

Husain, S. M., Wilks, M., Mupita, M., Reddy, S. P., Hennessy, E. M., Macfarlane, A. J. & Millar, M. R. (2014). Diversity and stability of cultured vaginal lactobacilli in pregnant women from a multi-ethnic urban UK population. *Journal Of Applied Microbiology*, 117(1), pp. 258-265. doi: 10.1111/jam.12506



**CITY UNIVERSITY
LONDON**

[City Research Online](#)

Original citation: Husain, S. M., Wilks, M., Mupita, M., Reddy, S. P., Hennessy, E. M., Macfarlane, A. J. & Millar, M. R. (2014). Diversity and stability of cultured vaginal lactobacilli in pregnant women from a multi-ethnic urban UK population. *Journal Of Applied Microbiology*, 117(1), pp. 258-265. doi: 10.1111/jam.12506

Permanent City Research Online URL: <http://openaccess.city.ac.uk/4865/>

Copyright & reuse

City University London has developed City Research Online so that its users may access the research outputs of City University London's staff. Copyright © and Moral Rights for this paper are retained by the individual author(s) and/ or other copyright holders. All material in City Research Online is checked for eligibility for copyright before being made available in the live archive. URLs from City Research Online may be freely distributed and linked to from other web pages.

Versions of research

The version in City Research Online may differ from the final published version. Users are advised to check the Permanent City Research Online URL above for the status of the paper.

Enquiries

If you have any enquiries about any aspect of City Research Online, or if you wish to make contact with the author(s) of this paper, please email the team at publications@city.ac.uk.

Diversity and stability of cultured vaginal lactobacilli in pregnant women from a multi-ethnic urban UK population

Journal:	<i>Applied Microbiology</i>
Manuscript ID:	JAM-2013-2385.R2
Journal Name:	Journal of Applied Microbiology
Manuscript Type:	JAM - Original Article
Date Submitted by the Author:	n/a
Complete List of Authors:	Husain, Shahid; Barts and the London School of Medicine & Dentistry, Centre for Paediatrics Wilks, Mark; Barts Health NHS Trust, Department of Microbiology Mupita, Mary; Homerton University Hospital, Department of Midwifery Reddy, Srinivasulu; Barts Health NHS Trust, Department of Microbiology Hennessy, Enid; Barts and the London School of Medicine and Dentistry, Wolfson Institute Macfarlane, Alison; City University London, School of Health Sciences Millar, Michael; Barts and the London NHS Trust, Department of Infection
Key Words:	Lactobacillus, Diagnosis, Disease processes

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

Title

Diversity and stability of cultured vaginal lactobacilli in pregnant women from a multi-ethnic urban UK population

Authors and addresses

Shahid M Husain, Blizard Institute, Barts and The London School of Medicine and Dentistry, 4 Newark Street, London E1 2AT and Neonatal Unit, Homerton University Hospital NHS Foundation Trust, Homerton Row, London E9 6SR, UK

Mark Wilks, Department of Microbiology, Pathology and Pharmacy Block, Barts Health NHS Trust, 80 Newark Street, London E1 2ES and Barts and The London School of Medicine and Dentistry, Blizard Institute, 4 Newark Street, London E1 2AT UK

Mary Mupita, Department of Midwifery, Homerton University Hospital NHS Foundation Trust, Homerton Row, London E9 6SR, UK

Srinivasulu P Reddy, Department of Microbiology, Pathology and Pharmacy Block, Barts Health NHS Trust, 80 Newark Street, London E1 2ES, UK

Enid M Hennessy, Wolfson Institute of Preventive Medicine, Barts and The London School of Medicine and Dentistry, Charterhouse Square, London EC1M 6BQ, UK

Alison J Macfarlane, School of Health Sciences, City University London, Northampton Square, London EC1V 0HB, UK

Michael R Millar, Department of Microbiology, Pathology and Pharmacy Block, Barts Health NHS Trust, 80 Newark Street, London E1 2ES and Barts and The London School of Medicine and Dentistry, Blizard Institute, 4 Newark Street, London E1 2AT UK

Running headline

Vaginal lactobacilli

Corresponding author

Shahid M Husain

Neonatal Unit

Homerton University Hospital NHS Foundation Trust

Homerton Row

London E9 6SR, UK

T: + 44 (0)20 8510 7952

1
2
3 31 F: + 44 (0)20 8510 7787

4 32 E: s.m.husain@qmul.ac.uk

5
6 33 **Abstract**

7
8 34 **Aims**

9
10 35 To determine the diversity and stability of cultured vaginal lactobacilli in a multi-ethnic population of
11 36 pregnant women.

12
13 37 **Methods and Results**

14
15 38 A single centre, prospective, cohort study was performed in a tertiary perinatal centre in East London,
16 39 UK. Self-collected vaginal swabs at 13 and 20 weeks gestation were obtained from women attending
17 40 for routine antenatal care and cultured for lactobacilli. In women who provided both swabs, 37 of 203
18 41 (18%) had no lactobacilli cultured at either time. Only 53 (26%) had the same species at both times.
19 42 Black women were less likely to have lactobacilli cultured at 13 weeks ($p = 0.014$) and Black and
20 43 Asian women were less likely to have lactobacilli cultured at 20 weeks ($p = 0.002$) compared with
21 44 those in the White and Other groups.

22
23 45 **Conclusions**

24 46 Significant differences exist between ethnic groups in the carriage and stability of vaginal lactobacilli.

25
26 47 **Significance and Impact of Study**

27 48 These differences have implications for the design of interventions aimed at normalising the vaginal
28 49 microbiota in pregnant women.

29
30 50 **Keywords**

31 51 Lactobacilli, vaginal microbiota, pregnancy, preterm birth

32
33 52
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

53 Introduction

54 Preterm birth (PTB) makes a major contribution to infant mortality and long-term disability (Moser *et al.*
55 2007; Saigal and Doyle 2008). The mechanisms and causes of spontaneous PTB are poorly
56 understood but known associations include ethnicity, low socio-economic status, a short interval
57 between pregnancies, poor nutritional status, previous history of PTB, intrauterine infection and
58 ethnicity (Goldenberg *et al.* 2008). In the US, the rate of PTB in Black women is 2-3 times that of white
59 mothers (Adams *et al.* 2000; Collins *et al.* 2007; Kistka *et al.* 2007; Goldenberg *et al.* 2008). Similar
60 but more complex patterns have been observed in Europe. The rate of PTB is higher in Black women
61 but differences in PTB are seen between Black Caribbean and Black African groups, and within Black
62 African subgroups. Studies in North Paris, East London, North West England, and England and Wales
63 as a whole have found higher rates of PTB among women from the Caribbean and West Africa
64 compared with women from Northern Africa (Zeitlin *et al.* 2004; Macfarlane *et al.* 2005; Balchin and
65 Steer 2007; Datta-Nemdharry *et al.* 2012). A review of ethnic disparities in PTB pointed out that both
66 social and biological factors are likely to play a part (Kramer and Hogue 2009).

67
68 Bacterial vaginosis (BV) is associated with PTB (Gibbs *et al.* 1992; Taylor *et al.* 1997). It is
69 characterised by both the absence of lactobacilli and by the presence of large numbers of anaerobic
70 species. Lactobacilli, principally the strains that produce higher levels of H₂O₂, appear to protect
71 against vaginal colonisation by pathogenic species, particularly those causing BV (Klebanoff *et al.*
72 1991; Hawes *et al.* 1996). There is some evidence that vaginal colonisation with H₂O₂ producing
73 lactobacilli reduces the risk of chorioamnionitis and PTB (Reid and Bocking 2003; Wilks *et al.* 2004;
74 Mosbah and Mesbah 2009). In the US, BV is commoner in Black women (Antonio *et al.* 2009; Uscher-
75 Pines and Hanlon 2009) and is significantly associated with PTB of a low birthweight baby in this
76 ethnic group (Hittie *et al.* 2007). Despite substantial evidence linking bacterial vaginosis with PTB, the
77 results of trials of antibiotic treatment of BV in pregnancy have not produced clear evidence of benefit
78 (Nygren *et al.* 2008; Brocklehurst 2013).

79
80 Ethnic differences in the vaginal microbiota of sexually-active, non-pregnant women have been
81 described in the US (Ravel *et al.* 2011). Previous cross-sectional (Wilks *et al.* 2004; Kiss *et al.* 2007;
82 Mosbah and Mesbah 2009) and longitudinal (Verstraelen *et al.* 2007; Verstraelen *et al.* 2009) studies

1
2
3 83 have reported on the presence and stability of vaginal lactobacilli in pregnant women who were
4 84 predominantly White. Similar studies on pregnant women from multi-ethnic backgrounds have not
5 85 reported before. The aims of this study were to determine the prevalent types and stability of vaginal
6 86 lactobacilli in pregnant women from a multi-ethnic population in East London, UK using standard
7 87 laboratory techniques.
8
9
10
11
12

13 88 14 89 **Material and Methods**

15 90 This single centre, prospective, cohort study was performed with the approval of the Redbridge &
16 91 Waltham Forest Local Research Ethics Committee which formed part of the UK National Research
17 92 Ethics Service (REC reference number 08/H0701/26). The study population consisted of women
18 93 attending the antenatal clinic at Homerton University Hospital NHS Foundation Trust (HUH), London
19 94 between September 2008 and February 2009. Women referred to the antenatal clinic at HUH received
20 95 an information leaflet about the study with the appointment letter for their first antenatal clinic visit.
21 96 Participation involved permitting access to hospital obstetric and neonatal records, contact with the GP
22 97 if required to enquire about prescribed medications, agreeing to self-collect vaginal swabs on two
23 98 occasions, and permission to retain the specimens. Antibiotic usage during the period of pregnancy
24 99 was determined by asking the participant. Ethnicity was self-defined by the participants and results
25 100 were analysed by grouping ethnicity into the categories used in England, based on categories used in
26 101 the 2001 population census: White (British, Irish and other White), Black (Caribbean, African, other
27 102 Black and mixed Black and White), Asian (Indian, Pakistani, Bangladeshi, other Asian and mixed
28 103 Asian and white) and Other (Chinese, other and not known).
29
30
31
32
33
34
35
36
37
38
39
40
41
42

43 104
44 105 The women in the study provided two self-collected swabs: the first at the time of the first antenatal
45 106 clinic appointment at approximately 13 weeks gestation (swab A) and the second when the women
46 107 attended for a routine ultrasound anomaly scan at approximately 20 weeks gestation (swab B).
47 108 Women were provided with a sheet of written instructions and diagrams that described how to self-
48 109 collect a vaginal swab. Briefly, women were asked to wash their hands, gently part their labia, remove
49 110 the sterile swab from its plastic tube, insert the 'cotton-bud' end of the swab into their vagina to
50 111 approximately half the swab length (about 6 cm), gently twist the swab about three times, part their
51 112 labia, remove the swab and place it back into its plastic tube. The swab was then extracted into 3 mls
52
53
54
55
56
57
58
59
60

1
2
3 113 brain heart infusion broth (BHI) containing 10% glycerol and 0.005% cysteine hydrochloride and
4 114 stored at -70°C. After the women gave birth, maternal and neonatal hospital records were reviewed
5
6 115 and data on maternal demographics, gestational age at birth, birth outcome, and birthweight collected.
7
8 116

10 117 ***Culture and identification of lactobacilli***

11 118 Members of staff performing the microbiological assays were blinded to the clinical characteristics of
12
13 119 the study population. Thawed vaginal secretions were vortexed for 10 secs, inoculated onto MRS agar
14
15 120 (Unipath, Basingstoke, UK) and incubated for 48 h at 35 °C in an atmosphere of 10% CO₂, 10% H₂
16
17 121 and 80% N₂. Single colonies from recovered cultures were subcultured onto blood agar plates (5%
18
19 122 horse blood, Oxoid, Basingstoke UK) and used for DNA extraction as described below and for
20
21 123 determination of H₂O₂ production. H₂O₂ production was measured using a semi-quantitative assay
22
23 124 (Merckoquant Peroxide Test, Merck, Leics, UK) as described previously.¹⁷ Results from this test are
24
25 125 expressed in bands of H₂O₂ production: negative, 1-3, 3-10, 10-30 and 30-100 mg l⁻¹.
26
27 126

28
29 127 Following DNA extraction using a QIAamp DNA minikit (Qiagen, Manchester, UK), lactobacilli were
30
31 128 identified to species level by 16S rDNA sequencing or matrix assisted laser desorption ionisation time
32
33 129 of flight (MALDI-TOF) analysis. For 16S rDNA sequencing, a 1,350-bp fragment of 16S rRNA gene
34
35 130 was amplified using oligonucleotide primers 5'-GAA CGC TGG CGG CGT GCC (Z1-forward) and 5'-
36
37 131 TCC GCG ATT ACT AGC GAT TCC (Z2-reverse). During the course of the study, MALDI-TOF mass
38
39 132 spectrometry was introduced into the laboratory and validated for the identification of lactobacilli using
40
41 133 standard strains. For MALDI-TOF analysis, a single colony of a fresh culture was lysed with 70%
42
43 134 ethanol, extracted with acetonitrile and formic acid, overlaid with hydroxy cinnamic acid matrix and
44
45 135 analysed using a Bruker Microflex mass spectrometer running MALDI-TOF Biotyper 2.0 analysis
46
47 136 software.
48

49 138 ***Statistics***

50
51 139 The data from the swabs and the clinical information were merged and checked for obvious errors.
52
53 140 The analyses were performed using Stata 10. Log_e transformations of H₂O₂ were analysed by the
54
55 141 Kruskal-Wallis test followed by Sidak's adjustment for multiple comparisons. Associations were tested
56
57 142 using chi-squared or Fisher's exact tests for tables. Logistic regression was used to investigate
58
59
60

1
2
3 143 associations with PTB, and any or specific lactobacilli carriage. For comparisons of White v Black, and
4 144 White v Asian, a Bonferroni correction assuming 3 potential comparisons was made. The other group
5 145 was not included because it is heterogeneous and small. This is a conservative correction. No
6
7 146 adjustments were made for White v all others or Black v all others. All p-values are two sided and
8
9 147 confidence intervals are 95%.
10
11
12 148

13 14 149 **Results**

15 150 The base line characteristics of the recruited women are shown in Table 1. Of the 293 women
16
17 151 recruited to the study, gestational age and birth weights of live births were unavailable in 46 women (9
18
19 152 had a miscarriage or termination of pregnancy and 37 moved out of area). A second swab was not
20
21 153 obtained from 90 women mainly because of researcher non-availability when these women attended
22
23 154 for their routine ultrasound anomaly scan.
24
25 155

26
27 156 Overall, 75% of women were colonised with any lactobacillus at either of the sampling times (Table 2).
28
29 157 The mean (SD) number of species of lactobacilli isolated from swab A was 1.15 (0.92) compared with
30
31 158 1.14 (0.87) from swab B (data not shown). The statistically significant effects of ethnic group on
32
33 159 isolation of any lactobacillus in swab A and in Swab B among mothers with both swabs is associated
34
35 160 with a significant reduction in carriage for Black compared to White mothers ($p = 0.006$ for both
36
37 161 comparisons). Indian women had very similar reduced carriage for any lactobacilli in swab B as Black
38
39 162 mothers, but the results are not significant because of smaller numbers ($p = 0.105$, chi-squared after
40
41 163 Bonferroni correction). Compared with the White women in the study, the reductions in lactobacilli
42
43 164 carriage appeared to be because fewer Black and Indian women were colonised with *L. crispatus* ($p =$
44
45 165 0.12 and $p = 0.19$, respectively), fewer Black women were colonised with *L. gasseri* ($p = 0.32$) and
46
47 166 fewer Indian women with *L. jensenii* ($p = 0.12$) but none of these associations were significant
48
49 167 (Fisher's exact test adjusted for 3 comparisons using Bonferroni's test for multiple comparisons).
50

51 169 Delivery of the fetus between 22⁺⁰ and 36⁺⁶ completed weeks of gestation occurred in 9 (5%) of 181
52
53 170 women who were lactobacillus positive at the first swab and 6 (9%) of 66 women who were negative
54
55 171 ($p = 0.23$). Delivery during this range of gestational age was lower in White women (2.4%) compared
56
57 172 to all others (9.9%) ($p = 0.016$, Fisher's exact test). Excluding 4 multiple pregnancies which are
58
59
60

1
2
3 173 themselves associated with PTB, non-White women were at increased risk with 9 PTBs (7.8%) from
4 174 144 births compared to White women with 2 PTBs(1.6%) from 122 births ($p = 0.030$, Fisher's exact
5
6 175 test). The odds ratio for PTB for non-White women after adjustment for lactobacilli carriage at swab A,
7
8 176 is 5.0 (CI 1.05 – 24, $p = 0.045$) while that for presence of any lactobacilli at swab A was not significant
9
10 177 (OR = 0.7, CI .21 - 3.7, $p=0.64$ after adjustment for non-White ethnic group).

11 178
12
13 179 The amount of H_2O_2 produced by *L. jensenii* was significantly higher than other common species
14
15 180 (Table 3). A one-way analysis of variance comparing the $\log_e H_2O_2$ produced showed that *L. jensenii*
16
17 181 was highly significantly different from *L. crispatus*, *gasserii* and *vaginalis*, ($p < 0.001$ after Sidak's
18
19 182 adjustment for multiple comparisons). Similarly, regression analysis of the $\log_e H_2O_2$ produced showed
20
21 183 that *L. jensenii* had nearly six times the level of H_2O_2 production as *L. crispatus*, *gasserii* and *vaginalis*
22
23 184 (5.9, CI 4.3 to 8.1, $p < 0.001$).

24
25 185
26
27 186 Isolates of the same species were assumed to be the same strain of that species and the data were
28
29 187 analysed to obtain basic information on the stability of lactobacillus carriage. The proportions of
30
31 188 women who had specific strains at swab A and swab B were very similar but this masks a high
32
33 189 turnover in species in individual women (Table 4). In women who provided both swab samples, 37
34
35 190 (18%) of 203 did not have lactobacilli isolated at either time, 53 (26%) had the same lactobacillus
36
37 191 species isolated at both times, 71 (35%) gained a new species, and 68 (45%) of 150 who had a
38
39 192 lactobacillus isolated at the first sampling time lost a species. In total, 90 of 203 (44%, CI 37 to 51%)
40
41 193 had the same strains (or none) at both time points. Using multivariate analysis, Black women were
42
43 194 less likely to gain a new species (OR 0.49, CI 0.25 to 0.98, $p = 0.043$) compared with all other ethnic
44
45 195 groups combined. There were significant differences in the proportions of different ethnic groups
46
47 196 losing either any species ($p = 0.008$) or all species ($p = 0.005$), with over 20% of Asian and Black
48
49 197 women losing all species compared with only 4% of White women.

50 198
51 199 Antibiotic usage occurred in the preceding month in 18 of 293 (6%) women who provided a swab A
52
53 200 and 11 of 203 (5%) of those who provided a swab B. The oral antibiotics used were amoxicillin,
54
55 201 cefalexin and co-amoxiclav. Of the 150 women who provided both swabs and had lactobacilli in swab
56
57 202 A, 6 received antibiotics between swabs A and B and none of them lost any strains, while 65 of the

1
2
3 203 other 144 who did not report antibiotic usage did lose a species. This difference is significant ($p =$
4 204 0.029, Fisher's exact test) and suggests that those receiving oral antibiotics were less likely to lose a
5
6 205 species. The binomial exact one-sided confidence interval for the proportions losing a strain if they
7
8 206 had received oral antibiotics is 0 - 46%. This suggests that of similar women given oral antibiotics
9
10 207 fewer than half would be expected to lose a strain of Lactobacilli over the period.
11
12 208

13 14 209 **Discussion**

15
16 210 In this study, we found significant differences in cultured vaginal lactobacilli between ethnic groups at
17
18 211 two time points during pregnancy. Black women were less likely to have vaginal lactobacilli at 13 and
19
20 212 20 weeks of gestation compared with White women. There was a high turnover of vaginal lactobacilli
21
22 213 species in individual women.
23

24
25 214
26 215 To our knowledge, this is the first report to present longitudinal data on vaginal lactobacillus
27
28 216 colonisation during pregnancy in an ethnically diverse population. Vaginal colonisation was
29
30 217 determined using standard laboratory techniques only. We did this because interventions involving the
31
32 218 administration of live lactobacilli (Vangelista *et al.* 2010; Yamamoto *et al.* 2013) require amongst other
33
34 219 properties that the strains are easily culturable to allow manufacture of adequate quantities of the
35
36 220 product and to allow the ready detection of the organism after administration not only to determine the
37
38 221 success of colonisation but also for reasons of safety. Therefore, no attempt was made to identify
39
40 222 strains such as *L. iners* that are often difficult to recover in culture and require molecular methods of
41
42 223 detection.
43

44 224
45 225 Our findings are in agreement with recent reports of ethnic variation in vaginal lactobacillus
46
47 226 colonisation in non-pregnant women (Zhou *et al.* 2007). Three quarters of women in our study were
48
49 227 found to be colonised with vaginal lactobacilli at both times of swabbing and this result is in agreement
50
51 228 with previous cross-sectional reports (Bayó *et al.* 2002; Zhou *et al.* 2007). However, Black women at
52
53 229 the time of both swabs A and B, and Asian women at the time of swab B, were less likely to have
54
55 230 vaginal lactobacillus colonisation. The ethnic differences in vaginal microbiota found in this study and
56
57 231 others may be due to a number of reasons including genetic influences on the immune system and
58
59 232 differences in nutritional factors and cultural practices. The distribution pattern of the most common
60

1
2
3 233 lactobacillus species varies between studies for reasons that are unclear. In earlier studies, the
4 234 unreliability of biochemical identification methods made reliable speciation of lactobacilli unreliable
5
6 235 (Wilks *et al.* 1984), but advances in the identification of lactobacilli by molecular methods such as 16S
7
8 236 rDNA sequencing or MALDI-TOF suggests that reported differences in detected species are not due
9
10 237 to technical factors.

11
12 238

13
14 239 In this study, mean gestational age of live births did not differ between the ethnic groups, although as
15
16 240 expected the birth weight of Asian babies was lower than that of the other groups (Leon and Moser
17
18 241 2012). PTB occurred significantly more frequently in non-White women but not significantly more in
19
20 242 the absence of lactobacilli in swab A. Reports in the literature suggest an association between preterm
21
22 243 labour and reduced frequency of vaginal lactobacillus colonisation or BV (Hitti *et al.* 2007; Donders *et*
23
24 244 *al.* 2009; Mosbah and Mesbah 2009). These findings have prompted trials both with antibiotics and
25
26 245 probiotics designed to modify the vaginal microbiota with the objective of improving pregnancy
27
28 246 outcome. Antibiotics administered to pregnant women can eradicate BV but are unable to reduce the
29
30 247 risk of preterm labour and birth (Lams *et al.* 2008; Brocklehurst *et al.* 2013). Oral or vaginal
31
32 248 administration with probiotic strains of lactobacilli has often been successful in establishing
33
34 249 colonisation of the vagina by the probiotic strain but studies have not been sufficiently powered to
35
36 250 determine an effect on preterm birth (Othman *et al.* 2007). If there are ethnic differences in the vaginal
37
38 251 microbiota, any interventions designed to restore the normal microbiota must take this into account in
39
40 252 addition to viability, dosage and strain/species of lactobacilli.

41
42 253

43
44 254 H₂O₂ production by vaginal lactobacilli is considered to be an important defence mechanism against
45
46 255 vaginal colonisation by undesirable microorganisms. In a previous study we showed that the presence
47
48 256 of H₂O₂ producing lactobacilli in the vagina of women who were at risk of PTB was associated with
49
50 257 reduced risk of adverse birth outcomes (Wilks *et al.* 2004). The explanation for this finding is unclear
51
52 258 because in vitro experiments have shown that the microbicidal activity of H₂O₂ is blocked by
53
54 259 cervicovaginal fluid and semen (O'Hanlon *et al.* 2010). However, these findings may not be applicable
55
56 260 in vivo where, for example, H₂O₂ producing lactobacilli may produce concentrations of H₂O₂ in their
57
58 261 immediate vicinity that are sufficiently high to prevent adherence of a potential pathogen to the vaginal
59
60 262 mucosa and thus prevent colonisation. In addition, it may be that H₂O₂ producing lactobacilli strains

1
2
3 263 produce other microbicidal factors such as lactic acid or bacteriocins that prevent proliferation of
4 264 pathogenic in the vagina.

5
6 265

7
8 266 In this study, approximately 5-6% of women received antibiotics in the month preceding either of the
9
10 267 swab samples. Our figures are similar to that reported in a longitudinal study in the UK which also
11
12 268 used self-reported data and showed that 8% of women reported antibiotic use in early pregnancy and
13
14 269 5% at 32 weeks gestation (Headley *et al.* 2004). By contrast, Petersen and colleagues used
15
16 270 prescribing information recorded in a primary care database in South West London and found that
17
18 271 14% of women received at least one antibiotic in each trimester (Petersen *et al.* 2010). Taken together
19
20 272 the data suggest that either the use of self-reporting underestimates the consumption of antibiotics
21
22 273 during pregnancy or regional differences exist in the prescribing habits of GPs. In a study of non-
23
24 274 pregnant women, use of antibiotics was associated with loss of vaginal lactobacillus strains (Vallor *et*
25
26 275 *al.* 2001). However, we found that vaginal lactobacillus colonisation was relatively unperturbed by
27
28 276 exposure to oral antibiotic administration even though lactobacilli show in vitro sensitivity to some of
29
30 277 the antibiotics ingested by the women in this study (Hamilton-Miller and Shah 1994).

31 278

32 279 While this observational study was not powered to detect independent effects of ethnicity and
33
34 280 lactobacillus colonisation on PTB, the combined results from this and previous studies warrant further
35
36 281 research to investigate their effects on PTB. Two significant advances in recent years have made it
37
38 282 more practical to undertake large studies in which multiple samples could be taken during pregnancy
39
40 283 from different ethnic groups. Firstly, the validity of collecting self-taken swabs, enabling easier patient
41
42 284 recruitment, is now well-established (Strauss *et al.* 2005; Srinivasan *et al.* 2010) and secondly the
43
44 285 ready availability of molecular methods for the in-depth analysis of samples at relatively low cost.
45
46 286 Further research along these lines will allow examination of the effects of ethnic, dietary and other
47
48 287 factors on the vaginal microbiota and provide a more robust framework for interventions.

49 288

50 289 **Acknowledgements**

51
52 290 We thank the women who participated in this study. We also thank the staff of the midwifery and
53
54 291 antenatal ultrasonography departments at Homerton University Hospital for their help in facilitating this
55
56 292 study. Angela Whiley, Simon Warwick and Najeema Begum helped with the culture and identification
57
58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

293 of the lactobacilli. This study was supported by funds allocated to Team Hackney from the UK
294 government's Neighbourhood Renewal Fund.

295

296 **Conflict of Interest**

297 No conflict of interest declared.

298

299 **References**

300 Adams, M.M., Elam-Evans, L.D., Wilson, H.G., and Gilbertz, D.A. (2000) Rates and factors associated
301 with recurrence of preterm delivery. *JAMA* **283**,1591-1596.

302 Antonio, M.A., Meyn, L.A., Murray, P.J., Busse, B. and Hillier, S.L. (2009) Vaginal colonization by
303 probiotic *Lactobacillus crispatus* CTV-05 is decreased by sexual activity and endogenous *Lactobacilli*.
304 *J Infect Dis* **199**,1506-1513.

305 Balchin, I. and Steer, P.J. (2007) Race, prematurity and immaturity. *Early Hum Dev* **83**,749-754.

306 Bayó, M., Berlanga, M. and Agut, M. (2002) Vaginal microbiota in healthy pregnant women and
307 prenatal screening of group B streptococci (GBS). *Int Microbiol* **5**,87–90.

308 Brocklehurst, P., Gordon, A., Heatley, E. and Milan, S.J. (2013) Antibiotics for treating bacterial
309 vaginosis in pregnancy. *Cochrane Database of Syst Rev* **Issue 1**,Art No: CD000262. DOI:
310 10.1002/14651858.CD000262.pub4.

311 Collins, J.W., Jr, David, R.J., Simon, D.M. and Prachand, N.G. (2007) Preterm birth among African
312 American and white women with a lifelong residence in high-income Chicago neighborhoods: an
313 exploratory study. *Ethn Dis* **17**,113-117.

314 Datta-Nemdharry, P., Dattani, N. and Macfarlane, A.J. (2012) Birth outcomes for African and
315 Caribbean babies in England and Wales: retrospective analysis of routinely collected data. *BMJ Open*
316 **2**,e001088. doi:10.1136/bmjopen-2012-001088.

317 Donders, G.G., Van Calsteren, K., Bellen, G., Reybrouck, R., Van den Bosch, T., Riphagen, I. and
318 Van Lierde, S. (2009) Predictive value for preterm birth of abnormal vaginal flora, bacterial vaginosis
319 and aerobic vaginitis during the first trimester of pregnancy *BJOG* **116**,1315–1324.

320 Gibbs, R.S., Romero, R., Hillier, S.L., Eschenbach, D.A. and Sweet, R.L. (1992) A review of
321 premature birth and subclinical infection. *Am J Obstet Gynecol* **66**,1515-1528.

- 1
2
3 322 Goldenberg, R.L., Culhane, J.F., Iams, J.D. and Romero, R. (2008) Epidemiology and causes of
4
5 323 preterm birth. *Lancet* **371**,75-84.
6
7 324 Hamilton-Miller, J.M.T. and Shah, S. (1994) Susceptibility patterns of vaginal lactobacilli to eleven oral
8
9 325 antibiotics. *J Antimicrob Chemother* **33**,1059-1060.
10
11 326 Hawes, S.E., Hillier, S.L., Benedetti, J., Stevens, C.E., Koutsky, L.A., Wølnner-Hanssen, P. and
12
13 327 Holmes, K.K. (1996) Hydrogen peroxide producing lactobacilli and acquisition of vaginal infections. *J*
14
15 328 *Infect Dis* **174**,1058-1063.
16
17 329 Headley, J., Northstone, K., Simmons, H. and Golding, G. (2004) Medication use during pregnancy:
18
19 330 Data from the Avon longitudinal study of parents and children. *Eur J Clin Pharmacol* **60**,355-361.
20
21 331 Hitti, J., Nugent, R., Boutain, D., Gardella, C., Hillier, S.L. and Eschenbach, D.A. (2007) Racial
22
23 332 disparity in risk of preterm birth associated with lower genital tract infection. *Paediatr Perinat Epidemiol*
24
25 333 **21**,330-337.
26
27 334 Kiss, H., Kögler, B., Petricevic, L., Sauerzapf, I., Klayraung, S., Domig, K., Viernstein, N. and Kneifel,
28
29 335 W. (2007) Vaginal Lactobacillus microbiota of healthy women in the late first trimester of pregnancy.
30
31 336 *BJOG* **114**,1402-1407.
32
33 337 Kistka, Z.A-F., Palomar, L., Lee, K.A., Boslaugh, S.E., Wangler, M.F., Cole, F.S., DeBaun, M.R. and
34
35 338 Muglia, L.J. (2007) Racial disparity in the frequency of recurrence of preterm birth. *Am J Obstet*
36
37 339 *Gynecol* **196**,131.e1-131.e6.
38
39 340 Klebanoff, S.J., Hillier, S.L., Eschenbach, D.A. and Waltersdorff, A.M. (1991) Control of the microbial
40
41 341 flora of the vagina by H₂O₂-generating lactobacilli. *J Infect Dis* **164**,94-100.
42
43 342 Kramer, M.R. and Hogue, C.R. (2009) What causes racial disparities in very preterm birth? A biosocial
44
45 343 perspective. *Epidemiol Rev* **31**,84-98.
46
47 344 Lams, J.D.F., Romero, R., Culhane, J.F. and Goldenberg, R.L. (2008) Primary, secondary, and tertiary
48
49 345 interventions to reduce the morbidity and mortality of preterm birth. *Lancet* **371**:164-175.
50
51 346 Leon, D.A. and Moser, K.A. (2012) Low birth weight persists in South Asian babies born in England
52
53 347 and Wales regardless of maternal country of birth. Slow pace of acculturation, physiological constraint
54
55 348 or both? Analysis of routine data. *J Epidemiol Community Health* **66**,544-551.
56
57 349 Macfarlane, A., Grant, H., Hancock, J., Hilder, L., Lyne, M., Costeloe, K. and Hird, M. (2005) Early life
58
59 350 mortality in East London: a feasibility study. Summary report. Fetal and Infant Death in East London.
60
351 London: City University.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

- 352 Mosbah, A. and Mesbah, M.R. (2009) A study of the role of hydrogen peroxide production by
353 lactobacilli in preterm labor. *Int J Med Med Sci* **1**,388-395.
- 354 Moser, K., Macfarlane, A., Chow, Y.H., Hilder, L. and Dattani, N. (2007) Introducing new data on
355 gestation-specific infant mortality among babies born in 2005 in England and Wales. *Health Statistics*
356 *Quarterly* **35**,13-27.
- 357 Nygren, P., Rongwei, F., Freeman, M., Bougatsos, C., Klebanoff, M. and Guise, J-M. (2008) Evidence
358 on the benefits and harms of screening and treating pregnant women who are asymptomatic for
359 bacterial vaginosis: An update review for the U.S. Preventive Services Taskforce. *Ann Intern Med*
360 **148**,220-233.
- 361 O'Hanlon, D.E., Lanier, B.R., Moench, T.R. and Cone, R.A. (2010) Cervicovaginal fluid and semen
362 block the microbicidal activity of hydrogen peroxide produced by vaginal lactobacilli. *BMC Infect Dis*
363 **10**,120.
- 364 Othman, M., Neilson, J.P. and Alfirevic, Z. (2007) Probiotics for preventing preterm labour. *Cochrane*
365 *Database of Syst Rev Issue 1*,Art No: CD005941. DOI: 10.1002/14651858.CD005941.pub2.
- 366 Petersen, I., Gilbert, R., Evans, S., Ridolfi, A. and Nazareth, I. Oral antibiotic prescribing during
367 pregnancy in primary care: UK population-based study. *J Antimicrob Chemother* **65**,2238–2246.
- 368 Ravel, J., Gajer, P., Abdo, Z., Schneider, G.M., Koenig, S.S.K., McCulle, S.L., Karlebach, S., Gorle,
369 R., Russell, J., Tacket, T.O., Brotman, R.M., Davis, C.C., Ault, K., Peralta, L. and Forney, L.J. (2011)
370 Vaginal microbiome of reproductive-age women. *Proc Natl Acad Sci USA* **108(suppl 1)**,4680-4687.
- 371 Reid, G. and Bocking, A. (2003) The potential for probiotics to prevent bacterial vaginosis and preterm
372 labour. *Am J Obstet Gynecol* **189**,1202-1208.
- 373 Saigal, S. and Doyle, L.W. (2008) An overview of mortality and sequelae of preterm birth from infancy
374 to adulthood. *Lancet* **371**,261-269.
- 375 Srinivasan, A., Liu, C., Mitchell, C.M., Fielder, T.L., Thomas, K.K., Agnew, K.J., Marrazzo, J.M. and
376 Fredricks, D.N. (2010) Temporal variability of human vaginal bacteria and relationship with bacterial
377 vaginosis. *PLoS ONE* **5**,e10197. DOI: 10.1371/journal.pone.0010197.
- 378 Strauss, R.A., Eucker, B., Savitz, D.A. and Thorp, Jr, J.M. (2005) Diagnosis of bacterial vaginosis from
379 self-obtained vaginal swabs. *Inf Dis Obstet Gynecol* **13**,31–35.
- 380 Taylor, D., Kenyon, S. and Tarnow-Mordi, W. (1997) Infection and preterm labour. *BJOG* **104**,1338-
381 1340.

- 1
2
3 382 Uscher-Pines, L. and Hanlon, A.L. (2009) Racial differences in bacterial vaginosis among pregnant
4
5 383 women: The relationship between demographic and behavioral predictors and individual BV-related
6
7 384 microorganism levels. *Matern Child Health J* **13**,512-519.
- 8
9 385 Vallor, A.C., Antonio, M.A., Hawes, S.E. and Hillier, S. (2001) Factors associated with acquisition of,
10
11 386 or persistent colonization by, vaginal lactobacilli: role of hydrogen peroxide production. *J Infect Dis*
12
13 387 **184**,1431-1436.
- 14
15 388 Vangelista, L., Secchi, M., Liu, X., Bachi, A., Jia, L., Xu, Q. and Lusso, P. (2010) Engineering of
16
17 389 *Lactobacillus jensenii* to secrete RANTES and a CCR5 antagonist analogue as live HIV-1 blockers.
18
19 390 *Antimicrob Agents Chemoter* **54**,2994-3001.
- 20
21 391 Verstraelen, H., Verhelst, R., Claeys, G., De Backer, E., Temmerman, M. and Vanechoutte, M.
22
23 392 (2007) Modified classification of Gram-stained vaginal smears to predict spontaneous preterm birth: a
24
25 393 prospective cohort study. *Am J Obstet Gynecol* **196**,528.e1-6.
- 26
27 394 Verstraelen, H., Verhelst, R., Claeys, G., De Backer, E., Temmerman, M. and Vanechoutte, M.
28
29 395 (2009) Longitudinal analysis of the vaginal microflora in pregnancy suggests that *L. crispatus*
30
31 396 promotes the stability of the normal vaginal microflora and that *L. gasseri* and/or *L. iners* are more
32
33 397 conducive to the occurrence of abnormal vaginal microflora. *BMC Microbiology* **9**,116.
- 34
35 398 Wilks M., Thin R.N. and Tabaqchali, S. (1984) Quantitative bacteriology of the vaginal flora in genital
36
37 399 disease. *J Med Microbiol* **18**, 217-231.
- 38
39 400 Wilks, M., Wiggins, R., Whiley, A., Hennessy, E., Warwick, S., Porter, H., Corfield, A. and Millar, M.
40
41 401 (2004) Identification and H₂O₂ production of vaginal lactobacilli from pregnant women at high risk of
42
43 402 preterm birth and relation with outcome. *J Clin Microbiol* **42**,713-717.
- 44
45 403 Yamamoto, H.S., Xu, Q. and Fichorova, R.N. (2013) Homeostatic properties of *Lactobacillus jensenii*
46
47 404 engineered as a live vaginal anti-HIV microbicide. *BMC Microbiol* **13**,4.
- 48
49 405 Zeitlin, J., Bucourt, M., Rivera, L., Topuz, B. and Papiernik, E. (2004) Preterm birth and maternal
50
51 406 country of birth in a French district with a multiethnic population. *BJOG* **111**,849-855.
- 52
53 407 Zhou, X., Brown, C.J., Abdo, Z., Davis, C.C., Hansmann, M.A., Joyce, P., Foster, J.A. and Forney, L.J.
54
55 408 (2007) Differences in the composition of vaginal microbial communities found in healthy Caucasian
56
57 409 and black women. *ISME J* **1**,121-33.
- 58
59
60

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49

Table 1 Maternal ethnicity and age, gestational age at time of vaginal swabs A and B, and gestational age and birth weight of live births

	White	Black	Asian	Other	All
Number recruited to study (%)	158 (54)	89 (30)	32 (11)	14 (5)	293 (100)
Maternal age (years)	31.4 (5.8)	28.9 (6.1)	28.3 (4.3)	31.7 (7.0)	30.3 (5.9)
Gestational age swab A (w)	12.5 (2.0)	12.8 (2.0)	13.4 (2.3)	14.4 (2.1)	12.9 (2.2)
Gestational age swab B (w)	19.3 (2.6)	19.8 (3.2)	20.5 (0.8)	20.2 (0.4)	19.7 (2.7)
Gestational age of live births (w)	40.0 (1.9)	39.3 (2.6)	38.8 (2.6)	39.8 (2.8)	39.1 (4.0)
Birth weight of live births (kg)	3.47 (0.50)	3.39 (0.53)	3.05 (0.50)	3.17 (0.55)	3.34 (0.57)

Data are shown as mean (SD) unless otherwise indicated

Table 2 Lactobacilli in women who provided swabs A and B

	White	Black	Asian	Other	Total	p-value *
Women with swab A	158	89	32	14	293	
Any lactobacillus in swab A	128 (81)	56 (63)	24 (75)	12 (86)	220 (75)	0.014
Women with swabs A and B	108	64	21	10	203	
Any lactobacillus in swab A	84 (78)	41 (64)	16 (76)	9 (90)	150 (74)	0.165
Any lactobacillus in swab B	89 (82)	39 (61)	13 (62)	10 (100)	151 (74)	0.002
<i>L. jensenii</i>						
Sample A	41 (49)	21 (51)	3 (19)	2 (22)	67 (45)	0.058
Sample B	36 (40)	18 (46)	1 (8)	2 (20)	57 (38)	0.051
<i>L. crispatus</i>						
Sample A	37 (44)	10 (24)	3 (19)	5 (56)	55 (37)	0.042
Sample B	40 (45)	10 (26)	2 (15)	3 (30)	55 (36)	0.062
<i>L. gasseri</i>						
Sample A	32 (38)	8 (20)	8 (50)	3 (33)	51 (34)	0.098
Sample B	34 (38)	9 (23)	7 (54)	3 (30)	53 (35)	0.174
<i>L. vaginalis</i>						
Sample A	14 (17)	9 (22)	3 (19)	0 (0)	26 (17)	0.514
Sample B	19 (21)	8 (21)	1 (8)	1 (10)	29 (19)	0.717
Other lactobacilli						
Sample A	14 (17)	9 (22)	6 (38)	1 (11)	30 (20)	0.307
Sample B	17 (19)	10 (26)	5 (38)	3 (30)	35 (23)	0.559

Data are shown as number (%). * chi-square test for types of lactobacilli; Fisher's exact test for individual 4 (ethnicity) x 2 (yes/no) tables for each row.

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49

Table 3 H₂O₂ production by Lactobacilli isolated from swab A

Lactobacillus species (isolates tested)	H ₂ O ₂ production *	
	Median	Interquartile range
<i>L. jensenii</i> (175)	10 - 30	3 – 10 to 30 – 100
<i>L. crispatus</i> (177)	1 - 3	0 – 1 to 3 – 10
<i>L. gasseri</i> (177)	1 - 3	1 – 3 to 3 – 10
<i>L. vaginalis</i> (68)	1 - 3	1 – 3 to 3 – 10
<i>Other strain</i> (115)	0 - 1	0 – 1 to 1 – 3

* There was significant interspecies variation in H₂O₂ production (p = 0.0001, Kruskal-Wallis test).

Table 4 Gain and loss of lactobacilli between swabs A and B

Ethnicity	White	Black	Asian	Other	Total	
Total number of women	108	64	21	10	203	
Gain of lactobacilli						p-value *
Any lactobacillus species	45 (41.7)	15 (23.4)	7 (33.3)	4 (40.0)	71 (35.0)	0.111
<i>L. jensenii</i>	7 (10.5)	1 (2.3)	0 (0.0)	1 (12.5)	9 (6.6)	0.204
<i>L. crispatus</i>	11 (15.5)	2 (3.7)	0 (0.0)	0 (0.0)	13 (8.8)	0.071
<i>L. gasseri</i>	12 (15.8)	4 (7.1)	2 (15.4)	0 (0.0)	18 (11.8)	0.336
<i>L. vaginalis</i>	12 (12.8)	3 (5.5)	1 (5.6)	1 (10.0)	17 (9.6)	0.473
Any other lactobacillus species	12 (11.1)	5 (7.8)	4 (19.1)	2 (20)	23 (11.3)	0.70
Loss of lactobacilli						
Any lactobacillus species	38 (45.2)	14 (34.2)	13 (81.3)	3 (33.3)	68 (45.3)	0.012
<i>L. jensenii</i>	12 (29.3)	4 (19.1)	2 (66.7)	1 (50.0)	19 (28.4)	0.316
<i>L. crispatus</i>	8 (21.6)	2 (20.0)	1 (33.3)	2 (40.0)	13 (23.6)	0.788
<i>L. gasseri</i>	10 (31.3)	3 (37.5)	3 (37.5)	0 (0.0)	16 (31.4)	0.648
<i>L. vaginalis</i>	7 (50.0)	4 (44.4)	3 (100.0)	0 (0.0)	14 (53.9)	0.226
Any other lactobacillus species	10 (71.4)	4 (44.4)	5 (83.3)	0 (0.0)	19 (63.3)	0.209

Data are shown as number of women (%). * Fisher's exact test for individual 4 (ethnicity) x 2 (yes/no) tables for each row.