

RESEARCH ARTICLE

Association between circulating levels of sex steroid hormones and esophageal adenocarcinoma in the FINBAR Study

Jessica L. Petrick^{1*}, Roni T. Falk¹, Paula L. Hyland¹, Patrick Caron², Ruth M. Pfeiffer¹, Shannon N. Wood¹, Sanford M. Dawsey¹, Christian C. Abnet¹, Philip R. Taylor¹, Chantal Guillemette², Liam J. Murray³, Lesley A. Anderson^{3☉}, Michael B. Cook^{1☉}

1 Division of Cancer Epidemiology and Genetics, National Cancer Institute, NIH, DHHS, Bethesda, Maryland, United States of America, **2** Pharmacogenomics Laboratory, Centre Hospitalier de l'Université Laval de Québec (CHU de Québec) Research Center and Faculty of Pharmacy, Laval University, Québec City, Québec, Canada, **3** Centre for Public Health, School of Medicine, Dentistry and Biomedical Sciences, Queens University Belfast, Belfast, Northern Ireland, United Kingdom

☉ These authors contributed equally to this work.

* jessica.petrick@nih.gov


 OPEN ACCESS

Citation: Petrick JL, Falk RT, Hyland PL, Caron P, Pfeiffer RM, Wood SN, et al. (2018) Association between circulating levels of sex steroid hormones and esophageal adenocarcinoma in the FINBAR Study. *PLoS ONE* 13(1): e0190325. <https://doi.org/10.1371/journal.pone.0190325>

Editor: John Green, University Hospital Llandough, UNITED KINGDOM

Received: June 26, 2017

Accepted: December 12, 2017

Published: January 17, 2018

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the [Creative Commons CC0](https://creativecommons.org/licenses/by/4.0/) public domain dedication.

Data Availability Statement: The authors confirm that data are stored at Queen's University Belfast and available upon request, adhering to the Office for Research Ethics Committees Northern Ireland agreements for patient confidentiality, from Dr Lesley Anderson (L.anderson@qub.ac.uk).

Funding: The study was supported by NIH Intramural Research Program, National Cancer Institute; Cancer Focus Northern Ireland (formerly the Ulster Cancer Foundation); the Northern Ireland R&D office; and the Health Research Board,

Abstract

Background

Esophageal adenocarcinoma (EA) is characterized by a strong male predominance. Sex steroid hormones have been hypothesized to underlie this sex disparity, but no population-based study to date has examined this potential association.

Methods

Using mass spectrometry and ELISA, we quantitated sex steroid hormones and sex hormone binding globulin, respectively, in plasma from males—172 EA cases and 185 controls—within the Factors Influencing the Barrett/Adenocarcinoma Relationship (FINBAR) Study, a case-control investigation conducted in Northern Ireland and Ireland. Multivariable adjusted logistic regression was used to calculate odds ratios (ORs) and 95% confidence intervals (CIs) for associations between circulating hormones and EA.

Results

Higher androgen:estrogen ratio metrics were associated with increased odds of EA (e.g., testosterone:estradiol ratio $OR_{Q4 \text{ v. } Q1} = 2.58$, 95%CI = 1.23–5.43; $P_{\text{trend}} = 0.009$). All estrogens and androgens were associated with significant decreased odds of EA. When restricted to individuals with minimal to no decrease in body mass index, the size of association for the androgen:estrogen ratio was not greatly altered.

Conclusions

This first study of sex steroid hormones and EA provides tentative evidence that androgen:estrogen balance may be a factor related to EA. Replication of these findings in prospective studies is needed to enhance confidence in the causality of this effect.

Ireland. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Abbreviations: ADT, androsterone; BE, Barrett's esophagus; CV, coefficient of variation; DHEA, dehydroepiandrosterone; DHT, dihydrotestosterone; EA, esophageal adenocarcinoma; FINBAR, Factors Influencing the Barrett/Adenocarcinoma Relationship; GERD, gastroesophageal reflux disease; SHBG, sex hormone binding globulin.

Introduction

Esophageal adenocarcinoma (EA) is more common in men than women worldwide, with a male-to-female ratio of approximately 4.4 [1]. However, the male-to-female ratio peaks between 50 to 54 years and then decreases [2], suggesting an androgenic effect and perhaps estrogenic protection. Additionally, established risk factors that vary in prevalence by sex, such as smoking and obesity, cannot fully explain the male predominance [3, 4]. Thus, sex steroid hormones have been proposed as a possible explanation of the sex disparity [5]. This hypothesis is supported by sex steroid hormone involvement in the inflammatory process, including associations between testosterone and inflammatory markers [6–9], sex steroid hormone receptor protein expression—specifically estrogen receptor β —in esophageal cancer tissue [10, 11], and lower rates of EA among men with prostate cancer, who are likely to receive anti-androgen therapies [12, 13]. Additionally, a small hospital-based study reported higher testosterone levels among EA cases ($n = 25$) than controls ($n = 8$) [14], and two studies of the EA precursor metaplasia, Barrett's esophagus (BE), reported positive associations for free androgens [15, 16]. Thus, we investigated whether circulating sex steroid hormone concentrations were associated with EA in the Factors Influencing the Barrett/Adenocarcinoma Relationship (FINBAR) study—a population-based case-control study of EA and BE conducted in Northern Ireland and the Republic of Ireland between 2002 and 2004 [17–36].

Materials and methods

Study population

The aim of the FINBAR study was to determine the stage along the esophageal inflammation-metaplasia-adenocarcinoma sequence that potential risk factors for BE and EA exert their effects [35]. To accomplish this, the study included individuals aged ≤ 85 years with a histologically confirmed EA ($N = 227$) and ≥ 3 cm of visible BE ($n = 224$). *In situ* cancers were excluded. In Northern Ireland, cases were identified from electronic pathology records from all laboratories in the province. In the Republic of Ireland, cases were identified from primary esophageal cancer treatment centers. The association between sex steroid hormones and BE has previously been reported [16]; therefore, only EA cases and controls are included in the current study.

Eligible controls were adults without a history of gastrointestinal cancer or BE ($N = 260$). Controls were selected at random from the General Practice Master Index in Northern Ireland and from four general practices in the Republic of Ireland and frequency-matched to cases on sex and 5-year age group.

For the current study of circulating hormones, we restricted the study population to males because there were too few females to provide adequate statistical power for a female-only analysis.

Thus, 172 EA cases and 185 controls are included in the current study. The FINBAR study was approved by the Research Ethics Committee of Queen's University Belfast, the Clinical Research Ethics Committee of Cork Teaching Hospitals, the Research Ethics Committee Board of St. James's Hospital, Dublin, and the National Institutes of Health Office of Human Subjects Research.

Data and sample collection

Information on demographics and risk factors was obtained by a structured computerized interview administered by trained interviewers. Anthropometric variables (i.e., height, weight, waist, and hip circumference) were measured at the time of interview and recalled from

5-years prior to study enrollment. Written informed consent was obtained from each participant before interview.

At the time of interview, a 30 ml sample of peripheral venous blood (non-fasting) was drawn, transported on ice, and then centrifuged within 2 hours for the majority of participants. Serum, plasma, and buffy coat were stored at -80°C . Plasma samples from EDTA-treated tubes were used for quantitation of circulating sex steroid hormones and sex hormone binding globulin (SHBG).

Laboratory assays and measurements

All assays were performed in 2015 at the Pharmacogenomics Laboratory of Laval University, Quebec, Canada. The plasma samples were never thawed prior to this study and have been stored since processing at -80°C . Prior studies have shown long-term stability of sex steroid hormones at temperatures less than or equal to -20°C [37–43]. Samples were quantitated for dehydroepiandrosterone (DHEA), androstenediol, androstenedione, progesterone, testosterone, dihydrotestosterone (DHT), androsterone (ADT), estrone, and estradiol using gas chromatography–mass spectrometry [44]. In each of the nine batches, three low and three high hormone concentration quality control replicates were included. All hormone coefficients of variation (CVs) were less than 10% (range: 3.5–7.4%).

SHBG was quantitated using ELISA (Diagnostics Biochem Canada, Inc.). In each of the fourteen batches, one low and one high concentration replicate were included and the CV was 8.8%.

In addition to individual hormones, we calculated parent estrogens (the sum of estrone and estradiol), testosterone:parent estrogens ratio, testosterone:estradiol ratio, androstenedione:estrone ratio, free estradiol [45], free testosterone [46], and free DHT [47]. Hormone levels were categorized in quartiles, based on the distributions among the control participants. Quartiles allowed examination of the associations without assumptions about the underlying dose-response relationship. Tests of linear trend were performed based on the quartile-specific medians of the hormone levels.

Statistical analysis

Unconditional logistic regression was used to calculate odds ratios (ORs) and corresponding 95% confidence intervals (CIs) for the associations between circulating hormones and EA odds. Potential confounders were variables associated with EA and with at least one exposure. These were entered into an age-adjusted multivariable stepwise logistic regression model (entrance p-value = 0.05 and removal p-value = 0.10). Final adjusted models included age (<54.1 , 54.1 – 65.8 , 65.9 – 72.2 , ≥ 72.3), education (<10 , 10 – 12 , ≥ 13 years), smoking status (ever, never), body mass index (BMI, <25.0 , 25.0 – 29.9 , ≥ 30.0 kg/m^2), frequent gastroesophageal reflux disease (GERD) symptoms (yes, no; defined as heartburn or acid reflux occurring weekly at least 5 years prior to the interview), and serologic *Helicobacter pylori* status (positive, negative) [34]. We additionally tested alcohol consumption, physical activity, occupation, waist-to-hip ratio, aspirin use, proton pump inhibitor use, and H2 receptor antagonist use, but these did not meet the inclusion criteria and were not included in the final models. Additionally, adjustment for age and BMI as continuous instead of categorical measures had negligible effects on the analysis. As androgens and estrogens are known to exhibit circadian variation [48], we also adjusted for blood draw time of day (as minutes and am/pm), and our results were not substantially altered. Effect measure modification of the relationship between hormones–modeled both as continuous and categorical–and EA by age, BMI, smoking, *H. pylori*, GERD symptoms, waist-to-hip ratio, proton pump inhibitor use, H2 receptor antagonist use, and geographic location was assessed using likelihood ratio tests [49]. The likelihood ratio tests

were corrected for multiple comparisons using a false discovery rate [50]. After correction, there were no effect-measure modifiers of the associations between hormones and EA.

All tests were two-sided. All analyses were conducted using SAS, version 9.3 (SAS Institute, Cary, NC).

Sensitivity analysis

As the measured hormone levels could be affected by cachexia—or weight loss due to underlying EA—we categorized the cases into three groups: 1) individuals without weight loss between BMI reported 5-years prior to interview and the BMI measured at interview (i.e., BMI change ≥ 0 kg/m²); among individuals with weight loss, we dichotomized BMI change at the mean into 2) BMI change < -3.8 kg/m² and 3) BMI change -3.8 to < 0 kg/m². This analysis was conducted using polytomous logistic regression. Sample size was limited; thus, we dichotomized the hormone levels. To further examine the potential residual confounding by age, we examined continuous hormone variables, which were standardized to half the value of the interquartile range, and adjusted for continuous age.

Results

Descriptive characteristics of controls and EA cases are presented in [S1 Table](#). Significant differences were noted for GERD symptoms (20% controls v. 48% EA cases), *H. pylori* positivity (63% v. 52%), manual occupation (51% v. 63%), ever-smoking (64% v. 84%), and education (11.7 v. 10.6 years). Additionally, significant differences in BMI were noted for both self-reported BMI 5-years prior to enrollment or diagnosis (27.2 v. 28.6 kg/m²) and measured at study enrollment (27.9 v. 26.3 kg/m²).

As shown in [Table 1](#), most of the individual hormones had higher measured concentrations in the controls than in the EA cases. For example, testosterone (12.07 v. 10.83 nmol/L, respectively)

Table 1. Mean circulating metabolite concentrations among esophageal adenocarcinoma cases and controls, in the FINBAR Study: 2002–2004.

Hormone	Controls			Esophageal Adenocarcinoma Cases			P-value ^a
	N	Median	Interquartile Range	N	Median	Interquartile Range	
DHEA, nmol/L	185	5.13	3.36–8.18	169	3.68	1.87–5.55	<0.0001
Androstenediol, pmol/L	183	1764.39	1315.76–2494.35	164	1286.03	848.78–1913.04	<0.0001
Androstenedione, nmol/L	185	2.72	2.03–3.39	171	2.34	1.68–3.28	0.06
Testosterone, nmol/L	184	12.07	9.67–15.20	170	10.83	7.52–14.77	0.01
DHT, pmol/L	184	1008.55	751.13–1372.98	171	746.69	548.68–1193.90	<0.0001
ADT, pmol/L	174	592.55	409.10–754.82	143	466.67	315.11–643.47	<0.0001
Estrone, pmol/L	185	104.23	81.11–129.60	165	89.99	65.21–126.75	0.0009
Estradiol, pmol/L	185	67.33	55.29–82.60	170	50.54	37.48–66.49	<0.0001
Progesterone, nmol/L	90	0.23	0.19–0.36	91	0.26	0.20–0.34	0.3
SHBG, nmol/L	184	53.70	39.50–69.35	172	71.10	48.65–100.90	<0.0001
Parent estrogens, pmol/L	185	175.22	142.98–210.75	165	139.37	106.03–196.09	<0.0001
Testosterone: Parent estrogens ratio	184	68.78	54.55–87.32	165	78.88	52.38–102.22	0.05
Androstenedione: Estrone ratio	185	24.49	20.59–31.03	165	26.16	18.98–37.99	0.2
Testosterone: Estradiol ratio	184	177.92	140.11–221.56	170	209.59	157.47–279.48	0.0002
Free testosterone, nmol/L	183	0.18	0.14–0.22	170	0.13	0.09–0.17	<0.0001
Free DHT, pmol/L	183	16.35	11.99–20.41	170	10.21	6.50–13.93	<0.0001
Free estradiol, pmol/L	184	1.51	1.27–1.85	170	0.99	0.73–1.42	<0.0001

^aWilcoxon-Mann-Whitney test comparing the medians of cases and controls.

<https://doi.org/10.1371/journal.pone.0190325.t001>

and estradiol (67.33 v. 50.54 pmol/L) concentrations were both higher in controls than cases. However, the androgen:estrogen ratio metrics were lower in controls compared to cases (e.g., testosterone:estradiol ratio 177.92 v. 209.59).

Individual sex steroid hormones, and calculated free estradiol, free testosterone, and free DHT, were associated with significantly decreased EA odds (Table 2). However, SHBG and the androgen:estrogen ratio metrics—particularly testosterone:estradiol—were associated with an increased EA odds. Comparing quartile 4 versus 1, SHBG was associated with 2.3-times the odds of EA (OR = 2.30, 95% CI = 1.10–4.80; $P_{\text{trend}} = 0.01$), and the testosterone:estradiol ratio was associated with 2.6-times the odds (OR = 2.58, 95% CI = 1.23–5.43; $P_{\text{trend}} = 0.009$). Adjustment for covariates had little effect on the estimates, as observed in the unadjusted results (S2 Table).

When we examined the cases by BMI change, the androgen:estrogen ratio metrics were still associated with an increased odds of EA among individuals with little to no BMI change during the 5 years preceding interview. Comparing individuals with \geq median versus $<$ median, testosterone:estradiol ratio was associated with 1.4 to 2.6-times the odds of EA among individuals with little to no BMI change (OR = 1.35, 95% CI = 0.54–3.39; $P_{\text{trend}} = 0.3$ among EA cases with a BMI change ≥ 0 kg/m² and OR = 2.61, 95% CI = 1.27–5.38; $P_{\text{trend}} = 0.0005$ among individuals with a BMI change of -3.8 to <0 kg/m²) (S3 Table). Additional adjustment of the continuous hormones and SHBG for continuous age did not substantially alter the results (S4 Table).

Discussion

This is the first population-based study to examine associations between circulating sex steroid hormones and EA and provides tentative evidence that androgen:estrogen balance may be a factor related to EA. In our analysis, a high ratio of androgens to estrogens—particularly testosterone:estradiol ratio—was more common in EA patients than controls, including after restriction to cases without weight loss in the previous 5 years. The highest quartile ratio of testosterone:estradiol was associated with a 2.4-times increased odds of EA, while androstenedione:estrone and testosterone:parent estrogens ratios were associated with a non-significant 1.6-times odds. However, many of the individual hormone metrics assessed were inversely associated with EA. Deciphering the extent to which these associations are due to reverse causation will require prospective studies with prediagnostic hormone quantitation.

In older men, obesity is strongly linked to SHBG and influences levels of circulating sex hormones. Specifically, higher BMI is associated with lower levels of DHEA, testosterone, and SHBG and higher levels of estrogens [51]. In our main analysis, only 23% of the cases did not experience weight loss during the 5-years prior to cancer diagnosis. Cancer patients often present with cachexia, which is associated with hypogonadism. Thus, we were concerned that our findings could be a consequence of the cancer itself. Indeed, studies suggest testosterone levels are lower and SHBG higher in male cancer cases compared to age-matched healthy men [52–54]. As a sensitivity analysis, we examined cases by BMI changes in the 5-years prior to study entry. The results showed that SHBG was associated with increased odds of EA among individuals with the most extreme BMI loss (i.e., <-3.8 kg/m²), but not among individuals with little to no BMI loss. Among individuals with little to no BMI change, the ratios of androgens:estrogens were associated with increased odds of EA, but not among individuals with extreme BMI loss. However, cautious interpretation is warranted, as these were *post hoc* analyses that used a proxy (i.e., BMI) of cancer's metabolic effects.

While it has long been hypothesized that sex steroid hormones may underlie the sex disparities in EA pathogenesis [5], no population-based study to date has examined circulating sex

Table 2. Adjusted^a odds ratios (ORs) and 95% confidence intervals (CIs) for the associations between circulating metabolite concentrations and esophageal adenocarcinoma incidence, in the FINBAR Study: 2002–2004.

Hormone	Control (n)	Esophageal Adeno. (n)	OR	95% CI
DHEA, nmol/L				
<3.36	45	70	Referent	
3.36 to <5.13	38	40	0.91	(0.46, 1.80)
5.13 to <8.18	46	29	0.43	(0.21, 0.90)
≥8.18	43	16	0.32	(0.14, 0.76)
<i>P trend^b</i>				0.003
Androstenediol, pmol/L				
<1315.76	41	79	Referent	
1315.76 to <1764.39	42	34	0.41	(0.21, 0.80)
1764.39 to <2494.35	44	27	0.33	(0.16, 0.68)
≥2494.35	43	12	0.13	(0.05, 0.30)
<i>P trend^b</i>				<0.0001
Androstenedione, nmol/L				
<2.03	45	57	Referent	
2.03 to <2.72	42	39	0.96	(0.48, 1.92)
2.72 to <3.39	41	26	0.57	(0.27, 1.19)
≥3.39	44	35	0.65	(0.32, 1.31)
<i>P trend^b</i>				0.1
Testosterone, nmol/L				
<9.67	42	66	Referent	
9.67 to <12.07	45	28	0.54	(0.27, 1.07)
12.07 to <15.20	42	30	0.45	(0.22, 0.92)
≥15.20	42	32	0.50	(0.25, 1.01)
<i>P trend^b</i>				0.05
DHT, pmol/L				
<751.13	42	80	Referent	
751.13 to <1008.55	43	29	0.39	(0.19, 0.78)
1008.55 to <1372.98	42	21	0.23	(0.11, 0.50)
≥1372.98	44	27	0.23	(0.11, 0.49)
<i>P trend^b</i>				<0.0001
ADT, pmol/L				
<409.10	38	51	Referent	
409.10 to <592.55	43	44	0.73	(0.37, 1.44)
592.55 to <754.82	42	21	0.40	(0.19, 0.87)
≥754.82	41	15	0.28	(0.12, 0.64)
<i>P trend^b</i>				0.0008
Estrone, pmol/L				
<81.11	41	72	Referent	
81.11 to <104.23	45	31	0.34	(0.17, 0.68)
104.23 to <129.60	42	19	0.24	(0.11, 0.52)
≥129.60	44	29	0.36	(0.18, 0.72)
<i>P trend^b</i>				0.003
Estradiol, pmol/L				
<55.29	46	97	Referent	
55.29 to <67.33	42	21	0.25	(0.12, 0.52)
67.33 to <82.60	41	16	0.20	(0.09, 0.44)

(Continued)

Table 2. (Continued)

Hormone	Control (n)	Esophageal Adeno. (n)	OR	95% CI
≥82.60	43	22	0.25	(0.12, 0.51)
<i>P trend^b</i>				<0.0001
Progesterone, nmol/L^c				
<0.15	86	74	Referent	
0.15 to <0.20	29	21	0.74	(0.35, 1.54)
0.20 to <0.29	29	33	1.58	(0.80, 3.14)
≥0.29	28	30	1.00	(0.49, 2.01)
<i>P trend^d</i>				0.9
SHBG, nmol/L				
<39.50	43	27	Referent	
39.50 to <53.70	42	28	0.97	(0.44, 2.13)
53.70 to <69.35	44	23	0.64	(0.28, 1.44)
≥69.35	42	80	2.30	(1.10, 4.80)
<i>P trend^b</i>				0.01
Parent estrogens, pmol/L				
<142.98	43	85	Referent	
142.98 to <175.22	43	23	0.29	(0.14, 0.61)
175.22 to <210.75	42	19	0.22	(0.10, 0.47)
≥210.75	44	24	0.26	(0.13, 0.54)
<i>P trend^b</i>				<0.0001
Testosterone: Parent estrogens ratio				
<54.55	45	35	Referent	
54.55 to <68.78	39	23	0.91	(0.42, 2.00)
68.78 to <87.32	44	31	0.93	(0.43, 1.99)
≥87.32	43	62	2.06	(1.00, 4.24)
<i>P trend^b</i>				0.03
Androstenedione: Estrone ratio				
<20.59	41	41	Referent	
20.59 to <24.49	43	18	0.54	(0.24, 1.20)
24.49 to <31.03	44	31	0.92	(0.44, 1.92)
≥31.03	44	61	1.56	(0.76, 3.22)
<i>P trend^b</i>				0.1
Testosterone: Estradiol ratio				
<140.11	44	28	Referent	
140.11 to <177.92	40	28	1.31	(0.59, 2.91)
177.92 to <221.56	44	26	1.10	(0.49, 2.48)
≥221.56	43	74	2.58	(1.23, 5.43)
<i>P trend^b</i>				0.009
Free testosterone, nmol/L				
<0.14	41	91	Referent	
0.14 to <0.18	45	28	0.31	(0.16, 0.61)
0.18 to <0.22	42	24	0.26	(0.12, 0.53)
≥0.22	42	13	0.16	(0.07, 0.37)
<i>P trend^b</i>				<0.0001
Free DHT, pmol/L				
<11.99	42	101	Referent	
11.99 to <16.35	41	32	0.24	(0.12, 0.49)

(Continued)

Table 2. (Continued)

Hormone	Control (n)	Esophageal Adeno. (n)	OR	95% CI
16.35 to <20.41	44	12	0.09	(0.04, 0.21)
≥20.41	43	11	0.07	(0.03, 0.18)
<i>P trend^b</i>				<0.0001
Free estradiol, pmol/L				
<1.27	44	106	Referent	
1.27 to <1.51	41	21	0.24	(0.12, 0.50)
1.51 to <1.85	44	16	0.20	(0.09, 0.42)
≥1.85	42	13	0.15	(0.07, 0.34)
<i>P trend^b</i>				<0.0001

^aLogistic regression models were adjusted for age at interview (quartiles), education (<10, 10–12, 13–20 years), smoking (ever/never), BMI at interview (<25, 25–<30, ≥30 kg/m²), gastroesophageal reflux disease symptoms (yes/no), and *H. pylori* seropositivity (yes/no).

^bTests of linear trend were calculated by assigning the median of each quartile as scores.

^cProgesterone values below the LOD form the referent with the subsequent three categories based on tertiles of the observed population distribution.

^dTest of linear trend for progesterone was calculated by assigning the categorical groups as scores.

<https://doi.org/10.1371/journal.pone.0190325.t002>

steroid hormones and EA. One small hospital-based study (n = 25 cases) that examined the association between fasting serum testosterone reported that EA cases had significantly higher testosterone concentrations compared with eight control subjects. However, post-esophagectomy testosterone levels in cases decreased to levels similar to controls, leading the authors to conclude that the cancer was causing higher circulating testosterone concentrations [14]. While stage information and treatment status was not uniformly collected in the current study, blood samples were collected prior to treatment for the majority of participants. Therefore, changes in testosterone levels post-treatment are unlikely to completely explain the inverse association between testosterone and EA in our main analysis.

Additionally, BE has been examined in relation to circulating sex steroid hormones in two studies—one from a U.S. military medical center and one from the FINBAR Study. Both studies determined that free androgens were associated with increased odds of BE, although this was limited to participants with a high waist-to-hip ratio in the FINBAR Study [15, 16]. Thus, free androgens may be important in the development of BE—possibly by delaying wound healing and allowing more time for metaplasia to develop—while the balance of androgens and estrogens is tentatively inferred from this study to be a factor for progression to invasive cancer.

In vitro studies of esophageal squamous cell carcinoma [55] and adenocarcinoma [56] have shown that administration of estradiol decreases cell growth. One of these studies further examined the effects of estradiol and DHT administration on EA tumors in mice and reported that estradiol injections resulted in a high estradiol:DHT ratio and significant inhibition of tumor growth [55]. This indirectly further supports that the androgen:estrogen balance may be important for EA development, whereby higher levels of androgens relative to estrogens may favor cellular proliferation and tumor growth.

A potential limitation of the current study is that the plasma for cases was collected at time of interview. While the esophagus is not an endocrine gland, there is the possibility that circulating hormone concentrations could be affected by tumor growth and reverse causation could be contributing to these findings [14]. Additionally, age and chronic disease status may alter hormone and SHBG levels. We adjusted the models for age and GERD, which has been suggested to be associated with altered hormone levels [57], but there still may be residual confounding. There is also a potential in this study for recall bias, or differential recall between

cases and controls, particularly for weight 5 years prior to study interview. A strength of the current study is that the FINBAR Study was designed specifically to examine risk factors for EA—such as GERD and *H. pylori*, and we were able to adjust for these. The current study also used robust quantitative technology for sex steroid hormone measurement.

In summary, this study provides tentative evidence that the balance of androgens to estrogens may be important in the development of EA. However, the large number of inverse associations detected make the possibility of reverse causation more likely and underscore the need for a cautious interpretation. Future studies need to be conducted using prospectively collected blood to provide additional confidence that these associations between sex steroid hormones and EA represent causal effects and not effects of cachexia.

Supporting information

S1 Table. Distribution of characteristics among control participants and esophageal adenocarcinoma cases, in the FINBAR Study 2002–2004. ^aP values based on t-test for continuous variables and χ^2 test for categorical variables. ^bDefined as at least 50 times per year or about once a week. ^cDefined as at least once weekly for 6 months or more. (DOCX)

S2 Table. Unadjusted odds ratios (ORs) and 95% confidence intervals (CIs) for the associations between circulating metabolite concentrations and esophageal adenocarcinoma incidence, in the FINBAR Study: 2002–2004. ^aTests of linear trend were calculated by assigning the median of each quartile as scores. ^bProgesterone values below the LOD form the referent with the subsequent three categories based on tertiles of the observed population distribution. ^cTest of linear trend for progesterone was calculated by assigning the categorical groups as scores. (DOCX)

S3 Table. Adjusted^a odds ratios (ORs) and 95% confidence intervals (CI) for associations between circulating sex steroid hormone concentrations and esophageal adenocarcinoma risk, stratified by body mass index (kg/m²) change in the 5 years preceding interview. ^aLogistic regression models were adjusted for age at interview (quartiles), education (<10, 10–12, 13–20 years), smoking (ever/never), BMI at interview (<25, 25–<30, \geq 30 kg/m²), gastroesophageal reflux disease symptoms (yes/no), and *H. pylori* seropositivity (yes/no). ^bTests of linear trend were calculated by assigning the median of each quartile as scores. ^cProgesterone values below the LOD form the referent with the subsequent three categories based on tertiles of the observed population distribution. ^dTest of linear trend for progesterone was calculated by assigning the categorical groups as scores. (DOCX)

S4 Table. Adjusted^a odds ratios (ORs) and 95% confidence intervals (CIs) for the associations between circulating metabolite concentrations^b and esophageal adenocarcinoma incidence, Factors Influencing the Barrett/Adenocarcinoma Relationship: 2002–2004. ^aLogistic regression models were adjusted for age at interview (continuous), education (<10, 10–12, 13–20 years), smoking (ever/never), BMI at interview (<25, 25–<30, \geq 30 kg/m²), gastroesophageal reflux disease symptoms (yes/no), and *H. pylori* seropositivity (yes/no). ^bStandardized to half the value of the interquartile range (e.g., for DHEA the OR is 0.70 for an increase of 2.41 nmol/L, which is 0.5* [8.18–3.36]), which approximates a single quartile increase in exposure. (DOCX)

Author Contributions

Conceptualization: Jessica L. Petrick, Roni T. Falk, Paula L. Hyland, Patrick Caron, Ruth M. Pfeiffer, Shannon N. Wood, Sanford M. Dawsey, Christian C. Abnet, Chantal Guillemette, Liam J. Murray, Lesley A. Anderson, Michael B. Cook.

Data curation: Jessica L. Petrick, Patrick Caron, Shannon N. Wood, Chantal Guillemette, Lesley A. Anderson, Michael B. Cook.

Formal analysis: Jessica L. Petrick, Roni T. Falk, Patrick Caron, Shannon N. Wood, Chantal Guillemette, Michael B. Cook.

Funding acquisition: Michael B. Cook.

Methodology: Jessica L. Petrick, Roni T. Falk, Paula L. Hyland, Patrick Caron, Ruth M. Pfeiffer, Shannon N. Wood, Sanford M. Dawsey, Christian C. Abnet, Philip R. Taylor, Chantal Guillemette, Liam J. Murray, Lesley A. Anderson, Michael B. Cook.

Resources: Liam J. Murray, Lesley A. Anderson, Michael B. Cook.

Supervision: Michael B. Cook.

Writing – original draft: Jessica L. Petrick.

Writing – review & editing: Jessica L. Petrick, Roni T. Falk, Paula L. Hyland, Patrick Caron, Ruth M. Pfeiffer, Shannon N. Wood, Sanford M. Dawsey, Christian C. Abnet, Philip R. Taylor, Chantal Guillemette, Liam J. Murray, Lesley A. Anderson, Michael B. Cook.

References

1. Arnold M, Soerjomataram I, Ferlay J, Forman D. Global incidence of oesophageal cancer by histological subtype in 2012. *Gut*. 2015; 64(3):381–7. <https://doi.org/10.1136/gutjnl-2014-308124> PMID: 25320104
2. Mathieu LN, Kanarek NF, Tsai HL, Rudin CM, Brock MV. Age and sex differences in the incidence of esophageal adenocarcinoma: results from the Surveillance, Epidemiology, and End Results (SEER) Registry (1973–2008). *Dis Esophagus*. 2014; 27(8):757–63. <https://doi.org/10.1111/dote.12147> PMID: 24118313
3. Freedman ND, Derakhshan MH, Abnet CC, Schatzkin A, Hollenbeck AR, McColl KE. Male predominance of upper gastrointestinal adenocarcinoma cannot be explained by differences in tobacco smoking in men versus women. *European journal of cancer*. 2010; 46(13):2473–8. Epub 2010/07/08. <https://doi.org/10.1016/j.ejca.2010.05.005> PMID: 20605442
4. Rutegard M, Nordenstedt H, Lu Y, Lagergren J, Lagergren P. Sex-specific exposure prevalence of established risk factors for oesophageal adenocarcinoma. *British journal of cancer*. 2010; 103(5):735–40. Epub 2010/08/12. <https://doi.org/10.1038/sj.bjc.6605804> PMID: 20700121
5. Lagergren J, Nyren O. Do sex hormones play a role in the etiology of esophageal adenocarcinoma? A new hypothesis tested in a population-based cohort of prostate cancer patients. *Cancer Epidemiol Biomarkers Prev*. 1998; 7(10):913–5. Epub 1998/10/31. PMID: 9796637
6. Schmidt M, Naumann H, Weidler C, Schellenberg M, Anders S, Straub RH. Inflammation and sex hormone metabolism. *Ann N Y Acad Sci*. 2006; 1069:236–46. Epub 2006/07/21. <https://doi.org/10.1196/annals.1351.021> PMID: 16855150
7. Maggio M, Basaria S, Ceda GP, Ble A, Ling SM, Bandinelli S, et al. The relationship between testosterone and molecular markers of inflammation in older men. *J Endocrinol Invest*. 2005; 28(11 Suppl Proceedings):116–9. Epub 2006/06/09.
8. Liao CH, Li HY, Yu HJ, Chiang HS, Lin MS, Hua CH, et al. Low serum sex hormone-binding globulin: marker of inflammation? *Clin Chim Acta*. 2012; 413(7–8):803–7. Epub 2012/02/02. <https://doi.org/10.1016/j.cca.2012.01.021> PMID: 22293276
9. Kupelian V, Chiu GR, Araujo AB, Williams RE, Clark RV, McKinlay JB. Association of sex hormones and C-reactive protein levels in men. *Clin Endocrinol (Oxf)*. 2010; 72(4):527–33. Epub 2009/09/23.
10. Rashid F, Khan RN, Iftikhar SY. Probing the link between oestrogen receptors and oesophageal cancer. *World J Surg Oncol*. 2010; 8:9. Epub 2010/02/12. <https://doi.org/10.1186/1477-7819-8-9> PMID: 20146809

11. Yang H, Sukocheva OA, Hussey DJ, Watson DI. Estrogen, male dominance and esophageal adenocarcinoma: is there a link? *World J Gastroenterol*. 2012; 18(5):393–400. Epub 2012/02/22. <https://doi.org/10.3748/wjg.v18.i5.393> PMID: 22346245
12. Cooper SC, Croft S, Day R, Thomson CS, Trudgill NJ. Patients with prostate cancer are less likely to develop oesophageal adenocarcinoma: could androgens have a role in the aetiology of oesophageal adenocarcinoma? *Cancer Causes Control*. 2009; 20(8):1363–8. <https://doi.org/10.1007/s10552-009-9359-2> PMID: 19455396
13. Cooper SC, Trudgill NJ. Subjects with prostate cancer are less likely to develop esophageal cancer: analysis of SEER 9 registries database. *Cancer Causes Control*. 2012; 23(6):819–25. PMID: 24251326
14. Awan AK, Iftikhar SY, Morris TM, Clarke PA, Grabowska AM, Waraich N, et al. Androgen receptors may act in a paracrine manner to regulate oesophageal adenocarcinoma growth. *Eur J Surg Oncol*. 2007; 33(5):561–8. <https://doi.org/10.1016/j.ejso.2006.12.001> PMID: 17254742
15. Cook MB, Wood SN, Cash BD, Young P, Acosta RD, Falk RT, et al. Association Between Circulating Levels of Sex Steroid Hormones and Barrett's Esophagus in Men: A Case-Control Analysis. *Clin Gastroenterol Hepatol*. 2014; 13(4):673–82. Epub 2014/08/28. <https://doi.org/10.1016/j.cgh.2014.08.027> PMID: 25158929
16. Cook MB, Wood SN, Hyland PL, Caron P, Drahos J, Falk RTP, R., et al. Sex steroid hormones in relation to Barrett's esophagus: An analysis of the FINBAR Study. *Andrology*. 2017; In Press.
17. Anderson LA, Johnston BT, Watson RG, Murphy SJ, Ferguson HR, Comber H, et al. Nonsteroidal anti-inflammatory drugs and the esophageal inflammation-metaplasia-adenocarcinoma sequence. *Cancer Res*. 2006; 66(9):4975–82. <https://doi.org/10.1158/0008-5472.CAN-05-4253> PMID: 16651456
18. Murphy SJ, Hughes AE, Patterson CC, Anderson LA, Watson RG, Johnston BT, et al. A population-based association study of SNPs of GSTP1, MnSOD, GPX2 and Barrett's esophagus and esophageal adenocarcinoma. *Carcinogenesis*. 2007; 28(6):1323–8. <https://doi.org/10.1093/carcin/bgm007> PMID: 17277236
19. Ferguson HR, Wild CP, Anderson LA, Murphy SJ, Johnston BT, Murray LJ, et al. No association between hOGG1, XRCC1, and XPD polymorphisms and risk of reflux esophagitis, Barrett's esophagus, or esophageal adenocarcinoma: results from the factors influencing the Barrett's adenocarcinoma relationship case-control study. *Cancer Epidemiol Biomarkers Prev*. 2008; 17(3):736–9. <https://doi.org/10.1158/1055-9965.EPI-07-2832> PMID: 18349297
20. Murphy SJ, Anderson LA, Ferguson HR, Johnston BT, Watson PR, McGuigan J, et al. Dietary antioxidant and mineral intake in humans is associated with reduced risk of esophageal adenocarcinoma but not reflux esophagitis or Barrett's esophagus. *J Nutr*. 2010; 140(10):1757–63. <https://doi.org/10.3945/jn.110.124362> PMID: 20702746
21. O'Rourke MA, Cantwell MM, Abnet CC, Brockman AJ, Murray LJ, Group FS. Toenail trace element status and risk of Barrett's oesophagus and oesophageal adenocarcinoma: results from the FINBAR study. *Int J Cancer*. 2012; 131(8):1882–91. <https://doi.org/10.1002/ijc.27434> PMID: 22262413
22. Cook MB, Wood S, Hyland PL, Caron P, Drahos J, Falk RT, et al. Sex steroid hormones in relation to Barrett's esophagus: an analysis of the FINBAR Study. *Andrology*. 2017; 5(2):240–7. <https://doi.org/10.1111/andr.12314> PMID: 28241109
23. Shivappa N, Hebert JR, Anderson LA, Shrubsole MJ, Murray LJ, Getty LB, et al. Dietary inflammatory index and risk of reflux oesophagitis, Barrett's oesophagus and oesophageal adenocarcinoma: a population-based case-control study. *Br J Nutr*. 2017; 117(9):1323–31. <https://doi.org/10.1017/S0007114517001131> PMID: 28571591
24. Dai Q, Cantwell MM, Murray LJ, Zheng W, Anderson LA, Coleman HG, et al. Dietary magnesium, calcium:magnesium ratio and risk of reflux oesophagitis, Barrett's oesophagus and oesophageal adenocarcinoma: a population-based case-control study. *Br J Nutr*. 2016; 115(2):342–50. <https://doi.org/10.1017/S0007114515004444> PMID: 26563986
25. Sharp L, Carsin AE, Cantwell MM, Anderson LA, Murray LJ, Group FS. Intakes of dietary folate and other B vitamins are associated with risks of esophageal adenocarcinoma, Barrett's esophagus, and reflux esophagitis. *J Nutr*. 2013; 143(12):1966–73. <https://doi.org/10.3945/jn.113.174664> PMID: 24132576
26. Denver P, Donnelly M, Murray LJ, Anderson LA. Psychosocial factors and their association with reflux oesophagitis, Barrett's oesophagus and oesophageal adenocarcinoma. *World J Gastroenterol*. 2013; 19(11):1770–7. <https://doi.org/10.3748/wjg.v19.i11.1770> PMID: 23555165
27. Mulholland HG, Murray LJ, Anderson LA, Cantwell MM, group Fs. Vitamin D, calcium and dairy intake, and risk of oesophageal adenocarcinoma and its precursor conditions. *Br J Nutr*. 2011; 106(5):732–41. <https://doi.org/10.1017/S0007114511000742> PMID: 21736847

28. O'Doherty MG, Cantwell MM, Murray LJ, Anderson LA, Abnet CC, Group FS. Dietary fat and meat intakes and risk of reflux esophagitis, Barrett's esophagus and esophageal adenocarcinoma. *Int J Cancer*. 2011; 129(6):1493–502. <https://doi.org/10.1002/ijc.26108> PMID: 21455992
29. O'Doherty MG, Abnet CC, Murray LJ, Woodside JV, Anderson LA, Brockman JD, et al. Iron intake and markers of iron status and risk of Barrett's esophagus and esophageal adenocarcinoma. *Cancer Causes Control*. 2010; 21(12):2269–79. <https://doi.org/10.1007/s10552-010-9652-0> PMID: 20936528
30. Ladanchuk TC, Johnston BT, Murray LJ, Anderson LA, group Fs. Risk of Barrett's oesophagus, oesophageal adenocarcinoma and reflux oesophagitis and the use of nitrates and asthma medications. *Scand J Gastroenterol*. 2010; 45(12):1397–403. <https://doi.org/10.3109/00365521.2010.503968> PMID: 20626305
31. Anderson LA, Cantwell MM, Watson RG, Johnston BT, Murphy SJ, Ferguson HR, et al. The association between alcohol and reflux esophagitis, Barrett's esophagus, and esophageal adenocarcinoma. *Gastroenterology*. 2009; 136(3):799–805. <https://doi.org/10.1053/j.gastro.2008.12.005> PMID: 19162028
32. Mulholland HG, Cantwell MM, Anderson LA, Johnston BT, Watson RG, Murphy SJ, et al. Glycemic index, carbohydrate and fiber intakes and risk of reflux esophagitis, Barrett's esophagus, and esophageal adenocarcinoma. *Cancer Causes Control*. 2009; 20(3):279–88. <https://doi.org/10.1007/s10552-008-9242-6> PMID: 18839322
33. Ferguson HR, Wild CP, Anderson LA, Murphy SJ, Johnston BT, Murray LJ, et al. Cyclooxygenase-2 and inducible nitric oxide synthase gene polymorphisms and risk of reflux esophagitis, Barrett's esophagus, and esophageal adenocarcinoma. *Cancer Epidemiol Biomarkers Prev*. 2008; 17(3):727–31. <https://doi.org/10.1158/1055-9965.EPI-07-2570> PMID: 18349295
34. Anderson LA, Murphy SJ, Johnston BT, Watson RG, Ferguson HR, Bamford KB, et al. Relationship between *Helicobacter pylori* infection and gastric atrophy and the stages of the oesophageal inflammation, metaplasia, adenocarcinoma sequence: results from the FINBAR case-control study. *Gut*. 2008; 57(6):734–9. <https://doi.org/10.1136/gut.2007.132662> PMID: 18025067
35. Anderson LA, Watson RG, Murphy SJ, Johnston BT, Comber H, Mc Guigan J, et al. Risk factors for Barrett's oesophagus and oesophageal adenocarcinoma: results from the FINBAR study. *World J Gastroenterol*. 2007; 13(10):1585–94. <https://doi.org/10.3748/wjg.v13.i10.1585> PMID: 17461453
36. Murphy SJ, Anderson LA, Johnston BT, Fitzpatrick DA, Watson PR, Monaghan P, et al. Have patients with esophagitis got an increased risk of adenocarcinoma? Results from a population-based study. *World J Gastroenterol*. 2005; 11(46):7290–5. <https://doi.org/10.3748/wjg.v11.i46.7290> PMID: 16437630
37. Holl K, Lundin E, Kaasila M, Grankvist K, Afanasyeva Y, Hallmans G, et al. Effect of long-term storage on hormone measurements in samples from pregnant women: the experience of the Finnish Maternity Cohort. *Acta Oncol*. 2008; 47(3):406–12. <https://doi.org/10.1080/02841860701592400> PMID: 17891670
38. Gislefoss RE, Grimsrud TK, Morkrid L. Stability of selected serum proteins after long-term storage in the Janus Serum Bank. *Clin Chem Lab Med*. 2009; 47(5):596–603. <https://doi.org/10.1515/CCLM.2009.121> PMID: 19290843
39. McGlynn KA, Graubard BI, Nam JM, Stanczyk FZ, Longnecker MP, Klebanoff MA. Maternal hormone levels and risk of cryptorchism among populations at high and low risk of testicular germ cell tumors. *Cancer Epidemiol Biomarkers Prev*. 2005; 14(7):1732–7. <https://doi.org/10.1158/1055-9965.EPI-05-0128> PMID: 16030109
40. Thomas HV, Key TJ, Allen DS, Moore JW, Dowsett M, Fentiman IS, et al. A prospective study of endogenous serum hormone concentrations and breast cancer risk in post-menopausal women on the island of Guernsey. *British journal of cancer*. 1997; 76(3):401–5. PMID: 9252211
41. Thomas HV, Key TJ, Allen DS, Moore JW, Dowsett M, Fentiman IS, et al. A prospective study of endogenous serum hormone concentrations and breast cancer risk in premenopausal women on the island of Guernsey. *British journal of cancer*. 1997; 75(7):1075–9. PMID: 9083346
42. Vatten LJ, Ursin G, Ross RK, Stanczyk FZ, Lobo RA, Harvei S, et al. Androgens in serum and the risk of prostate cancer: a nested case-control study from the Janus serum bank in Norway. *Cancer Epidemiol Biomarkers Prev*. 1997; 6(11):967–9. PMID: 9367072
43. Stroud LR, Solomon C, Shenassa E, Papandonatos G, Niaura R, Lipsitt LP, et al. Long-term stability of maternal prenatal steroid hormones from the National Collaborative Perinatal Project: still valid after all these years. *Psychoneuroendocrinology*. 2007; 32(2):140–50. <https://doi.org/10.1016/j.psyneuen.2006.11.008> PMID: 17270355
44. Caron P, Turcotte V, Guillemette C. A chromatography/tandem mass spectrometry method for the simultaneous profiling of ten endogenous steroids, including progesterone, adrenal precursors,

- androgens and estrogens, using low serum volume. *Steroids*. 2015; 104:16–24. <https://doi.org/10.1016/j.steroids.2015.07.009> PMID: 26254607
45. Sodergard R, Backstrom T, Shanbhag V, Carstensen H. Calculation of free and bound fractions of testosterone and estradiol-17 beta to human plasma proteins at body temperature. *J Steroid Biochem*. 1982; 16(6):801–10. PMID: 7202083
 46. Vermeulen A, Verdonck L, Kaufman JM. A critical evaluation of simple methods for the estimation of free testosterone in serum. *J Clin Endocrinol Metab*. 1999; 84(10):3666–72. <https://doi.org/10.1210/jcem.84.10.6079> PMID: 10523012
 47. Starka L, Pospisilova H, Hill M. Free testosterone and free dihydrotestosterone throughout the life span of men. *J Steroid Biochem Mol Biol*. 2009; 116(1–2):118–20. <https://doi.org/10.1016/j.jsbmb.2009.05.008> PMID: 19465126
 48. Leymarie P, Roger M, Castanier M, Scholler R. Circadian variations of plasma testosterone and estrogens in normal men. A study by frequent sampling. *J Steroid Biochem*. 1974; 5(2):167–71. PMID: 4407742
 49. Rothman KJ, Greenland S, Lash TL. *Modern epidemiology*. 3rd ed. Philadelphia: Wolters Kluwer Health/Lippincott Williams & Wilkins; 2008. x, 758 p. p.
 50. Benjamini Y, Hochberg Y. Controlling the False Discovery Rate—a Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society Series B-Methodological*. 1995; 57(1):289–300.
 51. Muller M, den Tonkelaar I, Thijssen JH, Grobbee DE, van der Schouw YT. Endogenous sex hormones in men aged 40–80 years. *Eur J Endocrinol*. 2003; 149(6):583–9. PMID: 14641001
 52. Burney BO, Garcia JM. Hypogonadism in male cancer patients. *J Cachexia Sarcopenia Muscle*. 2012; 3(3):149–55. <https://doi.org/10.1007/s13539-012-0065-7> PMID: 22528986
 53. Burney BO, Hayes TG, Smiechowaska J, Cardwell G, Papusha V, Bhargava P, et al. Low testosterone levels and increased inflammatory markers in patients with cancer and relationship with cachexia. *J Clin Endocrinol Metab*. 2012; 97(5):E700–9. <https://doi.org/10.1210/jc.2011-2387> PMID: 22419719
 54. Garcia JM, Li H, Mann D, Epner D, Hayes TG, Marcelli M, et al. Hypogonadism in male patients with cancer. *Cancer*. 2006; 106(12):2583–91. <https://doi.org/10.1002/cncr.21889> PMID: 16688773
 55. Ueo H, Matsuoka H, Sugimachi K, Kuwano H, Mori M, Akiyoshi T. Inhibitory effects of estrogen on the growth of a human esophageal carcinoma cell line. *Cancer Res*. 1990; 50(22):7212–5. PMID: 2224855
 56. Sukocheva OA, Wee C, Ansar A, Hussey DJ, Watson DI. Effect of estrogen on growth and apoptosis in esophageal adenocarcinoma cells. *Dis Esophagus*. 2013; 26(6):628–35. <https://doi.org/10.1111/dote.12000> PMID: 23163347
 57. Asanuma K, Iijima K, Shimosegawa T. Gender difference in gastro-esophageal reflux diseases. *World J Gastroenterol*. 2016; 22(5):1800–10. <https://doi.org/10.3748/wjg.v22.i5.1800> PMID: 26855539