

Synthetic Approaches to Phosphasugars (2-oxo-1,2-oxaphosphacyclanes) Using the Anomeric Alkoxy Radical β -Fragmentation Reaction as the Key Step

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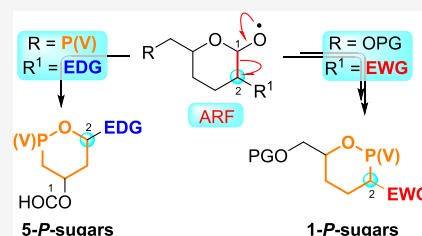


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ABSTRACT: The anomeric alkoxy radical β -fragmentation (ARF) of carbohydrates possessing an electron-withdrawing group (EWG) at C2, promoted by $\text{PhI}(\text{OAc})_2/\text{I}_2$, gives rise to an acyclic iodide through which a pentavalent atom of phosphorus can be introduced via the Arbuzov reaction. After selective hydrolysis and subsequent cyclization, the phosphonate or phosphinate intermediates can be converted into 2-deoxy-1-phosphahexopyranose and 2-deoxy-1-phosphapentopyranose sugars. The ARF of carbohydrates with an electron-donor group (EDG) at C2 proceeds by a radical-polar crossover mechanism, and the cyclization occurs by nucleophilic attack of a conveniently positioned phosphonate or phosphinate group to the transient oxocarbenium ion. This alternative methodology leads to 5-phosphasugars with a 4-deoxy-5-phosphapentopyranose framework. The structure and conformation of the 2-oxo-1,2-oxaphosphinane and 2-oxo-1,2-oxaphospholane ring systems in different carbohydrate models have been studied by NMR and X-ray crystallography.



INTRODUCTION

1-Phosphapyranosyl and 1-phosphafuranosyl carbohydrates possessing a 2-oxo-1,2-oxaphosphacyclane skeleton have attracted considerable attention from synthetic chemists due to their unusual non-natural structures and great diversity of biological activities.¹ Cyclic phosphonate and phosphinate esters with a six-membered 2-oxo-1,2-oxaphosphinane ring system, also known as phostones and phostines, respectively, have been prepared almost exclusively by acid- or base-catalyzed Pudovik–Abramov addition of the phosphorus atom to the sugar carbonyl.² Originally reported by Thiem et al.³ on a D-mannofuranose derivative, this methodology has been extended to other sugars giving always C1–P expanded 1,2-oxaphosphinane rings with respect to the original sugar.⁴ Generally, the reaction occurs with little stereoselectivity on the adjacent phosphorus and carbon centers, and a mixture of the four possible isomers is formed. Nevertheless, asymmetric Abramov hydrophosphonylation enables optimization of the synthesis of phostones for the formation of the manno-isomers.^{4f} The total synthesis of phostones using ring-closing metathesis has also been reported.⁵ What has been less well documented, however, is the preparation of 1-phosphafuranosyl carbohydrates. Cyclic phosphonates and phosphinates with a five-membered 2-oxo-1,2-oxaphospholane ring have been synthesized by the analogous unselective Abramov addition to the aldehyde in four-carbon-atom acyclic sugar derivatives.⁶ The total synthesis of racemic cyclic phosphonate analogs of 2-deoxy-ribose and 2-deoxy-xylose has also been accomplished.⁷ Some of these 2-

oxo-1,2-oxaphosphacyclanes of a carbohydrates origin have been shown to exhibit biological activities as anticancer⁸ and enzyme inhibitors.⁹

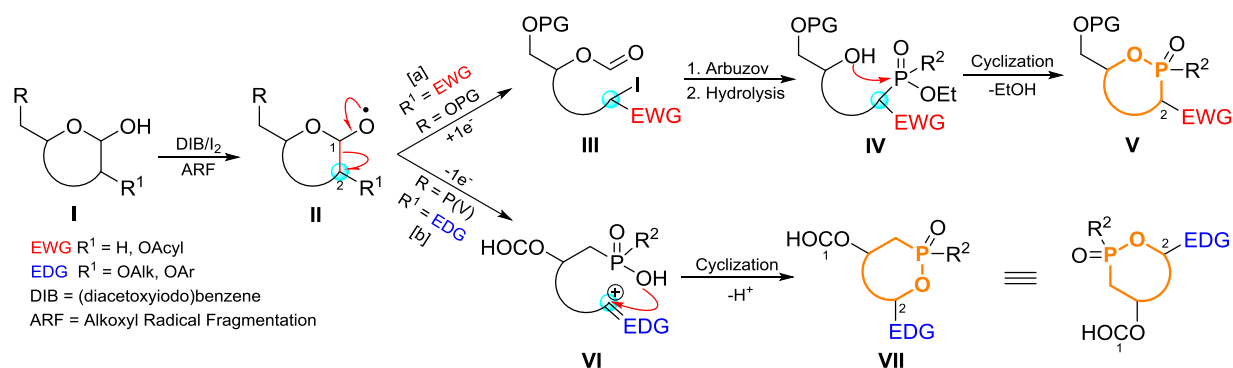
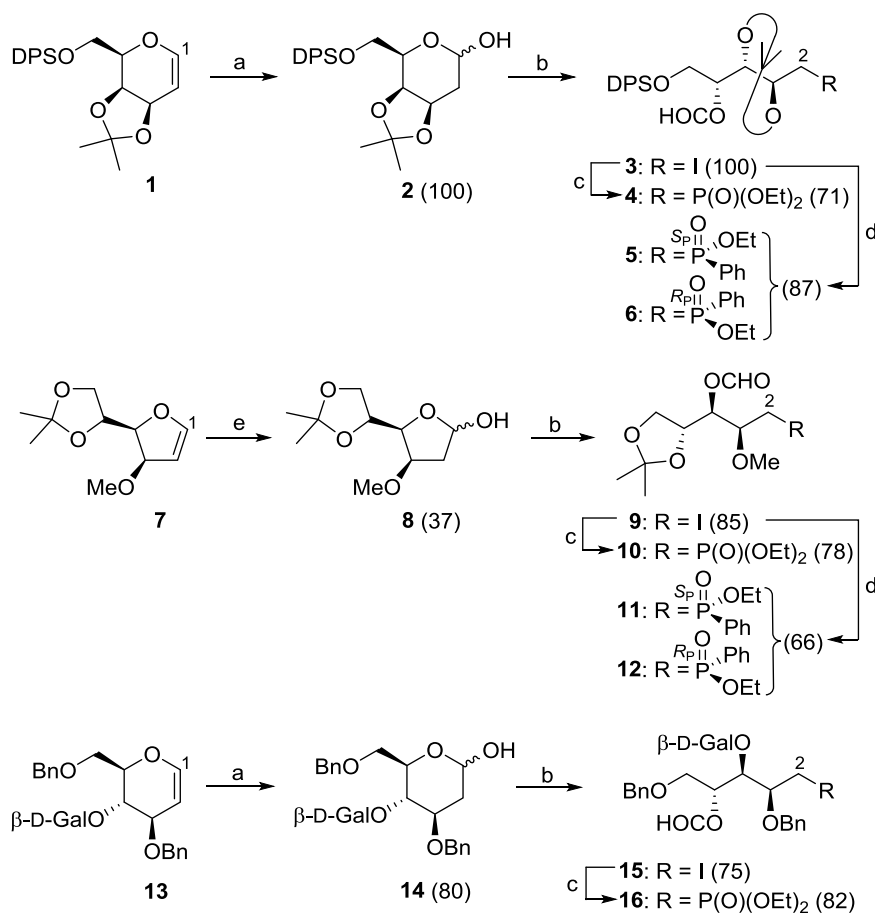
In previous papers, we have studied that the anomeric alkoxy radical (i.e., II, Scheme 1) β -fragmentation (ARF) of sugars initiated under oxidative¹⁰ or reductive¹¹ conditions. We have shown that with the system (diacetoxyiodo)benzene (DIB)/I₂, the reaction mechanism is strongly dependent on the polarity of the C2 substituent.¹² With an electron-withdrawing group (EWG), the C2 radical initially formed is trapped by the iodine atoms in the medium to give an open-chain iodide with one carbon less than the original carbohydrate (i.e., III). Notwithstanding, if the C2 substituent is an oxygen-containing electron-donor group (EDG), the intermediate radical is rapidly oxidized to an oxocarbenium ion by a radical-polar crossover mechanism,¹³ which may subsequently react inter- or intramolecularly with nucleophiles (i.e., VI).¹⁴

This unique behavior of the C2 radical during the ARF reaction motivated us to investigate the use of this dual

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Scheme 1. Approaches for the Synthesis of Phosphasugars

Scheme 2. Synthesis of Models for the Preparation of 1-Phosphasugars^a

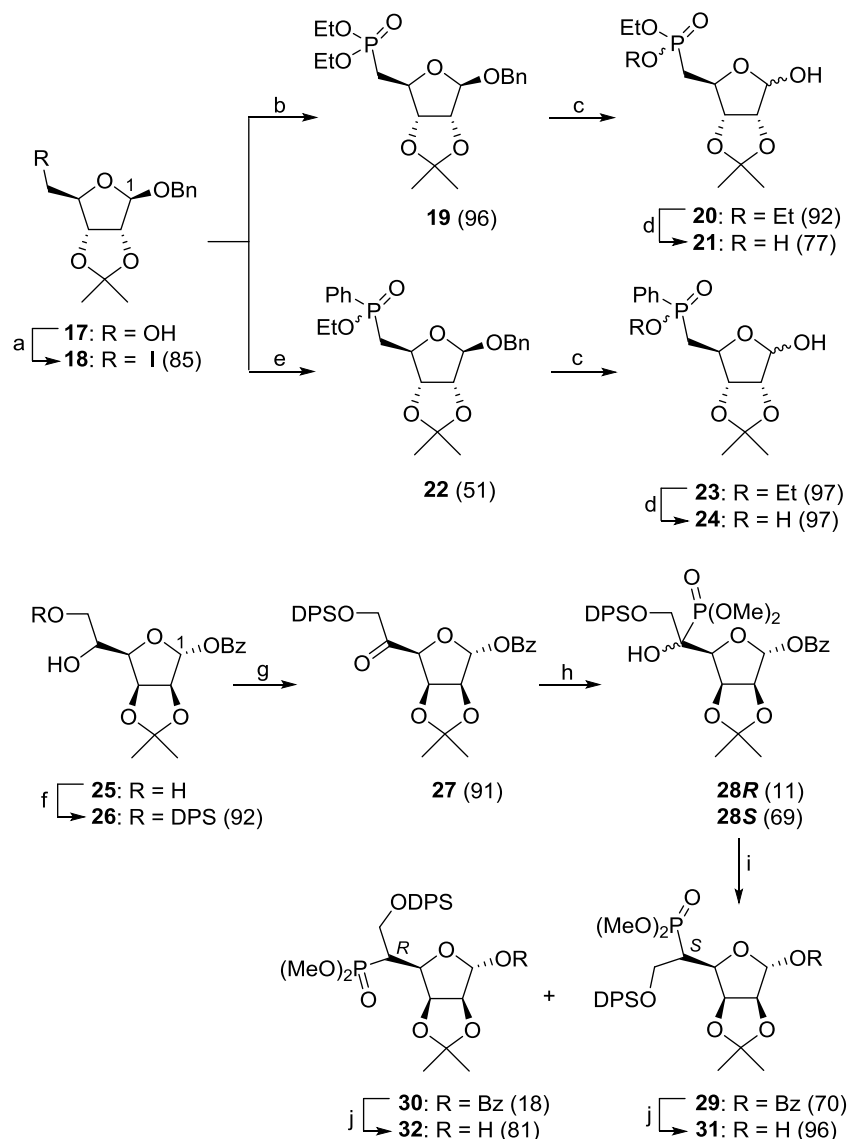
^aReagents and conditions: (a) $\text{Ph}_3\text{P}\cdot\text{HBr}$, tetrahydrofuran (THF), H_2O , 2–24 h; (b) $\text{PhI}(\text{OAc})_2/\text{I}_2$, CH_2Cl_2 , room temperature (rt), 1–2 h; (c) $(\text{EtO})_3\text{P}$, Δ , 3–6 h; (d) $(\text{EtO})_2\text{PPh}$, 160 °C, 6 h; (e) (i) $\text{Hg}(\text{OAc})_2$, THF, H_2O , rt, 1 h, (ii) KI , H_2O , 0 °C, 0.5 h, (iii) NaBH_4 , H_2O , 0 °C, 1.5 h. For brevity, β -D-Gal refers to the perbenzylated moiety of galactose.

67 methodology for the preparation of two completely different 2-oxo-1,2-oxaphosphacyclane ring systems.

69 In the first method (path a), the synthesis starts with a 2-deoxy-sugar (I, $R^1 = \text{H}$) and the phosphorus is introduced in the final steps of the synthesis by the Michaelis–Arbuzov reaction of the iodide generated by the ARF reaction (III).¹⁵ The obtained phosphinates or phosphonates after hydrolysis of the formyl group (IV, $R^2 = \text{Aryl, OAlkyl}$) can be conveniently cyclized to 2-oxo-1,2-oxaphosphinanes (V, $R^2 = \text{Aryl, OAlkyl}$). The final result of this method is the formation of 1-phosphasugars by net substitution of the anomeric carbon by

an atom of pentavalent phosphorus, maintaining the ring size and the stereochemistry of the original 2-deoxy-sugar.¹⁶

The second approach (path b) requires the placement of an EDG at C2 (I, $R^1 = \text{OAlkyl}$), and phosphorus is now introduced at earlier steps of the synthesis, before the ARF reaction. As can be seen, the same four steps of the previous methodology are now introduced in a different order: Arbuzov–hydrolysis–ARF–cyclization. The strategically located monophosphonate ester (VI, $R^2 = \text{OAlkyl}$) or phosphinic acid (VI, $R^2 = \text{Aryl}$) is cyclized by a tandem ARF–nucleophilic addition on the intermediate oxocarbenium

Scheme 3. Synthesis of Models for the Preparation of 5-Phosphasugars^a

^aReagents and conditions: (a) I_2 , PPh_3 , imidazole, PhH , Δ , 0.5 h; (b) $(EtO)_3P$, Δ , 28 h; (c) H_2 , Pd/C 10%, $EtOAc$, 2.5–20 h; (d) $NaOH$, $EtOH$, rt, 42–48 h; (e) $(EtO)_2PPh$, $160^\circ C$, 28 h; (f) $TBDPSiCl$, imidazole, 4-dimethylaminopyridine (DMAP), dimethylformamide (DMF), rt, 2 h; (g) pyridinium chlorochromate (PCC), $MS\ 3\ \text{\AA}$, CH_2Cl_2 , rt, 9 h; (h) $(MeO)_2P(O)H$, DBU , $0^\circ C$, 0.5 h; (i) (i) $MeO(CO)_2Cl$, DMAP, $MeCN$, $0^\circ C$, 1 h, (ii) Bu_3SnH , AIBN, $PhCH_3$, Δ , 2 h; (j) $NaOMe$, $MeOH$, rt, 2 h.

89 ion. In the new 2-oxo-1,2-oxaphosphinane (**VII**, $R^2 = \text{Aryl}$,
 90 OAlkyl) formed, the ether bridge is now linked to C2 and the
 91 ring has been expanded in relation to the original sugar. This
 92 structure can actually be considered to be a 4-deoxy-5-
 93 phosphapyranose sugar.

94 ■ RESULTS AND DISCUSSION

95 **Preparation of Carbohydrate Models.** Intermediates
 96 primary alkyl iodides **3**, **9**, and **15** were prepared from the
 97 corresponding 2-deoxy-sugars **2**, **8**, and **14** via anomeric
 98 alkoxy radical fragmentation with DIB and iodine in good to
 99 excellent yield (Scheme 2). The 2-deoxy-sugars **2** and **14** were
 100 in turn synthesized using a modification of the procedure
 101 developed by Mioskowski and Falck¹⁷ by catalytic hydration
 102 of glycols **1** and **13** employing triphenylphosphine hydro-
 103 bromide and water as nucleophile. This reaction failed in the
 104 case of 2-deoxy-pent-1-enitol **7**, and an oxymercuration-

demercuration protocol¹⁸ with the glycol **7** was utilized instead
 105 (Scheme 2). The required phosphonates **4**, **10**, and **16** and
 106 phosphinates **5**, **6**, **11**, and **12** were prepared by the Michaelis–
 107 Arbuzov reaction: by heating the corresponding primary
 108 iodides with triethyl phosphate and diethyl phenylphosphonite,
 109 respectively. 110

Although the high temperature required was feared to be a
 111 problem with these sensitive substrates, the reaction proceeded
 112 efficiently.¹⁹ The pairs of phosphorus-centered diastereomers
 113 of phenylphosphinates **5**, **6** and **11**, **12** could be separated by
 114 high-performance liquid chromatography (HPLC), although
 115 the absolute stereochemistry was more conveniently deter-
 116 mined at later stages after the cyclization step (vide infra). 117

The preparation of models **21**, **24**, **31**, and **32** for the
 118 synthesis of 1,2-oxaphosphinanes using the second method-
 119 ology (route b, Scheme 1) is described in Scheme 3. To access
 120 **5**-phosphonate **19** and **5**-phosphinate **22**, benzyl 2,3-*O*-
 121

Table 1. Synthesis of 1-Phosphosugars

entry	substrate	time (h)	products (yield)
1		102	
2		24	
3		24	
4		86.5	
	\xrightarrow{a} 10 : R = CHO \xrightarrow{b} 33 : R = H ^b		
5		94	
	\xrightarrow{a} 11 : R = CHO \xrightarrow{b} 34 : R = H ^b		
6		94	
	\xrightarrow{a} 12 : R = CHO \xrightarrow{b} 35 : R = H ^b		
7		27	

^aReagents and conditions per mmol of substrate: NaOEt/EtOH (0.01 M, 0.09 mmol), rt. ^bPyridinium *p*-toluenesulfonate (PPTS) (2.0 mmol), MS 4 Å, CHCl₃, reflux. ^cFor brevity, β-D-Gal refers to the perbenzylated moiety of galactose.

122 isopropylidene-β-D-ribofuranoside (**17**) was converted into the
 123 5-iodo derivative **18** in 85% yield by treatment with
 124 triphenylphosphine in the presence of iodine and imidazole.
 125 Iodide **18** was subsequently submitted to Michaelis–Arbuzov
 126 conditions, as described above, to afford 5-phosphonate **19** and
 127 5-phosphinate **22** derivatives, that were then debenzylated by
 128 catalytic hydrogenolysis in the presence of Pd/C 10% under 1
 129 atm of H₂ to deprotect the anomeric alcohols to give **20** and
 130 **23**, respectively. Selective hydrolysis of the phosphonate
 131 diester **20** and the phosphinate ester **23** was performed using
 132 sodium hydroxide in ethanol to furnish the required monoester
 133 **21** and phosphinic acid derivative **24**, respectively.
 134 Finally, the introduction of the 5-dimethoxyphosphoryl
 135 group in a mannofuranose skeleton was accomplished through

a methodology developed by Hanaya et al.,²⁰ which essentially
 136 consists of an Abramov addition to a 5-ulose carbohydrate
 137 (**Scheme 3**). The sequence was initiated by the selective
 138 protection of 1-O-benzoyl-O-isopropylidene-α-D-mannofura-
 139 nose (**25**) with ^tBuPh₂SiCl to give **26**. The 5-ulose **27** was
 140 efficiently obtained by the oxidation of the secondary alcohol
 141 with PCC, and the addition of dimethyl phosphite to the
 142 carbonyl group in the presence of DBU afforded a mixture of
 143 α-hydroxyphosphonate esters **28R** and **28S**. Both isomers can
 144 be separated by chromatography, but since the next step is the
 145 deoxygenation of the tertiary 5-hydroxyl group, the determi-
 146 nation of 5-stereochemistry at **28** was deemed to be
 147 unnecessary. Nevertheless, the stereochemistry has been 148

149 tentatively assigned in analogy with that observed in a similar
150 case.²⁰

151 The deoxygenation was carried out by the Dolan and
152 MacMillan protocol;²¹ the 5-*O*-methyl oxalyl derivative
153 prepared by methyl oxalyl chloride in the presence of DMAP
154 was then reduced with the *n*-Bu₃SnH/AIBN system to give a
155 mixture of **29** and **30** (88%, 80:20). The C-5 stereochemistry
156 was also more easily accomplished in the next step after the
157 cyclization. Debenzoylation with NaOMe in MeOH afforded
158 models **31** and **32** with the free anomeric alcohol prepared for
159 the ARF reaction.

160 **Synthesis of 1-Phosphasugars.** Open-chain diethoxy-
161 phosphoryl intermediate **4** cyclized by a base-catalyzed
162 intramolecular transesterification process furnished S_p and R_p
163 diastereomeric mixtures of 1-phospha-1-oxo-*D*-*lyxo*-hexopyra-
164 nose derivatives **36** and **37** (Table 1, entry 1). The sensitive δ-
165 formate ester was hydrolyzed rapidly under the reaction
166 conditions. The R_p-isomer **37** in which the ethyl group is
167 disposed at the less sterically congested α-side of the ring is the
168 major component. Nuclear Overhauser effect spectroscopy
169 (NOESY) experiments were used for the assignment of the
170 H2α and H2β protons and the relative configuration of the
171 phosphorus chiral center [see Figures S1 and S2 in the
172 Supporting Information (SI)]. The conformational situation of
173 the 1,2-oxaphosphacyclane ring was evaluated on the basis of
174 ³J_{HHβ}, ³J_{PHβ}, and ³J_{PC} vicinal NMR coupling constants suggesting
175 a preferential B_{2,5} conformation in both compounds.²² The
176 values of the ³J constants calculated on minimized structures of
177 **36** and **37**, using the corresponding Karplus equations,²³ agree
178 well with the experimental data (see Tables S1 and S2 in the
179 SI).

180 The cyclization proceeded analogously with pure phenyl-
181 phosphinates **5** and **6** to give 2-phenyl-1,2-oxaphosphinanes **38**
182 and **39**, respectively, in good overall yield (Table 1, entries 2
183 and 3). The ring conformations are similar to those previously
184 commented for compounds **36** and **37** and very close to a
185 theoretical B_{2,5} boat (Cremer-Pople parameters: φ = 290°, θ =
186 94°, Q = 0.7 Å) confirmed by the large ³J_{PCCH₃} (24 Hz)
187 coupling constant observed, consistent with an almost
188 antiperiplanar arrangement of the atoms (torsion angle ≈
189 170°) (Figures and Tables S3 and S4 in the SI). The *P*-
190 stereochemistry has also been established with the aid of
191 NOESY experiments. The NOESY spectrum of **38** unequiv-
192 ocaly related the phenylphosphinate protons to β-face
193 hydrogens (H2β and the endo methyl group of the
194 isopropylidene), while in the spectrum of **39**, the phenyl
195 protons are related to α-face hydrogens (H2α and H5). In this
196 context, the substantial deshielding of the endo methyl group
197 of the isopropylidene (ca. 0.26 ppm) is also significant, with
198 the consequent shielding of the 2Hα (−0.25 ppm) and 5H
199 (−0.58 ppm) protons observed on going from the ¹H NMR
200 spectrum of **38**–**39** caused by the anisotropic shielding effect
201 of the P–Ph group. Under these conditions, the cyclization
202 proceeded stereospecifically; only one isomer was formed in
203 each case, as detected by ³¹P NMR spectroscopy of the crude
204 reaction mixture. As described in the literature, the phosphorus
205 nucleophilic substitution mechanism is produced through an
206 intermediary with trigonal bipyramidal geometry, the final
207 result being a *P*-stereochemical inversion.²⁴ Consequently, the
208 configuration of open-chain precursors **5** and **6** can now be
209 determined as S_p and R_p, respectively.

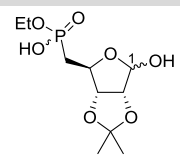
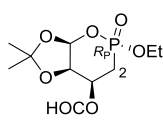
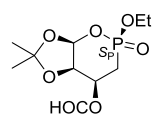
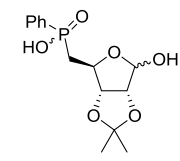
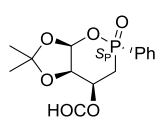
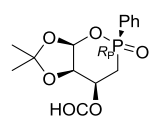
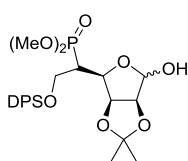
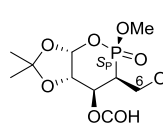
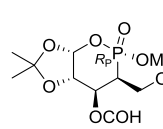
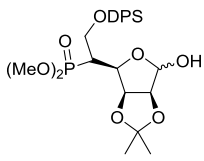
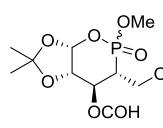
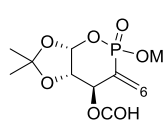
This methodology can readily be extended to the synthesis
of five-membered 1,2-oxaphospholanes ring compounds.
Starting from a hexose sugar in furanose form [e.g., 2-deoxy-
5,6-*O*-isopropylidene-3-*O*-methyl-*D*-*arabino*-hexofuranose (**8**),
Scheme 2], the alkoxy radical fragmentation led, across γ-
hydroxy-phosphonates (e.g., **33**) or -phosphinates (e.g., **34** and
35), finally to the required 1,2-oxaphospholanes (e.g., **40**–**43**).

In this particular model, the cyclization step was performed
using PPTS in anhydrous CHCl₃ after hydrolysis of the formyl
ester. This two-step method provides better yields than the
direct cyclization used previously for the six-membered rings. It
has been reported in the literature that the intramolecular
transesterification is highly favored in the case of five-
membered 1,2-oxaphospholanes.²⁵ This may be the reason
why small amounts of the corresponding cyclized compounds
can be detected during the hydrolysis of the formyl group in
the preparation of γ-hydroxy-precursors **33**–**35** (Table 1,
entries 4–6), or even in the Arbuzov reactions of iodide **9**.
Indeed, a mixture of cyclized **42** and **43** can be obtained
directly when heating **9** with PhP(OEt)₂ at 120 °C for a long
period of time (68 h), although in moderate yield (37%, 6:4,
three steps). The *P*-stereochemistries were assigned by analysis
of their NOESY spectra (Figures S5–S8 in the SI) and in the
case of 2-phenyl-1,2-oxaphospholanes **42** and **43** confirmed by
the chemical shift displacement of H2α (−0.32 ppm), H2β
(0.14 ppm), and 3β-methoxyl group (0.1 ppm) observed on
going from the ¹H NMR spectrum of **42**–**43** motivated by the
aromatic ring current effect. According to the ³J_{HHβ}, ³J_{PHβ}, and
³J_{PC} coupling constants, the 1,2-oxaphospholane rings in
compounds **40**–**43** adopt preferentially very similar ³T₂
conformation patterns. The experimental ³J values are
consistent with those calculated on a minimized ³T₂ structure.
This is also in accordance with a pseudorotational analysis of
the ³J_{HH} ring constants. The most populated conformers
appear at phase angles of P = 0–42° (³T₂–³E–³T₄) of the
northern region of the pseudorotational itinerary (see Tables
S5–S8 in the SI for details).²⁶

For the sake of completeness, this methodology was also
extended to a disaccharide model derived from perbenzylated
D-lactose [β-*D*-Galp-(1 → 4)-*D*-Glc]. The diethyl phospho-
nate **16** was effectively cyclized under similar reaction
conditions to give a 1,2-oxaphosphinane derivative as a
mixture of *P*-diastereomers **44** and **45** (62%, 1:2) (Table 1,
entry 7). The reaction can be expected to come under some
steric control and to lead to the major diastereomer **45** having
the ethyl group positioned on the less hindered α-side of the
ring. NOE interactions observed between this ethyl group and
the H2α and H3α protons are also in agreement with the R_p
stereochemistry assigned to this isomer (Figures S9 and S10 in
the SI). In this case, the 1,2-oxaphosphinane ring adopts
preferentially a ⁴C₁ chair conformation in both isomers.²⁷ This
is evident from the ³J_{HH} vicinal coupling values between the
axial ring protons (H2β, H3, H4, and H5) and further
supported by the relatively small ³J_{PCCH₃β} (ca. 2 Hz) observed,
which is consistent with a P–C₂–C₃–H dihedral angle of
approximately 60° (Table S9 and S10 in the SI).^{5b} This is the
first synthesis of a β-*D*-Galp-(1 → 4)-1-phospha-*D*-Arap
structure via direct introduction of the phosphorus at the
reducing unit on a disaccharide framework.²⁸

Synthesis of 5-Phosphasugars. The second approach
was first tested with compound **21**, which possesses a tethered
5-phosphonate monoester functionality on a *D*-ribofuranose

Table 2. Synthesis of 5-Phosphasugars^{abc}

entry	substrate	time (h)	products (yield)
1		2.5	 (32) +  (15)
2		3.5	 (29) +  (51)
3		2	 (26) ^d +  (42) ^d
4		5.5	 (n.d.) +  (13) ^d

^aReagents and conditions per mmol of substrate: DIB (2.0 mmol), I₂ (1.1 mmol), CHCl₃, rt. ^bDIB (2.2 mmol), I₂ (1.1 mmol), CH₂Cl₂, rt, *hν*. ^c(i) NaOH/MeOH (2 M, 5.3 mL), 40–43 °C, 30 h; (ii) DIB (2.1–2.2 mmol), I₂ (1.0–1.1 mmol), CH₂Cl₂, rt, *hν*. ^dTwo-step overall yield. n.d. = not detected.

272 skeleton. The reaction with PhI(OAc)₂ and iodine triggers the
273 alkoxy radical fragmentation and the intramolecular cycliza-
274 tion via a nucleophilic radical-polar crossover mechanism. In
275 consequence, a 2-ethoxy-1,2-oxaphosphinane 2-oxide ring
276 system was formed and isolated as a chromatographically
277 separable mixture of *P*-diastereomers **46** and **47** (Table 2,
278 entry 1). The 6-*exo* cyclization is highly stereoselective; only
279 one isomer at C5 is formed leaving the isopropylidene in *cis*
280 disposition. *P*-Stereochemistries were assigned by NOESY
281 analyses, and the rings adopt predominantly very similar *E*₃
282 conformations. The calculated vicinal coupling constants in
283 these *E*₃ minimized structures agreed reasonably well with the
284 experimental values (Figures and Tables S11 and S12 in the
285 SI). The ¹H NMR spectrum of **46** shows a ⁴J_w coupling (0.7
286 Hz, calcd 1.2 Hz)²⁹ between H2 α and H4 hydrogens, which
287 also supports the previously mentioned conformation.

288 Curiously, in the ¹H NMR spectra of both isomers, the
289 formyl hydrogen signal appears splitting as a doublet of
290 doublets with two long-range couplings: ⁴J_w with the H3 (ca. 1
291 Hz) and ⁵J_{PH} with the atom of phosphorus (ca. 2.8 Hz), as
292 determined by selective spin decoupling and ¹H{³¹P} NMR
293 spectroscopy, respectively.

294 The cyclized 2-phenyl-1,2-oxaphosphinanes **48** and **49** were
295 prepared analogously from the free phosphinic acid **24** (Table
296 2, entry 2). The stereochemical assignment of these examples
297 rests on spectroscopic evidences and on a single-crystal X-ray
298 crystallographic analysis of **48** (Figure S17 in the SI). The
299 solution conformation of the 1,2-oxaphosphinane ring,
300 elucidated from the experimental homo- and heteronuclear ³J

coupling constants of the ring protons, was practically identical
301 to the slightly distorted B_{3,0} boat conformation (Cremer-Pople
302 parameters: $\varphi = 188.9^\circ$, $\theta = 79.5^\circ$, $Q = 0.636 \text{ \AA}$) observed in
303 the crystalline structure of **48** (Figure and Table S13 in the SI).
304 The ⁴J_w coupling (0.9 Hz, calcd 1.2 Hz)²⁹ between H2 α and
305 H4 hydrogens provides additional support for this conforma-
306 tion. Interestingly, the ring of the R_p isomer **49** seems to adopt
307 in solution a conformation closer to a ⁴C₁ chair than to the B_{3,0}
308 boat (Figure and Table S14 in the SI). The most significant
309 difference was observed in the heteronuclear ³J_{POCH₃} coupling
310 constant [**48** (19.0 Hz, calcd 21.1 Hz, TA = -161.5°, B_{3,0}); **49**
311 (0.7 Hz, calcd 1.5 Hz, TA = -89.6°, ⁴C₁)] in full agreement
312 with the Karplus equation and the geometry of the rings.³⁰ 313

314 An example with the phosphorous on a secondary carbon of
the sugar side chain has also been accomplished. The 5-
315 dimethoxyphosphoryl-L-gulofuranose derivative (**31**) was
316 partially hydrolyzed to the monoester and immediately
317 subjected to the ARF reaction (Table 2, entry 3). The *P*-
318 configuration of the two 1,2-oxaphosphinane derivatives
319 obtained **50** and **51** was determined on the basis of NOE
320 interactions of the P-OMe with protons in close spatial
321 proximity to the β - and α -side of the ring, respectively (Figures
322 S15 and S16 in the SI). The ³J_{H2 α ,H3} coupling constant and the
323 NOE interaction of the H2 with the endo methyl group of the
324 isopropylidene established the stereochemistry at C2 and,
325 consequently, confirmed that of compounds **28**–**32**. Both
326 epimeric compounds appear to exist in solution predominantly
327 in a ³E conformation (Tables S15 and S16 in the SI). 328

329 However, the cyclization completely failed with the minor
330 isomeric 5-dimethoxyphosphoryl-D-mannofuranose derivative
331 (32) (Table 2, entry 4). The reaction is slower, and the
332 expected 5-phosphasugar 52 could not be detected in the
333 complex mixture of products obtained. Instead, a compound
334 eventually identified as the cyclized olefin 53 was obtained by
335 chromatography in a low yield. A hypothesis that can
336 rationalize this result is based on unfavorable steric interactions
337 between the large side chain and the crowded α -side of the
338 molecule that may hinder the cyclization step. The formation
339 of 53 may be explained by cyclization after desilylation and
340 subsequent elimination of the alcohol. The 3-methylene-1,2-
341 oxaphosphinane structure of 53 was characterized by
342 spectroscopic means. Most significantly, $^3J_{\text{PH6cis}} = 23.6$ Hz
343 and $^3J_{\text{PH6trans}} = 43.8$ Hz coupling constants of the methylene
344 protons are observed in the ^1H NMR spectrum.³¹

345 **Assignment of P-Stereochemistry.** Phosphorous stereo-
346 chemistry in cyclic phosphonates or phosphinates was
347 determined by studying one-dimensional (1D) and two-
348 dimensional (2D) NOE experiments on minimized structures
349 with the established conformation for the five- or six-
350 membered rings. In some cases, other ^1H and ^{31}P NMR
351 supporting data have also been taken into account. In general,
352 it has been observed that in hexopyranoses possessing a 2-
353 methoxy-1,2-oxaphosphinane 2-oxide ring system, the signal of
354 the $^{31}\text{P}\{^1\text{H}\}$ NMR for the axial α -P-OMe is 2–4 ppm upfield
355 from the corresponding equatorial β -P-OMe.^{4b,c,f,9b} This
356 trend has also been detected in our 2-deoxy compounds,
357 although the observed difference is smaller (ca. 1 ppm)
358 [compare ^{31}P -chemical shift displacements of compounds 36–
359 37 (1 ppm), 44–45 (1 ppm), and 47–46 (1.6 ppm)]. In some
360 previous cases, this decrease has been attributed to conforma-
361 tional changes in the chair ring, and this may be the reason for
362 the small difference observed in our compounds.^{4c} Five- or six-
363 membered cyclic phosphinates show a greater difference in the
364 ^{31}P NMR chemical shifts displacement between both P -
365 isomers [i.e., 38–39 (3.3 ppm), 42–43 (4.0 ppm), and 49–48
366 (3.2 ppm)]. This trend, which the small number of examples
367 does not allow us to generalize, however coincides with that
368 observed for another pair of this type of compounds described
369 in the literature.^{4h} The $^2J_{\text{PH2}}$ coupling constant has also been
370 used for the determination of the configuration at phosphorus
371 in the glucopyranose series. This constant is considerably
372 smaller in the α -P-OMe (2–5 Hz) than in the β -P-OMe
373 compounds (6–10 Hz).^{4f} As one might expect this correlation
374 of $^2J_{\text{PH2}}$ with phosphorus, stereochemistry does not seem to be
375 applicable to our 2-deoxy compounds, in which the $^2J_{\text{PH2}\alpha}$ and
376 $^2J_{\text{PH2}\beta}$ are in general very similar and considerably higher (ca.
377 16–20 Hz).

378 ■ CONCLUSIONS

379 In summary, we have successfully applied the anomeric ARF
380 reaction for the synthesis of 1-phosphahexopyranose, 1-
381 phosphahexofuranose, and 5-phosphapentopyranose sugars.
382 The process initiated by the 1-O-yl radical is strongly
383 influenced by the polarity of the C2 substituent. In the
384 presence of an EWG at C2, rapid β -fragmentation affords a
385 stabilized C2 radical, which is trapped by the iodine atoms
386 present in the medium. The acyclic iodide can be readily
387 transformed into 1-phosphasugars using a three-step sequence:
388 Arbuzov reaction, selective hydrolysis, and intramolecular
389 cyclization. Cyclic phosphonates and phosphinates with 2-

oxo-1,2-oxaphosphinane and 2-oxo-1,2-oxaphospholane ring
systems can be obtained using this methodology. The 1-
phosphasugar ring has the same number of atoms and
stereochemistry as the original carbohydrate. This sequence
has also been applied to the synthesis of β -D-Galp-
(1 \rightarrow 4)-1-phospha-D-Arap with a disaccharide structure.
Nevertheless, if the C2 substituent is an EDG, the C2 radical
intermediate undergoes one-electron oxidation causing a
crossover from the radical to an oxocarbenium ion that can
be trapped intramolecularly by tethered P -nucleophiles
strategically located in the carbohydrate skeleton. This
methodology using the same sequence of reactions, although
in a different order, provides access to ring-expanded
phosphinates and phosphonates with a hitherto unknown 5-
phosphapentopyranose skeleton. The five- and six-membered
cyclic phosphinates and phosphonates prepared herein present
a wide variety of solution conformations that have been
studied using 3J homo- and heteronuclear coupling constants.

■ EXPERIMENTAL SECTION

General Methods. Melting points were determined with a hot-
stage apparatus. Optical rotations were measured at the sodium line at
ambient temperature in CHCl_3 solutions. IR spectra were measured as
thin films on CHCl_3 solutions. NMR spectra were determined at 500
MHz for ^1H , 125.7 MHz for $^{13}\text{C}\{^1\text{H}\}$, and 162 or 202.5 MHz for
 $^{31}\text{P}\{^1\text{H}\}$ in CDCl_3 as stated. The chemical shifts are given in parts per
million (ppm) relative to TMS at δ 0.00 ppm or to residual CDCl_3 at
 δ 7.26 ppm for proton spectra, relative to CDCl_3 at δ 77.00 ppm for
carbon spectra, and relative to external phosphoric acid at δ 0.00 ppm
for phosphorus spectra. ^{13}C DEPT-90, –135 and 2D correlation
spectroscopy (COSY), heteronuclear single-quantum coherence
(HSQC), NOESY, and heteronuclear multiple bond correlation
(HMBC) experiments were performed routinely for all new
compounds. When necessary, J_{PH} coupling constants have been
determined using $^1\text{H}\{^{31}\text{P}\}$ NMR decoupled spectra. The DAISY
program as implemented in the TopSpin 4.0.6 software package was
used for the simulation of ^1H NMR spectra. Low- and high-resolution
mass spectra were recorded using electrospray ionization (ESI⁺) and
time-of-flight (TOF) analyzer. Merck silica gel 60 PF (0.063–0.2
mm) was used for column chromatography. Circular layers of 1 and 2
mm of Merck silica gel 60 PF₂₅₄ were used on a Chromatotron for
centrifugally assisted chromatography. HPLC separations were
undertaken using a semipreparative (10 mm \times 250 mm) Ascentis
Si normal-phase column. Commercially available reagents and
solvents were of analytical grade or were purified by standard
procedures prior to use. The spray reagents for TLC analysis were
used with 0.5% vanillin in H_2SO_4 –EtOH (4:1), with Hanessian's
stain,³² or alternatively with ninhydrin and further heating until
development of color.

6-*O*-*tert*-Butyldiphenylsilyl-2-deoxy-3,4-*O*-isopropylidene-*D*-lyxo-
hexopyranose (2).³³ A solution of glycal 1³⁴ (228.0 mg, 0.537 mmol)
in THF (11.7 mL) containing water (233 μL , 12.9 mmol) and Ph_3P -
HBr (29.5 mg, 0.086 mmol) was stirred at room temperature for 24 h.
The reaction mixture was then poured into a saturated solution of
 NaHCO_3 and extracted with CH_2Cl_2 . The combined organic extracts
were washed with a saturated solution of NaHCO_3 , dried over
 Na_2SO_4 , and concentrated under reduced pressure. The residue was
purified by silica gel column chromatography (hexanes–EtOAc,
70:30) to give 2 (237.1 mg, 0.536 mmol, 100%, anomeric mixture, α /
 β , 3.7:1) as a colorless oil: IR (CHCl_3) 3592, 2932, 1427, 1112 cm^{-1} ;
 ^1H NMR (500 MHz, CDCl_3 , major isomer) δ_{H} 1.08 (9H, s), 1.33
(3H, s), 1.42 (3H, s), 1.70 (1H, ddd, $J = 15.0, 6.8, 3.8$ Hz, H2a), 2.23
(1H, ddd, $J = 15.1, 5.0, 4.2$ Hz, H2b), 3.83 (1H, dd, $J = 10.1, 6.6$ Hz,
H6a), 3.90 (1H, dd, $J = 10.1, 6.3$ Hz, H6b), 3.99 (1H, ddd, $J = 6.6,$
6.6, 1.9 Hz, H5), 4.24 (1H, dd, $J = 7.3, 1.9$ Hz, H4), 4.51 (1H, ddd, J
 $= 7.3, 4.2, 4.2$ Hz, H3), 5.33 (1H, dd, $J = 6.9, 5.0$ Hz, H1), 7.36–7.43
(6H, m), 7.69–7.74 (4H, m); $^{13}\text{C}\{^1\text{H}\}$ NMR (125.7 MHz, CDCl_3 , 455

596 z: $[M + Na]^+$ calcd for $C_{33}H_{43}NaO_7PSi$ 633.2413; found 633.2404.
597 Anal. calcd for $C_{33}H_{43}O_7SiP$: C, 64.90; H, 7.10. Found: C, 64.76; H,
598 7.45. Compound 6: $[\alpha]_D -2.7$ (c, 0.15, $CHCl_3$); IR ($CHCl_3$) 1729,
599 1374, 1249 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ_H 1.06 (9H, s), 1.19
600 (3H, s), 1.25 (3H, s), 1.27 (3H, t, $J = 6.9$ Hz), 2.15–2.31 (2H, m),
601 H2a, H2b) 3.76 (1H, dd, $J = 10.7$, 5.0 Hz, H6a), 3.82 (1H, dd, $J =$
602 10.7, 6.9 Hz, H6b), 3.83–3.88 (1H, m), 4.03–4.11 (1H, m), 4.36
603 (1H, dd, $J = 5.8$, 3.6 Hz, H4), 4.63 (1H, dddd, $^3J_{PH} = 6.8$ Hz, $J = 6.8$,
604 6.8, 6.8 Hz, H3), 5.40 (1H, ddd, $J = 6.9$, 5.7, 4.1 Hz, H5), 7.38–7.46
605 (6H, m), 7.48–7.51 (2H, m), 7.55–7.58 (1H, m), 7.65–7.69 (4H,
606 m), 7.80–7.84 (2H, m), 8.07 (1H, s, CHO); $^{13}C\{^1H\}$ NMR (125.7
607 MHz, $CDCl_3$, the C-*ipso* was not observed) δ_C 16.4 (CH_3 , d, $^3J_{PC} =$
608 7.4 Hz), 19.1 (C), 25.5 (CH_3), 26.4 (CH_3), 26.7 ($3 \times CH_3$), 30.8
609 (CH_2 , d, $^1J_{PC} = 100.7$ Hz, C2), 60.8 (CH_2 , d, $^2J_{PC} = 6.4$ Hz), 62.9
610 (CH_2 , C6), 71.4 (CH, C3), 71.6 (CH, C5), 75.4 (CH, d, $^3J_{PC} = 8.5$
611 Hz, C4), 108.7 (C), 127.75 ($2 \times CH$), 127.78 ($2 \times CH$), 128.5 ($2 \times$
612 CH, d, $^3J_{PC} = 12.7$ Hz), 129.8 ($2 \times CH$), 131.9 ($2 \times CH$, d, $^2J_{PC} =$
613 10.6 Hz), 132.4 (CH, d, $^4J_{PC} = 2.1$ Hz), 132.9 ($2 \times C$), 135.5 ($2 \times$
614 CH), 135.6 ($2 \times CH$), 160.5 (CH, CHO); $^{31}P\{^1H\}$ NMR (162 MHz,
615 $CDCl_3$) δ_P 41.1 (P); MS (ESI⁺-TOF) m/z (%) 633 [$(M + Na)^+$,
616 100]; HRMS (ESI⁺-TOF) m/z : $[M + Na]^+$ calcd for $C_{33}H_{43}NaO_7PSi$
617 633.2413; found 633.2400. Anal. calcd for $C_{33}H_{43}O_7PSi$: C, 64.90; H,
618 7.10. Found: C, 65.11; H, 7.32.
619 (*R*_p)-6-*O*-*tert*-Butyldiphenylsilyl-1,2-dideoxy-3,4-*O*-isopropylidene-
620 1-phenyl-1-phospha-1-oxo-*D*-lyxo-hexopyranose (38). A sol-
621 ution of 5 (8.5 mg, 0.014 mmol) in NaOEt/EtOH 0.01 M (0.12 mL,
622 1.2×10^{-3} mmol) was stirred at room temperature under nitrogen for
623 24 h. The reaction mixture was concentrated under reduced pressure,
624 and the residue was purified by Chromatotron chromatography
625 (hexanes–EtOAc, 40:60) to give compound 38 (5.2 mg, 0.01 mmol,
626 70%) as a crystalline solid: mp 134.4–135.6 °C (from *n*-hexane–
627 EtOAc); $[\alpha]_D +11.9$ (c, 0.27, $CHCl_3$); IR ($CHCl_3$) 2996, 1428, 1243,
628 1229, 1113 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$, simulated coupling
629 constants using DAISY) δ_H 1.07 (9H, s), 1.37 (3H, s), 1.39 (3H, s),
630 2.36 (1H, ddd, $^2J_{PH} = 17.8$ Hz, $J = 16.3$, 4.5 Hz, H2a), 2.59 (1H, ddd,
631 $^2J_{PH} = 7.9$ Hz, $J = 16.3$, 4.5 Hz, H2b), 3.96 (1H, dd, $J = 9.8$, 6.0 Hz,
632 H6a), 4.05 (1H, dd, $J = 9.8$, 7.6 Hz, H6b), 4.52 (1H, ddd, $^3J_{PH} = 0.8$
633 Hz, $J = 6.9$, 1.9 Hz, H4), 4.72 (1H, dddd, $^3J_{PH} = 6.3$ Hz, $J = 7.6$, 6.0,
634 1.9 Hz, H5), 4.81 (1H, dddd, $^3J_{PH} = 24.6$ Hz, $J = 6.9$, 4.5, 4.5 Hz,
635 H3), 7.34–7.47 (8H, m), 7.53–7.56 (1H, m), 7.67–7.70 (2H, m),
636 7.72–7.75 (2H, m), 7.90–7.95 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz,
637 $CDCl_3$) δ_C 19.2 (C), 24.6 (CH_3), 26.2 (CH_3), 26.7 ($3 \times CH_3$), 28.1
638 (CH_2 , d, $^1J_{PC} = 85.8$ Hz, C2), 62.6 (CH_2 , d, $^2J_{PC} = 9.5$ Hz, C6), 70.8
639 (CH, d, $^2J_{PC} = 6.4$ Hz, C3), 72.1 (CH, d, $^3J_{PC} = 4.2$ Hz, C4), 73.7
640 (CH, d, $^2J_{PC} = 5.3$ Hz, C5), 108.8 (C), 127.6 ($2 \times CH$), 127.7 ($2 \times$
641 CH), 128.3 ($2 \times CH$, d, $^3J_{PC} = 13.8$ Hz), 129.7 (CH), 129.73 (CH),
642 130.7 (C, d, $^1J_{PC} = 142.0$ Hz), 131.6 ($2 \times CH$, d, $^2J_{PC} = 10.6$ Hz),
643 132.5 (CH, d, $^4J_{PC} = 3.2$ Hz), 133.0 (C), 133.2 (C), 135.6 ($2 \times CH$),
644 135.7 ($2 \times CH$); $^{31}P\{^1H\}$ NMR (162 MHz, $CDCl_3$) δ_P 37.2 (P); MS
645 (ESI⁺-TOF) m/z (%) 559 [$(M + Na)^+$, 100]; HRMS (ESI⁺-TOF) m/z :
646 z: $[M + Na]^+$ calcd for $C_{30}H_{37}O_5PSi$ 559.2046; found 559.2057.
647 Anal. calcd for $C_{30}H_{37}O_5PSi$: C, 67.14; H, 6.95. Found: C, 67.32; H,
648 6.92.
649 (*S*_p)-6-*O*-*tert*-Butyldiphenylsilyl-1,2-dideoxy-3,4-*O*-isopropylidene-
650 1-phenyl-1-phospha-1-oxo-*D*-lyxo-hexopyranose (39). A sol-
651 ution of 6 (4.8 mg, 0.0079 mmol) in NaOEt/EtOH 0.01 M (0.07 mL,
652 0.7×10^{-3} mmol) was stirred at room temperature under nitrogen for
653 24 h. The reaction mixture was concentrated under reduced pressure,
654 and the residue was purified by Chromatotron chromatography
655 (hexanes–EtOAc, 40:60) to give 39 (3.2 mg, 6.0×10^{-3} mmol, 75%)
656 as a crystalline solid: mp 171.8–173.2 °C (from *n*-hexane–EtOAc);
657 $[\alpha]_D +10.8$ (c, 0.12, $CHCl_3$); IR ($CHCl_3$) 2997, 1229, 1113 cm^{-1} ; 1H
658 NMR (500 MHz, $CDCl_3$, simulated coupling constants using DAISY)
659 δ_H 1.08 (9H, s), 1.35 (3H, s), 1.63 (3H, s), 2.11 (1H, ddd, $^2J_{PH} = 6.6$
660 Hz, $J = 15.9$, 4.0 Hz, H2a), 2.62 (1H, ddd, $^2J_{PH} = 14.8$ Hz, $J = 15.9$,
661 4.1 Hz, H2b), 3.99 (1H, ddd, $^4J_{PH} = 2.3$ Hz, $J = 10.7$, 5.5 Hz, H6a),
662 4.07 (1H, dd, $J = 10.7$, 6.8 Hz, H6b), 4.19 (1H, dddd, $^3J_{PH} = 3.3$ Hz, J
663 = 6.8, 5.5, 1.5 Hz, H5), 4.35 (1H, ddd, $^3J_{PH} = 0.9$ Hz, $J = 7.2$, 1.5 Hz,
664 H4), 4.78 (1H, dddd, $^3J_{PH} = 24.2$ Hz, $J = 7.2$, 4.0, 4.1 Hz, H3), 7.37–
665 7.41 (4H, m), 7.43–7.47 (4H, m), 7.53–7.57 (1H, m), 7.68–7.71

(4H, m), 7.76–7.80 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, $CDCl_3$, 666
the C-*ipso* was not observed) δ_C 19.2 (C), 24.8 (CH_3), 26.2 (CH_3), 667
26.8 ($3 \times CH_3$), 27.5 (CH_2 , d, $^1J_{PC} = 85.8$ Hz, C2), 63.6 (CH_2 , d, $^3J_{PC}$
668 = 9.5 Hz, C6), 71.4 (CH, d, $^2J_{PC} = 7.4$ Hz, C3), 72.4 (CH, d, $^3J_{PC} = 6.9$
669 6.4 Hz, C4), 77.7 (CH, d, $^2J_{PC} = 6.4$ Hz, C5), 110.0 (C), 127.7 ($2 \times$
670 CH), 127.8 ($2 \times CH$), 128.79 ($2 \times CH$, d, $^3J_{PC} = 13.8$ Hz), 129.82 (2
671 $\times CH$), 130.2 ($2 \times CH$, d, $^2J_{PC} = 9.5$ Hz), 132.2 (CH, d, $^4J_{PC} = 2.1$
672 Hz), 132.9 (C), 133.0 (C), 135.6 ($4 \times CH$); $^{31}P\{^1H\}$ NMR (162
673 MHz, $CDCl_3$) δ_P 33.9 (P); MS (ESI⁺-TOF) m/z (%) 559 [$(M +$
674 $Na)^+$, 100]; HRMS (ESI⁺-TOF) m/z : $[M + Na]^+$ calcd for
675 $C_{30}H_{37}NaO_5PSi$ 559.2046; found 559.2043. Anal. calcd for
676 $C_{30}H_{37}O_5PSi$: C, 67.14; H, 6.95. Found: C, 67.03; H, 6.96.
677
678 *2-Deoxy-5,6-O-isopropylidene-3-O-methyl-D-arabino-hexofura-*
679 *nose* (8). To 1,4-anhydro-2-deoxy-3-*O*-methyl-5,6-*O*-isopropylidene-
680 *D-arabino-hex-1-enitol* 7³⁵ (4 g, 20 mmol) in THF (56 mL) and water
681 (40 mL), Hg(OAc)₂ (7 g, 22 mmol) was added at room temperature.
682 The reaction mixture was stirred at this temperature for 1 h, and
683 afterward, the solution was kept at 0 °C for 15 min. To this reaction
684 mixture, an aqueous solution of KI (15.9 g, 96 mmol in 18 mL of
685 water) was added and the mixture was stirred for another 30 min. To
686 the resulting reaction mixture at 0 °C, an aqueous solution of NaBH₄
687 (831 mg, 22 mmol in 40 mL of water) was added dropwise and
688 stirred for 1.5 h at this temperature. The insoluble part was removed
689 by filtration through a bed of Celite and washed with EtOAc. The
690 filtrate was separated, and the organic layer was extracted with EtOAc,
691 dried over Na₂SO₄, and concentrated under reduced pressure to give
692 a residue (5 g), which was purified by column chromatography
693 (hexanes–EtOAc, 60:40) to obtain 8 (1.63 g, 7.48 mmol, 37%) as a
694 crystalline mixture of anomeric isomers (6:4); IR ($CHCl_3$) 3597,
695 2938, 2938, 1228, 1069 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$, minor
696 isomer) δ 1.37 (3H, s), 1.43 (3H, s), 2.00 (1H, ddd, $J = 14.2$, 5.7, 3.5
697 Hz, H2a), 2.26 (1H, ddd, $J = 14.3$, 5.6, 1.5 Hz, H2b), 3.33 (3H, s),
698 3.94 (1H, dd, $J = 8.8$, 6.1 Hz, H6a), 4.02 (1H, m, H3), 4.05 (1H, dd, J
699 = 8.6, 6.4 Hz, H6b), 4.19 (1H, dd, $J = 6.6$, 4.1 Hz, H4), 4.32 (1H,
700 ddd, $J = 6.3$, 6.3, 6.3 Hz, H5), 5.64 (1H, ddd, $J = 5.6$, 3.0, 3.0 Hz,
701 H1); $^{13}C\{^1H\}$ NMR (125.7 MHz, $CDCl_3$, minor isomer) δ 25.3
702 (CH_3), 26.6 (CH_3), 39.7 (CH_2 , C2), 57.2 (CH_3), 66.6 (CH_2 , C6),
703 73.3 (CH, C5), 80.6 (CH, C3), 80.9 (CH, C4), 98.0 (CH, C1) 108.7
704 (C); 1H NMR (500 MHz, $CDCl_3$, major isomer) δ 1.37 (3H, s), 1.42
705 (3H, s), 1.97 (1H, ddd, $J = 13.9$, 5.0, 4.1 Hz, H2a), 2.25 (1H, br d, $J =$
706 13.9 Hz, H2b), 3.45 (3H, s), 3.71 (1H, d, $J = 11.3$ Hz, OH), 3.85
707 (1H, dd, $J = 7.7$, 3.6 Hz, H4), 3.96 (1H, m, H3), 4.02 (1H, m, H6a),
708 4.13 (1H, dd, $J = 8.7$, 6.1 Hz, H6b), 4.37 (1H, ddd, $J = 7.9$, 5.8, 5.8
709 Hz, H5), 5.36 (1H, dd, $J = 11.3$, 5.0 Hz, H1); $^{13}C\{^1H\}$ NMR (125.7
710 MHz, $CDCl_3$, major isomer) δ 25.4 (CH_3), 26.8 (CH_3), 38.6 (CH_2 ,
711 C2), 57.8 (CH_3), 67.2 (CH_2 , C6), 73.8 (CH, C5), 80.1 (CH, C3),
712 83.8 (CH, C4), 99.0 (CH, C1), 108.9 (C); MS (ESI⁺-TOF) m/z (%)
713 241 [$(M + Na)^+$, 100]; HRMS (ESI⁺-TOF) m/z [$(M + Na)^+$] calcd for
714 $C_{10}H_{18}NaO_5$ 241.1052; found 241.1051. Anal. calcd for $C_{10}H_{18}O_5$: C,
715 55.03; H, 8.31. Found: C, 54.93; H, 8.24.
716 *1-Deoxy-3-O-formyl-1-iodo-4,5-O-isopropylidene-2-O-methyl-D-*
717 *arabinol* (9). To a solution of 8 (52 mg, 0.24 mmol) in dry CH_2Cl_2
718 (3.3 mL) were added PhI(OAc)₂ (93.4 mg, 0.29 mmol) and I₂ (61
719 mg, 0.24 mmol). The reaction mixture was stirred at room
720 temperature for 1.25 h, poured into an aqueous solution of
721 Na₂S₂O₃ (10%), and extracted with CH_2Cl_2 . The organic extracts
722 were dried over Na₂SO₄ and concentrated under reduced pressure.
723 The residue was purified by Chromatotron chromatography
724 (hexanes–EtOAc, 8:2) to give 9 (70 mg, 0.20 mmol, 85%) as a
725 colorless oil: $[\alpha]_D +11.7$ (c, 0.97, $CHCl_3$); IR ($CHCl_3$) 1733, 1372,
726 1226, 1164, 1077 cm^{-1} ; 1H NMR (500 MHz, $CDCl_3$) δ 1.36 (3H, s),
727 1.43 (3H, s), 3.07 (1H, dd, $J = 10.4$, 8.2 Hz, H6a), 3.29 (1H, dd, $J =$
728 10.4, 5.7 Hz, H6b), 3.52 (3H, s), 3.72 (1H, ddd, $J = 8.0$, 5.6, 2.5 Hz,
729 H3), 3.85 (1H, dd, $J = 8.8$, 5.7 Hz, H2a), 4.02 (1H, dd, $J = 8.8$, 6.0
730 Hz, H2b), 4.27 (1H, ddd, $J = 7.6$, 6.0, 6.0 Hz, H5), 5.35 (1H, dd, $J =$
731 7.3, 2.5 Hz, H4), 8.14 (1H, s, CHO); $^{13}C\{^1H\}$ NMR (125.7 MHz,
732 $CDCl_3$) δ 1.6 (CH_2 , C2), 25.4 (CH_3), 26.7 (CH_3), 59.3 (CH_3), 66.3
733 (CH_2 , C6), 72.9 (CH, C4), 74.1 (CH, C5), 79.8 (CH, C3), 109.6
734 (C), 160.1 (CH, CHO); MS (ESI⁺-TOF) m/z (%) 367 [$(M + Na)^+$,
735 100]; HRMS (ESI⁺-TOF) m/z [$(M + Na)^+$] calcd for $C_{10}H_{17}InaO_5$ 735

736 367.0018; found 367.0018. Anal. calcd for C₁₀H₁₇IO₅: C, 34.90; H, 4.98. Found: C, 34.88; H, 5.21.

738 *1-Deoxy-1-diethoxyphosphoryl-3-O-formyl-4,5-O-isopropylidene-2-O-methyl-D-arabinitol* (**10**). A solution of **9** (311 mg, 0.90 mmol) in (EtO)₃P (8.2 mL, 48 mmol) was stirred at reflux temperature for 3 h. The mixture was concentrated under high vacuum, and the residue was purified by Chromatotron chromatography (hexanes–EtOAc, 3:7 → 2:8) to give compound **10** (248 mg, 0.7 mmol, 78%). Small quantities of alcohol **33** (14 mg, 0.043 mmol, 5%) and cyclized compounds **40** (13 mg, 0.046 mmol, 5%) and **41** (15 mg, 0.054 mmol, 6%) were also obtained (*vide infra*). Compound **10**: colorless oil, [α]_D +1.6 (c, 0.87, CHCl₃); IR (CHCl₃) 2995, 1733, 1375, 1236, 1167, 1027 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.306 (3H, t, J = 6.9 Hz), 1.308 (3H, t, J = 6.9 Hz), 1.33 (3H, s), 1.38 (3H, s), 1.91 (1H, ddd, ²J_{PH} = 18.6 Hz, J = 15.4, 6.6 Hz, H2a), 2.02 (1H, ddd, ²J_{PH} = 19.2 Hz, J = 16.1, 6.3 Hz, H2b), 3.46 (3H, s), 3.83 (1H, dd, J = 8.7, 6.1 Hz, H6a), 3.92 (1H, dddd, ³J_{PH} = 9.5 Hz, J = 6.6, 6.6, 2.2 Hz, H3), 3.99 (1H, dd, J = 8.8, 6.3 Hz, H6b), 4.08 (4H, m), 4.26 (1H, dd, J = 6.2, 6.2, 6.2 Hz, H5), 5.12 (1H, dd, J = 6.5, 2.0 Hz, H4), 8.13 (1H, s, CHO); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 16.30 (CH₃, d, ³J_{PC} = 6.4 Hz), 16.32 (CH₃, d, ³J_{PC} = 6.4 Hz), 25.4 (CH₃), 26.5 (CH₃), 27.6 (CH₂, d, ¹J_{PC} = 140.9 Hz, C2), 58.7 (CH₃), 61.7 (CH₂, d, ²J_{PC} = 6.4 Hz), 61.9 (CH₂, d, ²J_{PC} = 6.4 Hz), 66.3 (CH₂, 75.9 C6), 74.0 (CH, C5), 74.8 (CH, C4), 74.9 (CH, d, ²J_{PC} = 9.5 Hz, C3), 109.3 (C), 160.3 (CH, CHO); ³¹P{¹H} NMR (202.5 MHz, CDCl₃) δ 27.5 (P); MS (ESI⁺-TOF) m/z (%) 377 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) m/z [M + Na]⁺ calcd for C₁₄H₂₇NaO₈P 377.1341; found 377.1344. Anal. calcd for C₁₄H₂₇O₈P: C, 47.44; H, 7.68. Found: C, 47.23; H, 7.40.

765 *1-Deoxy-1-diethoxyphosphoryl-4,5-O-isopropylidene-2-O-methyl-D-arabinitol* (**33**). A solution of **10** (48 mg, 0.135 mmol) in NaOEt/EtOH 0.011 M (1.1 mL, 0.012 mmol) was stirred at room temperature under nitrogen for 1.25 h. The mixture was concentrated under reduced pressure, and the residue was purified by Chromatotron chromatography (hexanes–EtOAc, 2:98) to give alcohol **33** (34 mg, 0.104 mmol, 77%). Small quantities of cyclized compounds **40** (2.8 mg, 0.01 mmol, 7%) and **41** (1.5 mg, 0.0054 mmol, 4%) were also obtained (*vide infra*). Compound **33**: colorless oil, [α]_D −21.3 (c, 0.92, CHCl₃); IR (CHCl₃) 3389, 2986, 1373, 1231, 1160, 1072, 1032 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.33 (6H, t, J = 7.2 Hz), 1.34 (3H, s), 1.39 (3H, s), 2.14 (1H, ddd, ²J_{HP} = 19.8 Hz, J = 15.5, 5.3 Hz, H2a), 2.23 (1H, ddd, ²J_{HP} = 18.3 Hz, J = 15.4, 7.6 Hz, H2b), 2.91 (1H, br d, J = 8.0 Hz, OH), 3.44 (3H, s), 3.56 (1H, ddd, J = 7.8, 7.8, 1.6 Hz, H4), 3.83 (1H, dddd, ³J_{HP} = 12.0 Hz, J = 7.5, 5.3, 1.6 Hz, H3), 4.09 (7H, m); ¹H NMR (500 MHz, C₆D₆) δ 1.03 (3H, t, J = 6.9 Hz), 1.06 (3H, t, J = 6.9 Hz), 1.31 (3H, t), 1.44 (3H, s), 2.12 (1H, ddd, ²J_{HP} = 19.9 Hz, J = 15.1, 5.0 Hz, H2a), 2.33 (1H, ddd, ²J_{HP} = 17.8 Hz, J = 15.3, 7.9 Hz, H2b), 2.42 (1H, m, OH), 3.26 (3H, s), 3.81 (1H, br dd, J = 7.0, 7.0 Hz, H4), 3.90 (4H, m), 3.98 (1H, m, H3), 4.10 (1H, dd, J = 8.2, 6.0 Hz, H6a), 4.23 (2H, m, H5 and H6b); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 16.3 (2 × CH₃, d, ³J_{PC} = 6.4 Hz), 25.4 (CH₃), 26.81 (CH₃), 27.2 (CH₂, d, ¹J_{PC} = 137.8 Hz, C2), 58.1 (CH₃), 61.7 (CH₂, d, ²J_{PC} = 7.4 Hz, 78.9 Hz, C6), 75.2 (CH₂, d, ²J_{PC} = 6.4 Hz), 67.3 (CH₂, C6), 74.2 (CH, d, ²J_{PC} = 6.4 Hz, C4), 75.2 (CH, C3), 75.4 (CH, C5), 109.1 (C); ¹³C{¹H} NMR (125.7 MHz, C₆D₆) δ 16.4 (2 × CH₃, d, ³J_{PC} = 6.4 Hz), 25.7 (CH₃), 27.2 (CH₃), 27.6 (CH₂, d, J = 137.8 Hz, C2), 57.8 (CH₃), 79.3 (1H, CH₂, d, ²J_{PC} = 6.4 Hz), 61.6 (CH₂, d, ²J_{PC} = 6.4 Hz), 68.0 (CH₂, C6), 74.7 (CH, d, ²J_{PC} = 5.3 Hz, C4), 76.0 (CH, C5), 76.1 (CH, C3), 109.3 (C); ³¹P{¹H} NMR (162 MHz, CDCl₃) δ 29.16 (1P, s); ³¹P{¹H} NMR (162 MHz, C₆D₆) δ 42.91 (1P, s); MS (ESI⁺-TOF) m/z (%) 349 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) m/z [M + Na]⁺ calcd for C₁₃H₂₇NaO₇P 349.1392; found 349.1395. Anal. calcd for C₁₃H₂₇O₇P: C, 47.85; H, 8.34. Found: C, 47.98; H, 7.96.

800 (*S_p*-1,2-Dideoxy-1-ethoxy-5,6-O-isopropylidene-3-O-methyl-1-oxo-1-phospha-D-arabino-hexofuranose (**40**) and (*R_p*)-1,2-Dideoxy-1-ethoxy-5,6-O-isopropylidene-3-O-methyl-1-oxo-1-phospha-D-arabino-hexofuranose (**41**). A solution of **33** (29 mg, 0.089 mmol) and pyridinium *p*-toluenesulfonate (PPTS) (45 mg, 0.18 mmol) in dry CHCl₃ (0.8 mL), containing molecular sieves (4 Å

powder, 30 mg), was stirred at reflux temperature under nitrogen for 86.5 h. The mixture was poured into brine and extracted with CHCl₃, 807 and the residue was purified by Chromatotron chromatography 808 (CHCl₃–EtOAc, 30:70) to give the cyclized compounds **40** (1.8 mg, 809 0.0064 mmol, 7%, 9% brsm) and **41** (10.4 mg, 0.037 mmol, 41%, 52% 810 brsm) while the starting material **33** (6 mg, 0.018 mmol, 20%) was 811 recovered. Compound **40**: colorless oil, [α]_D = −26.8 (c, 0.96, 812 CHCl₃); IR (CHCl₃) 2935, 1373, 1250, 1221, 1077, 1044, 1013 813 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 1.33 (3H, t, J = 7.1 Hz), 1.36 814 (3H, s), 1.42 (3H, s), 1.95 (1H, ddd, ²J_{PH} = 15.9 Hz, J = 15.9, 5.3 Hz, 815 H2 α), 2.24 (1H, ddd, ²J_{PH} = 11.8 Hz, J = 15.7, 1.2 Hz, H2 β), 3.38 816 (3H, s), 4.08 (4H, m), 4.18 (2H, m), 4.46 (1H, ddd, J = 7.3, 6.3, 5.0 817 Hz); ¹H NMR (500 MHz, C₆D₆, simulated coupling constants using 818 DAISY) δ 1.02 (3H, t, J = 7.0 Hz), 1.22 (3H, s), 1.32 (1H, ddd, ²J_{PH} 819 = 15.9 Hz, J = 15.7, 5.7 Hz, H2 α), 1.36 (3H, s), 1.80 (1H, ddd, ²J_{PH} = 820 11.7 Hz, J = 15.7, 1.2 Hz, H2 β), 2.83 (3H, s), 3.45 (1H, dddd, ³J_{PH} = 821 28.7 Hz, J = 5.7, 3.3, 1.2 Hz, H3), 3.83 (1H, dd, J = 7.3, 3.3 Hz, H4), 822 3.92 (1H, dd, J = 9.0, 6.5 Hz, H6a), 3.97 (2H, m), 4.07 (1H, dd, J = 823 9.0, 5.1 Hz, H6b), 4.42 (1H, ddd, J = 7.3, 6.5, 5.1 Hz, H5); ¹³C{¹H} 824 NMR (125.7 MHz, CDCl₃) δ 16.4 (CH₃, d, ³J_{PC} = 5.3 Hz), 25.3 825 (CH₃), 26.3 (CH₂, d, ¹J_{PC} = 124.0 Hz, C2), 26.9 (CH₃), 57.2 (CH₃), 826 62.4 (CH₂, d, ²J_{PC} = 6.4 Hz), 66.6 (CH₂, C6), 73.1 (CH, d, ³J_{PC} = 827 11.7 Hz, C5), 77.6 (CH, C3), 81.6 (CH, d, ²J_{PC} = 7.4 Hz, C4), 109.4 828 (C); ¹³C{¹H} NMR (125.7 MHz, C₆D₆) δ 16.5 (CH₃, d, ³J_{PC} = 5.3 829 Hz), 25.3 (CH₃), 26.2 (CH₂, d, ¹J_{PC} = 122.9 Hz, C2), 27.0 (CH₃), 830 56.6 (CH₃), 61.9 (CH₂, d, ²J_{PC} = 6.4 Hz), 66.9 (CH₂, C6), 73.6 (CH, 831 d, ³J_{PC} = 11.7 Hz, C5), 77.9 (CH, C3), 81.7 (CH, d, ²J_{PC} = 7.4 Hz, 832 C4), 109.2 (C); ³¹P{¹H} NMR (162 MHz, CDCl₃) δ 45.02 (1P, s); 833 ³¹P{¹H} NMR (162 MHz, C₆D₆) δ 57.0 (1P, s); MS (ESI⁺-TOF) m/ 834 z (%) 303 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) m/z [M + Na]⁺ 835 calcd for C₁₁H₂₁NaO₆P 303.0973; found 303.0970. Anal. calcd for 836 C₁₁H₂₁O₆P: C, 47.14; H, 7.55. Found: C, 47.16; H, 7.24. Compound 837 **41**: colorless oil, [α]_D = −49.5 (c, 0.98, CHCl₃); IR (CHCl₃) 2934, 838 1375, 1252, 1160, 1077, 1046, 1013 cm⁻¹; ¹H NMR (500 MHz, 839 C₆D₆, simulated coupling constants using DAISY) δ 1.01 (3H, t, J = 840 7.1 Hz), 1.23 (3H, s), 1.36 (3H, s), 1.53 (1H, ddd, ²J_{PH} = 16.7 Hz, J = 841 16.0, 5.3 Hz, H2 α), 1.67 (1H, ddd, ²J_{PH} = 10.6 Hz, J = 16.0, 0.9 Hz, 842 H2 β), 2.79 (3H, s), 3.40 (1H, dddd, ³J_{PH} = 30.7 Hz, J = 5.3, 4.1, 0.9 843 Hz, H3), 3.93 (1H, dd, J = 8.8, 6.9 Hz, H6a), 3.97 (2H, m, CH₂-P), 844 3.97 (1H, dd, J = 7.0, 4.1 Hz, H4), 4.04 (1H, dd, J = 8.8, 5.3 Hz, 845 H6b), 4.32 (1H, ddd, J = 7.0, 6.9, 5.3 Hz, H5); ¹³C{¹H} NMR (125.7 846 MHz, C₆D₆) δ 16.5 (CH₃, d, ³J_{PC} = 5.3 Hz), 25.4 (CH₃), 26.1 (CH₂, 847 ¹J_{PC} = 121.9 Hz, C2), 27.0 (CH₃), 56.6 (CH₃), 62.5 (CH₂, d, ²J_{PC} 848 = 6.4 Hz), 66.9 (CH₂, C6), 73.6 (CH, d, ³J_{PC} = 10.6 Hz, C5), 78.3 849 (CH, C3), 82.1 (CH, d, ²J_{PC} = 8.5 Hz, C4), 109.3 (C); ³¹P{¹H} NMR 850 (162 MHz, CDCl₃) δ 45.4 (1P, s); ³¹P{¹H} NMR (162 MHz, C₆D₆) 851 δ 57.3 (1P, s); MS (ESI⁺-TOF) m/z (%) 303 [(M + Na)⁺, 100]; 852 HRMS (ESI⁺-TOF) m/z [M + Na]⁺ calcd for C₁₁H₂₁NaO₆P 853 303.0973; found 303.0981. Anal. calcd for C₁₁H₂₁O₆P: C, 47.14; H, 854 7.55. Found: C, 47.20; H, 7.38. 855

(*S_p*)-1-Deoxy-1-ethoxyphenylphosphoryl-3-O-formyl-4,5-O-isopropylidene-2-O-methyl-D-arabinitol (**11**) and (*R_p*)-1-Deoxy-1-ethoxyphenylphosphoryl-3-O-formyl-4,5-O-isopropylidene-2-O-methyl-D-arabinitol (**12**). A solution of **9** (61 mg, 0.177 mmol) in PhP(OEt)₂ (360 μ L, 1.87 mmol) was stirred at 150 °C for 6 h under nitrogen. The mixture was concentrated under high vacuum, and the residue was purified by Chromatotron chromatography (toluene–EtOAc, 30:70) to give a mixture of *P*-isomers (49.6 mg, 0.128 mmol, 63% 1:1, 72%), which was separated by HPLC: Nucleosil 100-5 chiral-3, 864 250 mm × 4 mm, *n*-hexane–EtOAc, 40:60, 0.65 mL/min, **12** at 25 865 min (22 mg, 0.057 mmol, 32%), **11** at 31 min (23.5 mg, 0.061 mmol, 866 34%). Compound **12**: colorless oil, [α]_D = +24.9 (c, 0.94, CHCl₃); IR 867 (CHCl₃) 3017, 1730, 1228, 1170, 1123, 1075, 1036 cm⁻¹; ¹H NMR 868 (500 MHz, CDCl₃) δ 1.28 (3H, t, J = 6.9 Hz), 1.35 (3H, s), 1.41 (3H, 869 s), 2.15 (2H, dd, ²J_{PH} = 14.8 Hz, J = 6.6 Hz, H2), 3.29 (3H, s), 3.80 870 (1H, m), 3.85 (1H, dd, J = 8.7, 6.1 Hz, H6a), 4.01 (1H, dd, J = 8.7, 871 6.1 Hz, H6b), 4.06 (2H, m), 4.27 (1H, ddd, J = 6.5, 6.5, 6.5 Hz, H5), 872 5.19 (1H, dd, J = 6.6, 2.2 Hz, H4), 7.50 (2H, m), 7.56 (1H, m), 7.78 873 (2H, m), 8.17 (1H, s, CHO); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 874 16.4 (CH₃, d, ³J_{PC} = 7.4 Hz), 25.4 (CH₃), 26.54 (CH₃), 31.7 (CH₂, d,

$^1J_{PC} = 100.7$ Hz, C2), 58.7 (CH₃), 60.9 (CH₂, d, $^2J_{PC} = 6.4$ Hz), 66.3 (CH₂, C6), 74.0 (CH, CS), 74.5 (CH, C3), 75.5 (CH, d, $^3J_{PC} = 8.5$ Hz, C4), 109.4 (C), 128.7 ($2 \times$ CH, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.5 ($2 \times$ CH, d, $^2J_{PC} = 9.5$ Hz, C-*ortho*), 132.4 (CH, d, $^4J_{PC} = 3.2$ Hz, C-*para*), 160.5 (CH, CHO), (the C-*ipso* is not observed); 1H NMR (500 MHz, C₆D₆) δ 0.96 (3H, t, $J = 7.1$ Hz), 1.24 (3H, s), 1.37 (3H, s), 2.08 (1H, ddd, $^2J_{PH} = 12.0$ Hz, $J = 15.4$, 6.9 Hz, H2a), 2.21 (1H, ddd, $^2J_{PH} = 16.7$ Hz, $J = 15.1$, 6.0 Hz, H2b), 3.10 (3H, s), 3.52 (1H, m), 3.79 (1H, dd, $J = 8.7$, 6.1 Hz, H6a), 3.84 (1H, m), 3.90 (1H, dd, $J = 8.5$, 6.0 Hz, H6b), 4.14 (1H, m, H3), 4.16 (1H, ddd, $J = 6.3$, 6.3, 6.3 Hz, H5), 5.37 (1H, dd, $J = 6.5$, 3.0 Hz, H4), 7.07 (3H, m), 7.68 (1H, s, CHO), 7.79 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, C₆D₆) δ = 16.3 (CH₃, d, $^3J_{PC} = 6.4$ Hz), 25.6 (CH₃), 26.7 (CH₃), 31.9 (CH₂, d, $^1J_{PC} = 102.8$ Hz, C2), 58.2 (CH₃), 60.6 (CH₂, d, $^2J_{PC} = 6.4$ Hz), 66.4 (CH₂, C6), 74.7 (CH, C3 or C5), 75.1 (CH, C5 or C3), 75.7 (CH, d, $^3J_{PC} = 8.5$ Hz, C4), 109.3 (C), 128.6 ($2 \times$ CH, d, $^3J_{PC} = 11.7$ Hz, C-*meta*), 131.9 ($2 \times$ CH, d, $^2J_{PC} = 9.5$ Hz, C-*ortho*), 132.0 (CH, d, $^4J_{PC} = 3.2$ Hz, C-*para*), 132.7 (C, d, $^1J_{PC} = 124.0$ Hz, C-*ipso*), 160.5 (CH, CHO); $^{31}P\{^1H\}$ NMR (162 MHz, CDCl₃) δ 40.9 (1P, s); $^{31}P\{^1H\}$ NMR (162 MHz, C₆D₆) δ 52.5 (1P, s); MS (ESI⁺-TOF) m/z (%) 409 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) m/z [M + Na]⁺ calcd for C₁₈H₂₇NaO₇P 409.1392; found 409.1392. Anal. calcd for C₁₈H₂₇O₇P: C, 55.95; H, 7.04. Found: C, 55.71; H, 7.15. Compound 11: colorless oil, [α]_D = -9.8 (c, 0.89, CHCl₃); IR (CHCl₃) 3024, 1729, 1439, 1373, 1228, 1203, 1172, 1035 cm⁻¹; 1H NMR (500 MHz, CDCl₃) δ 1.30 (3H, t, $J = 6.9$ Hz), 1.33 (3H, s), 1.34 (3H, s), 2.04 (1H, ddd, $^2J_{PH} = 12.2$ Hz, $J = 15.4$, 5.5 Hz, H2a), 2.25 (1H, ddd, $^2J_{PH} = 15.9$ Hz, $J = 15.9$, 6.9 Hz, H2b), 3.47 (3H, s), 3.80 (1H, dd, $J = 8.5$, 6.3 Hz, H6a), 3.87 (1H, m), 3.99 (1H, dd, $J = 8.8$, 6.3 Hz, H6b), 4.01 (1H, m, H3), 4.09 (1H, m), 4.26 (1H, ddd, $J = 6.3$, 6.3, 6.3 Hz, H5), 5.00 (1H, dd, $J = 6.5$, 1.7 Hz, H4), 7.50 (2H, m), 7.57 (1H, m), 7.79 (2H, m), 8.05 (1H, s, CHO); $^{13}C\{^1H\}$ NMR (125.7 MHz, CDCl₃) δ 16.4 (CH₃, d, $^3J_{PC} = 7.4$ Hz), 25.4 (CH₃), 26.5 (CH₃), 32.0 (CH₂, d, $^1J_{PC} = 100.7$ Hz, C2), 58.7 (CH₃), 61.0 (CH₂, d, $^2J_{PC} = 6.4$ Hz), 66.2 (CH₂, C6), 74.0 (CH, C5), 74.2 (CH, C3), 75.5 (CH, d, $^3J_{PC} = 8.5$ Hz, C4), 109.3 (C), 128.8 ($2 \times$ CH, d, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.5 ($2 \times$ CH, d, $^2J_{PC} = 10.6$ Hz, C-*ortho*), 132.6 (CH, d, $^4J_{PC} = 3.2$ Hz, C-*para*), 160.4 (CH, CHO) (the C-*ipso* is not observed); 1H NMR (500 MHz, C₆D₆) δ 0.97 (3H, t, $J = 7.3$ Hz), 1.22 (3H, s), 1.29 (3H, s), 2.06 (1H, ddd, $^2J_{PH} = 11.7$ Hz, $J = 15.8$, 5.7 Hz, H2a), 2.26 (1H, ddd, $^2J_{PH} = 16.1$ Hz, $J = 16.1$, 6.3 Hz, H2b), 3.29 (3H, s), 3.57 (1H, m), 3.76 (1H, dd, $J = 8.5$, 6.6 Hz, H6a), 3.84 (1H, dd, $J = 8.2$, 6.6 Hz, H6b), 3.90 (1H, m), 4.05 (1H, m, H3), 4.15 (1H, ddd, $J = 6.2$, 6.2, 6.2 Hz, H5), 5.16 (1H, dd, $J = 6.0$, 2.5 Hz, H4), 7.07 (3H, m), 7.57 (2H, s, CHO), 7.78 (2H, m); 1H NMR (500 MHz, C₆D₆, 60 °C, simulated coupling constants using DAISY) δ 1.01 (3H, t, $J = 6.9$ Hz), 1.23 (3H, s), 1.29 (3H, s), 2.10 (1H, ddd, $^2J_{PH} = 11.5$ Hz, $J = 15.3$, 6.0 Hz, H2a), 2.25 (1H, ddd, $^2J_{PH} = 17.2$ Hz, $J = 15.3$, 6.3 Hz, H2b), 3.30 (3H, s), 3.65 (1H, m), 3.79 (1H, dd, $J = 8.6$, 6.3 Hz, H6a), 3.83 (1H, dd, $J = 8.6$, 6.3 Hz, H6b), 3.92 (1H, m), 4.03 (1H, ddd, $^3J_{PH} = 11.1$ Hz, $J = 6.3$, 6.0, 3.1 Hz, H3), 4.17 (1H, ddd, $J = 6.3$, 6.3, 6.2 Hz, H5), 5.15 (1H, dd, $J = 6.3$, 3.1 Hz, H4), 7.10 (3H, m), 7.64 (1H, s, CHO), 7.78 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, C₆D₆) δ 16.4 (CH₃, d, $^3J_{PC} = 6.4$ Hz), 25.6 (CH₃), 26.6 (CH₃), 32.1 (CH₂, d, $^1J_{PC} = 98.5$ Hz, C2), 58.1 (CH₃), 60.6 (CH₂, d, $^2J_{PC} = 7.4$ Hz), 66.4 (CH₂, C6), 74.7 (CH, C5), 74.8 (CH, C3), 75.7 (CH, d, $^3J_{PC} = 8.5$ Hz, C4), 109.3 (C), 128.8 ($2 \times$ CH, d, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.9 ($2 \times$ CH, d, $^2J_{PC} = 9.5$ Hz, C-*ortho*), 132.2 (CH, br. s., C-*para*), 160.5 (CH, CHO), (the C-*ipso* is not observed); $^{31}P\{^1H\}$ NMR (162 MHz, CDCl₃) δ 40.5 (1P, s); $^{31}P\{^1H\}$ NMR (162 MHz, C₆D₆) δ 52.3 (1P, s); MS (ESI⁺-TOF) m/z (%) 409 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) m/z [M + Na]⁺ calcd for C₁₈H₂₇NaO₇P 409.1392; found 409.1399. Anal. calcd for C₁₈H₂₇O₇P: C, 55.95; H, 7.04. Found: C, 55.83; H, 7.15.

(S_p)-1-Deoxy-1-ethoxyphenylphosphoryl-4,5-O-isopropylidene-2-O-methyl-D-arabinitol (34). A solution of 11 (7.5 mg, 0.019 mmol) in NaOEt/EtOH 0.011 M (160 μ L, 1.65×10^{-3} mmol) was stirred at room temperature under nitrogen for 1.25 h. The organic extracts were concentrated under reduced pressure, and the residue was

purified by purified by Chromatotron chromatography (CHCl₃-EtOAc, 30:70) to give alcohol 34 (3.2 mg, 0.009 mmol, 46%), and a small amount of the cyclized compound 42 (0.5 mg, 0.0016 mmol, 8%) is also obtained (vide infra). Compound 34: colorless oil, 1H NMR (500 MHz, CDCl₃) δ 1.30 (3H, t, $J = 7.1$ Hz), 1.34 (3H, s), 1.38 (3H, s), 2.33-2-38 (2H, m, H2), 3.29 (3H, s), 3.42 (1H, d, $J = 6.9$ Hz, OH), 3.56 (1H, ddd, $J = 7.6$, 7.6, 1.6 Hz, H4), 3.82-3.87 (1H, m), 3.87-3.93 (1H, m, H3), 3.96-4.01 (1H, m, H6a), 4.05-4.14 (3H, m, H5, H6b, P-O-CH₂), 7.45-7.53 (2H, m), 7.54-7.60 (1H, m), 7.77-7.85 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, CDCl₃) δ 16.4 (CH₃, d, $^3J_{PC} = 6.4$ Hz), 25.5 (CH₃), 26.9 (CH₃), 31.7 (CH₂, d, $J = 98.5$ Hz, C2), 57.9 (CH₃), 60.9 (CH₂, d, $^2J_{PC} = 6.4$ Hz), 67.5 (CH₂, C6), 74.7 (CH, d, $J = 4.2$ Hz, C4), 75.3 (CH), 75.4 (CH), 109.1 (C), 128.7 ($2 \times$ CH, d, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.6 ($2 \times$ CH, d, $^2J_{PC} = 9.5$ Hz, C-*ortho*), 132.5 (CH, d, $^4J_{PC} = 4.2$ Hz, C-*para*), (the C-*ipso* is not observed); 1H NMR (500 MHz, C₆D₆) δ 0.96 (3H, t, $J = 7.1$ Hz), 1.31 (3H, s), 1.44 (3H, s), 2.22 (1H, ddd, $^2J_{PH} = 17.0$ Hz, $J = 15.4$, 4.4 Hz, H2a), 2.32 (1H, ddd, $^2J_{PH} = 11.3$ Hz, $J = 15.4$, 8.2 Hz, H2b), 3.08 (3H, s), 3.48-3.57 (1H, m), 3.75 (1H, br dd, $J = 6.9$, 6.9 Hz, H4), 3.82-3.88 (1H, m), 3.90 (1H, br d, $J = 6.9$ Hz, OH), 4.04 (1H, dddd, $^3J_{PH} = 12.6$ Hz, $J = 7.9$, 4.4, 1.6 Hz, H3), 4.08 (1H, dd, $J = 8.2$, 6.0 Hz, H6a), 4.19 (1H, dd, $J = 8.5$, 5.0 Hz, H6b), 4.24 (1H, ddd, $J = 8.4$, 5.8, 5.8 Hz, H5), 6.98-7.09 (3H, m), 7.70-7.77 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, C₆D₆) δ 16.3 (CH₃, d, $^3J_{PC} = 5.3$ Hz), 25.6 (CH₃), 27.2 (CH₃), 31.8 (CH₂, d, $^1J_{PC} = 95.4$ Hz, C2), 57.4 (CH₃), 60.5 (CH₂, d, $^2J_{PC} = 5.3$ Hz), 68.0 (CH₂, C6), 75.3 (CH, d, $^3J_{PC} = 4.2$ Hz, C4), 75.9 (CH), 76.0 (CH), 109.1 (C), 128.5 ($2 \times$ CH, d, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.9 ($2 \times$ CH, d, $^2J_{PC} = 9.5$ Hz, C-*ortho*), 132.0 (CH, d, $^4J_{PC} = 3.2$ Hz, C-*para*), (the C-*ipso* is not observed); $^{31}P\{^1H\}$ NMR (162 MHz, CDCl₃) δ 42.6 (1P, s); $^{31}P\{^1H\}$ NMR (202.5 MHz, C₆D₆) δ 54.9 (1P, s); MS (ESI⁺-TOF) m/z (%) 381 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) m/z [M + Na]⁺ calcd for C₁₇H₂₇NaO₆P 381.1443; found 381.1443.

(R_p)-1-Deoxy-1-ethoxyphenylphosphoryl-4,5-O-isopropylidene-2-O-methyl-D-arabinitol (35). A solution of 12 (6.8 mg, 0.018 mmol) in NaOEt/EtOH 0.011 M (150 μ L, 1.65×10^{-3} mmol) was stirred at room temperature under nitrogen for 1.25 h. The organic extracts were concentrated under reduced pressure, and the residue was purified by Chromatotron chromatography (hexanes-EtOAc, 5:95) to give alcohol 35 (3.3 mg, 0.0092 mmol, 52%), and a small amount of the cyclized compound 43 (0.5 mg, 0.0016 mmol, 9%) is also obtained (vide infra). Compound 35: colorless oil, 1H NMR (500 MHz, CDCl₃) δ 1.29 (3H, t, $J = 7.1$ Hz), 1.33 (3H, s), 1.35 (3H, s), 2.26 (1H, ddd, $^2J_{PH} = 12.5$ Hz, $J = 15.4$, 5.5 Hz, H2a), 2.44 (1H, ddd, $^2J_{PH} = 17.5$ Hz, $J = 15.3$, 7.6 Hz, H2b), 2.99 (1H, br d, $J = 7.9$ Hz, OH), 3.32 (3H, s), 3.58 (1H, br dd, $J = 7.9$, 7.9 Hz, H4), 3.84 (2H, m, H3, P-O-CH₂), 4.00 (1H, m, H6), 4.08 (3H, m, H6, P-O-CH₂, C5), 7.51 (2H, m), 7.57 (1H, m), 7.80 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, CDCl₃) δ 16.4 (CH₃, d, $^3J_{PC} = 6.4$ Hz), 25.5 (CH₃), 26.9 (CH₃), 31.4 (CH₂, d, $^1J_{PC} = 99.6$ Hz, C2), 58.1 (CH₃), 60.8 (CH₂, d, $^2J_{PC} = 6.4$ Hz), 67.4 (CH₂, C6), 74.6 (CH, d, $^3J_{PC} = 5.3$ Hz, C4), 75.0 (CH, C3), 75.5 (CH, C5), 109.2 (C), 128.7 ($2 \times$ CH, d, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.5 ($2 \times$ CH, d, $^2J_{PC} = 10.6$ Hz, C-*ortho*), 132.4 (CH, d, $^4J_{PC} = 3.2$ Hz, C-*para*) (the C-*ipso* is not observed); 1H NMR (500 MHz, C₆D₆) δ 0.93 (3H, t, $J = 7.1$ Hz), 1.28 (3H, s), 1.40 (3H, s), 2.11 (1H, ddd, $^2J_{PH} = 12.3$ Hz, $J = 15.1$, 5.0 Hz, H2a), 2.48 (1H, ddd, $^2J_{PH} = 17.8$ Hz, $J = 15.1$, 8.0 Hz, H2b), 3.12 (3H, s), 3.40 (1H, br d, $J = 7.3$ Hz, OH), 3.56 (1H, ddq, $^2J_{PH} = 8.2$ Hz, $J = 10.1$, 6.9, 6.9 Hz), 3.79 (1H, br dd, $J = 7.1$, 7.1 Hz, H4), 3.85 (1H, ddq, $^2J_{PH} = 7.9$ Hz, $J = 10.1$, 6.9, 6.9 Hz), 3.94 (1H, dddd, $^3J_{PH} = 10.8$ Hz, $J = 7.9$, 4.9, 1.9 Hz, H3), 4.04 (1H, dd, $J = 8.0$, 5.5 Hz, H6a), 4.16 (1H, ddd, $J = 8.2$, 5.4, 5.4 Hz, H5), 4.19 (1H, dd, $J = 7.9$, 4.7 Hz, H6b), 7.08 (3H, m), 7.76 (2H, m); $^{13}C\{^1H\}$ NMR (125.7 MHz, C₆D₆) δ = 16.4 (CH₃, d, $^3J_{PC} = 6.4$ Hz), 25.7 (CH₃), 27.2 (CH₃), 31.6 (CH₂, d, $^1J_{PC} = 99.6$ Hz, C2), 57.7 (CH₃), 60.6 (CH₂, d, $^2J_{PC} = 6.4$ Hz), 68.0 (CH₂, C6), 75.1 (CH, d, $^3J_{PC} = 4.2$ Hz, C4), 75.8 (CH), 76.0 (CH), 109.3 (C), 128.7 ($2 \times$ CH, d, $^3J_{PC} = 12.7$ Hz, C-*meta*), 131.8 ($2 \times$ CH, d, $^2J_{PC} = 9.5$ Hz, C-*ortho*), 132.1 (CH, d, $^4J_{PC} = 2.1$ Hz, C-*para*) (the C-*ipso* is not observed); $^{31}P\{^1H\}$ NMR (162 MHz,

HRMS (ESI⁺-TOF) *m/z*: [M - H]⁺ calcd for C₁₄H₁₈O₆P 313.0841; found 313.0838. Anal. calcd for C₁₄H₁₈O₆P: C, 53.51; H, 6.09. Found: C, 53.27; H, 6.18.

(S_p)-4-Deoxy-3-O-formyl-1,2-O-isopropylidene-5-phenyl-5-phospha-5-oxo-α-D-erythro-pentopyranose (**48**) and (R_p)-4-Deoxy-3-O-formyl-1,2-O-isopropylidene-5-phenyl-5-phospha-5-oxo-α-D-erythro-pentopyranose (**49**). A solution of the crude alcohol **24** (42.4 mg, 0.135 mmol) in dry CH₂Cl₂ (7.4 mL) containing PhI(OAc)₂ (96.6 mg, 0.3 mmol) and I₂ (41.1 mg, 0.152 mmol) under nitrogen was stirred and irradiated with two 80 W tungsten-filament lamps at room temperature for 3.5 h. The reaction mixture was then poured into 10% aqueous Na₂S₂O₃, extracted with CH₂Cl₂, dried over Na₂SO₄, and concentrated. The residue was purified by Chromatotron chromatography (hexanes–EtOAc, 20:80) to give compounds **48** (12.2 mg, 0.039 mmol, 29%) and **49** (21.4 mg, 0.069 mmol, 51%). Compound **48**: crystalline solid, mp 166.2–167.4 °C (from *n*-hexane–EtOAc); IR (CHCl₃) 1729, 1229, 1139, 1036 cm⁻¹; [α]_D -12.7 (c 0.67, CHCl₃); ¹H NMR (500 MHz, CDCl₃, simulated coupling constants using DAISY) δ_H 1.46 (3H, s), 1.85 (3H, s), 2.58 (1H, dddd, ²J_{PH} = 10.1 Hz, J = 14.6, 5.1, ⁴J_w = 0.9 Hz, H2α), 2.79 (1H, ddd, ²J_{PH} = 21.3 Hz, J = 14.6, 12.6 Hz, H2β), 4.54 (1H, ddd, J = 3.8, 1.8 Hz, ⁴J_w = 0.9 Hz, H4), 5.39 (1H, dddd, ³J_{PH} = 2.0 Hz, J = 12.6, 5.1, 1.8 Hz, ⁴J_w = 1.1 Hz, H3) [Decoupling by selective irradiation at CHO (8.08 ppm) (1H, dddd, ³J_{PH} = 1.8 Hz, J = 12.2, 4.9, 1.8 Hz, H3)], 5.92 (1H, dd, ³J_{PH} = 19.0 Hz, J = 3.8 Hz, H5), 7.48–7.55 (2H, m), 7.56–7.62 (1H, m), 7.73–7.80 (2H, m), 8.08 (1H, dd, ⁵J_{PH} = 2.2 Hz, ⁴J_w = 1.1 Hz, CHO) [Decoupling by selective irradiation at H3 (5.39 ppm) (1H, d, ⁵J_{PH} = 2.2 Hz, H3)]; ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 26.0 (CH₃), 26.2 (CH₃), 26.9 (CH₂, d, ¹J = 80.5 Hz, C2), 66.3 (CH, d, ²J = 4.2 Hz, C3), 75.1 (CH, d, ³J = 5.3 Hz, C4), 99.4 (CH, d, ²J = 10.6 Hz, C5), 114.2 (C), 128.9 (2 × CH, d, ³J = 12.7 Hz), 130.3 (2 × CH, d, ²J = 11.7 Hz), 131.6 (C, d, ¹J = 142.0 Hz), 132.8 (CH, d, ⁴J = 2.1 Hz), 159.7 (C, C1); ³¹P{¹H} NMR (202.5 MHz, CDCl₃) δ_P 30.9 (P); MS (ESI⁺-TOF) *m/z* (%) 335 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) *m/z*: [M + Na]⁺ calcd for C₁₄H₁₇NaO₆P 335.0660; found 335.0658. Anal. calcd for C₁₄H₁₇O₆P: C, 53.85; H, 5.49. Found: C, 53.69; H, 5.68. Compound **49**: colorless oil, [α]_D -30.2 (c 0.61, CHCl₃); IR (CHCl₃) 1731, 1243, 1229, 1146, 1032 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, simulated coupling constants using DAISY) δ_H 1.49 (3H, s), 1.70 (3H, s), 2.37 (1H, dddd, ²J_{PH} = 12.9 Hz, J = 13.0, 4.2, ⁴J_w = 0.7 Hz, H2α), 2.50 (1H, ddd, ²J_{PH} = 10.0 Hz, J = 13.0, 12.8 Hz, H2β), 4.53 (1H, dd, J = 2.9, 2.3 Hz, ⁴J_w = 0.7 Hz, H4), 6.02 (1H, dddd, ³J_{PH} = 3.3 Hz, J = 12.8, 4.2, 2.9 Hz, ⁴J_w = 1.1 Hz, H3) [Decoupling by selective irradiation at CHO (8.10 ppm) (1H, dddd, ³J_{PH} = 3.2 Hz, J = 12.9, 4.1, 3.2 Hz, H3)], 6.04 (1H, br d, ³J_{PH} = 0.7 Hz, J = 2.3 Hz, H5), 7.50–7.55 (2H, m), 7.60–7.65 (1H, m), 7.80–7.86 (2H, m), 8.10 (1H, dd, ⁵J_{PH} = 2.1 Hz, ⁴J_w = 1.1 Hz, CHO) [Decoupling by selective irradiation at H3 (6.02 ppm) (1H, d, ⁵J_{PH} = 2.2 Hz, CHO)]; ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 25.6 (CH₃), 26.4 (CH₂, d, ¹J_{PC} = 83.7 Hz, C2), 27.9 (CH₃), 65.3 (CH, d, ²J_{PC} = 2.1 Hz, C3), 75.9 (CH, d, ⁴J_{PC} = 3.2 Hz, C4), 97.5 (CH, d, ²J_{PC} = 4.2 Hz, C5), 113.1 (C), 128.77 (2 × CH, d, ³J_{PC} = 13.8 Hz), 128.84 (C, d, ¹J_{PC} = 143.1 Hz), 131.1 (2 × CH, d, ²J_{PC} = 10.6 Hz), 133.3 (CH, d, ⁴J_{PC} = 3.2 Hz), 159.3 (C, C1); ³¹P{¹H} NMR (202.5 MHz, CDCl₃) δ_P 34.1 (P); MS (ESI⁺-TOF) *m/z* (%) 335 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) *m/z*: [M + Na]⁺ calcd for C₁₄H₁₇NaO₆P 335.0660; found 335.0656. Anal. calcd for C₁₄H₁₇O₆P: C, 53.85; H, 5.49. Found: C, 54.09; H, 5.76.

1-O-Benzoyl-6-O-tert-butylidiphenylsilyl-2,3-O-isopropylidene-α-D-mannofuranose (**26**). To a solution of 1-O-benzoyl-2,3-O-isopropylidene-α-D-mannofuranose **25**³⁹ (188 mg, 0.58 mmol) in dry DMF (2 mL) were added imidazole (79 mg, 1.16 mmol), DMAP (1 mg, 0.008 mmol), and TBDPSiCl (200 μL, 0.769 mmol) at room temperature under nitrogen. The mixture was then stirred for 2 h, poured into water, and extracted with CH₂Cl₂. The organic extracts were concentrated under reduced pressure, and the residue was purified by Chromatotron chromatography (hexanes–EtOAc, 85:15) to give **26** (300 mg, 0.532 mmol, 92%) as a colorless oil: [α]_D +8.1 (c 1.54, CHCl₃); IR (CHCl₃) 3564, 2934, 2858, 1728, 1427, 1259, 1091 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ_H 0.99 (9H, s), 1.39 (3H, s),

1.51 (3H, s), 2.79 (1H, d, J = 6.9 Hz, OH), 3.89 (2H, m, H6), 4.10 (1H, dddd, J = 7.4, 7.4, 3.7, 3.7 Hz, H5), 4.34 (1H, dd, J = 8.5, 3.5 Hz, H4), 4.88 (1H, d, J = 5.7 Hz, H2), 5.03 (1H, dd, J = 5.7, 3.5 Hz, H3), 6.43 (1H, s, H1), 7.31–7.46 (8H, m), 7.53–7.58 (1H, m), 7.59–7.69 (4H, m), 7.97–8.04 (2H, m); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ_C 19.2 (C), 24.8 (CH₃), 26.1 (CH₃), 26.7 (3 × CH₃), 64.9 (CH₂, C6), 69.4 (CH, C5), 79.9 (CH, C3), 81.0 (CH, C4), 85.0 (CH, C2), 101.4 (CH, C1), 113.2 (C), 127.7 (4 × CH), 128.4 (2 × CH), 129.5 (C), 129.7 (2 × CH), 129.8 (2 × CH), 132.7 (C), 133.0 (C), 133.4 (CH), 135.5 (2 × CH), 135.5 (2 × CH), 164.9 (C); MS (ESI⁺-TOF) *m/z* (%) 585 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) *m/z* [M + Na]⁺ calcd for C₃₂H₃₈NaO₇Si 585.2285; found 585.2290. Anal. calcd for C₃₂H₃₈O₇Si: C, 68.30; H, 6.81. Found: C, 68.21; H, 6.85.

1-O-Benzoyl-6-O-tert-butylidiphenylsilyl-2,3-O-isopropylidene-α-D-lyxo-hexofuranose-5-ulose (**27**). To a stirred solution of **26** (3 g, 5.3 mmol) in dry CH₂Cl₂ (160 mL) at room temperature were added MS 3 Å (5 g) and PCC (3 g, 13.9 mmol). After 9 h, isopropanol (2 mL) was added and the mixture diluted with Et₂O and filtered through Celite. The filtrate was concentrated under reduced pressure, and the residue was purified by silica gel column chromatography (hexanes–EtOAc, 90:10 → 85:15) to give **27** (2.68 g, 4.8 mmol, 91%) as a colorless oil: [α]_D -18.8 (c 1.22, CHCl₃); IR (CHCl₃) 2860, 1737, 1428, 1260, 1105 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.13 (9H, s), 1.31 (6H, s), 4.52 (2H, s, H6), 4.86 (1H, d, J = 5.7 Hz, H2), 4.99 (1H, d, J = 4.1 Hz, H4), 5.30 (1H, dd, J = 5.7, 4.1 Hz, H3), 6.48 (1H, s, H1), 7.36–7.48 (8H, m), 7.57–7.62 (1H, m), 7.65–7.73 (4H, m), 7.98–8.00 (2H, m); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 19.3 (C), 24.6 (CH₃), 25.7 (CH₃), 26.8 (3 × CH₃), 69.0 (CH₂, C6), 80.3 (CH, C3), 84.3 (CH, C2), 86.1 (CH, C4), 101.1 (CH, C1), 113.7 (C), 127.77 (2 × CH), 127.8 (2 × CH), 128.5 (2 × CH), 129.2 (C), 129.7 (2 × CH), 129.86 (CH), 129.93 (CH), 132.5 (C), 132.6 (C), 133.6 (CH), 135.48 (2 × CH), 135.51 (2 × CH), 164.6 (C), 202.3 (C, C5); MS (ESI⁺-TOF) *m/z* (%) 583 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) *m/z* [M + Na]⁺ calcd for C₃₂H₃₆NaO₇Si 583.2128; found 583.2128. Anal. calcd for C₃₂H₃₆O₇Si: C, 68.55; H, 6.47. Found: C, 68.70; H, 6.45.

1-O-Benzoyl-6-O-tert-butylidiphenylsilyl-5-C-dimethoxyphosphoryl-2,3-O-isopropylidene-α-D-mannofuranose (**28R**) and 1-O-Benzoyl-6-O-tert-butylidiphenylsilyl-5-C-dimethoxyphosphoryl-2,3-O-isopropylidene-β-L-gulofuranose (**28S**). To a solution of **27** (314 mg, 0.56 mmol) in (MeO)₂P(O)H (2.3 mL, 25.1 mmol) was added DBU (100 μL, 0.67 mmol) at 0 °C under nitrogen, and the mixture was stirred for 30 min at this temperature. An aqueous saturated solution of NH₄Cl (0.3 mL) was then added, and the stirring was continued for 4 h. The mixture was then extracted with CHCl₃, washed with water, and the organic extract was concentrated under reduced pressure. The residue was purified by silica gel column chromatography (hexanes–EtOAc, 50:50) to give **28R** (42 mg, 0.063 mmol, 11%) and **28S** (260 mg, 0.388 mmol, 69%). Compound **28R**: Colorless oil, [α]_D +36.5 (c 1.23, CHCl₃); IR (CHCl₃) 3469, 3001, 2959, 1729, 1428, 1260, 1059 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 1.11 (9H, s), 1.22 (3H, s), 1.45 (3H, s), 3.74 (3H, d, ³J_{PH} = 10.7 Hz), 3.80 (3H, d, ³J_{PH} = 10.7 Hz), 4.04 (1H, dd, ³J_{PH} = 18.6 Hz, J = 10.7 Hz, H6a), 4.20 (1H, dd, ³J_{PH} = 10.4 Hz, J = 10.4 Hz, H6b), 4.21 (1H, d, ³J_{PH} = 20.8 Hz, OH), 4.65 (1H, dd, ³J_{PH} = 3.6 Hz, J = 3.6 Hz, H4), 4.78 (1H, d, J = 6.0 Hz, H2), 4.86 (1H, dd, J = 5.7, 3.2 Hz, H3), 6.44 (1H, s, H1), 7.36–7.47 (8H, m), 7.57–7.60 (1H, s), 7.67–7.76 (4H, m), 7.99–8.01 (2H, m); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 19.3 (C), 24.3 (CH₃), 25.8 (CH₃), 26.8 (3 × CH₃), 53.3 (CH₃, d, ²J_{PC} = 7.4 Hz), 54.0 (CH₃, d, ²J_{PC} = 7.4 Hz), 66.1 (CH₂, d, ²J_{PC} = 2.1 Hz, C6), 76.7 (C, d, ¹J_{PC} = 167.4 Hz, C5), 80.1 (CH), 81.1 (CH, d, ¹J_{PC} = 10.6 Hz, C3), 85.0 (CH, C2), 100.6 (CH, C1), 113.5 (C), 127.7 (4 × CH), 128.5 (2 × CH), 129.4 (C), 129.7 (2 × CH), 129.85 (CH), 129.87 (CH), 132.7 (C), 132.8 (C), 133.5 (CH), 135.7 (2 × CH), 135.8 (2 × CH), 164.9 (C); ³¹P{¹H} NMR (202.5 MHz, CDCl₃) δ 22.7 (P); MS (ESI⁺-TOF) *m/z* (%) 693 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) *m/z* [M + Na]⁺ calcd for C₃₄H₄₃O₁₀NaPSi 693.2261; found 693.2264. Anal. calcd for C₃₄H₄₃O₁₀PSi: C, 60.88; H, 6.46. Found: C, 60.58; H, 6.54. Compound **28S**: Crystalline solid, mp 140.2–141.3 °C (from CH₂Cl₂–*n*-hexane); [α]_D +16.6 (c 1.23,

1573 CHCl₃; IR (CHCl₃) 3460, 3001, 2957, 1729, 1452, 1428, 1248, 1574 1090 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ 0.88 (9H, s), 1.40 (3H, 1575 s), 1.56 (3H, s), 3.78 (3H, d, ³J_{PH} = 10.4 Hz), 3.84 (3H, d, ³J_{PH} = 10.4 1576 Hz), 3.88 (1H, dd, ³J_{PH} = 10.1 Hz, J = 10.1 Hz, H6a), 4.03 (1H, dd, 1577 ³J_{PH} = 30.3 Hz, J = 9.8 Hz, H6b), 4.61 (1H, br s, OH), 4.90 (1H, d, J = 1578 = 2.8 Hz, H4), 4.97 (1H, d, J = 5.7 Hz, H2), 5.27 (1H, dd, J = 5.7, 2.8 1579 Hz, H3), 6.55 (1H, d, ³J_{PH} = 2.2 Hz, H1), 7.29–7.42 (8H, m), 7.47– 1580 7.53 (1H, m), 7.59–7.61 (2H, m), 7.70–7.74 (2H, m), 7.92–7.97 1581 (2H, m); ¹³C{¹H} NMR (125.7 MHz, CDCl₃) δ 19.0 (C), 24.7 1582 (CH₃), 26.0 (CH₃), 26.4 (3 × CH₃), 53.3 (CH₃, d, ²J_{PC} = 7.4 Hz), 1583 53.7 (CH₃, d, ²J_{PC} = 8.5 Hz), 64.3 (CH₂, d, ²J_{PC} = 2.1 Hz, C6), 76.8 1584 (C, d, ¹J_{PC} = 154.7 Hz, C5), 77.4 (CH, ²J_{PC} = 21.8 Hz, C4), 82.0 1585 (CH, C3), 86.2 (CH, C2), 100.7 (CH, C1), 113.7 (C), 127.55 (2 × 1586 CH), 127.63 (2 × CH), 128.3 (2 × CH), 129.3 (C), 129.57 (CH), 1587 129.60 (CH), 129.8 (2 × CH), 132.4 (C), 132.8 (C), 133.3 (CH), 1588 135.5 (2 × CH), 135.7 (2 × CH), 164.9 (C); ³¹P{¹H} NMR (202.5 1589 MHz, CDCl₃) δ 24.1 (P); MS (ESI⁺-TOF) *m/z* (%) 693 [(M + Na)⁺, 1590 100]; HRMS (ESI⁺-TOF) *m/z* [M + Na]⁺ calcd for C₃₄H₄₃O₁₀NaPSi 1591 693.2261; found 693.2255. Anal. calcd for C₃₄H₄₃O₁₀PSi: C, 60.88; 1592 H, 6.46. Found: C, 60.85; H, 6.41.

1593 **5-Deoxy-5-dimethoxyphosphoryl-1-O-benzoyl-6-O-tert-butylidiphenylsilyl-2,3-O-1-isopropylidene-β-L-gulofuranose (29) and 5-1595 Deoxy-5-dimethoxyphosphoryl-1-O-benzoyl-6-O-tert-butylidiphenylsilyl-2,3-O-1-isopropylidene-α-D-mannofuranose (30).** Methyl 1596 oxalyl chloride (110 μL, 1.2 mmol) was added to a solution of **28S** 1597 (161 mg, 0.24 mmol) and DMAP (146.6 mg, 1.2 mmol) in dry 1598 MeCN (2 mL) at 0 °C, and the mixture was stirred at this 1600 temperature for 30 min under nitrogen. Additional amounts of MeCN 1601 (1 mL), methyl oxalyl chloride (55 μL, 0.6 mmol), and DMAP (73.3 1602 mg, 0.6 mmol) were then added, and the stirring was continued for 30 1603 min. Most of the solvent was distilled off in vacuo, and the residue was 1604 dissolved with CHCl₃. The organic layer was washed with water, 1605 saturated NaHCO₃ solution and brine, dried with Na₂SO₄, and 1606 evaporated under reduced pressure to give methoxalyl derivatives as a 1607 pale yellow syrup. The crude was coevaporated with dry toluene 1608 under reduced pressure. A solution of the residue in toluene (2 mL) 1609 was treated with tributyltin hydride (97 μL, 0.36 mmol) and AIBN 1610 (5.9 mg, 0.036 mmol), and the mixture was stirred at reflux 1611 temperature for 2 h under nitrogen. Additional AIBN (5.9 mg, 0.036 1612 mmol) was added every 40 min. The mixture was then concentrated 1613 under reduced pressure, and the residue was partitioned between 1614 MeCN and hexane. The MeCN layer was extracted three times with 1615 hexane and then concentrated under reduced pressure. The residue 1616 was purified first by silica gel column chromatography (hexanes→ 1617 hexanes–EtOAc, 6:4) and then by Chromatotron chromatography 1618 (hexanes–EtOAc, 70:30) to give **29** (110 mg, 0.168 mmol, 70%) and 1619 **30** (28 mg, 0.043 mmol, 18%). Compound **29**: colorless oil, [α]_D 1620 +40.6 (c, 1.18, CHCl₃); IR (CHCl₃) 2957, 2859, 1728, 1428, 1261, 1621 1105, 1063 cm⁻¹; ¹H NMR (500 MHz, CDCl₃) δ ppm 1.10 (9H, s), 1622 1.21 (3H, s), 1.41 (3H, s), 2.63 (1H, dddd, ²J_{PH} = 18.0 Hz, J = 10.1, 1623 3.8, 3.8 Hz, H5), 3.70 (3H, d, ³J_{PH} = 11.0 Hz), 3.73 (3H, d, ³J_{PH} = 1624 11.0 Hz), 4.01 (1H, ddd, ³J_{PH} = 26.8 Hz, J = 10.4, 3.8 Hz, H6a), 4.21 1625 (1H, ddd, ³J_{PH} = 11.0 Hz, J = 11.0, 3.9 Hz, H6b), 4.48 (1H, dd, J = 1626 5.4, 3.5 Hz, H3), 4.61 (1H, ddd, ³J_{PH} = 6.7 Hz, J = 10.0, 3.2 Hz, H4), 1627 4.71 (1H, d, J = 5.7 Hz, H2), 6.37 (1H, s, H1), 7.35–7.48 (9 H, m), 1628 7.67–7.74 (4 H, m), 7.99–8.01 (2H, m); ¹³C{¹H} NMR (125.7 1629 MHz, CDCl₃) δ 19.3 (C), 24.8 (CH₃), 26.0 (CH₃), 26.8 (3 × CH₃), 1630 39.5 (CH, d, ¹J_{PC} = 143.1 Hz, C5), 52.4 (CH₃, d, ²J_{PC} = 6.4 Hz), 52.9 1631 (CH₃, d, ²J_{PC} = 7.4 Hz), 61.1 (CH₂, d, ²J_{PC} = 7.4 Hz, C6), 79.4 (CH, 1632 d, ³J_{PC} = 11.7 Hz, C3), 79.8 (CH, d, ²J_{PC} = 4.2 Hz, C4), 84.7 (CH, 1633 C2), 101.2 (CH, C1), 112.9 (C), 127.67 (2 × CH), 127.71 (2 × 1634 CH), 128.4 (2 × CH), 129.7 (2 × CH), 129.76 (CH), 129.77 (CH), 1635 133.0 (2 × C), 133.4 (CH), 135.7 (4 × CH), 164.9 (C) (the 1636 benzoate ipso carbon is missing); ³¹P{¹H} NMR (202.5 MHz, 1637 CDCl₃) δ 30.5 (P); MS (ESI⁺-TOF) *m/z* (%) 677 [(M + Na)⁺, 100]; 1638 HRMS (ESI⁺-TOF) *m/z* [M + Na]⁺ calcd for C₃₄H₄₃NaO₉PSi 1639 677.2312; found 677.2313. Anal. calcd for C₃₄H₄₃O₉PSi: C, 62.37; H, 1640 6.62. Found: C, 62.23; H, 6.63. Compound **30**: colorless oil, [α]_D 1641 +20.9 (c, 1.13, CHCl₃); IR (CHCl₃) 2956, 2857, 1727, 1428, 1260, 1642 1092 cm⁻¹; ¹H NMR (500 MHz, CDCl₃, simulated coupling

constants using DAISY) δ 0.87 (9H, s), 1.37 (3H, s), 1.47 (3H, s), 1643 2.60 (1H, dddd, ²J_{PH} = 20.0 Hz, J = 10.8, 3.3, 2.0 Hz, H5), 3.72 (3H, 1644 d, ³J_{PH} = 10.7 Hz), 3.77 (3H, d, ³J_{PH} = 11.0 Hz), 4.14 (1H, ddd, ³J_{PH} = 1645 = 34.5 Hz, J = 9.9, 3.3 Hz, H6a), 4.14 (1H, ddd, ³J_{PH} = 11.3 Hz, J = 1646 9.9, 2.0 Hz, H6b), 4.74 (1H, ddd, ³J_{PH} = 4.3 Hz, J = 10.8, 2.9, Hz, 1647 H4), 4.84 (1H, d, J = 5.7 Hz, H2), 4.92 (1H, dd, J = 5.7, 2.9 Hz, H3), 1648 6.36 (1H, br s, J = 0.4 Hz, H1), 7.23–7.35 (8H, m), 7.42–7.47 (1H, 1649 m), 7.54–7.59 (2H, m), 7.67–7.69 (2H, m), 7.89–7.93 (2H, m); ¹H 1650 NMR (500 MHz, C₆D₆, simulated coupling constants using DAISY) δ 1651 1.06 (9H, s), 1.15 (3H, s), 1.31 (3H, s), 2.61 (1H, dddd, ²J_{PH} = 20.0 1652 Hz, J = 10.9, 3.1, 1.8 Hz, H5), 3.45 (3H, d, ³J_{PH} = 10.7 Hz), 3.51 (3H, 1653 d, ³J_{PH} = 11.0 Hz), 4.31 (1H, ddd, ³J_{PH} = 34.7 Hz, J = 9.9, 3.1 Hz, 1654 H6a), 4.31 (1H, ddd, ³J_{PH} = 10.8 Hz, J = 9.9, 1.8 Hz, H6b), 4.63 (1H, 1655 d, J = 5.7 Hz, H2), 4.92 (1H, dd, J = 5.7, 2.9 Hz, H3), 5.03 (1H, ddd, 1656 ³J_{PH} = 4.2 Hz, J = 10.9, 2.9 Hz, H4), 6.73 (1H, br s, J = 0.9 Hz, H1), 1657 6.95–7.02 (2H, m), 7.04–7.09 (1H, m), 7.18–7.25 (2H, m), 7.27– 1658 7.32 (2H, m), 7.36–7.39 (2H, m), 7.90 (2H, d, J = 6.9 Hz), 8.00 (2H, 1659 d, J = 6.9 Hz), 8.04 (2H, d, J = 7.6 Hz); ¹³C{¹H} NMR (125.7 MHz, 1660 CDCl₃) δ 19.2 (C), 25.2 (CH₃), 26.3 (CH₃), 26.5 (3 × CH₃), 38.4 1661 (CH, d, ¹J_{PC} = 137.8 Hz, C5), 52.2 (CH₃, d, ²J_{PC} = 6.4 Hz), 52.6 1662 (CH₃, d, ²J_{PC} = 5.3 Hz), 60.0 (CH₂, d, ²J_{PC} = 8.5 Hz, C6), 77.9 (CH, 1663 d, ²J_{PC} = 8.5 Hz, C4), 80.3 (CH, C3), 85.8 (CH, C2), 100.9 (CH, 1664 C1), 112.8 (C), 127.6 (2 × CH), 127.6 (2 × CH), 128.3 (2 × CH), 1665 129.49 (CH), 129.54 (CH), 129.8 (2 × CH), 132.8 (2 × C), 133.3, 1666 (CH), 135.6 (4 × CH), 165.1 (C) (the benzoate ipso carbon is 1667 missing); ³¹P{¹H} NMR (202.5 MHz, CDCl₃) δ 30.0 (P); MS (ESI⁺- 1668 TOF) *m/z* (%) 677 [(M + Na)⁺, 100]; HRMS (ESI⁺-TOF) *m/z* [M 1669 + Na]⁺ calcd for C₃₄H₄₃NaO₉PSi 677.2312; found 677.2310. Anal. 1670 calcd for C₃₄H₄₃O₉PSi: C, 62.37; H, 6.62. Found: C, 62.09; H, 6.84. 1671

5-Deoxy-5-dimethoxyphosphoryl-6-O-tert-butylidiphenylsilyl-2,3-O-1-isopropylidene-L-gulofuranose (31). A solution of **29** (67 1672 mg, 0.102 mmol) in dry MeOH (2.7 mL) containing NaOMe (11 mg, 1674 0.204 mmol) was stirred at room temperature for 2 h. The mixture 1675 was neutralized with acid ion-exchange resin Amberlite IR120, 1676 filtered, and concentrated under reduced pressure. The residue was 1677 purified by Chromatotron chromatography (hexanes–EtOAc, 20:80) 1678 to give **31** (54 mg, 0.098 mmol, 96%, mixture of anomers, α - β , 1679 92:8): colorless oil, major isomer, α -anomer: ¹H NMR (500 MHz, 1680 CDCl₃) δ 1.08 (9H, s), 1.19 (3H, s), 1.36 (3H, s), 2.58 (1H, dddd, 1681 ²J_{PH} = 18.3 Hz, J = 10.5, 3.7, 3.7 Hz, H5), 3.72 (3H, d, ³J_{PH} = 11.0 1682 Hz), 3.75 (3H, d, ³J_{PH} = 11.0 Hz), 3.94 (1H, ddd, ³J_{PH} = 27.7 Hz, J = 1683 10.7, 3.5 Hz, H6a), 4.10 (1H, ddd, ³J_{PH} = 10.2 Hz, J = 10.2, 4.1 Hz, 1684 H6b), 4.41 (1H, dd, J = 5.4, 3.5 Hz, H3), 4.50 (1H, d, J = 5.7 Hz, 1685 H2), 4.56 (1H, ddd, ³J_{PH} = 6.5 Hz, J = 10.2, 3.2 Hz, H4), 5.40 (1H, s, 1686 H1), 7.36–7.46 (6H, m), 7.68–7.71 (4H, m); ¹³C{¹H} NMR (125.7 1687 MHz, CDCl₃) δ 19.2 (C), 24.8 (CH₃), 26.1 (CH₃), 26.8 (3 × CH₃), 1688 39.6 (CH, d, ¹J_{PC} = 140.9 Hz, C5), 52.4 (CH₃, d, J = 7.4 Hz), 52.8 1689 (CH₃, d, J = 6.4 Hz), 61.1 (CH₂, d, ²J_{PC} = 6.4 Hz, C6), 77.0 (CH, d, 1690 ²J_{PC} = 4.5 Hz, C4), 79.9 (CH, d, ³J_{PC} = 12.7 Hz, C3), 85.4 (CH, C2), 1691 100.9 (CH, C1), 112.0 (C), 127.6 (2 × CH), 127.7 (2 × CH), 129.68 1692 (CH), 129.73 (CH), 133.1 (C), 133.2 (C), 135.7 (4 × CH); ³¹P{¹H} 1693 NMR (202.5 MHz, CDCl₃) δ 32.2 (P, α -anomer), 31.3 (P, β - 1694 anomer); MS (ESI⁺-TOF) *m/z* (%) 573 [(M + Na)⁺, 100]; HRMS 1695 (ESI⁺-TOF) *m/z* [M + Na]⁺ calcd for C₂₇H₃₉NaO₈PSi 573.2050; 1696 found 573.2048. 1697

5-Deoxy-5-dimethoxyphosphoryl-6-O-tert-butylidiphenylsilyl-2,3-O-1-isopropylidene-D-mannofuranose (32). A solution of **30** 1698 (13.5 mg, 0.021 mmol) in dry MeOH (0.55 mL) containing NaOMe 1699 (2.3 mg, 0.042 mmol) was stirred at room temperature for 2 h. The 1700 mixture was neutralized with acid ion-exchange resin Amberlite 1701 IR120, filtered, and concentrated under reduced pressure. The residue 1702 was purified by Chromatotron chromatography (hexanes–EtOAc, 1703 30:70) to give **32** (9.4 mg, 0.017 mmol, 81%, mixture of anomers, 1705 α - β , 92:8): colorless oil, major isomer, α -anomer: ¹H NMR (500 1706 MHz, CDCl₃) δ 1.08 (9H, s), 1.31 (3H, s), 1.37 (3H, s), 1.63 (1H, br 1707 s), 2.57 (1H, dddd, ²J_{PH} = 19.9 Hz, J = 10.4, 4.1, 2.5 Hz, H5), 3.74 1708 (3H, d, J = 11.0 Hz), 3.76 (3H, d, J = 10.7 Hz), 4.15 (1H, ddd, ³J_{PH} = 1709 31.2 Hz, J = 9.8, 4.1 Hz, H6a), 4.26 (1H, ddd, ³J_{PH} = 13.6 Hz, J = 1710 10.2, 2.4 Hz, H6b), 4.57 (1H, d, J = 5.7 Hz, H2), 4.59 (1H, ddd, ³J_{PH} 1711 = 5.4 Hz, J = 10.4, 2.7, H4), 4.81 (1H, dd, J = 5.7, 2.8 Hz, H3), 5.16 1712

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1845 Notes

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