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1 Therapeutic potential of co-enzyme Q10 in retinal diseases

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30 **Abstract**

31 Coenzyme Q10 (CoQ10) plays a critical role in mitochondrial oxidative phosphorylation by
32 serving as an electron carrier in the respiratory electron transport chain. CoQ10 also functions
33 as a lipid-soluble antioxidant by protecting lipids, proteins and DNA damaged by oxidative
34 stress. CoQ10 deficiency has been associated with a number of human diseases including
35 mitochondrial diseases, neurodegenerative disorders, cardiovascular diseases, diabetes,
36 cancer, and with the ageing process. In many of these conditions CoQ10 supplementation
37 therapy has been effective in slowing or reversing pathological changes. Oxidative stress is a
38 major contributory factor in the process of retinal degeneration. In this brief review, we
39 summarize the functions of CoQ10 and highlight its use in the treatment of age-related
40 macular degeneration and glaucoma. In light of these data we propose that CoQ10 could have
41 therapeutic potential for other retinal diseases.

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43 **Keywords** co-enzyme Q10, oxidative stress, retina, age related macular degeneration,
44 glaucoma, retinitis pigmentosa, diabetic retinopathy, protection

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60 **1. Introduction**

61 Coenzyme Q10 (CoQ10), first identified by Crane et al, is a 1,4-benzoquinone-containing
62 molecule with a hydrophobic tail harbouring 10 isoprenyl units (1). CoQ10 is ubiquitously
63 distributed in various tissues and blood and presents in all cell membranes (2, 3). It is
64 synthesized in the mitochondrial matrix and at least 12 genes are required for its biosynthesis;
65 mutations in some of these genes have been reported to cause CoQ10 deficiencies (4). CoQ10
66 exists in more than one state within the body: oxidized (ubiquinone), partially reduced
67 (semiquinone radical) and reduced (ubiquinol) forms (Figure 1A); the ratio of oxidized and
68 reduced forms in various cellular membranes is dependent on the metabolic state of
69 individual cells. Within the inner mitochondrial membrane, the CoQ10 pool is found in two
70 main forms: approximately 30% is protein bound and principally participates in oxidative
71 phosphorylation; the remainder is not protein-bound and contributes to different functions,
72 the major one being as a lipophilic antioxidant (5). CoQ10 is required for cellular ATP
73 generation by shuttling electrons from complexes I and II to complex III in the mitochondrial
74 respiratory chain (Figure 1 B) (6). The oxidized form of CoQ10 is able to undergo two
75 electron reductions in a reaction involving complex I and complex II, resulting in the
76 formation of ubiquinol: subsequently, electrons are passed to complex III. Typically, tissues
77 that are heavily reliant on oxidative metabolism, such as the myocardium, present a high
78 concentration of CoQ10. It is the only naturally occurring endogenous lipid-soluble
79 antioxidant which, in its reduced and active form ubiquinol, may act as a direct free radical
80 scavenger, inhibiting the oxidation of lipids, proteins and DNA (6) or may act synergistically
81 with other antioxidants, such as vitamin E, regenerating its oxidised form, tocopheryl radical.
82 CoQ10 also demonstrates a regulatory role in the expression of genes involved in cell
83 signalling, metabolism and nutrition transport (7). Moreover, it has been shown to exert an
84 anti-inflammatory effect by reducing LPS-induced secretion of TNF- α , possibly via the
85 NFkB1-dependent pathway (8).

86 CoQ10 deficiency is mainly associated with encephalomyopathy, infantile multisystemic
87 disease, cerebellar ataxia, pure myopathy, and cardiofaciocutaneous syndrome. The causes of
88 CoQ10 deficiencies are primarily due to mutations in ubiquinone biosynthesis genes (*COQ2*,
89 *PDSS1* and 2, and *ADCK3*) or in genes indirectly related to CoQ10 biosynthesis (*APTX*,
90 *BRAF*, and *ETFDH*). However, the causes of CoQ10 deficiency still remain unknown in a
91 large number of patients (4). Lowered levels of CoQ10 have been reported in different
92 clinical conditions, including cardiovascular disease, diabetes, cancer, and neurodegenerative
93 disease. More generally, a subliminal deficit of CoQ10 might also be observed in

94 paraphysiological states such as ageing: synthesis in human is known to peak around the third
95 decade of life and subsequently decreases with age. Moreover, the use of commonly
96 prescribed drugs that interfere with the mevalonate pathway, such as statins, are known also
97 to impact cellular coenzyme Q10 level. Interestingly, intracellular content of CoQ10 is close
98 to the Km of the respiratory complexes, implying that even slight variations in the CoQ10
99 content translate into dramatic changes in the mitochondrial bioenergetics that is known to be
100 a major site of production of reactive oxygen species. Oral CoQ10 therapy has been applied
101 to different forms of CoQ10 deficiency, with resulting significant clinical improvements (4).
102 Oxidative stress plays an important role in the pathogenesis of vascular diseases, diabetes and
103 neurodegenerative disease. Due to its antioxidant properties, CoQ10 supplementation has
104 been beneficial in the treatment of the above diseases. Numerous studies have reported that
105 CoQ10 administration improved cardiovascular function (2,9,10). CoQ10 supplementation in
106 three separate clinical trials of dyslipidemic type 2 diabetic patients showed raised plasma
107 CoQ10 levels, improved endothelial function, and decreased blood pressure and glycosylated
108 haemoglobin (HbA1C) (11-13). CoQ10 has been used in different neurodegenerative diseases,
109 including Parkinson's disease, Huntington's disease, and Alzheimer's disease. CoQ10
110 supplementation seemed to slow progression of Parkinson's disease (9, 14).

111 CoQ10 is detectable in both choroid and retina, though levels are relative low when
112 compared to other oxygen-demanding tissues (15, 16). Similarly to other tissues, the level of
113 CoQ10 in the retina declines with age (15). There is increasing evidence that CoQ10 protects
114 retinal cells *in vitro* and *in vivo*, therefore the age-related CoQ10 decrease might exacerbate
115 the risk of retinal disease, while supplementation could have a preventative role. Here we
116 provide an overview of the therapeutic roles of CoQ10 in retinal diseases.

117

118 **2. Structure and function of mammalian retina**

119 The neural retina is a unique structure, consisting of three major cellular layers (outer
120 nuclear layer, ONL; inner nuclear layer, INL; ganglion cell layer, GCL), separated by
121 synaptic layers (Figure 2) (17). An outer monolayer, the retinal pigment epithelium (RPE),
122 underlies the retina and supports photoreceptor function. ONL contains the light-sensitive
123 photoreceptors, rods and cones. Rods are sensitive to dim light, whereas cones function in
124 bright light and colour vision. In the central retina of primates, there is a small cone-enriched
125 area, the 'macula', which is functionally specialised for high acuity vision. The central pit of
126 the macula is the fovea (Figure 2), which contains only cones and provides the sharpest
127 vision (18). In the retinae of most mammalian species, about 95% of photoreceptors are rods.

128 The adult human retina has about 91 million rods and 5 million cones (18). INL is composed
129 mainly of bipolar cells, although amacrine cells and horizontal cells are also localized in this
130 layer. Bipolar cells receive synaptic input from photoreceptors and are responsible for
131 transmitting the signals to ganglion cells directly or indirectly via amacrine cells. Horizontal
132 cells provide feedback to photoreceptor cells and possibly bipolar cells. Amacrine cells are
133 inhibitory neurons and interact with retinal ganglion cells via their dendritic arbors (17). The
134 ganglion cells have long axons, which form the optic nerve, and are responsible for the
135 transmission of signals from photoreceptors to brain.

136 The retina has the highest oxygen consumption rate (per gram tissue) in the body, which
137 results in the production of a large amount of reactive oxygen and nitrogen species (RONS)
138 that pose a risk for subsequent cellular damage (19, 20). The retina, particularly the macula,
139 is also subjected to high light exposure, making photosensitizing molecules, such as retinoids,
140 vulnerable to light damage. In addition, photoreceptor outer segments are extremely lipid-rich:
141 about 15% of wet weight content is lipid compared with 1% of wet weight in other types of
142 cells (21). The photoreceptor outer segments also have a high level of the very-long-chain
143 polyunsaturated fatty acid (PUFA), which is vulnerable to RONS and are easily oxidisable to
144 malondialdehyde (MDA) and 4-hydroxy-2-nonenal (HNE) (20). Cellular systems present
145 several defence lines against oxidative damage. However, oxidative imbalances occurring as
146 a result of the ageing process promote oxidative damage that might contribute to the
147 pathogenesis of retinal diseases.

148 Clinical data have demonstrated oxidative stress contributes the pathogenesis of retinal
149 diseases. Lower total antioxidant capacity has been reported in aqueous humor and sera from
150 patients with retinitis pigmentosa (RP) (22). In patients with diabetic retinopathy (DR), lipid
151 peroxidation in serum was significantly increased when compared to that of patients with
152 diabetes (but with no retinopathy) and there is positive correlation between lipid peroxidation
153 and disease severity (23, 24). Furthermore, patients with proliferative DR have a markedly
154 increased serum MDA level compared to that of non-proliferative DR patients (25). Recently
155 increased oxidative stress level and decreased antioxidant defence have been identified in the
156 sera from patients with primary open angle glaucoma, pseudoexfoliative glaucoma and
157 primary angle-closure glaucoma (26, 27). Due to the central role of oxidative stress in the
158 progression of these diseases, antioxidant therapies may play a role in counteracting retinal
159 degeneration. Actually antioxidant supplementation in patients with nonproliferative DR
160 demonstrated retardation of disease progress and maintenance of plasma antioxidant capacity
161 (28).

162 **3. Protection of retinal diseases by co-enzyme Q10**

163 **3.1 Age related macular degeneration**

164 Age-related macular degeneration (AMD) is the most common cause of blind registration in
165 the developed world (29). Early AMD is characterized by drusen formation and pigmentary
166 changes. Late AMD is characterized by geographic atrophy (dry AMD) and / or choroidal
167 neovascularisation (CNV, wet AMD). Wet AMD presents newly formed immature blood
168 vessels growing from the choroid through Bruch's membrane toward the outer retina. Wet
169 AMD accounts only for about 10-15% of cases, but for 80-90% of resultant blindness. Anti-
170 VEGF (vascular endothelial growth factor) therapies dramatically halt progression of CNV in
171 the majority of wet AMD patients but there is no effective treatment for AMD patients with
172 geographic atrophy. An important pathological feature of AMD is the accumulation of both
173 focal (drusen) and diffuse extracellular (basal) deposits in the macula, between the retinal
174 pigment epithelium (RPE) and the adjacent Bruch's membrane. These deposits lead to
175 dysfunction and subsequent death of RPE and associated photoreceptors (30). It is well
176 recognized that oxidative damage plays an important role in AMD and that antioxidant
177 supplementation can protect against the condition (31).

178 Blasi et al. measured CoQ10 in plasma and platelets of 19 patients with exudative AMD
179 and 19 age-matched controls (32). They found that most AMD patients had a lower level of
180 plasma CoQ10 than that of most controls, suggesting a link between CoQ10 level and AMD
181 (32). Fourteen early AMD patients treated with a mixture including polyunsaturated fatty
182 acids (1320 mg/day), acetyl-L-carnitine (500 mg/day), CoQ10 (30 mg/day), and vitamin E
183 (30 mg/day) showed slight improvement in visual functions after three months of treatment;
184 the improved visual functions remained relatively steady until 24 months. By contrast, the
185 visual functions of controls treated with vitamin E only (30 mg/day) slowly worsened (33).
186 The same research group continued to evaluate the treatment efficacy of a combination of
187 acetyl-L-carnitine, n-3 fatty acids and CoQ10 in early AMD patients for 12 months (34). 106
188 patients were randomly allocated to two groups: the treated group (51 patients) and the
189 placebo group (55 patients); four efficacy parameters including visual field mean defect
190 (VFMD), visual acuity, foveal sensitivity and fundus alteration were measured. The treated
191 group showed significant improvement in visual function, demonstrating a significant
192 difference in VFMD, visual acuity and foveal sensitivity when compared to that of the
193 placebo group. Only 2% of the treated group exhibited clinically related worsening in VFMD
194 while 17% of the control group showed further deterioration by the end of the trial (34).

195

196 **3.2 Glaucoma**

197 Glaucoma is a leading cause of irreversible blindness, affecting more than 70 million people
198 worldwide (35). It is characterized by the progressive degeneration of retinal ganglion cells.
199 Intraocular pressure (IOP) is higher in many glaucoma patients and regarded as an important
200 factor for initiating neuronal damage in these patients. Previous studies demonstrated that
201 elevated acute and chronic IOP induced oxidative stress in the retina (36-38), resulting in the
202 oxidative modification of proteins, lipids and DNA (39-41). Primary and secondary hypoxia
203 (the latter subsequent to elevated IOP) result in oxidative stress and glutamate excitotoxicity,
204 both of which contribute to ganglion cell dysfunction in glaucoma (42). Histological studies
205 on glaucomatous eyes from patients and different animal models demonstrated that ganglion
206 cells were degenerated through apoptosis (43-47). The death of ganglion cells is mainly
207 caused by oxidative damage via multiple pathogenic mechanisms (42,48). Antioxidants (n-3
208 PUFAs, α -Lipoic acid and mitochondrially-targeted SKQ1) treatment in glaucoma animal
209 models showed protection of retinal ganglion dysfunction (49-52).

210 CoQ10 has also been used to protect retinal ganglion cell function in the glaucomatous
211 condition. *In vitro* studies demonstrated that CoQ10 treatment increased survival of RGC-5
212 cells (a rat ganglion cell line) from apoptosis when exposed to H₂O₂, radiation, antimycin (the
213 complex III inhibitor) or serum starvation (53-55). Administration of CoQ10 in high
214 intraocular pressure-induced ischemia rat model prevented ganglion cell loss (56). The
215 protection of ganglion cell death by CoQ10 in ischemic retina was through ameliorating
216 oxidative stress, blocking apoptosis, preserving mitochondrial function and inhibiting
217 microglial activation (57). In untreated mouse ischemic retina, the level of superoxide
218 dismutase 2 (SOD2) and heme oxygenase 1 (HO-1) was significantly increased at 12h after
219 transient retinal ischemia when compared to non-ischemic control retina; however, CoQ10
220 treatment preserved SOD2 and HO-1 at levels similar to those of non-ischemic retina. The
221 level of apoptosis-associated protein Bax was significantly increased in ischemic retina, but
222 CoQ10 treatment markedly decreased Bax expression. In addition, the expression of glial
223 fibrillary acidic protein (GFAP, a marker for astroglial cells) and Iba-1 (a marker for
224 microglial cells) was significantly decreased in CoQ10-treated ischemic retina when
225 compared to that of non-treated ischemic retina, demonstrating the inhibition of astroglial and
226 microglial cell activation (57).

227 Glutamate excitotoxicity can cause ganglion cell death in glaucoma through the N—
228 methyl-D-aspartate (NMDA) receptor-activated influx of extracellular calcium into cells,
229 which regulates the activities of cell-death-associated enzymes (42,58). Significantly

230 increased retinal extracellular glutamate was detected in pressure-induced ischemic rat model;
231 intraocular administration of CoQ10 markedly minimized the increase (56). In an
232 intravitreally NMDA-injection-induced retinal damage mouse model, oral administration of
233 CoQ10 (10 mg/kg) for 14 days showed that CoQ10 exerted neuroprotective effects by
234 decreasing ganglion cell death significantly when compared to that of untreated ischemic
235 retina (53). When CoQ10 was administered as eye drops on mouse cornea, it reached the
236 choroid/ retina in a dose- and time-dependent manner (55). Moreover, in patients undergoing
237 vitrectomy CoQ10 administered by eye drops has been shown to penetrate the vitreous body,
238 where it could function on the retinas (59). In a retinal damage mouse model made by
239 intravitreal injection with kainite (a glutamate agonist), CoQ10 eye drop treatment
240 significantly reduced ganglion cell death by inhibiting caspase-dependent apoptosis (55). Lee
241 et al investigated the neuroprotective effects of CoQ10 in a glaucoma mouse model (DBA/2J)
242 by feeding the glaucomatous mice with CoQ10 for 6 months (60). The survival of ganglion
243 cells was markedly increased in the CoQ10 treated mouse retina when compared to that of
244 mouse fed with a control diet. Similar to the data from retinal ischemic mouse model (57), the
245 protection of ganglion cell death by CoQ10 also resulted from ameliorating glutamate
246 excitotoxicity, blocking oxidative stress, maintaining mitochondrial function and inhibiting
247 astroglial activation (60). Most recently, open-angle glaucoma (OAG) patients treated with
248 eye drops containing CoQ10 and vitamin E (in addition to a β -blocker monotherapy) for 12
249 months showed beneficial effects on function of the inner retina (assessed by pattern
250 electroretinogram) and enhanced visual cortical response (assessed by pattern visual-evoked
251 potential) (61).

252 Astrocytes are the major type of glial cell in the optic nerve head (ONH) and provide
253 support for axon function (61). During the progression of glaucoma, astrocytes become
254 activated and are involved in the ONH remodelling associated with the condition (62, 63).
255 Oxidative stress is known to reactivate ONH astrocytes and is implicated in the pathogenesis
256 of glaucoma (64, 65). When rat ONH astrocytes were treated with CoQ10 and H₂O₂, cell
257 viability and ATP in these cells were both significantly increased, while ROS production was
258 markedly reduced, when compared to that of cells treated with H₂O₂ alone. The GFAP,
259 SOD2 and HO-1 proteins in CoQ10 treated cells were significantly decreased compared to
260 those of H₂O₂-treated cells. CoQ10 treatment preserved mitochondrial morphology and
261 biogenesis by upregulating the expression of the mitofilin and PGC-1 α proteins, respectively
262 (66). These data suggest that CoQ10 can protect ONH astrocytes from oxidative stress mainly
263 through maintenance of mitochondrial function.

264 **4. Potential of CoQ10 for the treatment of retinitis pigmentosa and diabetic retinopathy**

265 As oxidative stress also plays a critical role in the pathogenesis of retinitis pigmentosa and
266 diabetic retinopathy, so CoQ10 has potential for the treatment of both diseases.

267 **4.1 Retinitis pigmentosa**

268 Retinitis pigmentosa (RP, MIM #268000) is a heterogeneous group of conditions involving
269 progressive degeneration of photoreceptor cells and affects 1/4000 individuals worldwide
270 (67). The early clinical feature of RP is night blindness, often starting in adolescence,
271 followed by progressive loss of peripheral vision and late loss of central vision. The
272 characteristically clinical feature is bone spicule pigment deposits presented in the retinal
273 fundus (Figure 3). RP may occur alone, as non-syndromic RP, without other clinical features,
274 or as syndromic RP with different clinical phenotypes. Most RP cases are presumed to result
275 from a mutation in one or more genes and may show autosomal dominant, recessive, X-
276 linked, or mitochondrial inheritance, although about one-half of all cases are sporadic.
277 Mutations in more than 62 genes have been reported to cause non-syndromic RP, including
278 23 genes associated with autosomal dominant RP, 36 genes associated with recessive RP, and
279 3 genes associated with X-linked RP (68).

280 Death of rod cells in RP occurs through both caspase-dependent and -independent
281 apoptosis, while death of cone cells occurs primarily through necrosis (69, 70). Oxidative
282 damage plays a critical role in the death of photoreceptors (69). Photoreceptors have one of
283 the highest rates of oxygen consumption in the body and this is particularly high in the
284 parafoveal region of primates, where rod density is highest (71). Our previous work showed
285 that severe oxidative stress was present in the retinas of four RP mouse models ($Pde6b^{rd1/rd1}$,
286 $Pde6b^{atrd1/atrd1}$, $Rho^{-/-}$ and $Prph2^{rds/rds}$) evidenced by significantly reduced retinal complex I
287 activities (14-29% of wildtype) at a stage when significant photoreceptor loss has not yet
288 occurred (72). In RP, oxidative damage is also a major contributing factor to cone death
289 subsequent to the death of rod cells. Further antioxidants have been shown to slow/reduce
290 cone cell death in different RP animal models (73). Upregulation of antioxidant defences by
291 over-expression of both superoxide dismutase 2 (SOD2) and catalase in photoreceptor
292 mitochondria also reduces cone cell death in RP mouse models (74).

293 It is desirable to develop new candidates with the potential of reducing reactive oxygen
294 species (ROS) production and/or upregulating antioxidant defences, which in turn can
295 potentially slow down retinal degeneration in RP.

296 **4.2 Diabetic retinopathy**

297 Diabetic retinopathy (DR) refers to the irreversible damage of retinal cells and structures as a
298 result of chronic diabetes. DR is a progressive disease that is influenced by the duration and
299 control of diabetes, and its development is believed to occur gradually with different degrees
300 of disease severity. DR is classified into five stages: no diabetic retinopathy, background
301 diabetic retinopathy, non-proliferative diabetic retinopathy (NPDR), proliferative diabetic
302 retinopathy (PDR) and diabetic macular edema (DME) (75, 76). The first stage, no diabetic
303 retinopathy (Figure 4A), is characterized by normal retinal histology with absence of any
304 abnormal neovascularization and microvascular abrasions. The second stage, background DR
305 (Figure 4B), is the earliest stage of DR and is associated with the presence of low grade of
306 microaneurysm, retinal hemorrhage and exudate. The third stage, non-proliferative diabetic
307 retinopathy (NPDR), itself includes three phases: mild NPDR (Figure 4C) which involves
308 microaneurysm; moderate NPDR (Figure 4D) which involves less severe microaneurysm,
309 intraretinal haemorrhage and microvascular occlusion; and severe NPDR (Figure 4E) which
310 is characterized by severe and increased rate of intraretinal haemorrhage, microvascular
311 abnormalities and venous beading. The fourth stage, proliferative diabetic retinopathy (PDR,
312 Figure 4F), is considered to be a severe phase and is defined by retinal ischemia and
313 increased rate of abnormal neovascularization in the retina, optic disc and iris with vitreous or
314 pre-retinal haemorrhage. The fifth stage, diabetic macular edema (DME, Figure 4G), is
315 associated with relatively increased retinal thickness at the centre of the macula, vascular
316 permeability and leakage, hard exudate, breakdown of blood-retina barrier (BRB) and retinal
317 detachment (75, 76).

318 Oxidative stress is a common characteristic of DR secondary to hyperglycemia.
319 Mitochondria are the principal source of energy production. Under normal conditions,
320 mitochondria provide energy through the electron transport chain (ETC) in which oxygen (O₂)
321 is utilized as the main electron donor and then reduced to ROS to maintain cellular functions;
322 any increase of ROS level is neutralized by a specialized antioxidant defence system (77).
323 Mitochondria are the main source of ROS production during diabetes, and studies have
324 shown that hyperglycemia induces mitochondrial ROS overproduction in response to
325 increased activation of the polyol pathway, AGEs, PKC pathway, hexosamine biosynthesis,
326 and poly (ADP-ribose) polymerase. Under physiological conditions excess ROS is
327 eliminated by specific antioxidant scavengers and balanced by mitochondria maintaining
328 redox (77). Several antioxidant scavengers such as catalase, superoxide dismutases (SODs)
329 and glutathione peroxidases (GPXs) have been reported to be involved in oxidative stress
330 during DR (78-81). Accumulated data from diabetic patients, diabetic animal models and

331 high glucose treated cells has revealed that these anti-oxidants may exhibit different gene
332 expression patterns and activity; the activity of catalase, SODs and GPXs were reported to be
333 low in diabetic patients, animal models and high glucose treated cells compared to normal
334 control (78-81). Thus, a therapeutic strategy to directly decrease ROS production and
335 enhance expression of these anti-oxidants will protect the retina from oxidative stress damage
336 during DR.

337

338 **5. Conclusion**

339 Oxidative stress causes damage to protein, lipid and DNA, which results in retinal cell
340 dysfunction and death. Mitochondria are the major source of oxidative stress. CoQ10
341 functions as an electron carrier in the mitochondrial respiratory chain and as an intracellular
342 antioxidant that offers therapeutic potential for neurodegenerative diseases. Furthermore, due
343 to its antioxidant properties CoQ10 has demonstrated a protective role in the neuroretina by
344 counteracting oxidative stress, inhibiting microglia cell activation and maintaining
345 mitochondrial function. In particular, the role of CoQ10 in modulating mitochondrial
346 permeability transition pore has been linked to its beneficial effects in preventing the
347 glutamate-induced cytotoxicity that may contribute to neural degeneration. CoQ10 topical
348 eye preparation has been shown to be an effective means of delivery to the vitreous cavity
349 and retina. However, until now only oxidised CoQ10 (ubiquinone) has been tested. The
350 recent availability of the stable formation of the reduced and active form of CoQ10
351 (ubiquinol) might represent a ground-breaking innovation in the field. In fact, cellular
352 metabolism is able to efficiently promote reduction of exogenously provided coenzyme Q10,
353 while the activity of reducing systems declines with age.

354 In conclusion, a significant body of evidence supports a role for CoQ10 in promoting eye
355 health through inhibiting ROS production and protecting neuroretinal cells from oxidative
356 damages (Figure 5), although further studies are required to evaluate potential beneficial
357 effects of ubiquinol eye-drop treatment for patients with retinal diseases, including AMD,
358 DR, RP and glaucoma, which are major causes of blindness in the world.

359

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364

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599 **Figure legends**

600 **Figure 1** Coenzyme Q10 is a redox existing in the cellular membranes and consisting of a
601 quinone ring and 10-isoprenoid-unit tail. There are three states of Coenzyme Q10: fully
602 oxidized form (ubiquinone), semiquinone (ubisemiquinone) and fully reduced form
603 (ubiquinol) (A). Coenzyme Q10 is soluble in phospholipid bilayer of the inner mitochondrial
604 membrane. It is an essential component of the mitochondrial respiratory chain (B).

605 Ubiquinone can adopt one or two electrons from inner mitochondrial membrane complex I
606 and II, transforming into semiquinone or ubiquinol by Q10 reductases. Then Q10 transfers
607 the electrons to complex III. The electron is then passed to complex IV through cytochrome
608 C. A component of Complex III can convert ubiquinol to ubiquinone to recycle Q10.

609 **Figure 2** The structure of the retina. (A) Cross-sectional image of the healthy retina obtained
610 by optical coherence tomography (OCT). Scans were taken with the upper panel showing the
611 64th scan and the lower panel showing the 256th scan. (B) Histological structure of mouse
612 retina obtained by hematoxylin-eosin staining (left panel) and by immunostaining with 1D4
613 antibody (labelling the rod outer segments, right panel). GCL, ganglion cell layer; INL, inner
614 nuclear layer; IS, inner segment; IPL, inner plexiform layer; L, lens; NFL, nerve fiber layer;
615 ONL, outer nuclear layer; OS, outer segments; RPE, retinal pigment epithelium.

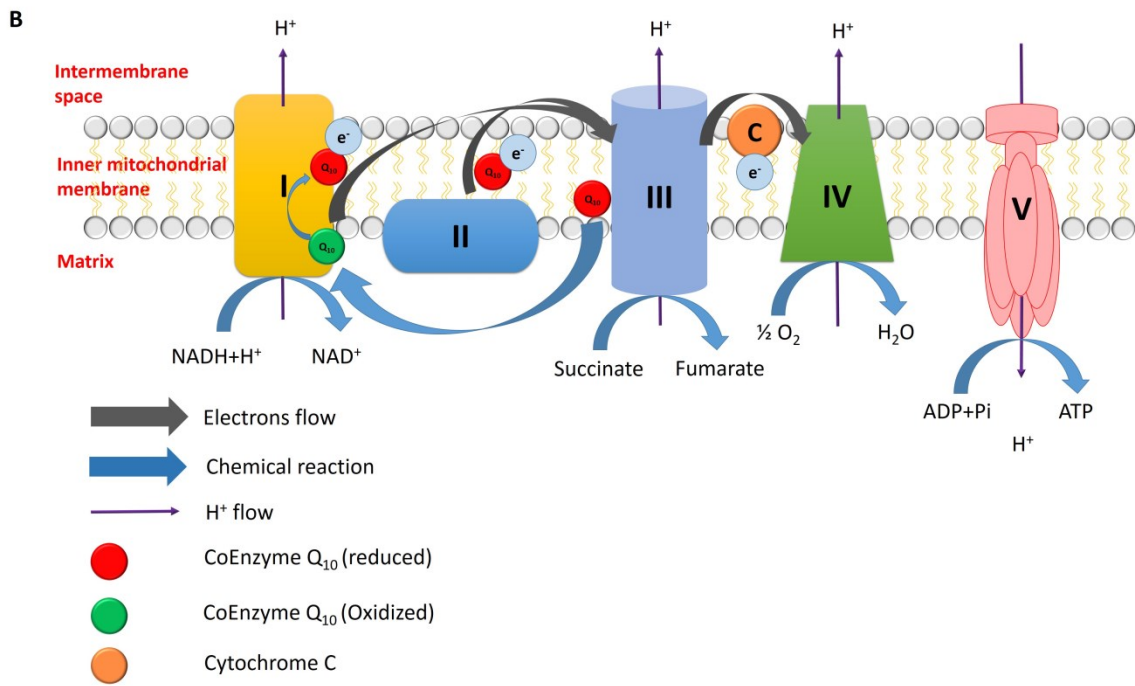
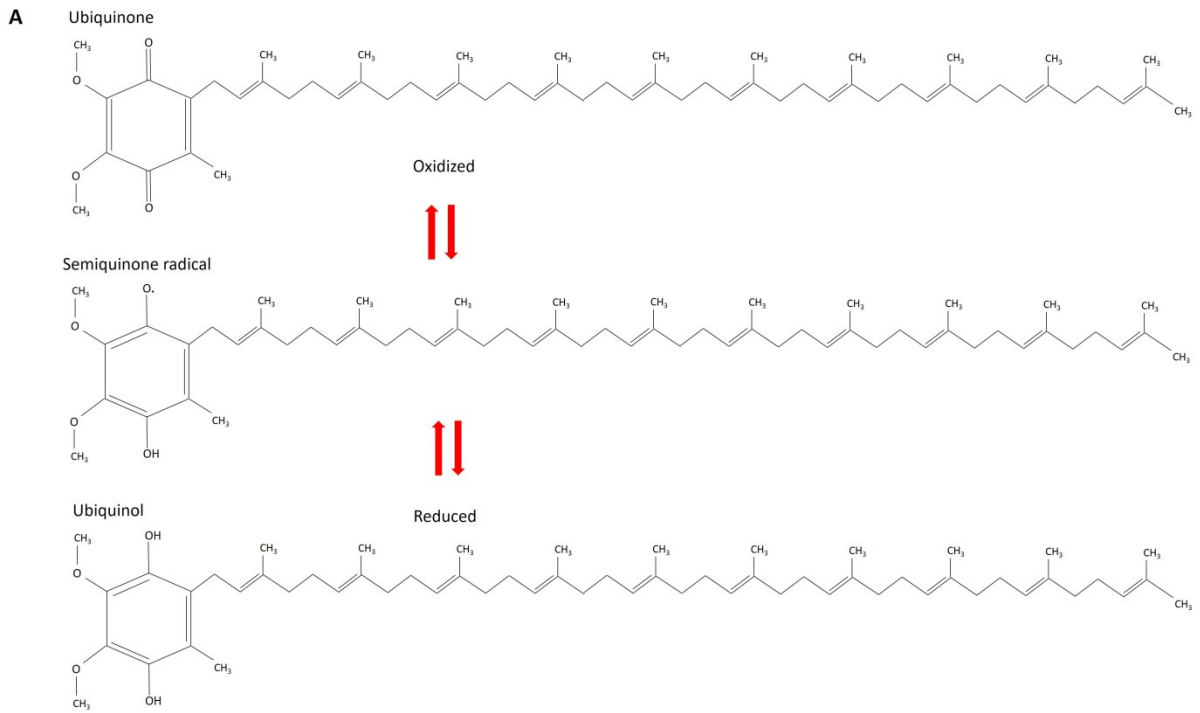
616 **Figure 3** Bone spicule pigment deposits are present in the fundus of retinitis pigmentosa
617 patient (right side). Fundus of healthy individual is on the left side. Adapted from Raghpathy
618 et al. (Ref 81)

619 **Figure 4** Clinical classification of diabetic retinopathy (DR) determined by ophthalmoscopy
620 (fundoscopy). (A) Healthy retina. (B) Background DR. (C) Mild non-proliferative diabetic
621 retinopathy (NPDR). (D) Moderate NPDR. (E) Severe NPDR. (F) Proliferative diabetic
622 retinopathy (PDR). (G) Diabetic macular edema (DME). Adapted from El-Bab et al., 2012
623 (Ref 82) and Shotliff and Duncan, 2006 (Ref 83).

624 **Figure 5** Diagram illustrating protection of co-enzyme Q10 (CoQ10) via inhibiting ROS
625 production. CoQ10 (ubiquinol) blocks the production of ROS and subsequently attenuates
626 oxidative damage and inflammation, which reduce death of retinal cells (photoreceptors,
627 retinal pigment epithelium cells and ganglion cells) and delays the progression of retinal
628 diseases (age-related macular degeneration, AMD; diabetic retinopathy, DR; retinitis
629 pigmentosa, RP; glaucoma).

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633 **Figure 1**



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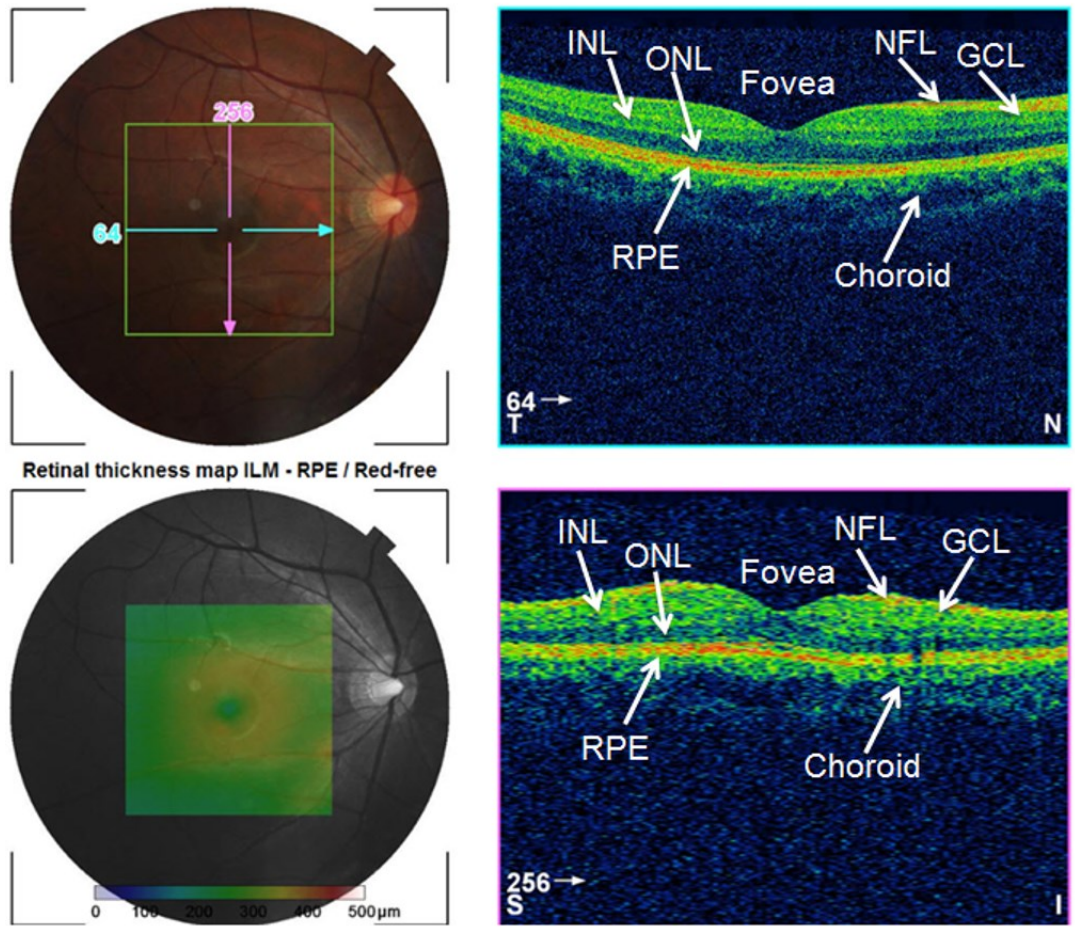
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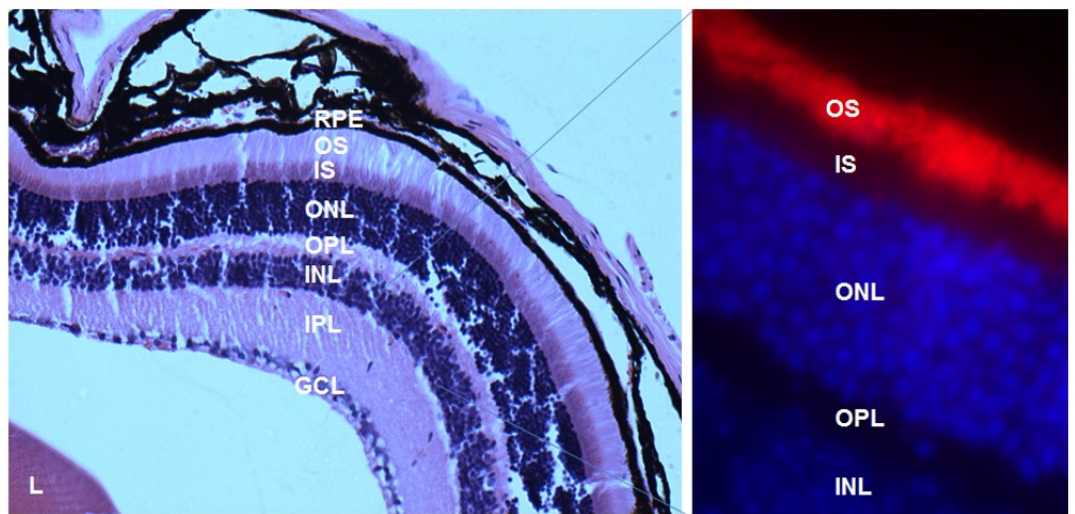
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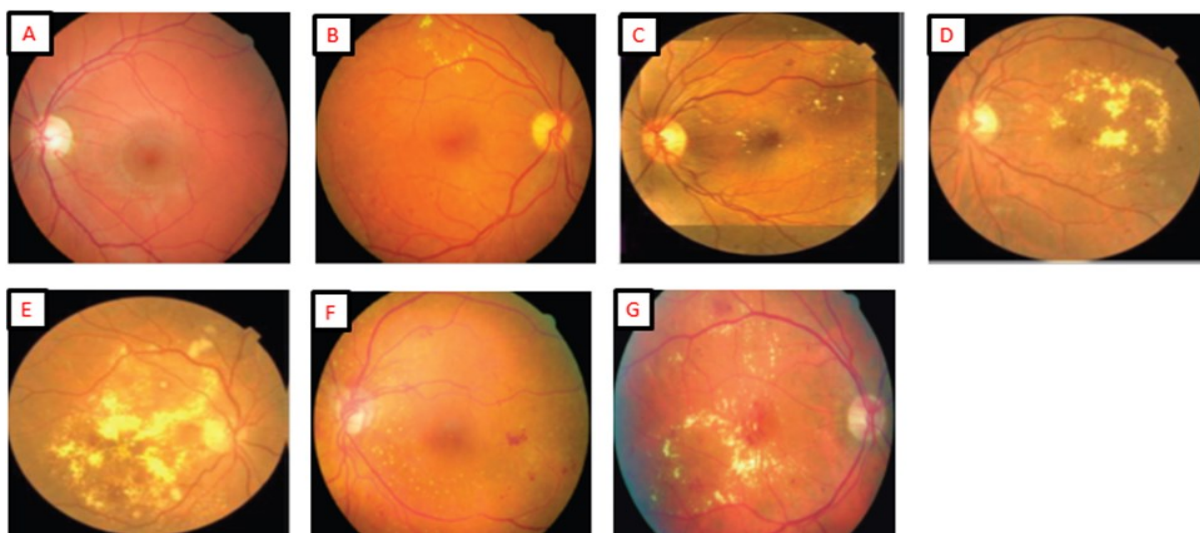
643 **Figure 3**



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646 **Figure 4**



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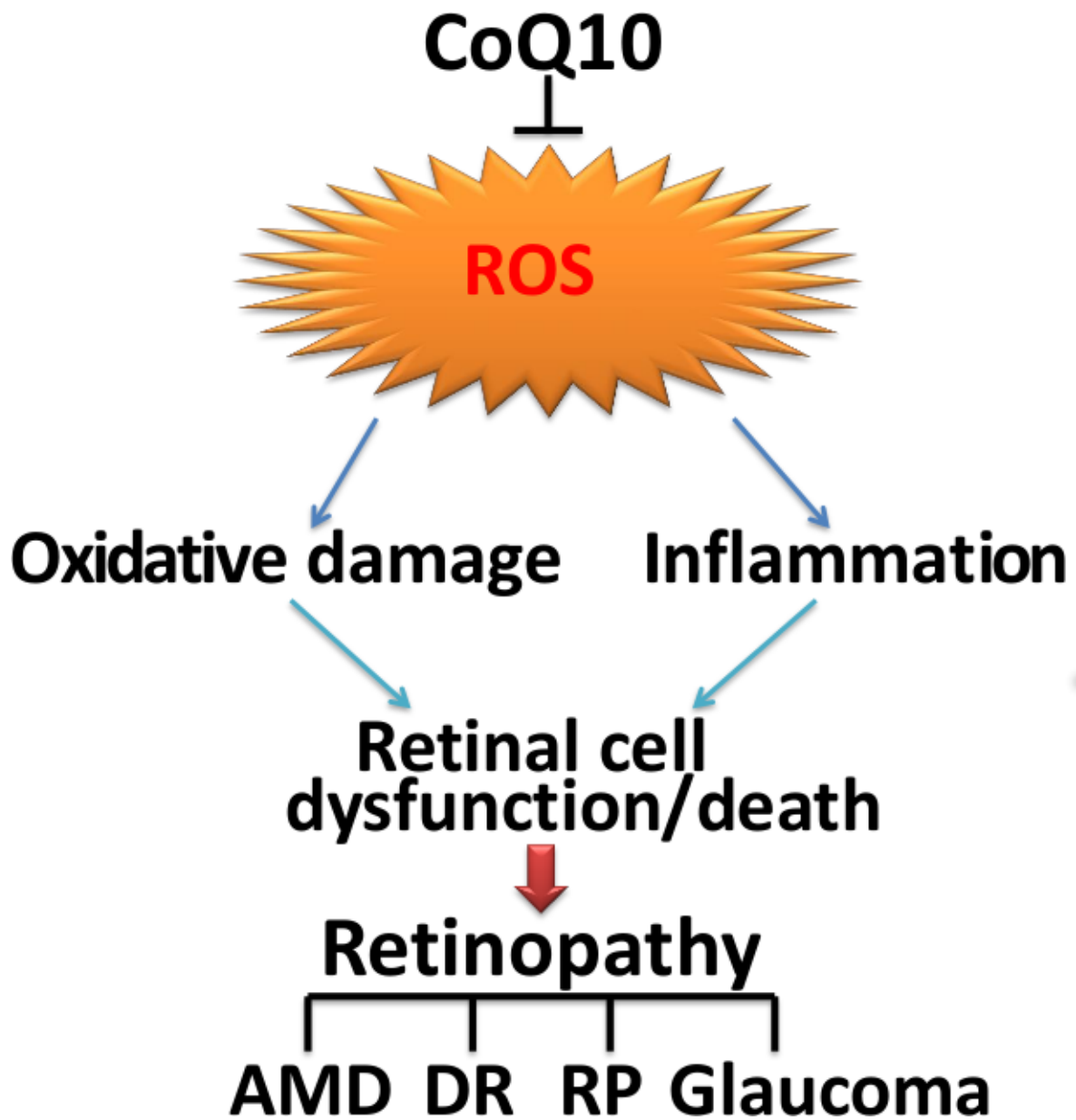
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