Accepted Manuscript

Preparation of Azide Biosynthetic Surrogates of myo-Inositol

Sandip Pasari, Shareef M. Ismail, Markus R. Wenk, Martin J. Lear

PII:	\$0040-4039(15)00623-1
DOI:	http://dx.doi.org/10.1016/j.tetlet.2015.04.009
Reference:	TETL 46146

To appear in: Tetrahedron Letters

Received Date:23 January 2015Revised Date:29 March 2015Accepted Date:2 April 2015



Please cite this article as: Pasari, S., Ismail, S.M., Wenk, M.R., Lear, M.J., Preparation of Azide Biosynthetic Surrogates of *myo*-Inositol, *Tetrahedron Letters* (2015), doi: http://dx.doi.org/10.1016/j.tetlet.2015.04.009

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Graphical Abstract

To create your abstract, type over the instructions in the template box below. Fonts or abstract dimensions should not be changed or altered.





Tetrahedron Letters

journal homepage: www.elsevier.com

Preparation of Azide Biosynthetic Surrogates of myo-Inositol

Sandip Pasari^a, Shareef M. Ismail,^b Markus R. Wenk^{b,c}, Martin J. Lear*^{a,d}

^a Department of Chemistry and Medicinal Chemistry Program, Faculty of Science, Science Drive 3, National University of Singapore, Singapore 117543

^b National University of Singapore (NUS) Graduate School for Integrative Sciences and Engineering (NGS), Centre for Life Sciences, 28 Medical Drive, Singapore 117456

^c Department of Biochemistry and Department of Biological Science, Yong Loo School of Medicine, National University of Singapore, Singapore 117543 ^d School of Chemistry, University of Lincoln, Brayford Pool, Lincoln, Lincolnshire, LN6 7TS, UK

ARTICLE INFO

Received in revised form

Article history:

Available online

Received

Accepted

Keywords:

Deoxy sugars

Biosynthesis

Glycolipids

Inositols

ABSTRACT

As a prelude to biomolecular incorporation studies, practical routes to a series of four regioisomeric azido-deoxy derivatives of inositol that mimic the natural *myo*-stereochemistry are described. Starting from commercially available *myo*-inositol, the regioselective and stereoselective introduction of azide functionality was achieved at the C-2, C-3, C-4 and C-5 positions via azide displacement of the corresponding *O*-sulfonates of suitably protected *scyllo*-, *chiro*-, *epi*- and *neo*-inositols, respectively. Notably, a final one-pot acetolysis method conveniently allowed for rapid access to pentaacetate azido-deoxy inositols. Investigations on the metabolic incorporation of these *myo*-inositol azide surrogates in both acetate and free alcohol forms are in progress.

2009 Elsevier Ltd. All rights reserved.

Due to its putative prebiotic history and innate chemical versatility, inositol is a natural carbohydrate central to a plethora of cellular signaling processes and recognition events, both inside and outside cells, and even between different cell types.¹ Typically through initial phosphorylation events, inositol derivatives (e.g., inositol triphosphates, IP₃, and lipidated phosphatidylinositols, PIs) play key roles in modulating calcium signaling, membrane functioning, cell death, cell division and cell-to-cell communication.^{2,3} Additional glycosylation and lipidation events eventually yield higher order glycolipids, such as the glycosylphosphatidylinositols (GPIs) and polymannosylated lipids, including the phosphatidyl-myo-inositol mannosides (PIMs) and lipoarabinomannans (LAMs). These glycolipids are essential components in anchoring functional proteins on extracellular membrane surfaces and intracellular trafficking processes, and are key to host-cell recognition, immunological processes, and pathogenic infections.⁴

In this study, we have a view to prepare biosynthetic mimics of *myo*-inositol (azide-surrogates) that have the potential to be metabolically incorporated into a chosen cell type (e.g., yeast). For this purpose, the modification or introduction of bioorthogonal functional groups ("click" functionalities) was necessary. Herein, we chose to introduce azide groups to replace the different hydroxyl groups of *myo*-inositol (1) (Figure 1). For symmetry reasons, this would result in four regioisomeric series of *myo*-cyclohexanols 2-5, of which two (the 2-series and 5-series) would be *meso* (achiral).





A few routes to the synthesis of optically active azido and amino inositols have been reported. Examples of routes start from *p*-benzoquinone⁵ or conduritol- E^6 via chemo-enzymatic resolution, and also from chiral sources such as L-quebrachitol.⁷ Among various synthetic protocols developed for the synthesis of myo-inositol intermediates and its analogues, commercially available *myo*-inositol **1** is the most commonly preferred starting material due to its low cost and pre-defined relative stereochemistry.⁸ For *myo*-inositol, positions C_1 , C_3 and C_4 , C_6 are equivalent and unsymmetrical protection leads to racemates. The chemical synthesis of optically active inositol analogues would thus necessitate the resolution of racemic inositol intermediates (chemically, enzymatically or via desymmetrization techniques)⁹ or by starting with an alternative chiral material (e.g., via the Ferrier carbo-cyclization of sugars).¹⁰ For our biological studies, the racemic azido-inositol series 3 and 4, and the *meso* series 2 and 5, were considered sufficient to test our hypothesis of metabolic selection and incorporation into live cells. Our synthetic routes to make 2-5 are described herein.

^{*} Corresponding author. Tel.: +44-1522-837411; fax: +44-1522-837411; e-mail: mlear@lincoln.ac.uk

2

ACCEPTED MANUSCRIP1

Tetrahedron

Synthesis of (±)-4-Deoxy-4-Azido myo-Inositols 4:

We first decided to pursue the dicyclohexylidene diketal protection approach reported by Angyal and co-workers¹¹ as a starting material over diacetals or diisopropylidene derivatives.^{12,13} This allows for the practical, differential hydroxyl protection of *myo*-inositol (1) as relatively stable *trans* and *cis* cyclohexylidenes that not only tolerate multi-step synthesis, but also result in reactivity differences between the two remaining hydroxyl groups. The 1,2,4,5-dicyclohexylidene *myo*-inositol **6** was isolated as a white solid in 22% yield after recrystallisation from 1:9 acetone/petroleum ether.¹⁴ The remaining two isomeric ketals (2,3,4,5- and 3,4,5,6-dicyclohexylidenes) can be converted to **6** in a sequence of partial deprotection-reprotection steps.^{11b} Although yields are low, this method is scalable to multi-grams and is convenient in practice.



Scheme 1. Synthesis of (±)-4-azido *myo*-inositol analogues 4a,b. Reagents and conditions: i. Cyclohexanone, toluene/DMF (1:2), pTSA, 110 °C, 22%; ii. BaO, Ba(OH)₂·8H₂O, BnBr, DMF, 60%; iii. DMP, DCM, rt, 93%; iv. NaBH₄, EtOH, 0 °C, 76%; v. Tf₂O, Py, DCM 0 °C to rt, 73%; vi. 11, AcOH, Ac₂O, 10% H₂SO₄ in Ac₂O, rt; vii. NaOMe, MeOH, rt, 99%.

Barium oxide and barium hydroxide chelation-mediated benzylation of **6** produced the monobenzylated *myo*-inositol derivative **7** as the major product in 60% yield¹⁴ (Scheme 1). The free alcohol at the C₆ position of the *myo*-inositol derivative **7** was subsequently oxidised under Dess-Martin periodinane (DMP) conditions to give the desired ketone **8** in excellent yield (93%). Stereoselective reduction of **8** by sodium borohydride produced the *epi*-alcohol **9**¹⁵ exclusively. The free C₆ alcohol of *epi*-inositol **9** was subsequently activated as its triflate **10** using trifluoromethane sulfonic acid anhydride in pyridine.¹⁶ In accordance with the report of Schlewer *et al.*,¹⁵ azide displacement of triflate **10** gave the desired *myo*-product **11** together with trace amounts of the enol ether side-product **12** via 1,2-*anti* elimination under the basic conditions.

After several methods were explored, global deprotection of **11** in the presence of the azide group was eventually achieved in one step by acetolysis¹⁷ using a mixture of sulphuric acid, acetic anhydride and acetic acid to afford the 4-deoxy-4-azido-*myo*-inositol pentaacetate **4a**^{5.6} in 65% yield. Minor amounts of the partially acetolysed product 3-benzyl-6-azido-*myo*-inositol tetraacetate **13** was also obtained, which could be transformed under the same conditions to yield **4a**. Further methanolysis in the presence of a catalytic amount of sodium methoxide afforded the 4-deoxy-4-azido-*myo*-inositol analogue **4b**. Single crystals of pentacetate **4a**, from 1:1 hexane/ethyl acetate, confirmed the stereochemistry unambiguously by X-ray analysis (Figure 2).



Figure 2. X-ray structure of racemic pentaacetate 4-azide analogue 4a.

Synthesis of meso-5-Deoxy-5-Azido myo-Inositols 5:

Starting with the dicyclohexylidene diol 6, benzylation of the two remaining hydroxyl groups afforded the fully protected inositol derivative 14 in excellent yield¹⁸ (Scheme 2). The kinetically labile and slightly distorted trans ketal of the compound 14 was cleaved selectively by controlled acid hydrolysis using acetyl chloride in DCM/MeOH (3:1) to afford the diol 15 in 68% yield.¹⁹ The diol 15 was regioselectively benzylated under phase transfer catalyst (tetrabutyl ammonium hydrogen sulphate) conditions, resulting in a separable mixture of benzylated products 16 and 17.20 Due to difficulties to distinguish 16 from 17, both alcohols were converted to their corresponding methylated derivatives 18a,b for identification purposes. Subsequent removal of the *cis*-ketal unit and global benzylation of 18a,b provided the fully protected inositol derivatives 20a and 20b, respectively. The ¹H and ¹³C NMR of the derived mesocompound (20b) clearly indicated the structure to be 5-O-methyl-1,2,3,4,6-penta-O-benzyl myo-inositol. Thus, the corresponding starting alcohol 17 was confirmed to possess the C5-free alcohol for the synthesis of the desired 5-series azido-analogues 5.



Scheme 2. Synthesis of *meso*-5-azido *myo*-inositol analogues 5a,b. Reagents and conditions: i. NaH, DMF, BnBr 0 °C, 97%; ii. CH₃COCl, DCM, MeOH (3:1), 68%; iii. BnBr, Bu4_NHSO₄, DCM, 5% NaOH, reflux; iv. NaH, DMF, MeI, 0 °C to rt, 90%; v. DCM/MeOH (1:1), CH₃COCl, rt, 89%; vi. (i); vii. DMP, NaHCO₃, DCM, rt, 85%; viii. NaBH₄, EtOH, rt, 60%; ix. Tf₂O, Py, 0 °C, 71%; x. NaN₃, DMF, rt, 83%; xi. Ac₂O, AcOH, 10% H₂SO₄ in Ac₂O, 60 °C, 64%; xii. NaOMe, MeOH, rt, 98%.

Having identified the C₅-OH derivative, the C₅stereochemistry of **17** was inverted by DMP oxidation followed by reduction of the ketone **21** to give the alcohol **22** in the *neo*configuration (Scheme 2). In the reduction step, the original alcohol **17** was also formed in 36 % yield, presumably due to similar sterics being encountered by the hydride reagent when approaching from either the α or β face in **21**. The alcohol **22** was then converted into its trifluoromethane sulfonate **23** by treatment with Tf₂O in pyridine. Next, the azide group was installed by a clean substitution in an S_N2 fashion with sodium azide to

regenerate the *myo*-configured product **24** in good yield. Interestingly, no elimination (enol) product was detected, presumably due to 1,3-diaxial sterics preventing E₂-elimination. Compound **24** was then subjected to exhaustive acetolysis to form the 5-deoxy-5-azido-*myo*-inositol pentaacetate **5a**.¹⁷ In this case, an elevated temperature 60 °C was required for complete conversion to the pentaacetate **5a**.⁶ The isomeric, partially acetolysed, benzyl ether products were also converted to **5a** under the same acetolysis conditions. Deacetylation of **5a** was accomplished by methanolysis in the presence of catalytic amounts of sodium methoxide to yield 5-deoxy-5-azido-*myo*inositol **5b**. Single crystals of compound **5a** from 1:1 hexane/ether confirmed the structure unambiguously by X-ray analysis (Figure 3).



Figure 3. X-ray structure of meso pentaacetate 5-azide analogue 5a.

Synthesis of (±)-3-Deoxy-3-Azido myo-Inositols 3:

Inspired by the report of Watanabe *et al.*^{16b} for an efficient $S_N 2$ substitution of C_3 -inositol triflates, the synthesis of the 3-series **3a,b** was studied by consecutive double substitution at the C_3 position of an inositol intermediate **26**, which was obtained from the previously synthesized inositol derivative **7** by subsequent methoxy methyl (MOM) ether protection to form the fully protected **25** and Pd-C hydrogenolysis (Scheme 3).²¹ The C_3 -free alcohol **26** was treated with trifluoromethane sulfonic acid anhydride in pyridine to yield the corresponding triflate **27**, which was immediately reacted with potassium acetate in DMA to furnish the C_3 -inverted *chiro*-acetate **28** in excellent yield (96%)^{16b} without elimination, presumably due to the less basic nature of the acyl anion as compared to azide species.

Deacetylation of chiro-28 in the presence of catalytic amounts of sodium methoxide gave the axial alcohol 29 (Scheme 3). After completion, acidic resin was added to the reaction mixture to remove sodium ions by ion exchange. Unexpectedly, the trans ketal unit of 29 rearranged into a more stable cis C3/C4-ketal (30) by ketal migration under the slightly acidic conditions. Hence, a basic aqueous work up procedure using ethylacetatewater was employed while scaling up. The free axial alcohol in 29 was subsequently treated with triflic anhydride and pyridine to form the triflate 31. Excess pyridine led to formation of an eliminated product (35) in minor amounts. Two equivalents of pyridine in dichloromethane, however, generated the trifluoromethane sulfonylated product 31 in 90% yield without elimination. On the other hand, the free axial alcohol of 29 could be smoothly mesylated in pyridine as the solvent to form 32 without elimination, presumably due to the lower leaving group aptitude of OMs as compared to OTf.²² The isolated triflate 31 was examined first. Treatment with sodium azide in DMF generated the myo-configured substitution product 33 in low yield (15%) together with a minor S_N1 substitution product 34 (10%) and the elimination product 35 in 56% yield.²³ In comparison, the mesylate derivative 32 failed to undergo azide displacement with sodium azide in DMF, even upon heating at 70 °C. Also, Mitsunobu reaction of the alcohol 29 in order to achieve direct azide displacement was unsuccessful.



Scheme 3. Synthesis of (±)-3-azido *myo*-inositol analogues **3a,b**. Reagents and conditions: i. MOMCl, DIPEA, 0 °C to rt, 87%; ii. EtOAc, THF, 20% Pd-C, H₂, 60%; iii. Tf₂O, Py, DCM, 0 °C, 78%; iv. KOAc, DMA, 70 °C, 96%; v. NaOMe, MeOH, rt, 97%; vi. (iii), 90%; vii. MeSO₂Cl, py, 94%; viii. NaN₃, DMF, rt; ix. **31**, TMSN₃, TBAF, THF, 43%.

We thus opted to optimise the azide displacement of triflate **31** by screening differing azide reagents and solvents: NaN₃/DMA, NaN₃/Me₂CO/H₂O, ⁿBu₄NN₃/DCM, TMSN₃/TBAF/THF. Eventually, the combination of excess TMSN₃ (8 equiv.) in the presence of (0.5 equiv.) of TBAF in THF medium provided *myo*-azide **33** in a 43% optimal yield.²⁴ To complete the 3-series, global acetolysis of compound **33** formed the desired 3-deoxy-3-azido *myo*-inositol pentaacetate **3a**⁵ in moderate yield (Scheme 3) under mildly acidic conditions (2% H₂SO₄ in acetic anhydride). Treatment of **33** in dry, ethereal HCl further allowed all acid labile protecting groups to be cleaved in one step and 3-deoxy-3-azido-*myo*-inositol **3b** could be isolated cleanly as a white solid after an ether wash.

Synthesis of meso-2-Deoxy-2-Azido myo-Inositols 2:

Following the orthoformate protection approach of Kishi et al.25, which allows for large differences in reactivity between equatorial and axial hydroxyl groups of inositol in terms of sterics and electronics,²⁶ a route to the 2-series of azide analogues was developed (Scheme 4). The symmetric dibenzyl mvo-inositol orthoformate 37^{27} was prepared from the dibenzoate derivative 36 through a prolonged silver(I) oxide chelation mediated bisbenzylation²⁸ and aminolysis sequence. The equatorial free hydroxyl group of 37 was oxidised under DMP condition to give the ketone 38 in excellent yield. Next, stereoselective reduction of ketone 38 produced the inverted axial alcohol 39 in the scylloconfiguration exclusively.²⁵ The rigidity of the orthoformate unit presumably accounts for this highly stereoselective reduction step. The axially positioned free alcohol of 39 was then sulfonylated²² by treatment with methane sulfonyl chloride in pyridine to give 40 in excellent yield. At this stage, an S_N^2 attack at C_2 of compound **40** was presumed to be sterically challenging. Hence, orthoformate cleavage of compound 40 was performed first, and the triol 41 was formed by mild methanolysis with pTSA.²⁹ Next, the mesylate derivative **41** was heated with sodium azide in DMF, which formed the myo-configured azide substituted product 42 in moderate yield, along with minor amounts of the epoxide 43 and some unreacted starting material 41. No elimination product was identified in this case; however, the reaction required heating at 80 °C, which led to decomposition of some material. Final exhaustive acetolysis of 42 completed the synthesis of the desired 2-deoxy-2-azido-myoinositol pentaacetate 2a in good yield. Clean methanolysis under

Tetrahedron

basic conditions regenerated the free hydroxyl groups to produce *meso* 2-deoxy-2-azido-*myo*-inositol **2b**.



Scheme 4. Synthesis of 2-azido *myo*-inositol analogues 2a,b. Reagents and conditions: i. (a) Ag_2O , BnBr, DMF, rt, (b) (CH₃)₂CHCH₂NH₂, MeOH, reflux, 70%; ii. DMP, DCM, rt, 95%; iii. MeOH, THF, NaBH₄, 87%; iv. CH₃SO₂Cl, py, 92%; v. *p*TSA, MeOH, 97%; vi. NaN₃, DMF, 80 °C; vii. 42, Ac₂O, AcOH, 15% H₂SO₄ in Ac₂O, 50 °C, 64%; viii. NaOMe, MeOH, 99%.

Summary

In this letter, we have described straighforward routes to various azido-deoxy inositol analogues through azide installation. As a common strategy, we adopted a consecutive $S_N 2$ doubleinversion approach on suitably protected inositol derivatives, which were derived via convenient oxidation-reduction sequences from myo-inositol. A final, global acetolysis step in the presence of the azide group (2-15% sulphuric acid in acetic anhydride/acid) enabled the direct and convenient synthesis of azido-inositol pentaacetate analogues 2a-5a. Methanolysis by treatment with catalytic sodium methoxide subsequently provided the fully unprotected azido myo-inositol surrogates 2b-5b cleanly. The routes are convenient and provide sufficient material for biological study. Having genuine azide-surrogates of myo-inositol in hand, a more concise and diversified strategy is under investigation. In the meantime, the metabolic incorporation of these modified azido inositol analogues into various inositol lipids of yeast cells are in progress, and lipid profiling and biosynthetic cell compatabilities will be reported in due course.

Acknowledgments

We thank the National University of Singapore for graduate Scholarships (to S.P. and S.M.I.). This work was funded (to M.R.W. and M.J.L.) by the Competitive Research Program (CRP) of the National Research Foundation (NRF) of Singapore (NRF-G-CRP 2007-04).

Supplementary data

Supplementary data associated with this article can be found, in the online version, at ...

References and notes

- Agranoff, B. W.; Bradley, R. M.; Brady, R.O. J. Biol. Chem. 1958, 233, 1077.
- 2. Shears, S. B. Mol. Cell. Biol. L 1998, 1436, 49.
- Hammond, G.; Thomas, C. L.; Schiavo, G. Curr. Top. Microbiol. 2004, 282, 177.

- (a) Paulick, M. G.; Bertozzi, C. R. *Biochemistry* 2008, 47, 6991; (b) Baker, G. R.; Billington, D. C.; Gani, D. *Bioorg. Med. Chem. Lett.* 1991, 1, 17.
- Podeschwa, M. A. L.; Plettenburg, O.; Altenbach, H. J. Org. Biomol. Chem. 2003, 1, 1919.
- (a) Sanfilippo, C.; Patti, A.; Piattelli, M.; Nicolosi, G. *Tetrahedron* Asymm. **1998**, *9*, 2809; (b) Serrano, P.; Llebaria, A.; Delgado, A. J. Org. Chem. **2005**, *70*, 7829.
- (a) Johnson, S. C.; Dahl, J.; Shih, T. L.; Schedler, D. J.; Anderson, L.; Benjamin, T. L.; Baker, D. C. *J. Med. Chem.* **1993**, *36*, 3628; (c) De Almeida, M. V.; Figueiredo, R. M.; Dos Santos, H. F.; Da Silva, A. D.; De Almeida, W. B. *Tetrahedron Lett.* **2001**, *42*, 2767.
- (a) Sureshan, K. M.; Ikeda, K.; Asano, N.; Watanabe, Y. *Tetrahedron Lett.* 2004, 45, 8367; (b) Riley, A. M.; Jenkins, D. J.; Potter, B. V. L. *Carbohyd. Res.* 1998, 314, 277; (c) Gurale, B. P.; Shashidhar, M. S.; Gonnade, R. G. J. Org. Chem. 2012, 77, 5801; (d) Murali, C.; Gurale, B. P.; Shashidhar, M. S. *Eur. J. Org. Chem.* 2010, 755.
- (a) Bruzik, K. S.; Myers, J.; Tsai, M.-D. Tetrahedron Lett. 1992, 33, 1009; (b) Martin, S. F.; Josey, J. A.; Wong, Y.-L.; Dean, D. W. J. Org. Chem. 1994, 59, 4805; (c) Rudolf, M. T.; Schultz, C. Liebigs Ann. 1996, 533 and references therein; (d) Mayer, T. G.; Schmidt, R. R. Liebigs Ann. 1997, 859; (e) Suzuki, T.; Tanaka, S.; Yamada, I.; Koashi, Y.; Yamada, K.; Chida, N. Org. Lett. 2000, 2, 1137; (f) Riley, A. M.; Correa, V.; Mahon, M. F.; Taylor, C. W.; Potter, B. V. L. J. Med. Chem. 2001, 44, 2108; (g) Sureshan, K. M.; Yamasaki, T.; Hayashi, M.; Watanabe, Y. Tetrahedron Asymm. 2003, 14, 1771.
- (a) Estevez, V. A.; Prestwich, G. D. J. Am. Chem. Soc. 1991, 113, 9885;
 (b) Chen, J.; Feng, L.; Prestwich, G. D. J. Org. Chem. 1998, 63, 6511;
 (c) Collins, P.; Ferrier, R. In Monosaccharides; Their chemistry and their roles in natural products: John Wiley & Sons Ltd.: West Sussex, England, 1995; pp. 449.
- (a) Angyal, S. J.; Tate, M. E.; Gero, S. D. J. Chem. Soc. 1961, 4116; (b) Angyal, S. J.; Irving, G. C.; Rutherfo, D.; Tate, M. E. J. Chem. Soc. 1965, 6662.
- (a) Angyal, S. J.; Macdonald, C. G. J. Chem. Soc. 1952, 686; (b) Leiserow, L.; Rabinovi, D. J. Chem. Soc. A 1969, 2367; (c) Gigg, J.; Gigg, R.; Payne, S.; Conant, R. Carbohyd. Res. 1985, 142, 132; (d) Desai, T.; Gigg, J.; Gigg, R.; Martinzamora, E.; Schnetz, N. Carbohyd. Res. 1994, 258, 135.
- 13. Baker, G. R.; Billington, D. C.; Gani, D. Tetrahedron 1991, 47, 3895.
- (a) Vacca, J. P.; Desolms, S. J.; Huff, J. R.; Billington, D. C.; Baker, R.; Kulagowski, J. J.; Mawer, I. M. *Tetrahedron* **1989**, *45*, 5679; (b) Dreef, C. E.; Tuinman, R. J.; Lefeber, A. W. M.; Elle, C. J. J.; Vandermarel, G. A.; Vanboom, J. H. *Tetrahedron* **1991**, *47*, 4709.
- Ballereau, S.; Guedat, P.; Poirier, S. N.; Guillemette, G.; Spiess, B.; Schlewer, G. J. Med. Chem. 1999, 42, 4824.
- (a) Tagliaferri, F.; Wang, S. N.; Berlin, W. K.; Outten, R. A.; Shen, T. Y. *Tetrahedron Lett.* **1990**, *31*, 1105-1108; (b) Sureshan, K. M.; Ikeda, K.; Asano, N.; Watanabe, Y. *Tetrahedron* **2008**, *64*, 4072.
- (a) van Dijkum, E.; Danac, R.; Hughes, D. J.; Wood, R.; Rees, A.; Wilkinson, B. L.; Fairbanks, A. J. Org. Biomol. Chem. 2009, 7, 1097; (b) Cao, Y.; Okada, Y.; Yamada, H. Carbohyd. Res. 2006, 341, 2219.
- Ozaki, S.; Watanabe, Y.; Ogasawara, T.; Kondo, Y.; Shiotani, N.; Nishii, H.; Matsuki, T. *Tetrahedron Lett.* **1986**, *27*, 3157.
- (a) Lu, P. J.; Gou, D. M.; Shieh, W. R.; Chen, C. S. *Biochemistry* 1994, 33, 11586; (b) Wang, D. S.; Hsu, A. L.; Song, X. Q.; Chiou, C. M.; Chen, C. S. J. Org. Chem. 1998, 63, 5430.
- 20. Jiang, C.; Moyer, J. D.; Baker, D. C. J. Carbohyd. Chem. 1987, 6, 319.
- 21. Lin, G.; Tsai, M. D. J. Am. Chem. Soc. 1989, 111, 3099.
- Fabris, F.; Rosso, E.; Paulon, A.; De Lucchi, O. *Tetrahedron Lett.* 2006, 47, 4835.
- Johnson, S. C.; Dahl, J.; Shih, T. L.; Schedler, D. J. A.; Anderson, L.; Benjamin, T. L.; Baker, D. C. *J. Med. Chem.* **1993**, *36*, 3628.
- Anilkumar, G.; Nair, L. G.; Olsson, L.; Daniels, J. K.; Fraser-Reid, B. *Tetrahedron Lett.* **2000**, *41*, 7605.
- 25. Lee, H. W.; Kishi, Y. J. Org. Chem. 1985, 50, 4402.
- Sureshan, K. M.; Shashidhar, M. S.; Praveen, T.; Das, T. Chem. Rev. 2003, 103, 4477.
- 27. Praveen, T.; Shashidhar, M. S. Carbohyd. Res. 2001, 330, 409.
- 28. Banerjee, T.; Srikantiah, S. M. Tetrahedron Lett. 1994, 35, 8053.
- Sureshan, K. M.; Das, T.; Shashidhar, M. S.; Gonnade, R. G.; Bhadbhade, M. M. *Eur. J. Org. Chem.* **2003**, 1035.

- Practical routes to all azido-deoxy • derivatives of myo-inositol are described.
- Regio- and stereo-controlled introduction of • azide functionality were achieved.
- Key scyllo-, chiro-, epi- and neo-inositols ٠ were prepared as intermediates.
- Acceleration A final, one-pot acetolysis gave convenient ٠