# Terahertz-based system for dehydration analysis of hydrogel contact lenses

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The use of terahertz-based techniques has grown very fast since they are capable of performing evaluations at molecular level, being very suitable for the analysis of biological samples and biomaterials such those for contact lenses. These biomaterials are continuously evolving to enhance the lens wearer's comfort by improving their hydration state and surface wettability. Therefore, this study examines a novel terahertz system for the assessment of the temporary in-vitro dehydration of hydrogel contact lenses, which provides a new index to assess their state of hydration. Several conventional and silicone hydrogel contact lenses and lens care solutions were analysed. Traditional methods such as the gravimetric determination of water content and the measurement of the static contact lenses obtained with the proposed system correlated with the values provided by traditional methods. As a whole, conventional hydrogel contact lenses exhibited the lowest values for dehydration rate. The tests conducted on various solutions showed a correlation between the wetting action of the solution and the dehydration rate of the contact lens material.

Keywords: terahertz radiation, terahertz systems, hydrogel contact lens dehydration, water content, lens care solutions.

## 1. Introduction

The use of new methods based on terahertz (THz) radiation is growing fast in biomedical research, mainly because it is a non-invasive technique with non-ionizing properties [1]. THz radiation has a spectral range between 100 GHz and 10 THz, corresponding to wavelengths from 3 mm in the microwave domain to 30  $\mu$ m in the far infrared. Specifically, THz energy corresponds to the intermolecular oscillations, which can identify and differentiate between different polymorphic crystals or crystal orientations of the same molecule.

THz radiation is currently used to determine the hydration state of living cells and biomaterials since the dielectric responses are selectively sensitive to bulk water [2]. Any other polar liquid can also absorb THz radiation and can thus be used in remote sensing of any process that results from changes in the water content (WC) of the sample [1]. For instance, it is commonly applied to non-invasive inspection of the WC in leaves [3,4], drying paint [5,6] and to evaluate variations in WC when a healthy tissue becomes cancerous [7,8].

The development of hydrogel biomaterials for contact lens (CL) application has been driven by the need to increase the biocompatibility of CLs as a means to extend the wearing time. Biocompatibility is dependent on multiple factors, which can be classified in 2 main categories: (1) bulk parameters, such as oxygen permeability, Dk, and equilibrium WC; (2) interfacial phenomena, such as wettability of the polymeric surface. In essence, biocompatibility of CLs refers to the characteristics of both bulk and surface properties when interacting with the tear film and the lens care solution.

Conventional hydrogel biomaterials for CLs were based in the synthetic biocompatible material poly(hydroxyethylmethacrylate) (PHEMA), which presents a matrix surface with hydrophilic character that allows sufficient surface wettability at the interface between the CL and the tear film. The increased equilibrium WC of the hydrogel polymeric matrix has led to the development of new PHEMA-based hydrogel materials with higher Dk [9]. On the other hand, the first generation of silicone hydrogel biomaterials for cCLs was formulated by block

polymerization of tris(trimethylsiloxy)silylpropylmethacrylate (TRIS), polydimethylsiloxane (PDMS) and dimethylacrylamide macromers [10] that resulted in higher Dk values than conventional hydrogel CLs. However, the hydrophobic character of the polymeric surface requires treatment to ensure wettability. This is usually achieved by increasing the WC of the hydrogel matrix without decreasing its Dk, while also lowering the tensile modulus of the first silicone hydrogel generation materials to improve comfort [11,12].

With regard to the evaluation of WC and CL dehydration, gravimetric [13-16] and refractometric [17,18] methods have been widely applied to silicone and conventional hydrogel CLs for in-vitro evaluation, since they can determine the dehydration rate in different environments. Wettability of silicone and conventional hydrogel materials has also been assessed by means of the contact angle of pure water and surfactant solutions for CLs preserved in different solutions [19,20].

In this work, we propose a novel THz system for the in-vitro evaluation of the dehydration process of different materials used for conventional and silicone hydrogel CLs and the influence of conditioning solutions with different surfactants and lubricant agents. The gravimetric WC and wettability of the CL polymeric surface were also assessed to validate the performance of the system. The main goal was to provide a more precise analysis of water loss considering the bulk and superficial performance of CL materials by means of a THz-based approach that, to the authors' knowledge, has not been used previously for this purpose. Furthermore, the system can contribute to a better understanding of user's wearability of hydrogel CLs as a function of the material, WC and the solution used to achieve the hydrated state of the hydrogel as complementary and valuable information not available in the literature can be obtained by means of the proposed technique.

# 2. Material and methods

## 2.1 Dehydration measurements

## 2.1.1 Experimental setup

The dehydration process of CLs was evaluated with a novel THz-based system (Figure 1), which consisted of a mechanically tuned Gunn oscillator SOM-94301313-10-SC coupled to a voltage regulator SOR-R3, and an amplitude GaAs Schottky detector SFD-753114-10SF-N from SAGE Millimeter, Inc. The Gunn oscillator was fed with 7.3 V and emitted a linearly polarized, free space radiation at 94 GHz with approximately 1 GHz frequency modulate bandwidth. Additionally, 2 SGH-26-WR10 B horn antennas from Anteral, S.L. were attached to the oscillator and detector; a polarizer FPOL-MMW-20 from the same company was also placed in front of the oscillator-antenna assembly in order to maximize the signal. A NI USB-3211 acquisition card from National Instruments Corp. was incorporated to retrieve the signal detected by the sensor.



Fig. 1. (a) General view of the setup and (b) ray tracing from the emitter to the detector. L1, L2, L3 and L4 refer to the four Teflon<sup>TM</sup> lenses.

The beam propagation from the emitter to the detector was handled by a configuration of 4 Teflon<sup>TM</sup> lenses from Thorlabs GmbH, which included 2 collimating lenses of 100-mm focal length (L1 and L4) and two aspherical lenses with a focal length of 50 mm (L2 and L3). In order to guarantee the perpendicular positioning of the CL with respect to the optical path, a transparent plastic holder with a semi-A sphere of 8-millimeter radius in the centre that simulated the cornea was used.

#### 2.1.2 Hydration index (HI)

Dehydration was measured in terms of transmission by comparing the voltage detected with and without the CL in the optical path. This transmitted voltage was expressed as a percentage through a hydration index (*HI*) as follows:

$$HI = 100 - \frac{(V_0 - V_i)}{V_0} \cdot 100, \tag{1}$$

 $V_0$  refers to the initial value of voltage and  $V_i$  indicates the voltage for each individual measurement over time. The computation of each voltage value (either  $V_0$  or  $V_i$ ,) was calculated as follows,

$$V_i = 1 - \frac{R_i - \Delta R}{R_H} \tag{2}$$

where  $R_i$  is the raw voltage value obtained when measuring the transmission of the CL and its holder,  $R_H$  is the voltage transmitted only using the holder; the influence of the holder on the total transmission is thus avoided. Moreover, dividing by  $R_H$  also benefits the minimization of the effect of environmental conditions such as variations of humidity, i.e., water vapour present in the air, which where monitored during measurements, showed fluctuations below 2%. The term  $\Delta R$  is calculated as  $R_n$ -  $R_H$  and was added to obtain *HI* values of zero when the signal was constant, which meant that the CL was completely dehydrated at time *n*. At this time, some CLs showed a constant signal but with values slightly above zero, which might have been caused by the inner structure of the material. In relation to the assessment of different lens care solutions, the  $\Delta R$ remained constant for each CL material but was different among the solutions. In this case, the  $\Delta R$  selected as a reference was the  $\Delta R$  of the CL preconditioned in MilliQ<sup>®</sup> water (DW). This voltage ratio was finally subtracted from 1 to obtain a curve decreasing over time in consonance with the expected behaviour of the *HI*.

## 2.1.3 CLs preconditioning

With regard to the assessments of the different solutions, once the CLs had been 24 hours in DW and before the THz measurements, they were immersed in the solution under test for 24 additional hours. Next, the residual water was eliminated with lens cleaning tissue and the sample was weighted to establish the initial mass. The time needed for these processes was about 1 minute to minimize dehydration before THz measurements. The dehydration was measured until the complete stabilization of the signal, indicated by constant and very low values of voltage. Each CL was tested once, but the transmitted voltage measurement was repeated at least twice, with new samples for each material and preconditioning process, in order to assure the repeatability of the *HI* index. Other factors also considered during the preconditioning processes were the use of silicone tweezers and nitrile gloves to avoid any contamination of the CLs. Room temperature and relative humidity were kept constant at  $22\pm1^{\circ}$ C and  $32\pm2$  %, respectively.

#### 2.1.4 Gravimetric determination of the WC

The WC of the different materials was gravimetrically quantified by means of a PCE-AB 100 analytical balance from PCE Instruments<sup>TM</sup>, with a precision of 0.1 mg. The following formula was used to calculate the WC as a percentage:

$$WC = \frac{(m_0 - m_n)}{m_0} \cdot 100,\tag{3}$$

where  $m_0$  refers to the initial value of mass measured just after the preconditioning process, and  $m_n$  indicates the final mass calculated when the CL is completely dehydrated, i.e., when the THz signal becomes stable.

#### 2.2 CL materials and solutions

Eight CLs were analysed: 5 silicone hydrogel (SiHy) and 3 conventional hydrogel (Hy), i.e., poly(hidroxyethylmethacrylate)-based, CLs (Table 1). All had the same optical power (-4.00 D) to prevent differences among them. Concerning hydration, 3 CL materials with high (> 50%) and 5 with low WC (< 50%) were included in the study. With regard to the material characteristics, 2 were ionic and 6 non-ionic. This sample set consisted of monthly and biweekly replacement CLs except for the etafilcon A, which is a material for daily use only.

Table 1. Material name (United States Generic name), principal monomers, nominal water content ( $WC_{Nominal}$ ), ionic character (FDA), surface treatment, and brand name of the contact lenses evaluated.

Material	Principal monomers	WC <sub>nomial</sub> (%)	Ionic character	Surface treatment	Brand name
lotrafilcon B	DMA, TRIS, fluorine containing siloxame macromer	33	Non-ionic SiHy	Plasma coating	Air Optix
senofilcon A	mPDMS, DMA, HEMA, SiGMA, TEGDMA, PVP	38	Non-ionic SiHy	None	Acuvue Oasys
asmofilcon A	Silicone methacrylates, silicone acrylates, DMA, pyrrolidone derivative	40	Non-ionic SiHy	Plasma coating	Menicon PremiO
comfilcon A	M3U, FMM, TAIC, IBM, NMNVA, NVP, HOB	48	Non-ionic SiHy	None	Biofinity
balafilcon A	NPV, TPVC, NVA, PBVC	36	Ionic SiHy	Plasma oxidation	Pure Vision
omafilcon A	HEMA, PC	62	Non-Ionic Hy	-	Proclear
hilafilcon B	HEMA, NVP	59	Non-Ionic Hy	-	Acuvue 1- Day
etafilcon A	HEMA, MA	58	Ionic Hy	-	SofLens59

DMA: N,N-dimethylacrilamida; PVP: poly(vinyl pyrrolidona); TRIS: methacriloxypropyl tris(trimetrhyl siloxy)silane; mPDMS: monofuncional methacryloxypropyl terminated polydimethylsiloxane; HEMA: 2-hydroxyethylmethacrylate; SiGMA: 2-propenoic acid-2-methyl-2hydroxy-3(3(1,3,3,3-tetramethyl-1-(trimethylsilyl)oxy)disiloxanyl)propoxypropyl ester; TEGDMA: tetraethyleneglycol dimethacrylate; NVP: N-vynilpirrolidone; PC: phosphorylcoline; MA: methacrilyc acid; M3U:  $\alpha \omega$ -bis(methacryloyloxiethyl iminocarboxy ethyloxypropyl)-poly(dimethylsiloxane)-poly(trifluoropropylmethylsiloxane)-poly(methoxy-poly(ethyleneglycol)propylmethyl-siloxane; FMM:  $\alpha$ -methacryloyloxiethyl iminocarboxyethyloxypropyl-poly(dimethylsiloxy)-buthyldimethylsilane; TAIC: 1,3,5-triallyl-1,3,5-triazine-2,4,6(1H,3H,5H)-trione; IBM: isobornyl methacrylate; HOB: 2-hydroxybuthyl methacrylate; NMNVA: N-methyl-Nvinylacetamide; HOB:

isobornyl methacrylate; HOB: 2-hydroxybuthyl methacrylate; NMNVA: N-methyl-Nvinylacetamide; HOB:
2.hydroxybuthyl methacrylate; TPVC; tris-(trinethyl siloxysilyl) propylvinyl carbamate; NVA: N-vinyl amino acid;
PBVC: poly(dimethylsiloxy) di (silylbutanol) bis (vinyl carbamate).
Borate buffered saline solution was prepared using analytical grade reagents from Sigma-

Aldrich Co and deionized ultrapure MilliQ<sup>®</sup>. The BBS solution (pH =  $7.40\pm0.02$  and osmolarity of  $304 \pm 0.5$  mOsm/L) contained boric acid (0.01M), sodium tetraborate decahydrate (0.16 mM) and sodium chloride (0.14 M). BBS solutions containing in addition hyaluronic acid sodium salt (HA, molecular weight of 1.5-1.8  $10^3$  kDa) and/or purified Poloxamer-407 (P, molecular weight

12.6 kDa,) were also prepared. The BBS-HA, BBS-P, and BBS-HA-P solutions contained 0.01 % wt/vol HA, 0.2 % wt/vol P and 0.01 % wt/vol HA + 0.2 % wt/vol P, respectively. Finally, the commercial multi-purpose solution Biotrue<sup>®</sup> from Bausch&Lomb (pH =  $7.50\pm0.02$  and osmolarity =  $285\pm1.0$  mOsm/L) [21, 22] was also evaluated.

## 2.3 Static contact angle evaluation

The static contact angles ( $\theta$ ) of water drops on the surface of CLs were measured by means of the sessile drop method using a DSA100 drop shape analyser from KRÜSS GmbH. All measurements were also conducted at a room temperature of  $22\pm1^{\circ}$ C. Prior to these measurements, the CLs were removed from the blister packages and immersed in BBS solution during two minutes, followed by the immersion in the test solution for 48 hours.

The methodology followed to obtain the contact angle measurements was based on previous works [19, 20]. Firstly, the CL was removed from the storage solution using silicone tweezers and repeatedly placed on a lens cleaning tissue to collect any residual surface liquid. Next, the CL was placed on a holder and a 4  $\mu$ l water drop controlled by a micrometre pass dosage was transferred to the CL surface, ensuring a total time of exposure to air of 2 minutes. This process was captured in the digital video by the camera integrated in the drop shape analyser, controlled and analysed by the software DSA4. A tangent method was used to measure the contact angles. Three independent fittings were performed to provide three mean contact angles of the same drop image and the average ( $\theta_m$ ) was determined. This procedure was repeated for a second video of the same CL solution to verify the reproducibility of the measurement.

# 3. Results and discussion

## 3.1. Dehydration evaluation of different CL materials preconditioned in DW

Figure 2 shows the dehydration process, indicating the *HI* index for the eight CLs preconditioned in DW at different times (Figure 2a) and for 5000 s (Figure 2b). The first time of 20 s was selected to simulate natural eye conditions, where the average tear film break up time (TFBUT) is 17 s, approximately<sup>29</sup>. On the other hand, between 4000 s and 5000 s, the voltage of all CLs remained constant and at minimum values; therefore, 5000 s were thus assumed to be enough for all CLs to dehydrate completely.



Fig. 2. Dehydration evolution for the eight contact lenses conditioned in MilliQ<sup>®</sup> water (DW) analysed at different times (a) and during 5000 s (b).

The dehydration process within the first time interval (0 s, 20 s) in Figure 2b revealed that the *HI* was not dependent on the tested WC of the CL material (Table 2). Six CL materials showed a mean dehydration under 2 % (balafilcon A, comfilcon A, omafilcon A, hilafilcon B and lotrafilcon

B); 2 other materials exhibited values between 2 % and 3 % (etafilcon A and senofilcon A); and only one led to a mean dehydration of almost 5 % (asmofilcon A). In the second time interval (20 s, 1000 s), the dehydration was more dependent on the tested WC and the CL polymeric matrix material. All Hy CLs had dehydration rates lower than SiHy materials, as expected due to their higher WC, with the exception of comfilcon A. The three Hy CLs and comfilcon A (SiHy) showed a  $HI \ge 40\%$  at t = 1000 s. Among all Hy CLs evaluated, the hilafilcon A material presented the lowest dehydration rate, i.e., the highest HI in this range; the ionic character did not seem to influence the dehydration process. In the case of SiHy materials, comfilcon A reached the highest HI values up to 2000s, which corresponds to its inherent wettability as a third generation SiHy CL [10]. The remaining SiHy materials showed HI < 40% at t = 1000 s. The ionic material balafilcon A exhibited the slowest dehydration rate, even though its dehydration behaviour is very similar to that of lotrafilcon B, both belonging to the first generation of SiHy CLs. Although the HI decrease of senofilcon A was initially faster, it presented the slowest dehydration decay beyond 1000 s (third time interval) due to the PVP internal wetting agent. Beyond 3000 s, most CL materials were completely dehydrated. The materials with the longest dehydration times were etafilcon B and senofilcon A, while asmofilcon A and omafilcon A presented the shortest dehydration times, with  $HI \sim 0$  at 2500 s and 2750 s, respectively.

Table 2 shows the WC provided by the manufacturers (WC<sub>Nominal</sub>) and the mean and standard deviation (SD) values of WC experimentally measured as previously described in equation (3) (WC<sub>Tested</sub>) for the eight CLs and calculated at t = 5000 s. A good Pearson correlation was obtained between both WC<sub>Tested</sub> and WC<sub>Nominal</sub> values (R<sup>2</sup>= 0.9854, p<0.0001), with a decreasing tendency in the WC<sub>Tested</sub> values for the CLs conditioned in DW respect to the WC<sub>Nominal</sub> values measured in standard saline solution (following ISO 18369-3: 2017 for measuring methods in ophthalmic optics).

Material	WC <sub>Nominal</sub>	$WC_{Tested} \pm SD$
lotrafilcon B	33	$32.7\pm0.6$
senofilcon A	38	$37.0\pm0.0$
asmafilcon A	40	$41.3\pm1.2$
comfilcon A	48	$45.0\pm1.7$
balafilcon A	36	$36.3\pm0.6$
omafilcon A	62	$61.7\pm1.2$
hilafilcon B	59	$57.7\pm4.6$
etafilcon A	58	$58.0 \pm 1.7$

Table 2. Nominal and tested WC of the eight contact lenses. Both are expressed as a percentage.

In general, the THz evaluation showed good inter-blink hydration since the mean *HI* of all CL materials was above 95% during the first 20 s. Because of their slower dehydration decay, hilafilcon B, etafilcon A and comfilcon A were shown to be the most comfortable materials for long-time use. These results agreed with other studies where conventional hydrogel materials also presented lower dehydration decay than silicone-containing hydrogels [15] and CLs of higher WC dehydrated at a slower rate [23].

#### 3.2. Dehydration evaluation of CLs preconditioned in different lens care solutions

Additionally, the dehydration of balafilcon A and senofilcon A materials was studied after preconditioning the CLs in the different prepared solutions and in the commercial Biotrue<sup>®</sup> solution. Balafilcon A and senofilcon A CLs were chosen to account for materials with similar WC belonging to different SiHy generations and with different ionic character. The objective of this second test was to study the behaviour of CL dehydration when using solutions with different lubricant or wetting agents.

The latest formulations of multipurpose CL solutions [24] try to mimic the physicochemical properties of natural tear film (pH, osmolarity, viscosity and surface tension) [25-27]. This is the

case of the Biotrue<sup>®</sup> and some of the prepared solutions, which incorporate HA and poloxamers or poloxamines agents. HA is a natural polysaccharide present in biological fluids and tissues, including human tears, that is commonly used in CL solutions for its wetting and lubricant properties. Poloxamers and poloxamines are non-ionic surfactants [28].

Figures 3 and 4 show the evolution of the *HI* index of both materials conditioned in the different solutions at different times and during 5000 s, respectively. In both figures the dehydration in MilliQ<sup>®</sup> DW was also included as reference.



Fig. 3. Dehydration evolution of the balafilcon A (a) and the senofilcon A (b) contact lens materials conditioned in the different lens care solutions at different times.



Fig. 4. Dehydration evolution of the balafilcon A (a) and the senofilcon A (b) contact lens materials conditioned in the different lens care solutions during 5000 s.

The assessments of balafilcon A revealed that for the first time interval (0 s, 20 s), the mean *HI* was above 96% for all solutions. The highest mean *HI* was obtained in the case of DW, BBS and the multipurpose Biotrue<sup>®</sup> solutions (Figure 3a). In the second interval (20 s, 1000 s), the DW led to higher mean *HI* than BBS and both of them presented higher mean *HI* than the rest of solutions (Figures 3a and 4a). Moreover, the decrease of the *HI* was constant for the Biotrue<sup>®</sup>, BBS-HA-P, BBS-HA and BBS-P solutions until around 95% of water had been lost, presenting a change in the dehydration profile respect to that of DW. For all solutions and DW, balafilcon A became completely dehydrated around 3000 s (Figure 4a) and the values of WC tested were

almost identical to the nominal WC (Table 3), except for the Biotrue<sup>®</sup> that presented a value slightly higher and for the BBS-P slightly lower. This allowed us to conclude that the total equilibrium WC of balafilcon A is few affected by the presence of surfactant or lubricant agents in the lens care solutions. However, in contrast with DW, the osmolarity change and the addition of these agents in the preconditioning solution affected the kinetics of the dehydration process increasing the dehydration rate. This phenomenon was less noticeable during the first 20 s of the dehydration process since a similar performance with a decrease of the *HI* below 6 % was observed for all solutions.

On the other hand, the tests for senofilcon A in the interval (0 s, 20 s) showed that the mean HI was above 94% for all solutions. The highest mean HI values were obtained for the BBS-HA, BBS-HA-P and Biotrue® (Figure 3b). In the interval (20 s, 1000 s) BBS-P, BBS-HA, BBS-HA-P, and Biotrue<sup>®</sup> solutions produced very similar dehydration curves (Figure 4b), always above those of DW and BBS, with higher mean HI values at t=1000 s (Figure 3b). Senofilcon A became completely dehydrated around 4000 s in the case of DW and BBS, and at 4500 s in the case of BBS-P; in all cases, the dehydration process lasted longer than that of balafilcon A. For the remaining solutions, an apparent residual HI value under 5% was observed when the signal was considered stable (5000 s). However, the values of WC tested at 5000 s (Table 3) indicated that in all cases the senofilcon A CLs were completely dehydrated since these values essentially coincided with the nominal WC. On the one hand, it can be concluded that the total equilibrium WC of senofilcon A behaves similar to that found for balafilcon A, i.e., is almost independent from the presence of these agents. On the other hand, the THz data transmitted by the senofilcon A material in a dehydrated state seemed to be affected by the presence of HA, P and/or poloxamine in the BBS solution, since a remaining amount of HI persisted up to 5000 s. It probably occurred because these agents were adsorbed on the polymeric matrix surface during the preconditioning process. In order to obtain more in depth information about this fact, the contact angle of both balafilcon A and senofilcon A conditioned in DW and in the different care solutions was measured.

The gravimetric determination of the WC and the static contact angle were also experimentally evaluated for this second set of measurements. The contact angles shown in Table 3 indicate the effect of the lens care solutions on the polymeric surface wettability of the CLs. As previously mentioned, the balafilcon A material incorporates a surface treatment to increase its wettability, which was confirmed by the contact angle values when conditioned in DW or BBS. The presence of only one agent (HA or P) in the BBS solution did not produce noticeable changes in surface wettability. However, the inclusion of two agents as in the BBS-HA-P (HA and P) and the Biotrue® solutions (HA and poloxamine) achieved an improvement of surface wettability. Comparing the contact angle measurements with the mean HI values in the interval time (0 s, 20 s) in Figure 3a for the 4 solutions containing surfactant and lubricant agents, the BBS-HA and Biotrue<sup>®</sup> showed slightly higher HI values. The selection of the most suitable solution for balafilcon A is complex because this material showed a very good hydration performance when using only DW and BBS. However, the wettability of the surface is also important in the ocular environment, since it determines how the tear film is spread over the surface of the CL. Considering the results for these two parameters, BBS-HA-P and Biotrue® would be the best solutions due to their high mean HI value during the first 20 s and the low contact angles, although long-term HI values were below those of DW and BBS.

S alasti an	balafilcon A		senofilcon A		
Solution	WC tested $\pm$ SD	$\theta_m \pm SD$	WC tested $\pm$ SD	$\theta_m \pm SD$	
DW	$36.3\pm0.6$	$29.9\pm2.6$	$37.0\pm0.0$	$97.2 \pm 1.2$	
BBS	$35.0\pm0.0$	$33.6\pm1.3$	$38.0\pm0.0$	$55.1 \pm 1.7$	
BBS-P	$33.5\pm2.4$	$31.0\pm0.8$	$36.5\pm0.7$	$36.8\pm2.1$	
BBS-HA	$36.5\pm0.7$	$36.7 \pm 1,9$	$38.0\pm0.0$	$26.8 \pm 1.1$	
BBS-HA-P	$35.0\pm0.0$	$25.1\pm0.6$	$36.0\pm0.0$	$39.0\pm2.6$	
Biotrue®	$39.0\pm0.0$	$26.6\pm0.3$	$38.0\pm0.0$	$39.5 \pm 1.6$	

Table 3. Tested WC in percentage and static contact angles ( $\theta_m$ , degrees) of the balafilcon A and the senofilcon A contact lenses for the different lens care solutions.

Senofilcon A incorporates PVP to increase the hydrophilicity of the surface [2]. Despite this fact, the contact angle of senofilcon A conditioned in DW and BBS solution indicated a low hydrophilic surface character (Table 3). Consequently, the presence of wetting or lubricant agents in the BBS solution improved the wettability considerably, especially in the case of HA. On the other hand, the hydrophobic character of the senofilcon A surface was shown to depend on the non-ionic surfactant agents added to the lens care solutions such as BBS-P, BBS-HA-P and Biotrue<sup>®</sup>. This behaviour indicated that non-ionic surfactants P and poloxamine were less effective than using HA without surfactants. The contact angles for senofilcon A also correlated with the mean *HI* values during the first 20 s; the lower the static contact angle, the higher the HI values. In addition, the stronger adsorption of HA and HA+P or HA+poloxamine on the senofilcon A surface correlated with the residual water content that was observed for these solutions at 5000 s (Figure 4b). Considering the results of *HI* and contact angle, the most suitable solution for senofilcon A CLs is the BBS-HA, since it showed the best hydration throughout time and wettability, with a contact angle below  $30^\circ$ .

## 4. Conclusions

A THz system was developed to evaluate the dehydration of conventional and silicone hydrogel CLs and to analyse the performance of different lens care solutions by means of a THz-based system. This presents a significant extension of existing works in this field as THz technology is used for the first time for the CLs study. Based on THz measurements, a new expression for the quantification of the hydration state of CLs was defined. The results of the new system correlated with results obtained with traditional methods.

The assessment of the in-vitro dehydration process of different CL materials conditioned in MilliQ<sup>®</sup> DW revealed different dehydration kinetics in the first stages (0s, 20s) compared to the dehydration kinetics at longer times (20 s, 5000s). Although the dehydration profile of the different materials was similar within the Hy group and among SiHy CLs of the same generation, each dehydration profile presented specific features that could be used as a fingerprint for material identification.

The study of the effect of different solutions on the in-vitro dehydration profile of CLs was analysed for the balafilcon A and senofilcon A materials. The solutions used were based on BBS in combination with wetting and/or lubricant components. For each solution, a correlation was observed between the measurements of dehydration by means of the *HI* and of the static contact angle.

To conclude, the THz-based system contributed to improve the accuracy in the study of hydrogel CL materials and their interaction with lens care solutions. The system evaluated the performance of biomaterials and solutions in terms of dehydration and can be a useful technique to evaluate new hydrogel biomaterials and their behaviour prior to in-vivo tests.

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#### References

- [1] Yang X, Zhao X, Yang K, Liu Y, Liu Y, Fu W, Luo Y (2016) Biomedical Applications of Terahertz Spectroscopy and Imaging. Trends Biotech 34:810-824.
- [2] Shiraga K, Suzuki T, Kondo N, Tanaka K, Ogawa Y (2015) Hydration state inside HeLa cell monolayer investigated with terahertz spectroscopy. Appl Phys Lett 106:253701-1-5.
- [3] Gente R, Koch M (2015) Monitoring leaf water content with THz and sub-THz waves. Plant Methods 11:15. https://doi.org/10.1186/s13007-015-0057-7.
- [4] Breitenstein B, Scheller M, Shakfa MK, Kinder T, Müller-Wirts T, Koch M, Selmar D (2001) Introducing terahertz technology into plant biology: A novel method to monitor changes in leaf water status. J Appl Bot Food Qual 84:158-161.
- [5] Shen YC (2014) Terahertz Sensor for Noncontact and Non-destructive Inspection of Automotive Paints. Sensor Netw Data Commun 4:e103. doi:10.4303/2090-4886.1000e103
- [6] Fukunaga K, Hosako I (2010) Innovative non-invasive analysis techniques for cultural heritage using terahertz technology. C R Phys 11:519-526.
- [7] Yu C, Fan S, Sun Y, Pickwell-MacPherson E (2012) The potential of terahertz imaging for cancer diagnosis: A review of investigations to date. Quant Imaging Med Surg 2: 33-45.
- [8] Rahman A, Rahman AK, Rao B (2016) Early detection of skin cancer via terahertz spectral profiling and 3D imaging. Biosens Bioelectron 82:64-70.
- [9] Nicolson PC, Vogt J. Soft contact lens polymers: an evolution. Biomaterials 2001;22:3273–3283.
- [10] Jacob JT. Biocompatibility in the Development of Silicone-Hydrogel Lenses. Eye Contact Lens 2013;39:13-19.
- [11] Tighe BT (2013) A decade of Silicone Hydrogel Development: Surface properties, Mechanical Properties, and Ocular Compatibility. Eye Contact Lens 39:4-12.
- [12] Hutter JC, Green JA, Eydelman MB (2012) Proposed Silicone Hydrogel Contact lens Grouping System for Lens Care Product Compatibility System testing. Eye Contact Lens 38:358-362.
- [13] Jones L, May C, Nazar L, Simpson T (2002) In vitro evaluation of the dehydration of silicone hydrogel and conventional hydrogel contact lens materials. Contact Lens & Anterior Eye 25:147–156.
- [14] González-Méijome JM, López-Alemany A, Almedia JB, Parafita MA (2009) Dynamic in vitro dehydration patterns of unworn and worn silicone hydrogel contact lenses. J Biomed Mater Res B 90B:250–258.
- [15] Krysztofiak K, Szyczewski A (2014) Study of dehydration and water states in new and worn soft contact lens materials. Optica Applicata XLIV:237-250
- [16] Rajchel D, Krysztofiak K, Szyczewski A (2016) Influence of sodium hyaluronate on dehydration and water distribution in soft contact lenses. Optica Applicata, XLVI:483-496.
- [17] Morgan PB, Efron N, Morgan SL, Little SA (2004) Hydrogel contact lens dehydration in controlled environmental conditions. Eye Contact Lens 30:99–102.
- [18] Alemany AL, Refojo MF, (2000) Comparative study of the hydration of hydrophilic contact lenses by refractive index and gravimetry. CLAO J 26:200-203.
- [19] Ketelson HA, Meadows DL, Stone RP (2005) Dynamic wettability properties of a soft contact lens hydrogel. Colloid Surface B 40:1-9.
- [20] Maldonado-Codina C, Morgan PB (2007) In vitro wettability of silicone hydrogel contact lenses determined using the sessile drop band captive Hubble techniques. J Biomed Mater Res A 83A: 496-502.
- [21] Contact lens Spectrum, Supplement: Bringing new inspiration to contact lens care, September (2010)
- [22] Liu XM, Harmon PS, Maziarz EP, Rah MJ, Merchea MM (2014) Comparative studies of Hyaluronan in marketed ophthalmic products. Optom Vis Sci 91: 32-38.
- [23] Efron N, Young G (1988) Dehydration of hydrogel contact lenses in vitro and in vivo. Ophthalmic Physiol Opt 8:253–6.
- [24] Dalton K, Subraman LN, Rogers R., Jones L (2008) Physical properties of soft contact lens solutions, Optom Vis Sc 85:122-128.
- [25] Benelli U, Nardi M, Posarelli C, Albert TG (2010) Tear osmolarity measurement using the TearLabTM Osmolarity System in the assessment of dry eye treatment effectiveness. Contact Lens & Anterior Eye 33:61–67.
- [26] Nagyová B, Tiffany J M (1999) Components responsible for the Surface tension of human tears. Current Eye Research, 19:4-11.
- [27] Gouveia SM, Tiffany J M (2005) Human tear viscosity: An interactive role for proteins and lipids. Biochimica et Biophisica Acta 1753:155-163.
- [28] Moghimi SM, Hunter AC (2000) Poloxamers and poloxamines in nanoparticle engineering and experimental medicine. Trends Biotech 18:412-420.