



UNIVERSIDADE ESTADUAL DE CAMPINAS
FACULDADE DE ODONTOLOGIA DE PIRACICABA

JOÃO GABRIEL SILVA SOUZA

EFEITO DO PRÉ-BOCHECHO COM CÁLCIO NA POTENCIALIZAÇÃO
DA INIBIÇÃO DA DESMINERALIZAÇÃO DO ESMALTE POR
BOCHECHO FLUORETADO

EFFECT OF CALCIUM PRERINSE ON THE ENHANCEMENT OF
FLUORIDE MOUTHRINSE ON THE INHIBITION OF ENAMEL
DEMINERALIZATION

PIRACICABA

2016

JOÃO GABRIEL SILVA SOUZA

EFEITO DO PRÉ-BOCHECHO COM CÁLCIO NA POTENCIALIZAÇÃO
DA INIBIÇÃO DA DESMINERALIZAÇÃO DO ESMALTE POR
BOCHECHO FLUORETADO

EFFECT OF CALCIUM PRERINSE ON THE ENHANCEMENT OF
FLUORIDE MOUTHRINSE ON THE INHIBITION OF ENAMEL
DEMINERALIZATION

Dissertação apresentada à Faculdade de Odontologia de Piracicaba da Universidade Estadual de Campinas como parte dos requisitos exigidos para a obtenção do título de Mestre em Odontologia, na Área de Cariologia.

Dissertation presented to the Piracicaba Dental School of the University of Campinas in partial fulfillment of the requirements for the degree of Master in Dentistry, in Cariology area.

Orientadora: Profa. Dra. Livia Maria Andaló Tenuta

ESTE EXEMPLAR CORRESPONDE Á VERSÃO FINAL DA DISSERTAÇÃO DEFENDIDA PELO ALUNO JOÃO GABRIEL SILVA SOUZA, E ORIENTADA PELA PROFA. DRA. LIVIA MARIA ANDALÓ TENUTA

PIRACICABA

2016

Agência(s) de fomento e nº(s) de processo(s): FAPESP, 2014/00799-1

Ficha catalográfica
Universidade Estadual de Campinas
Biblioteca da Faculdade de Odontologia de Piracicaba
Marilene Girello - CRB 8/6159

So89e Souza, João Gabriel Silva, 1991-
Efeito do pré-bochecho com cálcio na potencialização da inibição da desmineralização do esmalte por bochecho fluoretado / João Gabriel Silva Souza. – Piracicaba, SP : [s.n.], 2016.

Orientador: Lívia Maria Andaló Tenuta.
Dissertação (mestrado) – Universidade Estadual de Campinas, Faculdade de Odontologia de Piracicaba.

1. Flúor. 2. Fluoreto de cálcio. 3. Biofilme. 4. Desmineralização do dente. I. Tenuta, Lívia Maria Andaló, 1976-. II. Universidade Estadual de Campinas. Faculdade de Odontologia de Piracicaba. III. Título.

Informações para Biblioteca Digital

Título em outro idioma: Effect of calcium prerinse on the enhancement of fluoride mouthrinse on the inhibition of enamel demineralization

Palavras-chave em inglês:

Fluorine

Calcium fluoride

Biofilms

Tooth demineralization

Área de concentração: Cariologia

Titulação: Mestre em Odontologia

Banca examinadora:

Lívia Maria Andaló Tenuta [Orientador]

Branca Heloísa de Oliveira Martins Vieira

Cassiano Kuchenbecker Rosing

Data de defesa: 26-02-2016

Programa de Pós-Graduação: Odontologia



UNIVERSIDADE ESTADUAL DE CAMPINAS
Faculdade de Odontologia de Piracicaba



A Comissão Julgadora dos trabalhos de Defesa de Dissertação de Mestrado, em sessão pública realizada em 26 de Fevereiro de 2016, considerou o candidato JOÃO GABRIEL SILVA SOUZA aprovado.

PROF^a. DR^a. LÍVIA MARIA ANDALÓ TENUTA

PROF^a. DR^a. BRANCA HELOÍSA DE OLIVEIRA MARTINS VIEIRA

PROF. DR. CASSIANO KUCHENBECKER ROSING

A Ata da defesa com as respectivas assinaturas dos membros encontra-se no processo de vida acadêmica do aluno.

AGRADECIMENTOS

Agradeço a Faculdade de Odontologia de Piracicaba (FOP) e a Universidade Estadual de Campinas (UNICAMP) por me proporcionar a oportunidade para condução deste projeto e formação adquirida. Agradeço também o Programa de Pós-graduação em Odontologia, em especial o Departamento de Ciências Fisiológicas, no qual esta inserida a área de Cariologia, pela estrutura, oportunidade e contribuição na minha formação profissional e realização desta pesquisa.

Agradeço a Fundação de Amparo à Pesquisa do Estado de São Paulo pela concessão da bolsa de mestrado durante o curso.

Agradeço imensamente a área de Cariologia da FOP-Unicamp por todo conhecimento e valores passados a mim.

A minha orientadora, a Profa. Livia Tenuta, meu muito obrigado por todos ensinamentos e valores passados a mim e pela orientação na condução desta pesquisa e importante contribuição na minha formação profissional. Quero expressar aqui a minha grande admiração pela profissional que ela é, seja como professora ou como pesquisadora, sou grato e honrado por ter tido a oportunidade de ter sido orientado por alguém tão dedicada e competente.

Agradeço ao professor Jaime Cury por toda contribuição e colaboração na concepção deste trabalho, e também por compartilhar com seus alunos valores e conhecimentos relacionados a pesquisa científica e, especificamente, aqueles relacionados a área de Cariologia. Meu muito obrigado também a professora Altair Antoninha Del Bel Cury por toda sua contribuição na concepção e execução desta pesquisa, agradeço também os muitos conselhos dados durante as coletas. Agradeço também a professora Cinthia Tabchoury por todo conhecimento passado a mim durante as disciplinas cursadas.

Agradeço aos técnicos do laboratório de Bioquímica Oral, Sr. Waldomiro e Alfredo, por todo apoio e amizade durante o mestrado.

Muito obrigado aos voluntários que colaboraram e tornaram possível a realização desta pesquisa: Renan, Lyvia, Della, Joao Leme, Carolina, Diego, Thais Priscila e Thiago.

Agradeço aos alunos de iniciação científica Mateus, Renan e Larissa, por toda a ajuda na condução da pesquisa e pelos momentos de descontração.

Meu muito obrigado aos amigos e colegas por todo apoio durante o mestrado e que fizeram essa experiência ainda melhor: Dayse, Juliana, Emanuele, Lina e Pablo. Muito obrigado também para as baianas inseridas na Cariologia: Aline e Samilly. Ao Diego, meu muito obrigado por toda ajuda e colaboração durante a execução do projeto. Em especial, quero agradecer todo o apoio e amizade as colegas que entraram no curso junto comigo: Carol, Barbara e Mayara, sem vocês este processo não teria sido o mesmo, muito obrigado por tudo!

Ao Antônio Pedro, muito obrigado por toda ajuda durante a execução do projeto, por todas as duvidas sanadas e pelos importantes conselhos dados.

Agradeço minha professora de graduação e orientadora durante minha iniciação científica e hoje uma parceira de pesquisa (que ainda tem muito a me ensinar), a professora Andrea Eleutério, por ter despertado em mim a paixão pela pesquisa científica.

Por fim, agradeço em especial a minha família. Aos meus pais (Ivani e Djalma) meu muito obrigado por todo apoio dado, educação e valores passados a mim, que sem dúvidas iram refletir na minha postura profissional, não teria chegado ate aqui sem o apoio de você, muito obrigado! Estendo estes agradecimentos também aos meus irmãos (Tamirys e Andrey) por todo apoio durante este processo. Aos meus sobrinhos (Arthur, Caio e Bernardo) que apesar de ainda não entenderem todo esse processo e sentirem falta do tio que mora em SP, são fundamentais na minha vida.

No mais, um muito obrigado a todos que contribuíram de alguma forma na condução desse projeto e na minha formação como Mestre.

RESUMO

Visando potencializar o efeito anticárie do fluoreto, aumentando sua retenção na cavidade bucal (saliva, biofilme dental) após o uso de dentífrícios ou bochechos, a utilização prévia de um bochecho contendo cálcio tem sido preconizada. Embora esse pré-bochecho, seguido do uso de solução fluoretada, esteja associado a um aumento significativo na concentração de fluoreto na saliva e biofilme dental, seu efeito anticárie ainda não foi comprovado experimentalmente. Assim, o objetivo deste trabalho foi verificar se um pré-bochecho contendo cálcio potencializa o efeito do fluoreto na redução da desmineralização do esmalte dentário. Para tal, foi realizado um experimento *in situ*, duplo cego e cruzado, conduzido durante 4 fases de 14 dias cada, no qual 10 voluntários utilizaram dispositivos intrabucais palatinos contendo blocos de esmalte bovino hígidos. Sacarose a 20% foi gotejada sobre os blocos na frequência de 8 vezes ao dia simulando um alto desafio cariogênico. Duas vezes por dia os voluntários realizaram bochechos com: (1) pré-bochecho placebo para cálcio (lactato de sódio) seguido de bochecho placebo para fluoreto (água purificada) (controle negativo); (2) pré-bochecho placebo para cálcio seguido de bochecho contendo 250 ppm F; (3) pré-bochecho placebo para cálcio seguido de bochecho contendo 1000 ppm F; (4) pré-bochecho com cálcio (150 mM de lactato de cálcio), seguido de bochecho contendo 250 ppm F. As variáveis-resposta estudadas foram: concentrações de fluoreto no fluido e na porção sólida do biofilme; nos blocos dentais: porcentagem de perda de dureza de superfície (%PDS), área de lesão de cárie e concentração de fluoreto fracamente e firmemente ligado. O grupo Ca + F não diferiu do grupo 1000 ppm F na perda mineral resultante (ANOVA, $p > 0,05$), mas ambos diferiram dos demais grupos ($p < 0,05$). A concentração de F na porção sólida do biofilme exposto ao tratamento Ca + F foi 22 vezes maior quando comparado ao uso de apenas F (250 ppm F), sendo significativamente mais alto do que todos os grupos ($p < 0,05$). No entanto, no fluido, o aumento de 2 vezes na concentração de F na comparação do grupo Ca+ F e 250 ppm F não foi significativo ($p > 0,05$). Não houve efeito significativo do grupo Ca+F no aumento das concentrações de F no esmalte. Conclui-se que o pré-bochecho com cálcio potencializou o efeito do fluoreto na redução da desmineralização, o que parece estar ligado à sua capacidade de aumentar a concentração de fluoreto no biofilme.

Palavras-chave: Flúor. Fluoreto de Sódio. Biofilmes. Desmineralização do dente.

ABSTRACT

A calcium (Ca) pre-rinse has been recommended to enhance the anticaries effect of fluoride by increasing its retention in the oral cavity (saliva, dental biofilm) after the use of toothpastes or mouthrinses. Although this pre-rinse, followed by the use of fluoride rinse, has been associated with a significant increase in the fluoride concentration in saliva and dental biofilm, its anticaries effect has not yet been demonstrated experimentally. Thus, the objective of this study was to verify if a calcium pre-rinse potentiates the effect of fluoride on the reduction of enamel demineralization. In a double-blind, crossover, in situ study in 4 phases of 14 days each, ten volunteers wore a palatal appliance containing enamel slabs. Sucrose solution (20%) was dripped onto the slabs 8 times/day to simulate a high cariogenic challenge. Twice/day volunteers rinsed with: (1) A Ca placebo pre-rinse (sodium lactate) followed by a distilled water rinse (negative control); (2) A placebo pre-rinse followed by a 250 ppm F rinse; (3) A placebo pre-rinse followed by a 1000 ppm F rinse; or (4) A Ca pre-rinse (150 mM Ca as Ca lactate), followed by a 250 ppm F rinse. Response variables were: in the biofilm: F concentrations in the fluid and solid; in enamel slabs: percentage of surface hardness loss (%SHL), cross-sectional hardness and F concentration in enamel (loosely-bound and firmly-bound). The Ca+ F group resulted in similar mineral loss than the 1000 ppm F group (ANOVA, $p > 0.05$), but both differed significantly from the other groups ($p < 0.05$). The F concentration in the solids of the biofilm treated with Ca + F was ≈ 22 times greater when compared to using only F (250 ppm F), being significantly higher than all other groups ($p < 0.05$). However, in the biofilm fluid, the twice increase in F concentration caused by the Ca + F treatment when compared to the 250 ppm F group was not significant. The Ca pre-rinse did not increase F concentrations in the enamel. In conclusion, the Ca pre-rinse followed by an F rinse enhances the effect of F on the inhibition of enamel demineralization, which seems to be linked to its ability to increase the F concentration in the biofilm.

Keywords: Fluoride; Calcium fluoride; Biofilm; Demineralization.

SUMÁRIO

1	INTRODUÇÃO	11
2	ARTIGO: <i>Ca pre-rinse before F rinse increases biofilm F concentration and reduces enamel demineralization</i>	13
3	CONCLUSÃO	29
	REFERÊNCIAS	30
	ANEXOS	
	Anexo 1 – Comprovante de submissão	32
	Anexo 2 – Dose-resposta modelo	33
	Anexo 3 – Análise biofilme	34

1 INTRODUÇÃO

A cárie dentária é conceituada como uma doença biofilme-açúcar dependente que resulta na perda mineral da estrutura dentária (Fejerskov, 2004; Fejerskov e Kidd, 2005). Em contrapartida a esses fatores negativos (biofilme e açúcar), o uso de fluoreto (F) tem sido indicado como fator determinante positivo para o controle da cárie dentária, reduzindo a desmineralização e potencializando a remineralização dental, culminando, portanto, com redução da progressão e aceleração da reversão de lesões de cárie (Ekstrand et al., 1990; Fejerskov e Kidd, 2005; Cury e Tenuta, 2008). O declínio de cárie observado nas últimas décadas tem sido atribuído ao amplo uso de fluoretos (Brathall et al., 1996). Sua eficácia anticárie tem sido demonstrada por revisões sistemáticas da literatura avaliando diferentes meios de utilização do F, como dentifrícios e bochechos fluoretados (Marinho et al., 2003a; 2003b).

O mecanismo pelo qual o fluoreto controla o desenvolvimento de lesões de cárie está baseado na sua manutenção nos fluidos bucais, principalmente no fluido do biofilme, onde ocorre o processo de perda mineral (Featherstone, 2000; Vogel, 2011). A concentração de fluoreto no fluido do biofilme aumenta logo após a exposição a agentes fluoretados (Cury et al., 2010; Vale et al., 2011), mas pode também ser mantida elevada pela liberação a partir de reservatórios de fluoreto formados durante a aplicação desses produtos. No biofilme, o fluoreto pode estar retido de duas formas: ligado a superfície bacteriana através de íons cálcio (chamado de reservatório biológico) (Rose et al., 1996); ou precipitado na forma de minerais, como fluoreto de cálcio ("CaF₂") e fluorapatita (Vogel, 2011). Quando na forma ligada a superfície bacteriana, a concentração de fluoreto será função da sua concentração no fluido do biofilme, liberando o íon mediante sua diminuição no biofilme (Vogel, 2011). No entanto, o fluoreto retido na forma de "CaF₂" pode ser um reservatório mais duradouro de F, liberando-o lentamente para agir nos processos de des- e remineralização dentária (Vogel, 2011). Porém, a formação de reservatórios do tipo "CaF₂" no biofilme dental não tem sido alcançada a partir do uso de veículos de uso diário, como os bochechos fluoretados (Vogel et al., 2010), devido a baixa concentração prévia de cálcio presente no fluido do biofilme (Vogel, 2011), que é inferior a concentração de fluoreto disponibilizada pelos bochechos.

Recentemente foi proposta na literatura uma forma de superar esta limitação e aumentar a formação desses reservatórios de fluoreto no biofilme dental (Vogel et al., 2008a, 2008b, 2014), principalmente na forma de "CaF₂" (Vogel et al., 2014). Este fluoreto de cálcio é formado no biofilme dental por um bochecho com solução contendo cálcio, visando aumentar a concentração de cálcio livre na cavidade bucal, previamente a utilização de um produto fluoretado, e consequentemente favorecer a formação de fluoreto de cálcio (Vogel et al., 2014). Estudos in vivo avaliando a capacidade de enriquecimento do biofilme dental com fluoreto utilizando esse protocolo (pré-bochecho com cálcio seguido de bochecho com solução fluoretada) demonstraram aumentos na concentração de fluoreto da ordem de 12 vezes no biofilme total e 5 vezes no fluido do biofilme (Vogel et al., 2008a; Vogel et al., 2014). Porém, o efeito anticárie deste protocolo no esmalte dental ainda não tem sido bem esclarecido na literatura.

Atualmente, os únicos estudos disponíveis que avaliaram o efeito de um pré-bochecho com cálcio na des- e remineralização do esmalte (Magalhães et al., 2007; Furlani et al., 2009) não comprovaram a superioridade desse tratamento em relação ao uso isolado do F. Estes estudos identificaram que a utilização prévia de produto contendo cálcio resulta em um aumento significativo da concentração de fluoreto no biofilme. No entanto, na avaliação de perda ou ganho de mineral, o grupo controle, sem F, não diferiu do grupo utilizando apenas fluoreto (Magalhães et al., 2007; Furlani et al., 2009), indicando falta de poder para diferenciar os grupos experimentais pela falta de dose-resposta ao fluoreto do modelo de estudo utilizado. Ressalta-se ainda que estes estudos utilizaram o fluoreto em forma de dentifício, porém, estudos prévios evidenciaram o aumento dos reservatórios de fluoreto de cálcio no biofilme dental a partir da utilização de um pré-bochecho com cálcio seguido do uso de solução fluoretada em forma de bochecho (Vogel et al., 2008a, 2008b, 2014). Dentifícios contêm detergentes, como o lauryl sulfato de sódio, que diminuem a formação de "CaF₂" (Barkvoll et al., 1988), o que pode ter influenciado os resultados e impede a avaliação isolada do efeito da combinação entre cálcio e fluoreto.

Assim, o objetivo deste trabalho foi verificar o potencial de inibição de desmineralização do esmalte de um pré-bochecho com cálcio, seguido de um bochecho fluoretado, utilizando um modelo de estudo que apresente dose-resposta ao F, bem como investigar os mecanismos que explicam seu efeito.

2 ARTIGO***Ca pre-rinse before F rinse reduces enamel demineralization – an in situ caries study**

João G.S. **SOUZA**^a, Livia M.A. **TENUTA**^a, Altair A. **DEL BEL CURY**^a, Diego F. **NÓBREGA**^a, Renan R. **BUDIN**^a, Mateus X. de **QUEIROZ**^a, Gerald L. **VOGEL**^b,
Jaime A. **CURY**^a

^a Piracicaba Dental School, UNICAMP, Piracicaba, Brazil

^b ADA Foundation, Volpe Research Center, Gaithersburg, USA

Short Title: Ca pre-rinse enhances the fluoride anticaries potential

Key words: dental caries, fluoride, rinse, dentifrice, dental plaque

Corresponding author:

Prof. Livia M A Tenuta

Piracicaba Dental School

CP 52

13414-903 Piracicaba, SP, Brazil

E-mail litenuta@fop.unicamp.br

Tel. +55 19 21065393

Declaration of Interests

G.L.V. (ADA Foundation) holds a patent for the Ca pre-application technique: U.S. Patent No. 9,011,823 on April 21, 2015. There are no other conflicts of interest with respect to the authorship and/or publication of this article.

* Artigo submetido na revista Caries Research (Anexo 1)

ABSTRACT

A calcium (Ca) pre-rinse before a fluoride (F) rinse has been shown to increase oral F levels. We tested the anti-caries effect of this combination in a dose-response in situ caries model. In a double-blind, crossover experiment, 10 volunteers carried enamel slabs in palatal appliances for 14 days, during which they rinsed twice/day with one of the four rinse combinations: (1) A placebo pre-rinse (sodium lactate) followed by a distilled water rinse (negative control); (2) A placebo pre-rinse followed by a 250 ppm F rinse; (3) A placebo pre-rinse followed by a 1000 ppm F rinse; (4) A Ca pre-rinse (150 mM Ca, as calcium lactate), followed by a 250 ppm F rinse. Sucrose solution was dripped onto the slabs 8x/day to simulate a high cariogenic challenge. The reduction of surface hardness loss (SHL) caused by the Ca pre-rinse used with the 250 ppm F rinse (%SHL=38.0±21.0) was significantly higher than that caused by the F rinse alone (%SHL=59.5±24.1) and similar to the 1000 ppm F rinse (%SHL=42.0±18.3). Compared to the 250 ppm F rinse, the Ca pre-rinse increased biofilm fluid F only twice (non-significant), however, in accordance with previous studies, it greatly increased F in biofilm solids (≈22x). The Ca pre-rinse had little effect on loosely- or firmly-bound enamel F. The results showed an increased level of protection against demineralization by the use of a Ca pre-rinse, which seems to be caused by the enhancement of F concentration in the biofilm.

Introduction

The worldwide decline in dental caries of recent decades [Nadanosvsky and Sheiham, 1995; Marthaler, 2004; Narvai et al., 2006] has been primarily attributed to widespread fluoride (F) use [Bratthall et al., 1996]. However, caries still remains as an important oral health problem [Kassebaum et al., 2015].

The anti-caries effect of F is primarily based on obtaining an elevated F concentration in oral fluids, especially in the fluid phase of the biofilm (biofilm fluid) [Vogel et al., 1990; Margolis and Moreno, 1990]. These increased fluid F levels are able to reduce dental demineralization and enhance remineralization. However, the long-term maintenance of such elevated oral F levels is problematic given the poor oral retention (about 2-3 h) of F following topical administration [Zero et al., 1988].

A Ca pre-application followed by topical F treatment has been shown to produce significantly, persistent increases in F levels in saliva and biofilm [Vogel et al., 2006, 2008a, 2008b, 2014; Chen et al., 2014]. Specifically, when compared to F rinse used alone, the Ca pre-application has been shown to increase F concentration by 12 times in the whole biofilm and 5 times in the biofilm fluid or salivary samples 1 h after the use [Vogel et al., 2006; 2008a]. In samples collected in the morning after evening application, the Ca pre-rinse increased salivary F concentration by 5 times [Vogel et al., 2008b]. Surprisingly, in view of these large increases in oral levels of F, previous *in situ* studies on the inhibition of enamel demineralization or enhancement of remineralization [Magalhães et al., 2007; Furlani et al., 2009] failed to demonstrate a significant effect of the Ca pre-rinse when used before a F dentifrice. The lack of effect can be explained by the fact that the *in situ* model used by these authors did not show a dose-response effect for different F concentrations. Furthermore the dentifrices used in these studies contain high concentration of sodium lauryl sulfate that may have precipitated or inactivated the Ca applied by the pre-rinse [Vogel et al., 2006].

The “Piracicaba *in situ* model” [Benelli et al., 1993; Cury et al., 1997; 2000] has been successfully used to test the anticaries effect of F toothpastes [Cury et al., 2001; Paes Leme et al., 2004; Cury et al., 2010] and products for professional F use [Vale et al., 2011; Calvo et al., 2012]. Unfortunately, the sensitivity of this model to high strength products [Proskin, 1995] has not been evaluated. Therefore, the aims of this study were to test the anticaries effect of the Ca pre-rinse using the

“Piracicaba in situ model” in a “dose response” experimental design that also demonstrates that his model was sensitive to the increased levels of oral fluoride anticipated with this procedure. We hypothesized that the use of a Ca pre-rinse followed by an F rinse would be able to increase the F concentrations in the biofilm fluid and solids, and would also reduce enamel demineralization.

Materials and Methods

Ethical aspects and volunteers

The study was approved by the Piracicaba Dental School (UNICAMP) Research and Ethics Committee (protocol 020/2014). All volunteers signed an informed consent, according to Brazilian ethical regulations. Volunteers, aged 18 to 30 years, were selected based on good general and oral health, as well as normal unstimulated (>0.1 mL/min) and stimulated (>0.7 mL/min) salivary flow rate. Included volunteers did not use antibiotics 1 month prior to the study and were not under orthodontic treatment.

Experimental design

In a double-blind, crossover, in situ design, 10 volunteers wore, in 4 phases of 14 days, a palatal appliance containing four sound bovine enamel slabs covered by a plastic mesh to allow the accumulation of biofilm. The slabs were treated with 20% sucrose solution eight times per day to simulate a high cariogenic challenge. In order to assess the enhancement of F effect by a Ca pre-rinse, and the dose-response of the model to the high levels of F anticipated with the pre-rinse, the following set of two-rinse treatments were tested: 1. a 150 mM sodium lactate pre-rinse (placebo solution for the Ca pre-rinse) followed by a purified water (0 ppm F) rinse (negative control); 2. a 150 mM sodium lactate pre-rinse followed by a 250 ppm F (as NaF) rinse (250 ppm F group); 3. a 150 mM sodium lactate pre-rinse followed by a 1000 ppm F rinse (1000 ppm F group); and 4. a 150 mM calcium lactate pre-rinse followed by a 250 ppm F rinse (Ca+ 250 ppm F group). The rinses were performed twice per day (in the morning and at night), after the oral hygiene with non-F toothpaste, for 1 min each. After 14 days, 10 h after the last exposure to sucrose and treatment solutions, the biofilm formed and the slabs were collected for analyses. Response variables were biofilm fluid F concentration, total concentration of F in the biofilm

solids, percent surface hardness loss (%SHL), area of surface hardness loss (ΔS), and F concentration in enamel as loosely or firmly-bound F. Wash-out periods of at least 7 days were allowed between each phase. ANOVA followed by tukey test was used to compare all groups.

Preparation of enamel slabs and appliance

Bovine teeth, stored in formaldehyde solution 2% (pH 7.0) for at least 30 days [Cury et al., 2000], were used to prepare 4 x 4 x 2 mm enamel slabs. These were polished flat and selected for the study based on their baseline Knoop hardness values [Cury et al., 1997] (average 338.5 ± 17.8), determined using a Future Tech FM-7 microhardness tester, as previously described [Cury et al., 2003]. The slabs were accommodated on cavities in custom-made, acrylic resin appliances, and covered by a plastic mesh to promote biofilm accumulation [Hara et al., 2003].

Treatments

In order to simulate a high cariogenic challenge, volunteers dripped a 20% sucrose solution extra orally on the slabs eight times per day (8:00, 9:30, 11:00, 13:30, 15:00, 16:30, 18:00, 19:30 h) and replaced the appliance in the mouth, after 5 min [Ccahuana-Vásquez et al., 2007]. The Ca pre-rinse was prepared from calcium lactate (Sigma® - C8356) at concentration of 150 mM. Its placebo was a sodium lactate solution (Sigma® L7022) at concentration of 150 mM. The F rinse was prepared from sodium F (Merck® - 106449) at concentrations of 250 and 1000 ppm F (13.16 and 52.63 mM F, respectively). These set of rinses (15 mL each) were used in the morning, before the first sucrose exposure, and at night, after the last sucrose exposure, after oral hygiene with non-F toothpaste. A 30-min period was allowed to pass between morning and evening toothbrushing and the rinses to avoid the influence of the detergent present in toothpaste in the treatments (rinses). Each rinsing solution was sequentially swished in the mouth for 1 minute and then expectorated, with the second rinse being used immediately after the first.

Biofilm analyses

On the 15th day of each experimental phase, in the morning, 10 h after the last rinse, the biofilm and slabs were collected for analysis. The plastic meshes were removed and the biofilm samples covering the enamel slabs were collected, and

immediately immersed in mineral oil to avoid evaporation. The biofilm fluid and solids were separated by centrifugation and the fluid collected using a micropipette, as previously described [Vogel et al., 1992, 2000, 2008a]. A F ion-specific electrode adapted for microanalysis was used to determine F concentration in the biofilm fluid [Vogel et al., 1983, 1990, 2000; Tenuta et al., 2006]. After the fluid separation, the biofilm solids were extracted twice with 0.5 M HCl [Cury et al., 2000; Tenuta et al., 2006] for 60 min. The separate extract was neutralized with NaOH, diluted with TISAB and analyzed using the same methodology used for the fluid [Vogel et al., 1990, 2000; Tenuta et al., 2006]. The concentration in the 2 extracts was summed.

Enamel analyses

The slabs were collected for determination of enamel demineralization and F concentration in enamel as loosely or firmly-bound F. Of the four slabs present in each appliance: (1) all were analyzed for the surface hardness; (2) two were also analyzed for cross-sectional microhardness; (3) the other two slabs were used to determine F concentration in enamel.

Enamel demineralization determination

Surface hardness of all enamel slabs was re-assessed at 100- μ m distance from the initial indentations to determine enamel demineralization. The percentage of surface hardness loss (%SHL) was calculated by: $(\text{baseline hardness} - \text{final hardness}) / \text{baseline hardness} \times 100$.

Two of these slabs were further sectioned to assess cross-sectional hardness, by measuring hardness at 14 distances (from 10 to 300 μ m) from the enamel surface. The hardness at each depth was plotted in a graph and the area of hardness loss (ΔS) was calculated using the trapezoidal rule [Cury et al., 2010].

Enamel loosely and firmly-bound F determination

Two slabs were isolated with wax, leaving only the surface area free. They were then immersed in 1M KOH and agitated for 24 h [Caslavská et al., 1975] for extraction of enamel loosely-bound F. After that, slabs were treated with 0.5 M HCl and agitated for 30 s for extraction of enamel firmly-bound F [Koo and Cury, 1998]. F concentrations in both extracts were determined using an ion-specific electrode, after appropriate neutralization and buffering with TISAB II, using standards prepared in

the same conditions of the samples. The results for both loosely and firmly-bound F were calculated as $\mu\text{g F/cm}^2$.

Statistical analysis

The normality of errors and homocedasticity of data was checked for each response variable, and when necessary the data were transformed: the area of hardness loss (ΔS), F in the biofilm fluid, F in the biofilm solids and F in enamel (loosely- and firmly-bound) were transformed using \log_{10} . Groups were compared using ANOVA, considering the volunteers as statistical blocks, followed by post hoc Tukey test. For the transformed data, the geometric mean and 95% confidence intervals (calculated from the transformed data and then converted to the non-log scale) are presented. One volunteer did not complete one experimental phase while in the control group, so for all variables $n=9$ for this group. The SAS system 9.4 (SAS Institute, Cary, USA) was used in these calculations and a significance level of 5% was adopted.

Results

Significant differences in the %SHL were found among groups 0, 250 and 1000 ppm F (fig. 1a). The %SHL in the Ca + 250 ppm F group was not different from that in the 1000 ppm F group, but was significantly higher than in the 250 ppm F rinse (fig. 1a). In relation to area of hardness loss (ΔS) no difference was observed between any of the F rinse groups ($p>0.05$), but all F rinse groups resulted in significantly lower ΔS values when compared to the negative control group ($p<0.05$) (fig. 1b).

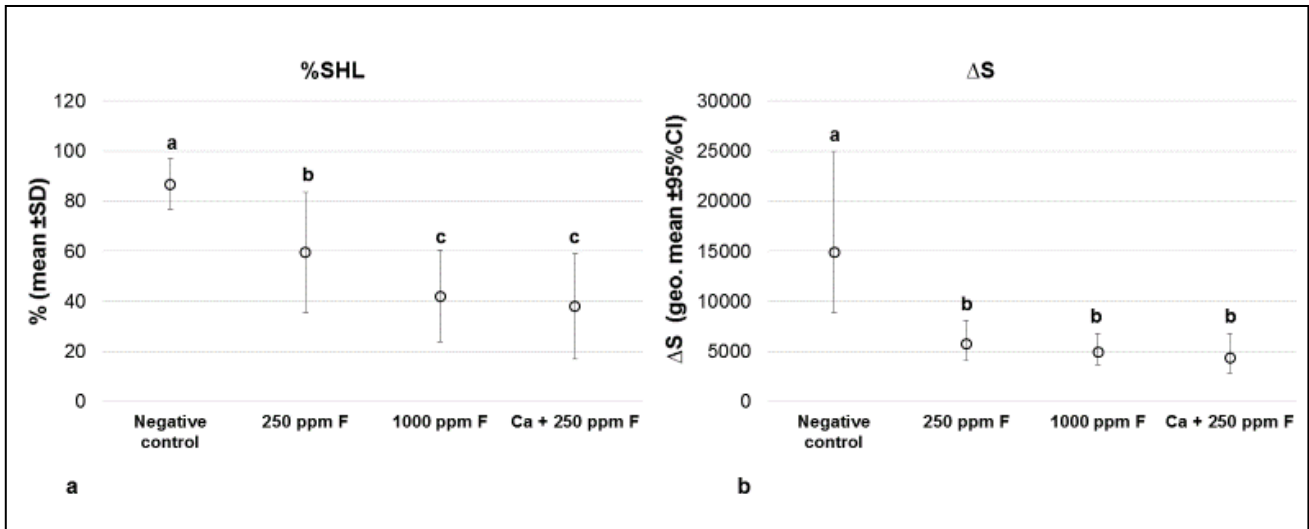


Figure 1 – Analysis of enamel demineralization, according to the treatment groups. (a) %SHL (mean ± SD); (b) ΔS (geometric mean ± 95% CI). Different letters represent statistical differences ($p < 0.05$).

All F containing rinses increased F concentrations in the fluid relative to the placebo, non-F rinse. However, although the Ca pre-rinse increased biofilm F (~2x), only the 1000 ppm F rinse produced a significant (4x) increase from above the 250 ppm F rinse. The Ca pre-rinse increased F in the biofilm solids (22x above the 250 ppm F rinse), being significantly higher than the other groups, including the 1000 ppm F rinse (fig. 2).

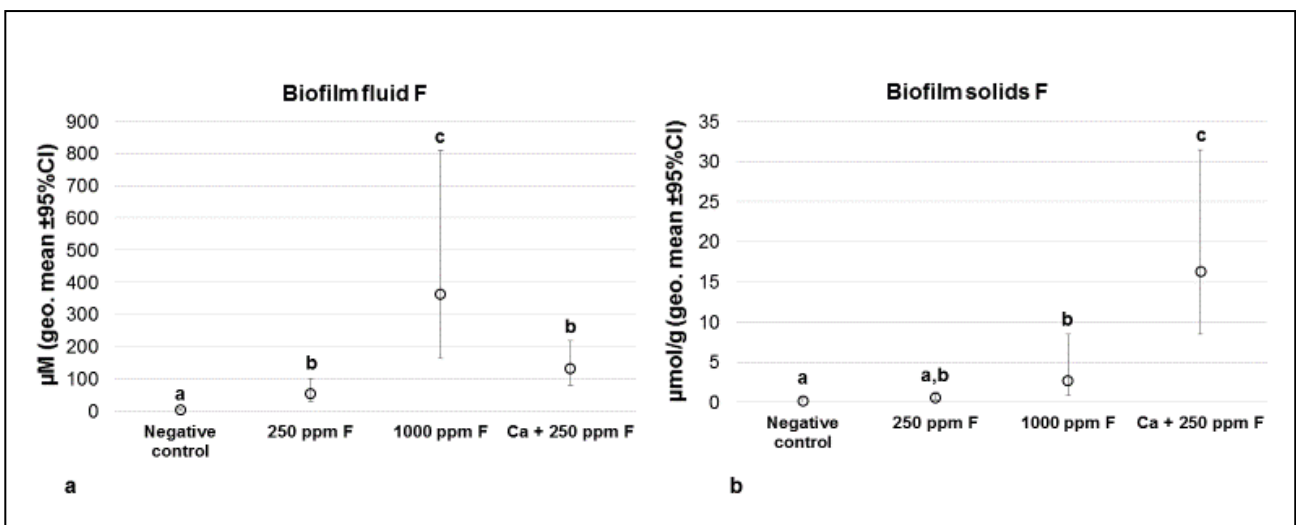


Figure 2 – Analysis of the biofilm fluid and solids (geometric mean ± 95% CI), according to the treatment groups. (a) F in the biofilm fluid; (b) total F in the biofilm

solids. Different letters represent statistical differences ($p < 0.05$). For F in the biofilm fluid, one outlier (value = $7.70 \mu\text{M}$) was removed from the 1000 ppm F group ($n=9$).

The loosely-bound F concentration in enamel increased significantly with the increasing F rinse concentration, however only the 1000 ppm F rinse was significantly different from the other F-containing rinses. The firmly-bound F concentrations were all significantly higher than the negative control (fig. 3), with the 250 ppm F and 1000 ppm F groups having the highest values. The Ca pre-rinse group did not differ significantly from the 1000 ppm F group in this concentration (fig. 3).

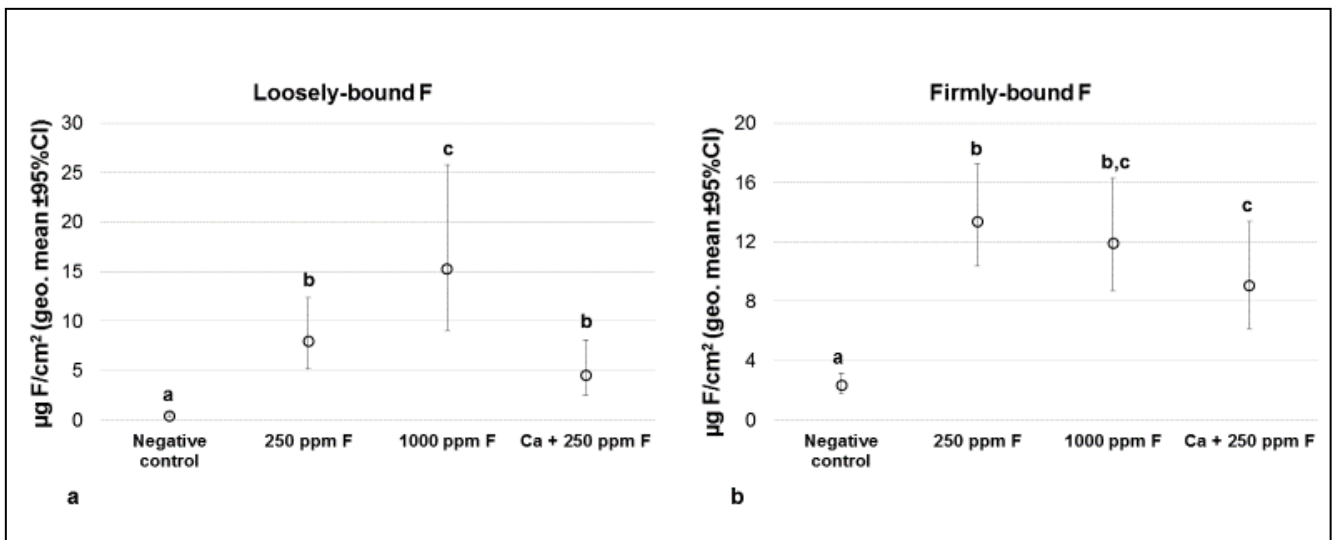


Figure 3 – Analysis of loosely-bound (a) and firmly-bound (b) fluoride in enamel (geometric mean \pm 95% CI), according to the treatment groups. Different letters represent statistical differences ($p < 0.05$).

Discussion

As shown in Fig. 1a, with respect to surface hardness, the in situ model was sensitive to increases in F concentration to at least 1000 ppm F. This provides confidence in the data of this figure demonstrating that the Ca + 250 ppm F group significantly reduce the %SHL, when compared with the 250 ppm F rinse alone: the inhibition of mineral loss was similar to that found with the 1000 ppm F rinse. Surface hardness data is taken as the only indicator of mineral loss [Cury et al., 2003; 2010; Kusano et al., 2011; Vale et al., 2011] in this study since, as shown in Fig. 1b, the area of hardness loss was not sensitive to levels above 250 ppm F.

Surprising, given its central role in the mechanism of F in reducing caries [Vogel et al., 1990; Margolis and Moreno, 1990; Vogel 2011], overnight biofilm fluid F levels only increased about twice with the use of the Ca pre-rinse, and this increase was not significantly higher ($p=0.068$) than the 250 ppm F rinse alone. Only the 1000 ppm F rinse produced a 4 times, significant increase in this parameter. The fact that nearly 10 h elapsed since the rinses suggest that biofilm fluid F should be highly elevated by the Ca pre-rinse procedure at shorter time periods: as noted above, 1 h biofilm samples collected from human teeth [Vogel et al., 2008a] showed nearly 5 x increase.

The ~22x increase in the biofilm solids F (Fig. 2b) with use of the Ca pre-rinse is greater than then the 6x increase observed in samples also collected from an in situ model when the Ca pre-rinse was used with a dentifrice [Magalhães et al., 2007]. Differences in protocol, such as the use of extra oral or intra oral rinses, and the presence of calcium binding moieties in dentifrices [Vogel et al., 2006], appear to explain these differences. In any case, the very large increase in biofilm F in both these studies, as well as the 12x increase in this parameter found in samples of biofilm collected from human teeth 1 h after use of this same Ca pre-rinse/250 ppm F rinse combination [Vogel et al., 2008a], appear to explain the high level of protection against demineralization observed here. Such biofilm F trapping is consistent with the fixing of fluoride by Ca mechanism by which the rinse has been postulated to increase oral levels of fluoride [Rose et al., 1996; Vogel et al., 2008a, 2008b, 2011, 2014]. Although the nature of such biofilm reservoirs requires further investigation, recent studies have suggested that that unlike a conventional F rinse [Vogel et al., 2010], the Ca pre-rinse has been shown to deposit nearly 30% of its F as calcium fluoride-like precipitates [Vogel et al., 2014]. With the 1000 ppm F rinse, the higher levels of biofilm fluid F coupled with a lower level of F in the biofilm solids suggest that the biofilm F deposits induced by this rinse are more labile than those produced by the Ca pre-rinse. Specifically, they may be in the form of calcium mediated biological/bacterial bound F reservoirs [Rose et al., 1996] that have previously been found to be the form that F is held in plaque 1 h after use of a 250 ppm F rinse [Vogel et al., 2010].

Loosely-bound F may be an important source of F on the enamel surface which can slowly release F to oral fluids to induce remineralization and reduce demineralization during periods of caries attack, but it is mostly formed after the use

of professional methods of F use [Tenuta et al., 2008]. Firmly-bound F appears to represent the F incorporated into the tooth mineral, mainly as a result of re-precipitation of fluoridated apatites from enamel mineral dissolved during the periods of low pH in the presence of F [Margolis and Moreno, 1990]. Both these reservoirs were increased by the F regimen used here (fig. 3). However, in spite of the fact that the Ca + 250 ppm F group greatly reduced demineralization, this rinse combination did not significantly enhance either of these F reservoirs compared with the 250 ppm F alone. In fact, the Ca pre-rinse appears to decrease these parameters (significantly in the case of firmly-bound F). This suggests that, in accordance with the very high levels of biofilm F observed with the Ca + 250 ppm F group, the F in this combination of rinses appears to have been trapped in the plaque and hence did not reach the surface of the enamel to form these F reservoirs. Thus, when a conventional F rinse is being used, at least some of the fluid F at tooth surface may be released from loosely-bound enamel F, but when a Ca pre-rinse/F rinse combination is used, the very high levels of F found in the biofilm appears to be the primary source of biofilm fluid F.

Some inhibition of bacterial metabolism by the high F concentrations found in the biofilm fluid could also explain the lower demineralization in the Ca pre-rinse/F rinse group, with no increase in firmly-bound F by re-precipitation. However, the F concentration found in the biofilm fluid in this group, although high, is not enough to significantly inhibit bacterial metabolism [Bradshaw et al., 2002]. It may be, however, that immediately after the F rinses and for some time after that, the F concentration reached inhibitory concentration of at least 10 ppm F (526.3 μM); this would be important for the morning rinse, which was performed before the first sugar exposure of the day. However, such effect would be even higher in the 1000 ppm F group than in the Ca pre-rinse + F group, considering that higher resting fluid F values were observed for the first.

In conclusion, the results showed that a Ca pre-rinse greatly increases the deposition of F in the biofilm, and that this increased biofilm F deposition appears to be responsible for the large decrease in tooth enamel demineralization observed in this study after use of this pre-rinse.

Acknowledgements

We thank the volunteers for their valuable participation, FAPESP (São Paulo Research Foundation) for the scholarship granted to J.G.S.S. (2014/00799-1), and CNPq for the scholarships granted to R.R.B. and M.X.Q. This study was partially funded by FUNCAMP (4887.1). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Author Contributions

Conceived and designed the experiment: L.M.A.T., A.A.D.B.C., G.L.V., J.A.C.; performed the experiment: J.G.S.S., L.M.A.T., A.A.D.B.C., D.F.N., R.R.B., M.X.Q.; analyzed the data: J.G.S.S., L.M.A.T.; wrote the draft manuscript: J.G.S.S., L.M.A.T.; reviewed and approved the final manuscript: all authors

References

Bradshaw DJ, Marsh PD, Hodgson RJ, Visser JM: Effects of glucose and fluoride on competition and metabolism within in vitro dental bacterial communities and biofilms. *Caries Res* 2002;36:81-86.

Bratthall D, Hänsel-Petersonn G, Sundberg H: Reasons for the caries decline: what do experts believe? *Eur J Oral Sci* 1996;104:416-422.

Benelli EM, Serra MC, Rodrigues AL Jr, Cury JA: In situ anticariogenic potential of glass ionomer cement. *Caries Res* 1993;27:280-284.

Calvo AF, Tabchoury CP, Del Bel Cury AA, Tenuta LM, da Silva WJ, Cury JA: Effect of acidulated phosphate fluoride gel application time on enamel demineralization of deciduous and permanent teeth. *Caries Res* 2012;46:31-37.

Caslavska V, Moreno EC, Brudevold F: Determination of calcium fluoride formed from in vitro exposure of human enamel of fluoride solutions. *Arch. Oral Biol* 1975;20:333-339.

Ccahuana-Vásquez RA, Tabchoury CPM, Tenuta LMA, Del Bel Cury AA, Vale GC, Cury JA: Effect of frequency of sucrose exposure on dental biofilm composition and enamel demineralization in the presence of fluoride. *Caries Res* 2007;41:9-15.

Chen MS, Strömberg E, Vogel GL, Sandborgh-Englund G: A randomized controlled trial: the efficacy of fluoride rinse combined with calcium pre-rinse to increase overnight salivary fluoride. *Acta Odontol Scand* 2014;72:557-560.

Cury JA, Rebelo MAB, Del Bel Cury AA: In situ relationship between sucrose exposure and the composition of dental plaque. *Caries Res* 1997; 31:356-360.

Cury JA, Rebelo MAB, Del Bel Cury AA, Derbyshire MTV, Tabchoury CPM: Biochemical Composition and Cariogenicity of Dental Plaque Formed in the Presence of Sucrose or Glucose and Fructose. *Caries Res* 2000;34:491-497.

Cury JA, Hashizume LN, Del Bel Cury AA, Tabchoury CP: Effect of dentifrice containing fluoride and/or baking soda on enamel demineralization/remineralization: an in situ study. *Caries Res* 2001;35:106-110.

Cury JA, Francisco SB, Simões GS, Del Bel Cury AA, Tabchoury CPM: Effect of a Calcium Carbonate-Based Dentifrice on Enamel Demineralization in situ. *Caries Res* 2003;37:194-199.

Cury JA, do Amaral RC, Tenuta LMA, Del Bel Cury AA, Tabchoury CPM: Low-fluoride toothpaste and deciduous enamel demineralization under biofilm accumulation and sucrose exposure. *Eur J Oral Sci* 2010;118:370-375.

Furlani TA, Magalhães AC, Iano FG, Cardoso VES, Delbem ACB, Buzalaf MAR: Effect of Calcium Pre-rinse and Fluoride Dentifrice on Enamel and on Dental Plaque Formed In Situ. *Oral Health Prev Dent* 2009;7:23–28.

Hara AT, Queiroz CS, Paes Leme AF, Serra MC, Cury JA: Caries progression and inhibition in human and bovine root dentine in situ. *Caries Res* 2003;37:339-344.

Kassebaum NJ, Bernabé E, Dahiya M, Bhandari B, Murray CJ, Marcenes W: Global burden of untreated caries: a systematic review and metaregression. *J Dent Res* 2015;94:650-658.

Koo RH, Cury JA: Soluble calcium/SMFP dentifrice: effect on enamel fluoride uptake and remineralization. *Am J Dent* 1998;11:173-176.

Kusano SC, Tenuta LM, Cury AA, Cury JA: Timing of fluoride toothpaste use and enamel-dentin demineralization. *Braz Oral Res* 2011;25:383-387.

Magalhães AC, Furlani TA, Italiani FM, Iano FG, Delbem ACB, Buzalaf MAR: Effect of calcium pre-rinse and fluoride dentifrice on remineralisation of artificially demineralised enamel and on the composition of the dental biofilm formed in situ. *Arch Oral Biol* 2007;52:1155-1160.

Margolis HC, Moreno EC: Physicochemical perspectives on the cariostatic mechanisms of systemic and topical fluorides. *J Dent Res* 1990;69:606–613.

Marthaler TM: Changes in dental caries 1953-2003. *Caries Res* 2004;38:173-181.

Nadanovsky P, Sheiham A: Relative contribution of dental services to the changes in caries levels of 12-year-old children in 18 industrialized countries in the 1970s and early 1980s. *Community Dent Oral Epidemiol* 1995;23:331-339.

Narvai PC, Frazão P, Roncalli AG, Antunes JL: Dental caries in Brazil: decline, polarization, inequality and social exclusion. *Rev Panam Salud Publica* 2006;19:385-393.

Paes Leme AF, Dalcico R, Tabchoury CP, Del Bel Cury AA, Rosalen PL, Cury JA: In situ effect of frequent sucrose exposure on enamel demineralization and on plaque composition after APF application and F dentifrice use. *J Dent Res* 2004;83:71-75.

Proskin HM: Statistical considerations related to the use of caries model systems for the determination of clinical effectiveness of therapeutic agents. *Adv Dent Res* 1995;9:270-278

Rose RK, Shellis RP, Lee AR: The role of cation bridging in microbial fluoride binding. *Caries Res* 1996;30:458-464.

Tenuta LM, Del Bel Cury AA, Bortolin MC, Vogel GL, Cury JA: Ca, Pi, and F in the fluid of biofilm formed under sucrose. *J Dent Res* 2006;85:834-838.

Tenuta LM, Cerezetti RV, Del Bel Cury AA, Tabchoury CP, Cury JA: Fluoride release from CaF₂ and enamel demineralization. *J Dent Res*. 2008;87:1032-1036.

Vale GC, Tabchoury CP, Del Bel Cury AA, Tenuta LM, ten Cate JM, Cury JA: APF and dentifrice effect on root dentin demineralization and biofilm. *J Dent Res* 2011;90:77-81.

Vogel GL, Chow LC, Brown WE: A microanalytical procedure for the determination of calcium, phosphate and fluoride in enamel biopsy samples. *Caries Res* 1983;17:23-31.

Vogel GL, Carey CM, Chow LC, Tatevossian A: Micro-analysis of plaque fluid from single-site fasted plaque. *J Dent Res* 1990;69:1316–1323.

Vogel GL, Carey CM, Ekstrand J: Distribution of fluoride in saliva and plaque fluid after a 0.048 mol/L NaF rinse. *J Dent Res* 1992;71:1553-1557.

Vogel GL, Zhang Z, Chow LC, Carey CM, Schumacher GE, Banting DW: Effect in vitro acidification on plaque fluid composition with and without a NaF or a controlled-release fluoride rinse. *J Dent Res* 2000;79:983-990.

Vogel GL, Shim D, Schumacher GE, Carey CM, Chow LC, Takagi S: Salivary fluoride from fluoride dentifrices or rinses after use of a calcium pre-rinse or calcium dentifrice. *Caries Res* 2006;40:449-454.

Vogel GL, Schumacher GE, Chow LC, Takagi S, Carey CM: Ca pre-rinse greatly increases plaque and plaque fluid F. *J Dent Res* 2008a;87:466-469.

Vogel GL, Chow LC, Carey CM. Calcium pre-rinse greatly increases overnight salivary fluoride after a 228 ppm fluoride rinse. *Caries Res* 2008b;42:401-404.

Vogel GL, Tenuta LM, Schumacher GE, Chow LC: No calcium-fluoride-like deposits detected in plaque shortly after a sodium fluoride mouthrinse. *Caries Res* 2010;44:108-115.

Vogel GL: Oral fluoride reservoirs and the prevention of dental caries. *Monogr Oral Sci* 2011;22:146-157.

Vogel GL, Tenuta LM, Schumacher GE, Chow LC: A calcium pre-rinse required to form calcium fluoride in plaque from a sodium fluoride rinse. *Caries Res* 2014;48:174-178.

Zero DT, Fu J, Espeland MA, Featherstone JD: Comparison of fluoride concentrations in unstimulated whole saliva following the use of a fluoride dentifrice and a fluoride rinse. *J Dent Res* 1988;67:1257-1262.

3 CONCLUSÃO

O presente estudo demonstrou que a utilização de um pré-bochecho com cálcio, antes do uso de bochecho fluoretado, é capaz de reduzir significativamente a perda mineral do esmalte submetido a um alto desafio cariogênico em relação ao uso isolado do bochecho fluoretado na mesma concentração, o que está ligado a sua capacidade de aumentar a concentração de fluoreto no biofilme dental.

REFERÊNCIAS

- Barkvoll P, Rølla G, Lagerlöf F. Effect of sodium lauryl sulfate on the deposition of alkali-soluble fluoride on enamel in vitro. *Caries Res.* 1988;22(3):139-144.
- Bratthall D, Hänsel-Petersonn G, Sundberg H. Reasons for the caries decline: what do experts believe? *Eur J Oral Sci.* 1996; 104:416–422.
- Cury JA, Tenuta LMA. How to Maintain a Cariostatic Fluoride Concentration in the Oral Environment. *Adv Dent Res* 2008;20:13-16.
- Cury JA, do Amaral RC, Tenuta LM, Del Bel Cury AA, Tabchoury CP. Low-fluoride toothpaste and deciduous enamel demineralization under biofilm accumulation and sucrose exposure. *Eur J Oral Sci.* 2010;118(4):370-5.
- Ekstrand J, Spak CJ, Vogel G. Pharmacokinetics of fluoride in man and its clinical relevance. *J Dent Res* 1990;69:550-555.
- Featherstone JD. The science and practice of caries prevention. *J Am Dent Assoc.* 2000;131:887–899.
- Fejerskov O, Kidd E. *Cárie dentária: a doença e seu tratamento clínico.* Editora Santos: São Paulo; 2005.
- Fejerskov O. Changing paradigms in concepts on dental caries: consequences for oral health care. *Caries Res.* 2004;38(3):182-91.
- Furlani TA, Magalhães AC, Iano FG, Cardoso VES, Delbem ACB, Buzalaf MAR. Effect of Calcium Pre-rinse and Fluoride Dentifrice on Enamel and on Dental Plaque Formed In Situ. *Oral Health Prev Dent.* 2009;7:23–28.
- Magalhães AC, Furlani TA, Italiani FM, Iano FG, Delbem ACB, Buzalaf MAR. Effect of calcium pre-rinse and fluoride dentifrice on remineralisation of artificially demineralised enamel and on the composition of the dental biofilm formed in situ. *Arch Oral Biol.* 2007;52:1155-1160.
- Marinho VC, Higgins JP, Sheiham A, Logan S. Fluoride toothpastes for preventing dental caries in children and adolescents. *Cochrane Database Syst Rev* 2003a;(1):CD002278.
- Marinho VC, Higgins JP, Logan S, Sheiham A. Fluoride mouthrinses for preventing dental caries in children and adolescents. *Cochrane Database Syst Rev.* 2003b;(3):CD002284.
- Rose RK, Shellis RP, Lee AR. The role of cation bridging in microbial fluoride binding. *Caries Res.* 1996;30(6):458-64.

- Vale GC, Tabchoury CP, Del Bel Cury AA, Tenuta LM, ten Cate JM, Cury JA. APF and dentifrice effect on root dentin demineralization and biofilm. *J Dent Res.* 2011;90(1):77-81.
- Vogel GL, Schumacher GE, Chow LC, Takagi S, Carey CM. Ca Pre-rinse Greatly Increases Plaque and Plaque Fluid F. *J Dent Res.* 2008a;87(5):466-469.
- Vogel GL, Chow LC, Carey CM. Calcium pre-rinse greatly increases overnight salivary fluoride after a 228 ppm fluoride rinse. *Caries Res.* 2008b;42(5):401-404.
- Vogel GL, Tenuta LMA, Schumacher GE, Chow LC. A Calcium Prerinse Required to Form Calcium Fluoride in Plaque from a Sodium Fluoride Rinse. *Caries Res.* 2014;48:174–178.
- Vogel GL, Tenuta LMA, Schumacher GE, Chow LC. No Calcium-Fluoride-Like Deposits Detected in Plaque Shortly after a Sodium Fluoride Mouthrinse. *Caries Res.* 2010;44:108–115.
- Vogel GL. Oral Fluoride Reservoirs and the Prevention of Dental Caries. In: Buzalaf MAR (ed): *Fluoride and the Oral Environment.* Monogr Oral Sci. Basel, Karger; 2011; pp 146–157.

ANEXOS

Anexo 1

Comprovante de submissão

- Computer-Generated E-Mail: Caries Research - Account Created in Manuscript Central
-

● **david.beighton@kcl.ac.uk**

Jan 19 em 10:13 PM 1

Para jgabriel.ssouza@yahoo.com.br

19-Jan-2016

Dear Ms. Souza:

A manuscript titled Ca pre-rinse before F rinse increases biofilm F concentration and reduces enamel demineralization (201601010) has been submitted by Ms. João Gabriel Souza to Caries Research.

You are listed as a co-author for this manuscript. The online peer-review system, Manuscript Central, automatically creates a user account for you. Your USER ID and PASSWORD for your account is as follows:

Site URL: <https://mc.manuscriptcentral.com/cre>

USER ID: jgabriel.ssouza@yahoo.com.br

PASSWORD: For security reasons your password is not contained in this email. To set your password click the link below.

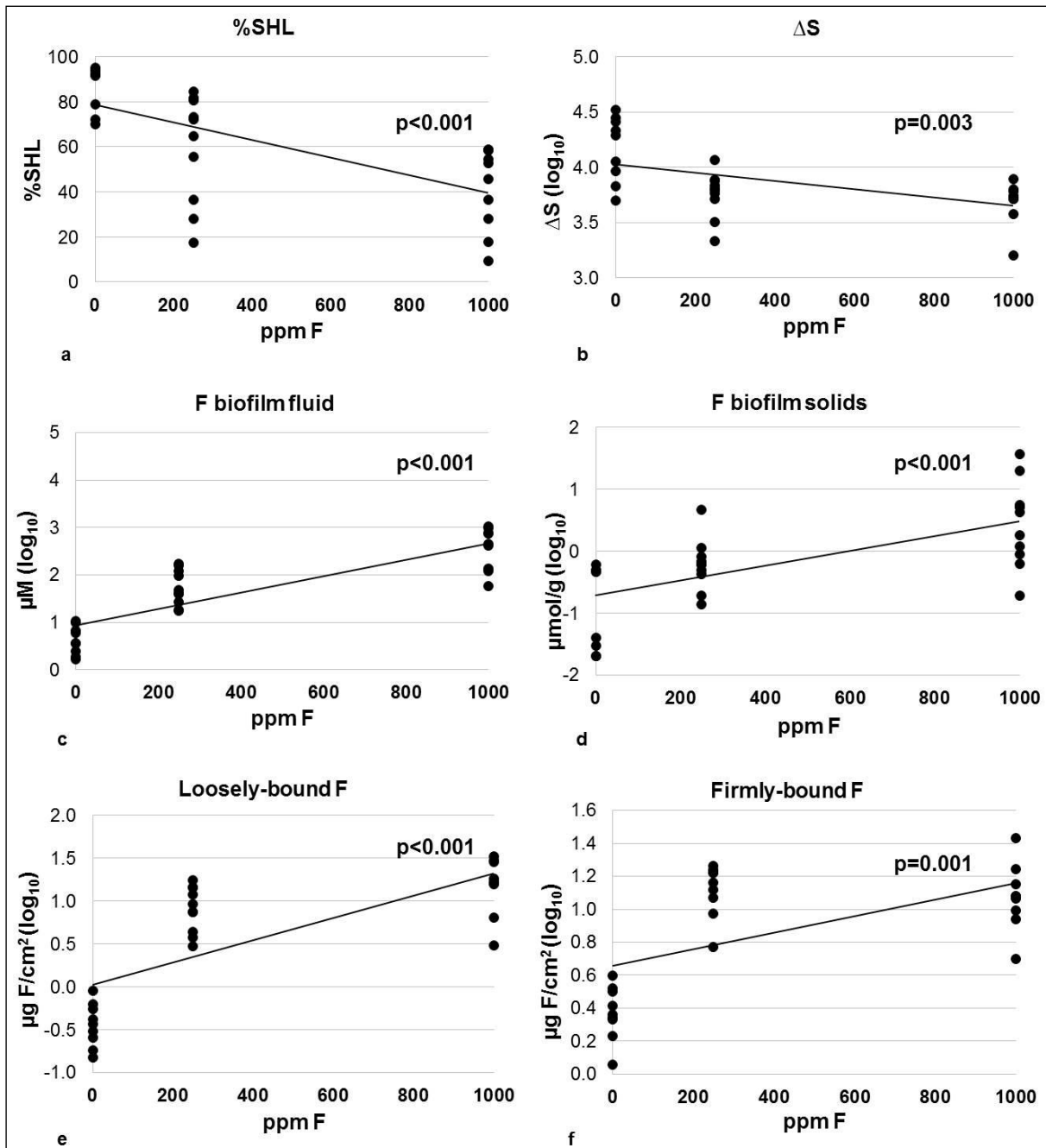
https://mc.manuscriptcentral.com/cre?URL_MASK=a0b297d7ba894005bd3fa1939b902727

You can use the above USER ID and PASSWORD (once set) to log in to the site and check the status of papers you have authored/co-authored. Please log in to <https://mc.manuscriptcentral.com/cre> to update your account information via the edit account tab at the top right.

Thank you for your participation.

Anexo 2 –Dose-resposta modelo

Análise de regressão linear considerando apenas os grupos 0, 250 e 1000 ppm F para verificar o efeito dose-resposta do “modelo in situ de Piracicaba” para diferentes concentrações de bochecho fluoretado.



Anexo 3 – Análise biofilme

Análise da porção fluida e sólida do biofilme, considerando as concentrações de F (descrita no artigo), Ca iônico livre e pH do fluido do biofilme, determinado de acordo com metodologia previamente padronizada (Vogel et al., 2000), utilizando eletrodos de íon específico. Determinou-se também as concentrações de F (descrita no artigo), Ca e fósforo (Pi) na porção sólida do biofilme. As concentrações de Ca e Pi foram realizadas utilizando reagentes colorimétricos (Arsenazo III e verde malaquita, respectivamente) (Vogel et al., 1983; Tenuta et al., 2006).

