidade Estadual de Campinas. Faculdade de Odontologia de Piracicaba. III. Título.

Informações para Biblioteca Digital

Título em outro idioma: Comparative study of the expression of p-Akt1, COX-2 and mitochondrial markers in cutaneous and mucosal melanomas Palavras-chave em inglês: Melanoma Ciclooxigenase 2 Prognosis Área de concentração: Patologia Titulação: Doutor em Estomatopatologia Banca examinadora: Jacks Jorge Junior [Orientador] Bruno Augusto Benevenuto de Andrade Marcia Martins Margues Roseana de Almeida Freitas Sergio Roberto Peres Line Data de defesa: 17-01-2020 Programa de Pós-Graduação: Estomatopatologia

Identificação e informações acadêmicas do(a) aluno(a)

ORCID do autor: https://orcid.org/0000-0002-6861-6640
 Currículo Lattes do autor: http://lattes.cnpq.br/7961127069977379



UNIVERSIDADE ESTADUAL DE CAMPINAS Faculdade de Odontologia de Piracicaba

A Comissão Julgadora dos trabalhos de Defesa de Tese de Doutorado, em sessão pública realizada em 17 de Janeiro de 2020, considerou o candidato CIRO DANTAS SOARES aprovado.

PROF. DR. JACKS JORGE JUNIOR

PROF^a. DR^a. ROSEANA DE ALMEIDA FREITAS

PROF^a. DR^a. MARCIA MARTINS MARQUES

PROF. DR. BRUNO AUGUSTO BENEVENUTO DE ANDRADE

PROF. DR. SERGIO ROBERTO PERES LINE

A Ata da defesa, assinada pelos membros da Comissão Examinadora, consta no SIGA/Sistema de Fluxo de Dissertação/Tese e na Secretaria do Programa da Unidade.

AGRADECIMENTOS

À Universidade Estadual de Campinas, na pessoa do Magnífico Reitor, Prof. Dr. Marcelo Knobel.

À Faculdade de Odontologia de Piracicaba, na pessoa de seu Diretor, Prof. Dr. Francisco Haiter Neto e seu Diretor Associado, Prof. Dr. Flávio Henrique Baggio Aguiar.

À Profa. Dra. Cínthia Pereira Machado Tabchoury, Coordenadora Geral da Pós-Graduação da Faculdade de Odontologia de Piracicaba.

Ao Coordenador do Programa de Pós-Graduação em Estomatopatologia, Prof. Dr. Márcio Ajudarte Lopes.

O presente trabalho foi realizado com apoio da Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP), processos 2015/25905-1, 2017/16102-8, e 2017/18134-4, agradecemos todo apoio financeiro da instituição citada.

AGRADECIMENTOS ESPECIAIS

Em primeiro lugar gostaria de agradecer à Deus, que me concedeu forças para trilhar o caminho da pós-graduação e por sempre ser meu apoio nos momentos difíceis e minha inspiração nos momentos felizes.

Agradeço, particularmente ao meu orientador, Prof. Dr. Jacks Jorge Junior, um grande ser humano. Em especial, gostaria de citar que a sua compreensão, altruísmo e responsabilidade fizeram com que o percurso do curso de doutorado fosse mais tranquilo. Sempre serei grato por me ter oportunizado, ajudado nos momentos mais difíceis e inclusive ser um conselheiro de vida.

Aproveito também para agradecer a todos os professores do programa de pós-graduação em Estomatopatologia que de forma direta ou indireta contribuíram para a minha formação: Profs. Alan Roger, Edgard Graner, Márcio Lopes, Oslei Almeida, Pablo Vargas e Ricardo Coletta.

De maneira muito especial, gostaria de agradecer ao Prof. Oslei Paes de Almeida que me acolheu como um verdadeiro filho, e nossos laços se estenderam para além da academia. O Prof. Oslei me ajudou a criar um pensamento crítico, além de ter concedido várias oportunidades que seriam difíceis de listá-las aqui. Cito de maneira especial por ter me oportunizado estar com tantos seres humanos incríveis que acrescentaram tanto à minha formação profissional e pessoal: Dr. Román Carlos, Dr. Adalberto Mosqueda-Taylor, Dr. Juan Carlos Hernandez-Guerrero, Dr. José Manuel Aguirre-Urizar, Dr. Wilson Delgado Azañero.

Ao Dr. Román Carlos, toda minha gratidão por ter sido uma pessoa tão acolhedora na minha estadia na Guatemala e por ter contribuído com a grande maioria dos casos dessa tese. Agradeço também de maneira especial aos Drs. Adalberto Mosqueda e Juan Carlos Hernandez Guerrero, por serem tão acolhedores em todas as vezes que estive no México e por contribuírem de forma tão essencial para minha formação pessoal e profissional.

Faltariam palavras para agradecer todo apoio, renúncias e todo o amor que recebi de minha família (apesar de citar alguns nomes, sou e serei grato a todos meus tios, primos, avós). Meus pais Cícero e Josélia que sempre estiveram ao meu lado e me ensinaram que a vida não é construída com coisas e valores materiais, mas com muito amor e esforço. De verdade, com vocês aprendi o verdadeiro valor de construir uma família, e isso hoje é um dos meus maiores sonhos, ser pai. Sempre me senti amparado

por pais honestos, acima de tudo, sinceros com os valores que pregam. Vocês são inspiração e sustento para minha vida, sempre serei grato ao amor de vocês. Madja e Cicinho (como carinhosamente chamamos a Cícero Júnior), obrigado por aceitar o desafio de suportar-me e dividir comigo os melhores momentos da minha vida. Minha eterna gratidão por me ensinarem o verdadeiro significado da palavra "irmãos", para sempre meus "pequenos". Vocês, junto com minha pequenina Maryanne, são peças fundamentais em minha vida e sempre serão as pessoas que me farão ir além. Eu amo vocês. Gostaria de agradecer a minha namorada Jocelyn. Você se tornou uma peça fundamental para entender que o verdadeiro amor não precisa de explicações, apenas necessita ser vivido. Sempre serei grato ao universo por ter conhecido uma pessoa tão sincera, espontânea e acima de tudo, com um amor próprio invejável. Obrigado por decidir estar ao meu lado, obrigado por não desistir de mim.

Aos meus amigos. Eu, verdadeiramente, poderia não citar nomes apenas por assumir o risco de esquecer alguém, no entanto, esquecer de citar um nome tampouco seria uma causa de não amar alguém de maneira sincera. Assim, Rafael Sales, Carla Rodrigues, Jeny Kesia, João Paulo Queiroz, Alex Paiva, Léo Pereira, Júlio Alves, Rodrigo Macedo, Alan e Alandson Lacerda, vocês foram e são meu "porto seguro", em quem posso me apoiar quando me faltam as forças. Obrigado por tudo.

Aos professores de hoje e de sempre. Em especial, gostaria de dedicar todo meu carinho as professoras Rejane Carvalho e Goretti Carvalho, que de forma sempre muito especial me educaram e me ajudaram a trilhar os caminhos da verdadeira ciência. Vocês são um exemplo a ser seguido, são pessoas que devem ser sempre ressaltadas por suas características de pesquisadoras de altíssimo nível, além de serem humanas que adjetivo em nenhuma língua seria capaz de descrevê-las! Obrigado por serem as pessoas mais importantes na minha formação. Agradeço de maneira muito especial as professoras Albina Altemani e Maria Letícia Cintra, que sem dúvidas foram peças fundamentais em minha formação acadêmica. Agradeço também aos doutores e doutoras: Fernanda Mariano, Marcelo Brum Corrêa, Rodrigo Reis e Luciana Schultz Amorim pela valiosa contribuição com a casuística e dados clínicos desse estudo.

Por último, mas não menos importante, gostaria de agradecer aos meus amigos que compartilharam comigo a desgastante jornada da pós-graduação. Aos que já foram e aos que ficam, muita força para concluir esse processo, que é muitas vezes difícil, mas é muito gratificante.

RESUMO

Melanomas são neoplasias malignas agressivas que ocorrem principalmente na pele. Os melanomas mucosos de cabeça e pescoço, por outro lado, que ocorrem nas mucosas da cavidade bucal e nasal, são neoplasias malignas extremamente agressivas; e devido sua raridade, estudos mais completos sobre possíveis marcadores prognósticos ainda são escassos na literatura. O presente trabalho foi organizado em 5 capítulos que dissertam sobre a expressão de proteína B quinase fosforilada (p-Akt1), ciclooxigenase 2 (COX-2) e marcadores mitocondriais em uma série de melanomas cutâneos e mucosos de cabeca e pescoço. Como principais resultados, podemos destacar, as conclusões de cada capítulo conforme organizado abaixo. No primeiro capítulo, demonstramos que a proteína p-Akt1 teve maior expressão em melanomas mucosos que nos cutâneos e foi identificada como um fator prognóstico independente em melanomas mucosos. No segundo capítulo, foi estudada a expressão de COX-2 em melanomas orais, demonstrando que essa enzima é um fator prognóstico independente para pacientes com estes tumores. No terceiro capítulo, apresentamos uma série de melanomas amelanóticos e foi possível concluir que a apresentação clínica e microscópica desses tumores pode mimetizar outros tipos de lesões, e que, portanto, podem ser mais difíceis de ser diagnosticados. No quarto capítulo, foi comparada a expressão de COX-2 e de Ki67 em melanomas orais amelanóticos e melanomas orais convencionais. Como principal conclusão, a expressão de COX-2 e o índice de proliferação foram maiores em melanomas orais amelanóticos, indicando provavelmente uma maior agressividade destes tumores. No quinto capítulo, foi avaliada a expressão de três marcadores mitocondriais em melanomas cutâneos e mucosos. Como destaque desse último capítulo podemos citar que a quantidade de mitocôndrias nas células tumorais parece desempenhar um papel importante na patogênese dos melanomas. E que, a proteína de fissão 1 (FIS1) pode ser considerada um fator prognóstico em melanomas orais, enquanto a proteína de fusão 2 (mitofusin 2) está associada com pior prognóstico em pacientes com melanomas cutâneos. Em resumo, os cinco capítulos da presente tese contribuíram para o melhor entendimento dos melanomas mucosos e em conjunto apresentam uma série de possíveis marcadores prognósticos que podem ser estudados e indicados como candidatos para terapias-alvo.

Palavras-chave: Melanomas cutâneos, melanomas mucosos de cabeça e pescoço, p-Akt1, COX-2, marcadores mitocondriais, prognóstico.

ABSTRACT

Melanomas are tumors with an aggressive behavior that occur mainly in the skin. Mucosal head and neck melanomas, on the other hand occur in the mucous membranes of the oral and nasal cavity. These tumors are extremely aggressive; and due to its rarity, studies with these tumors regarding possible prognostic markers are still scarce in the literature. The present PhD thesis was organized in 5 chapters that discuss the phosphorylated serine/threonine-protein kinase expression of 1 (p-Akt1). cyclooxygenase 2 (COX-2) and mitochondrial markers in a series of cutaneous and mucous melanomas of the head and neck. We have highlighted the main conclusions of each chapter, as organized below. In the first chapter, we demonstrated that p-Akt1 protein was overexpressed in mucosal than in cutaneous melanomas and it was identified as an independent prognostic factor for mucosal melanomas. In the second chapter, we have studied the COX-2 expression in oral melanomas, demonstrating that this enzyme is an independent prognostic factor for patients with these tumors. In the third chapter, we present a series of amelanotic melanomas and the main conclusion was: the clinical and microscopic presentation of these tumors may mimic other types of lesions, and therefore they may be more difficult to diagnose. In the fourth chapter, the expression of COX-2 and Ki67 in amelanotic and conventional oral melanomas was compared. As a main conclusion, COX-2 expression was higher in amelanotic oral melanomas, as well as cell proliferation index in these tumors than in conventional oral melanomas. In the fifth chapter, the expression of three mitochondrial markers in cutaneous and mucosal melanomas was evaluated. We have demonstrated that mucosal melanomas have a higher expression of mitochondrial markers than cutaneous ones. Thus, the high number of mitochondria in tumor cells seems to play an important role in the pathogenesis of melanomas. The fission protein 1 (FIS1) was considered a prognostic factor in oral melanomas, whereas mitofusin 2 is associated with worse prognosis in patients with cutaneous melanomas. In summary, the five chapters of this thesis have contributed to a better understanding of mucosal melanomas and together present a series of possible prognostic markers that can be studied and indicated as candidates for target therapies.

Keywords: Cutaneous melanomas, head and neck mucosal melanomas, p-Akt1, COX-2, mitochondrial markers, prognosis.

SUMÁRIO

1 INTRODUÇÃO

Melanomas são neoplasias malignas extremamente agressivas que ocorrem principalmente na pele (Leonardi *et al.*, 2018). Sua etiologia está intrinsicamente ligada a alterações genéticas em células melanocíticas de pacientes com histórico de larga exposição à radiação solar (Curtin *et al.*, 2005). O prognóstico de pacientes com melanoma cutâneo melhorou significativamente com o advento da imunoterapia, um novo tipo de tratamento baseado na reprogramação do sistema imunológico para combater células malignas (Lugowska, Teterycz and Rutkowski, 2017). Ainda assim, alguns casos mais avançados – com metástase à distância, por exemplo – possuem prognóstico ruim (Li *et al.*, 2007).

Os melanomas mucosos de cabeça e pescoço, por outro lado, que ocorrem nas mucosas da cavidade bucal e nasal, são neoplasias malignas extremamente agressivas; e devido sua raridade, estudos mais completos sobre possíveis marcadores prognósticos ainda são escassos na literatura (de Andrade *et al.*, 2012; de-Andrade *et al.*, 2012; Bishop and Olszewski, 2014; Lourenço *et al.*, 2014; Mochel *et al.*, 2015; de Souza do Nascimento *et al.*, 2016). Os fatores etiológicos dos melanomas mucosos de cabeça e pescoço ainda não são conhecidos, o que também dificulta o entendimento da patogenia desses tumores agressivos.

As diferenças biológicas entre melanomas mucosos e cutâneos são objetivo de debate há muito tempo (Luna-Ortiz *et al.*, 2016). No entanto, poucos estudos conseguiram determinar que realmente há uma diferença na biologia tumoral dessas lesões. O pior prognóstico das lesões mucosas quase sempre é associado unicamente ao diagnóstico tardio, que obviamente representa um papel importante, mas talvez não seja o único fator associado com péssimo prognóstico para estes pacientes (Kuk *et al.*, 2016; Moya-Plana *et al.*, 2019).

A biologia tumoral é muito complexa, e envolve vários passos que culminam com a progressão tumoral e o processo metastático. Algumas proteínas desempenham papéis importantes no contexto de migração, invasão e proliferação celular. Dentre elas, a proteína B kinase (também denominada Akt), que é uma serina-treonina com três isoformas bem conhecidas: Akt1, Akt2 e Akt3 (Franks *et al.*, 2016). A isoforma Akt1 é amplamente conhecida por sua atividade no metabolismo celular, ativando vias glicolíticas, apoptóticas e inclusive modulando a atividade de proliferação celular. Esses efeitos têm sido demonstrados em diversos tipos de tumores (Cicenas, 2008; Liu *et al.*, 2010; Agarwal, Brattain and Chowdhury, 2013; Wu *et al.*, 2014). Estudo recente

também demonstrou que a Akt1 desempenha um papel muito relevante no desenvolvimento de metástases de melanoma em um modelo de camundongos (Cho *et al.*, 2015). Em estudo de sequenciamento genético do nosso grupo (dados ainda não publicados), foi possível observar uma diferença na expressão de Akt1 entre melanomas cutâneos e mucosos de cabeça e pescoço.

A ciclooxigenase 2 (COX-2) é uma enzima pró-inflamatória envolvida diretamente no metabolismo do ácido araquidônico (Fitzpatrick, 2005). As consequências da expressão e da atividade de COX-2 nas células tumorais é bem conhecida: ativa vias de angiogênese, facilitando o processo de disseminação tumoral e consequentemente metástases à distância; modulação do processo apoptótico; evasão do sistema imune, e muitas vezes também associado com resistência quimioterápica (Guo *et al.*, 2015; Xia *et al.*, 2015; Hu *et al.*, 2017; Hashemi Goradel *et al.*, 2019). A expressão de COX-2 já foi estudada em melanomas de cavidade oral, e interessantemente, a ausência de COX-2 em nevos orais e cutâneos indica que essa enzima poderia desempenhar um papel central na biologia tumoral dos melanomas (de Souza do Nascimento *et al.*, 2016).

Mitocôndrias são organelas responsáveis pela produção de energia no ambiente intracelular e desempenham um papel crucial durante praticamente todos os processos biológicos. No contexto das células tumorais, vários processos biológicos requerem o ATP como fonte de energia através da fosforilação oxidativa (Corazao-Rozas *et al.*, 2016). Como consequência da fosforilação oxidativa, as mitocôndrias também produzem a maioria das espécies reativas de oxigênio (ERO). As EROs estão implicadas no processo carcinogênico, danificando importantes proteínas e macromoléculas, causando desregulação do ciclo celular e outros processos relevantes para o desenvolvimento e metástase do tumor (Liou and Storz, 2010). Além disso, as mitocôndrias desempenham um papel essencial na apoptose, regulando as vias críticas de sinalização para sua ativação ou inibição (Suen, Norris and Youle, 2008; Srinivasan *et al.*, 2017). Esses eventos ainda não foram estudados em melanomas.

De acordo com o que foi exposto, há uma clara escassez de estudos comparativos com melanomas cutâneos e mucosos de cabeça e pescoço, particularmente, no que se diz respeito a marcadores prognósticos. Na era de terapiasalvo, é sumamente importante entender a biologia tumoral, e assim, aplicar esses conceitos no desenvolvimento de terapias complementares que possam melhorar o prognóstico dos pacientes. Nesse sentido, o objetivo desse estudo foi, avaliar e comparar a expressão de p- Akt1, COX-2 e marcadores mitocondriais em uma larga série de melanomas cutâneos e mucosos de cabeça e pescoço, bem como validar possíveis marcadores com utilidade prognóstica nestes tumores.

2 ARTIGOS

2.1 Artigo: Phosphorylated Akt1 expression is associated with poor prognosis in cutaneous, oral and sinonasal melanomas

Artigo publicado no periódico Oncotarget DOI: 10.18632/oncotarget.26458 (Anexo 1)

Ciro Soares¹, Thayná Melo de Lima Morais¹, Roman Carlos², Fernanda Viviane Mariano^{1,3}, Albina Altemani^{1,3}, Maria Goretti Freire de Carvalho⁴, Marcelo Brum Corrêa⁵, Rodrigo Ribas Dias dos Reis⁶, Luciana Schultz Amorim⁷, Oslei Paes de Almeida¹, and Jacks Jorge¹

¹Department of Oral Diagnosis, Area of Pathology, Piracicaba Dental School, University of Campinas, Piracicaba, São Paulo, Brazil
²Pathology Division, Centro Clínico de Cabeza y Cuello/Hospital Herrera Llerandi, Guatemala City, Guatemala
³Department of Pathology, Faculty of Medical Sciences, University of Campinas, Campinas, São Paulo, Brazil
⁴Private Pathology Service, Natal, Rio Grande do Norte, Brazil
⁵Head and Neck Surgery Department - Oncology Center (CEON), Fornecedores de Cana Hospital, Piracicaba, São Paulo, Brazil
⁶Oncology Surgery Department - Cancer Center (CECAN), Santa Casa Hospital, Piracicaba, São Paulo, Brazil

Corresponding author:

Ciro Dantas Soares Department of Oral Diagnosis. Piracicaba Dental School, University of Campinas Address: Avenida Limeira, 901, Areião - 13414-903 Piracicaba/SP, Brazil, Telefone: (84) 99601-1270 E-mail: ciro.dss@gmail.com

Abstract

Melanomas are highly aggressive tumours derived from melanocytes, which occur most commonly in the skin. Occasionally, these tumours may appear in oral and sinonasal mucous membranes. In this study, we performed a comparative analysis of the Phosphorylated Akt1 (p-Akt1) expression in 144 patients affected by cutaneous (CM), 34 oral cavity (OM), and 31 sinonasal melanomas (SNM). Similar to the metastatic cutaneous melanomas, p-Akt1 was overexpressed in 17/34 of the oral cavity and 20/31 of the sinonasal melanomas. In addition, the p-Akt1-nuclear expression was associated with poorer cancer-specific survival in cutaneous (P < .0001), oral (P < .0001), and sinonasal (P = .001) melanomas. Multivariate analysis showed p-Akt1 to be an independent prognostic marker in oral (P = .041) and sinonasal (P < .0001) melanomas patients. In conclusion, p-Akt1 overexpression is an independent prognostic marker in mucosal melanomas and is significantly up-regulated in sinonasal melanomas. As both mucosal and metastatic cutaneous melanomas showed high frequency of p-Akt1 expression, these findings suggest that mucosal melanomas have a biological behaviour, similar to the aggressive cutaneous melanomas.

Keywords: Mucosal melanomas, cutaneous melanomas, prognostic markers, p-Akt1

Introduction

Cutaneous melanomas represent about 1.6% of all cancers, and discreet advances in their treatment have been made over the last decades. Nevertheless, this tumour still remains deathly, especially the metastatic disease [1]. It is estimated that melanomas with lymph nodal metastases are responsible for 59,782 global deaths, with mean overall survival rate of 13% in 5 years [2]. Controlling the advanced-stage disease is the major problem for the treatment of melanomas. Thus, for the development of targeted therapies, making progress in understanding the molecular factors that influence the aggressiveness of the metastatic melanomas is essential.

Until now, comparative studies with melanomas from different anatomic sites that assessed the potential differences and similarities between these tumours are scarce [3–6]. Overall, the etiologic factors and the clinical and biological behaviour of cutaneous melanomas are very distinct from the other melanomas [7, 8]. For example, mucosal melanomas constitute a particular subset of all melanomas characterized by high-aggressive behaviour, tendency to metastasize and consequently association with marked worse prognosis. Sinonasal melanomas in some series have presented a high-mortality index (near 100%) [8, 9]. Clinicopathological parameters determine the prognosis and staging of the melanomas from different anatomic sites [10].

The use of biological markers, especially by immunohistochemistry, for predict prognosis is still poorly explored in mucosal melanomas. The serine/threonine protein B Kinase, known as Akt, has an oncogenic function in several tissues by regulating cell proliferation, migration and invasion [11]. The overexpression of p-Akt1 is correlated with adverse outcome in breast [12], gastric [13] and oesophageal squamous cell carcinomas [14]. In the context of melanoma, an overgrowing interest in determining as p-Akt1 acts during melanoma progression [15, 16] has been recently observed, and its inhibition has emerged as an interesting targeted therapy for these tumours [17].

Although some studies have demonstrated the role of p-Akt1 in several cancers, including melanoma metastases in a mice model [18], no studies have evaluated the prognostic value of the immunohistochemical expression of p-Akt1 in different subsets of melanomas. In the present study, we evaluated the immunohistochemical expression and the prognostic value of p-Akt1 expression in a large cohort of cutaneous, oral, and sinonasal melanomas.

Material and Methods

Ethical issues

The study protocol was approved by the National Commission for Ethics in Research (CONEP-Brazil, CAAE: 72077517.1.0000.5418) and all procedures were in accordance with the Declaration of Helsinki.

Patient samples and data collection

Cutaneous, sinonasal, and oral samples from the surgical samples of patients with melanoma who were referred to four Brazilian and Guatemalan cancer centers since 1997 were collected and further analysed. Melanoma diagnoses were confirmed by three experienced pathologists based on the microscopic examination of haematoxylin–eosin–stained slides, and on the S-100 protein, MART-1/Melan A and gp-100 (HMB-45) expressions.

In total, 209 samples, including cutaneous (n = 144), oral (n = 34) and sinonasal (n = 31) melanomas. The first patient was included in May 1997 and the last patient in February 2016. Surviving patient follow-up was censored on September 10, 2017. Clinical data were collected from the patient's medical records which included age, sex, tumour location, metastases (presence/absence and site), clinical-stage, treatment options, and follow-up data. Tumour stage was determined according to the seventh edition of the staging manual of the American Joint Committee on Cancer [25].

Tissue microarray construction

For construction of the 7 tissue microarray blocks, a previously reported method was followed [41, 54]. Duplicate cores of 2 mm² were collected from the original blocks of cutaneous, oral, and sinonasal melanoma patients.

Immunohistochemistry (IHC) and digital scoring

IHC manual technique was performed using 3-mm thick formalin-fixed paraffinembedded melanomas sections mounted on silane-coated glass slides. An anti-Phospho-Akt1 (clone D7F10, 1:100 dilution; Cell Signalling, Danvers, Massachusetts, USA) was used. The antigen detection was achieved using the ADVANCE[™]/HRP (code K406889-2; Dako, Carpinteria, CA, USA), revealed with the 3,3'-diaminobenzidinetetrahydrochloride chromogen and counterstained with Carazzi's haematoxylin.

All slides were scanned into high-resolution images with Aperio Scanscope CS Slide Scanner (Aperio Technologies Inc., Vista, California, USA). In order to calculate the scores of positivity expression of p-Akt1, digital analyses were performed, as previously described [41, 55]. Nuclear p-Akt1 expression was scored based on the percentage [%] of positive nuclei, assessed digitally with the Nuclear Algorithm (Aperio Technologies Inc.).

Statistical analyses

For the statistical analyses, all collected data were recorded in a passwordprotected computer database. All statistical analyses were carried out using the SPSS software, version 18.0 (SPSS Inc., Chicago, IL, USA). In the univariate model, the cancer-specific survival (CSS) (defined as the time between the start date of the treatment and the date of death due the disease) and disease-free survival (DFS) (defined as the time between treatment and recurrence) were estimated comparing the Kaplan–Meier survival curves using the log-rank test. The multivariate Cox regression was performed for prognostic significance determination of the p-Akt1 expression Index. For both univariate and multivariate models, the association between the variables age, sex, primary tumour site and p-Akt1 expression and CSS and DFS were evaluated. p-value $\leq .05$ was considered statistically significant.

Results

Cutaneous melanomas

The age range of the cohort of 144 patients affected by cutaneous melanomas was from 20 to 88 years old at diagnosis (mean age, 56). The three most common sites were the trunk (64/144), followed by the lower limbs (40/144) and the head and neck area (22/144) (Table 1). The median follow-up period was 62 months, ranging from 14 months to 18 years. The 3- and 5-year cancer-specific survival (CSS) rates were 89.9% and 69.3%, respectively. Detailed clinicopathological data from the cutaneous melanomas is shown in Supplementary Table 1.

 Table 1: The relationship between clinicopathological characteristics of patients

 with cutaneous melanomas, p-Akt1 expression and cancer-specific survival

| Factors | Sample | CSS | (%) | Univariate | Multivariate | |
|------------|--------|---------|---------|--------------|--------------------|-----------------|
| Factors | n (%) | 3-years | 5-years | P (log-rank) | HR (95% CI) | <i>P</i> -value |
| Age | | | | | | |
| <56 | 78 | 94.8% | 87.6% | .013 (6.118) | 2.029 (.969–4.250) | .061 |
| ≥56 | 66 | 82.5% | 70.4% | | | |
| Ulceration | | | | | | |

| Factors | Sample | CSS | (%) | Univariate | Multivariate | |
|---------------------|--------|---------|---------|-----------------|------------------------|-----------------|
| Factors | n (%) | 3-years | 5-years | P (log-rank) | HR (95% CI) | <i>P</i> -value |
| Present | 61 | 76.5% | 60.3% | <.0001 (39.745) | 1.569 (.499–4.933) | .441 |
| Absent | 83 | 98.8% | 94.4% | | | |
| Breslow's Thickness | | | | | | |
| <1.55 | 72 | 100% | 98.4% | <.0001 (42.747) | 10.855 (1.101–107.057) | .041 |
| ≥1.55 | 72 | 78.7% | 61.7% | | | |
| Mitotic rate | | | | | | |
| <3 | 60 | 98.3% | 94.2% | <.0001 (17.848) | 1.311 (.373–4.606) | .673 |
| ≥3 | 84 | 82.9% | 70.1% | | | |
| Clark's level | | | | | | |
| I and II | 47 | 100% | 100% | .002 (9.706) | .667 (.216–2.063) | .482 |
| III, IV and V | 97 | 84.7% | 71.4% | | | |
| Distant metastasis | | | | | | |
| Present | 34 | 67.5% | 41.2% | <.0001 (62.087) | 2.277 (.512–10.135) | .280 |
| Absent | 110 | 96.3% | 92.8% | | | |
| AJCC-stage | | | | | | |
| In situ | 15 | 100% | 100% | <.0001 (71.219) | 1.438 (.650–3.181) | .369 |
| Ι | 51 | 100% | 100% | | | |
| П | 42 | 90.4% | 80.1% | | | |
| III | 23 | 69.3% | 55.4% | | | |
| IV | 13 | 69.2% | 27.7% | | | |
| p-Akt1 | | | | | | |
| High expression | 69 | 80.8% | 64.2% | <.0001 (24.206) | 2.766 (.952-8.038) | .062 |
| Low expression | 75 | 98.6% | 94.1% | | | |

CSS – Cancer–specific survival, HR – Hazard Ratio, AJCC – American Joint Committee on Cancer. Bold values indicate statistical significance (P < .05).

Cancer-specific survival (CSS) rates for cutaneous melanomas based on clinicopathological parameters are shown in Table 1. Significant prognostic factors for

CSS using a univariate model were Age (P = .013), Ulceration (P < .0001), Breslow's Thickness (P < .0001), Mitotic rate (P < .0001), Clark's level (P = .002), Distant metastasis (P < .0001), AJCC-stage (P < .0001) and p-Akt1 expression (P < .0001) Figure 1; while the unique independent prognostic factor in a multivariate analysis was Breslow Thickness (HR = 10.855, P = .041).



Figure 1: p-Akt1 expression in cutaneous melanomas. (A) Representative example of non-metastatic cutaneous melanoma in a tissue microarray, which was focally positive for p-Akt1. (B) Representative example of metastatic cutaneous melanoma in a tissue microarray, which was strongly positive for p-Akt1. (C) The association of p-Akt1nuclear expression and cancer-specific survival in cutaneous melanomas (logrank = 24.206, P < .0001). (D) The association of metastasis and cancer-specific survival in cutaneous melanomas (logrank = 62.087, P < .0001). The scale bars represent 1 mm (top) and 100 µm (bottom).

p-Akt1 is an independent prognostic marker in Oral melanomas

For oral melanomas, 34 patients were enrolled in this study, with a mean age of 52 years, and regarding sex, 18 were men and 16 women. The main locations for oral melanomas were the hard palate (15/34), the tooth alveoluls (5/34), and other areas (14/34). The OM clinicopathological data are listed in Supplementary Table 2. The

median follow-up period was 23.5 months, ranging from 2 months to 14 years. The 3and 5-year CSS rates were 49.1% and 35.8%, respectively.

Using the multivariate analysis, the mitotic rate (P = .001), presence of vascular invasion (P < .0001), neural invasion (P = .011), epithelioid cellular morphology (P = .001), and p-Akt1 nuclear expression were considered prognostic factors for CSS in oral melanomas (P < 0.001, Log-rank = 28.086; Figure 2). More interestingly, in the Cox regression model against established pathological prognostic factors such as mitotic rate, cellular morphology, vascular and neural invasion, p-Akt1 expression demonstrated to be an independent prognostic factor for oral melanomas Table 2 (HR = 11.397, P = .041).



Figure 2: p-Akt1 expression in oral melanomas. (A) Representative example of oral melanoma in a tissue microarray, which was focally positive for p-Akt1. (B) Representative example of oral melanoma in a tissue microarray, which was strongly positive for p-Akt1. (C) The association of p-Akt1-nuclear expression and cancerspecific survival in oral melanomas (log-rank = 28.086, P < .0001). (D) The association of AJCC-stage and cancer-specific survival in oral melanomas (log-rank = 1.68, P = .682). The scale bars represent 1 mm (top) and 100 µm (bottom).

| Factors | Sample | CSS (%) | | Univariate | nivariate Multivariate | | |
|------------------------------|-----------|---------|--------|---------------|------------------------|-----------------|--|
| ractors | (%) | 3-year | 5-year | P (log-rank) | HR (95% CI) | <i>P</i> -value | |
| Age | | | | | | | |
| <47 | 18 (53.9) | 53.8% | 38.5% | .926 (.009) | 1.407 (.237–8.361) | .707 | |
| ≥47 | 16 (46.1) | 41.5% | 41.5% | | | | |
| Gender | | | | | | | |
| Female | 16 (46.1) | 38.4% | 38.4% | .374 (.791) | .718 (.196–2.628) | .617 | |
| Male | 18 (53.9) | 58.2% | 38.8% | | | | |
| Treatment | | | | | | | |
| Only surgery | 22 (64.7) | 55.4% | 55.4% | .070 (3.273) | .689 (.129–3.674) | .663 | |
| Surgery + chemo/radiotherapy | 11 (32.4) | 38.1% | 12.7% | | | | |
| Anatomical site | | | | | | | |
| Palate | 15 (44.1) | 34.2% | 34.2% | .113 (4.355) | _ | _ | |
| Others | 19 (55.9) | 68.8% | 55% | | | | |
| Mitotic rate | | | | | | | |
| <1 | 14 (41.2) | 85.1% | 85.1% | .001 (10.116) | 1.372 (.154–12.212) | .777 | |
| ≥1 | 20 (58.8) | 26.9% | 13.5% | | | | |
| Vascular invasion | | | | | | | |

Table 2: The relationship between clinicopathological characteristics of patientswith oral melanomas, p-Akt1 expression and cancer-specific survival

| Factors | Sample | CSS (%) | | Univariate | Multivariate | | |
|---------------------|-----------|---------|--------|-----------------|------------------------|-----------------|--|
| Factors | (%) | 3-year | 5-year | P (log-rank) | HR (95% CI) | <i>P</i> -value | |
| Absent | 21 (61.8) | 80.4% | 58.6% | <.0001 (18.408) | 1.485 (.191–11.579) | .706 | |
| Present | 13 (38.2) | 0% | 0% | | | | |
| Neural invasion | | | | | | | |
| Absent | 26 (76.5) | 42.9% | 42.9% | .011 (6.467) | 4.622 (.640–33.373) | .129 | |
| Present | 8 (23.5) | 15.6% | 15.6% | | | | |
| Cellular morphology | | | | | | | |
| Epithelioid | 22 (64.7) | 30.3% | 20.2% | .001 (12.052) | .927 (.085–10.121) | .951 | |
| Non-epithelioid | 12 (35.3) | 90% | 90% | | | | |
| AJCC-stage | | | | | | | |
| III | 9 (26.5) | 55.6% | 37% | .682 (.168) | .885 (.211–3.706) | .867 | |
| IVa, b and c | 25(73.5) | 46% | 36.8% | | | | |
| p-Akt1 | | | | | | | |
| High expression | 17 (50) | 11.8% | 5.9% | <.0001 (28.086) | 11.397 (1.102–117.855) | .041 | |
| Low expression | 17 (50) | 93.3% | 93.3% | | | | |

CSS – Cancer–specific survival, HR – Hazard Ratio, AJCC – American Joint Committee on Cancer. Bold values indicate statistical significance (P < .05)

p-Akt1 is an independent prognostic marker in sinonasal melanomas

Regarding the sinonasal melanomas (SNM), out of 31 patients, 15 were women and 16 men of ages between 24 and 82 (mean age, 55). The tumours were located at nasal cavity (41.9%), maxillary sinus (38.7%), and rhinopharynx (19.4%). Detailed clinicopathological data for SNM are shown in Supplementary Table 3. The median follow-up for SNM patients was 30 months, ranging from 3 months to 18.6 years, and the CSS 3-years and 5-years survival rates were 56.9% and 31.8%, respectively. In a univariate model, vascular invasion (P = .042) and fusiform cellular morphology (P = .001) were associated with poorer CSS for patients with SNM (Table 3). p-Akt1 was associated with poor CSS for SNM in the univariate (P = .001, log-rank = 11.079) and multivariate (HR = 65.726, P < .0001) models Figure 3. In addition, the clinical grade was also an independent prognostic marker for SNM (HR = 7.351, P = .019, Table 3).

| Factors | SampleCSS (%) | | Univariate | Multivariate | | |
|------------------------------|---------------|--------|------------|--------------|---------------------|---------|
| ractors | (%) | 3-year | 5-year | P (log-rank) | HR (95% CI) | P-value |
| Age | | | | | | |
| <58 | 16 (51.6) | 60.6% | 30.3% | .701 (.147) | 1.935 (.479–7.820) | .354 |
| ≥58 | 15 (48.4) | 51.3% | 38.5% | | | |
| Gender | | | | | | |
| Female | 15 (48.4) | 67% | 41.9% | .319 (.994) | _ | _ |
| Male | 16 (51.6) | 49.2% | 25.3% | | | |
| Treatment | | | | | | |
| Only surgery | 13 (41.9) | 63.5% | 25.4% | .604 (.268) | .473 (.132–1.694) | .250 |
| Surgery + chemo/radiotherapy | 17 (54.8) | 52.7% | 36.1% | | | |
| Anatomical site | | | | | | |
| Nasal cavity | 13 (41.9) | 56.6% | 45.3% | .239 (2.860) | _ | _ |
| Others | 18 (58.1) | 50% | 0% | | | |
| Mitotic rate | | | | | | |
| <1 | 20 (64.5) | 60.2% | 50.2% | .401 (.707) | .361 (.103–1.260) | .110 |
| ≥1 | 11 (35.5) | 54.5% | 10.9% | | | |
| Vascular invasion | | | | | | |
| Absent | 22 (70.9) | 63.9% | 45.7% | .042 (4.151) | .280 (.072–1.080) | .065 |
| Present | 9 (28.1) | 44.4% | 11.1% | | | |
| Neural invasion | | | | | | |
| Absent | 27 (87.1) | 63.1% | 38.5% | .072 (3.230) | 3.223 (.616–16.857) | .166 |
| Present | 4 (12.9) | 25% | 0% | | | |
| Cellular morphology | | | | | | |

 Table 3: The relationship between clinicopathological characteristics of patients

 with sinonasal melanomas, p-Akt1 expression and cancer-specific survival

| Factors | Sample | CSS | (%) | Univariate | Multivariate | |
|-----------------|-----------|--------|--------|---------------|------------------------|---------|
| Factors | (%) | 3-year | 5-year | P (log-rank) | HR (95% CI) | P-value |
| Epithelioid | 9 (29.1) | 100% | 62.5% | .001 (17.600) | .582 (.325–1.040) | .068 |
| Fusiform | 13 (41.9) | 15.4% | 7.7% | | | |
| AJCC-stage | | | | | | |
| III | 9 (28.1) | 85.7% | 64.3% | .060 (3.544) | 7.351 (1.392–38.821) | .019 |
| IVa, b and c | 22 (70.9) | 45% | 20.2% | | | |
| p-Akt1 | | | | | | |
| High expression | 20 (64.5) | 42.2% | 12.1% | .001 (11.079) | 65.726 (6.491–665.549) | <.0001 |
| Low expression | 11 (35.5) | 88.9% | 88.9% | | | |

CSS – Cancer–specific survival, HR – Hazard Ratio, AJCC – American Joint Committee on Cancer. Bold values indicate statistical significance (P < .05)



Figure 3: p-Akt1 expression in sinonasal melanomas. (A) Representative example of sinonasal melanoma in a tissue microarray, which was focally positive for p-Akt1. (B) Representative example of sinonasal melanoma in a tissue microarray, which was strongly positive for p-Akt1. (C) The association of p-Akt1-nuclear expression and

cancer-specific survival in sinonasal melanomas (log-rank = 11.079, P = .001). (D) The association of AJCC-stage and cancer-specific survival in sinonasal melanomas (logrank = 3.544, P = .060). The scale bars represent 1 mm (top) and 100 μ m (bottom).

p-Akt1 expression is associated with clinicopathological parameters in melanomas

In the cutaneous melanomas patients, p-Akt1 showed positive association with ulceration (P < .0001), growth phase (P < .0001), Breslow's Thickness (P < .0001), mitotic rate (P = .003), Clark's Level (P < .0001), metastasis (P = .008), AJCC-stage (P < .0001), and recurrence (P < .0001), as shown in Table 4. For oral melanomas, p-Akt1 expression showed association with vascular invasion (P = .001), mitotic rate (P < .0001), and epithelioid cellular morphology (P < .0001), as displayed in Table 5. In addition, in the SNM cohort (Table 6), vascular invasion (P = .008), mitotic rate (P = .023), and fusiform cellular morphology (P = .009) were significantly associated with the positive p-Akt1-nuclear expression.

Table4:The relationship between p-Akt1 nuclear expression andclinicopathological characteristics of 144 patients with cutaneous melanomas

| Clinicopathological characteristics | p-Akt1 low | p-Akt1 high | P- value |
|-------------------------------------------------------|--------------|---------------|-------------|
| Age (<56/≥56) | 44/31 | 34/35 | .259 |
| Gender (Female/Male) | 39/36 | 32/37 | .500 |
| Site (Trunk/Head and neck/Upper limbs/Lower Limbs) | 33/11/9/22 | 31/11/9/18 | .975 |
| Ulceration (Absent/Present) | 63/12 | 20/49 | <.0001 |
| Growth phase (Radial/Vertical) | 40/35 | 15/54 | <.0001 |
| Breslow's Thickness (<1.55/≥1.55) | 59/16 | 13/56 | <.0001 |
| Mitotic rate (<3/≥3) | 40/35 | 20/49 | .003 |
| Clark's Level (I and II/III, IV and V) | 40/35 | 7/62 | <.0001 |
| Metastasis (Absent/Present) | 64/11 | 46/23 | .008 |
| AJCC-stage (in situ/I/II/III/IV) | 15/39/10/8/3 | 0/12/32/15/10 | <.0001 |

| Clinicopathological characteristics | p-Akt1 low | p-Akt1 high | P- value |
|-------------------------------------|------------|-------------|-------------|
| Recurrence (Absent/Present) | 61/14 | 26/43 | <.0001 |

Table 5: The relationship between p-Akt1 nuclear expression andclinicopathological characteristics of 34 patients with oral melanomas

| Clinicopathological characteristics Age (<47/≥47) Gender (Female/Male) ite (Palate/alveolus/others) Creatment (only surgery/surgery plus chemo adiotherapy) Vascular invasion (Absent/Present) Meural invasion (Absent/Present) Meural invasion (Absent/Present) Mecrosis (Absent/Present) Cellular morphology (Non-epithelioid/Epithelioid) Recurrence (No/Yes) | p-Akt1 | p-Akt1 bigb | P- |
|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--------|----------------|--------|
| | 10 W | ingil | value |
| Age (<47/≥47) | 8/9 | 10/7 | .492 |
| Gender (Female/Male) | 8/9 | 8/9 | 1.000 |
| Site (Palate/alveolus/others) | 6/2/9 | 9/3/5 | .379 |
| Treatment (only surgery/surgery plus chemo- radiotherapy) | 13/4 | 9/7 | .151 |
| Vascular invasion (Absent/Present) | 15/2 | 6/11 | .001 |
| Neural invasion (Absent/Present) | 14/3 | 12/5 | .419 |
| Mitotic rate (<1/≥1) | 12/5 | 2/15 | <.0001 |
| Necrosis (Absent/Present) | 14/3 | 10/7 | .132 |
| Cellular morphology (Non-epithelioid/Epithelioid) | 12/5 | 0/17 | <.0001 |
| Recurrence (No/Yes) | 14/3 | 13/4 | .671 |

Table6:The relationshipbetweenp-Akt1nuclearexpressionandclinicopathological characteristics of 31 patients with sinonasal melanomas

| Cliniconothological characteristics | p-Akt1 | p-Akt1 | P- |
|--------------------------------------------------|--------|--------|-----------|
| Chincopathological characteristics | low | high | value |
| Age (<58/≥58) | 5/6 | 11/9 | .611 |
| Gender (Female/Male) | 7/4 | 8/12 | .208 |
| Site (Nasal Cavity/Maxillary sinus/Rhinopharynx) | 6/5/0 | 7/7/6 | .126 |

| Cliniconsthelogical characteristics | | p-Akt1 | <i>P</i> - |
|------------------------------------------------------------------------------|-------|--------|------------|
| Chincopathological characteristics | low | high | value |
| Treatment (only surgery/surgery plus chemo- radiotherapy) | 5/6 | 8/11 | .741 |
| Vascular invasion (Absent/Present) | 11/0 | 11/9 | .008 |
| Neural invasion (Absent/Present) | 11/0 | 16/4 | .112 |
| Mitotic rate (<1/≥1) | 10/1 | 10/10 | .023 |
| Necrosis (Absent/Present) | 8/3 | 10/10 | .220 |
| Cellular morphology (Epithelioid/Fusiform/Plasmacytoid- Undifferentiated) | 6/2/3 | 3/11/6 | .009 |
| Recurrence (No/Yes) | 7/4 | 12/8 | .842 |

p-Akt1 expression is higher in metastatic cutaneous melanomas and mucosal melanomas than in non-metastatic cutaneous melanomas

p-Akt1 expression was tested in a large cohort of cutaneous, oral, and sinonasal melanomas, and it was expressed in tumour cells and occasionally in inflammatory cells surrounding the tumour. We also compared the scores of p-Akt1-positive nuclei into four groups: (a) primary (cutaneous melanoma) without metastatic history; and (b) primary (cutaneous melanoma) with metastatic history, (c) oral melanomas, and (d) sinonasal melanomas.

The p-Akt1 expression was predominantly detected in the nucleus, and nonmetastatic cutaneous melanomas have a mean of 5.6% of the nuclei positive (ranging from 0% to 50.15%). Primary metastatic cutaneous melanomas have a mean of 15.7% (ranging from 1.7% to 41.3%). Cutaneous melanomas with low and high p-Akt1nuclear expression are illustrated in Figure 1. The representative photomicrographs for p-Akt1 expression in oral and sinonasal melanomas are illustrated in Figures 2 and 3, respectively. Oral melanomas have scores of p-Akt1-positivity (mean = 19.45%, ranging from 2.6% to 44.4%) similar to the cutaneous melanomas with metastatic history. The sinonasal melanomas demonstrated a higher number of nuclei positive for p-Akt1 (mean = 56.15%, ranging from 15.1% to 82.6%), statistically different from the other subtypes of melanomas analysed in this cohort (Figure 4). The nuclear p-Akt1expression was closely related with invasive cells in all tumours, as illustrated in Figure





Figure 4: p-Akt1 expression in invasive cells and in different subgroups of melanomas. (A) Invasive cells strongly express p-Akt1 in the nucleus of a case of sinonasal melanoma. (B) Graphical representation of the median, minimum, and maximum scores of p-Akt1-nuclear positivity in the different subgroups of melanomas (***indicates statistically significant difference between the groups, ANOVA test, *P* value < .00001). The scale bar represents 100 μ m.

Discussion

The poor prognosis of patients with mucosal melanomas and their ineffective treatment options evidence that these tumours have a distinct biological signature,

which is different from the other subtypes of melanomas [5, 10, 19]. Although some studies have extensively addressed the clinicopathological profile of the mucosal melanomas and their immunoprofile, these tumours have lacked predictive markers for tailored treatment [20–24]. Currently, the insufficient knowledge regarding molecular pathways that are selectively activated in different melanomas subtypes ensures comparative studies in mucosal and cutaneous melanomas. Herein, the authors proposed this comparative study to assess the p-Akt1 prognostic value in a series of cutaneous, oral, and sinonasal melanomas.

We used the AJCC system [25] for cutaneous melanomas staging. The AJCCstage of cutaneous melanoma's patients significantly affects the outcome, indicating that it is a reproducible method for suggesting the prognosis and therapeutic options. In this study, we also used the AJCC 7th edition staging system for the mucosal melanomas. This system is more useful in prognosis prediction than the Ballantyne's staging system, which is mostly applied [19]. However, we used a dichotomized model (III/IV AJCC-stages), and the AJCC-staging had no impact on survival rates for oral melanoma patients, in the multivariate model. The relatively small number of cases, one limitation of our study, may explicate this finding. However, as mucosal melanomas of the head and neck are very rare tumours, the authors have provided novel information regarding sixty-five patients with oral and sinonasal melanomas.

Melanomas with predominant epithelioid morphology have been considered tumours of poor prognosis, as these neoplasms have greater DNA ploidy abnormalities when compared with other cellular morphologies in cutaneous melanomas [26]. Interestingly, in the current study, the epithelioid cellular morphology was found to be associated with a poor prognosis in oral melanomas, and it is in accordance with the findings from previous studies [27]. We also demonstrated a statistically significant association of the epithelioid cellular morphology with p-Akt1 overexpression, indicating a probable correlation between these two features. On the other hand, in sinonasal melanomas, only the undifferentiated cell morphology has been reported to confer poorer prognosis [28]. However, in this study, the spindle cell morphology was associated with worse prognosis for sinonasal melanoma patients.

The protein kinase B is serine/threonine-specific protein kinase involved in several biological activities in a wide range of cells, such as glucose metabolism and apoptosis [29]. To date, three isoforms of this protein have been well-described, AKT1, AKT2, and AKT3 [30]. AKT1 is widely documented for its activity in cellular

31

metabolism of several human cancers [29] and when phosphorylated, p-Akt1 is described as playing an important role in the redox modulation of cell cycle progression [31]. On the other hand, Akt2 seems to play a critical role in cell proliferation through increased glycogen synthesis [32] and Akt3 expression is reported in normal tissues, including the brain, heart, kidney, and fat [33].

Previous studies have demonstrated that p-Akt1-nuclear expression is closely associated with worse prognosis in breast [12], gastric [13], and oesophageal squamous carcinoma [14]. In opposition, it is associated with a favourable outcome in patients with pancreatic cancer [34]. The tumorigenic activity of p-Akt1 has also been investigated in several tumours [35-37] and in the context of tumour cells, Akt1 upregulates cell proliferation, invasion, and migration [38, 39]. Contrastively, its deletion prevents lung tumorigenesis in mice models [40]. One previous study shows the putative role of p-Akt1 in metastatic cutaneous melanomas [41]. In this study, the nuclear p-Akt1 expression was associated with the presence of distant metastasis in patients with cutaneous melanomas. In fact, several studies have addressed the role of p-Akt1 in cancer progression and metastasis [42], including its important part in the development of melanoma metastases in a mice model [18]. In addition, many studies have demonstrated the role of Akt1 in metastases development in several malignancies, such as colorectal [43] and lung [44] cancers. Although a correlation between p-Akt1 expression and the metastatic status in cutaneous melanomas have been found for these tumours, the p-Akt1 showed prognostic values only in the univariate model, but it was lost in the multivariate model. In addition, p-Akt1 expression was correlated with important clinicopathologic features in the cutaneous melanomas, such as Breslow's thickness, Clark's level, mitotic rate, and AJCC-stage, indicating that p-Akt1 is correlated with melanoma tumorigenesis and affects the clinical outcome of these patients, emerging as a possible target for personalized therapies.

For oral and sinonasal melanomas, univariate and multivariate analysis identified p-Akt1 as an independent prognostic factor. Our results also showed that p-Akt1 expression was significantly higher in sinonasal melanomas than in oral and cutaneous melanomas. Taken together, these data support the hypothesis that sinonasal melanomas have distinct molecular signature and biological behaviour from its cutaneous counterpart. Importantly, we have demonstrated by immunohistochemistry that p-Akt1 is overexpressed in the nuclei of tumour cells and is closely correlated with poor outcome in a subset of mucosal melanomas, whereas the cutaneous melanomas

demonstrated lower p-Akt1 expression compared with the oral and sinonasal melanomas. In fact, several studies have addressed the differences concerning etiologic factors and molecular pathways involved in the pathogenesis of cutaneous vs mucosal melanomas [3–5, 45]. For example, c-KIT aberrations were reported in tumours that have no correlation with chronic sun exposure (mainly mucosal melanomas). In opposition, the cutaneous counterpart frequently harbours mutations in the BRAF gene, given the fact that chronic sun exposure is important for its pathogenesis [9, 46]. It has been previously demonstrated that c-KIT is a key molecule for p-Akt1 activation through PI3K phosphorylation [46]. Therefore, particularly in mucosal melanomas, a correlation between p-Akt1 overexpression and c-KIT mutation may help to understand how these pathways are important for melanoma pathogenesis and patient outcomes.

In the cohort of sinonasal melanomas analysed in this study, p-Akt1 overexpression showed a correlation with higher number of mitosis, presence of vascular invasion, and with spindle and undifferentiated cellular morphology. These findings provide new insights about the probable role of p-Akt1 in the aggressive genotype and phenotype of the sinonasal melanomas. In these particular tumours, necrosis was associated with overall survival in one previous report [47]. However, in this study's cohort, necrosis has not demonstrated prognostic predicting value, corroborating with other previous studies [48, 49]. No correlation between p-Akt1 and necrosis was observed in this study.

Overall, the treatment of choice for both mucosal and cutaneous melanoma is the wide surgical resection [50–52]. Although it has been demonstrated that melanomas are not radiosensitive, some tumours have been treated with adjuvant radiotherapy, and the use of these combined therapies are not standardized [10]. Regarding the mucosal melanomas, approximately 30% of the patients from the sample with oral melanomas and 50% of the patients with sinonasal melanomas received chemotherapy or radiotherapy as adjuvant treatment. Nevertheless, no differences in the cancer-specific survival were observed in both groups. The use of adjuvant therapies with surgical resection still needs to be investigated as a option to treat patients with mucosal melanomas [53].

In conclusion, the results of the current study demonstrated that p-Akt1 overexpression is an independent prognostic marker in mucosal melanomas and is significantly up-regulated in sinonasal melanomas. As both mucosal and metastatic cutaneous melanomas showed high frequency of p-Akt1 expression, our findings

suggest that mucosal melanomas have a biological behaviour that is similar to the one identified in aggressive cutaneous melanomas.

Abbreviations

CM: cutaneous melanomas; OM: oral melanomas; SNM: sinonasal melanomas; HR: Hazard Ratio; AJCC: American Joint Committee on Cancer; p-Akt1: Phosphorylated Akt1, or protein B kinase 1; IHC: immunohistochemistry; CSS: Cancer-specific survival; DFS: disease-free survival.

Author contributions

Conception and design: CDS, JJ, OPA. Provision of study materials and/or patients: RC, FVMBC, MBC, RRDR, LSA, AMMA, MGFC, OPA. Collection and assembly of data: CDS, TMLM, MGFC, OPA, JJ. Data analysis and interpretation: CDS, TMLM, FVMBC, AMMA, JJ, Manuscript writing: CDS, TMLM. Final approval of manuscript: All authors approved the definitive version of this manuscript.

ACKNOWLEDGMENTS

The authors thank Dr. Gabriel Silva (MD) and Dr. Carlos Galvão (MD) for the support in the clinical and follow-up data.

CONFLICTS OF INTEREST

The authors have no conflicts of interest to disclose.

FUNDING

This work was supported by São Paulo Research Foundation (FAPESP), grants #2015/25905-1, #2017/16102-8 and #2017/18134-4. None of the funding sources had any part in the study design, data collection/analysis, data interpretation, or manuscript writing.

References

1. Foletto MC, Haas SE. Cutaneous melanoma: new advances in treatment. An Bras Dermatol. 2014; 89:301–10.

2. Karimkhani C, Green AC, Nijsten T, Weinstock MA, Dellavalle RP, Naghavi M, Fitzmaurice C. The global burden of melanoma: results from the Global Burden of Disease Study 2015. Br J Dermatol. 2017; 177:134–40.

3. Korabiowska M, Brinck U, Hoenig JF, Bartkowski SB, Mirecka J, Schauer A. An application of MIB antibody to the retrospective study of melanomas of oral mucosa and facial skin. J Cancer Res Clin Oncol. 1994; 120:365–8.

4. Bishop KD, Olszewski AJ. Epidemiology and survival outcomes of ocular and mucosal melanomas: A population-based analysis. Int J Cancer. 2014; 134:2961–71.

5. Kuk D, Shoushtari AN, Barker CA, Panageas KS, Munhoz RR, Momtaz P, Ariyan CE, Brady MS, Coit DG, Bogatch K, Callahan MK, Wolchok JD, Carvajal RD, et al. Prognosis of Mucosal, Uveal, Acral, Nonacral Cutaneous, and Unknown Primary Melanoma From the Time of First Metastasis. Oncologist. 2016; 21:848–54.

6. Alaeddini M, Etemad-Moghadam S. Immunohistochemical profile of oral mucosal and head and neck cutaneous melanoma. J Oral Pathol Med. 2015; 44:234–8.

7. Moreno MA, Roberts DB, Kupferman ME, Demonte F, El-Naggar AK, Williams M, Rosenthal DS, Hanna EY. Mucosal melanoma of the nose and paranasal sinuses, a contemporary experience from the M. D. Anderson cancer center. Cancer. 2010; 116:2215–23.

8. Patel SG, Prasad ML, Escrig M, Singh B, Shaha AR, Kraus DH, Boyle JO, Huvos AG, Busam K, Shah JP. Primary mucosal malignant melanoma of the head and neck. Head Neck. 2002; 24:247–57.

9. de Mendonça UB, Cernea CR, Matos LL, Toscano de Mendonça UB, Monteiro de Araujo Lima RR. Analysis of KIT gene mutations in patients with melanoma of the head and neck mucosa: a retrospective clinical report. Oncotarget. 2018; 9:22886–94. <u>https://doi.org/10.18632/oncotarget.25094</u>.

10. Luna-Ortiz K, Villavicencio-Valencia V, Martinez Said H. Comparative study of head and neck mucosal melanoma in 66 patients vs 226 patients with cutaneous melanoma: A survival analysis. Clin Otolaryngol. 2018; 43:691–696.

11. Cicenas J. The potential role of Akt phosphorylation in human cancers. Int J Biol Markers. 2008; 23:1–9.

12. Liu J, Wei XL, Huang WH, Chen CF, Bai JW, Zhang GJ. Cytoplasmic Skp2 Expression Is Associated with p-Akt1 and Predicts Poor Prognosis in Human Breast Carcinomas. PLoS One. 2012; 7: e52675.

13. Cao F, Zhang C, Han W, Gao XJ, Ma J, Hu YW, Gu X, Ding HZ, Zhu LX, Liu Q.
p-Akt as a potential poor prognostic factor for gastric cancer: a systematic review and meta-analysis.
Oncotarget.
2017;
8:59878–
88. https://doi.org/10.18632/oncotarget.17001.

14. Zhu Z, Yu W, Fu X, Sun M, Wei Q, Li D, Chen H, Xiang J, Li H, Zhang Y, Zhao W, Zhao K. Phosphorylated AKT1 is associated with poor prognosis in esophageal squamous cell carcinoma. J Exp Clin Cancer Res. 2015; 34: 95.

15. Vivanco I, Chen ZC, Tanos B, Oldrini B, Hsieh WY, Yannuzzi N, Campos C, Mellinghoff IK. A kinase-independent function of AKT promotes cancer cell sur vival. eLife. 2014; 3:3.

16. Möser CV, Meissner M, Laarmann K, Olbrich K, King-Himmelreich TS, Wolters MC, Geisslinger G, Niederberger E. The protein kinase IKKepsilon contributes to tumour growth and tumour pain in a melanoma model. Biochem Pharmacol. 2016;103:64–73.

17. Montenegro RC, de Vasconcellos MC, Barbosa GS, Burbano RM, Souza LG, Lemos TL, Costa-Lotufo LV, de Moraes MO. A novel o-naphtoquinone inhibits N-cadherin expression and blocks melanoma cell invasion via AKT signaling. Toxicol *In Vitro*. 2013; 27:2076–83.

18. Cho JH, Robinson JP, Arave RA, Burnett WJ, Kircher DA, Chen G, Davies MA, Grossmann AH, VanBrocklin MW, McMahon M, Holmen SL. AKT1 Activation Promotes Development of Melanoma Metastases. Cell Rep. 2015;13:898–905.

19. Luna-Ortiz K, Aguilar-Romero M, Villavicencio-Valencia V, Zepeda-Castilla E, Vidrio-Morgado H, Peteuil N, Mosqueda-Taylor A. Comparative study between two

different staging systems (AJCC TNM VS BALLANTYNE'S) for mucosal melanomas of the head & neck. Med Oral Patol Oral Cir Bucal. 2016;21: e425–30.

20. Prasad ML, Busam KJ, Patel SG, Hoshaw-Woodard S, Shah JP, Huvos AG. Clinicopathologic differences in malignant melanoma arising in oral squamous and sinonasal respiratory mucosa of the upper aerodigestive tract. Arch Pathol Lab Med. 2003;127:997–1002.

21. Lourenço SV, A MS, Sotto MN, Bologna SB, Giacomo TB, Buim ME, Coutinho-Camillo CM, Silva SD, Landman G, Soares FA, Simonsen Nico MM. Primary oral mucosal melanoma: A series of 35 new cases from south America. Am J Dermatopathol. 2009; 31:323–30.

22. Benevenuto de Andrade BA, Piña AR, León JE, Paes de Almeida O, Altemani A. Primary nasal mucosal melanoma in Brazil: clinicopathologic and immunohistochemical study of 12 patients. Ann Diagn Pathol. 2012;16:344–9.

23. de Andrade BAB, León JE, Carlos R, Delgado-Azañero W, Mosqueda-Taylor A, de Almeida OP. Immunohistochemical expression of p16, p21, p27 and cyclin D1 in oral nevi and melanoma. Head Neck Pathol. 2012; 6:297–304.

24. de-Andrade BAB, Toral-Rizo VH, León JE, Contreras E, Carlos R, Delgado-Azañero W, Mosqueda-Taylor A, de-Almeida OP. Primary oral melanoma: a histopathological and immunohistochemical study of 22 cases of Latin America. Med Oral Patol Oral Cir Bucal. 2012; 17: e383-8.

25. Edge SB, Compton CC. The american joint committee on cancer: The 7th edition of the AJCC cancer staging manual and the future of TNM. Ann Surg Oncol. 2010; 17:1471–4.

26. Chi HI, Uyeda Y, Umebayashi Y, Otsuka F. Epithelioid cell melanomas have greater DNA ploidy abnormalities than spindle cell melanomas: cytological evidence for a higher malignant potential of the former. Arch Dermatol Res. 1993; 285:410–4.

27. Song H, Wang L, Lyu J, Wu Y, Guo W, Ren G. Loss of nuclear BAP1 expression is associated with poor prognosis in oral mucosal melanoma. Oncotarget. 2017; 8:29080–90. <u>https://doi.org/10.18632/oncotarget.16175</u>.
28. Narasimhan K, Kucuk O, Lin HS, Heilbrun LK, Carron M, Venkatramanamoorthy R, Mathog R. Sinonasal mucosal melanoma: a 13-year experience at a single institution. Skull Base. 2009; 19:255–62.

29. Carpten JD, Faber AL, Horn C, Donoho GP, Briggs SL, Robbins CM, Hostetter G, Boguslawski S, Moses TY, Savage S, Uhlik M, Lin A, Du J, et al. A transforming mutation in the pleckstrin homology domain of AKT1 in cancer. Nature. 2007; 448:439–44.

30. Santi SA, Lee H. The Akt isoforms are present at distinct subcellular locations. Am J Physiol Cell Physiol. 2010; 298:C580–91.

31. Arciuch VGA, Galli S, Franco MC, Lam PY, Cadenas E, Carreras MC, Poderoso JJ. Akt1 intramitochondrial cycling is a crucial step in the redox modulation of cell cycle progression. PLoS One. 2009;4:e7523.

32. Garofalo RS, Orena SJ, Rafidi K, Torchia AJ, Stock JL, Hildebrandt AL, Coskran T, Black SC, Brees DJ, Wicks JR, McNeish JD, Coleman KG. Severe diabetes, agedependent loss of adipose tissue, and mild growth deficiency in mice lacking Akt2/PKBβ. J Clin Invest. 2003;112:197–208.

33. Easton RM, Cho H, Roovers K, Shineman DW, Mizrahi M, Forman MS, Lee VMY, Szabolcs M, de Jong R, Oltersdorf T, Ludwig T, Efstratiadis A, Birnbaum MJ. Role for Akt3/Protein Kinase B in Attainment of Normal Brain Size. Mol Cell Biol. 2005;25:1869–78.

34. Liu J, Cheng Sun SH, Sun SJ, Huang C, Hu HH, Jin YB, Qiu ZJ. Phosph-Akt1 expression is associated with a favourable prognosis in pancreatic cancer. Ann Acad Med Singapore. 2010;39:548–7.

35. Shen G, Rong X, Zhao J, Yang X, Li H, Jiang H, Zhou Q, Ji T, Huang S, Zhang J, Jia H. MicroRNA-105 suppresses cell proliferation and inhibits PI3K/AKT signaling in human hepatocellular carcinoma. Carcinogenesis. 2014;35:2748–55.

36. Wu Y, Kim J, Elshimali Y, Sarkissyan M, Vadgama JV. Activation of Akt1 accelerates carcinogen-induced tumorigenesis in mammary gland of virgin and postlactating transgenic mice. BMC Cancer. 2014;14:266. 38. Yang L, Xiao L, Ma X, Tang M, Weng X, Chen X, Sun L, Cao Y. Effect of DNAzymes targeting Akt1 on cell proliferation and apoptosis in nasopharyngeal carcinoma. Cancer Biol Ther. 2009;8:366–71.

39. Chen L, Kang QH, Chen Y, Zhang YH, Li Q, Xie SQ, Wang CJ. Distinct roles of Akt1 in regulating proliferation, migration and invasion in HepG2 and HCT 116 cells. Oncol Rep. 2014; 31:737–44.

40. Hollander MC, Maier CR, Hobbs EA, Ashmore AR, Linnoila RI, Dennis PA. Akt1 deletion prevents lung tumorigenesis by mutant K-ras. Oncogene. 2011; 30:1812–21.

41. Soares CD, Borges CF, Sena-Filho M, Almeida OP, Stelini RF, Cintra ML, Graner E, Zecchin KG, Jorge J. Prognostic significance of cyclooxygenase 2 and phosphorylated Akt1 overexpression in primary nonmetastatic and metastatic cutaneous melanomas. Melanoma Res. 2017; 27:448–56.

42. Ooms LM, Binge LC, Davies EM, Rahman P, Conway JRW, Gurung R, Ferguson DT, Papa A, Fedele CG, Vieusseux JL, Chai RC, Koentgen F, Price JT, et al. The Inositol Polyphosphate 5-Phosphatase PIPP Regulates AKT1-Dependent Breast Cancer Growth and Metastasis. Cancer Cell. 2015; 28:155–69.

43. Agarwal E, Brattain MG, Chowdhury S. Cell survival and metastasis regulation by Akt signaling in colorectal cancer. Cell Signal. 2013; 25:1711–9.

44. Yu S, Zhang C, Dong F, Zhang Y. miR-99a Suppresses the Metastasis of Human Non-Small Cell Lung Cancer Cells by Targeting AKT1 Signaling Pathway. J Cell Biochem. 2015; 116:268–76.

45. Turri-Zanoni M, Medicina D, Lombardi D, Ungari M, Balzarini P, Rossini C, Pellegrini W, Battaglia P, Capella C, Castelnuovo P, Palmedo G, Facchetti F, Kutzner H, et al. Sinonasal mucosal melanoma: Molecular profile and therapeutic implications from a series of 32 cases. Head Neck. 2013; 35:1066–77.

46. Lourenço SV, Fernandes JD, Hsieh R, Coutinho-Camillo CM, Bologna S, Sangueza M, Nico MMS. Head and neck mucosal melanoma: A review. Am J Dermatopathol. 2014; 36:578–87.

47. Mochel MC, Duncan LM, Piris A, Kraft S. Primary mucosal melanoma of the sinonasal tract: a clinicopathologic and immunohistochemical study of thirty-two cases. Head Neck Pathol. 2015; 9:236–43.

48. Thompson LDR, Wieneke JA, Miettinen M. Sinonasal tract and nasopharyngeal melanomas: a clinicopathologic study of 115 cases with a proposed staging system. Am J Surg Pathol. 2003; 27:594–611.

49. Prasad ML, Patel SG, Huvos AG, Shah JP, Busam KJ. Primary mucosal melanoma of the head and neck. Cancer. 2004; 100:1657–64.

50. Mendenhall WM, Amdur RJ, Hinerman RW, Werning JW, Villaret DB, Mendenhall NP. Head and neck mucosal melanoma. Am J Clin Oncol. 2005; 28:626–30.

51. McLean N, Tighiouart M, Muller S. Primary mucosal melanoma of the head and neck. Comparison of clinical presentation and histopathologic features of oral and sinonasal melanoma. Oral Oncol. 2008; 44:1039–46.

52. Warszawik-Hendzel O, Słowińska M, Olszewska M, Rudnicka L. Melanoma of the oral cavity: pathogenesis, dermoscopy, clinical features, staging and management. J

Dermatol Case Rep. 2014; 8:60-6.

53. Cheng YF, Lai CC, Ho CY, Shu CH, Lin CZ. Toward a Better Understanding of Sinonasal Mucosal Melanoma: Clinical Review of 23 Cases. J Chin Med Assoc. 2007; 70:24–9.

54. Montezuma MAP, Fonseca FP, Benites BM, Soares CD, do Amaral-Silva GK, de Almeida OP, Soares FA, Pagano RL, Fregnani ER. COX-2 as a determinant of lower disease-free survival for patients affected by ameloblastoma. Pathol Res Pract. 2018; 214:907–913.

55. Fonseca FP, Bingle L, Santos-Silva AR, Lopes MA, de Almeida OP, de Andrade BA, Mariano FV, Kowalski LP, Rangel AL, Martins MD, Meurer L, Speight PM, Vargas PA. Semaphorins and neuropilins expression in salivary gland tumors. J Oral Pathol Med. 2016; 45:119–26.

| Factors | Category | Frequency |
|---------------------------|---------------|-----------|
| | | n (%) |
| Sex | Female | 71 (49.3) |
| | Male | 73 (50.7) |
| Age (years) | Range; median | 20-88; 56 |
| | Mean (SD) | 56 (15.7) |
| Site | Head and neck | 22 (15.3) |
| | Trunk | 64 (44.4) |
| | Upper limbs | 18 (12.5) |
| | Lower limbs | 40 (27.8) |
| T-stage | In situ | 15 (10.4) |
| | Ι | 51 (35.4) |
| | II | 42 (29.2) |
| | III | 23 (16.0) |
| | IV | 13 (9.0) |
| Clark's level | I and III | 47 (32.6) |
| | III, IV and V | 97 (67.4) |
| Ulceration | Present | 61 (42.4) |
| | Absent | 83 (57.6) |
| Mitotic index | <3 | 60 (41.7) |
| (number/mm ²) | <u>≥</u> 3 | 84 (58.3) |
| Breslow's | <1.55 | 72 (50) |
| thickness (mm) | ≥1.55 | 72 (50) |
| Distant metastasis | Present | 34 (23.6) |
| | Absent | 11 (76.4) |

Supplementary Table 1. Clinicopathologic features of 144 patients with cutaneous melanomas.

| Factors | Category | Frequency |
|---------------------------|------------------|-----------|
| | 0 1 | n (%) |
| Sex | Female | 16 (47.1) |
| | Male | 18 (52.9) |
| Age (years) | Range; median | 19–89; 47 |
| | Mean (SD) | 52 (19.5) |
| Site | Palate | 15 (44.1) |
| | Inferior gingiva | 5 (14.7) |
| | Other | 14 (41.2) |
| Cell morphology | Epithelioid | 22 (64.7) |
| | Non-Epithelioid | 12 (35.3) |
| Vascular invasion | Present | 13 (38.2) |
| | Absent | 21 (61.8) |
| Neural invasion | Present | 8 (23.5) |
| | Absent | 26 (76.5) |
| Necrosis | Present | 10 (29.4) |
| | Absent | 24 (70.6) |
| Mitotic index | <1 | 14 (41.2) |
| (number/mm ²) | ≥1 | 20 (58.8) |
| Treatment | Only surgery | 22 (64.7) |
| | Surgery plus | 11 (32.4) |
| | chemotherapy or | |
| | radiotherapy | |
| | Other/missing | 1 (2.9) |
| Clinical Stage | III | 9 (26.5) |
| ~ | IVa | 16 (47.1) |
| | IVb | 4 (11.8) |
| | IVc | 5 (14.7) |

Supplementary Table 2. Clinicopathologic features of 34 patients with oral melanomas.

| Factors | Category | Frequency |
|---------------------------|------------------|-----------|
| | | n (%) |
| Sex | Female | 15 (48.4) |
| | Male | 16 (51.6) |
| Age (years) | Range; median | 24-82; 58 |
| | Mean (SD) | 55 (17) |
| Site | Nasal cavity | 13 (41.9) |
| | Maxillary sinus | 12 (38.7) |
| | Rhinopharynx | 6 (19.4) |
| Cell morphology | Epithelioid | 9 (29.0) |
| | Fusiform | 13 (41.9) |
| | Plasmacytoid | 4 (12.9) |
| | Undifferentiated | 5 (16.1) |
| Vascular invasion | Present | 9 (28.1) |
| | Absent | 22 (71.9) |
| Neural invasion | Present | 4 (12.9) |
| | Absent | 27 (87.1) |
| Necrosis | Present | 13 (41.9) |
| | Absent | 18 (58.1) |
| Mitotic index | <1 | 20 (64.5) |
| (number/mm ²) | ≥1 | 11 (35.5) |
| Treatment | Only surgery | 13 (41.9) |
| | Surgery plus | 17 (54.8) |
| | chemotherapy or | |
| | radiotherapy | |
| | Other/missing | 1 (3.3) |
| Clinical stage | III | 9 (29.0) |
| | IVa | 11 (35.5) |
| | IVb | 4 (12.9) |
| | IVc | 7 (22.6) |

Supplementary Table 3. Clinicopathologic features of 31 patients with sinonasal melanomas.

2.2 Artigo: Cyclooxygenase 2 is associated with aggressiveness and unfavorable survival in oral melanoma

Ciro Dantas Soares¹, Lucas Lacerda de Souza², Roman Carlos³, Oslei Paes de Almeida¹, Maria Goretti Freire de Carvalho⁵, Madja Ruanna Soares Macedo⁵, Juan Carlos Hernandez-Guerrero⁶, Adalberto Mosqueda-Taylor⁷, Jacks Jorge¹

1. Department of Oral Diagnosis, Area of Pathology, Piracicaba Dental School, University of Campinas, Piracicaba, São Paulo, Brazil.

2. Department of Oral Pathology, University Hospital João de Barros Barreto, Federal University of Pará, Belém, Pará, Brazil.

3. Pathology Division, Centro Clínico de Cabeza y Cuello/Hospital Herrera Llerandi, Guatemala City, Guatemala.

5. Department of Nursing, Federal University of Rio Grande do Norte, Natal, Rio Grande do Norte, Brazil.

6. Immunology Laboratory, Faculty of Dentistry, National University Autonomous of Mexico

7. Health Care Department, Universidad Autónoma Metropolitana, Xochimilco, Mexico

Running tittle: COX-2 expression in oral melanoma.

Corresponding author:

Ciro Dantas Soares

Department of Oral Diagnosis, Piracicaba Dental School, University of Campinas

Avenida Limeira, 901, Areião - 13414-903 Piracicaba/SP, Brazil

E-mail address: ciro.dss@gmail.com

ABSTRACT

Objectives: The aim of this study was to investigate the association of cyclooxygenase 2 (COX-2) expression with clinicopathological data and survival. *Materials and methods:* immunohistochemical (IHC) staining was used to measure the expression of COX-2 on the tissue microarray from 30 patients with oral melanoma. A digital analysis was performed by slides scanning and scores of staining were assessed. The immunostaining scores were correlated with clinical parameters. Survival data were evaluated by logrank test and COX proportional hazards regression analysis. *Results:* All oral melanomas were positive for COX-2 in different intensities. COX-2 expression was significantly associated with overall survival in a multivariate model [HR = 6.6475 (2.0466-21.5919); P-value = 0.0016]. *Conclusions:* Our results suggest that COX-2 may be considered a valuable prognostic marker for oral melanoma. These findings may help the development of target-based therapies focusing on COX-2 inhibitors as effective single or combined therapy against oral melanoma.

Keywords: Melanoma; oral; COX-2; prognosis.

INTRODUCTION

Cyclooxygenases (COXs) are enzymes responsible by converting the first step of arachidonic acid to prostaglandins and constitutes two subtypes, wherein COX-1 is mostly observed in normal tissue due to its association with physiological processes, while COX-2 is not detected in normal tissues, but is immediately induced by inflammatory reactions [1,2]. COX-2 is also expressed in several tumour types and its level of expression are associated with proliferation in non-small cell lung cancer [3], angiogenesis in pancreatic ductal adenocarcinoma [4], cell migration and invasion in hepatocellular carcinoma [5] and promotes apoptotic resistance in colorectal cancer [6]. COX-2 expression has already been demonstrated in oral and cutaneous metastatic melanoma [1,2,8], but the clinical significance of COX-2 expression in oral melanomas was not addressed in these articles.

Therefore, in this current study we sought to elucidate the prognostic significance of COX-2 overexpression in oral melanomas.

METHODS

This protocol of this study followed the ethical principles stated in the Declaration of Helsinki and was approved by the National Commission for Ethics in Research (CONEP-Brazil, CAAE: 72077517.1.0000.5418). We have included 30 patients with oral melanoma from four pathology laboratories which presented complete clinical information and follow-up. The lesions were diagnosed following the 4th edition for Mucosal Melanoma of the World Health Organization Classification of Head and Neck Tumours criteria [9]. In amelanotic or doubt cases, S-100, MelanA, and HMB45 immunostainings were performed. The staging classification was performed according to the 8th edition of the American Joint Committee on Cancer for Melanoma Staging [10].

For the immunohistochemical (IHC) polymer-based method accordingly protocols stablished in our laboratory [8,11]. We used the primary antibody COX-2 (Clone: CX-294; Dako Corporation, Carpinteria, CA, USA). Following the IHC reactions, the slides were scanned into high-resolution images and digitally assessed as described previously [8,11]. The immunostaining scores were correlated with clinical data using contingency tables and Chi- Square or Fisher's exact test. Survival curves were calculated according to the Kaplan-Meier method. The Log-Rank test was applied for patients with low and high markers expression (cut off value: median of positivity scores). COX proportional hazards regression analysis was performed to test the statistical independence and significance between pathological, molecular, and clinical variables. All statistical tests were carried out in the SPSS software, version 22.0 (SPSS Inc., Chicago, IL, USA) with a 95%-confidence level (P-value ≤ 0.05).

RESULTS

The current samples have already been used in other previous studies [8,11]. The mean age was 54 years old (range of 22-89 years-old); females were slight more affected than males (M:F ratio of 1:1.14). Regarding the anatomical site, palate was more affected (13 cases; 43.4%), followed by gingiva (5 cases; 16.6%). In addition, patients were mainly treated with only surgery (20 cases; 66.6%) and surgery plus chemotherapy or radiotherapy (9 cases; 30%). Regarding the tumour's AJCC-stage classification, the cases were stage III in 43.4%, IVa in 36.6% and IVb/IVc in 20%.

COX-2-positive cells showed reddish granules in the cytoplasm, evidenced by a variable cytoplasmic staining in all analysed samples. Its high expression was

significantly associated with vascular invasion (P-value: 0.0018), high mitotic index (P-value: 0.0344), epithelioid cellular morphology (P-value: 0.0032) and advanced AJCC-stage level (P-value: 0.0043). The univariate and multivariate Cox model revealed that none of the analysed factors was significantly associated with DFS. Concerning OS, the univariate model evaluation revealed that anatomical site (P-value: 0.0239), treatment (P-value: 0.0004), vascular invasion (P-value: < 0.0001), neural invasion (P-value: 0.0341), mitotic index (P-value: 0.0002), epithelioid cellular morphology (P-value: 0.0018), advanced AJCC-stage (P-value: 0.0007) and high COX-2 expression (P-value: 0.0003) showed significant results. The multivariate Cox model revealed that only high COX-2 expression was significantly associated with overall survival (P-value: 0.0016) (Table 1).



Figure 1. Immunohistochemical expression of COX-2 in oral melanomas. A-B: Positive expression in the cytoplasm of tumour cells of a predominantly spindle cell oral melanoma – a (weak), b (moderate). C-D: COX-2 demonstrated intense positivity in all epithelioid oral melanomas.

| | Categories | | DFS univariate (%) | | Multiva | Multivariate | | variate 6) | | Multivariate | | |
|-------------------|-----------------------------|-----------|-----------------------|---------|-----------|----------------------|---------|---------------|---------|--------------|----------------------|---------|
| Variables | | n (%) | 3-years | 5-years | P-value | HR (95% | P-value | 3-years | 5-years | P-value | HR (95% | P-value |
| | | | | | (logrank) | CI) | | | | (logrank) | CI) | |
| Age | <54 | 16 (53.3) | 88.8 | 71.6 | 0.0873 | 4.0731 | 0.8309 | 53.3 | 40.1 | 0.7634 | 1.0568 | 0.9654 |
| | ≥54 | 14 (46.7) | 63.5 | 35.8 | | (2.4287- 6.8313) | | 43.8 | - | | (0.0867- 12.8837) | |
| Gender | Female | 16 (53.3) | 78.7 | 78.7 | 0.7574 | 127.4771 (7.2946- | 0.9415 | 38.3 | 38.3 | 0.3318 | 1.7371 (0.2743- | 0.5575 |
| | Male | 14 (46.7) | 82 | 46.8 | - | 228.1245) | | 64.3 | 42.8 | | 11.0008) | |
| Anatomical site | Palate | 13 (43.4) | 63 | 63 | 0.1808 | 0.0011 | 0.8407 | 27.7 | 27.7 | 0.0239* | 0.3580 | 0.1184 |
| | Gingiva | 5 (16.6) | 50 | 50 | | (2.0369- | | 40 40 | 40 | | (0.0986- | |
| | Others | 12 (40) | 100 | 66.6 | | 9.6351) | | 81.8 | 65.4 | | 1.2998) | |
| Treatment | Only surgery | 20 (66.6) | 86.2 | 71.9 | 0.3679 | 484.1991 (4.3047- | 0.8654 | 57.4 | 57.4 | 0.0004* | 1.9265 (0.5707- | 0.2907 |
| | Surgery plus CH or RT | 9 (30) | 64.3 | 42.8 | | 5.4464) | | 44.4 | 14.8 | | 6.5024) | |
| | Others | 1 (0.4) | - | - | | | | - | - | | | |
| Vascular invasion | Absent | 19 (63.3) | 87.4 | 56.6 | 0.1161 | 5.8936 | 0.8709 | 83.6 | 60.9 | < | 1.0944 | 0.9425 |
| | Present | 11 (36.6) | - | - | | (2.8031- 1.2392) | | - | _ | 0.0001* | (0.0943- 12.6992) | |
| Neural invasion | Absent | 24 (80) | 82.2 | 53.3 | 0.4965 | 1596.5338 | 0.9562 | 60.4 | 52.8 | 0.0341* | 1.4340 | 0.7052 |
| | Present | 6 (20) | - | - | | (6.5659- 3.8821) | | - | - | | (0.2214- 9.2879) | |
| Mitotic index | <1 | 13 (43.4) | 90 | 72 | 0.1558 | 23.7592 | 0.9613 | 91.6 | 91.6 | 0.0002* | 10.9201 | 0.1047 |
| | ≥1 | 17 (56.6) | 72.2 | 36 | | (4.9228- | | 23.5 | 17.6 | | (0.6079- | |

Table 1. Clinicopathological characteristics and survival analysis of all patients with oral melanoma.

| | | | | | | 1.1467) | | | | | 196.1630) | |
|------------|-------------|-----------|------|------|--------|-----------|--------|------|------|---------|-----------|---------|
| Necrosis | Absent | 22 (73.3) | 74.2 | 49.9 | 0.4002 | 0.0002 | 0.8731 | 51.2 | 41 | 0.5963 | 0.6026 | 0.7307 |
| | Present | 8 (26.6) | 100 | 100 | _ | (1.0136- | | 50 | 50 | | (0.0337- | |
| | | | | | | 8.2176) | | | | | 10.7745) | |
| Cellular | Non- | 10 (33.3) | 76.2 | 76.2 | 0.1756 | 0.000001 | 0.8468 | 88.8 | 88.8 | 0.0018* | 2.4372 | 0.4841 |
| morphology | epithelioid | | | | | (1.5589- | | | | | (0.2008- | |
| | Epithelioid | 20 (66.6) | 84.8 | 42.4 | | 6.6615) | | 35 | 23.3 | | 29.5684) | |
| AJCC-stage | III | 13 (43.4) | 80.2 | 50.1 | 0.4554 | 6384.8337 | 0.7957 | 68.3 | 68.3 | 0.0007* | 2.5116 | 0.1550 |
| | IVa | 11 (36.6) | 88.8 | 59.2 | | (9.8415- | | 60.6 | 40.4 | | (0.7057- | |
| | IVb/IVc | 6 (20) | - | - | | 4.1423) | | - | - | | 8.9386) | |
| COX-2 | Low | 19 (63.3) | 86.5 | 67.3 | 0.1875 | 3.7241 | 0.2145 | 78.3 | 68.5 | 0.0003* | 6.6475 | 0.0016* |
| | High | 11 (36.6) | - | - | | (0.4669- | | - | - | | (2.0466- | |
| | - | | | | | 29.6992) | | | | | 21.5919) | |

*Statistically significant difference DFS – disease-free survival; OS – overall survival

DISCUSSION

Despite the better understanding of the mechanisms responsible for melanoma progression in recent years, the five-year survival rate of melanoma remains less than 25% throughout the world [1,6,10]. Particularly, mucosal melanomas represent a highly aggressive subset of all melanomas and due their rarity, in the last decades, it has been observed slightly advance on understanding the molecular pathways of interest in these tumours [12-14]. In the present study, we evaluated the expression of COX-2 in oral melanomas and its association with clinicopathological factors and survival rate.

A higher expression of COX-2 was significantly related with presence of vascular, high mitotic index and neural invasion. Previous studies have demonstrated a positive correlation between COX-2 expression with cellular proliferation and angiogenesis, since inflammation provides a conducive environment for tumour progression [3,4]. These factors are known to determine tumour progression and these results are consistent with previous literature which showed that COX-2 overexpression is associated with advanced disease [15].

The Cox multivariate model demonstrated that COX-2 expression was an independent prognostic factor for overall survival, evidencing the most significant result of our study. Previously, de Souza do Nascimento and collaborators showed COX-2 is a useful marker to differentiate oral nevi and oral melanoma [2]. In addition, Panza *et al.* [1] observed that high COX-2 expression is a negative prognostic factor in lymph node metastatic melanoma. To the best of our knowledge, this is the first study to show that COX-2 expression is associated with OS in oral melanoma. Our findings propose a useful tool to argue for a possible therapeutic use of prophylactic anti-inflammatory use in the treatment of oral melanoma [16-19]. It has been recognized a synergic effect of COX-2 inhibitors and chemotherapy and/or radiotherapy in some molecular targets, such as tumour growth and proliferation, including VEGFR or aromatase inhibitors [20-23].

In conclusion, we suggest COX-2 as valuable immunohistochemical prognostic marker for oral melanoma. Future studies addressing the molecular signals that regulate COX-2 in oral melanoma are necessary to evaluate the effect of novel therapeutic agents in the modulation of cell immune responses.

REFERENCES

[1] Panza E, De Cicco P, Ercolano G, Armogida C, Scognamiglio G, Anniciello AM, et al. Differential expression of cyclooxygenase-2 in metastatic melanoma affects progression free survival. Oncotarget. 2016 Aug 30;7(35):57077-57085.

[2] de Souza do Nascimento J, Carlos R, Delgado-Azañero W, Mosqueda Taylor A, de Almeida OP, Romañach MJ et al. Immunohistochemical expression of cyclooxygenase-2 (COX-2) in oral nevi and melanoma. J Oral Pathol Med. 2016 Jul;45(6):440-3.

[3] Xia M, Duan ML, Tong JH, Xu JG. MiR-26b suppresses tumor cell proliferation, migration and invasion by directly targeting COX-2 in lung cancer. Eur Rev Med Pharmacol Sci. 2015 Dec;19(24):4728-37.

[4] Hu H, Han T, Zhuo M, Wu LL, Yuan C, Wu L, et al. Elevated COX-2 Expression Promotes Angiogenesis Through EGFR/p38-MAPK/Sp1-Dependent Signalling in Pancreatic Cancer. Sci Rep. 2017 Mar 28;7(1):470.

[5] Guo Z, Jiang JH, Zhang J, Yang HJ, Yang FQ, Qi YP, et al. COX-2 Promotes Migration and Invasion by the Side Population of Cancer Stem Cell-Like Hepatocellular Carcinoma Cells. Medicine (Baltimore). 2015 Nov;94(44):e1806.

[6] Semaan J, Pinon A, Rioux B, Hassan L, Limami Y, Pouget C, et al. Resistance to 3-HTMC-Induced Apoptosis Through Activation of PI3K/Akt, MEK/ERK, and p38/COX-2/PGE2 Pathways in Human HT-29 and HCT116 Colorectal Cancer Cells. J Cell Biochem. 2016 Dec;117(12):2875-2885.

[7] Iacono D, Cinausero M, Gerratana L, Angione V, Scott CA, De Maglio, et al. Tumour-infiltrating lymphocytes, programmed death ligand 1 and cyclooxygenase-2 expression in skin melanoma of elderly patients: clinicopathological correlations. Melanoma Res. 2018 Dec;28(6):547-554.

[8] Soares CD, Borges CF, Sena-Filho M, Almeida OP, Stelini RF, Cintra ML, et al. Prognostic significance of cyclooxygenase 2 and phosphorylated Akt1 overexpression in primary nonmetastatic and metastatic cutaneous melanomas. Melanoma Res. 2017 Oct;27(5):448-456.

[9] Williams MD. Update from the 4th Edition of the World Health Organization Classification of Head and Neck Tumours: Mucosal Melanomas. Head Neck Pathol. 2017 Mar;11(1):110-117.

[10] Gershenwald JE, Scolyer RA. Melanoma Staging: American Joint Committee on Cancer (AJCC) 8th Edition and Beyond. Ann Surg Oncol. 2018 Aug;25(8):2105-2110.

[11] Soares CD, Morais TML, Carlos R, de Almeida OP, Mariano FV, Altemani A, et al. Prognostic importance of mitochondrial markers in mucosal and cutaneous head and neck melanomas. Hum Pathol. 2019 Mar;85:279-289.

[12] Sun CZ, Chen YF, Jiang YE, Hu ZD, Yang AK, Song M. Treatment and prognosis of oral mucosal melanoma. Oral Oncol. 2012 Jul;48(7):647-52.

[13] Moya-Plana A, Mangin D, Dercle L, Taouachi R, Casiraghi O, Ammari S, et al. Risk-based stratification in head and neck mucosal melanoma. Oral Oncol. 2019 Aug 9;97:44-49.

[14] Shujiao L, Lilin H, Yong S. Cyclooxygenase-2 expression and association with skin cancer: A meta-analysis based on Chinese patients. J Cancer Res Ther. 2016 Dec;12(Supplement):C288-C290.

[15] Zhou P, Qin J, Li Y, Li G, Wang Y, Zhang N, et al. Combination therapy of PKCζ and COX-2 inhibitors synergistically suppress melanoma metastasis. J Exp Clin Cancer Res. 2017 Sep 2;36(1):115.

[16] Muraki C, Ohga N, Hida Y, Nishihara H, Kato Y, Tsuchiya K, et al. Cyclooxygenase-2 inhibition causes antiangiogenic effects on tumor endothelial and vascular progenitor cells. Int J Cancer. 2012 Jan 1;130(1):59-70.

[17] Soslow RA, Dannenberg AJ, Rush D, Woerner BM, Khan KN, Masferrer J, et al. COX-2 is expressed in human pulmonary, colonic, and mammary tumors. Cancer 2000;89:2637–2645.

[18] Sun RH, Gao P, Chen L, Ma DL, Wang JM, Oppenheim JJ, et al. Protein kinase C zeta is required for epidermal growth factor-induced chemotaxis of human breast cancer cells. Cancer Res. 2005;65:1433–41.

[19] Liao Z, Komaki R, Milas L, Yuan C, Kies M, Chang JY, et al. A phase I clinical trial of thoracic radiotherapy and concurrent celecoxib for patients with unfavorable performance status inoperable/unresectable non-small cell lung cancer. Clin Cancer Res. 2005;11:3342–3348.

[20] Penas-Prado M, Hess KR, Fisch MJ, Lagrone LW, Groves MD, Levin VA, et al. Randomized phase II adjuvant factorial study of dose-dense temozolomide alone and in combination with isotretinoin, celecoxib, and/or thalidomide for glioblastoma. Neuro Oncol. 2015;17:266–273.

[21] Zhang H, Li Z, Wang K. Combining sorafenib with celecoxib synergistically inhibits tumor growth of non-small cell lung cancer cells in vitro and in vivo. Oncol Rep. 2014;31:1954–1960.

[22] Ahmed M, Hussain AR, Siraj AK, Uddin S, Al-Sanea N, Al-Dayel F, et al. Cotargeting of cyclooxygenase-2 and FoxM1 is a viable strategy in inducing anticancer effects in colorectal cancer cells. Mol Cancer. 2015;14:131.

[23] Lustberg MB, Povoski SP, Zhao W, Ziegler RM, Sugimoto Y, Ruppert AS, et al. Phase II trial of neoadjuvant exemestane in combination with celecoxib in postmenopausal women who have breast cancer. Clin Breast Cancer. 2011;11:221–227.

2.3. Artigo: Oral amelanotic melanomas: clinicopathologic features of 8 cases and review of the literature

Ciro Dantas Soares¹, Román Carlos², Bruno Augusto Benevenuto de Andrade³, John Lennon Silva Cunha¹, Michelle Agostini³, Mário José Romañach³, Juan Carlos

Hernandez-Guerrero⁴, Adalberto Mosqueda-Taylor⁵, Oslei Paes de Almeida¹, Jacks

Jorge¹

- Department of Oral Diagnosis, Area of Pathology, Piracicaba Dental School, University of Campinas, Piracicaba, São Paulo, Brazil
- 2. Pathology Division, Centro Clínico de Cabeza y Cuello/Hospital Herrera Llerandi, Guatemala City, Guatemala
- 3. Oral Pathology, School of Dentistry, Federal University of Rio de Janeiro (UFRJ), Rio de Janeiro, Brazil
- Universidad Nacional Autónoma de México UNAM, Facultad de Odontología, Laboratory of Immunology, México City, México
- Health Care Department, Universidad Autónoma Metropolitana, Xochimilco, Mexico

Running tittle: A series of amelanotic oral melanomas.

Corresponding author: Ciro Dantas Soares; Department of Oral Diagnosis, Piracicaba Dental School, University of Campinas; Avenida Limeira, 901, Areião - 13414-903 Piracicaba/SP, Brazil; E-mail address: <u>ciro.dss@gmail.com</u>

Funding source: The authors acknowledge FAPESP for their support on this research (grants #2015/25905-1 and #2017/16802-8).

Competing Interests: The authors declare they have no conflict of interest.

Ethical Approval: The National Commission for Ethics in Research approved the study protocol (CONEP-Brazil, CAAE: 72077517.1.0000.5418).

ABSTRACT

Background: Mucosal melanomas are very aggressive tumors, rarely observed in the oral cavity. Usually, it is identified based on the clinical and microscopical features, including variable amounts of melanin-pigmentation. However, when melanin is absent, the tumors are denominated amelanotic, presenting a tendency to misdiagnosis and delayed treatment.

Objective: To describe the clinicopathological characteristics of a series of oral amelanotic melanomas (OAM).

Methods: Cases diagnosed as OAM were retrospectively retrieved from the files of four south and central America institutions from January 2002 to January 2019. Data on clinical features, morphological aspects, immunohistochemical reactions, treatment and follow-up status were also collected from the patient's medical records.

Results: The study group included six male and two females (ratio of 3:1) ranging in age from 33 to 77 years (mean 53.6 years). Clinically, masses or ulcerated swellings were reported. The most common intraoral locations of the tumors were gingiva and palate. Cervical lymph node metastasis was observed in three patients at the time of diagnosis. Three patients were treated with surgery plus chemotherapy, two with only surgery and one patient received adjuvant radiotherapy. All but one patient died from complications of the tumors after a mean follow-up period of 8.5 months. (ranging from 2 to 23 months).

Conclusion: OAM is a very aggressive malignant tumor, it demands especial concern to early diagnosis which may achieve successful treatment, improving the patient's prognosis. Similarly to occurs in the cutaneous melanomas, the oral amelanotic counterpart seems to have a different biology from the pigmented ones.

INTRODUCTION

Oral melanomas are extremely rare malignant melanocytic neoplasms, representing about 2–8% of all melanomas [1,2]. Although the microscopic recognition of melanomas is usually possible by the combination of cytomorphological and architectural features, its diagnosis remains a challenge, especially in amelanotic or hypopigmented cases [3]. Amelanotic melanomas are exceedingly rare and can, clinically and microscopically, resemble other cancer types or even mimic benign tumors [1,4].

Recent evidences have demonstrated that cutaneous amelanotic melanomas have poorer prognosis than pigmented tumors [5]. However, few retrospective cohort studies have been carried out to evaluate the clinicopathological features of oral amelanotic melanomas (OAM). The aim of the present study was to describe the clinical, histological and immunoprofile features of a series of 8 OAM.

PATIENTS AND METHODS

Cases diagnosed as oral amelanotic melanomas were retrieved from the files of 2 Brazilian, 1 Mexican and 1 Guatemalan oral pathology services. Clinical data were retrieved from the pathology reports and follow-up information was obtained from the referring physicians. Five-micrometer hematoxylin and eosin–stained sections were reviewed for histological description of the lesions. Immunohistochemical reactions were performed in 3- μ m tissue sections in silanized slides, using standard protocols for the following antibodies: S-100 (polyclonal, dilution 1:10.000), HMB-45 (clone HMB-45, dilution 1:200), Melan A (clone A103, dilution 1:800), pan-Cytokeratin (clone AE1/AE3, dilution 1:400), Vimentin (clone Vim 3B4, dilution 1:400), α -SMA (clone 1A4, dilution 1:400), CD45 LCA (clone 2B11+PD7/26, dilution 1:200), and Ki-67 (clone MIB-1, dilution 1:100). All antibodies were obtained from Dako (Glostrup, Denmark).

This study was carried out in accordance with the Helsinki Declaration of 1964 and was approved by the Piracicaba Dental School ethics board (#72077517.1.0000.5418).

RESULTS

Clinical features

The patients included 6 men and 2 women, with a mean age at presentation of 53.6 years (range 33–77 years). The patients were white (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2), black (n = 5) or unknown (n = 2) or unkno

= 1) race. The locations of the tumors were upper and lower gingiva (n=3), tongue (n=1), lip (n=1), upper alveolar ridge (n=1), and palate (n=2). The tumors presented as reddish swellings or ulcerations and lacked brown or dark pigmentation in oral cavity. One patient complained of bleeding from the lesion, and three patients (37.5%) had lymph node involvement or metastasis at the time of diagnosis (Table 1).

| Case | Age (years) | Gender | Location | Treatment | Follow-up | Survival time (months) | Lymph node involvement |
|------|----------------|--------|-------------------|-----------|--------------|------------------------------|------------------------------|
| 1 | 34 | М | Soft palate | SX+CX+RX | died | 2 | Yes |
| 2 | 77 | М | Tongue | SX+CX | died | 11 | No |
| 3 | 56 | М | Lip | SX | disease-free | 23 | No |
| 4 | 55 | М | Inferior Gingiva | SX | died | 5 | No |
| 5 | 52 | F | Superior Gingiva | SX+CX | died | 8 | Yes |
| 6 | 33 | F | Inferior Gingiva | SX+CX | died | 2 | No |
| 7 | 68 | М | Superior alveolar | N/A | lost | N/A | N/A |
| | | | ridge | | | | |
| 8 | 54 | М | Palate | N/A | lost | N/A | Yes |

Table 1. Summary of clinical features of 8 cases of oral amelanotic melanomas.

M – male, F – Female, SX – surgery, CX – chemotherapy, RX – radiotherapy, N/A – Not available.

Clinically, case 1 presented as a diffuse and painless asymptomatic swelling (approximately 3.0 cm of size) covered by erythematous lining mucosa. The tumor extended to the oropharynx region and caused difficulty in swallowing and phonation. In addition, extraoral clinical examination showed ipsilateral nodal metastasis at the time of diagnosis. Case 6 presented as a small asymptomatic sessile nodule, 0.5 cm, bleeding on palpation, in the region of the incisive papilla between lower central incisors with a clinical diagnosis of pyogenic granuloma. The case 7 presented as an asymptomatic swelling showing areas of ulceration in the upper left alveolar ridge. A panoramic radiograph showed slight erosion and alteration of the trabecular bone of the affected region. The patient reported being a smoker and a chronic alcoholic (Fig. 1).



Figure 1. A: Clinical presentation of the case 1, a diffuse and painless asymptomatic swelling (approximately 3.0 cm of size) covered by erythematous lining mucosa. The tumor extended to the oropharynx region and caused difficulty in swallowing and phonation. B: In this case 6, the patient presented a small asymptomatic sessile nodule, 0.5 cm, bleeding on palpation, in the region of the incisive papilla between lower central incisors with a clinical diagnosis of pyogenic granuloma. C: In the case 7, the patient presented an asymptomatic swelling showing areas of ulceration in the upper left alveolar ridge.

Differential clinical diagnoses for all lesions included: infectious diseases, malignant neoplasms such as squamous cell carcinoma, lymphomas and others malignant tumors. For the incisive papilla lesion, periodontal disease and pyogenic granuloma were raised. In none of the 8 cases the dentist specifically mentioned melanoma as part of the clinical differential diagnosis. Some cases that had the

diagnosis of benign lesions were completely excised; whereas cases with clinical appearance of malignant lesions were submitted to incisional biopsy.

Pathologic features

Histologically all cases demonstrated a diffuse proliferation of pleomorphic rhabdoid, spindle and epithelioid melanocytes with a variable number of mitosis and absent or very focal melanin deposition (Figs. 2 and 3). In two cases, undifferentiated small blue round cells were observed. In addition, nuclear inclusions and stromal desmoplasia areas were also observed in one case. Large central amphophilic or eosinophilic nucleoli were observed in 5 cases. Lymphomas, carcinomas, and sarcomas not otherwise specified were considered as the differential diagnosis. Regarding cell morphology, 3 (37.5%) presented a predominance of undifferentiated cells, 3 (37.5%) epithelial/rhabdoid and 2 (25.0%) with predominant spindle cell morphology (Table 2). Morphological features of the cases 5 and 7 are illustrated in the Figure 2 and 3, respectively.

 Table 2. Microscopical and immunohistochemical features of 8 oral amelanotic melanomas.

| Case | Predominant morphology | Vim | AE1/AE3 | LCA | a-SMA | S-100 | Melan A | HMB-45 | Ki-67 |
|------|---------------------------|-----|---------|-----|-------|-------|----------|--------|-------|
| 1 | Undifferentiated | + | neg | neg | neg | + | weak/neg | + | >70% |
| 2 | Epithelioid | + | neg | neg | neg | + | + | + | >70% |
| 3 | Spindle cell | + | neg | neg | neg | + | + | + | 22.1% |
| 4 | Undifferentiated | + | neg | neg | neg | + | weak/neg | + | >70% |
| 5 | Epithelioid | + | neg | neg | neg | + | + | + | 34.2% |
| 6 | Spindle cell | + | neg | neg | neg | + | + | + | >90% |
| 7 | Undifferentiated | + | neg | neg | neg | + | + strong | + | >70% |
| 8 | Epithelioid | + | neg | neg | neg | + | + | + | 51.4% |

Vim – vimentin, AE1/AE3 – pan-cytokeratin, LCA – leucocyte common antigen, α -SMA α -smooth muscle actin, neg – negative



Figure 2. A: Microscopic findings of the case 7, demonstrating a proliferation of spindle and epithelioid melanocytes with a variable number of mitosis and absent or very focal melanin deposition. B: Areas of necrosis and vascular invasion were also observed. The IHC results showed intense immunoreactivity for S-100 (C), MelanA (D), HMB45 (E) and high-proliferative index demonstrated by nuclear Ki67 expression (F).



Figure 3. Morphological features of the case 7. A: The tumor was composed predominantly by epithelioid cells arranged in nests; the nuclei were hypercromatic. B: A perivascular pattern was observed in several areas. The IHC results showed intense immunoreactivity for MelanA (C), HMB45 (D), S-100 (E), and very high-proliferative index demonstrated by nuclear Ki67 expression.

Immunohistochemical analysis was subsequently performed for confirmation of the phenotype of tumor cells (Fig. 2 and 3). The results of the immunohistochemical analysis of the evaluated cases are presented in Table 2. The tumor cells were strongly and diffusely positive in all cases for vimentin, protein S-100, and HMB-45. Melan A was considered as focal/weak positive in two cases. In addition, all the cases were negative for smooth muscle actin (α -SMA), pan-cytokeratin AE1/AE3, and anti-leukocyte common antigen (CD45). Ki67-positivity index was >70% in three cases and the case 6 achieved more than 90%.

The patients were treated with surgery and/or chemotherapy (paclitaxel and cisplatin). One of them remains disease-free, 5 died and 2 lost follow-ups (Table 1).

| Case | Author, year | Age | Gender | Oral Location | Predominant morphology | Immunohistochemical studies | Cervical node metastases | Treatment | Survival from presentation (months) |
|------|-----------------------|-----|--------|-------------------|--------------------------------|-----------------------------------------------------------------------------------|--------------------------------|---------------------------------------------------------------------------------------|----------------------------------------------|
| 1 | Saku, 1983 | 59 | F | Palate | Desmoplastic | N/A | N/A | N/A | N/A |
| 2 | Kumar, 2013 | 50 | F | Tongue | Spindle cells/undifferentiated | CK –, VIM –, desmin–, HMB45+ | No | Sx, Rtx | DOD, 6 |
| 3 | Chu, 1993 | 79 | F | Upper gingiva | Spindle cells/undifferentiated | S100+ | Yes | N/A | N/A |
| 4 | Boyd, 2011 | 25 | М | Upper gingiva | Small round blue cells | CK-, CD45-, MelanA +, HMB45+,. CD99-, CD138- | Yes | Sx: Chemoterapy (carboplatin, paclitaxel, and pegfilgrastim) and Rtx | DOD, 13 |
| 5 | Cicconetti, 2009 | 65 | F | Upper gingiva | Spindle cells | CK–,. S-100+, HMB45+ | N/A | Sx, Rtx | AWD, 36 |
| 6 | Pandiar, 2013 | 50 | М | Lower gingiva | Epithelioid/rhabdoid | VIM+, Desmin+, S-100+, Melan-A +, HMB45+, CD68 -, enolase -, myoglobin - | No | Sx, Rtx | N/A |
| 7 | Ducic, 2001 | 68 | F | Soft palate | Spindle cells | HMB45+,. S-100+ | No | Sx | AWD, 12 |
| 8 | Godoy, 2009 | 42 | М | Lower lip | Spindle cells | S100+,. HMB45+, CK- | Yes | Sx, Rtx | AWD, N/A |
| 9 | Saghravanian, 2014 | 27 | М | Upper gingiva | Epithelioid | S-100+ and HMB45+, LCA–, CD68–, and Desmin– | No | Sx, ChT, | N/A |
| 10 | Kao, 2001 | 80 | М | Palate | Undifferentiated/epithelioid | CK-, S100+, HMB45+ | No | No treatment | N/A |
| 11 | Venugopal, 2013 | 19 | М | Tongue | Epithelioid/spindle cells | Desmin–, VIM+, EMA–, CD34–, HMB45+ | Yes | N/A | N/A |
| 12 | Moshe, 2018 | 58 | F | Lower gingiva | N/A | N/A | Yes | N/A | DOD, 6 |
| 13 | Notani, 2002 | 67 | М | Upper gingiva | Spindle cells | S100+, HMB45+ | Yes | Sx, ImT (dacarbazine, 1- methyl-3-nitrosourea hydrochloride and vincristine) | DOD, 25 |
| 14 | Notani, 2002 | 78 | F | Palate | Spindle cells | S100+, HMB45+ | Yes | Sx | DOD, 15 |
| 15 | Notani, 2002 | 69 | М | Retromolar region | Epithelioid | S100+, HMB45+ | Yes | Sx, chemotherapy (cisplatin, dacarbazine | AWD, 27 |

 Table 3. Summary of published cases of amelanotic melanomas affecting the oral cavit

| | | | | | | | | and vindesine) | |
|----|---------------|----|---|------------------|---------------------------|---------------------------------------------|-----|-----------------------------------------------------------------------------------------------------------------------------|---------|
| 16 | Ohnishi, 2015 | 80 | М | Upper gingiva | Epithelioid/Spindle cells | S100+ /.HMB45+ / MelanA + | No | Surgery, ChT: dacarbazine, nimustine and vincristine, and local intracutaneous interferon β therapy (DAV-Feron) | N/A |
| 17 | Paulo, 2015 | 33 | F | Lower gingiva | Epithelioid | S100 + / Vimentina +/ HMB45 + / MelanA + | Yes | Sx, ChT | AwD, 60 |
| 18 | Tanaka, 2004 | 82 | F | Upper gingiva | N/A | S100+, HMB45+ | Yes | Rx | DOD, 3 |
| 19 | Tanaka, 2004 | 46 | М | Upper gingiva | N/A | S100+, HMB45+ | Yes | Rtx, ChT, ImT | DOD, 8 |
| 20 | Tanaka, 2004 | 71 | F | Upper gingiva | N/A | S100+, HMB45+ | Yes | Sx, ChT, ImT | DOD, 49 |
| 21 | Tanaka, 2004 | 70 | F | Upper gingiva | N/A | S100+, HMB45+ | No | Sx, ChT, ImT | DOD, 9 |
| 22 | Tanaka, 2004 | 84 | F | Upper gingiva | N/A | S100+, HMB45+ | No | Sx, Rtx | DOD, 29 |

ChT chemotherapy, Sx Surgery, Rtx Radiotherapy, ImT immunotherapy, AWD alive with disease, AwD alive without disease, DOD died of disease, N/A not available

DISCUSSION

Oral amelanotic melanomas (OAM) are exceedingly rare with less than 35 cases reported in the English literature to date [6–8]. In this study, we present the first large series of eight patients with OAM. Most were male (3:1) and older than 50 years (75%). Four patients presented with relatively advanced disease and three (37.5%) patients had lymph node involvement or metastatic disease at the time of diagnosis. The male predominance in our patients with OAM agrees with several studies [1,3,9]; however, others found a higher predilection of OAM in females [10–12] or no sex predilection [13]. We reviewed 22 cases of OAM with clinical information available. Table 3 lists the demographic and clinical information for these cases. The table does not include eight cases reported by Bachar et al. (2008)[8] and Andrade (2012)[4] because these were series of patients with melanotic and amelanotic oral melanomas and did not have demographic and clinical information about the specific cases. From the data of the reviewed literature, most patients presented neck lymphadenopathy at the time of diagnosis (54.5%), and the majority of the cases occurred in gingiva (59.1%), with only 1 case occurred in soft palate.

Mucosal melanomas of the head and neck region are very aggressive tumors and often are diagnosed at advanced stages [14]. Its occurrence in oral cavity is characterized by detection of pigmented swelling or ulcerated lesion; and microscopically, a proliferation of atypical melanocytes with variable melanin deposition help to establish the diagnosis [4,15]. However, amelanotic tumors present a challenge to clinical and microscopical diagnosis. In fact, the absence of melanin pigmentation makes the clinical diagnosis of these tumors extremely challenging due to the great clinical similarity with several other conditions that affects oral cavity, including reaction lesions, non-neoplastic proliferative processes, infectious diseases, and other oral malignancies. Indeed, microscopically, the difficulty in identifying this differential diagnosis lies in the high variable sarcomatous-like presentation of this lesion, since melanoma can mimic rhabdomyosarcoma, spindle cell carcinoma, lymphomas, poorly differentiated carcinomas, neuroendocrine tumors and poorly differentiated sarcomas. When melanin pigmentation is present, melanomas usually can be diagnosed by morphological analysis. However, when the lesion is amelanotic, the immunohistochemistry analysis is an essential tool for confirming the tumor phenotype and establishing the correct diagnosis. In both situations, positive immunohistochemical reactions for S-100, HMB-45, and Melan-A confirm the diagnosis.

As mentioned above, the differential diagnosis of amelanotic melanomas is very broad because its morphological appearance includes different cell types including epithelioid, spindle and plasmacytoid morphology [16,17]. In our series undifferentiated/epithelioid cells predominated in most cases. Cases with fusiform and rhabdoid predominant morphology were less common. In addition, some other tumors presented a very polymorphous morphology. An extensive IHC panel with markers of neural, neuroendocrine, lymphoid or epithelioid differentiation may be done to rule out other tumors. Neural and vascular invasion were commonly observed in most cases.

Mucosal melanomas frequently show a very diverse morphology, that include fusiform, epithelioid, rhabdoid or even undifferentiated cells. Previous studies have demonstrated no correlation of the type cell with prediction of tumor behavior [18,19]. Nevertheless, some studies have indicated a poor prognosis for tumors with epithelioid morphology [20]. Epithelioid cell type and undifferentiated morphology was markedly prevalent in the amelanotic tumors. These factors may explain the aggressiveness of these tumors; however, other studies can confirm these early observations. According to the review of the literature, epithelioid and spindle cells morphology are the most common presentation for OAM (more than 60%). Rhabdoid and undifferentiated morphology is reported in few cases, representing about less than 9% of all cases.

Similarly to the conventional oral melanoma, OAM presented as masses and/or ulcerated lesions in the gingiva and palate. Only two tumors occurred in other locations, including tongue and lip. The case that occurred in lip presented a non-aggressive course, and the patient remains disease-free 23 months after the surgery. In fact, the intraoral tumors usually show an aggressive behavior probably associated with a very worse prognosis.

Another interesting finding that deserve commentaries is the clinical presentation of OAM in gingiva. These particular cases have clinical diagnosis presumptive of pyogenic granuloma or even other reactive processes. Other series have been reported the possibility of melanomas mimic pyogenic granulomas [1,21]. These factors can be influence in the delayed diagnosis and reinforce the idea that even lesions with a benign clinical presentation must be examined microscopically.

Prasad et al.[2] proposed a microstaging system based in the depth of histologic invasion, considering: level I as exclusive melanoma in situ (very rare for mucosal melanomas), level II as invasion into lamina propria only and level III as deep tissue invasion into adjacent structures including skeletal muscle, bone, or cartilage. All cases

were classified as II/III Prasad-based staging. Our results reinforce the reproducibly of this classification and also confirm that AOM are tumors with particular aggressiveness, indicating a worse outcome for these patients. Several therapeutic options were used to treat mucosal melanomas, including surgery associated or not with chemotherapy/radiotherapy [22]. As particular observation of the authors, it is possible to affirm the management of these tumors remains a challenge and the different therapeutic approach highlight the necessity of standard protocols for oral melanomas.

The prognosis of OAM seems to be extremely poor which may be largely due to the advanced stage of disease at the time of clinical presentation, associated with difficult in stablish an accurate diagnosis [23,24]. Based on previous literature cutaneous amelanotic melanomas have poor prognosis than pigmented ones [5]. So far, only a few cases were reported concerning OAM. In our study, 5 of 8 patients died few months after the first consultation, indicating that oral amelanotic melanomas may have a worse prognosis when compared with the pigmented ones. Overall, the 20% 1-year survival rate in our cohort of OAM (calculated only for patients with follow-up information available) demonstrated the highly aggressive outcome of these tumors that agree with previous report. However, our small sample size did not allow a strong conclusion about this point. Extrapolating from the patients reported in the literature, 45.4% of the patients died with local or disseminated disease (n = 10 of 22 with data), an average of 21.1 months after initial melanoma diagnosis, with 18.2 % of patients alive with local disease (n = 4; mean 25 months follow-up). By contrast, 4.5 % of patients were alive or had died without evidence of disease (n = 1 of 22 with data), with an average follow-up of 27 months. Data were not available for 7 patients.

In conclusion, this study represents one of the largest series of cases of OAM, which seems to exhibit a biological behavior different from the pigmented tumors with metastatic foci present at time of diagnosis. OAM may mimic both clinically and microscopically several benign and malignant asymptomatic conditions of oral cavity, and therefore, immunohistochemical analysis is essential for proper diagnosis, particularly for S-100, HMB-45 and Melan-A. Comparative studies may help to determine specific pathways involved in the pathogenesis of these tumors, as well as to find biological markers to development of targeted therapies.

According to the results of the present cases and previous reports, it is reasonable to suspect that amelanotic melanomas will have an overall poor prognosis because of the aggressiveness of the tumor coupled with a delay in establishing a correct diagnosis.

Early diagnosis by histological examination together with immunochemistry are the keys to improving the survival for patients with oral amelanotic melanoma.

REFERENCES

[1] Moshe M, Levi A, Ad-El D, Ben-Amitai D, Mimouni D, Didkovsky E, et al. Malignant melanoma clinically mimicking pyogenic granuloma: comparison of clinical evaluation and histopathology. Melanoma Res 2018;28:363–7. https://doi.org/10.1097/CMR.00000000000451.

[2] Prasad ML, Patel SG, Huvos AG, Shah JP, Busam KJ. Primary Mucosal Melanoma of the Head and Neck: A Proposal for Microstaging Localized, Stage I (Lymph Node-Negative) Tumors. Cancer 2004;100:1657–64. https://doi.org/10.1002/cncr.20201.

[3] Cheung WL, Patel RR, Leonard A, Firoz B, Meehan SA. Amelanotic melanoma: a detailed morphologic analysis with clinicopathologic correlation of 75 cases. J Cutan Pathol 2012;39:33–9. https://doi.org/10.1111/j.1600-0560.2011.01808.x.

[4] de-Andrade B-A-B, Toral-Rizo V-H, León J-E, Contreras E, Carlos R, Delgado-Azañero W, et al. Primary oral melanoma: a histopathological and immunohistochemical study of 22 cases of Latin America. Med Oral Patol Oral Cir Bucal 2012;17:e383-8. https://doi.org/10.4317/MEDORAL.17588.

[5] Strazzulla LC, Li X, Zhu K, Okhovat JP, Lee SJ, Kim CC. Clinicopathologic, misdiagnosis, and survival differences between clinically amelanotic melanomas and pigmented melanomas. J Am Acad Dermatol 2019;80:1292–8. https://doi.org/10.1016/j.jaad.2019.01.012.

[6] Tanaka N, Mimura M, Kimijima Y, Amagasa T. Clinical investigation of amelanotic malignant melanoma in the oral region. J Oral Maxillofac Surg 2004;62:933–7. https://doi.org/10.1016/j.joms.2004.01.017.

[7] Notani K, Shindoh M, Yamazaki Y, Nakamura H, Watanabe M, Kogoh T, et al. Amelanotic malignant melanomas of the oral mucosa. Br J Oral Maxillofac Surg 2002;40:195–200. https://doi.org/10.1054/bjom.2001.0713.

[8] Bachar G, Kwok SL, O'Sullivan B, Goldstein D, Wood S, Brown D, et al. Mucosal melanomas of the head and neck: The Princess Margaret Hospital experience. Head Neck 2008;30:1325–31. https://doi.org/10.1002/hed.20878. [9] Moreau JF, Weissfeld JL, Ferris LK. Characteristics and survival of patients with invasive amelanotic melanoma in the USA. Melanoma Res 2013;23:408–13. https://doi.org/10.1097/CMR.0b013e32836410fe.

[10] McClain SE, Mayo KB, Shada AL, Smolkin ME, Patterson JW, Slingluff CL. Amelanotic melanomas presenting as red skin lesions: a diagnostic challenge with potentially lethal consequences. Int J Dermatol 2012;51:420–6. https://doi.org/10.1111/j.1365-4632.2011.05066.x.

[11] Thomas NE, Kricker A, Waxweiler WT, Dillon PM, Busam KJ, From L, et al. Comparison of Clinicopathologic Features and Survival of Histopathologically Amelanotic and Pigmented Melanomas. JAMA Dermatology 2014;150:1306. https://doi.org/10.1001/jamadermatol.2014.1348.

[12] Huvos AG, Shah JP, Goldsmith HS. A clinicopathologic study of amelanotic melanoma. Surg Gynecol Obstet 1972;135:917–20. https://doi.org/10.1097/00006534-197306000-00052.

[13] Giuliano AE, Cochran AJ, Morton DL. Melanoma from unknown primary site and amelanotic melanoma. Semin Oncol 1982;9:442–7.

[14] Thompson LDR, Wieneke JA, Miettinen M. Sinonasal tract and nasopharyngeal melanomas: a clinicopathologic study of 115 cases with a proposed staging system. Am J Surg Pathol 2003;27:594–611.

[15] Lourenço S V., Fernandes JD, Hsieh R, Coutinho-Camillo CM, Bologna S, Sangueza M, et al. Head and neck mucosal melanoma: A review. Am J Dermatopathol 2014;36:578–87. https://doi.org/10.1097/DAD.00000000000035.

[16] Benevenuto de Andrade BA, Piña AR, León JE, Paes de Almeida O, Altemani A. Primary nasal mucosal melanoma in Brazil: clinicopathologic and immunohistochemical study of 12 patients. Ann Diagn Pathol 2012;16:344–9. https://doi.org/10.1016/j.anndiagpath.2012.02.001.

[17] Mendenhall WM, Amdur RJ, Hinerman RW, Werning JW, Villaret DB, Mendenhall NP. Head and neck mucosal melanoma. Am J Clin Oncol 2005;28:626–30. https://doi.org/10.1097/01.coc.0000170805.14058.d3.

[18] Barker BF, Carpenter WM, Daniels TE, Kahn MA, Leider AS, Lozada-Nur F, et al. Oral mucosal melanomas: the WESTOP Banff workshop proceedings. Western Society of Teachers of Oral Pathology. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1997;83:672–9. https://doi.org/10.1016/s1079-2104(97)90318-8.

[19] Lourenço SV, Martin Sangüeza A, Sotto MN, Bologna SB, Giacomo TB di,
Buim ME, et al. Primary oral mucosal melanoma: A series of 35 new cases from south
America. Am J Dermatopathol 2009;31:323–30.
https://doi.org/10.1097/DAD.0b013e3181a0d37c.

[20] Soares C, De Lima Morais TM, Carlos R, Mariano FV, Altemani A, De Carvalho MGF, et al. Phosphorylated Akt1 expression is associated with poor prognosis in cutaneous, oral and sinonasal melanomas. Oncotarget 2018;9:37291–304. https://doi.org/10.18632/oncotarget.26458.

[21] Zaballos P, Rodero J, Serrano P, Cuellar F, Guionnet N, Vives JM. Pyogenic granuloma clinically and dermoscopically mimicking pigmented melanoma. Dermatol Online J 2009;15:10.

[22] Moya-Plana A, Aupérin A, Obongo R, Baglin AC, Ferrand FR, Baujat B, et al. Oncologic outcomes, prognostic factor analysis and therapeutic algorithm evaluation of head and neck mucosal melanomas in France. Eur J Cancer 2019;123:1–10. https://doi.org/10.1016/j.ejca.2019.09.007.

[23] Kumar V, Shukla M, Goud U, Ravi DK, Kumar M, Pandey M. Spindle Cell Amelanotic Lesion of the Tongue: a Diagnostic and Therapeutic Challenge. Indian J Surg 2013;75:394–7. https://doi.org/10.1007/s12262-012-0575-8.

[24] Boyd BC, Au J, Aguirre A, Votta TJ. Rapidly enlarging nodular lesion of the anterior maxilla. Oral Surgery, Oral Med Oral Pathol Oral Radiol Endodontology 2011;112:626–31. https://doi.org/10.1016/j.tripleo.2011.06.033.

2.4. Artigo: Comparative expression of cyclooxygenase 2 and Ki67 in amelanotic and conventional oral melanomas

Ciro Dantas Soares¹, Juan Carlos Hernandez-Guerrero², Bruno Augusto Benevenuto de Andrade³, Mario Romanach³, Adalberto Mosqueda-Taylor⁴, Román Carlos⁵, Madja Ruanna Soares Macedo⁶, Oslei Paes de Almeida¹, Jacks Jorge¹

- Department of Oral Diagnosis, Area of Pathology, Piracicaba Dental School, University of Campinas, Piracicaba, São Paulo, Brazil.
- 2. Immunology Laboratory, Faculty of Dentistry, National University Autonomous of Mexico
- Oral Pathology, School of Dentistry, Federal University of Rio de Janeiro (UFRJ), Rio de Janeiro, Brazil
- Health Care Department, Universidad Autónoma Metropolitana, Xochimilco, Mexico
- 5. Pathology Division, Centro Clínico de Cabeza y Cuello/Hospital Herrera Llerandi, Guatemala City, Guatemala.
- Department of Nursing, Federal University of Rio Grande do Norte, Natal, Rio Grande do Norte, Brazil.

Running tittle: COX-2 and Ki67 in amelanotic oral melanoma.

Corresponding author: Ciro Dantas Soares; Department of Oral Diagnosis, Piracicaba Dental School, University of Campinas; Avenida Limeira, 901, Areião - 13414-903 Piracicaba/SP, Brazil; E-mail address: <u>ciro.dss@gmail.com</u>

Funding source: The authors acknowledge FAPESP for their support on this research (grants #2015/25905-1 and #2017/16802-8).

Competing Interests: The authors declare they have no conflict of interest.

Ethical Approval: The National Commission for Ethics in Research approved the study protocol (CONEP-Brazil, CAAE: 72077517.1.0000.5418).

Abstract

Background: Oral melanomas have some histopathological resemblance with its cutaneous counterpart; however, an aggressive behavior is more common in tumors that occur in the oral cavity. Several markers have been suggested as indicative of tumoral progression and aggressiveness, such as cyclooxygenase 2 (COX-2) and Ki67.

Methods: In this study, we have compared the expression of COX-2 and Ki67 in a series of amelanotic (n=7) and melanotic oral melanomas (n=22). The cases were selected from 4 pathology laboratories and submitted to the immunohistochemical (IHC) reactions. We analyzed the IHC staining based on a qualitative – using visual scores; and a computer-assisted method (quantitative) using scanned slides and software for digital analysis.

Results: COX-2 was expressed in all oral melanomas; however, its intensity was significantly higher in the amelanotic ones (P<0.001). Similarly, a high Ki67-positivity index was observed in the amelanotic than melanotic ones (P<0.001).

Conclusion: Based on these results, we suggest that amelanotic oral melanomas have marked pro-inflammatory and high-proliferative phenotype, justifying their more aggressive behavior compared with the melanotic ones.

Keywords: oral melanoma, amelanotic tumor, cyclooxygenase 2, Ki67

Introduction

Cutaneous and mucosal melanomas origin from cells derivate of the neural crest, the melanocytes.¹ Clinically and microscopically, some tumors do not produce melanin, a brownish pigment that characterize these cells, and are named amelanotic tumors.² In the cutaneous counterpart, the absence of melanin is considered a sign of aggressiveness.³ However, in the tumors of the oral cavity, the significance of this biological event is poorly understood.

Cyclooxygenase 2 (COX-2) is an enzyme involved in several inflammatory pathways. Its main function is the conversion of arachidonic acid to prostaglandin.⁴ COX-2 mediate several cellular functions, and in the context of the cancer, its higher expression is associated with modulation of angiogenesis, cellular migration, invasion and proliferation, as well as apoptotic resistance, events that favors tumor progression.⁵ In fact, COX-2 is associated with aggressiveness in non-small cell lung cancer⁶, pancreatic ductal adenocarcinoma⁷, hepatocellular carcinoma⁸ and colorectal cancer.⁹

Previous studies of our group have demonstrated that COX-2 expression is associated with angiogenesis in ameloblastoma¹⁰ and cellular proliferation in cutaneous melanomas.¹¹ However, to date, comparative studies of oral melanotic and amelanotic melanomas are not available in the English language literature. The aim of the present study was to evaluate the immunohistochemical expression of COX-2 and Ki67 in oral melanotic and amelanotic melanomas.

Material and Methods

Formalin-fixed, paraffin-embedded tissue blocks and clinical information of 29 oral melanomas (7 amelanotic and 22 conventional melanotic) were obtained from the charts of four oral pathology laboratories from Latin America (Guatemala, Mexico and 2 from Brazil). Hematoxylin and eosin–stained slides were used for reviewing all diagnoses, and difficult cases were submitted to immunohistochemical analysis (HMB-45, S-100 and MelanA). To classify the melanomas selected in this study, we used a method proposed by Prasad et al.¹².

Accordingly laboratory, for previous protocols used in our immunohistochemical staining, 3-µm-thick sections were deparaffinized, rehydrated in graded ethanol solutions and after antigen retrieval with EDTA/Tris buffer (pH 9.0) in a pressure cooker; endogenous peroxidase activity was blocked with 20% H2O2 during 15 min. We used a method of 2-hours incubation with the primary antibodies: COX-2 (Clone: CX-294, dilution 1:100; Dako Corporation, Carpinteria, CA, USA) and Ki67 (Clone: MIB-1, dilution 1:100; Dako Corporation). Both diluted in BSA (bovine serum albumin). The secondary antibody conjugated with polymer dextran marked with peroxidase (Dako EnVision Labelled Polymer; Dako, Glostrup, Denmark) was applied for 1 hour. The reaction was developed with Permanent Red (Permanent Red Substrate System; Dako) and counterstained with Carazzi hematoxylin. Sections of oral squamous cell carcinoma were included in all reactions as positive control for both markers. Negative controls of reactions were performed by omitting the primary antibody. Only cytoplasmic staining was considered as positive for COX-2, and only nuclear reactivity was considered for Ki67. The quantification of the immunoreactivity was determined using a digital method previously validated by our group.^{10,13}

Results

Amelanotic melanomas were more common in men (5 males, 2 females), with a mean age of 51.5 years. The most common location was gingiva (3 cases), followed by palate (2 cases) and tongue and lip with one case. Five patients died from complications of the disease, 1 lost follow-up and 1 is disease-free. All but one patient was treated with surgery and complementary chemotherapy.

The conventional melanotic melanomas occurred predominantly in women (14 females, 8 males), with a mean age of 58 years. The locations were palate (n=14), gingiva (n=4) and other anatomical sites (n=4). Regarding treatment, 15 were treated with only surgery and 7 received complementary chemotherapy. Twelve patients died from complications of the tumor and 10 are alive.

COX-2 was expressed in the cytoplasm of tumor cells and occasionally in inflammatory cells. All melanomas were positive for COX-2 in different degrees. In the present series COX-2 expression had the highest positivity index for amelanotic (scores ranging from 143 to 294, mean 206), followed by melanotic melanomas (scores ranging from 101 to 144, mean 122.5). Representative photomicrographs are illustrated in the Figure 1. The means were statistically different (t test, P<0.001).



Figure 1. COX-2 expression in amelanotic (A) and melanotic melanomas (B). Graphical representation of the scores of immunopositivity.

Ki-67 presented a nuclear expression mainly in tumor cells. The index was digitally calculated and were higher for amelanotic (ranging from 34 to 92%, mean 64%) than melanotic melanomas (ranging from 11 to 59%, mean 30.8%). These results are described in the Figure 2. The means were statistically different (t test, P<0.001).


Figure 2. Ki67-nuclear expression in amelanotic (A) and melanotic melanomas (B). Graphical representation of the cellular proliferative index based on Ki67 expression.

Discussion

To the best of our knowledge this is the first study to compare the expression of COX-2 and Ki67 in amelanotic versus conventional melanotic oral melanomas. In the present study we found that both studied markers demonstrated higher expression in amelanotic melanomas than melanotic ones. The main theory for these results, is that the amelanotic tumors represent a most undifferentiated neoplasm, and the presence of melanin may represent a more differentiated phenotype. Other theory is that amelanotic tumors represent a de-differentiated conventional melanoma. In our opinion, it is very difficult to affirm which theory is more adequate. We only can confirm that amelanotic tumors really present a more aggressive behavior.

Tumor cell proliferation index is widely recognized to indicate the degree of aggressiveness of a tumor. In addition, Ki67-index has been used as prognostic factor for several tumors, including breast cancer¹⁴ and mucosal melanomas.¹⁵ Although our data suggest a relationship between COX-2 expression and higher cell proliferation, other features must be considered including the idea that amelanotic tumors are more undifferentiated, have higher frequency of necrosis and mitosis than melanotic melanomas.

In fact, the higher expression of COX-2 in amelanotic corroborates with the hypothesis that these tumors have a poorer clinical prognosis, once COX-2 is involved

in several pro-tumorigenic events, including angiogenesis⁷, cell proliferation and migration⁶ and association with a metastatic phenotype.¹¹ Additional studies are necessary to determine the biological differences between amelanotic and melanotic oral melanomas.

In the cutaneous melanomas, the absence of melanin is an indicator of aggressiveness. Cheung et al.³ have demonstrated that a higher mitotic index in amelanotic melanoma correlates with greater nuclear atypia and a worse prognosis. Similarly, our results demonstrated that oral amelanotic melanomas have a higher Ki67-index than the pigmented ones.

In summary, amelanotic melanomas had a higher COX-2 expression and higher cellular proliferative than melanotic ones. These data suggest that amelanotic melanomas differ from the melanotic subtype and more studies are necessary to better understand the role that the absence of melanin deposition may cause in the tumoral biology. It is important to keep in mind that, due to the rarity of these neoplasms, the data presented here must be interpreted with caution.

REFERENCES

1. Lourenço S V., Fernandes JD, Hsieh R, et al. Head and neck mucosal melanoma: A review. Am J Dermatopathol. 2014;36(7):578-587. doi:10.1097/DAD.000000000000035

2. Moshe M, Levi A, Ad-El D, et al. Malignant melanoma clinically mimicking pyogenic granuloma: comparison of clinical evaluation and histopathology. Melanoma Res. 2018;28(4):363-367. doi:10.1097/CMR.00000000000451

3. Cheung WL, Patel RR, Leonard A, Firoz B, Meehan SA. Amelanotic melanoma: a detailed morphologic analysis with clinicopathologic correlation of 75 cases. J Cutan Pathol. 2012;39(1):33-39. doi:10.1111/j.1600-0560.2011.01808.x

4. Fitzpatrick F. Cyclooxygenase Enzymes: Regulation and Function. Curr Pharm Des. 2005;10(6):577-588. doi:10.2174/1381612043453144

5. Hashemi Goradel N, Najafi M, Salehi E, Farhood B, Mortezaee K. Cyclooxygenase-2 in cancer: A review. J Cell Physiol. 2019;234(5):5683-5699. doi:10.1002/jcp.27411

6. Xia M, Duan M-L, Tong J-H, Xu J-G. MiR-26b suppresses tumor cell proliferation, migration and invasion by directly targeting COX-2 in lung cancer. Eur

RevMedPharmacolSci.2015;19(24):4728-4737.http://www.ncbi.nlm.nih.gov/pubmed/26744864.Accessed December 11, 2019.

7. Hu H, Han T, Zhuo M, et al. Elevated COX-2 Expression Promotes Angiogenesis Through EGFR/p38-MAPK/Sp1-Dependent Signalling in Pancreatic Cancer. Sci Rep. 2017;7(1). doi:10.1038/s41598-017-00288-4

8. Guo Z, Jiang JH, Zhang J, et al. COX-2 Promotes Migration and Invasion by the Side Population of Cancer Stem Cell-Like Hepatocellular Carcinoma Cells. Med (United States). 2015;94(44):e1806. doi:10.1097/MD.00000000001806

9. Semaan J, Pinon A, Rioux B, et al. Resistance to 3-HTMC-Induced Apoptosis Through Activation of PI3K/Akt, MEK/ERK, and p38/COX-2/PGE2 Pathways in Human HT-29 and HCT116 Colorectal Cancer Cells. J Cell Biochem. December 2016:2875-2885. doi:10.1002/jcb.25600

 Montezuma MAP, Fonseca FP, Benites BM, et al. COX-2 as a determinant of lower disease-free survival for patients affected by ameloblastoma. Pathol Res Pract.
 2018. doi:10.1016/j.prp.2018.03.014

11. Soares CD, Borges CF, Sena-Filho M, et al. Prognostic significance of cyclooxygenase 2 and phosphorylated Akt1 overexpression in primary nonmetastatic and metastatic cutaneous melanomas. Melanoma Res. 2017;27(5):448-456. doi:10.1097/CMR.00000000000368

12. Prasad ML, Patel SG, Huvos AG, Shah JP, Busam KJ. Primary mucosal melanoma of the head and neck: a proposal for microstaging localized, Stage I (lymph node-negative) tumors. Cancer. 2004;100(8):1657-1664. doi:10.1002/cncr.20201

13. Soares C, De Lima Morais TM, Carlos R, et al. Phosphorylated Akt1 expression is associated with poor prognosis in cutaneous, oral and sinonasal melanomas. Oncotarget. 2018;9(99):37291-37304. doi:10.18632/oncotarget.26458

14. Soliman NA, Yussif SM. Ki-67 as a prognostic marker according to breast cancer molecular subtype. Cancer Biol Med. 2016;13(4):496-504. doi:10.20892/j.issn.2095-3941.2016.0066

15. Ma X, Wu Y, Zhang T, et al. Ki67 proliferation index as a histopathological predictive and prognostic parameter of oral mucosal melanoma in patients without distant metastases. J Cancer. 2017;8(18):3828-3837. doi:10.7150/jca.20935

2.5 Artigo: Prognostic importance of mitochondrial markers in head and neck mucosal and cutaneous melanomas

Artigo publicado no periódico Human Pathology DOI: 10.1016/j.humpath.2018.11.009 (Anexo 3)

Ciro Dantas Soares¹, Thayná Melo de Lima Morais¹, Roman Carlos², Oslei Paes de Almeida¹, Fernanda Viviane Mariano Brum Corrêa^{1,3}, Albina Messias de Almeida Milani Altemani^{1,3}, Maria Goretti Freire de Carvalho⁴, Marcelo Brum Corrêa⁵, Rodrigo Ribas Dias dos Reis⁶, Luciana Schultz Amorim⁷, Jacks Jorge¹

- Department of Oral Diagnosis, Area of Pathology, Piracicaba Dental School, University of Campinas, Piracicaba, São Paulo, Brazil.
- 2. Pathology Division, Centro Clínico de Cabeza y Cuello/Hospital Herrera Llerandi, Guatemala City, Guatemala.
- Department of Pathology, Faculty of Medical Sciences, University of Campinas, Campinas, São Paulo, Brazil.
- 4. Private Pathology Service, Natal, Rio Grande do Norte, Brazil.
- 5. Head and Neck Surgery Department, Oncology Center (CEON), Fornecedores de Cana Hospital, Piracicaba, São Paulo, Brazil.
- 6. Oncology Surgery Department, Cancer Center (CECAN), Santa Casa Hospital, Piracicaba, São Paulo, Brazil.
- 7. Institute of Pathological Anatomy, Piracicaba, São Paulo, Brazil.

Running Head: Mitochondrial markers predict survival in melanomas

Disclose any potential conflict of interest: The authors state no conflict of interest. Word Count: 2608. Number of Figures = 3. Number of Tables = 6.

Corresponding author: Ciro Dantas Soares Department of Oral Diagnosis, Piracicaba Dental School, University of Campinas Avenida Limeira, 901, Areião - 13414-903 Piracicaba/SP, Brazil E-mail address: ciro.dss@gmail.com / c162617@g.unicamp.br

ABSTRACT

Mitochondrial dysfunction is caused by an imbalance in the processes of fission and fusion, and it has been implicated in the pathogenesis of several human cancers. However, the role of mitochondrial markers in melanomas still remain poorly understood. In this study, we assessed the expression of three mitochondrial markers [antimitochondrial (AMT), fission protein 1 (FIS1) and mitofusin 2 (MFN2)] in a series of head and neck mucosal and cutaneous melanoma (CM). One hundred and twelve patients with cutaneous (n=56), and mucosal [oral - n=30 and sinonasal - n=26] melanomas of the head and neck region were enrolled in this study. Clinical and followup data were retrieved from the medical records. The expression of three mitochondrial markers was assessed by the immunohistochemistry, digitally quantified and correlated with clinicopathological data and outcome information. In the multivariate model, high mitochondrial content was identified as an independent prognostic value for diseasefree survival (DFS) in cutaneous melanomas and overall survival (OS) in oral melanomas. FIS1 expression was significantly associated with lower OS rates in patients with oral melanomas, and strictly correlated with vascular invasion in mucosal melanomas. MFN2 was identified as an independent prognostic marker for DFS and OS in patients with CM, and it was associated with a high risk of distant metastasis in these tumors. In summary, we demonstrated that mitochondrial content and FIS1 and MFN2 expressions are correlated with important clinicopathologic characteristics in patients with cutaneous and mucosal melanomas of the head and neck region.

Keywords: head and neck melanomas, cutaneous melanomas, mucosal melanomas, mitochondria, FIS1, MFN2, prognosis.

INTRODUCTION

Mitochondria are highly dynamic organelles involved in the energy production of all human cells [1]. Several biological processes require the ATP as an energy source through oxidative phosphorylation (OXPHOS) [2]. As a consequence of the OXPHOS, mitochondria also produce the majority of reactive oxygen species (ROS). ROS are implicated in the carcinogenic process, damaging important proteins and macromolecules, causing a deregulation in the cell cycle and other relevant process for the tumor development and metastasis [3]. In addition, mitochondria play an essential role in apoptosis, regulating critical signaling pathways for its activation or inhibition [4].

Currently, there has been described the predictive value of mitochondrial markers in recurrence, metastasis and tamoxifen-resistance of breast cancer patients [5]. In the intracellular compartment, mitochondria constantly change its number and shape in an event named mitochondrial dynamics, which involves two main processes: fusion and fission [1]. Although these processes are widely investigated, their role in cancer cells and consequently in the context of clinical behavior, tumorigenesis and metastasis remains poorly studied in melanoma.

Melanoma is an aggressive tumor that arises most commonly on the skin or mucous membrane. Prognosis of primary melanoma is based on morphological parameters such as tumor thickness and mitotic index [6,7]. Considering that cutaneous melanomas (CM) is strictly associated with sun exposure, whereas in mucosal melanomas (MM) it has no effect in their pathogenesis, comparative studies are necessary to clarify the differences in signaling pathways involved in the different subtypes of melanomas [8,9]. In addition, there is currently demonstrated that CM and MM harbor different genetic alterations and few studies have compared mucosal and cutaneous melanomas of the head and neck (H&N) region [10]. Oral and sinonasal melanomas are uncommon tumors that arise in the mucosa of these anatomical sites, presenting an aggressive behavior, tendency to metastasize and consequently worse prognosis than cutaneous melanomas [11,12]. To date, few studies have attempted to describe proteins with prognostic predictive value for these tumors.

The role of metabolic markers in several cancers is well stablished [13–15]. Moreover, high levels of ROS are associated with tumor development and progression, and as the main source of ROS are mitochondria, we hypothesized that these organelles might play an essential role in tumorigenesis and impacts in the biological behavior of several cancers [16]. However, in melanomas, it is not fully clarified. In the current study, we compare the expression of three mitochondrial markers in oral, sinonasal and cutaneous melanomas of the H&N region.

METHODS

Case selection

We retrospectively collected 112 H&N cutaneous and mucosal melanoma cases with follow-up and complete clinical information from four pathology laboratories. Formalin-fixed, paraffin-embedded tissue blocks were retrieved; the diagnoses of the melanomas were reviewed and confirmed by three pathologists. In addition, we performed S-100, MelanA and HMB45 immunostainings to all cases.

Immunohistochemistry

For immunohistochemical (IHC) polymer-based method, 3-µm-thick sections mounted on silanized slides were used. The sections were deparaffinized, rehydrated in graded ethanol solutions and submitted to antigen retrieval with EDTA/Tris buffer (pH

9.0) in an electric pressure cooker for 15 minutes. Following that, endogenous peroxidase activity was blocked with 20% H2O2 by single incubation of 15 minutes.

The sections were then incubated with the diluted primary antibodies for two hours at room temperature. We used two high-sensitive visualization systems: ADVANCE[™]/HRP (code K406889-2; Dako, Carpinteria, CA, USA) and EnVision G|2 System/AP, Rabbit/Mouse (Permanent Red) (code K535521-2, Dako). The IHQ reactions were revealed with Permanent Red (Dako) or DAB (Sigma Aldrich, St. Louis, MO, USA) and counterstained with Carazzi's hematoxylin. Detailed information about the primary antibodies, dilution, manufacturer and respective positive controls is listed in Supplementary Table 1.

Digital analysis

Following the IHC reactions, the slides were scanned into high-resolution images and digitally assessed as described previously with the scores of positivity ranged from 100 (very weak) to 300 (strongly positive) [17]. One calibrated pathologist selected 10 different regions per case to assess the digital IHC quantification, these areas were from the superior and inferior portions of the tumors and not included highly melanin-pigmented areas.

Statistical methods

Briefly, for the statistical methods, we divided the cutaneous melanomas into two groups based in the Clark's level: (1) I, II and III and (2) IV and V. The scores of immunostaining were correlated with clinical data from all types of melanomas using contingency tables and Chi-Square or Fisher's exact test. Survival curves were calculated according to the Kaplan-Meier method. The Log-Rank test was applied for patients with low and high expression of the markers (cutoff value: median of the scores of positivity). COX proportional hazards regression analysis was performed to test the statistical independence and significance between pathological, molecular and clinical variables. All statistical tests were carried out in the SPSS software, version 22.0 (SPSS Inc., Chicago, IL, USA) with a 95% confidence level (P-value ≤ 0.05).

The National Commission for Ethics in Research approved the study protocol (CONEP-Brazil, CAAE: 72077517.1.0000.5418). This study was conducted according to the Declaration of Helsinki Principles.

RESULTS

The immunostaining scores were quantitatively assessed by digital analysis. All mitochondrial markers demonstrated a high expression in tumor cells. Overall, expression of AMT was higher in mucosal than in cutaneous melanomas (Figure 1); FIS1 immunostaining was high in oral melanomas and closely correlated with tumor cells in areas of vascular and neural invasion (Figure 2) and expression of MFN2 was very higher in cutaneous and sinonasal melanomas than in oral melanomas (Figure 3). The median scores for cutaneous melanomas were 199.4 (range 248.8–118.4 for AMT), 134.5 (range 175.9–101.9 for FIS1) and 211.9 (range 255.7–156.2 for MFN2). For oral melanomas, 224.4 (range 268.1–109.3 for AMT), 179.7 (range 230.0–116.1 for FIS1) and 182.7 (range 226.4–142.0 for MFN2). For sinonasal melanomas, 229.4 (range 258.9–173.5 for AMT), 124.1 (range 181.0–112.3 for FIS1) and 239.8 (range 259.4–208.7 for MFN2).



Figure 1. AMT expression in cutaneous and mucosal head and neck melanomas. A: Medium cytoplasmic positivity in malignant melanocytes in cutaneous melanoma. B: Strong positivity in a predominant fusiform oral melanoma. C: Strong positivity in a spindle/epithelioid sinonasal melanoma. D: Perinuclear pattern of AMT immunostaining in sinonasal melanomas, undifferentiated type. The magnification scale is 100 μ m for all figures.



Figure 2. FIS1 expression in cutaneous and mucosal head and neck melanomas. A: Medium cytoplasmic positivity in macrophages, and very weak FIS1 expression in cutaneous melanoma. B: Weak cytoplasmic expression in tumor cells of cutaneous melanoma. C: Strong FIS1 expression in malignant melanocytes of oral melanoma. D: FIS1-positive cells during mitosis (blue arrows). E: Strong FIS1 positivity in vascular and neural invasion areas. F: Weak FIS1 immunostaining in tumor cells of sinonasal melanomas,

occasional macrophages are positive. The magnification scale is 100 μm for all figures, except for C, which represents 200 $\mu m.$



Figure 3. MFN2 expression in cutaneous and mucosal head and neck melanomas. A: Medium MFN2 immunostaining in cutaneous melanoma. B: Positivity of MFN2 was focal and in individual cells of oral melanomas. C: Perinuclear immunopositivity of MFN2 was observed in some cases of sinonasal melanomas. D: Strong positivity of MFN2 in a case of sinonasal melanoma (stage IVc). The magnification scale is 100 μ m for all figures.

Cutaneous melanomas

A total of fifty-six patients with cutaneous melanomas of the head and neck region (25 female, 31 male) were included. The cases distribution of AJCC-stages was as follows: stage 0, In situ 5.4% (3/56), IA 17.9% (10/56), IB 8.8% (5/56), IIA 10.7% (6/56), IIB 12.5% (7/56), IIC 8.9% (5/56), IIIA 5.4% (3/56), IIIB 5.4% (3/56), IIIC 7.1% (4/56) and IV 17.9% (10/56). The mean of Breslow's thickness was 3.0 mm (ranging from 0 to 38 mm).

All mitochondrial markers studied here demonstrated a correlation with the presence of ulceration, Breslow's thickness and mitotic index. AMT and FIS1 were associated with AJCC-stage, whereas FIS1 and MFN2 were correlated with Clark's level. High expression of FIS1 was also correlated with vertical growth phase (Table 1).

| | | AMT [n (%)] | | P-value | FIS1 [| FIS1 [n (%)] | | MFN2 [n (%)] | | P-value |
|----------------------------------|------------|-------------|-----------|-----------|-----------|--------------|-----------|--------------|-----------|-----------|
| Variables | Categories | Low | High | | Low | High | | Low | High | |
| Age | <56 | 14 (51.9) | 15 (51.7) | 0.9923 | 14 (50.0) | 15 (53.6) | 0.7891 | 13 (46.4) | 16 (57.1) | 0.4223 |
| | ≥56 | 13 (48.1) | 14 (48.3) | | 14 (50.0) | 13 (46.4) | | 15 (53.6) | 12 (42.9) | |
| Sex | Female | 13 (48.1) | 12 (41.4) | 0.6106 | 13 (46.4) | 12 (42.9) | 0.7880 | 13 (46.4) | 12 (42.9) | 0.7880 |
| | Male | 14 (51.9) | 17 (58.6) | | 15 (53.6) | 16 (57.1) | | 15 (53.6) | 16 (57.1) | |
| Ulceration | Absent | 21 (77.8) | 13 (44.8) | 0.0116* | 24 (85.7) | 10 (35.7) | 0.0001* | 24 (85.7) | 10 (35.7) | 0.0001* |
| | Present | 6 (22.2) | 16 (55.2) | | 4 (14.3) | 18 (64.3) | | 4 (14.3) | 18 (64.3) | |
| Growth phase | Radial | 10 (37.0) | 12 (41.4) | 0.7395 | 15 (53.6) | 7 (25.0) | 0.0286* | 13 (46.4) | 9 (32.1) | 0.2737 |
| | Vertical | 17 (63.0) | 17 (58.6) | | 13 (47.4) | 21 (75.0) | | 15 (53.6) | 19 (67.9) | |
| Breslow's thickness (mm) | <3.3 | 23 (85.2) | 16 (55.2) | 0.0146* | 25 (89.3) | 14 (50.0) | 0.0013* | 24 (85.7) | 15 (53.6) | 0.0089* |
| | ≥3.3 | 4 (14.8) | 13 (44.8) | | 3 (10.7) | 14 (50.0) | | 4 (14.3) | 13 (46.4) | |
| Clark's level | I, II, III | 15 (55.6) | 10 (34.5) | 0.1129 | 22 (78.6) | 3 (10.7) | < 0.0001* | 17 (60.7) | 8 (28.6) | 0.0155* |
| | IV, V | 12 (44.4) | 19 (65.5) | | 6 (21.4) | 25 (89.3) | | 11 (39.3) | 20 (71.4) | |
| Mitotic index (mm ²) | <3 | 16 (59.3) | 8 (27.6) | 0.0167* | 19 (67.9) | 5 (17.9) | 0.0001* | 19 (67.9) | 5 (17.9) | 0.0001* |
| | <u>≥</u> 3 | 11 (40.7) | 21 (72.4) | | 9 (32.1) | 23 (82.1) | | 9 (32.1) | 23 (82.1) | |
| AJCC-stage | I and II | 24 (88.9) | 9 (31.0) | < 0.0001* | 19 (67.9) | 14 (50.0) | 0.1744 | 24 (85.7) | 9 (32.1) | < 0.0001* |
| | III and IV | 3 (11.1) | 20 (69.0) | | 9 (32.1) | 14 (50.0) | | 4 (14.3) | 19 (67.9) | |

Table 1. Relationship of AMT, FIS1, and MFN2 expressions with clinicopathological characteristics of all patients with HN cutaneous melanomas (n=56).

AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2; HN, Head and Neck; AJCC, American Joint Committee on Cancer. Asterisks (*) indicate statistical significance (P<0.05).

| Variables | Categ | Categories | | ivariate | P-value | Multivari | ate | OS univariate | | P-value | Multivaria | te |
|----------------|------------|------------|---------|----------|------------|-----------------|---------|---------------|---------|------------|------------------|---------|
| | | | (% | %) | (log-rank) | HR (95% CI) | P-value | (% | 5) | (log-rank) | HR (95% CI) | P-value |
| | | n (%) | 3-years | 5-years | | | | 3-years | 5-years | | | |
| Age | <56 | 29 (51.8) | 93.1 | 77.3 | 0.29 | _ | _ | 93.1 | 81.6 | 0.21 | _ | _ |
| - | ≥56 | 27 (48.2) | 92.3 | 59.9 | (1.09) | | | 92.3 | 67.6 | (1.57) | | |
| Sex | Female | 25 (44.6) | 95.7 | 77.0 | 0.39 | — | _ | 95.7 | 82.2 | 0.22 | _ | _ |
| | Male | 31 (55.4) | 90.3 | 62.0 | (0.71) | | | 90.3 | 68.9 | (1.46) | | |
| Ulceration | Absent | 34 (60.7) | 97.1 | 87.4 | < 0.01* | 3.27 | 0.08 | 97.1 | 90.9 | < 0.01* | 2.23 | 0.34 |
| | Present | 22 (39.3) | 85.4 | 30.1 | (24.17) | (0.86 - 12.35) | | 85.4 | 45.4 | (14.76) | (0.42 - 11.76) | |
| Growth phase | Radial | 22 (39.3) | 95.5 | 71.3 | 0.47 | _ | _ | 95.5 | 76.4 | 0.24 | _ | _ |
| | Vertical | 34 (60.7) | 87.7 | 67.5 | (0.51) | | | 87.8 | 74.4 | (1.34) | | |
| Breslow's | <3.3 | 39 (69.5) | 97.4 | 86.4 | < 0.01* | 4.61 | 0.02* | 97.4 | 86.4 | < 0.01* | 4.593 | 0.052 |
| thickness (mm) | ≥3.3 | 17 (30.5) | 81.6 | 30.1 | (13.43) | (1.24 - 17.11) | | 81.6 | 48.3 | (7.69) | (0.98 - 21.43) | |
| Clark's level | I, II, III | 25 (44.6) | 95.8 | 77.9 | 0.02* | 4.32 | 0.44 | 100.0 | 82.8 | 0.23 | _ | _ |
| | IV, V | 31 (55.4) | 86.6 | 57.0 | (4.79) | (0.1 - 184.66) | | 86.6 | 68.2 | (1.40) | | |
| Mitotic index | <3 | 24 (42.8) | 95.8 | 91.7 | < 0.01* | 5.168 | 0.04* | 95.8 | 91.7 | < 0.01* | 2.87 | 0.20 |
| (mm^2) | ≥3 | 32 (57.2) | 90.2 | 50.9 | (16.32) | (1.06 - 24.98) | | 90.2 | 61.1 | (7.84) | (0.55 - 14.82) | |
| AJCC-stage | I and II | 33 (58.9) | 96.6 | 85.5 | < 0.01* | 2.469 | 0.18 | 100.0 | 96.4 | < 0.01* | 11.21 | 0.03* |
| | III and IV | 23 (41.1) | 82.6 | 41.1 | (24.48) | (0.65 - 9.343) | | 82.6 | 46.2 | (22.48) | (1.18 - 106.003) | |
| AMT | Low | 27 (48.2) | 100.0 | 95.5 | < 0.01* | 24.07 | 0.012* | 100.0 | 95.5 | < 0.01* | 1.32 | 0.79 |
| | High | 29 (51.8) | 85.9 | 46.0 | (26.72) | (2.03 - 284.66) | | 85.9 | 56.5 | (12.67) | (0.15 - 11.34) | |
| FIS1 | Low | 28 (50.0) | 96.4 | 81.3 | < 0.01* | 0.04 | 0.14 | 96.4 | 85.6 | 0.06 | 0.24 | 0.13 |
| | High | 28 (50.0) | 88.7 | 49.8 | (8.92) | (0.001 - 2.989) | | 88.7 | 62.5 | (3.32) | (0.041 - 1.52) | |
| MFN2 | Low | 28 (50.0) | 100.0 | 88.7 | < 0.01* | 1.14 | 0.87 | 100.0 | 92.7 | < 0.01* | 2.09 | 0.49 |
| | High | 28 (50.0) | 85.1 | 45.9 | 24.03 | (0.21 - 6.181) | | 85.1 | 53.7 | (15.06) | (0.25 - 17.31) | |

Table 2. Relationship of clinicopathological characteristics of patients with HN cutaneous melanomas (n=56) and AMT, FIS1, and MFN2expressions with DFS and OS rates in univariate and multivariate models.

AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2; HN, Head and Neck; AJCC, American Joint Committee on Cancer; DFS, Disease-free survival; OS, overall survival, HR, Harzard Ratio; CI, Confidence interval. Asterisks (*) indicate statistical significance (P<0.05).

| | | AMT [n (%)] | | P-value | FIS1 [| FIS1 [n (%)] | | MFN2 | [n (%)] | P-value |
|---------------------|--------------------|-------------|-----------|---------|-----------|--------------|----------|-----------|-----------|---------|
| Variables | Categories | Low | High | | Low | High | | Low | High | |
| Age | <48 | 6 (40.0) | 10 (66.7) | 0.1432 | 7 (46.7) | 9 (60.0) | 0.4642 | 7 (46.7) | 9 (60.0) | 0.4642 |
| - | ≥48 | 9 (60.0) | 5 (33.3) | | 8 (53.3) | 6 (40.0) | | 8 (53.3) | 6 (40.0) | |
| Sex | Female | 9 (60.0) | 7 (46.7) | 0.4642 | 6 (40.0) | 10 (66.7) | 0.1432 | 10 (66.7) | 6 (40.0) | 0.1432 |
| | Male | 6 (40.0) | 8 (53.3) | | 9 (60.0) | 5 (33.3) | | 5 (33.3) | 9 (60.0) | |
| Site | Palate | 6 (40.0) | 7 (46.7) | 0.8706 | 5 (33.3) | 8 (53.3) | 0.0641 | 8 (53.3) | 5 (33.3) | 0.5418 |
| | Gingiva | 3 (20.0) | 2 (13.3) | | 1 (6.7) | 4 (26.7) | | 2 (13.3) | 3 (20.0) | |
| | Others | 6 (40.0) | 6 (40.0) | | 9 (60.0) | 3 (20.0) | | 5 (33.4) | 7 (46.7) | |
| Treatment | Only Surgery | 12 (80.0) | 8 (53.2) | 0.2465 | 12 (80.0) | 8 (53.3) | 0.2465 | 9 (60.0) | 11 (73.3) | 0.5191 |
| | Surgery plus CH/RT | 3 (20.0) | 6 (40.0) | | 3 (20.0) | 6 (40.0) | | 5 (33.3) | 4 (26.7) | |
| | No treatment | 0 (0.0) | 1 (6.7) | | 0 (0.0) | 1 (6.7) | | 1 (6.7) | 0 (0.0) | |
| Vascular invasion | Absent | 10 (66.7) | 9 (60.0) | 0.7047 | 15 (100) | 4 (26.7) | < 0.001* | 9 (60.0) | 10 (66.7) | 0.7047 |
| | Present | 5 (33.3) | 6 (40.0) | | 0 (0.0) | 11 (73.3) | | 6 (40.0) | 5 (33.3) | |
| Neural invasion | Absent | 13 (86.7) | 11 (73.3) | 0.3613 | 14 (93.3) | 10 (66.7) | 0.0678 | 12 (80.0) | 12 (80.0) | 1.000 |
| | Present | 2 (13.3) | 4 (26.7) | | 1 (6.7) | 5 (33.3) | | 3 (20.0) | 3 (20.0) | |
| Mitotic index | <1 | 9 (60.0) | 4 (26.7) | 0.0654 | 11 (73.3) | 2 (13.3) | < 0.001* | 6 (40.0) | 7 (46.7) | 0.7125 |
| | ≥1 | 6 (40.0) | 11 (73.3) | | 4 (26.7) | 13 (86.7) | | 9 (60.0) | 8 (53.3) | |
| Cellular morphology | Non-epithelioid | 8 (53.3) | 2 (13.3) | 0.0201* | 7 (46.7) | 3 (20.0) | 0.1213 | 5 (33.3) | 5 (33.3) | 1.000 |
| | Epithelioid | 7 (46.7) | 13 (86.7) | | 8 (53.3) | 12 (80.0) | | 10 (66.7) | 10 (66.7) | |
| AJCC-stage | Ī | 8 (53.3) | 5 (33.3) | 0.4843 | 8 (53.3) | 5 (33.3) | 0.0233* | 8 (53.3) | 5 (33.3) | 0.0201* |
| - | IVa | 5 (33.3) | 6 (40.0) | | 7 (46.7) | 4 (26.7) | | 2 (13.4) | 9 (60.0) | |
| | IVb and IVc | 2 (13.4) | 4 (26.7) | | 0 (0.0) | 6 (40.0) | | 5 (33.3) | 1 (6.7) | |

Table 3. Relationship of AMT, FIS1, and MFN2 expressions with clinicopathological characteristics of all patients with oral melanomas (n=30).

AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2; AJCC, American Joint Committee on Cancer; CH, chemotherapy; RT, radiotherapy.

| Variables | Categories | | OS univariate | | P-value | Multivari | ate |
|---------------|--------------------|-----------|---------------|---------|---------|-----------------|---------|
| | _ | | (% | (%) | | HR (95% CI) | P-value |
| | | n (%) | 3-years | 5-years | | | |
| Age | <48 | 16 (53.3) | 56.3 | 40.2 | 0.76 | _ | _ |
| - | ≥48 | 14 (46.7) | 43.8 | 43.8 | (0.09) | | |
| Sex | Female | 16 (53.3) | 38.4 | 38.4 | 0.33 | _ | _ |
| | Male | 14 (46.7) | 64.3 | 42.9 | (0.94) | | |
| Anatomical | Palate | 13 (43.3) | 27.7 | 0.0 | 0.02* | 0.33 | 0.02* |
| site | Gingiva | 5 (16.6) | 40.0 | 40.0 | (7.46) | (0.13 - 0.85) | |
| | Others | 12 (40.1) | 81.8 | 65.5 | | | |
| Treatment | Only surgery | 20 (66.7) | 57.4 | 57.4 | 0.21 | _ | _ |
| | Surgery plus CH/RT | 9 (30.0) | 44.4 | 14.8 | (1.57) | | |
| | No treatment | 1 (3.3) | _ | _ | | | |
| Vascular | Absent | 19 (63.3) | 83.6 | 61.0 | < 0.01* | 1.61 | 0.69 |
| invasion | Present | 11 (36.7) | 0.0 | 0.0 | (20.69) | (0.14 - 17.97) | |
| Neural | Absent | 24 (80.0) | 60.4 | 44.1 | 0.03* | 0.82 | 0.81 |
| invasion | Present | 6 (20.0) | 16.7 | 16.7 | (4.48) | (0.17 - 3.93) | |
| Mitotic index | <1 | 13 (43.3) | 91.7 | 91.7 | < 0.01* | 6.75 | 0.18 |
| (mm^2) | ≥1 | 17 (56.7) | 23.5 | 11.7 | (13.93) | (0.42 – 109.16) | |
| Necrosis | Absent | 22 (73.3) | 51.3 | 41.0 | 0.59 | _ | _ |
| | Present | 8 (26.7) | 50.0 | 25.0 | (0.28) | | |
| Cellular | Non-epithelioid | 10 (33.3) | 88.9 | 88.9 | < 0.01* | 8.56 | 0.11 |
| morphology | Epithelioid | 20 (66.7) | 35.0 | 0.0 | (9.76) | (0.59 – 122.71) | |
| AJCC-stage | III | 13 (43.3) | 68.4 | 51.3 | < 0.01* | 1.25 | 0.61 |
| | IVa | 11 (36.7) | 60.6 | 40.4 | (14.66) | (0.52 - 3.05) | |
| | IVb and IVc | 6 (20.0) | 0.0 | 0.0 | | | |
| AMT | Low | 15 (50.0) | 62.7 | 62.7 | 0.03* | 12.97 | 0.01* |
| | High | 15 (50.0) | 40.0 | 13.3 | (4.48) | (1.73 – 97.09) | |
| FIS1 | Low | 15 (50.0) | 92.9 | 63.7 | < 0.01* | 22.99 | 0.03* |
| | High | 15 (50.0) | 13.3 | 13.3 | (15.45) | (1.32 - 401.12) | |
| MFN2 | Low | 15 (50.0) | 52.5 | 28.0 | 0.49 | _ | _ |
| | High | 15 (50.0) | 50.3 | 50.3 | (0.47) | | |

Table 4. Relationship of clinicopathological characteristics of patients with oral melanomas (n=30) and AMT, FIS1, and MFN2 expressions with OS rates in univariate and multivariate models.

AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2; AJCC, American Joint Committee on Cancer; OS, overall survival, HR, Harzard Ratio; CI, Confidence interval.

| | | AMT [n (%)] | | P-value | FIS1 [| n (%)] | P-value | MFN2 | [n (%)] | P-value |
|---------------------|----------------------|-------------|----------|---------|-----------|-----------|---------|-----------|-----------|---------|
| Variables | Categories | Low | High | | Low | High | | Low | High | |
| Age | <59 | 6 (46.2) | 7 (53.8) | 0.6948 | 7 (53.8) | 6 (46.2) | 0.6948 | 4 (30.8) | 9 (69.2) | 0.0498* |
| | ≥59 | 7 (53.8) | 6 (46.2) | | 6 (46.2) | 7 (53.8) | | 9 (69.2) | 4 (30.8) | |
| Sex | Female | 6 (46.2) | 8 (61.5) | 0.4314 | 8 (61.5) | 6 (46.2) | 0.4314 | 10 (76.9) | 4 (30.8) | 0.0182* |
| | Male | 7 (53.8) | 5 (38.5) | | 5 (38.5) | 7 (53.8) | | 3 (23.1) | 9 (69.2) | |
| Site | Nasal cavity | 6 (46.2) | 5 (38.4) | 0.3180 | 7 (53.8) | 4 (30.8) | 0.4920 | 4 (30.8) | 7 (53.8) | 0.4920 |
| | Paranasal sinuses | 6 (46.2) | 4 (30.8) | | 4 (30.8) | 6 (46.2) | | 6 (46.1) | 4 (30.8) | |
| | Rinopharynx | 1 (7.6) | 4 (30.8) | | 2 (15.4) | 3 (23.0) | | 3 (23.1) | 2 (15.2) | |
| Treatment | Only Surgery | 6 (46.2) | 5 (38.5) | 0.5024 | 7 (53.8) | 4 (30.8) | 0.3492 | 5 (38.5) | 6 (46.2) | 0.5024 |
| | Surgery plus CH/RT | 6 (46.2) | 8 (61.5) | | 6 (46.2) | 8 (61.5) | | 8 (61.5) | 6 (46.2) | |
| | No treatment | 1 (7.6) | 0 (0.0) | | 0 (0.0) | 1 (7.7) | | 0 (0.0) | 1 (7.6) | |
| Vascular invasion | Absent | 10 (76.9) | 9 (69.2) | 0.6583 | 12 (92.3) | 7 (53.8) | 0.0270* | 11 (84.7) | 8 (61.5) | 0.1847 |
| | Present | 3 (23.1) | 4 (30.8) | | 1 (7.7) | 6 (46.2) | | 2 (15.3) | 5 (38.5) | |
| Neural invasion | Absent | 11 (84.6) | 13 (100) | 0.1410 | 13 (100) | 11 (84.7) | 0.1410 | 13 (100) | 11 (84.7) | 0.1410 |
| | Present | 2 (15.4) | 0 (0.0) | | 0 (0.0) | 2 (15.3) | | 0 (0.0) | 2 (15.3) | |
| Mitotic index | <1 | 10 (76.9) | 8 (61.5) | 0.3954 | 10 (76.9) | 8 (61.5) | 0.3954 | 8 (61.5) | 10 (76.9) | 0.3954 |
| | ≥1 | 3 (23.1) | 5 (38.5) | | 3 (23.1) | 5 (38.5) | | 5 (38.5) | 3 (23.1) | |
| Cellular morphology | Plasmacytoid/Others | 4 (30.8) | 6 (46.2) | 0.4201 | 7 (53.8) | 3 (23.1) | 0.1068 | 9 (69.2) | 1 (7.7) | 0.0012* |
| | Fusiform/Epithelioid | 9 (69.2) | 7 (53.8) | | 6 (46.2) | 10 (76.9) | | 4 (30.8) | 12 (92.3) | |
| AJCC-stage | III | 5 (38.5) | 4 (30.8) | 0.2110 | 5 (38.5) | 4 (30.8) | 0.6969 | 5 (38.4) | 4 (30.8) | 0.8948 |
| - | IVa | 6 (46.2) | 3 (23.1) | | 5 (38.4) | 4 (30.8) | | 4 (30.8) | 5 (38.4) | |
| | IVb and IVc | 2 (15.3) | 6 (46.1) | | 3 (23.1) | 5 (38.5) | | 4 (30.8) | 4 (30.8) | |

Table 5. Relationship of AMT, FIS1, and MFN2 expressions with clinicopathological characteristics of all patients with sinonasal melanomas (n=26).

AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2; AJCC, American Joint Committee on Cancer; CH, chemotherapy; RT, radiotherapy.

| Variables | Categories | | OS univariate | | P-value | Multivari | iate |
|--------------------|----------------------|-----------|---------------|---------|------------|---------------|---------|
| | C C | | (%) | | (log-rank) | HR (95% CI) | P-value |
| | | n (%) | 3-years | 5-years | | | |
| Age | <59 | 13 (50.0) | 59.2 | 39.5 | 0.67 | _ | _ |
| C | ≥59 | 13 (50.0) | 53.9 | 40.5 | (0.17) | | |
| Sex | Female | 14 (53.8) | 66.2 | 41.4 | 0.53 | _ | _ |
| | Male | 12 (46.2) | 48.6 | 36.5 | (0.38) | | |
| Anatomical | Nasal cavity | 11 (42.3) | 57.7 | 57.7 | 0.32 | _ | _ |
| site | Maxillary sinus | 10 (38.5) | 54.9 | 54.9 | (2.24) | | |
| | Rinopharynx | 5 (19.2) | 60.0 | 0.0 | | | |
| Treatment | Only surgery | 11 (42.3) | 57.1 | 28.6 | 0.24 | _ | _ |
| | Surgery plus CH/RT | 14 (53.8) | 60.6 | 50.5 | (1.36) | | |
| | No treatment | 1 (3.9) | _ | _ | | | |
| Vascular | Absent | 19 (73.1) | 70.1 | 50.1 | 0.01* | 2.16 | 0.15 |
| invasion | Present | 7 (26.9) | 28.6 | 14.3 | (5.60) | (0.75 - 6.19) | |
| Neural | Absent | 24 (92.3) | 63.0 | 43.0 | 0.12 | _ | — |
| invasion | Present | 2 (7.7) | 0.0 | 0.0 | (2.33) | | |
| Mitotic index | <1 | 18 (69.2) | 55.8 | 55.8 | 0.79 | _ | — |
| (mm ²) | ≥1 | 8 (30.8) | 62.5 | 15.6 | (0.06) | | |
| Necrosis | Absent | 16 (61.5) | 61.3 | 42.0 | 0.27 | _ | — |
| | Present | 10 (38.5) | 50.0 | 33.3 | (1.21) | | |
| Cellular | Plasmacytoid/others | 10 (38.5) | 100.0 | 83.3 | < 0.01* | 12.840 | 0.02* |
| morphology | Epithelioid/Fusiform | 16 (61.5) | 37.5 | 22.5 | (10.335) | (1.30–126.37) | |
| AJCC-stage | III | 9 (34.6) | 85.7 | 64.3 | 0.21 | _ | — |
| | IVa | 9 (34.6) | 43.8 | 29.2 | (3.12) | | |
| | IVb and IVc | 8 (30.8) | 37.5 | 18.8 | | | |
| AMT | Low | 13 (50.0) | 33.8 | 33.8 | 0.07 | _ | _ |
| | High | 13 (50.0) | 76.2 | 48.4 | (3.14) | | |
| FIS1 | Low | 13 (50.0) | 80.0 | 53.3 | 0.057 | _ | _ |
| | High | 13 (50.0) | 35.9 | 23.9 | (3.63) | | |
| MFN2 | Low | 13 (50.0) | 80.2 | 50.1 | 0.02* | 0.88 | 0.85 |
| | High | 13 (50.0) | 38.5 | 28.8 | (5.21) | (0.23 - 3.25) | |

Table 6. Relationship of clinicopathological characteristics of patients with sinonasal melanomas (n=26), AMT, FIS1, and MFN2 expressions with OS rates in univariate and multivariate models.

AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2; AJCC, American Joint Committee on Cancer; OS, overall survival, HR, Harzard Ratio; CI, Confidence interval.

Univariate Cox analysis of all cutaneous cases showed a significant correlation between lower disease-free survival (DFS) rates and presence of ulceration, Breslow's thickness, Clark's level, mitotic index, AJCC-stage, AMT, FIS1 and MFN2 expressions. While, in the multivariate Cox analysis of all cutaneous cases (including all variables with significance in the univariate model) only Breslow's thickness, mitotic index and AMT expression were significant for DFS. With respect to overall survival (OS), in the univariate model, ulceration, Breslow's thickness, mitotic index, AJCCstage, AMT and MFN2 expressions demonstrated significant prognostic value. All variables, except AJCC-stage, lost the prognostic value within a multivariate model (summarized in Table 2).

Moreover, the predictive value of tumor ulceration status, Breslow's thickness, mitotic rate, Clark's level, and the expression of AMT, FIS1 and MFN2 expressions for distant metastasis risk was assessed by Cox analysis. Only presence of ulceration (HR: 4.29, 95% CI: 1.19 - 15.37, *P*=0.025) and MFN2 expression (HR: 38.88, 95% CI: 3.42 - 441.93, *P*=0.003) were significantly correlated with high risk for distant metastasis, in a multivariate model (Supplementary Table 2).

Oral melanomas

Thirty patients with oral melanomas (16 female, 14 male) were included. Regarding the site distribution 13 cases occurred in the palate, 5 in the inferior gingiva and 12 cases in other localizations that include the tongue (1 case) and floor of the mouth (1 case), six cases have no information about the specific site. The distribution of AJCC-stages was as follows: III 43.3% (13/30); IVa 36.7% (11/30); IVb and IVc 20% (6/30).

High AMT expression was associated with epithelioid cellular morphology, whereas high FIS1 expression was correlated with the presence of vascular invasion, mitotic index and AJCC-stage. MFN2 expression was significantly associated only with AJCC-stage, with most cases with IVb/IVc stages presenting low expression of this marker (Table 3). Regarding the OS probabilities, some clinicopathologic characteristics were significantly correlated with lower OS, including anatomical site (palate), the presence of vascular and neural invasion, high mitotic index, epithelioid cellular morphology, advanced AJCC-stages, AMT and FIS1 expressions. High MFN2 expression demonstrated a better prognosis, without statistical significance. In the multivariate model, anatomical site (palate), AMT and FIS1 were significantly correlated with lower OS rates (summarized in Table 4).

Sinonasal melanomas

Twenty-six patients with oral melanomas (14 female, 12 male) were included. Regarding the site distribution 11 cases occurred in the nasal cavity, 10 in the maxillary sinus and 5 cases in the rinopharynx. The distribution of AJCC-stages was as follows: III 34.6% (9/26), IVa 34.6% (9/26); IVb and IVc 30.8% (8/26).

High expression of FIS1 was significantly correlated with the presence of vascular invasion, whereas MFN2 expression was associated with age (high expression in young patients), sex (high expression in men), and fusiform/epithelioid cellular morphology (Table 5). Lower OS rates were significantly associated with the presence of vascular invasion and fusiform/epithelioid cellular morphology. Whereas, regarding the mitochondrial markers, only MFN2 was associated with lower OS probability. In the multivariate model, fusiform/epithelioid cellular morphology showed predictive value for sinonasal melanomas (summarized in Table 6).

DISCUSSION

There are increasing shreds of evidence that mitochondrial dynamics regulate melanogenesis and probably has an important role in the malignant transformation of melanocytes [18]. Besides that, increased metabolism of tumor cells is a hallmark of cancer progression, and mitochondria are directly linked with energy production in human cells [19]. Corroborating with this, there has been studied a new tool for predicting melanoma metastasis using a magnetic resonance imaging method is based in the mitochondrial redox ratio, and it has an important value for predict and diagnosis of melanoma metastases [20]. In cancer cells, mitochondria constantly modify their number, shape and function mainly through two main processes (fission and fusion) and generically named mitochondrial dynamics [1,4]. The inhibition of mitochondrial dynamics-related proteins is also associated with reduced growth tumor [21]. However, limited research exists on the immunohistochemical expression of the mitochondrial markers and their prognostic value in mucosal and cutaneous melanomas. In the current study, we carried out a retrospective study to assess the expression of three mitochondrial markers in a series of cutaneous and mucosal (oral and sinonasal) melanomas. Furthermore, we evaluated the potential prognostic value of the mitochondrial content (assessed by AMT expression), and two proteins involved in the fission (FIS1) and fusion (MFN2) processes.

One interesting finding of this current study was the high mitochondrial content observed in MM than in CM, indicating that probably the mitochondrial mass may influence in the poor prognosis of MM. In agreement with prior reports high mitochondrial content is associated with poor prognosis in several human cancers such as head and neck squamous cell carcinomas [22], gallbladder [23], prostate [24] and breast carcinomas [25]. In addition, some studies have demonstrated a crucial role of the mitochondrial mass in acquisition of a malignant phenotype and in tumor chemoresistance [26,27]. Our findings also demonstrated that high mitochondrial content is an independent prognostic marker for DFS in cutaneous and OS in oral melanomas. In cutaneous melanomas, high expression of AMT was associated with AJCC-stage and Breslow's thickness, indicating a possible role of mitochondrial mass in cutaneous melanoma pathogenesis. High expression of AMT is also correlated with ulceration, indeed in ulcerated lesions; we observed a subpopulation of macrophages strongly positive for AMT. Thus, we hypothesize that the inflammatory infiltrate that overexpress AMT may be associated with the high mitochondrial content. As demonstrated previously, mitochondrial biogenesis plays a crucial role during cell cycle progression [28], and it corroborates with a positive correlation between high expression of AMT and mitotic index in cutaneous melanomas.

With respect to the fission process, several proteins are involved in its signaling, including dynamin-related protein 1 (DRP1) and fission protein 1 (FIS1) [29]. Recently, several studies have addressed the oncogenic role of DRP1 in cellular responses to MAPK inhibition [30]. In addition, another study showed that the induction of DRP1 in nevi and melanoma contributed to BRAFV600E disease [31]. Thus, it was hypothesized that the fission process could drive essential events for melanoma progression. Regarding FIS1 expression, in this study, we demonstrated a correlation between Clark's levels with increased FIS1 immunostaining. In fact, different Clark's levels comprise an interesting example to study cutaneous melanoma progress. Besides that, FIS1 expression was correlated with critical prognostic parameters for cutaneous melanomas, such as Breslow's thickness and mitotic index. In oral melanomas, FIS1 expression was associated with AJCC-stage and strictly correlated with vascular invasion. We have also identified FIS1 as an independent prognostic factor for OS in

patients with oral melanomas in both univariate and multivariate models. Importantly, our data suggest that FIS1 plays an essential role in the pathogenesis of cutaneous and oral melanomas, and consequently has identified as a possible prognostic biomarker for these tumors, mainly oral melanomas.

The exact role of MFN2 in cancer is not well established, with contradictory data, in some studies has demonstrated an oncogenic activity and other studies have studied its antioncogenic effect [13,32]. Concerning MFN2 expression in our cohort of head and neck melanomas, it was associated with lower DFS and OS in the multivariate model for patients with cutaneous melanomas. Furthermore, high MFN2 expression demonstrated a correlation with high risk for distant metastasis in a multivariate model. In oral melanomas, cases with high MFN2 expression have a better prognosis than cases with lower expression. Although this data was not significant, we consider important to highlight a possible antioncogenic effect of MFN2 in these particular tumors. It is in accordance with a previous study, in which, B16F10 melanoma cells knockdown for MFN2 demonstrated a higher number of lung metastasis [32]. On the other hand, for sinonasal melanomas, MFN2 was also associated with worse OS rates in the univariate model, corroborating with other studies [13,15]. Additional studies to determine the role of MFN2 in different human cancers types are highly suggested.

Current understanding of the clinical and pathological characteristics of mucosal melanomas is mainly based on the analysis of individual cases or small series [33–35]. However, in this study, we collected 56 oral and sinonasal melanomas samples with strict follow-up and complete clinicopathologic data. To the clinicopathologic parameters for oral melanomas, we identified a worse outcome for lesions in the palate and demonstrated a prognostic value of AMT and FIS1 for these tumors. In a univariate model, variables as vascular and neural invasion, mitotic index and AJCC-stage were

significantly correlated with lower OS rates. Importantly, epithelioid cellular morphology has emerged as a valuable prognostic tool, corroborating with previous studies. Regarding the sinonasal melanomas, only an epithelioid/fusiform cellular morphology was identified as an independent prognostic factor. Whereas in the univariate model, vascular invasion and MFN2 were correlated with lower OS probability. The lack of predictive value of these clinicopathologic parameters probably is due to the heterogeneity of these tumors, and future studies must be able to study the melanomas of the nasal cavity separately and from other localizations such as paranasal sinuses and rinopharynx.

In conclusion, high expression of AMT and FIS1 are associated with the aggressive characteristics and poor prognosis in oral melanomas. We also demonstrated the predictive value of MFN2 for high metastatic risk in cutaneous melanomas of the head and neck region. The novel mitochondrial markers that we identified may help in the development of new drugs targeting the mitochondria and modulating the mitochondrial dysfunction that underlies the pathogenesis of mucosal and cutaneous melanomas to prevent tumor recurrence and distant metastasis.

CONFLICTS OF INTEREST

The authors state no conflict of interest.

FUNDING

This work was supported by FAPESP grant numbers #2015/25905-1 and #2017/16802-

8.

AUTHOR CONTRIBUTIONS

Conception and design: CDS, JJ, OPA

Provision of study materials or patients: FVMBC, MBC, RRDR, LSA, RC, AMMA,

MGFC, OPA

Collection and assembly of data: CDS, MGFC, OPA, TMLM

Data analysis and interpretation: CDS, TMLM

Manuscript writing: CDS

Final approval of manuscript: All authors approved the final version of this manuscript.

REFERENCES

- Friedman JR, Nunnari J. Mitochondrial form and function. Nature 2014;505:335–43. doi:10.1038/nature12985.
- [2] Corazao-Rozas P, Guerreschi P, André F, Gabert P-E, Lancel S, Dekiouk S, et al. Mitochondrial oxidative phosphorylation controls cancer cell's life and death decisions upon exposure to MAPK inhibitors. Oncotarget 2016;7:39473–85. doi:10.18632/oncotarget.7790.
- [3] Liou G-Y, Storz P. Reactive oxygen species in cancer. Free Radic Res 2010;44:479–96. doi:10.3109/10715761003667554.

- [4] Suen DF, Norris KL, Youle RJ. Mitochondrial dynamics and apoptosis. Genes Dev 2008;22:1577–90. doi:10.1101/gad.1658508.
- [5] Sotgia F, Fiorillo M, Lisanti MP. Mitochondrial markers predict recurrence, metastasis and tamoxifen-resistance in breast cancer patients: Early detection of treatment failure with companion diagnostics. Oncotarget 2017;8:68730–45. doi:10.18632/oncotarget.19612.
- [6] Dickson P V, Gershenwald JE. Staging and prognosis of cutaneous melanoma.
 Surg Oncol Clin N Am 2011;20:1–17. doi:10.1016/j.soc.2010.09.007.
- [7] Foletto MC, Haas SE. Cutaneous melanoma: new advances in treatment. An Bras Dermatol 2014;89:301–10. doi:10.1590/abd1806-4841.20142540.
- [8] Curtin JA, Fridlyand J, Kageshita T, Patel HN, Busam KJ, Kutzner H, et al. Distinct Sets of Genetic Alterations in Melanoma. N Engl J Med 2005;353:2135– 47. doi:10.1056/NEJMoa050092.
- [9] Satzger I, Schaefer T, Kuettler U, Broecker V, Voelker B, Ostertag H, et al. Analysis of c-KIT expression and KIT gene mutation in human mucosal melanomas. Br J Cancer 2008;99:2065–9. doi:10.1038/sj.bjc.6604791.
- [10] Alaeddini M, Etemad-Moghadam S. Immunohistochemical profile of oral mucosal and head and neck cutaneous melanoma. J Oral Pathol Med 2015;44:234–8. doi:10.1111/jop.12235.
- [11] de-Andrade B-A-B, Toral-Rizo V-H, León J-E, Contreras E, Carlos R, Delgado-Azañero W, et al. Primary oral melanoma: a histopathological and immunohistochemical study of 22 cases of Latin America. Med Oral Patol Oral Cir Bucal 2012;17:e383-8. doi:10.4317/MEDORAL.17588.
- [12] Lourenço S V., Fernandes JD, Hsieh R, Coutinho-Camillo CM, Bologna S,Sangueza M, et al. Head and neck mucosal melanoma: A review. Am J

Dermatopathol 2014;36:578-87. doi:10.1097/DAD.00000000000035.

- [13] Lou Y, Li R, Liu J, Zhang Y, Zhang X, Jin B, et al. Mitofusin-2 over-expresses and leads to dysregulation of cell cycle and cell invasion in lung adenocarcinoma. Med Oncol 2015;32:132. doi:10.1007/s12032-015-0515-0.
- [14] Kannan A, Wells RB, Sivakumar S, Komatsu S, Singh KP, Samten B, et al. Mitochondrial Reprogramming Regulates Breast Cancer Progression. Clin Cancer Res 2016;22:3348–60. doi:10.1158/1078-0432.CCR-15-2456.
- [15] Ahn SY, Li C, Zhang X, Hyun Y-M. Mitofusin-2 Expression Is Implicated in Cervical Cancer Pathogenesis. Anticancer Res 2018;38:3419–26. doi:10.21873/anticanres.12610.
- [16] Sotgia F, Whitaker-Menezes D, Martinez-Outschoorn UE, Flomenberg N, Birbe RC, Witkiewicz AK, et al. Mitochondrial metabolism in cancer metastasis: visualizing tumor cell mitochondria and the "reverse Warburg effect" in positive lymph node tissue. Cell Cycle 2012;11:1445–54. doi:10.4161/cc.19841.
- [17] Soares CD, Borges CF, Sena-Filho M, Almeida OP de, Stelini RF, Cintra ML, et al. Prognostic significance of cyclooxygenase 2 and phosphorylated Akt1 overexpression in primary nonmetastatic and metastatic cutaneous melanomas. Melanoma Res 2017;27:448–56. doi:10.1097/CMR.00000000000368.
- [18] Kim ES, Park SJ, Goh M-J, Na Y-J, Jo DS, Jo YK, et al. Mitochondrial dynamics regulate melanogenesis through proteasomal degradation of MITF via ROS-ERK activation. Pigment Cell Melanoma Res 2014;27:1051–62. doi:10.1111/pcmr.12298.
- [19] Ward PS, Thompson CB. Metabolic Reprogramming: A Cancer Hallmark Even Warburg Did Not Anticipate. Cancer Cell 2012;21:297–308.

doi:10.1016/j.ccr.2012.02.014.

- [20] Li LZJ, Zhou R, Zhong T, Moon L, Kim EJ, Qiao H, et al. Predicting melanoma metastatic potential by optical and magnetic resonance imaging. Adv Exp Med Biol 2007;599:67–78.
- [21] Pal HC, Prasad R, Katiyar SK. Cryptolepine inhibits melanoma cell growth through coordinated changes in mitochondrial biogenesis, dynamics and metabolic tumor suppressor AMPKα1/2-LKB1. Sci Rep 2017;7:1498. doi:10.1038/s41598-017-01659-7.
- [22] Huebbers CU, Adam AC, Preuss SF, Schiffer T, Schilder S, Guntinas-Lichius O, et al. High glucose uptake unexpectedly is accompanied by high levels of the mitochondrial B-F1-ATPase subunit in head and neck squamous cell carcinoma. Oncotarget 2015;6:36172–84. doi:10.18632/oncotarget.5459.
- [23] Sun J, Yang Z, Miao X, Zou Q, Li J, Liang L, et al. ATP5b and β2-microglobulin are predictive markers for the prognosis of patients with gallbladder cancer. J Mol Histol 2015;46:57–65. doi:10.1007/s10735-014-9597-9.
- [24] Grupp K, Jedrzejewska K, Tsourlakis MC, Koop C, Wilczak W, Adam M, et al.
 High mitochondria content is associated with prostate cancer disease progression.
 Mol Cancer 2013;12:145. doi:10.1186/1476-4598-12-145.
- [25] Gonidi M, Athanassiadou A-M, Patsouris E, Tsipis A, Dimopoulos S, Kyriakidou V, et al. Mitochondrial UCP4 and bcl-2 expression in imprints of breast carcinomas: Relationship with DNA ploidy and classical prognostic factors. Pathol - Res Pract 2011;207:377–82. doi:10.1016/j.prp.2011.03.007.
- [26] Vlashi E, Lagadec C, Vergnes L, Reue K, Frohnen P, Chan M, et al. Metabolic differences in breast cancer stem cells and differentiated progeny. Breast Cancer Res Treat 2014;146:525–34. doi:10.1007/s10549-014-3051-2.

- [27] Farnie G, Sotgia F, Lisanti MP, Farnie G, Sotgia F, Lisanti MP, et al. High mitochondrial mass identifies a sub-population of stem-like cancer cells that are chemo-resistant. Oncotarget 2015;6:30472–86. doi:10.18632/oncotarget.5401.
- [28] Lee S, Kim S, Sun X, Lee J-H, Cho H. Cell cycle-dependent mitochondrial biogenesis and dynamics in mammalian cells. Biochem Biophys Res Commun 2007;357:111–7. doi:10.1016/j.bbrc.2007.03.091.
- [29] Smirnova E, Griparic L, Shurland DL, van der Bliek AM. Dynamin-related protein Drp1 is required for mitochondrial division in mammalian cells. Mol Biol Cell 2001;12:2245–56. doi:10.1091/mbc.12.8.2245.
- [30] Serasinghe MN, Wieder SY, Renault TT, Elkholi R, Asciolla JJ, Yao JL, et al. Mitochondrial division is requisite to RAS-induced transformation and targeted by oncogenic MAPK pathway inhibitors. Mol Cell 2015;57:521–36. doi:10.1016/j.molcel.2015.01.003.
- [31] Wieder SY, Serasinghe MN, Sung JC, Choi DC, Birge MB, Yao JL, et al. Activation of the Mitochondrial Fragmentation Protein DRP1 Correlates with BRAF(V600E) Melanoma. J Invest Dermatol 2015;135:2544–7. doi:10.1038/jid.2015.196.
- [32] Xu K, Chen G, Li X, Wu X, Chang Z, Xu J, et al. MFN2 suppresses cancer progression through inhibition of mTORC2/Akt signaling. Sci Rep 2017;7:41718. doi:10.1038/srep41718.
- [33] Narasimhan K, Kucuk O, Lin H-S, Heilbrun LK, Carron M, Venkatramanamoorthy R, et al. Sinonasal mucosal melanoma: a 13-year experience at a single institution. Skull Base 2009;19:255–62. doi:10.1055/s-0028-1115321.
- [34] Thompson LDR, Wieneke JA, Miettinen M. Sinonasal tract and nasopharyngeal

melanomas: a clinicopathologic study of 115 cases with a proposed staging system. Am J Surg Pathol 2003;27:594–611.

[35] Lourenço SV, Martin Sangüeza A, Sotto MN, Bologna SB, Giacomo TB di, Buim ME, et al. Primary oral mucosal melanoma: A series of 35 new cases from south America. Am J Dermatopathol 2009;31:323–30. doi:10.1097/DAD.0b013e3181a0d37c.

Supplementary Tables

| Clone | Dilution | Source | Positive control |
|-------|-------------------------------|----------------------------------------|-----------------------------------------------------------------------------------------------------------------------------|
| 113-1 | 1:100 | BioGenex ^a | Liver |
| B-5 | 1:50 | Santa Cruz | Oncocytoma |
| | | Biotechnology ^b | |
| XX-1 | 1:50 | Santa Cruz | Warthin's tumor |
| | | Biotechnology ^b | |
| | Clone 113-1 B-5 XX-1 | CloneDilution113-11:100B-51:50XX-11:50 | CloneDilutionSource113-11:100BioGenexaB-51:50Santa CruzBiotechnologybXX-11:50XX-11:50Santa CruzBiotechnologybBiotechnologyb |

Supplementary Table 1. List of primary antibodies, clone, dilutions, source and positive controls used in this study.

^aFremont, CA, USA; ^bDallas, TX, USA.

Supplementary Table 2. Multivariate analysis of risk factors for metastasis of head and neck cutaneous melanomas based on clinicopathological characteristics and AMT, FIS1 and MFN2 expressions.

| Factors | <i>P</i> -value | HR | 95% | 5 CI |
|---------------------|-----------------|--------|----------|----------|
| | | | Superior | Inferior |
| Ulceration | 0.025* | 4.292 | 1.198 | 15.374 |
| Breslow's thickness | 0.388 | 1.777 | 0.482 | 6.545 |
| Mitotic index | 0.439 | 0.620 | 0.185 | 2.081 |
| Clark's level | 0.776 | 1.522 | 0.084 | 27.614 |
| AMT | 0.112 | 3.856 | 0.730 | 20.359 |
| FIS1 | 0.262 | 0.149 | 0.005 | 4.146 |
| MFN2 | 0.003* | 38.883 | 3.421 | 441.930 |

HR, Hazard ratio; CI, confidence interval; AMT, mitochondrial antibody; FIS1, fission 1 protein; MFN2, mitofusin 2.

3 DISCUSSÃO

Visto que, em cada seção específica do artigo, os aspectos mais relevantes já foram discutidos, nesse espaço apenas ressaltaremos alguns resultados que consideramos importantes de cada capítulo e apresentaremos uma visão geral das principais conclusões desse trabalho.

Melanomas cutâneos e mucosos são neoplasias totalmente distintas do ponto de vista biológico. De fato, esses tumores são bastante agressivos e apresentam uma grande tendência a metástases (Braeuer *et al.*, 2014). Nesse estudo, nosso principal objetivo foi avaliar o papel biológico e o valor prognóstico de p-Akt1, COX-2 e marcadores mitocondriais nestas neoplasias. O primeiro capítulo deste estudo consistiu em avaliar o papel prognóstico da proteína p-Akt1 em melanomas cutâneos e mucosos. Estudos anteriores já haviam demonstrado que o AKT1 está associado a um fenótipo de melanoma mais metastático em modelo experimental (Cho *et al.*, 2015), no entanto, nós demonstramos pela primeira vez, que os melanomas mucosos (orais e nasais) apresentam maior expressão de p-Akt1 do que os melanomas cutâneos. Além disso, em conjunto com os dados demonstrados por estudos anteriores e, o fato de p-Akt1 ser um fator prognóstico independente para os melanomas mucosos pode indicar que novas terapias-alvo sejam desenvolvidas a partir desses resultados.

O segundo capítulo trata de um estudo determinou o papel da enzima COX-2 como um marcador prognóstico imuno-histoquímico importante para melanoma oral. Assim, estudos adicionais que estudem as diferentes vias de sinalização que ativam COX-2 em melanoma oral são necessários, para validar essa enzima como um potencial alvo terapêutico em melanomas mucosos.

No capítulo 3 e 4, apresentamos uma série de casos de melanomas orais amelanóticos e comparamos a expressão de COX-2 e de Ki67 nesses tumores e em melanomas orais convencionais, com deposição de melanina. Os melanomas amelanóticos parecem ter maior agressividade tumoral. Tanto os aspectos clínicos, como os aspectos patológicos confirmam essa hipótese. Os melanomas amelanóticos cutâneos também são considerados mais agressivos que os convencionais e vários fatores tem sido atribuídos a maior agressividade tumoral, incluindo que estes tumores podem representar uma variante pouco diferenciada e portanto, maior tendência a metástase e pior prognóstico (Thomas *et al.*, 2014; Moshe *et al.*, 2018).

No capítulo 5, foi estudada a expressão de marcadores mitocondriais em melanomas cutâneos e mucosos. Dentre eles, a proteína anti-mitocondrial (AMT), proteína de fissão 1 (FIS1) e proteína de fusão 2 (MFN2). A quantidade de mitocôndria, avaliada subjetivamente pela intensidade de AMT nas células tumorais parece desempenhar um papel importante na patogênese de melanomas; particularmente, os melanomas mucosos que apresentaram uma maior expressão de AMT. A proteína FIS1 teve maior expressão em melanomais orais e interessantemente pode ser considerada um fator prognóstico independente nesses tumores. Por outro lado, os melanomas nasais apresentaram alta expressão de MFN2 em comparação com os melanomas orais e cutâneos e também demonstrou relação com pior prognóstico em pacientes com melanomas cutâneos (Soares *et al.*, 2018).

Confirmamos em todos os capítulos da presente tese que os melanomas de mucosa oral e nasal tem um comportamento biológico distinto dos melanomas cutâneos. De fato, os melanomas mucosos apresentam uma expressão aumentada de p-Akt1, COX-2 e de marcadores mitocondriais comparado aos melanomas cutâneos. Esperamos que esses resultados, em conjunto, possam auxiliar no desenvolvimento de novas terapias-alvo objetivando ampliar o arsenal de opções terapêuticas dessas neoplasias malignas extremamente agressivas e com prognóstico sombrio.

4 CONCLUSÃO

O presente estudo confirmou que os melanomas de mucosa oral e nasal tem uma biologia muito distinta dos melanomas cutâneos convencionais. Características clínicas e patológicas como idade, tamanho do tumor, tipo celular e estadiamento podem influenciar negativamente a sobrevida dos pacientes. De maneira mais relevante, demonstramos que p-Akt1 e COX-2 podem ser usados como marcadores prognósticos em melanomas mucosos, e consequentemente podem desempenhar papel importante na biologia tumoral e no desenvolvimento de terapias-alvo.

REFERÊNCIAS*

Agarwal E, Brattain MG, Chowdhury S. Cell survival and metastasis regulation by Akt signaling in colorectal cancer. Cell Signal. 2013;25(8):1711–9.

Bishop KD, Olszewski AJ. Epidemiology and survival outcomes of ocular and mucosal melanomas: A population-based analysis. Int J Cancer. 2014;134(12):2961–71.

Braeuer RR, Watson IR, Wu CJ, Mobley AK, Kamiya T, Shoshan E, et al. Why is melanoma so metastatic? Pigment Cell Melanoma Res. 2014 Jan;27(1):19-36.

Cho JH, Robinson JP, Arave RA, Burnett WJ, Kircher DA, Chen G, et al. AKT1 Activation Promotes Development of Melanoma Metastases. Cell Rep. 2015;13(5):898–905.

Cicenas J. The potential role of Akt phosphorylation in human cancers. Int J Biol Markers. 2008;23(1):1–9.

Corazao-Rozas P, Guerreschi P, André F, Gabert P-E, Lancel S, Dekiouk S, et al. Mitochondrial oxidative phosphorylation controls cancer cell's life and death decisions upon exposure to MAPK inhibitors. Oncotarget. 2016;7(26):39473–85.

Curtin JA, Fridlyand J, Kageshita T, Patel HN, Busam KJ, Kutzner H, et al. Distinct Sets of Genetic Alterations in Melanoma. N Engl J Med. 2005;353(20):2135–47.

de Andrade BAB, León JE, Carlos R, Delgado-Azañero W, Mosqueda-Taylor A, de Almeida OP. Immunohistochemical expression of p16, p21, p27 and cyclin D1 in oral nevi and melanoma. Head Neck Pathol. 2012;6(3):297–304.

de Souza do Nascimento J, Carlos R, Delgado-Azañero W, Mosqueda Taylor A, de Almeida OP, Romañach MJ, et al. Immunohistochemical expression of cyclooxygenase-2 (COX-2) in oral nevi and melanoma. J Oral Pathol Med. 2016;45(6):440–3.

de-Andrade B-A-B, Toral-Rizo V-H, León J-E, Contreras E, Carlos R, Delgado-Azañero W, et al. Primary oral melanoma: a histopathological and immunohistochemical study of 22 cases of Latin America. Med Oral Patol Oral Cir Bucal. 2012;17(3):e383-8.

Fitzpatrick F. Cyclooxygenase Enzymes: Regulation and Function. Curr Pharm Des. 2005 23;10(6):577–88.

Franks ES, Briah R, Jones RA, Moorehead RA. Unique roles of Akt1 and Akt2 in IGF-IR mediated lung tumorigenesis. Oncotarget. 2016;7(3):3297–316.

Guo Z, Jiang JH, Zhang J, Yang HJ, Yang FQ, Qi YP, et al. COX-2 Promotes Migration and Invasion by the Side Population of Cancer Stem Cell-Like Hepatocellular Carcinoma Cells. Med (United States). 2015 Nov 1;94(44):e1806.

^{*}De acordo com as normas da UNICAMP/FOP, baseadas na padronização do International Committee of Medical Journal Editors - Vancouver Group. Abreviatura dos periódicos em conformidade com o PubMed.

Hashemi Goradel N, Najafi M, Salehi E, Farhood B, Mortezaee K. Cyclooxygenase-2 in cancer: A review. J Cell Physiol. 2019 May;234(5):5683-5699.

Hu H, Han T, Zhuo M, Wu LL, Yuan C, Wu L, et al. Elevated COX-2 Expression Promotes Angiogenesis Through EGFR/p38-MAPK/Sp1-Dependent Signalling in Pancreatic Cancer. Sci Rep. 2017 Dec 1;7(1).

Kuk D, Shoushtari AN, Barker CA, Panageas KS, Munhoz RR, Momtaz P, et al. Prognosis of Mucosal, Uveal, Acral, Nonacral Cutaneous, and Unknown Primary Melanoma From the Time of First Metastasis. Oncologist. 2016;21(7):848–54.

Leonardi GC, Falzone L, Salemi R, Zanghì A, Spandidos DA, Mccubrey JA, et al. Cutaneous melanoma: From pathogenesis to therapy (Review). Int J Oncol. 2018 Apr;52(4):1071-1080.

Li LZJ, Zhou R, Zhong T, Moon L, Kim EJ, Qiao H, et al. Predicting melanoma metastatic potential by optical and magnetic resonance imaging. Adv Exp Med Biol. 2007;599:67–78.

Liou G-Y, Storz P. Reactive oxygen species in cancer. Free Radic Res. 2010;44(5):479–96.

Liu J, Cheng Sun SH, Sun SJ, Huang C, Hu HH, Jin YB, et al. Phosph-Akt1 expression is associated with a favourable prognosis in pancreatic cancer. Ann Acad Med Singapore. 2010;39(7):548–7.

Lourenço S V., Fernandes JD, Hsieh R, Coutinho-Camillo CM, Bologna S, Sangueza M, et al. Head and neck mucosal melanoma: A review. Am J Dermatopathol. 2014 Jul;36(7):578-87.

Lugowska I, Teterycz P, Rutkowski P. Immunotherapy of melanoma. Wspolczesna Onkol. 2017;2(1A):61–7.

Luna-Ortiz K, Aguilar-Romero M, Villavicencio-Valencia V, Zepeda-Castilla E, Vidrio-Morgado H, Peteuil N, et al. Comparative study between two different staging systems (AJCC TNM VS BALLANTYNE'S) for mucosal melanomas of the head & neck. Med Oral Patol Oral Cir Bucal . 2016;21(4):e425–30.

Mochel MC, Duncan LM, Piris A, Kraft S. Primary mucosal melanoma of the sinonasal tract: a clinicopathologic and immunohistochemical study of thirty-two cases. Head Neck Pathol. 2015;9(2):236–43.

Moshe M, Levi A, Ad-El D, Ben-Amitai D, Mimouni D, Didkovsky E, et al. Malignant melanoma clinically mimicking pyogenic granuloma: comparison of clinical evaluation and histopathology. Melanoma Res. 2018 May;28(4):363–7.

Moya-Plana A, Aupérin A, Obongo R, Baglin AC, Ferrand FR, Baujat B, et al. Oncologic outcomes, prognostic factor analysis and therapeutic algorithm evaluation of head and neck mucosal melanomas in France. Eur J Cancer. 2019;123:1–10.

Soares C, De Lima Morais TM, Carlos R, Mariano FV, Altemani A, De Carvalho MGF, et al. Phosphorylated Akt1 expression is associated with poor prognosis in cutaneous, oral and sinonasal melanomas. Oncotarget. 2018;9(99):37291–304.

Soares CD, Borges CF, Sena-Filho M, Almeida OP de, Stelini RF, Cintra ML, et al. Prognostic significance of cyclooxygenase 2 and phosphorylated Akt1 overexpression in primary nonmetastatic and metastatic cutaneous melanomas. Melanoma Res. 2017;27(5):448–56.

Soares CD, de Lima Morais TM, Carlos R, de Almeida OP, Corrêa FVMB, de Almeida Milani Altemani AM, et al. Prognostic importance of mitochondrial markers in mucosal and cutaneous head and neck melanomas. Hum Pathol 2019 Mar;85:279-289.

Srinivasan S, Guha M, Kashina A, Avadhani NG. Mitochondrial dysfunction and mitochondrial dynamics-The cancer connection. Biochim Biophys Acta - Bioenerg. 2017;1858(8):602–14.

Suen DF, Norris KL, Youle RJ. Mitochondrial dynamics and apoptosis . Genes Dev. 2008 Jun 15;22(12):1577-90.

Thomas NE, Kricker A, Waxweiler WT, Dillon PM, Busam KJ, From L, et al. Comparison of Clinicopathologic Features and Survival of Histopathologically Amelanotic and Pigmented Melanomas. JAMA Dermatology. 2014 Dec 1;150(12):1306.

Wu Y, Kim J, Elshimali Y, Sarkissyan M, Vadgama J V. Activation of Akt1 accelerates carcinogen-induced tumorigenesis in mammary gland of virgin and post-lactating transgenic mice. BMC Cancer. 2014;14(1):266.

Xia M, Duan M-L, Tong J-H, Xu J-G. MiR-26b suppresses tumor cell proliferation, migration and invasion by directly targeting COX-2 in lung cancer. Eur Rev Med Pharmacol Sci . 2015;19(24):4728–37.
ANEXOS

ANEXO 1 - VERIFICAÇÃO DE ORIGINALIDADE E PREVENÇÃO DE PLÁGIO

ESTUDO COMPARATIVO DA EXPRESSÃO DE p-AKT1, COX-2 E MARCADORES MITOCONDRIAIS EM MELANOMAS CUTÂNEOS E MUCOSOS

| ORIGINALITY REPORT | | | | | |
|--------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------|-------------------------------------------------------------------|--|
| 1 SIMILA | 1% ARITY INDEX | 5% INTERNET SOURCES | 11% PUBLICATIONS | 3% STUDENT PAPERS | |
| PRIMAR | Y SOURCES | | | | |
| 1 | Ciro Dant Morais, R et al. "Pro markers i neck mel Publication | tas Soares, Thay Roman Carlos, O ognostic importai in mucosal and c anomas", Humai | ná Melo de Li slei Paes de A nce of mitocho utaneous hea n Pathology, 2 | ma 29 Imeida Indrial d and 018 | |
| 2 | de Souza do Nascimento, Juliana, Román Carlos, Wilson Delgado-Azañero, Adalberto Mosqueda Taylor, Oslei Paes de Almeida, Mário José Romañach, and Bruno Augusto Benevenuto de Andrade. "Immunohistochemical expression of cyclooxygenase-2 (COX-2) in oral nevi and melanoma", Journal of Oral Pathology and Medicine, 2015. Publication | | | án 29 berto da, Mário chemical 2) in oral athology | |
| 3 | Michael Moshe, Assi Levi, Dean Ad-El, Dan Ben-Amitai, Daniel Mimouni, Elena Didkovsky, Meora Feinmesser, Moshe Lapidoth. "Malignant | | | Dan 1 s kovsky, lalignant | |

melanoma clinically mimicking pyogenic

ANEXO 2 – PARECER CONSUBSTANCIADO DA COMISSÃO NACIONAL DE ÉTICA EM PESQUISA (CONEP)

COMISSÃO NACIONAL DE ÉTICA EM PESQUISA



PARECER CONSUBSTANCIADO DA CONEP

DADOS DO PROJETO DE PESQUISA

Título da Pesquisa: PROJETO INVESTIGATIVO PARA CARACTERIZAR O PERFIL MICROBIÔMICO E A RESPOSTA IMUNOLÓGICA EM MELANOMAS PRIMÁRIOS E METASTÁTICOS DE PELE, CAVIDADE ORAL, E NASOFARINGE

Pesquisador: CIRO DANTAS SOARES

Área Temática: A critério do CEP

Versão: 5

CAAE: 72077517.1.0000.5418

Instituição Proponente: Faculdade de Odontologia de Piracicaba - Unicamp

Patrocinador Principal: FUNDACAO DE AMPARO A PESQUISA DO ESTADO DE SAO PAULO

DADOS DO PARECER

Número do Parecer: 2.524.225

Situação do Parecer: Aprovado

ANEXO 3 – PÁGINA INICIAL DO ARTIGO 2.1

| www.oncotarget.com Onco | target, 2018, Vol. 9, (No. 99), pp: 37291-37304 | | | | |
|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--|--|--|--|
| Phosphorylated Akt1 expres prognosis in cutaneous, oral a | Research Paper ssion is associated with poor nd sinonasal melanomas | | | | |
| Ciro Soares ¹ , Thayná Melo de Lima M Mariano ^{1,3} , Albina Altemani ^{1,3} , Maria G Corrêa ⁵ , Rodrigo Ribas Dias dos Reis ⁶ Almeida ¹ and Jacks Jorge ¹ | lorais ¹ , Roman Carlos ² , Fernanda Viviane ioretti Freire de Carvalho ⁴ , Marcelo Brum , Luciana Schultz Amorim ⁷ , Oslei Paes de | | | | |
| ¹ Department of Oral Diagnosis, Area of Pathology, Piracica Brazil | ba Dental School, University of Campinas, Piracicaba, São Paulo, | | | | |
| ² Pathology Division, Centro Clínico de Cabeza y Cuello/Hospital Herrera Llerandi, Guatemala City, Guatemala | | | | | |
| ³ Department of Pathology, Faculty of Medical Sciences, University of Campinas, Campinas, São Paulo, Brazil | | | | | |
| ⁴ Private Pathology Service, Natal, Rio Grande do Norte, Brazil | | | | | |
| ⁵ Head and Neck Surgery Department - Oncology Center (CEON), Fornecedores de Cana Hospital, Piracicaba, São Paulo, Brazil | | | | | |
| ⁶ Oncology Surgery Department - Cancer Center (CECAN), | Santa Casa Hospital, Piracicaba, São Paulo, Brazil | | | | |
| 7Institute of Pathological Anatomy, Piracicaba, São Paulo, Brazil | | | | | |
| Correspondence to: Ciro Soares, email: ciro.dss@gmail.com | | | | | |
| Keywords: cutaneous melanomas; mucosal melanomas; p-Akt1; immunohistochemistry; prognosis | | | | | |
| Received: August 08, 2018 Accepted: November 26, 2 | 2018 Published: December 18, 2018 | | | | |
| Copyright: Soares et al. This is an open-access article distribut 3.0 (CC BY 3.0), which permits unrestricted use, distribution, of source are credited. | rted under the terms of the Creative Commons Attribution License and reproduction in any medium, provided the original author and | | | | |
| ABSTRACT | | | | | |
| Melanomas are highly aggressive occur most commonly in the skin. Occasi sinonasal mucous membranes. In this st the Phosphorylated Akt1 (p-Akt1) expri (CM), 34 oral cavity (OM), and 31 sinonas cutaneous melanomas, p-Akt1 was overe: of the sinonasal melanomas. In addition, with poorer cancer-specific survival in and sinonasal (P = .001) melanomas. M independent prognostic marker in oral (P patients. In conclusion, p-Akt1 overexp in mucosal melanomas and is significa As both mucosal and metastatic cutan p-Akt1 expression, these findings sugge | tumours derived from melanocytes, which onally, these tumours may appear in oral and udy, we performed a comparative analysis of ession in 144 patients affected by cutaneous al melanomas (SNM). Similar to the metastatic opressed in 17/34 of the oral cavity and 20/31 the p-Akt1-nuclear expression was associated o cutaneous ($P < .0001$), oral ($P < .0001$), lultivariate analysis showed p-Akt1 to be an = .041) and sinonasal ($P < .0001$) melanomas ression is an independent prognostic marker ntly up-regulated in sinonasal melanomas. eous melanomas showed high frequency of st that mucosal melanomas have a biological | | | | |

INTRODUCTION

Cutaneous melanomas represent about 1.6% of all cancers, and discreet advances in their treatment have been made over the last decades. Nevertheless, this tumour still remains deathly, especially the metastatic disease [1]. It is estimated that melanomas with lymph nodal metastases

behaviour, similar to the aggressive cutaneous melanomas.

are responsible for 59,782 global deaths, with mean overall survival rate of 13% in 5 years [2]. Controlling the advanced-stage disease is the major problem for the treatment of melanomas. Thus, for the development of targeted therapies, making progress in understanding the molecular factors that influence the aggressiveness of the metastatic melanomas is essential.

www.oncotarget.com

37291

Oncotarget

ANEXO 4 – PÁGINA INICIAL DO ARTIGO 2.5



Human Pathology Volume 85, March 2019, Pages 279-289



Original contribution

Prognostic importance of mitochondrial markers in mucosal and cutaneous head and neck melanomas * **

Ciro Dantas Soares DDs, MSc, PhD student ^a, ^A ^{III}, Thayná Melo de Lima Morais DDs, MSc, PhD student ^a, Roman Carlos DDs ^b, Oslei Paes de Almeida DDs, MSc, PhD ^a, Fernanda Viviane Mariano DDs, MSc, PhD ^a, ^c, Albina Altemani MD, PhD ^a, ^c, Maria Goretti Freire de Carvalho MD, PhD ^d, Marcelo Brum Corrêa MD ^e, Rodrigo Ribas Dias dos Reis MD ^f, Luciana Schultz Amorim MD, PhD ^g, Jacks Jorge DDs, MSc, PhD ^a

E Show more

https://doi.org/10.1016/j.humpath.2018.11.009

Get rights and content

Highlights

- High content of mitochondria may play an important role in melanoma pathogenesis.
- FIS1 expression predicts lower overall survival rates in oral melanoma patients.
- FIS1 expression is associated with vascular invasion in mucosal melanoma patients.
- MFN2 expression predicts metastasis in cutaneous melanoma patients.