



## Genes involved in cell division in mycoplasmas

Frank Alarcón<sup>1</sup>, Ana Tereza Ribeiro de Vasconcelos<sup>1</sup>, Lucia Yim<sup>2</sup> and Arnaldo Zaha<sup>3</sup>

<sup>1</sup>Laboratório Nacional de Computação Científica / Ministério da Ciência e Tecnologia, Petrópolis, RJ, Brazil.

<sup>2</sup>Instituto de Biologia Molecular do Paraná, Curitiba, PR, Brazil.

<sup>3</sup>Centro de Biotecnologia, Universidade Federal do Rio Grande do Sul, Porto Alegre, RS, Brazil.

### Abstract

Bacterial cell division has been studied mainly in model systems such as *Escherichia coli* and *Bacillus subtilis*, where it is described as a complex process with the participation of a group of proteins which assemble into a multiprotein complex called the septal ring. Mycoplasmas are cell wall-less bacteria presenting a reduced genome. Thus, it was important to compare their genomes to analyze putative genes involved in cell division processes. The division and cell wall (*dcw*) cluster, which in *E. coli* and *B. subtilis* is composed of 16 and 17 genes, respectively, is represented by only three to four genes in mycoplasmas. Even the most conserved protein, FtsZ, is not present in all mycoplasma genomes analyzed so far. A model for the FtsZ protein from *Mycoplasma hyopneumoniae* and *Mycoplasma synoviae* has been constructed. The conserved residues, essential for GTP/GDP binding, are present in FtsZ from both species. A strong conservation of hydrophobic amino acid patterns is observed, and is probably necessary for the structural stability of the protein when active. *M. synoviae* FtsZ presents an extended amino acid sequence at the C-terminal portion of the protein, which may participate in interactions with other still unknown proteins crucial for the cell division process.

*Key words:* cell division, *Mycoplasma* spp, genomes.

Received: May 3, 2006; Accepted: October 16, 2006.

### Introduction

Cell division in bacteria is a complex process involving the coordinated participation of a group of proteins which assemble at the division site into a multiprotein complex called the septal ring (for reviews see Errington *et al.*, 2003; Weiss, 2004; Goehring and Beckwith, 2005). This process has been well studied in some bacterial model systems, such as *Escherichia coli* and *Bacillus subtilis*.

The tubulin-like protein FtsZ, virtually present in all eubacteria, several archaeas, chloroplasts of plants and some mitochondria, plays a central role in cell division (for review see Margolin, 2005). This protein polymerizes to form the Z-ring, a structure associated to the cytosolic face of the inner membrane at midcell and essential for recruitment of other proteins to the division site. In *E. coli*, the following proteins are also components of the septal ring: FtsA, ZipA, ZapA, FtsEX, FtsK, FtsQ, FtsL, FtsB, FtsW, FtsI (PBP3), FtsN, AmiC and EnvC (Weiss, 2004). FtsA, ZipA and ZapA proteins assemble early to the division ring,

are mainly cytosolic and are involved in Z-ring stabilization. The remaining proteins assemble later to the ring, have membrane and/or periplasmic localization and except for FtsEX (unknown function) and FtsK (chromosome segregation, see below), participate in the synthesis of septal peptidoglycan.

FtsK is involved in chromosome partitioning during the last steps of DNA segregation, and is highly conserved almost throughout the eubacteria. It is a very large protein with an N-terminal transmembrane domain essential for septum formation and a cytoplasmic C-terminal domain which shows ATP dependent DNA translocase activity (Bigot *et al.*, 2004). Recently, it was demonstrated that FtsK interacts with topoisomerase IV, an enzyme responsible for chromosome decatenation (Espeli *et al.*, 2003).

In *B. subtilis*, besides FtsZ, the proteins FtsA, FtsW, DivIB (FtsQ homolog), DivIC (FtsL-like), and PBP2B (PBP 3 homolog) have been implicated in cell division, and their localization to the division site has been shown to be FtsZ-dependent (Beall and Lutkenhaus, 1992; Katis *et al.*, 1997, 1999; Daniel and Errington, 2000; Daniel *et al.*, 2000; Feucht *et al.*, 2001). In this bacterium, EzrA has been identified as a negative, and ZapA as a likely positive regu-

lator of FtsZ polymerization (Levin *et al.*, 1999; Gueiros-Filho and Losick, 2002).

FtsH and FtsY, even if they were initially identified by mutants that showed a filamentation phenotype, they are in fact involved in general processes that have pleiotropic effects on cell division (Tomoyasu *et al.*, 1993; Seluanov and Bibi, 1997).

Several other proteins are involved in different steps of the cell division process such as site selection (Min proteins, Noc, SlmA) and chromosome segregation (Smc proteins) (for review see Goehring and Beckwith, 2005).

Mycoplasmas belong to the class *Mollicutes*, a group of bacteria characterized by the absence of a cell wall. These bacteria present AT-rich genomes with sizes ranging from 580,070 bp in *Mycoplasma genitalium* to 1,358,633 bp in *Mycoplasma penetrans*. Most of them are parasites of a wide group of organisms, including mammals, birds, reptiles, arthropods, fish, and plants (Razin *et al.*, 1998). Only a few studies on *Mollicutes* have been dedicated to genes involved in cell division, particularly the gene *ftsZ* (Wang and Lutkenhaus, 1996; Kukekova *et al.*, 1999; Momynaliev *et al.*, 2002; Benders *et al.*, 2005). This gene is present in all mycoplasma genomes analyzed so far, with the exception of *Ureaplasma urealyticum* serovar 3 and *Mycoplasma mobile* (Glass *et al.*, 2000, Jaffe *et al.*, 2004). Mycoplasmas present genomes with reduced sizes, and the *M. genitalium* genome is considered to be the smallest for a self-replicating organism (Fraser *et al.*, 1995).

Presently, there are twelve completely sequenced mycoplasma genomes. Since these bacteria do not present a cell wall and present reduced genome sizes, it was important to compare their genomes to analyze putative genes involved in cell division processes. This work presents the results of such analysis and shows that a reduction in size of mycoplasma genomes leads to the loss of most genes involved in cell division. We also show that *ftsZ* is present in most of the mycoplasma genomes, and that its sequence and structure do not present regions involved in known interactions with other cell division proteins.

## Material and Methods

### DNA sequence analysis

The data on the complete genomes were downloaded from the Entrez genome except for the *Mycoplasma synoviae* and *Mycoplasma hyopneumoniae* strain J and *Mycoplasma hyopneumoniae* strain 7448, which were analyzed within the Brazilian National Genome Sequencing Consortium and the Southern Genome Investigation Program (PIGS). The comparison of CDS among the studied genomes was done through the SABIA (System for Automated Bacterial Integrated (genome) Annotation) software (Almeida *et al.*, 2004). Genes possibly involved in cell division in mycoplasmas were identified by blastp analysis using cell division genes from *E. coli* and *B. subtilis*.

Sequences presenting hits with E value of E-05 or less and a coverage higher than 30% were selected.

### Protein homology modeling

Multiple sequence alignment for modeling was prepared using *Mycoplasma* sp. sequences taken from SWISSPROT (ODonovan *et al.*, 2002) and then aligned using ClustalW (Thompson *et al.*, 1994). Theoretical structural framework models for FtsZ proteins from *M. synoviae* and *M. hyopneumoniae* 7448 were constructed using the atomic coordinates of FtsZ proteins from other bacterial species (1FSZ and 1W5B from *Methanococcus jannaschii*) (Lowe and Amos., 1998; Oliva *et al.*, 2004), 1OFU from *Pseudomonas aeruginosa* (Cordell *et al.*, 2003), and 1RLU from *Mycobacterium tuberculosis* (Leung *et al.*, 2004) and is available with these codes at RCSB website (Berman *et al.*, 2000). After the construction of a structural sequence alignment between FtsZ templates, models and their parts were calculated using the program MODELLER 8v1 (Sali and Blundell., 1993; Marti-Renom *et al.*, 2000) by the previous calculation of pairwise and multiple alignments between the template structures/sequences and the target sequences (FtsZ sequences from *M. synoviae* and *M. hyopneumoniae* 7448). Models that matched the lowest objective function score, as well as those which satisfied visual and stereochemical inspection, were chosen to produce the final models used in this work. Stereochemical validation was done using the package PROCHECK (Morris *et al.*, 1992, Laskowski *et al.*, 1993).

## Results and Discussion

### Putative genes involved in cell division in mycoplasmas

The genes that may be involved in cell division in ten species of mycoplasma are shown in Table 1. The gene encoding FtsZ is present in all except *U. urealyticum* serovar 3 (Glass *et al.*, 2000) and *M. mobile* (Jaffe *et al.*, 2004). FtsK, a protein involved in chromosome partitioning, is present only in *Mycoplasma gallisepticum* and *U. urealyticum* serovar 3. Species of the clade Pneumoniae (*M. genitalium*, *M. gallisepticum*, *Mycoplasma pneumoniae* and *U. urealyticum*) seem to have conserved more genes involved in cell division than members of the clade Hominis (*M. hyopneumoniae*, *M. synoviae*, *Mycoplasma pulmonis* and *M. mobile*). Orthologs of the structural maintenance of chromosomes (SMC) proteins, involved in chromosome segregation, are present in all genomes. The gene encoding ParA, a protein involved in chromosome partitioning, is present in *M. genitalium*, *M. pneumoniae*, *M. penetrans*, and *M. gallisepticum*. *M. penetrans*, *M. pulmonis* and *U. urealyticum* serovar 3 present genes encoding recombinases (xerD/xerC family) involved in resolution of the chromosomes. EzrA, a protein identified as a negative regulator of FtsZ polymerization,

**Table 1** - Putative genes involved in cell division in mycoplasmas. Genomes of ten different *Mycoplasma* species have been compared. Data from three different strains of *Mycoplasma hyopneumoniae* are presented.

Species	Length	Gene*	Product*
<i>M. genitalium</i>	612	smcC	P115 protein
	982	soj	soj protein
	326	ftsZ	cell division protein
<i>M. pneumoniae</i>	270	soj	ParA family of ATPases involved in chromosome partition
	982	smc	SMC family, chromosome/DNA binding/protecting functions
	380	ftsZ	cell division protein
<i>M. penetrans</i>	275	soj	Soj protein
	984	smc	structural maintenance of chromosomes SMC superfamily proteins
	253	MYPE2900	recombinase (xerD/xerC family)
	476	ftsZ	cell division protein FtsZ
<i>M. gallisepticum</i>	263	parA	ParA/Soj
	983	smc-like	Smc-like
	1931	Ezra	conserved hypothetical
	462	ftsZ	FtsZ
	655	ftsK	FtsK
<i>U. urealyticum</i> serovar 3	981	p115 (smc)	p115 protein
	251	codV	integrase-recombinase protein
	250	xerC	integrase-recombinase protein (xerD/xerC family)
	351	ftsY	SRP family of GTP-binding proteins - cell division protein
<i>M. hyopneumoniae</i> 232	979	p115 (smc)	p115 protein
	327	ftsZ	cell division protein
<i>M. hyopneumoniae</i> 7448	980	smc	ABC transporter ATP-binding protein P115-like
	322	ftsZ	cell division protein
<i>M. hyopneumoniae</i> J	980	smc	ABC transporter ATP-binding protein P115-like
	322	ftsZ	cell division protein
<i>M. pulmonis</i>	979	smc	P115-Like - ABC transporter ATP-binding protein
		MYPU5310	integrase/recombinase (xerD/xerC family)
	390	ftsZ	cell division protein
<i>M. synoviae</i>	981	smc	ABC transporter ATP-binding protein P115-like
	543	ftsZ	cell division protein
<i>M. mobile</i>	974	smc	segregation of chromosomes protein
<i>M. mycoides</i>	472	smc	conserved hypothetical protein

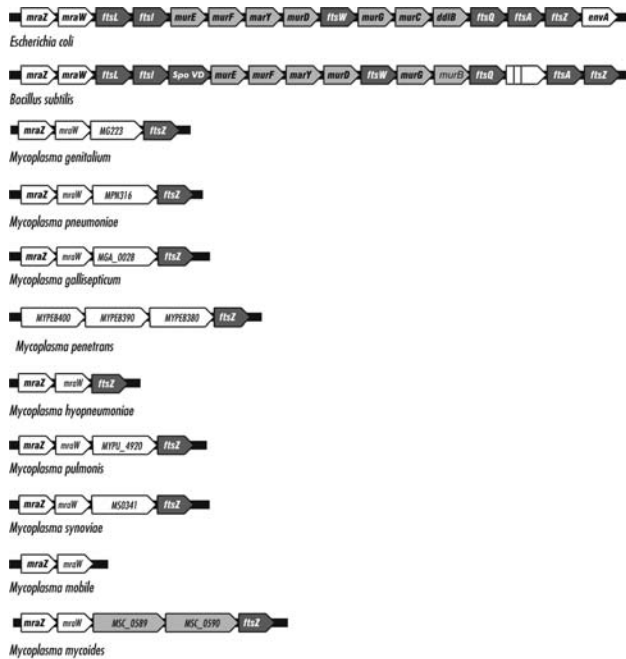
\* The information about the genes or gene products is based on the original annotation of the genomes.

was observed only in the *M. gallisepticum* genome. The genes encoding FtsH and FtsY are present in all species analyzed, and although they may affect cell division, they are not directly involved in this process.

### The *ftsZ* gene cluster

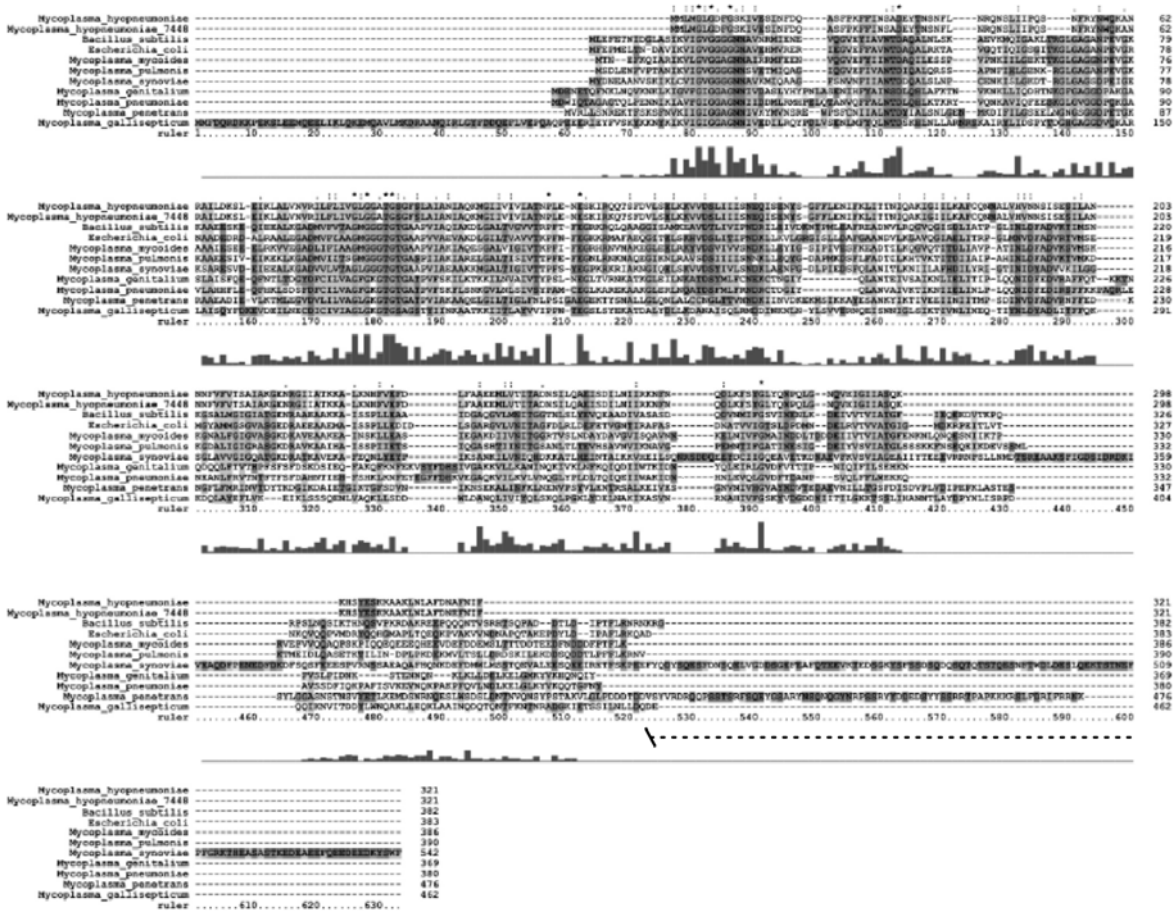
In bacteria, several genes involved in cell division and synthesis of the cell wall precursors are organized in the *dcw* (division and cell wall) cluster, a highly conserved group of about 16 genes, including *ftsZ* (Margolin, 2000;

Mingorance *et al.*, 2004). The functions of the first two genes in the cluster (*mraZ* and *mraW*) are unknown. The *mraW* gene encodes an S-adenosyl-methionine-dependent methyltransferase, and although the structure has been resolved, no function has been defined for this protein in cell division. Six *fts* genes (*ftsL*, *ftsI*, *ftsW*, *ftsQ*, *ftsA* and *ftsZ*) are also present in this cluster, and the products of these genes are involved in the formation of the division ring. Several genes involved in synthesis of murein precursors also belong to this cluster: genes *murBCDEFG*, *mraY* and

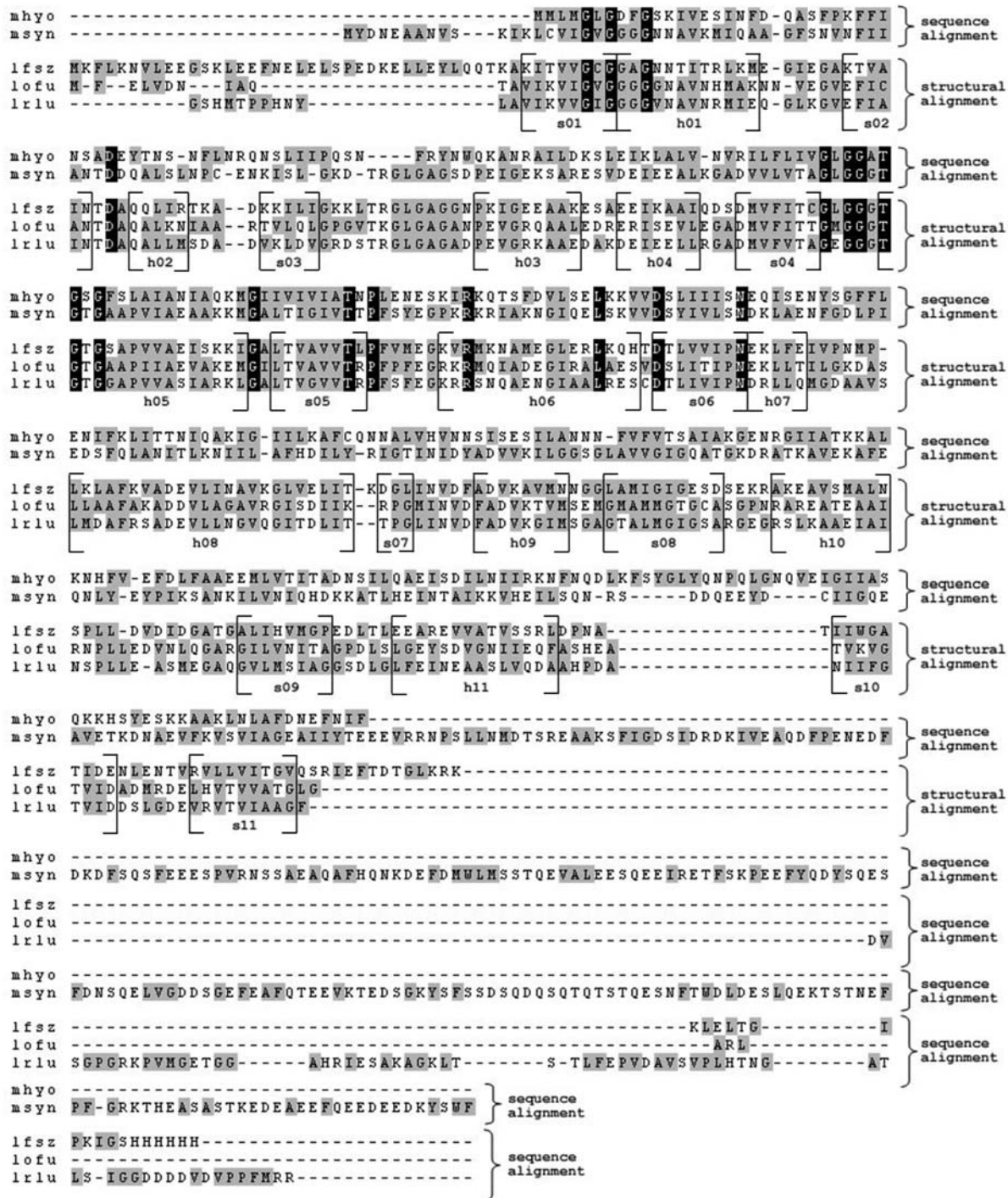


**Figure 1** - Comparison of the organization of genes of the *dcw* cluster in mycoplasmas. The organization of the genes in *E. coli* and *B. subtilis* is also presented for comparison.

*ddlB*. In mycoplasmas, which lack a cell wall, the number of genes in this cluster is much lower (Benders *et al.*, 2005). The *ftsL*, *ftsI*, *ftsW* and *ftsQ* gene products are all transmembrane proteins with the bulk of the proteins extended to the periplasm, and are involved in peptidoglycan synthesis during division (Weiss, 2004). The absence of a cell wall in mycoplasmas might explain why these genes are absent in their genomes. The same reasoning could be extended to the genes *murBCDEFG*, *mraY* and *ddlB*. The comparison of the *dcw* cluster in mycoplasmas shows that the organization of the genes in this group of bacteria is conserved (Figure 1). The *M. genitalium*, *M. pneumoniae* and *M. gallisepticum* *dcw* cluster is composed of four genes, *mraZ*, *mraW*, a conserved hypothetical CDS, and *ftsZ*. A similar organization was observed in *M. pulmonis* and *M. synoviae*, but the conserved hypothetical CDSs are not related to each other. In *M. hyopneumoniae*, the cluster is composed of three CDSs: *mraZ*, *mraW* and *ftsZ*. The *mraW* gene is present in *M. penetrans*, but in another position of the genome (424743..425681). No sequence related to the *mraZ* gene was observed in the *M. penetrans* genome, and the three CDSs located close to the *ftsZ* gene are conserved



**Figure 2** - Sequence alignment of FtsZ proteins from different species of mycoplasmas. The extended amino acid sequence concentrated at the C-terminal portion of the *M. synoviae* FtsZ protein is indicated by  $\backslash$ -----/.



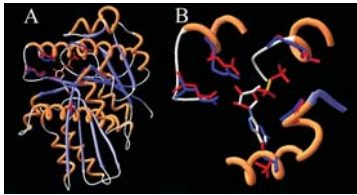
**Figure 3** - Structural alignment of FtsZ proteins. FtsZ templates (pdb codes) lfsz: *M. jannaschii*; lofu: *P. aeruginosa*; lrlu: *M. tuberculosis*) used to construct structural models of FtsZ proteins for *M. synoviae* (msyn) and *M. hyopneumoniae* 7448 (mhyo). Helices (h) and strands (s) in the template structures are represented along their amino acid sequences. The sequence alignment of mycoplasma amino acid sequences is represented above the template structural alignment.

hypothetical sequences with low similarity to other sequences in the nr database.

#### FtsZ protein in mycoplasmas

In mycoplasmas, the gene encoding FtsZ was initially characterized in *M. pulmonis*. The FtsZ protein of this or-

ganism was able to bind GTP (Wang and Lutkenhaus, 1996). It was also shown that its extreme carboxy tail, which is highly conserved between the *E. coli* and *B. subtilis* FtsZ proteins, is not as well conserved in *M. pulmonis* FtsZ (Wang and Lutkenhaus, 1996). We have extended the FtsZ comparative analysis to other mycoplasma



**Figure 4** - FtsZ structural model for *M. synoviae*. (A) Cartoon representation of FtsZ theoretical model and its GTP/GDP binding site. Amino acids that interact through hydrogen bonds with GTP/GDP are shown. The long C-terminal tail in FtsZ from *M. synoviae* is not shown. (B) Comparison of interacting amino acids from FtsZ with GTP/GDP molecule; Red amino acids (G21, G22, G107, T108, E138, R142 and N185) are those from the *M. synoviae* protein; Blue amino acids (G21, G22, G108, T109, E139, R143 and D187) are those from the *Pseudomonas aeruginosa* protein (1OFU). There are no significant differences in spatial configuration nor amino acid composition.

species. The alignment of mycoplasma FtsZ sequences shows that the conserved C-terminal portion, which appears in several eubacteria species, including *E. coli* and *B. subtilis*, is absent in this group of bacteria (Figure 2). Despite the low similarity of *M. synoviae* and *M. hyopneumoniae* 7448 FtsZ amino acid sequences with the FtsZ templates used in this work (around 20-30% of sequence identity), it was possible to suggest reasonable carbon alpha amino acid framework models for *M. synoviae* and *M. hyopneumoniae* 7448 FtsZ proteins, based on identities and similarities present in the structural alignment of templates (Figures 3 and 4). According to the models, no structural differences were observed for the main domains of FtsZ and its relationships. The conserved residues essential for GTP/GDP binding, typical for the FtsZ family, are present in proteins from both species. There is strong conservation of hydrophobic amino acid patterns along the sequences, which are probably necessary for the structural stability of the protein when active (Figures 3 and 4).

Despite the suggested structural uniformity for mycoplasma FtsZ proteins compared with the template's structures used during modeling, the *Mycoplasma* sp. amino acid sequences show some particularities. *M. synoviae* FtsZ presents an extended amino acid sequence (more than 200 amino acids) concentrated at the C-terminal portion of the protein (Figure 2). In several species this region is essential for interaction with other proteins, but not for the protofilament formation, since the central core of the protein (GTP binding site and neighbour sites) is involved in these processes. Since Mycoplasma genomes lack other important proteins involved in cell division processes and Z-ring formation (like ZipA and FtsA, if compared with other eubacteria), this amino acid tail is probably involved with other still unknown proteins crucial for cell division processes. It is well known that ZipA and FtsA help in the anchoring of FtsZ to the membrane during Z-ring formation (Hale and de Boer, 1997; Ma *et al.*, 1997). These proteins interact with FtsZ mainly through the C-terminal peptide. As *M. synoviae* does not present these proteins or similar

ones, a possible explanation for the function of the long tail located at the C-terminus of FtsZ could be its ability to fold in a domain that could interact directly with the inner bacterial membrane or with a transmembrane protein inserted into it (allowing the attachment necessary for the Z-ring formation). Actually, the deletion of C-terminal segments of FtsZ proteins blocks FtsZ functions, probably by preventing its interaction with other proteins important to the stability of the Z-ring (Ma and Margolin, 1999; Redick *et al.*, 2005). It was not possible to suggest a structural model for this C-terminal region as there is no solved structure similar enough to be used as a template. Both *M. synoviae* and *M. hyopneumoniae* 7448 amino acid sequences show a smaller amount of amino acids between strand03 and helix03, when compared with the template sequences (Figure 3). The region is spatially close to the GTP binding site, in some way contributing to the chemical environment and stability that directly affects the GTP molecule. As this region is smaller in mycoplasma sequences, it is reasonable to assume that the GTP molecule will be more exposed than normally. This situation could contribute positively or negatively to the GTP hydrolysis and/or protofilament formation, since both processes are highly dependent on GTP stability/accessibility or GTP binding site protection (Margolin, 2005).

Conversely, mycoplasma amino acid sequences were larger than usual between helix11 and strand10 (Figure 3). The extra amino acids are located on the protein surface and are highly exposed. There are no data to suggest that these amino acids could play an important role in the function of the protein. However, as several typical proteins involved in cell division processes are not observed in mycoplasmas, the importance of these fragments in the interaction with other proteins cannot be discarded.

The cell division process in mycoplasma involves a lower number of proteins, compared to the machinery described for *E. coli* and *B. subtilis*. However, the possible involvement of other proteins, with different amino acid sequences, cannot be ruled out. The molecular mechanisms of the cell division process in mycoplasma remain unclear.

## Acknowledgments

We would like to thank Vicente da Araujo Calfo and Rangel Celso Souza for helping us with the figures. The present and former staffs of the Ministério da Ciência e Tecnologia (MCT)/Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) are gratefully acknowledged for their strategic vision and enthusiastic support. This work was undertaken by the Brazilian National Genome Program (Southern Network for Genome Analysis and Brazilian National Genome Project Consortium) with funding provided by MCT/CNPq and SCT/FAPERGS (RS).

## References

- Almeida LG, Paixão R, Souza RC, Costa GC, Barrientos FJ, Santos MT, Almeida DF and Vasconcelos AT (2004) A system for automated bacterial (genome) integrated annotation -SABIA. *Bioinformatics* 20:2832-2833.
- Beall B and Lutkenhaus J (1992) Impaired cell division and sporulation of a *Bacillus subtilis* strain with the *ftsA* gene deleted. *J Bacteriol* 174:2398-2403.
- Benders GA, Powell BC and Hutchison CA 3<sup>rd</sup> (2005) Transcriptional analysis of the conserved *ftsZ* gene cluster in *Mycoplasma genitalium* and *Mycoplasma pneumoniae*. *J Bacteriol* 187:4542-4551.
- Berman HM, Westbrook J, Feng Z, Gilliland G, Bhat TN, Weissig H, Shindyalov IN and Bourne PE (2000) The protein data bank. *Nucleic Acids Res* 28:235-242.
- Bigot S, Corre J, Louarn JM, Cornet F and Barre FX (2004) FtsK activities in Xer recombination, DNA mobilization and cell division involve overlapping and separate domains of the protein. *Mol Microbiol* 54:876-886.
- Cordell SC, Robinson EJ and Lowe J (2003) Crystal structure of the SOS cell division inhibitor Sula and in complex with Ftsz. *Proc Nat Acad Sci USA* 100:7889-7894.
- Daniel RA and Errington J (2000) Intrinsic instability of the essential cell division protein FtsL of *Bacillus subtilis* and a role for DivIB protein in FtsL turnover. *Mol Microbiol* 36:278-289.
- Daniel RA, Harry EJ and Errington J (2000) Role of penicillin-binding protein PBP 2B in assembly and functioning of the division machinery of *Bacillus subtilis*. *Mol Microbiol* 35:299-311.
- Errington J, Daniel RA and Scheffers DJ (2003) Cytokinesis in bacteria. *Microbiol Mol Biol Rev* 67:52-56.
- Espeli O, Lee C and Mariani KJ (2003) A physical and functional interaction between *Escherichia coli* FtsK and topoisomerase IV. *J Biol Chem* 278:44639-44644.
- Feucht A, Lucet I, Yudkin MD and Errington J (2001) Cytological and biochemical characterization of the FtsA cell division protein of *Bacillus subtilis*. *Mol Microbiol* 40:115-125.
- Fraser CM, Gocayne JD, White O, Adams MD, Clayton RA, Fleischmann RD, Bult CJ, Kerlavage AR, Sutton G, Kelley JM, Fritchman RD, Weidman JF, Small KV, Sandusky M, Fuhrmann J, Nguyen D, Utterback TR, Saudek DM, Phillips CA, Merrick JM, Tomb JF, Dougherty BA, Bott KF, Hu PC, Lucier TS, Peterson SN, Smith HO, Hutchison CA 3<sup>rd</sup> and Venter JC (1995) The minimal gene complement of *Mycoplasma genitalium*. *Science* 270:397-403.
- Glass JI, Lefkowitz EJ, Glass JS, Heiner CR, Chen EY and Cassell GH (2000). The complete sequence of the mucosal pathogen *Ureaplasma urealyticum*. *Nature* 407:757-762.
- Goehring NW and Beckwith J (2005) Diverse paths to midcell: Assembly of the bacterial cell division machinery. *Curr Biol* 15:R514-R526.
- Gueiros-Filho FJ and Losick R (2002) A widely conserved bacterial cell division protein that promotes assembly of the tubulin-like protein FtsZ. *Genes Dev* 16:2544-2556.
- Hale CA and de Boer PA (1997) Direct binding of FtsZ to ZipA, an essential component of the septal ring structure that mediates cell division in *E. coli*. *Cell* 88:175-185.
- Jaffe JD, Stange-Thomann N, Smith C, DeCaprio D, Fisher S, Butler J, Calvo S, Elkins T, FitzGerald MG, Hafez N, Kodira CD, Major J, Wang S, Wilkinson J, Nicol R, Nusbaum C, Birren B, Berg HC and Church GM (2004) The complete genome and proteome of *Mycoplasma mobile*. *Genome Res* 14:1447-1461.
- Katis VL and Wake RG (1999) Membrane-bound division proteins DivIB and DivIC of *Bacillus subtilis* function solely through their external domains in both vegetative and sporulation division. *J Bacteriol* 181:2710-2718.
- Katis VL, Harry EJ and Wake RG (1997) The *Bacillus subtilis* division protein DivIC is a highly abundant membrane-bound protein that localizes to the division site. *Mol Microbiol* 26:1047-1055.
- Kukekova AV, Malinin AY, Ayala JA and Borchsenius SN (1999) Characterization of *Acholeplasma laidlawii* *ftsZ* gene and its gene product. *Biochem Biophys Res Commun* 262:44-49.
- Laskowski RA, MacArthur MW, Moss DS and Thornton JM (1993) PROCHECK: A program to check the stereochemical quality of protein structures. *J Appl Cryst* 26:283-291.
- Leung AK, Lucile White E, Ross LJ, Reynolds RC, DeVito JA and Borhani DW (2004) Structure of *Mycobacterium tuberculosis* FtsZ reveals unexpected, G protein-like conformational switches. *J Mol Biol* 342:953-970.
- Levin PA, Kurtser IG and Grossman AD (1999) Identification and characterization of a negative regulator of FtsZ ring formation in *Bacillus subtilis*. *Proc Natl Acad Sci USA* 96:9642-9647.
- Lowe J and Amos LA (1998) Crystal structure of the bacterial cell-division protein FtsZ. *Nature* 391:203-206.
- Ma X, Sun Q, Wang R, Singh G, Jonietz EL and Margolin W (1997) Interactions between heterologous FtsA and FtsZ proteins at the FtsZ ring. *J Bacteriol* 179:6788-6797.
- Ma X and Margolin W (1999) Genetic and functional analyses of the conserved C-terminal core domain of *Escherichia coli* FtsZ. *J Bacteriol* 181:7531-7544.
- Marti-Renom MA, Stuart A, Fiser A, Sánchez R, Melo F and Sali A (2000) Comparative protein structure modeling of genes and genomes. *Ann Rev Biophys Biomol Struct* 29:291-325.
- Margolin W (2000) Themes and variations in prokaryotic cell division. *FEMS Microbiol Rev* 24:531-548.
- Margolin W (2005) FtsZ and the division of prokaryotic cells and organelles. *Nat Rev Mol Cell Biol* 11:862-871.
- Mingorance J, Tamames J and Vicente M (2004) Genomic channeling in bacterial cell division. *J Mol Recognit* 17:481-487.
- Momynaliev KT, Smirnova OV, Lazyrev VN, Akopian TA, Chelysheva VV, Ayala JA, Simankova AN, Borchsenius SN and Govorun VM (2002) Characterization of the *Mycoplasma hominis* *ftsZ* gene and its sequence variability in mycoplasma clinical isolates. *Biochem Biophys Res Commun* 293:155-162.
- Morris AL, MacArthur MW, Hutchinson EG and Thornton JM (1992) Stereochemical quality of protein structure coordinates. *Proteins* 12:345-364.
- O'Donovan C, Martin MJ, Gattiker A, Gasteiger E, Bairoch A and Apweiler R (2002) High-quality protein knowledge resource: SWISS-PROT and TrEMBL. *Brief Bioinform* 3:275-284.
- Oliva MA, Cordell SC and Lowe J (2004) Structural insights into Ftsz protofilament formation. *Nat Struct Mol Biol* 11:1243-1250.

- Razin S, Yogev D and Naot Y (1998) Molecular biology and pathogenicity of mycoplasmas. *Microbiol Mol Biol Rev* 62:1094-1156.
- Redick SD, Stricker J, Briscoe G and Erickson HP (2005) Mutants of FtsZ targeting the protofilament interface: Effects on cell division and GTPase activity. *J Bacteriol* 187:2727-2236.
- Sali A and Blundell TL (1993) Comparative protein modelling by satisfaction of spatial restraints. *J Mol Biol* 234:779-815.
- Seluanov A and Bibi E (1997) FtsY, the prokaryotic signal recognition particle receptor homologue, is essential for biogenesis of membrane proteins. *J Biol Chem* 272:2053-2055.
- Thompson JD, Higgins DG and Gibson TJ (1994) CLUSTAL W: Improving the sensitivity of progressive multiple sequence alignment through sequence weighting, positions-specific gap penalties and weight matrix choice. *Nucleic Acids Res* 22:4673-4680.
- Tomoyasu T, Yuki T, Morimura S, Mori H, Yamanaka K, Niki H, Hiraga S and Ogura T (1993) The *Escherichia coli* FtsH protein is a prokaryotic member of a protein family of putative ATPases involved in membrane functions, cell cycle control, and gene expression. *J Bacteriol* 175:1344-1351.
- Wang X and Lutkenhaus J (1996) Characterization of the *ftsZ* gene from *Mycoplasma pulmonis*, an organism lacking a cell wall. *J Bacteriol* 178:2314-2319.
- Weiss DS (2004) Bacterial cell division and the septal ring. *Mol Microbiol* 54:588-597.

### Internet Resources Section

<http://www.ncbi.nlm.nih.gov> -GenBank Database (9/12/2006).

*Associate Editor: Darcy F. de Almeida*