

Why Do Elastin-Like Polypeptides Possibly Have Different Solvation Behaviors in Water–Ethanol and Water–Urea Mixtures?

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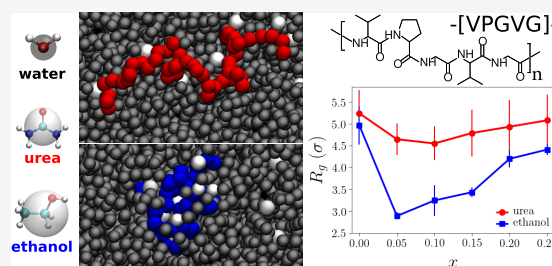
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ABSTRACT: The solvent quality determines the collapsed or the expanded state of a polymer. For example, a polymer dissolved in a poor solvent collapses, whereas in a good solvent it opens up. While this standard understanding is generally valid, there are examples when a polymer collapses even in a mixture of two good solvents. This phenomenon, commonly known as co-non-solvency, is usually associated with a wide range of synthetic (smart) polymers. Moreover, recent experiments have shown that some biopolymers, such as elastin-like polypeptides (ELPs) that exhibit lower critical solution behavior T_1 in pure water, show co-non-solvency behavior in aqueous ethanol mixtures. In this study, we investigate the phase behavior of elastin-like polypeptides (ELPs) in aqueous binary mixtures using molecular dynamics simulations of all-atom and complementary explicit solvent generic models. The model is parameterized by mapping the solvation free energy obtained from the all-atom simulations onto the generic interaction parameters. For this purpose, we derive segment-based (monomer level) generic parameters for four different peptides, namely proline (P), valine (V), glycine (G), and alanine (A), where the first three constitute the basic building blocks of ELPs. Here, we compare the conformational behavior of two ELP sequences, namely -(VPGGG)- and -(VPGVG)-, in aqueous ethanol and -urea mixtures. Consistent with recent experiments, we find that ELPs show co-non-solvency in aqueous ethanol mixtures. Ethanol molecules have preferential binding with all ELP residues, with an interaction contrast of 6–8 $k_B T$, and thus driving the coil-to-globule transition. On the contrary, ELP conformations show a weak variation in aqueous urea mixtures. Our simulations suggest that the glycine residues dictate the overall behavior of ELPs in aqueous urea, where urea molecules have a rather weak preferential binding with glycine as observed from the all atom simulations, i.e., less than $k_B T$. This weak interaction dilutes the overall effect of other neighboring residues and thus ELPs exhibit a different conformational behavior in aqueous urea in comparison to aqueous ethanol mixtures. While the validation of the latter findings will require a more detailed experimental investigation, the results presented here may provide a new twist to the present understanding of cosolvent interactions with peptides and proteins.



1. INTRODUCTION

Solvation of macromolecules in water and especially in a mixture of solvents is of central relevance for many areas of chemical physics, polymer physics, soft matter science, and material research.^{1–7} Indeed, solvation effects are the driving force underlying various macromolecular processes ranging from the responsiveness of hydrogels to external stimuli or concentration gradients of the solvents (“smart polymers”) to denaturation of proteins. Furthermore, the relevant energy scale in these systems is of the order of thermal energy $k_B T$, where k_B is the Boltzmann constant and $T = 300$ K, thus, the properties of macromolecules are dictated by large conformational and compositional fluctuations. Therefore, entropy (or generic physical laws) becomes as crucial as energy (or specific chemical details) for the study of these complex systems. Admittedly, understanding this entropy–energy balance is at the heart of soft matter science.^{8–10}

The flexibility of macromolecules provides a suitable platform for the tunable design of advanced functional

materials.^{11–16} Furthermore, because of the carbon-based microscopic architectures, they often create severe environmental problems. To circumvent this problem, recent interests have been directed toward the “so-called” green chemistry,¹⁷ more specifically, making use of macromolecular structures that are biocompatible¹⁸ and/or biodegradable¹⁹ and at the same time are also thermal,^{16,18} (co-)solvent,^{20–30} and photoresponsive.^{31,32} While most of these systems are homopolymers, recent interest has been directed to a variety of copolymer architectures.^{33–42} Here, polypeptides and synthetic peptide-based polymers have attracted great inter-

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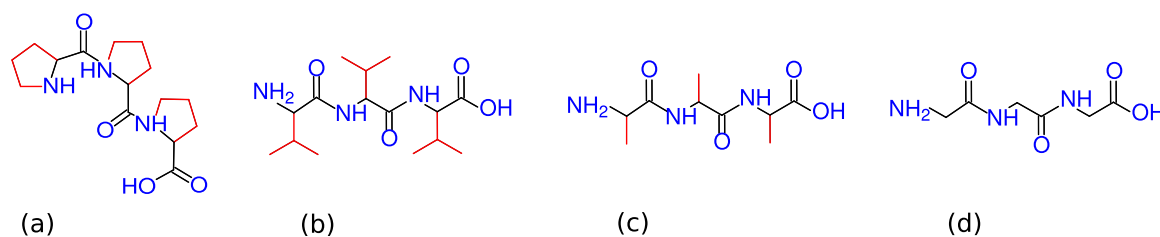


Figure 1. Schematic representation of the chemical structures of all four peptides, namely (a) triproline, (b) trivaline, (c) trialanine, and (d) triglycine. The hydrophilic parts are highlighted in blue, while the side chains are indicated in red.

est.^{39,42} In this context, elastin-like polypeptides (ELPs) represent a new class of stimuli-responsive synthetic polypeptides that show vibrant phase behavior.^{34,36,39,43,44} Additionally, because of the biocompatible nature, ELPs are used in many medicinal applications, such as tissue scaffolding,⁴³ cancer therapy,⁴⁵ and protein purification.⁴⁶

ELPs, similar to many known smart polymers,^{1–7} exhibit rich and tunable phase diagrams in water^{36,38,47} and in aqueous mixtures.⁴⁴ Furthermore, because of the hydrogen bonding nature of the microscopic interaction, these polymers are often water-soluble and, therefore, confers an expanded configuration of a chain for $T < T_l$, with T_l being the lower critical solution temperature (LCST). When $T > T_l$, a certain number of bound water molecules are released from the polymer solvation volume destabilizing an expanded polymer conformation.^{9,10}

There has been considerable interest in studying polypeptides, ELPs, and smart polymers using experiments,^{21–26,47–51} theory,^{2,24,52} and computer simulations.^{27–29,38,41} While most studies of ELPs focus on their behavior in water,^{34,38,46,47} a recent study has also investigated the phase behavior of ELPs in aqueous ethanol mixtures.⁴⁴ The latter study has shown that starting from an expanded conformation of ELPs (for $T < T_l$) in water, addition of ethanol molecules first collapses the chain. When ethanol concentration increases above a critical value, the chain again opens up. This coil-to-globule-to-coil transition in (miscible) binary mixtures is known as co-non-solvency, a name originally coined for the study of polystyrene in cyclohexane–dimethylformamide (DMF) mixtures⁵³ and later popularized for poly(*N*-isopropyl acrylamide) (PNIPAM) in aqueous alcohol mixtures.^{21,22} The molecular origin of this phenomenon has attracted intense debate in the literature. Here, various mechanisms have been proposed to be the driving force for the phenomenon of co-non-solvency, namely the cooperativity effect,^{24,54,55} solvent–cosolvent interactions,^{52,56} preferential cosolvent–monomer interaction,^{2,20,29,57,58} and the kosmotropic effect.⁵⁹

Even when the coil-to-globule transitions of polymers in aqueous alcohol mixtures are prevalent cases, they also show interesting conformational behavior in water–urea mixtures. For example, PNIPAM also collapses under the influence of urea.²⁵ The origin of the urea induced collapse of PNIPAM was shown to be due to the urea induced bridging of two NIPAM monomers topologically far along a polymer backbone.^{25,60}

In this work, we study and compare conformational behaviors of ELPs in water–ethanol and water–urea mixtures using molecular dynamics simulations of all-atom and complementary explicit solvent generic models. Going beyond simulation works dealing with implicit solvent generic model of ELPs under aqueous environments,⁶¹ our generic solvent

models are derived by mapping solvation free energies obtained from the all-atom simulations onto the generic explicit solvent model parameters for ELPs in binary solvents. We derive segment-based model parameters for four different amino acids relevant for ELPs, namely glycine (G), alanine (A), proline (P), and valine (V) in aqueous urea and aqueous ethanol mixtures. The chemical structures for the trimers of these amino acids are shown in Figure 1. The model parameters are tested to reasonably reproduce the phase behavior of two ELP sequences consisting of (VPGGG) and (VPGVG). Note that here we do not attempt to address the secondary structures of polypeptides and/or copolymer of peptides.⁶² Moreover, because ELPs can be classified as intrinsically disordered proteins,³⁹ their conformations can be described within the standard framework of polymer science,^{8,10} which is the motivation behind this study.

The remainder of the article is organized as follows: the details of the all-atom simulations and the generic model parameterization are presented in Section 2. The conformations of generic polypeptides and ELPs in binary solution are shown in Section 3. Finally, we draw our conclusions in Section 4.

2. METHOD AND MODEL

The generic model parameters are derived from solvation free-energy data obtained from all-atom simulations. We also emphasize here that the generic model parameters for the ELPs are obtained at the segment level, i.e., the parameters for different amino acids are obtained separately and then these are used to simulate different ELP sequences.^{19,41,63} It should be noted that this approximation is generally valid for neutral monomers, which applies to the amino acids V, P, G, and A. For charged monomers, it is necessary to refine the calculation of the solvation structure and the relative solvent–cosolvent coordination. Furthermore, V, P, G, and A are all very similar, and only the size of side carbon groups dictates their relative hydrophobicity; see Figure 1. This similarity eliminates cross-correlation between different monomer units, thus validating our segment-based approach.^{19,41,63} Therefore, we start by describing the all-atom model used in this study.

2.1. All-Atom Simulations. All-atom simulations are performed using the GROMACS molecular dynamics package.⁶⁴ These simulations are performed in the isobaric ensemble (*NPT*), where *N* is the number of particles, *P* is the isotropic pressure, and *T* is the temperature. $T = 300$ K is set using a velocity rescaling thermostat⁶⁵ with a coupling constant of 0.1 ps. Pressure is kept at 1 bar using a Parrinello–Rahman barostat⁶⁶ with a coupling constant of 2 ps. Electrostatics are treated with the particle mesh Ewald (PME) method.⁶⁷ The interaction cutoff for the nonbonded interactions is chosen as 1.0 nm and the equations of motion

are integrated using the leap-frog integrator with a time step of $\delta t = 2$ fs. All bonds were constrained with the LINCS.⁶⁸

We investigate four trimers, namely triglycine, trialanine, tripoline, and trivaline, in aqueous urea and aqueous ethanol mixtures (see peptide structures in Figure 1). Specific peptides are chosen because they constitute the monomeric building blocks of ELPs. Furthermore, we have only chosen trimers because the center monomer of a trimer gives a reasonable estimate of the solvation structure and relative solvent–cosolvent coordination, while not having to deal with conformation changes upon change in relative (co-)solvent compositions.⁴¹

For the trimers, we have used the GROMOS43a1 force field.⁶⁹ For water, we use the extended simple point charge (SPC/E)⁷⁰ model and the Kirkwood–Buff (KB) derived force field of urea.⁷¹ Note that the urea force field was parameterized on a GROMOS based model. Therefore, for consistency, we have also used GROMOS43a1 for trimers. We consider five different urea mole fractions, x_u : 0.0382, 0.0809, 0.1292, 0.1844, and 0.2495. The total number of water and urea molecules are taken exactly the same as in ref 72 that ensures solvent equilibrium within the simulation domain, i.e., system sizes are large enough to neglect finite-size effects.

Ethanol force field parameters are taken from ref 73. The ethanol mole fractions x_e are varied from pure water $x_e = 0.0$ to 0.25. For aqueous ethanol solutions, we have taken a total number of 616 ethanol and 15 528 water molecules at an ethanol molar concentration $x_e = 0.0382$, 1232 ethanol and 14 000 water at $x_e = 0.0809$, 1848 ethanol and 12 456 water at $x_e = 0.1292$, 2464 ethanol and 10 896 water at $x_e = 0.1844$, and 3080 ethanol and 9263 water at $x_e = 0.2495$.

2.2. Generic Simulations. Beyond the generic polymer model, we will describe the parameterization of the bulk binary solution, polymer–solvent interactions, and the model peptides in binary solutions. Note that while we will describe the polymer model and polymer–solvent (polymer–water) interactions in this section, polymer–cosolvent (polymer–urea and polymer–ethanol) parameterization will be described whenever it is discussed in this article.

2.2.1. Polymer Model. To describe a polymer, we have used the well-known bead-spring model.⁷⁴ In this model, monomers of a generic chain consist of Lennard-Jones (LJ) spheres. Bond connectivity between adjacent monomers is introduced by a finite extensible nonlinear elastic (FENE) potential. A bead-spring chain is solvated in mixtures of model water (solvent) and model ethanol or urea (cosolvent) molecules, also modeled as LJ spheres. The data are described in units of LJ diameter σ , LJ energy ϵ , and mass m of a monomer. This gives a time unit of $\tau = \sigma\sqrt{m/\epsilon}$ and pressure ϵ/σ^3 . We map one amino acid onto one generic monomer. Given that all four peptides investigated in this study have very similar sizes, this mapping scheme is reasonable. Furthermore, if we look at the monomer units (see Figure 1), they have typical sizes between 0.5 and 0.6 nm. This gives a length scale mapping of about $1\sigma \approx 0.5$ nm.

Generic simulations are performed using the ESPResSo++⁷⁵ and LAMMPS⁷⁶ molecular dynamics packages. Equations of motion are integrated with a velocity Verlet algorithm with a time step $\delta t = 0.005\tau$. The damping coefficient Γ of the Langevin thermostat is taken as $1.0\tau^{-1}$ to control the temperature at $T = 1.0\epsilon/k_B$.

Nonbonded monomers also interact with an attractive 6–12 LJ potential with a cutoff $r_c = 2.5\sigma$. Details of monomer–

monomer interactions will be described in the appropriate section in this article. We have chosen a chain length $N_1 = 50$ solvated in a solvent box consisting of 5×10^4 particles.

2.2.2. Bulk Solution. In this study, we have used very simple spherically symmetric models for binary mixtures without any specific chemical details. Moreover, these model parameters give correct miscibilities known from the all-atom data of aqueous ethanol⁵⁷ and aqueous urea.⁷²

2.2.2.1. Water–Ethanol Mixtures. To model the bulk solution, we consider that the size of water molecules (~ 0.28 nm) is typically half the size of the peptides, and the size of ethanol molecules (~ 0.50 nm) is about 1.8 times the size of water molecules. Therefore, we choose the sizes of water and ethanol in our generic model to be $\sigma_w = 0.5\sigma$ and $\sigma_e = 0.9\sigma$, respectively. For $x_e = 0$, we choose number density $\rho_w = 5.5\sigma^{-3}$, while the SPC/E water has $\rho_w = 32$ nm⁻³. The specific choice of ρ_w is motivated by the fact that if we choose 32 nm⁻³ = $5.5\sigma^{-3}$, this will lead to $1\sigma \approx 0.55$ nm, which is consistent with the length scale mapping described above. The generic water and ethanol molecules interact with each other via the repulsive LJ interactions with $\epsilon_{ij} = 1\epsilon$, $\sigma_{ij} = (\sigma_i + \sigma_j)/2$, and a cutoff $2^{1/6}\sigma_{ij}$. With these parameters and for $x_e = 0$, the typical pressure of the generic model is about $32\epsilon/\sigma^3$. We have adjusted total ρ with x_e such that the pressure is kept constant. In Figure 2, we show a comparative plot of the normalized

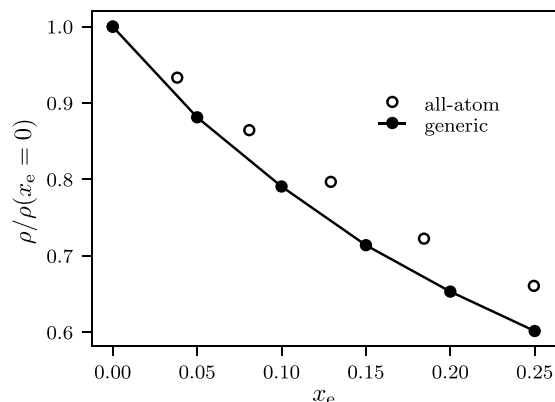


Figure 2. Normalized total number density $\rho/\rho(x_e = 0)$ as a function of ethanol mole fraction x_e . Data are shown for all-atom and generic simulations. The line is drawn to guide the eye.

density $\rho/\rho(x_e = 0)$ as a function of x_e between all-atom and generic simulations. ρ is consistent in both models as a function of x_e .

2.2.2.2. Water–Urea Mixtures. Similar to the parameterization of water–ethanol mixtures, we have also parameterized aqueous urea mixtures. For this purpose, we consider the size of urea molecules (~ 0.42 nm) to be about 1.5 times the size of water molecules. Therefore, we choose $\sigma_w = 0.5\sigma$ and $\sigma_u = 0.75\sigma$. The generic water and urea molecules interact with each other via the repulsive LJ interactions with $\epsilon_{ij} = 1\epsilon$, $\sigma_{ij} = (\sigma_i + \sigma_j)/2$, and a cutoff $2^{1/6}\sigma_{ij}$. We have adjusted the total ρ with x_u such that the pressure is kept constant at $32\epsilon/\sigma^3$. In Figure 3, we show normalized density $\rho/\rho(x_u = 0)$ as a function of x_u comparing all-atom and generic simulations.

3. RESULTS AND DISCUSSION

3.1. Elastin-Like Polypeptides in Aqueous Ethanol.

Before describing the conformation of ELPs in aqueous

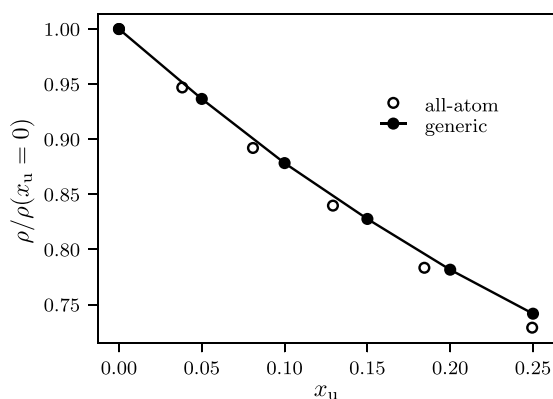


Figure 3. Same as Figure 2, however, for aqueous urea mixtures.

ethanol mixtures, we will first start our discussion by describing the polymer–(co-)solvent interaction.

3.1.1. Good Solvent Case of Peptide–Solvent Interactions. One of the most important factors in modeling ELPs in solution is to properly capture the relative affinities of different peptides in pure water (solvent). In this context, to investigate the coil-to-globule transition of ELPs in aqueous ethanol mixtures, we consider that the ELP chain is under a good solvent condition in pure water, i.e., when $T < T_1$. To obtain model parameters that reasonably satisfy the good solvent condition of ELPs in pure water, we have estimated the possible number of hydrogen bonds (H-bonds) between an amino acid and water molecules N_w from the all-atom simulations. H-bonds are calculated using the standard GROMACS subroutine, where a H-bond exists when the donor–acceptor distance is ≤ 0.35 nm and the acceptor–donor–hydrogen angle is $\leq 30^\circ$. The data are summarized in column 3 of Table 1. It can be appreciated that

$$N_w^{\text{proline}} < N_w^{\text{valine}} \approx N_w^{\text{alanine}} < N_w^{\text{glycine}}$$

Table 1. Number of Hydrogen Bonds between the Center Monomer of the Trimers with Water N_w and with Ethanol N_e ^a

x_e		0.0000	0.0382	0.0809	0.1292	0.1844	0.2495
P	N_w	0.806	0.519	0.346	0.200	0.134	0.107
	N_e		0.109	0.163	0.196	0.195	0.199
V	N_w	1.617	0.987	0.601	0.449	0.311	0.247
	N_e		0.501	0.706	0.881	0.872	0.915
A	N_w	1.660	1.097	0.756	0.524	0.369	0.257
	N_e		0.534	0.748	0.870	0.971	1.060
G	N_w	2.094	1.244	0.920	0.617	0.463	0.341
	N_e		0.419	0.655	0.865	0.946	1.029

^aData are shown for different ethanol mole fractions x_e .

To model the above described relative affinities, we have used LJ interaction parameters described in Table 2. These specific parameter choices ensure that a chain consisting of model amino acids remains expanded in pure solvent with attractive affinity with the solvent (water) molecules via hydrogen bonding, as is known from the ELP chain conformation below its T_1 .⁴⁴ Furthermore, it should be noted that this parameter space is not restricted and similar solvation conditions can also be achieved with different sets of parameters as long as relative monomer–monomer and monomer–solvent interactions are considered consistently.

3.1.2. Peptide–Ethanol Interactions. For the parameterization of the model to study ELPs in aqueous ethanol mixtures, we map the solvation free energies G_p obtained from the all-atom simulations of amino acids onto the generic model.^{2,6} To obtain G_p , we have used the Kirkwood–Buff theory of solutions,⁷⁷ which connects the fluctuation in the grand canonical ensemble μVT with the pairwise solution structure of complex fluids using the “so-called” Kirkwood–Buff integral (KBI)

$$G_{ij}^{\mu VT} = V \left[\frac{\langle N_i N_j \rangle - \langle N_i \rangle \langle N_j \rangle}{\langle N_i \rangle \langle N_j \rangle} - \frac{\delta_{ij}}{\langle N_i \rangle} \right] \\ = 4\pi \int_0^\infty [g_{ij}^{\mu VT}(r) - 1] r^2 dr \quad (1)$$

Here $G_{ij}^{\mu VT}$ and $g_{ij}^{\mu VT}(r)$ are the KBI and the radial distribution functions between i and j solution components in the μVT ensemble, respectively. μ is the chemical potential. $\langle \cdot \rangle$ gives the ensemble average, δ_{ij} is the Kronecker delta, and N_i is the number of particles of type i . In the thermodynamic limit, $G_{ij}^{\mu VT} \approx G_{ij}^{NVT/NPT}$. Here, however, we obtain G_{ij} from $4\pi \int_0^\infty [g_{ij}(r) - 1] r^2 dr$ with $r_0 = 1.5$ nm. Note that within the finite simulation domains, this is a safe approximation given that the typical correlation length in the aqueous systems is within the range 1.5–2.0 nm.²⁸ Furthermore, all system sizes are chosen to be the same as our earlier works that ensure well converged G_{ij} .^{28,57,72} There are also more accurate methods to obtain G_{ij} directly from fluctuations,^{78,79} here, we take the rather simple route of estimating G_{ij} from the convergence of KBIs.

G_{ij} can be used to derive the solvation free energy. When peptides (p) under infinite dilution are dissolved in a mixture of water (w) and ethanol (e), the shift in the solvation free energy ΔG_p can be calculated using

$$\lim_{\rho_p \rightarrow 0} \left(\frac{\partial \Delta G_p}{\partial x_e} \right)_{p,T} = \frac{RT(\rho_w + \rho_e)^2}{\eta} (G_{pw} - G_{pe}) \quad (2)$$

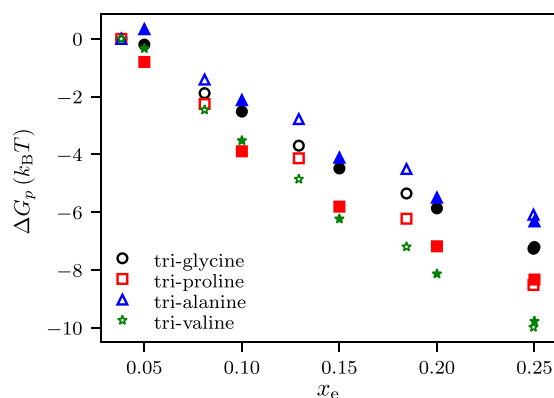
Here ρ_i is the number density and $\eta = \rho_w + \rho_e + \rho_w \rho_e (G_{ww} + G_{ee} - 2G_{we})$. Additionally, the preferential solvation parameter ($G_{ww} + G_{ee} - 2G_{we}$) gives the direct measure of the miscibility in bulk (binary) solution. Here, we find $(G_{ww} + G_{ee}) \approx 2G_{we}$ for both all-atom and generic simulations over a full range of x_e , indicating almost perfect miscibility as shown earlier.^{28,57}

In Figure 4, we show ΔG_p with changing x_e obtained from all-atom simulations (see empty symbols). We tune monomer–cosolvent (peptide–ethanol) interactions in our generic model to reproduce this shift in ΔG_p , as seen from the open symbols in Figure 4. The details of the model parameters are described in Table 2. Furthermore, as demonstrated in Table 1, we find $N_e^{\text{proline}} < N_e^{\text{valine}} < N_e^{\text{alanine}} \approx N_e^{\text{glycine}}$. Therefore, we incorporate the above conditions in our generic model via $\epsilon_w^{\text{proline}} < \epsilon_w^{\text{valine}} < \epsilon_w^{\text{alanine}} < \epsilon_w^{\text{glycine}}$ and $\epsilon_e^{\text{proline}} < \epsilon_e^{\text{valine}} < \epsilon_e^{\text{alanine}} < \epsilon_e^{\text{glycine}}$; see Table 2. Figure 4 also shows that ethanol has a preferential interaction with all amino acids, which is about 6–8 $k_B T$ more than the peptide–water interactions.

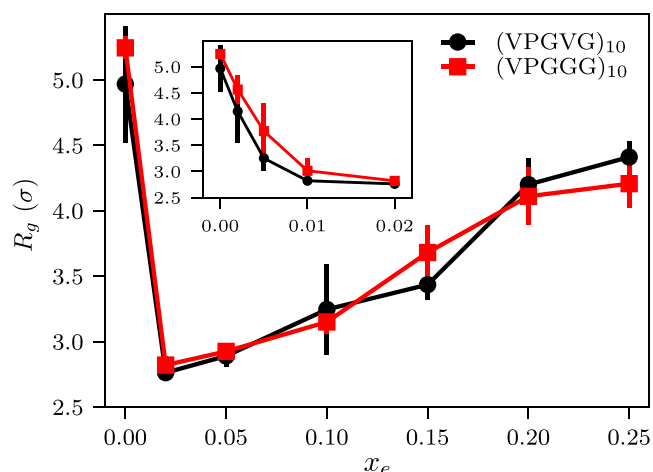
3.1.3. Conformation of Elastin-Like Polypeptides in Aqueous Ethanol. Using the generic model of peptides in aqueous ethanol described above, we will now investigate ELP conformations in aqueous ethanol mixtures. In this context, ELPs are one of the most intriguing classes of polymers that are genetically engineered having the properties of polymer random coil and at the same time are biocompatible because of

Table 2. Table Listing the Lennard-Jones (LJ) Length σ_{ij} and Energy ϵ_{ij} Parameters for All Pairs of Particles in the Generic Model of Valine (V), Proline (P), Alanine (A), and Glycine (G) in Water–Ethanol Mixtures

	water		ethanol		G		P		A		V	
	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}
water	0.500	1.000	0.700	1.000	0.750	0.680	0.750	0.480	0.750	0.670	0.750	0.500
ethanol	0.700	1.000	0.900	1.000	0.950	2.150	0.950	1.900	0.950	2.110	0.950	2.100
G	0.750	0.680	0.950	2.150	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500
P	0.750	0.480	0.950	1.900	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500
A	0.750	0.670	0.950	2.110	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500
V	0.750	0.500	0.950	2.100	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500

**Figure 4.** Shift in solvation free energy ΔG_p per monomer as a function of ethanol mole fraction x_e . Data are shown for four different trimers, namely triglycine, triproline, trialanine, and trivaline. The all-atom data are shown by empty symbols and the filled symbols correspond to the generic model. ΔG_p is calculated with respect to the center monomer of a trimer.

their amino acid-based monomeric building blocks. Here, ELPs usually have the sequence VPG-X-G, where X can be any amino acid except proline.⁴⁷ Because of the dominant H-bond nature of the interaction between amino acid and water molecules, ELPs show LCST behavior. Here, T_1 can be tuned by varying ELP sequences. For example, $T_1 \approx 300$ – 305 K for X = valine,⁸⁰ while $T_1 \approx 305$ – 310 K for X = glycine (i.e., more hydrophilic residue).^{44,80} This is identical to the typical LCST based copolymers, where T_1 can be tuned by changing hydrophobic or hydrophilic units along the backbone.^{19,40,41,63} Therefore, for this study, we investigate two sequences, namely -(VPGVG)- and -(VPGGG)-, for $T < T_1$. Note that both these systems remain expanded at around $T \approx 300$ K,^{44,80} where our generic models are parameterized. Using the default parameters (see Table 2), ELP conformations are studied with changing ethanol concentrations. In Figure 5, we show R_g for two sequences as a function of x_e . It can be seen that starting from an expanded chain in pure water ($x_e = 0$), increasing x_e first collapses a chain between $0.05 < x_e < 0.15$, and upon further increase of $x_e \geq 0.15$, an ELP chain reopens. This coil-to-globule-to-coil transition, often referred to as co-non-solvency,^{21,22} is a well-known phenomenon of standard smart polymers.^{26,81–84} Moreover, a recent experiment has also shown that ELPs can exhibit co-non-solvency in aqueous ethanol mixtures. In this context, our results are in good agreement with the experimental data.⁴⁴ Furthermore, not only that the conformational behavior observed in simulations is consistent with experiments, but also the window of collapse is in reasonable agreement with the experimental measurement at 300 K, i.e., $0.05 < x_e < 0.14$.⁴⁴

**Figure 5.** Gyration radius R_g of two ELP sequences, namely -(VPGVG)- and -(VPGGG)-, as a function of ethanol mole fraction x_e . Data are shown from the simulations using the parameters presented in Table 2. Inset shows the variation of R_g within the interval $0.00 \leq x_e \leq 0.02$.

While the microscopic origin of the co-non-solvency phenomenon is a matter of intense debate, it has been previously shown that the preferential binding of the better solvents (in this case ethanol) with the monomers drives the polymer collapse.^{2,20,29,58} When a small amount of ethanol is added into the aqueous solution of ELPs, these molecules preferentially bind to more than one amino acids to reduce the binding free energy. This leads to a typical case where a certain number of ethanol molecules form sticky contacts between different amino acids, thus initiating ELP collapse. Furthermore, it was also discussed that this collapse cannot be explained within the standard Flory–Huggins like mean-field picture, where the solvent–monomer and cosolvent–monomer interactions are dominant in comparison to the bulk solution χ parameter.⁸⁵ This also justifies our choice of spherically symmetric particles representing bulk solution components, which only requires $\chi \approx 0$ as known from the most common solvent mixtures where co-non-solvency is observed.²¹ Additionally, the interaction between pure solvent and pure cosolvent with the monomer should be $\sim 4 k_B T$ to observe co-non-solvency.²⁸ When this contrast reduces to $\leq 2 k_B T$, no co-non-solvency is observed.⁸⁶ In this context, we find that all four amino acids have very strong preferential binding (6 – $8 k_B T$) with ethanol in comparison to water (see Figure 4). Therefore, it is expected that an ELP shows the standard co-non-solvency in aqueous ethanol. We would like to mention that if the residue X is replaced with A, a -(VPGAG)- sequence will also show similar conformational behavior as shown in

Figure 5 because A has a very similar contrast of ΔG_p in aqueous ethanol as G or V (see Figure 4).

To further investigate the ELP collapse, we have also calculated the single-chain form factor $S(q)$ in Figure 6. While

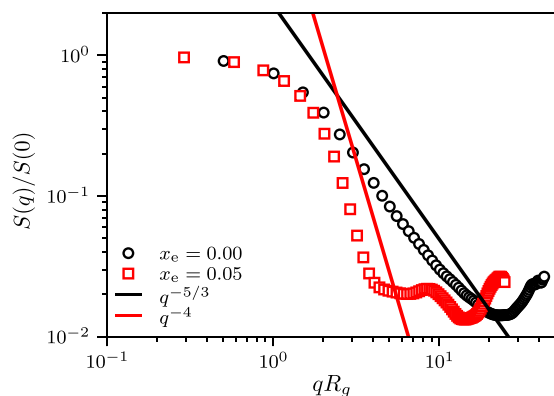


Figure 6. Normalized single-chain form factor $S(q)/S(0)$ of a -(VPGVG)- sequence for $x_e = 0.0$ (black) and $x_e = 0.05$ (red). Lines are power-law fits with scaling exponents $q^{-5/3}$ and q^{-4} .

the data for the pure solvent (i.e., $x_e = 0.0$) show $q^{-5/3}$ scaling as expected for a good solvent chain, data for $x_e = 0.05$ show q^{-4} behavior until $qR_g \sim 4.0$ and then deviates for $qR_g > 4.0$. It is noticeable that the ELP for $x_e = 0.05$ does not show a perfect sphere scattering as expected from $S(q)$ of a collapsed polymer globule.^{8–10} In this context, it is worthwhile to mention that even when a polymer collapses in a mixture of two good solvents, it is not a standard poor solvent collapse dictated by depletion interactions.⁶ Instead, solvent quality becomes better and better with increasing x_e , as evident from the ever-decreasing variation of ΔG_p with x_e (see Figure 4). This implies that even though a polymer remains collapsed under the influence of binary good solvents, it consists of good solvent blobs. Ideally speaking, for long chains, a cross-over from q^{-4} to $q^{-5/3}$ gives the typical blob size.⁸⁷ Here, however, our chain length is rather small and, therefore, we do not observe any cross-over scaling.

Having discussed the conformation of ELPs in aqueous ethanol mixtures, we now want to investigate a broader implication of ELP conformations in other binary mixtures, such as aqueous urea mixtures. In this context, it should be mentioned that one of the most studied polymers that show co-non-solvency in aqueous alcohol mixtures is PNIPAM. Here, PNIPAM not only collapses in aqueous alcohol but also shows an interesting coil-to-globule transition in aqueous urea solution.^{25,60} In these studies, it was shown that the strong H-bonding between the hydrophilic group of NIPAM monomers and urea drives the collapse of a chain. Here, the mechanism of

polymer collapse was shown to be driven by urea molecules forming sticky contacts between distant monomers far along the polymer backbone. Therefore, it is worth investigating if urea can also confer the collapse of an ELP, which is the motivation behind the next section.

3.2. Elastin-Like Polypeptides in Aqueous Urea. Urea is a well-known denaturant for proteins or peptides.⁸⁸ Here, however, we will investigate the effect of urea on the possible folding transition of good solvent ELPs in pure water. To mimic the good solvent case, we have taken the same monomer–solvent (amino acid–water) parameters as presented in Table 2. Consistently, the generic monomer–cosolvent (amino acid–urea) interaction parameters are obtained by mapping ΔG_p onto the all-atom data, using the same protocol presented in Section 3.1.2. In Table 3, we present the full list of generic monomer–(co-)solvent interactions. This parameter set also ensures that the shift in ΔG_p is well reproduced in the generic model as known from the all-atom simulations (see Figure 7). Figure 7 also shows

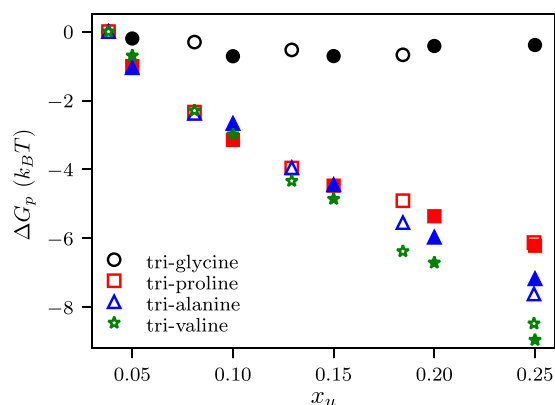


Figure 7. Shift in solvation free energy ΔG_p as a function of urea mole fraction x_u . Data are shown for four different trimers, namely triglycine, triproline, trialanine, and trivaline. The all-atom data are shown by empty symbols and the filled symbols correspond to the generic model. The all-atom data for triglycine are taken from ref 72.

that the relative preferability of urea with glycine is almost negligible as indicated by $|G_p(x_u = 0) - G_p(x_u = 0.2495)| < k_B T$. The all-atom data for glycine are taken from our earlier work,⁷² which is consistent with other experimental⁸⁹ and simulation data.⁹⁰ For the other three amino acids, the shift is between 6 and 8 $k_B T$, thus showing that the urea interaction is highly preferred with these amino acids in comparison to water. It should also be noted that urea molecules not only interact with an amino acid with preferential H-bonds but also interact with the hydrophobic residues of different amino acids (see red parts in Figure 1a–c) via van der Waals interactions.⁹¹

Table 3. Same as Table 2, However, for Peptides in Water and Urea Mixtures

	water		urea		G		P		A		V	
	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}	σ_{ij}	ϵ_{ij}
water	0.500	1.000	0.625	1.000	0.750	0.680	0.750	0.480	0.750	0.670	0.750	0.500
urea	0.625	1.000	0.750	1.000	0.875	1.100	0.875	1.350	0.875	1.800	0.875	1.760
G	0.750	0.680	0.875	1.100	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500
P	0.750	0.480	0.875	1.350	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500
A	0.750	0.670	0.875	1.800	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500
V	0.750	0.500	0.875	1.760	1.000	0.500	1.000	0.500	1.000	0.500	1.000	0.500

In Figure 8, we show the conformational behavior of an ELP sequence of -(VPGVG)- in aqueous urea mixtures. For

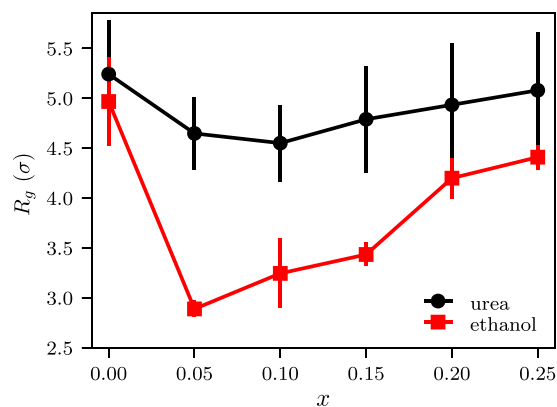


Figure 8. Gyration radius R_g of -(VPGVG)- sequence with 10 repeat units in aqueous urea (black) and aqueous ethanol (red) mixtures as a function of their molar concentrations x .

comparison, we have also included the data for aqueous ethanol mixtures; see Figure 5. It can be appreciated by the black data set in Figure 8 that -(VPGVG)- shows weak swelling–collapse–swelling behavior in aqueous urea in comparison to the aqueous ethanol mixtures (see red data set in Figure 8). It is still important to mention that valine and proline residues have $6\text{--}8 k_B T$ interaction contrast and thus a chain should collapse.⁸⁷ Here, however, the effect is diluted because of the dominant effect of the glycine residues that are in majority and almost have no preferentiability between water and urea (see Figure 7). In this context, it is important to mention that it requires a certain concentration of hydrophobic residues along the backbone to initiate the polymer collapse, which is more than 50% in most cases.^{41,63} Therefore, while -(VPGVG)- still shows weak collapse, conformation of -(VPGGG)- shows no noticeable change with x_u . While validating this scenario would require detailed experimental work, this already highlights that if glycine can have a strong preference with a cosolvent, one can observe a standard coil-to-globule transition as shown earlier in Section 3.2.

We would also like to clarify why glycine has a higher interaction strength with ethanol than with urea. Here, we observe that ethanol molecules bind with a glycine via a preferential H-bond. This leads to a typical case where the hydrophobic $-\text{CH}_2\text{CH}_3$ part of ethanol molecules get exposed to the bulk water forming large hydrophobic patches along the ELP backbone. These hydrophobic patches can then confer a collapsed conformation through the hydrophobic interactions. However, in the case of urea and water, there are only dominant H-bonds, leading to the complete mixing of all solution species.

4. CONCLUSIONS

We have derived explicit solvent generic models to study the conformation of peptides and ELPs in aqueous mixtures. The parameterization procedure is done by mapping the solvation free energies obtained from the all-atom simulations onto the generic model interaction parameters with changing cosolvent concentration. The mapping is performed at the monomer level for different peptides, namely proline (P), valine (V), glycine (G), and alanine (A), where the first three are typical building blocks of ELPs. These models are used to study the

conformational behavior of ELPs in aqueous ethanol and aqueous urea mixtures. Note that by conformation, we only mean the size of an ELP without attempting to describe the protein secondary structures. We find that ELPs show a fascinating co-non-solvency behavior in aqueous ethanol, as observed in recent experimental work.⁴⁴ We rationalize this result in terms of the preferential ethanol interactions with all peptide residues of ELPs. By contrast, the degree of collapse of ELPs in aqueous urea is rather weak. This distinct behavior can be attributed to the difference in glycine interactions in aqueous ethanol in comparison to aqueous urea. While some of our results are in direct agreement with the existing experimental data, we also make predictions. Indeed, we present the first set of simulations on ELPs in binary mixtures giving direct evidence toward a more robust and tunable phase behavior of biocompatible systems. Therefore, these results may provide new directions to the advanced materials' design.

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Notes

The authors declare no competing financial interest.

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