

## SHORT COMMUNICATION

**Thermal Constant of an Experimental Population of *Muscina stabulans* (Fallén 1817) (Diptera: Muscidae) in the Laboratory**Luciene Maura Mascarini/<sup>+</sup>, Ângelo Pires do Prado\*

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A study was conducted to investigate the thermal constant in an experimental population of *Muscina stabulans* (Fallén 1817). A natural population was obtained from a livestock and their  $F_1$  generations were maintained at four constant temperatures (16°C, 20°C, 26°C and 31°C). The thermal constant was calculated by the hyperbole method using the base temperature,  $t_b = 4.4^\circ\text{C}$  and the thermal constant  $K = 35.3 \text{ GD}$ . Ricker's geometric regression, considering  $t_b = 7.8^\circ\text{C}$  and  $K = 28.9 \text{ GD}$  was also calculated.

Key words: *Muscina stabulans* - thermal constant - constant temperature

*Muscina stabulans* (Fallén 1817) (Diptera: Muscidae) a common eusynanthropic fly in farms and orchards, may be associated with human dwellings (Smith 1986). In addition, it can play a role in the control of some Diptera species because of its predatory habits on their larvae (Legner & Dietrich 1989). The growth pattern and development of insects are influenced by temperature and can be measured under laboratory conditions. It is possible to estimate the number of generations per year and the potential ability of population increase of at a specific temperature by measuring developmental period, mortality and oviposition pattern under constant temperature conditions (Howe 1967). The thermal requirements of an insect can be appraised by the thermal constant (K), expressed in degree-days (Haddad & Parra 1984). This method has been utilized for many years in forecasting the occurrence of infestations in agriculture and the thermal constant can be used as indicators of the distribution and abundance of insect populations. The objective of this study was to establish a thermal constant for a population of *M. stabulans*.

Experimental populations of *M. stabulans* were obtained from livestock farm Capuavinha (26°57'S, 47°19'W), town of Monte-Mor, SP, Brazil. These flies were transferred to cages (30 cm x 15 cm x 15 cm), where they were fed yeast sugar and water and maintained at  $27 \pm 30^\circ\text{C}$ . Two thousand eggs were collected and divided into 40 pots (9.5 cm x 7.5 cm) with 50 eggs/pots and 10 pots (replicates) at each temperature: 16°C, 20°C, 26°C and 31°C, taining 200 g of fermented of Labina-Purina mouse ration. To these pots, a further 300 g of mouse ration was added and the larvae allowed to grow until pupation. Pu-

pae were removed and transferred to other cages, where they hatched. The total development time was recorded. The basic development temperature to calculate the thermal constant was determined by the hyperbole method (Bean 1961), using the total development times from the four constant temperature groups. Thermal constant was obtained from the equation  $K = D(T - T_b)$ , where K = constant thermal, D = development time (hours), T = temperature in which the insect grew and  $T_b$  = basic temperature of the insect, obtained from the Healthy linear regression program (1986) and from Ricker's geometric average (1986) regression program (Krebs 1989).

The Table shows the total development time from larval to adult stage. Average larval development times decreased with increasing temperature. This was inversely proportional to increase in temperature, except at 31°C when development time was longer than at 26°C. The data obtained at 26°C are similar to those obtained by Kobayashi and Mizushina (1937) with 14.06 days of total development time, Vinogradova and Marcenko (1984), 12.9 days, and Krzywinski (1993) who analyzed the thermal constant at 28°C, obtaining an average of 12.9 to 14 days. The total development time, observed by Lewallén (1952) at 26.7°C, was longer (20–25) that obtained in our study (12.8 days). Thermal requirements were estimated by determining the thermal constant K and expressed in degree-days. Fig. 1 shows the regression analysis, which showed  $T_b = 4.4^\circ\text{C}$  and  $K = 35.3 \text{ GD}$ , and Fig. 2 shows Ricker's regression with  $T_b = 7.8^\circ\text{C}$  and  $K = 28.9 \text{ GD}$ .

Insect body temperature is probably more important than environmental temperature in influencing behavior (Chapman 1983). Temperature limits can be established by inactivity and by the destruction of some enzymes (Howe 1967). Lethal temperature limits are metabolically regulated by enzymes. Metabolic pattern is measured by oxygen consumption, and when ambient temperature is close to the maximum limit, metabolism is reduced, probably due to denaturation of enzymes (Chapman 1983). The knowledge of the thermal requirements of insects estimated by basic temperature and thermal constant permits,

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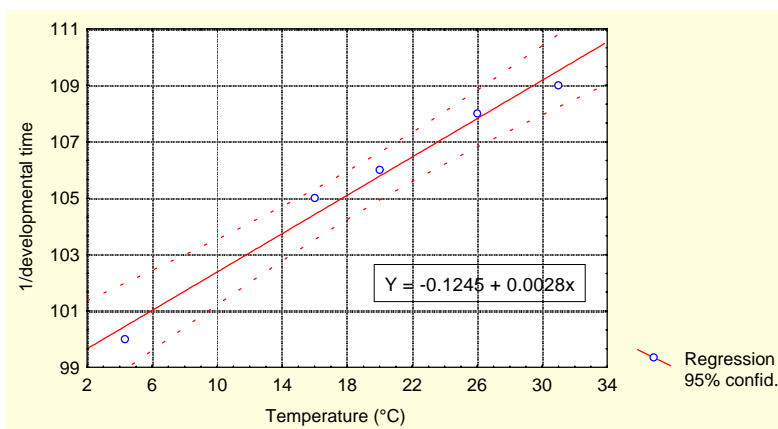


Fig. 1: standard regression analysis of 1/developmental time (temperature) for *Muscina stabulans*.  $T_b = 4.4^\circ\text{C}$ ;  $K = 35.3 \text{ GD}$

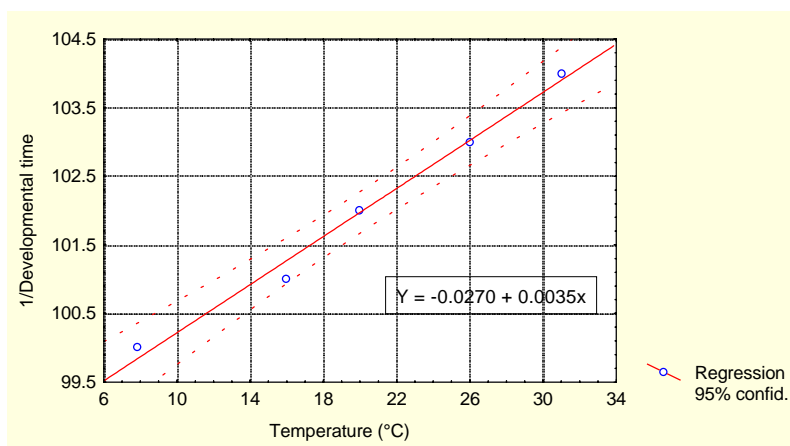


Fig. 2 : Ricker's geometric regression analysis of 1/developmental time (temperature) for *Muscina stabulans*.  $T_b = 7.8^\circ\text{C}$ ;  $K = 28.9\text{GD}$

TABLE

Development total time of *Muscina stabulans* at four constant temperatures (egg-adult)

Temperature	Total time of development (egg-adult) in hours (days) <sup>a</sup>
16°C	980 h (40.8 days) A
20°C	500 h (20.8 days) B
26°C	308 h (12.8 days) C
31°C	380 h (15.8 days) D

<sup>a</sup>: n = 10. The different letters show statistical significant difference ( $p = 0.05$ )

temporarily, store insects under certain artificial conditions, facilitating study program and field work (Aguiar-Valgorde & Milward Azevedo 1992). The threshold and thermal constant can also be used as indicators of distribution and abundance of insect populations (Messenger 1959). From these models, temperature occupies a prominent place, because it is a metodologic component that affects insect populations and is easy to predict.

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