



**UNIVERSIDADE ESTADUAL DE CAMPINAS
SISTEMA DE BIBLIOTECAS DA UNICAMP
REPOSITÓRIO DA PRODUÇÃO CIENTÍFICA E INTELLECTUAL DA UNICAMP**

Versão do arquivo anexado / Version of attached file:

Versão do Editor / Published Version

Mais informações no site da editora / Further information on publisher's website:

<http://www.ncbi.nlm.nih.gov/pubmed/7787799>

DOI: 0

Direitos autorais / Publisher's copyright statement:

©1994.0 by Associação Brasileira de Divulgação Científica. All rights reserved.

DIRETORIA DE TRATAMENTO DA INFORMAÇÃO

Cidade Universitária Zeferino Vaz Barão Geraldo

CEP 13083-970 – Campinas SP

Fone: (19) 3521-6493

<http://www.repositorio.unicamp.br>

The kinetics of chondroitin 4-sulfate release from stimulated platelets and its relation to thromboxane A₂ formation and granule secretion

J.L. Donato¹, M.D. Nogueira²,
S. Marcondes¹, E. Antunes¹, H.B. Nader²,
C.P. Dietrich² and G. de Nucci¹

¹*Departamento de Farmacologia, Faculdade de Ciências Médicas,
UNICAMP, 13081-970 Campinas, SP, Brasil*

²*Departamento de Bioquímica, Escola Paulista de Medicina,
04023-900 São Paulo, SP, Brasil*

1. In platelet rich plasma (PRP), chondroitin 4-sulfate release from platelets occurred after stimulation with ADP (5 μ M), collagen (5-10 μ g/ml), or adrenaline (10 μ M). Release started within 60 s and maximum release (0.7-2.0 mg/l) was reached within 180 s. TXA₂ formation and dense granule release reached a maximum within 90 s after stimulation.

2. Using washed platelets (1.5 x 10⁸ cells/ml), the platelet responses were faster. Release of chondroitin 4-sulfate and TXA₂ started within 20-30 s after thrombin addition (100 mU/ml). Maximum release was reached within 60 s in both cases. Dense granule release started in the first 5 s of stimulation (34.6 \pm 12.4%) reaching maximum secretion (74.4 \pm 8.7%) within 60 s.

3. Our results demonstrate that maximal chondroitin 4-sulfate release occurs after the dense granule release reaction in both PRP and washed platelets. This observation suggests that chondroitin 4-sulfate is unlikely to be stored in the dense granules but may be stored in the α -granules.

Key words: chondroitin 4-sulfate, thromboxane A₂, 5-HT release, platelet aggregation.

Presented at "SIMEC 94" - 3rd Brazilian Symposium on Extracellular Matrix, Angra dos Reis, RJ, Brasil, September 11-14, 1994.

Publication supported by FAPESP.

Correspondence: G. de-Nucci, Departamento de Farmacologia, Faculdade de Ciências Médicas, UNICAMP, Caixa Postal 6111, 13081-970 Campinas, SP, Brasil.

Introduction

Platelets contain large amounts of proteoglycans in their membranes and in their storage granules. In the latter case, the proteoglycans are present in the same subcellular fraction as platelet factor-4 (Barber et al., 1972). Chondroitin 4-sulfate, which accounts for more than 90% of the platelet proteoglycan content (Barber et al., 1972; Nader, 1991), is thought to be the carrier molecule for platelet factor-4 (Barber et al., 1972; Luscombe et al., 1981, Huang et al., 1982) and has been implicated in cell-cell interactions (Sampaio and Dietrich, 1981). In addition, a high molecular weight chondroitin sulfate (Mr 240,000) is known to be released from platelets during aggregation induced by several agonists including ADP, adrenaline, noradrenaline, serotonin, collagen and thrombin (Nader, 1991).

We describe the time course of chondroitin 4-sulfate release during platelet activation and aggregation induced by various stimuli. We have also studied the correlation between this release and the biosynthesis of thromboxane A₂ (TXA₂) and the liberation of dense granule contents.

Material and Methods

Platelet preparation and stimulation

Human blood was anticoagulated with 3.8% sodium citrate (1:10) and platelet rich plasma (PRP) was prepared by differential centrifugation. Washed platelets were prepared as described by Radomski and Moncada (1983). Platelet aggregation was monitored using a Born-type aggregometer.

Release reaction

Platelets were labelled in PRP by incubation with 0.5 mM [¹⁴C]5-HT (50 Ci/mmol) for 30 min at 37°C. Prior to stimulation, the platelets were incubated with imipramine (1 mM) for at least 15 min in order to minimize the reuptake of released 5-HT. After addition of the agonist, aggregation was allowed to proceed for 5 min and the reaction was terminated by adding 100 µl of 100 mM EDTA. The aggregated platelet suspensions were then centrifuged at 12,000 g for 2 min. The extent of granule [¹⁴C]5-HT release was determined by quantifying the radioactivity in 100 µl of the resulting supernatant. The results are reported as percent of the total [¹⁴C]5-HT content in 100 µl of platelet suspension (Holmsen and Weiss, 1979).

TXB₂ determination

TXA₂ formation was quantified by radioimmunoassay of its stable degradation product TXB₂ (Salmon, 1978). Following aggregation, the platelet samples were centrifuged as described above and the supernatants removed and stored at -20°C until assayed. The results are reported as ng TXB₂/ml platelet suspension.

Quantification of chondroitin 4-sulfate

The amount of chondroitin 4-sulfate released was quantified in both the soluble fraction and in the platelet pellet obtained after the centrifugation of stimulated platelets as described above. The procedure for the separation and identification of the glycosaminoglycans obtained from the platelets after proteolysis has been described elsewhere (Nader, 1991).

Results and Discussion

In PRP, chondroitin 4-sulfate release from platelets occurred after stimulation with ADP (5 µM), collagen (5-10 µg/ml), or adrenaline (10 µM). With these stimuli, release started within 60 s and was maximal (0.7-2.0 mg/l) within 180 s. Simultaneous with the increase in the supernatant level of chondroitin 4-sulfate, there was a decrease in the level of this proteoglycan in the platelet pellet. The release of chondroitin 4-sulfate occurred after TXB₂ formation and the secretion of the dense granule contents, as shown by the [¹⁴C]5-HT release which reached a maximum within 90 s after stimulation (Figure 1). No chondroitin 4-sulfate, TXB₂ formation or dense granule release was observed when a low dose of ADP (1.5 µM), which was unable to induce the second phase of platelet aggregation, was used.

With washed platelets (1.5 x 10⁸ cells/ml), the release of chondroitin 4-sulfate and TXB₂ was faster and started within 20-30 s after thrombin (100 mU/ml) addition. The maximum amount released in both cases was reached within 60 s. The dense granule release in the presence of thrombin started within 5 s of stimulation (34.6 ± 12.4%) and reached a maximum of 74.4 ± 8.7% within 60 s (Figure 1). In the first 15 s after stimulation there was no chondroitin 4-sulfate release from washed platelets. Hagen (1972) has previously shown that platelets stimulated with thrombin (4 U/ml) and latex released both adenine nucleotides, which are also stored in platelet dense granules, and glycosaminoglycans with the same time course. This discrepancy most likely reflects the fact that Hagen used

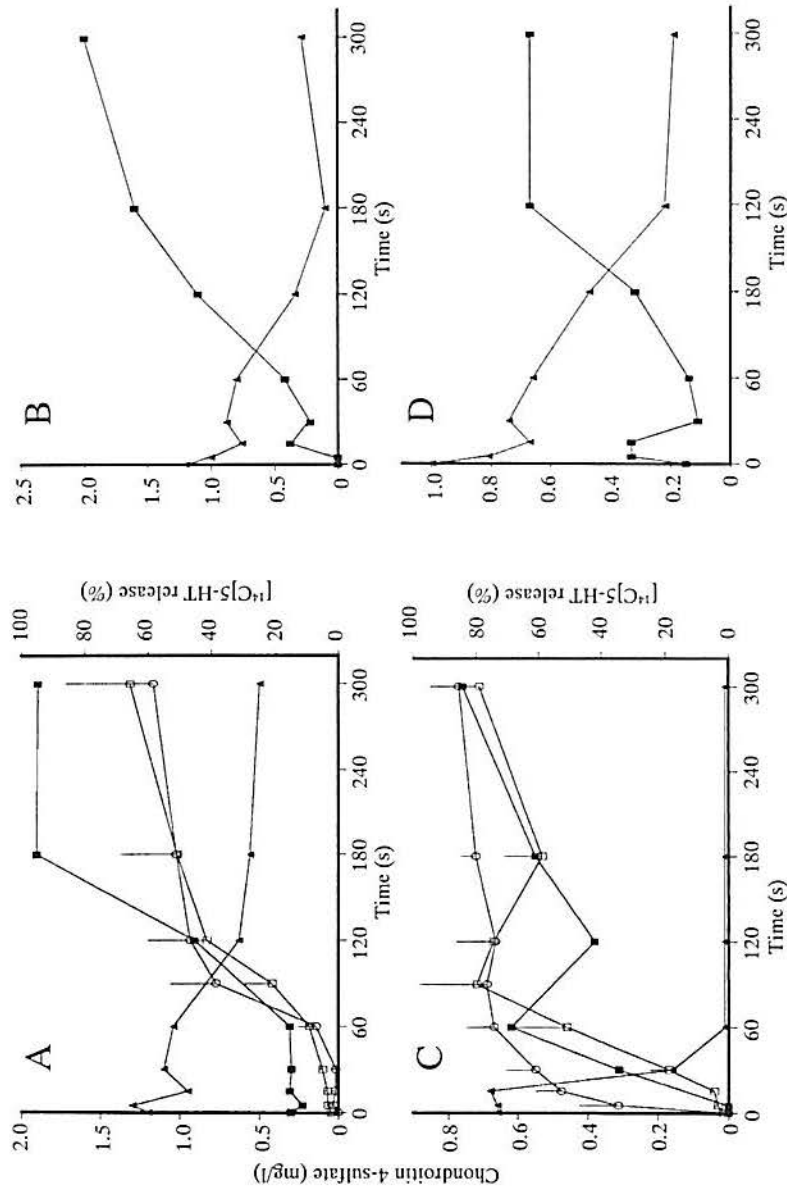


Figure 1 - The time course of chondroitin 4-sulfate release, thromboxane B_2 formation and dense granule release during platelet aggregation. *A*) Platelets aggregated with $5\ \mu\text{M}$ ADP. *B*) Platelets aggregated with $8\ \mu\text{g/ml}$ of collagen. *C*) Washed platelets stimulated with $100\ \text{mU}$ thrombin/ml. *D*) Platelets aggregated with $10\ \mu\text{M}$ adrenaline. (■), Chondroitin 4-sulfate in the supernatant; (Δ), chondroitin 4-sulfate in the platelet pellet; (○), $[^{14}\text{C}]5\text{-HT}$ release into the supernatant; (□), thromboxane B_2 release into the supernatant.

40-fold more thrombin than used here. Under such conditions, the granule release reaction and other biochemical events are very rapid and it becomes impossible to dissociate them in a time-course experiment.

It is known that when platelets are stimulated, the dense granule contents are released first, followed by the α -granule release (Rendu and Dupuy, 1991). Our results demonstrate that maximal chondroitin 4-sulfate release occurs after the dense granule release reaction in both PRP and washed platelets. This observation suggests that chondroitin 4-sulfate is unlikely to be stored in the dense granules but may be stored in the α -granules.

References

- Barber AJ, Kaser-Glanzmann R, Jakábová M & Lüscher EF (1972). Characterization of a chondroitin 4-sulfate proteoglycan carrier for heparin neutralizing activity (platelet factor 4) released from human blood platelets. *Biochimica et Biophysica Acta*, 286: 312-329.
- Hagen I (1972). The release of glycosaminoglycans during exposure of human platelets to thrombin and polystyrene latex particles. *Biochimica et Biophysica Acta*, 273: 141-148.
- Holmsen H & Weiss HJ (1979). Secretory pools in platelets. *Annual Review of Medicine*, 30: 119-130.
- Huang SS, Huang JS & Deuel TF (1982). Proteoglycan carrier of human platelet factor 4. *Journal of Biological Chemistry*, 257: 11546-11550.
- Luscombe M, Marshall SE, Pepper DS & Holbrook JJ (1981). The transfer of platelet factor 4 from its proteoglycan carrier to natural and synthetic polymers. *Biochimica et Biophysica Acta*, 678: 137-142.
- Nader HB (1991). Characterization of a heparan sulfate and a peculiar chondroitin 4-sulfate proteoglycan from platelets. *Journal of Biological Chemistry*, 266: 10518-10523.
- Radomski MW & Moncada S (1983). An improved method for washing human platelets with prostacyclin. *Thrombosis Research*, 30: 383-389.
- Rendu F & Dupuy E (1991). Hereditary disorders of platelet function. In: Harris JR (Editor), *Blood Cell Biochemistry*. Vol. 2. Plenum Press, New York, 77-112.
- Salmon JA (1978). A radioimmunoassay for 6-keto-prostaglandin $\text{F}_{1\alpha}$. *Prostaglandins*, 15: 383-397.
- Sampaio LO & Dietrich CP (1981). Changes of sulfated mucopolysaccharides and mucopolysaccharidases during fetal development. *Journal of Biological Chemistry*, 256: 9205-9210.

Received June 21, 1994
Accepted August 9, 1994