



Cavaliere, S., Malik, B. R., & Hodge, J. J. L. (2013). KCNQ channels regulate age-related memory impairment: KCNQ regulates age-related memory. PLoS ONE, 8(4), [e62445]. 10.1371/journal.pone.0062445

Link to published version (if available):
[10.1371/journal.pone.0062445](https://doi.org/10.1371/journal.pone.0062445)

[Link to publication record in Explore Bristol Research](#)
PDF-document

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
<http://www.bristol.ac.uk/pure/about/ebr-terms.html>

Take down policy

Explore Bristol Research is a digital archive and the intention is that deposited content should not be removed. However, if you believe that this version of the work breaches copyright law please contact open-access@bristol.ac.uk and include the following information in your message:

- Your contact details
- Bibliographic details for the item, including a URL
- An outline of the nature of the complaint

On receipt of your message the Open Access Team will immediately investigate your claim, make an initial judgement of the validity of the claim and, where appropriate, withdraw the item in question from public view.

KCNQ Channels Regulate Age-Related Memory Impairment

Sonia Cavaliere, Bilal R. Malik, James J. L. Hodge*

School of Physiology and Pharmacology, University of Bristol, Bristol, Avon, United Kingdom

Abstract

In humans KCNQ2/3 heteromeric channels form an M-current that acts as a brake on neuronal excitability, with mutations causing a form of epilepsy. The M-current has been shown to be a key regulator of neuronal plasticity underlying associative memory and ethanol response in mammals. Previous work has shown that many of the molecules and plasticity mechanisms underlying changes in alcohol behaviour and addiction are shared with those of memory. We show that the single KCNQ channel in *Drosophila* (dKCNQ) when mutated show decrements in associative short- and long-term memory, with KCNQ function in the mushroom body α/β neurons being required for short-term memory. Ethanol disrupts memory in wildtype flies, but not in a KCNQ null mutant background suggesting KCNQ maybe a direct target of ethanol, the blockade of which interferes with the plasticity machinery required for memory formation. We show that as in humans, *Drosophila* display age-related memory impairment with the KCNQ mutant memory defect mimicking the effect of age on memory. Expression of KCNQ normally decreases in aging brains and KCNQ overexpression in the mushroom body neurons of KCNQ mutants restores age-related memory impairment. Therefore KCNQ is a central plasticity molecule that regulates age dependent memory impairment.

Citation: Cavaliere S, Malik BR, Hodge JLL (2013) KCNQ Channels Regulate Age-Related Memory Impairment. PLoS ONE 8(4): e62445. doi:10.1371/journal.pone.0062445

Editor: Giorgio F. Gilestro, Imperial College London, United Kingdom

Received: January 25, 2013; **Accepted:** March 21, 2013; **Published:** April 30, 2013

Copyright: © 2013 Cavaliere et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by grants from the EU FP7 Marie-Curie (IRG200632), Royal Society (2008/R1), BBSRC (BB/G008973/1), and from a Wellcome PhD program (083361). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: james.hodge@bristol.ac.uk

Introduction

KCNQ (Kv7) channels mediate a range of important physiological functions and are a hotspot of genetic diseases and therefore target for existing and novel drug treatments. In human cardiac muscle, *KCNQ1* loss of function mutations result in the most common form of cardiac arrhythmia, Long QT syndrome, while gain of functions mutations cause Short QT and atrial fibrillation [1,2]. *KCNQ1* mutations also result in adult onset type II diabetes [3,4]. In the nervous system *KCNQ3* can heteromultimerise with either *KCNQ2* or *KCNQ5* subunits to form a channel that mediates a M-current, a current that is suppressed by muscarinic acetylcholine receptor activation. Because the M-current operates at resting membrane potential it is well poised to regulate membrane excitability so that when it is open it acts as a brake on action potential firing while if it is suppressed it increases neural activity and neurotransmitter release [5,6]. These features and its broad neuronal expression allow KCNQ channels to have an important function in synaptic plasticity and memory, alcohol response and nociception [2,7,8]. *KCNQ2* or *KCNQ3* loss-of-function mutations result in a developmental form of epilepsy called Benign familial neonatal convulsions [2,5]. *KCNQ4* loss-of-function mutations are a common cause of autosomal dominant deafness and age-dependent hearing impairment [9,10]. M-current inhibitors increase excitability of cholinergic neurons and have shown some promise as cognitive enhancers in models of dementia. Conversely, M-current openers are of great interest as anti-convulsants, analgesics and treatments of psychiatric diseases

[2,5]. In mice expression of human dominant negative *KCNQ2* transgene in hippocampal neurons increases neural excitability and results in associative memory deficits [7].

Drosophila has a single KCNQ (dKCNQ) channel that is most highly expressed in the nervous system [11–13], but like mammalian *KCNQ1* [2] is also expressed in the heart. dKCNQ encodes a slowly activating and deactivating Kv current that can be suppressed by muscarinic acetylcholine receptor agonists and hence is an M-current [12,14]. dKCNQ has been shown to have important age-dependent cardiac function, with hearts from young *dKCNQ* loss-of-function mutant flies showing arrhythmias similar to those seen in aged wildtype flies, whose hearts shows age dependent reduction in *dKCNQ* expression [13]. dKCNQ has many features of the M-current including conserved acute block by low concentrations of ethanol and broad neuronal expression [15]. Furthermore, targeted expression of *KCNQ-RNAi* in *Drosophila* neurons increased neural excitability, while KCNQ overexpression decreased excitability *in vivo*. *dKCNQ* loss-of-function mutant flies increased ethanol sensitivity and tolerance with acute activation of dopaminergic neurons by heat-activated TRP channel or *KCNQ-RNAi* expression shown to produce ethanol hyperexcitability [15]. In this study we characterise the role of *dKCNQ* mutants on memory, showing that *dKCNQ* expression decreases in the brain with aging and is linked to age-dependent cognitive deficits.

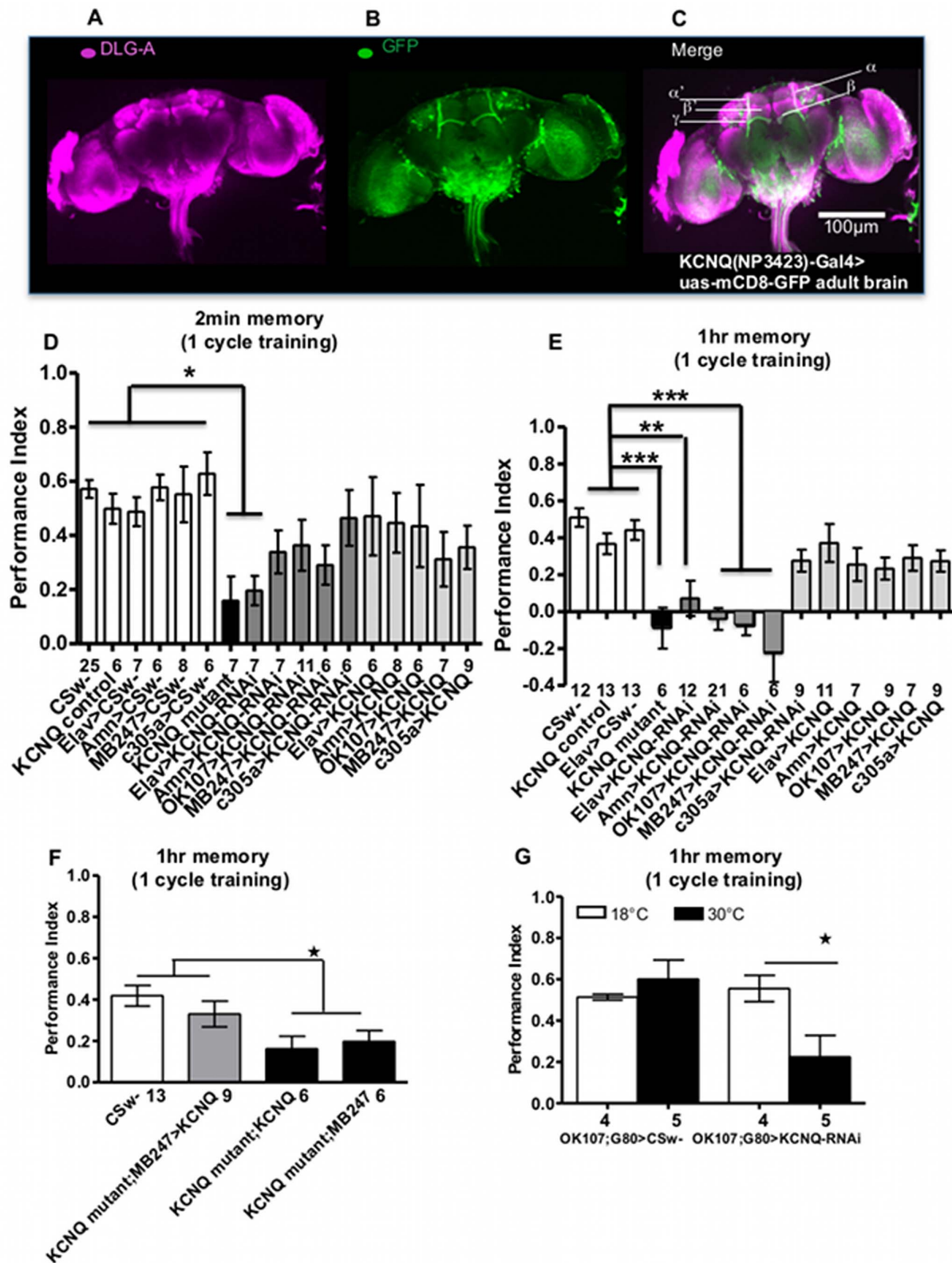


Figure 1. KCNQ signalling is required in the mushroom body α and β neurons for short-term memory. **A–C.** Adult brains containing a Gal4 enhancer trap (KCNQ^{NP3423}) in the KCNQ gene locus revealed broad neuronal expression of KCNQ (labelled by membrane bound GFP in green (**B**)) especially in the fly memory structures of the mushroom body α and β neurons and surrounding neurons known to be visualised by DLG-A (Ruiz-Cañada et al., 2002) staining (in magenta (**A**), co-localised structures in white (**C**)). **D.** Initial (2 min) memory was reduced in the KCNQ mutant (black bar) and flies with reduced KCNQ levels (dark grey bars) in all neurons (Elav-Gal4, uas-KCNQ-RNAi) ($p < 0.05$) compared with controls (CSw-, KCNQ control, and Gal4, +, white bars) but did not lead to any change in memory ($p > 0.05$) between the remaining genotypes. **E.** KCNQ mutants and flies with reduced KCNQ in the mushroom body (OK107-Gal4 or MB247-Gal4, uas-KCNQ-RNAi), DPM (amn-Gal4, uas-KCNQ-RNAi) ($p < 0.001$) or all (Elav-

Gal4, $p < 0.01$) neurons have a significant reduction in 1 hr STM compared to controls (CSw-, KCNQ control and Gal4, +), while KCNQ overexpression (light grey bars) had no effect ($p > 0.05$) with these promoters. **F.** Mushroom body α/β neuron expression of the KCNQ transgene in the KCNQ mutant background (KCNQ mutant; MB247-Gal4, *uas-KCNQ*) rescued the KCNQ mutant memory deficit with its memory being greater ($p < 0.05$) than KCNQ mutant with Gal4 or *uas* alone (KCNQ mutant; MB247-Gal4 and KCNQ mutant; *uas-KCNQ*) but statistically indistinguishable ($p > 0.05$) from control (CSw- wildtype) levels. Data in D-F were analysed by 1-way ANOVA with Bonferroni post-hoc test. **G.** 1 hr memory was measured in OK107-Gal4, Gal80^{ts}, *uas-KCNQ-RNAi* and OK107-Gal4, Gal80^{ts}, CSw- control flies raised at 18°C throughout development and then tested at 18°C conditions that prevent transgene expression (white bars). These scores were compared to the 1 hr memory of the same genotypes raised at 18°C throughout development and then shifted to 30°C allowing KCNQ transgene expression 2 days prior and during behavioural testing (black bars). 2-way ANOVA indicates significant differences due to interaction between temperature and genotype ($p = 0.0195$). Post-hoc analysis showed OK107-Gal4, Gal80^{ts}, *uas-KCNQ-RNAi* had less ($p < 0.05$) memory at 30°C compared to flies at 18°C flies (~100 flies per n). In this and all subsequent figures, error bars represent SEM with no asterisk $p > 0.05$, * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$. n is denoted by the number between the x axis and genotype names with experiments performed on multiple different days (~100 flies were used per n, unless otherwise stated). doi:10.1371/journal.pone.0062445.g001

Materials and Methods

Drosophila Stocks

The KCNQ deletion mutant contained an imprecise excision of the *EP2074* element (*KCNQ*¹⁸⁶) that removes all the 5' and transmembrane regions of the channel and therefore is a null [13]. The KCNQ control was a precise excision of the element (*KCNQ*⁹⁷) leaving the gene completely intact [13]. *uas-KCNQ* flies allowed Gal4 promoter driven overexpression of KCNQ [13] while *uas-KCNQ-RNAi* (Bloomington stock 27252) allowed Gal4 targeted knockdown of the channel. The KCNQ stocks were kind gifts of Dr Rolf Bodmer. *Elav-Gal4*, *uas-mCD8-GFP* and *OK107-Gal4* [16] were gifts from Dr Leslie Griffith. *OK107-Gal4*, *Gal80^{ts}* [17] was a gift from Dr Yi Zhong. *c305a-Gal4* [18], *MB247-Gal4* [19] and *Ann(c316)-Gal4* [20] stocks were gifts of Dr Scott Waddell. Wildtype flies were *Canton S w-* (*CSw-*) from a stock previously maintained in the Waddell lab. All KCNQ mutant, Gal4 and *uas* lines were out crossed with the relevant *CSw-* line prior to behavioural analysis. All genotypes and all other crosses were raised on corn-meal agar medium at 22±2°C and 60±10% humidity under 12:12 hr light-dark cycle.

Immunohistochemistry

Adult fly brains were dissected in HL3.1 (70 mM NaCl, 5 mM KCl, 10 mM NaHCO₃, 115 mM sucrose, 4 mM MgCl₂, 5 mM trehalose, 1.5 mM CaCl₂, and 5 mM HEPES, pH 7.3) and isolated brains were fixed in 4% paraformaldehyde in HL3.1 for 30 min before being washed in HL3.1 [21]. The samples were permeabilised in HL3.1 with 0.1% triton X (HL3.1-Tx) for 1 hr, and then blocked for 1 hr in HL3.1-Tx with 0.1% BSA and 2% normal donkey serum (HL3.1-Tx-BSA-NDS). In order to the visualise the mushroom body and antennal lobe, brains were

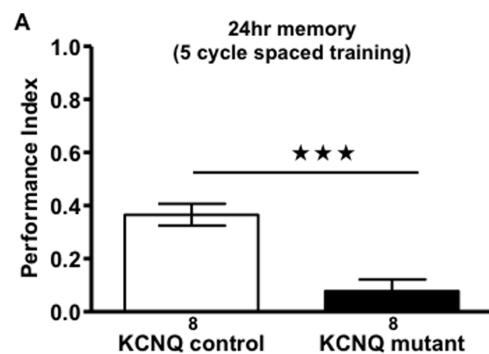


Figure 2. KCNQ signalling is required for long-term memory. A. 5 cycles of spaced training produces 24 hr LTM in KCNQ control flies that is absent ($p < 0.001$) in the KCNQ mutants. Data were analysed with unpaired t-test. doi:10.1371/journal.pone.0062445.g002

incubated with (1:2000) rabbit anti-*Drosophila* DLG (PDZ1-2) a protein known to be highly expressed in these memory-related structures [22] overnight at 4°C in HL3.1-Tx-BSA-NDS. After washing three times in HL3.1-Tx for 20 min, the brains were incubated with anti-rabbit Alexa-648 conjugated secondary antibody (1:400 in HL3.1-Tx-BSA-NDS) for 2 hr at room temperature. Finally the brains were washed three times HL3.1-Tx before being mounted in Vectorshield (Vector Laboratories). Samples were stored at 4°C in the dark until examination using a Leica TCS SP5 confocal microscope. The endogenous KCNQ expression pattern was determined by visualising membrane targeted GFP expressed using *KCNQ-Gal4* reporter lines (*KCNQ*^{NP3423}-*Gal4*, *uas-mCD8-GFP*).

Olfactory Aversive Conditioning

All experiments were performed at 25°C and 70% humidity under red light using the olfactory aversive conditioning protocol [23]. Groups of ~100 1–4 day old male and female flies received either 1 cycle of training during which they were exposed sequentially to one odour (conditioned stimulus, CS+; 3-octanol (1:74) or 4-methylcyclohexanol (1:57) diluted in mineral oil) for 1 min paired with electric 60 V DC shock (US) and then to a second odour (CS-; the reciprocal odour) for 1 min without electric shock separated by a 30 sec rest period when they were exposed to fresh air. Memory was measured after 1 (~2 min memory) training session at the choice point of the T-maze. To measure STM, flies were trained with 1 training cycle were stored for 1 hr and then allowed to distribute in the T-maze. LTM was assessed by giving flies either 5 cycles of spaced training cycles separated by 15 min rest intervals and then storing the flies for 24 hr before distribution in the T-maze. A performance index (PI) was calculated as the number of flies that distributed in the CS- arm minus the flies in the CS+ arm, divided by the total number of flies. Therefore a PI of 1.0 would be equivalent of 100:0 distribution where all the flies avoided the CS+ (perfect memory), while a 50:50 distribution would give a PI of zero (no memory). To test the effect of ethanol on *Drosophila* learning, flies were kept in bottles containing instant media (Formula 4–24 (R); Carolina biological supply company, Burlington, NC, USA) containing 10% ethanol in water containing a small amount of blue dye (0.05% Bromophenol blue) ~12 hr before testing. The controls were given the same water-blue dye solution but lacking ethanol. The blue dye was used to monitor whether the flies had actually drunk the ethanol solution; this was confirmed as all the flies had blue abdomens prior to the test. For *OK107-Gal4*, *Gal80^{ts}* experiments [17] flies were raised at 18°C and then shifted to 30°C allowing KCNQ transgene expression 1–2 days prior and during behavioural testing. Olfactory acuity was quantified by exposing naive flies to the odour versus air in the T-maze during a 2 min test trial. The performance index was calculated by counting the number of flies avoiding odour divided by total

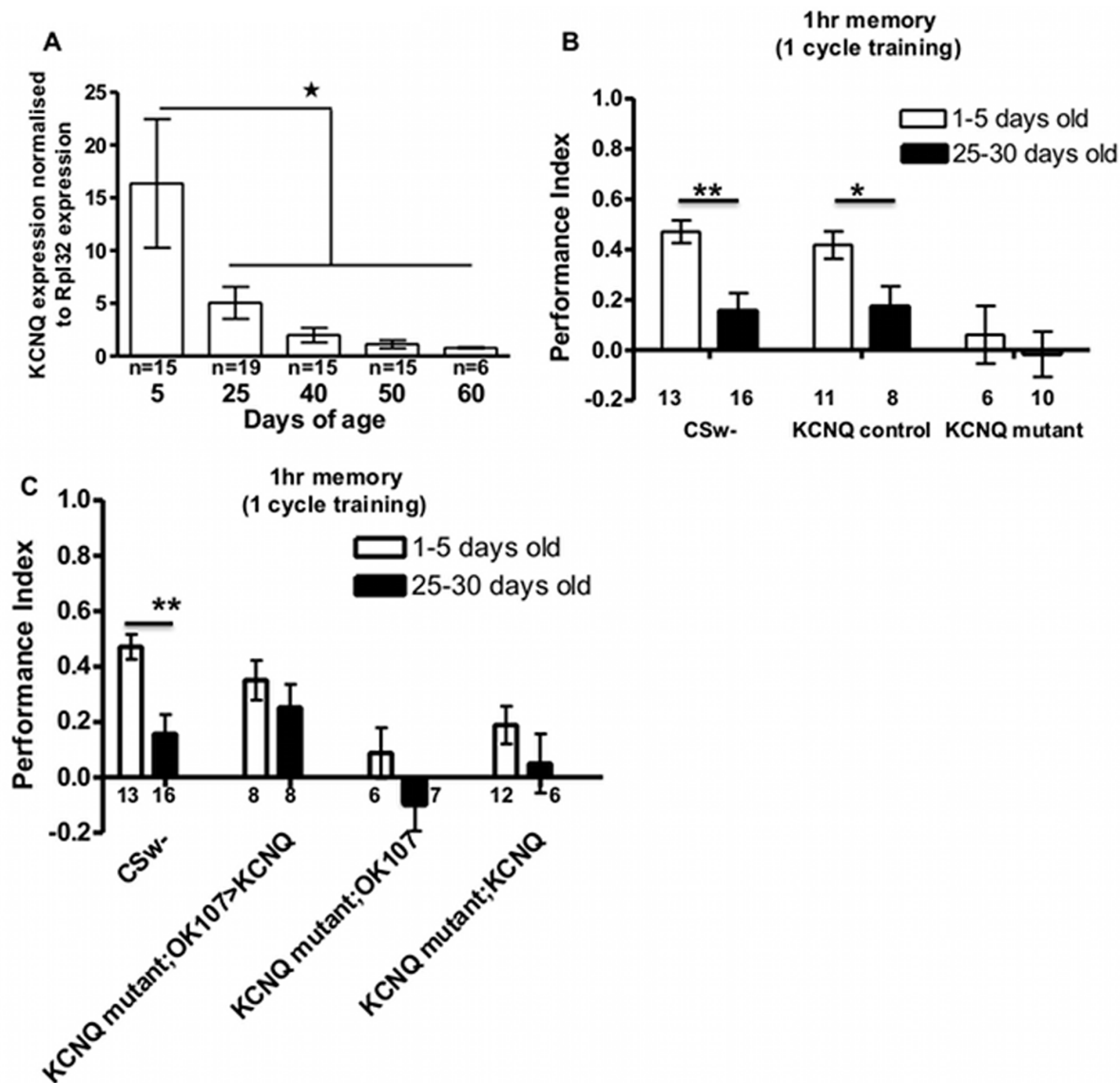


Figure 3. KCNQ mediates age-related memory impairment. **A.** Quantitative RT-PCR data show a dramatic age dependent reduction ($p < 0.05$) in KCNQ expression in adult brains (20 flies per n). **B.** 1 hr memory after 1 cycle training was compared between young (1–5 days old, white bars) and aged (25–30 days, black bars) adults. 2-way ANOVA indicates significant differences in memory due to age ($p = 0.0013$) and genotype ($p = 0.0008$). Post-hoc analysis revealed that memory becomes significantly impaired in aged as opposed to young CSw- wildtype ($p < 0.01$) and KCNQ control ($p < 0.05$) flies. KCNQ mutant flies had equally low ($p > 0.05$) memory whether young or old. **C.** Overexpression of KCNQ in the mushroom body rescues memory impairment of young and old KCNQ mutant flies. 2-way ANOVA indicates significant differences in memory due to age ($p < 0.01$) and genotype ($p < 0.001$). Post-hoc analysis revealed that memory becomes significantly impaired in aged as opposed to young CSw- wildtype ($p < 0.01$), while the memory of KCNQ mutant; OK107-Gal4, *uas-KCNQ* rescue flies stays similarly high ($p > 0.05$) in young and old flies as opposed to KCNQ mutant with Gal4 or *uas* alone (KCNQ mutant; OK107-Gal4 or KCNQ mutant; *uas-KCNQ*) whose memory was similarly low in young and old flies ($p > 0.05$).

doi:10.1371/journal.pone.0062445.g003

number flies. Shock reactivity was quantified by placing grids in each arm of the T-maze, and applying shock via the grid in one arm of the maze during a 2 min test trial. The performance index was calculated by counting the number of flies avoiding shock divided by total number flies [24]. Ethanol avoidance was quantified by placing a solution of 40% ethanol in the odour cup of one arm of the T-maze during a 2 min test trial. The performance index was calculated by counting the number of flies avoiding 40% ethanol divided by total number flies. All statistical analysis for behavioural data were performed and plotted with Graphpad Prism software.

Quantitative RT-PCR

Age-matched flies were frozen in liquid nitrogen and decapitated by vortexing. Heads were collected and an equal number of heads from each genotype were homogenised. Trizol was added directly in to the homogenised heads and RNA was extracted according to manufacturer's instructions (Invitrogen). RNA was DNAase treated (Ambion Inc) and reverse-transcribed (Retroscrip, Ambion). *KCNQ* mRNA was measured using a TaqMan Kit (*KCNQ* - Dm01846741_g1) and was normalised to *Rpl23* (*Rpl23*-Dm02151827_g1) mRNAs as a control allowing standardisation between the samples for aging experiments. The cDNA

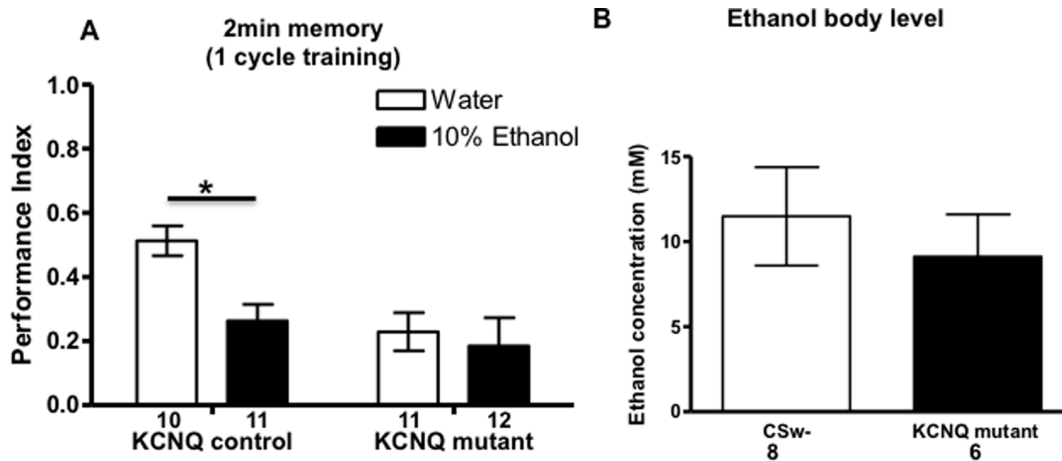


Figure 4. Ethanol disrupts memory in wildtype flies an effect removed by the KCNQ mutation. **A.** Flies received an overnight (~12 hr) exposure to 10% ethanol and were tested for 2 min memory. 2-way ANOVA showed a significant effect due to genotype and ethanol ($p < 0.05$). Post-hoc analysis showed that in KCNQ control ethanol caused a reduction ($p < 0.05$) in memory compared to water. The reduction in memory was removed by the KCNQ mutation that had similarly low memory with or without ethanol ($p > 0.05$). **B.** Ethanol content of KCNQ mutant and control (CSw- wildtype) flies exposed to 10% ethanol solution for ~12 hr was similar ($p > 0.05$, unpaired t-test, 20 flies per n). doi:10.1371/journal.pone.0062445.g004

concentration was measured using Roche's Light Cycler system and using multiplexing on a Stratagene Mx3000P system (Stratagene). All statistical analysis of data were performed and plotted with Graphpad Prism software.

Results

KCNQ Signalling Regulates Mushroom Body Dependent Associative Memory

Adult flies were tested for associative memory using the olfactory aversive conditioning [23]. Anatomically, neurons critical for *Drosophila* memory include those of the mushroom body that are labelled by *OK107-Gal4* [16]; these can be subdivided into α , β and γ neurons labelled by *MB247-Gal4* (which expresses strongly in α/β neurons but only weakly in γ neurons, [19]) and α'/β' neurons labelled by *c305a-Gal4* [18]. The mushroom body-associated dorsal paired medial (DPM) neurons are also important for memory. They express the memory gene *amnesiac* (*amn*) and can be labelled by *amn(c316)-Gal4* that expresses in the DPM [20]. KCNQ is broadly expressed in the brain [12,15]. Using a *Gal4* promoter enhancer trap within the KCNQ gene to drive GFP expression (Figure 1B), it appears *KCNQ* is expressed in the adult mushroom body α/β and surrounding neurons (Figure 1C), structures that are known to mediate memory formation and ethanol behaviour [24–26]. Compared with controls (Figure 1D), the *KCNQ* mutant had reduced initial memory 2 min after training, as did flies with pan-neural *KCNQ* knockdown. *KCNQ* mutants and flies with pan-neural, DPM or mushroom body neuron deficient *KCNQ* completely lacked the ability to form short-term memory (STM) assessed 1 hr after training (Figure 1E).

In order to map this STM phenotype further, we selectively knocked-down *KCNQ* in different parts of the mushroom body, finding that the α/β neurons that appear to express KCNQ (Figure 1C) were required for KCNQ's role in STM (as opposed to α'/β' neurons (Figure 1E) which do not seem to express KCNQ (Figure 1C)). *KCNQ* overexpression in any part of the mushroom body, DPM or all neurons did not change memory measured at 2 min or 1 hr (Figure 1D–E). We also found that expression of *KCNQ* in α/β neurons using *MB247-Gal4* in a fly otherwise completely lacking *KCNQ* rescued the *KCNQ* mutant STM defect

to normal (Figure 1F). Acute reduction of *KCNQ* levels in the mushroom body was sufficient to decrease 1 hr memory compared with controls (Figure 1G), showing that KCNQ is required post developmentally to mediate physiological changes underlying memory. We then wished to determine the role of KCNQ in long-term memory (LTM) which is formed after spaced training and lasts about 7 days and is protein synthesis and CREB dependent [17,27]. We found that the *KCNQ* mutant showed a drastic reduction in LTM (Figure 2A). No difference in ability of the flies to sense the odour (Figure S1A–B) or shock (Figure S1C) was observed between genotypes, showing that *KCNQ* mutants do not change peripheral sensory processing, but rather the memory defects are due to loss of KCNQ function in the mushroom body.

Drosophila Display Age Dependent Memory Deficits that are Rescued by Mushroom Body KCNQ Expression

Fly associative memory is known to decrease with age [28] and *KCNQ* expression decreases in old fly hearts leading to age related cardiac impairments [13]. Therefore we decided to test whether or not KCNQ was involved in age dependent memory decline. We first determined *KCNQ* expression in heads over the lifespan of *Drosophila* (~50 days, [28]) using quantitative RT-PCR. We found that by 25 days *KCNQ* expression had declined to about 10% of the level of 5 day olds (Figure 3A). Therefore, we tested STM of young (1–5 day old) as opposed to aged (25–30 day old) flies. Whereas control flies displayed an age dependent decrease in 1 hr STM (Figure 3B), *KCNQ* nulls were completely unable to form STM whether they were young or old. Previous experiments have implicated the *amn* DPM and mushroom body neurons in mediating the effect of age on 1 hr memory [28–30], with expression of a *PKA* transgene in mushroom body neurons restoring age-related memory impairment. We therefore over-expressed KCNQ in the mushroom body (Figure 3C) of *KCNQ* mutants and demonstrate rescue of age-dependent memory impairment. These experiments are consistent with decreases in KCNQ signalling being central forage dependent decrements in memory. This experiment also confirms that in young flies expression of *KCNQ* using mushroom body *Gal4* lines (Figure 1F and 3C) in a fly otherwise completely lacking *KCNQ* rescues the *KCNQ* mutant STM defect to normal.

Ethanol Disrupts Memory an Effect Mimicked by KCNQ Mutation

Previous work has shown that many of the molecules and plasticity mechanisms underlying changes in ethanol behaviour and addiction are shared with those of associative memory with ethanol known to disrupt synaptic plasticity and memory in humans [29,30]. Furthermore ethanol has been demonstrated to directly inhibit the M-current in dopaminergic neurons of the ventral tegmental area (VTA) a region of the brain important for ethanol reinforcement [26,31]. Likewise it has been shown that dKCNQ shows conserved blockade by ethanol, with reduction in KCNQ function causing increased ethanol sensitivity and tolerance via changes in dopamine neurons [15]. Consequently we investigated whether or not ethanol disrupted fly memory. Wildtype flies were exposed to 10% ethanol solution for ~12 hr and then immediately tested for 2 min memory (Figure 4A), although the flies did not appear sedated or intoxicated after the exposure or during the memory test, ethanol was found to reduce their memory. As KCNQ maybe a direct target of ethanol that is required for memory, we tested the *KCNQ* mutant and found that this resulted in a loss of the reduction in memory. This suggests that KCNQ is the plasticity molecule blocked by ethanol interfering with memory. No change in ethanol content was found between genotypes and was ~10 mM at the time of the memory test (Figure 4B), this would be sufficient to block a significant proportion of neural KCNQ channels ($IC_{50} = 19.8$ mM, [15]).

Discussion

Drosophila KCNQ displays conserved electrophysiological and pharmacological properties with mammalian neuronal KCNQ2/3 channels, with both mediating a slowly activating and non-inactivating Kv current called a M-current due to its suppression by muscarinic acetylcholine receptors [12,14]. In the hippocampus, expression of a dominant negative human KCNQ2 transgene was found to suppress the M-current [7]. This resulted in a decrease in the afterhyperpolarization (after a train of action potentials, the increase in Ca^{2+} activates K^+ channels leading to a pronounced hyperpolarization) and caused deficits in associative memory. Kv channels have also been implicated in plasticity underlying fly memory [32–35] consistent with the conspicuous expression of Kv currents in mushroom body neurons [34,36,37]. We have extended these studies by showing dKCNQ has a role in immediate memory, STM and LTM without peripheral defects. Targeted reduction of *KCNQ* in the mushroom body α/β neurons was sufficient to reduce memory, while expressing *KCNQ* in the same neurons in a fly otherwise completely lacking *KCNQ* rescued the *KCNQ* mutant memory phenotype to normal (Figure 1F). These results show KCNQ is required in the mushroom body α/β neurons in order for the fly to form STM. We found that the KCNQ memory phenotype resulted from reduced KCNQ function in the adult mushroom body (Figure 1G), showing that the role of KCNQ in formation is an acute physiological one as opposed to a developmental one. Mushroom body and DPM neurons have been suggested to respond to acetylcholine [27,38–42] and muscarinic acetylcholine receptor activation is known to suppress the dKCNQ current [12,14]. Therefore cholinergic stimulation would be expected to close dKCNQ channels causing depolarization and increasing firing and/or release from the DPM and mushroom body neurons. This might be expected to result in induction of plasticity and strengthening of recurrent activity in the DPM-mushroom body loop that could potentially consolidate memory. In addition the DPM neurons are thought to be

serotonergic with serotonin playing an important role in the DPM mediated memory [43], interestingly serotonin has been shown inhibit KCNQ currents in mammalian neurons [44,45], it is not known if this is also true for KCNQ currents in *Drosophila* neurons.

The effect of KCNQ mutation on LTM maybe through disruption of appropriate changes in resting membrane potential of memory neurons, which are required to remove the Mg^{2+} block of *Drosophila* NMDA glutamate receptors that is necessary for LTM and CREB-dependent gene expression [46].

On the basis of our data, high ethanol levels would be expected to cause significant or complete KCNQ blockade [15], disrupting synaptic plasticity and memory formation, thereby leading to alcohol induced amnesia or blackout. Consistent with this proposition we found that ethanol disrupts fly memory, an effect that was removed in the *KCNQ* mutant background (Figure 4A). This suggests that KCNQ is a key molecule that ethanol interacts with in the plasticity machinery involved in memory. Given the conserved role of mammalian KCNQ in ethanol response and memory [15], we suggest that it is likely this will also be the case for mammals.

Fly memory is known to reduce with age [28], as does *KCNQ* expression and function in the heart [13]. We found that *KCNQ* brain expression dramatically decreases with age and this is accompanied by an age-dependent decrease in memory (Figure 3A). Young *KCNQ* mutant flies that have low levels of *KCNQ* comparable to the low levels of *KCNQ* in aged wildtype flies have comparably reduced levels of memory. As mushroom body neurons are known to be important for mediating changes in memory performance with age [28,29] and mushroom body knockdown of *KCNQ* completely removed 1 hr memory (Figure 1E), we overexpressed a *KCNQ* transgene in the mushroom body (Figure 3C) of a fly otherwise completely lacking *KCNQ* and found that this restored age-related memory impairment with young and old flies having similarly high memory. In summary we show reduction in KCNQ function in mushroom body neurons mediates age-dependent cognitive decline.

In mammals, KCNQ specific modulators have been suggested to alleviate memory deficits associated with age related memory diseases such as Alzheimer's disease [2]. It is not clear how KCNQ-mediated mechanisms may affect memory in aged animals. However, based on the contribution of KCNQ2/3 to hippocampal afterhyperpolarizations and memory [7], one candidate mechanism would involve reduced neuronal KCNQ, as this modulates afterhyperpolarization duration that is known to change with memory and in aged animals [47–49]. Recently, KCNQ channels have been implicated in age-dependent decrements in the memory of primates [50], suggesting that KCNQ function in cognitive impairments accompanying aging are likely conserved from flies to humans. Furthermore, PKA signalling has been implicated in age-related memory impairment in flies and mammals [28,29,50,51] with the mammalian KCNQ channel open state being increased by PKA [50,52], suggesting that the mushroom body neuron KCNQ-mediated memory and age-dependent memory defects maybe due to an interaction with PKA.

The genetic and experimental tractability of *Drosophila* combined with its ~50 day lifespan and molecular conservation with human make it a convenient and powerful genetic model [28] to study further the age-dependent KCNQ cognitive deficits. We have shown that KCNQ neuronal function in memory and ethanol response are evolutionarily conserved with mammals, allowing further development of *Drosophila* models of KCNQ neuronal function and channelopathies to elucidate KCNQ

signalling networks, mechanisms of aging and potential screening for new disease therapies.

Supporting Information

Figure S1 A. *KCNQ* channel mutants display normal olfactory acuity and shock reactivity. Experimental and control (*Csw*-wildtype or *KCNQ* control) flies similarly ($p > 0.05$) avoided OCT **A.** and MCH **B.** odour used in the memory assay. **C.** Experimental and control (*Csw*-wildtype or *KCNQ* control) flies similarly ($p > 0.05$) avoided the arm of the T-maze delivering 60 V DC electric shock. Data in A-C were analysed by 1-way ANOVA with Bonferroni post-hoc test. (TIFF)

References

- Ashcroft FM (2006) From molecule to malady. *Nature* 440: 440–7.
- Wulff H, Castle NA, Pardo LA (2009) Voltage-gated potassium channels as therapeutic targets. *Nat Rev Drug Disc* 8: 982–1001.
- Unoki H, Takahashi A, Kawaguchi T, Hara K, Horikoshi M, et al. (2008) SNPs in *KCNQ1* are associated with susceptibility to type 2 diabetes in East Asian and European populations. *Nat Genet* 40: 1098–102.
- Yasuda K, Miyake K, Horikawa Y, Hara K, Osawa H, et al. (2008) Variants in *KCNQ1* are associated with susceptibility to type 2 diabetes mellitus. *Nat Genet* 40: 1092–1097.
- Delmas P, Brown DA (2005) Pathways modulating neural *KCNQ/M* (Kv7) potassium channels. *Nat Rev Neurosci* 6: 850–862.
- Huang H, Trussell LO (2011) *KCNQ5* channels control resting properties and release probability of a synapse. *Nat Neurosci* 4: 840–848.
- Peters HC, Hu H, Pongs O, Storm JF, Isbrandt D (2005) Conditional transgenic suppression of M channel in mouse brain reveals functions in neuronal excitability, resonance and behaviour. *Nat Neurosci* 8: 51–60.
- Koyama S, Brodie MS, Appel SB (2007) Ethanol inhibition of M-current and ethanol induced direct excitation of Ventral Tegmental Area dopamine neurons. *J Neurophysiol* 97: 1977–85.
- Kubisch C, Schroeder BC, Friedrich T, Lutjohann B, El Amraoui A, et al. (1999) *KCNQ4*, a novel potassium channel expressed in sensory outer hair cells, is mutated in dominant deafness. *Cell* 96: 437–446.
- Van Eyken E, Van Laer L, Franssen E, Topsakal V, Lemkens N, et al. (2006) *KCNQ4*: a gene for age-related hearing impairment? *Human Mutation* 27: 1007–1016.
- Littleton JT, Ganetzky B (2000) Ion channels and synaptic organization: analysis of the *Drosophila* genome. *Neuron* 26: 35–43.
- Wen H, Weiger TM, Ferguson TS, Shahidullah M, Scott SS, et al. (2005) A *Drosophila* *KCNQ* channel essential for early embryonic development. *J Neurosci* 25: 10147–10156.
- Ocorr K, Reeves NL, Wessells RJ, Fink M, Chen HS, et al. (2007) *KCNQ* potassium channel mutations cause cardiac arrhythmias in *Drosophila* that mimic the effects of aging. *Proc Natl Acad Sci USA* 104: 3943–3948.
- Cavaliere S, Hodge JJ (2011) *Drosophila* *KCNQ* channel displays evolutionarily conserved electrophysiology and pharmacology with mammalian *KCNQ* channels. *PLoS One* 6: e23898.
- Cavaliere S, Gillespie JM, Hodge JJ (2012) *KCNQ* channels show conserved ethanol block and function in ethanol behaviour. *PLoS One* 6: e23898.
- Connolly JB, Roberts IJ, Armstrong JD, Kaiser K, Forte M, et al. (1996) Associative learning disrupted by impaired Gs signaling in *Drosophila* mushroom bodies. *Science* 274: 2104–2107.
- Shuai Y, Lu B, Hu Y, Wang L, Sun K, et al. (2010) Forgetting is regulated through Rac activity in *Drosophila*. *Cell* 140: 579–589.
- Krashes MJ, Keene AC, Leung B, Armstrong JD, Waddell S (2007) Sequential use of mushroom body neuron subsets during *Drosophila* odor memory processing. *Neuron* 53: 103–115.
- Zars T, Fischer M, Schulz R, Heisenberg M (2000) Localization of a short-term memory in *Drosophila*. *Science* 288: 672–675.
- Waddell S, Armstrong JD, Kitamoto T, Kaiser K, Quinn WG (2000) The amnesiac gene product is expressed in two neurons in the *Drosophila* brain that are critical for memory. *Cell* 103: 805–811.
- Hodge JJ, Mullasseril P, Griffith LC (2006) Activity-dependent gating of CaMKII autonomous activity by *Drosophila* CASK. *Neuron* 51: 327–337.
- Ruiz-Cañada C, Koh YH, Budnik V, Tejedor FJ (2002) DLG differentially localizes Shaker K⁺-channels in the central nervous system and retina of *Drosophila*. *J Neurochem* 82: 1490–1501.
- Tully T, Quinn WG (1985) Classical conditioning and retention in normal and mutant *Drosophila* melanogaster. *J Comp Physiol A* 157: 263–277.
- Dura JM, Preat T, Tully T (1993) Identification of *linotte*, a new gene affecting learning and memory in *Drosophila melanogaster*. *J Neurogenet* 9: 1–14.

Acknowledgments

We thank Dr Rolf Bodmer, Leslie Griffith, Scott Waddell and Yi Zhong for flies. Additional stocks were from the Bloomington, Kyoto and Vienna stock centers. We are grateful to Dr Vivian Budnik for rabbit anti-*Drosophila* DLG (PDZ1-2). Confocal microscopy was performed in the Bristol University Wolfson bioimaging facility. We acknowledge Dr Jon Brown, Jules Hancox, Leslie Griffith, Neil Marrion, and Ralf Stanewsky for comments on the manuscript and Dr Stephen Henderson and Emma Robinson for advice on statistics.

Author Contributions

Conceived and designed the experiments: JLLH. Performed the experiments: SC BM JLLH. Analyzed the data: SC BM JLLH. Wrote the paper: SC BM JLLH.

- King I, Tsai LTY, Pflanz R, Voigt A, Lee S, et al. (2011) *Drosophila tao* controls mushroom body development and ethanol-stimulated behaviour through *par-1*. *J Neurosci* 31: 1139–1148.
- Kaun KR, Azanchi R, Maung Z, Hirsh J, Heberlein U (2011) A *Drosophila* model of alcohol reward. *Nat Neurosci* 14: 612–621.
- Keene AC, Stratmann M, Keller A, Perratt PN, Vosshall LB, et al. (2004) Diverse odor-conditioned memories require uniquely timed dorsal paired medial neuron output. *Neuron* 44: 521–533.
- Tamura T, Chiang AS, Ito N, Liu HP, Horiuchi J, et al. (2003) Aging specifically impairs amnesiac-dependent in *Drosophila*. *Neuron* 40: 1003–1011.
- Yamazaki D, Horiuchi J, Miyashita T, Saitoe M (2010) Acute inhibition of PKA activity at old ages ameliorates age-related memory impairment in *Drosophila*. *J Neurosci* 30: 15573–15577.
- Tonoki A, Davis RL (2012) Aging impairs intermediate-term behavioural memory by disrupting the dorsal paired medial neuron memory trace. *Proc Natl Acad Sci USA* 109: 6319–24.
- Spanagel R (2009) Alcoholism: A systems approach from molecular physiology to addictive behaviour. *Physiol Rev* 89: 649–705.
- Cowan TM, Siegel RW (1986) *Drosophila* mutations that alter ionic conduction disrupt acquisition and retention of a conditioned odor avoidance response. *J Neurogenet* 3: 187–201.
- Griffith LC, Wang J, Zhong Y, Wu F, Greenspan RJ (1994) Calcium/calmodulin dependent protein II and potassium channel subunit eag similarly affect plasticity in *Drosophila*. *Proc Natl Acad Sci USA* 91: 10044–10048.
- Gasque G, Labarca P, Reynaud E, Darszon A (2005) Shal and shaker differential contribution to the K⁺ currents in the *Drosophila* neurons. *J Neurosci* 25: 2348–58.
- Gasque G, Labarca P, Delgado R, Darszon A (2006) Bridging behaviour and physiology: ion-channel perspective on mushroom body-dependent olfactory learning and memory in *Drosophila*. *J Cell Physiol* 209: 1046–1053.
- Wright NJ, Zhong Y (1995) Characterization of K⁺ currents and the cAMP dependent modulation in cultured *Drosophila* mushroom body neurons identified by lacZ expression. *J Neurosci* 15: 1025–34.
- Rogero O, Hammerle B, Tejedor FJ (1997) Diverse expression and distribution of Shaker potassium channels during the development of the *Drosophila* nervous system. *J Neurosci* 17: 5108–18.
- Gorczyca MG, Hall JC (1987) Immunohistochemical localization of choline acetyltransferase during development and in *Cha*^{ts} mutants of *Drosophila melanogaster*. *J Neurosci* 7: 1361–9.
- Yu D, Baird GS, Tsien RY, Davis RL (2003) Detection of calcium transients in *Drosophila* mushroom body neurons with camgaroo reporters. *J Neurosci* 23: 64–72.
- Gu H, O'Dowd DK (2006) Cholinergic synaptic transmission in adult *Drosophila* Kenyon cells *in situ*. *J Neurosci* 26: 265–272.
- Martin JR, Rogers KL, Chagneau C, Brulet P (2007) *In vivo* bioluminescence imaging of Ca imaging in the brain of *Drosophila*. *PLoS One* 2: e275.
- Tsydzik V, Wright NJ (2009) Dopamine modulation of the *in vivo* acetylcholine response in the *Drosophila* mushroom body. *Dev Neurobiol* 69: 705–714.
- Lee PT, Lin HW, Change YH, Fu TF, Dubnau J, et al. (2011) Serotonin-mushroom body circuit modulating the formation of anesthesia-resistant memory in *Drosophila*. *Proc Natl Acad Sci USA* 108: 13794–9.
- Hansen HH, Waroux O, Seutin V, Jentsch TJ, Aznar S, et al. (2008) Kv7 channels: interaction with dopaminergic and serotonergic neurotransmission in the CNS. *J Physiol* 586: 1823–32.
- Roepke TA, Smith AW, Ronnekleiv OK, Kelly MJ (2012) Serotonin 5-HT2C receptor-mediated inhibition of the M-current in hypothalamic POMC neurons. *Am J Physiol Endocrinol Metab* 302: E1399–406.
- Miyashita T, Oda Y, Horiuchi J, Yin JCP, Morimoto T, et al. (2012) Mg²⁺ block of *Drosophila* NMDA receptors is required for long-term memory formation and CREB-dependent gene expression. *Neuron* 74: 887–898.

47. Tombaugh GC, Rowe WB, Rose GM (2005) The slow afterhyperpolarization in hippocampal CA1 neurons covaries with spatial learning ability in aged Fisher 344 rats. *J Neurosci* 25: 2609–2616.
48. Tzingounis A, Nicoll RA (2008) Contribution of KCNQ2 and KCNQ3 to the mediaband slow afterhyperpolarization currents. *Proc Natl Acad Sci USA* 105: 19974–19979.
49. Kaczorowski CC, Disterhoft JF (2009) Memory deficits are associated with impaired ability to modulate neuronal excitability in middle-aged mice. *Learn Mem* 16: 362–366.
50. Wang M, Gamo NJ, Yang Y, Jin LE, Xang XJ, et al. (2011) Neuronal basis of age-related working memory decline. *Nature* 476: 210–3.
51. Ramos BP, Birnbaum SG, Lindenmayer I, Newton SS, Dunman RS, et al. (2003) Dysregulation of protein kinase A signalling in the aged prefrontal cortex: new strategy for treating age-related cognitive decline. *Neuron* 40: 835–845.
52. George MS, Abbott LF, Siegelbaum SA (2009) Hyperpolarization-activated HCN channels inhibit subthreshold EPSPs through voltage-dependent interactions with M-type K⁺ channels. *Nat Neurosci* 12: 577–584.