

OPINION
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## The Good, the Bad and the Tick

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How tick-borne pathogens (TBPs) could help us understand cancer? The diversity of pathogens transmitted by ticks is higher than that of any other known arthropod vector and includes protozoa (e.g., Babesia spp. and Theileria spp.), bacteria (e.g., intracellular Rickettsia spp. and extracellular Borrelia spp.), viruses (e.g., Tick-borne encephalitis (TBE) and Crimean-Congo hemorrhagic fever (CCHF) virus), helminths (e.g., Cercopithifilaria) and, although less known, fungi (e.g., Dermatophilus) (Otranto et al., 2013; Brites-Neto et al., 2015; de la Fuente et al., 2017). TBPs have complex life cycles that involve vertebrate hosts and the ticks. Intracellular TBP infection triggers cellular and molecular responses that change host cell physiology in fundamental ways. Within vertebrate host cells, the apicomplexan parasites Theileria parva and Theileria annulata activate molecular pathways that result in increased production of reactive oxygen species (ROS), cell immortalization, cancer and host death. In contrast, infection by the rickettsia Anaplasma phagocytophilum inhibits apoptosis, block the production of ROS and results in a self-limiting infection that rarely is lethal for the host. Theileria spp. and A. phagocytophilum modulates host cell response by inducing transcriptional reprogramming of their vertebrate host cells, leukocytes. Transcriptional reprogramming is induced by pathogen-encoded effector proteins that modify host epigenetic pathways that affect not only gene transcription but also protein levels. The complexity of molecular pathways modulated by TBP infection in vertebrate host cells parallel that of cancer which offers a unique opportunity for comparative studies to understand complex health problems such as cancer. Identification of differences between the molecular pathways hijacked by Theileria spp. and A. phagocytophilum with those leading to non-infectious cancer will provide insights into proteins, pathways and biological processes (BP) associated with malignant transformation.

This hypothesis is based in the following rationality: (i) *Theileria* spp. (Cheeseman and Weitzman, 2015), *A. phagocytophilum* (Sinclair et al., 2014) and oncogenic factors (González-Herrero et al., 2018) behave as "epigenators" (Berger et al., 2009; Cheeseman and Weitzman, 2015) because they have the potential to trigger intracellular signaling pathways that lead to changes in chromatin status and gene expression, (ii) transcriptional reprograming and proteome modulation are hallmarks of infection by *Theileria* spp. (Kinnaird et al., 2013) and *A. phagocytophilum* (de la Fuente et al., 2005; Lee et al., 2008), and oncogenesis (González-Herrero et al., 2018), (iii) transcriptional reprograming and proteome modulation in *Theileria* spp. and *A. phagocytophilum* infections and oncogenesis are associated with similar molecular and cellular processes including apoptosis (Borjesson et al., 2005; Brown and Attardi, 2005; Hayashida et al., 2010; Ayllón et al., 2015), metabolic reprograming (Medjkane and Weitzman, 2013; Yu et al., 2018; Cabezas-Cruz et al., 2019; Masui et al., 2019), oxidative stress and ROS production (IJdo and Mueller, 2004; Medjkane et al., 2014; Takaki et al., 2019) among others. To compare the cell response to

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Theileria spp. and A. phagocytophilum infections and carcinogens we propose the combination of quantitative proteomics and network analysis (Figure 1). Networks of proteins and BPs clustered in Emerging Biological Pathways (i.e. network modules resulting from the clustering of proteins and BPs in response to different stimuli) can represent the topology of the specific cell response to Theileria spp. and A. phagocytophilum infection and exposure to carcinogens. The significance of proteins and processes can be then ranked and hierarchized by indexes representing the centrality of proteins and processes in the networks.

# TICK-BORNE PATHOGENS AS MODELS IN CANCER RESEARCH

Infection-induced malignant transformation accounted for 17.8% (1.9 million cases) of the global cancer burden in the year 2002 (Parkin, 2006). The contribution of infectious diseases to cancer epidemiology increased in 2008 to  $\sim$ 2 million new cancer cases attributable to infection with viruses, platyhelminthes, and bacteria (Oh and Weiderpass, 2014). The loss of cellular identity and the transformation of normal into tumor cells is a central and challenging problem in cellular biology. Major advances have been made in understanding the genetic basis and phenotypic changes underlining the continuum from normal cell to tumor cell to malignant transformation (Hanahan and Weinberg, 2000, 2011; Vogelstein et al., 2013). DNA mutations are observed in all types of cancer (Vogelstein et al., 2013). A significant proportion of cancer patients, however, do not have known coding driver mutations and several non-coding mutations affecting not gene function but gene transcription have been identified in cancer (Fredriksson et al., 2014; Zhang et al., 2018; Reyna et al., 2019). In consequence, the attention has been shifted to phenotypic changes induced by aberrant gene expression that also drive tumor and malignant transformation (Guo et al., 2017; Karki et al., 2017; Parfett and Desaulniers, 2017).

In contrast to virus-induced tumorigenesis that include DNA mutations in somatic cells (Ji et al., 2014), Theileria-induced tumorigenesis in bovines does not involve changes in DNA sequence (Cheeseman and Weitzman, 2015; Tretina et al., 2015). Instead, epigenetic mechanisms underlie phenotypic changes associated with Theileria-induced malignant transformation (Cheeseman and Weitzman, 2015). Theileria is consider as a good model to study the molecular basis of phenotypic changes associated with transformation (Cheeseman and Weitzman, 2015; Marsolier et al., 2015). Comparing the T. annulata genome with that of Toxoplasma gondii (as a control of intracellular and non-transforming apicomplexan parasite), Marsolier et al. (2015) identified 33 Theileria-specific proteins among which they found a homolog of mammalian Pin1, a Peptidyl Prolyl Isomerase that regulates cell proliferation, pluripotency, and survival (Marsolier et al., 2015). The human homolog of Pin1 is overexpressed in breast cancer, increases the transcriptional activity of c-Jun and promotes tumor growth (Wulf et al., 2001). It turned out that by interacting and inducing the degradation of FBW7 protein, which degrades c-Jun, Theileria's Pin1 induces c-Jun accumulation and activates the oncogenic c-Jun pathway which in turn promote transformation (Marsolier et al., 2015; Fernandes et al., 2018).

Thus, comparing the genomes of transforming (i.e., *Theileria*) and non-transforming (i.e., Toxoplasma) parasites proved a valid strategy for the identification of Pin1 as a protein relevant in cell transformation and tumor growth (Wulf et al., 2001; Marsolier et al., 2015). Oncogenic viruses such as Kaposi's Sarcoma Herpesvirus also activates c-Jun activity in host cells via virus-encoded proteins (An et al., 2004; Hamza et al., 2004; Xie et al., 2005). However, the complexity of the mechanisms leading tumor transformation is revealed by the fact that infections by non-transforming pathogens (e.g., Reovirus and Staphylococcus aureus) also induce and activate c-Jun transcriptional activity (Clarke et al., 2001; Borjesson et al., 2005). Staphylococcus aureus infection induces JUN (the gene encoding for c-Jun) expression in neutrophils (Borjesson et al., 2005), and staphylococcal α-Toxin activates c-Jun by inducing phosphorylation of its serine 73 (Moyano et al., 2018). Reoviruses can also activate c-Jun activity (Clarke et al., 2001) and were even proposed as cancer therapy (Harrington et al., 2010). Another more general example is that in both Theileria-induced and non-infectious neoplastic transformation, apoptosis is inhibited (Fernald and Kurokawa, 2013; Dasgupta et al., 2016). Apoptosis inhibition is therefore considered a hallmark of cancer (Hanahan and Weinberg, 2000, 2011). However, A. phagocytophilum infection also inhibits apoptosis but, as mentioned above, it does not result in malignant cell immortalization. One conclusion can be reached from these simple comparisons; c-Jun activation, or apoptosis inhibition, alone do not suffice to transform normal cells in tumor cells. What other pathway or pathways have to be modified in a cell to become a tumor cell? A comprehensive comparison between the timing and totality of cell molecular pathways modulated by Theileria spp., A. phagocytophilum and carcinogens may provide an integrative view of the molecular pathways leading to malignant transformation.

## SEMANTIC NETWORKS TO FIND THE KEYWORDS

In graph theory, a network is a set of nodes that are connected by edges (also known as links). In networks representing food webs (Dunne et al., 2013) or host-parasite interactions (Lafferty et al., 2006; Estrada-Peña et al., 2015), nodes are the organisms and the links represent interactions between them. The directionality and strength of the interactions can be measured as the "weight of interaction" (e.g., the number of times a parasite has been found on a host). We proposed to build "semantic networks" (Estrada-Peña et al., 2018) to capture the changes in cell response induced by different stimuli, Theileria spp., A. phagocytophilum and carcinogens (Figure 1). In such framework, two type of nodes can be distinguished [i.e., proteins with Gene Ontology (GO) annotation (Villar et al., 2014) and BPs], and the links between them would be the participation of proteins in one or more BPs (Estrada-Peña et al., 2018). In this regard, semantic networks are directed because a 'source' (i.e., the protein) is Cabezas-Cruz et al. Good, Bad and the Tick

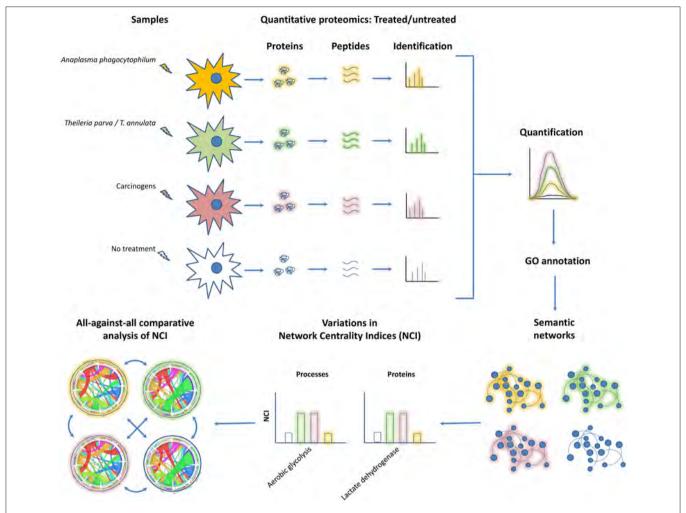


FIGURE 1 Integration of quantitative proteomics with network analysis to elucidate proteins and BPs key to malignant transformation. The figure displays a workflow previously proposed for the characterization of cell response to *A. phagocytophilum* infection (Estrada-Peña et al., 2018). Samples can be prepared using different stimuli, in this case, a non-transforming bacterium (i.e., *A. phagocytophilum*, colored in orange), a transforming apicomplexan parasite (i.e., *T. parva* and/or *T. annulata*, colored in green) and carcinogens (i.e., know chemical compounds capable of causing cancer, colored in brown). Following standard quantitative proteomics and GO annotation, the names of proteins (nodes type 1) are linked to the biological process (BP) (nodes type 2) in which that protein participates using bipartite networks. The links between nodes type 1 and 2 have strength equivalent to protein expression levels (expressed as normalized Peptide-Spectrum Matches) in treated and untreated cells (no colored). In these networks, BPs with many proteins connected to it and proteins with high relative expression will play a key role in keeping the structure of the network. After comparing with untreated cells, three types of phenotypic changes would be uncovered, those related with infectious transformation (in networks resulting from *Theileria* spp. infection), non-infectious transformation (in networks resulting from *A. phagocytophilum* will reveal key players in non-infectious transformation. Likewise, comparing the centrality indices of proteins and BPs in cells infected with *Theileria* spp. and *A. phagocytophilum* will reveal key players in non-infectious transformation. Likewise, comparing the centrality indices of proteins and BPs in cells infected with *Theileria* spp. and *A. phagocytophilum* will provide proteins and BPs critical to infectious transformation. Other comparisons following this logic can provide further insights on infection and cancer.

linked to a "destination" (i.e., the BP). In addition, the links have weight equivalent to the protein levels measured by quantitative proteomics and the Degree of each node is proportional to either the protein level or the sum of links reaching a BP.

Initially, semantic networks were used to describe the global cell transformation in response to *A. phagocytophilum* infection (Estrada-Peña et al., 2018). The results demonstrated that the resulting interactions between proteins and BP can be used to calculate the centrality indices of each node of the network (Estrada-Peña et al., 2018). Centrality indices are fundamental

measures of the structure of a network and account for intimate changes in the relative importance of key functions. In addition, centrality indices can be used to identify both proteins and BP that are "central" and therefore occupy prominent positions in the cellular response to different stimuli. The argument here is that centrality indices (e.g., Degree centrality, Weighted Degree, and Betweenness Centrality) are powerful indicators of subtle changes in the proteome, which could be missed when standard protein representation analysis is used. Comparison of centrality indices between *A. phagocytophilum*-infected and

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non-infected human and tick cells revealed (i) that infection by this pathogen rewires the network of cell processes and changes the relative importance of biological pathways and (ii) that tick and human cells respond differently to A. phagocytophilum infection (Estrada-Peña et al., 2018). More importantly, the ras-related protein Rab14, with a high centrality in infected tick cells, was selected for functional validation by gene knockdown. Rab14 knockdown resulted in a significant decrease in A. phagocytophilum infection levels, suggesting that A. phagocytophilum increases the relative importance of Rab14 in the proteome to facilitate infection (Estrada-Peña et al., 2018). The identification of Rab14 as a key protein in A. phagocytophilum infection shows that in addition to reveal the global cell response to stimuli, semantic networks can be also used to identify individual proteins that change the relative importance of different BPs and can be validated in further laboratory experiments (Estrada-Peña et al., 2018).

Network analysis has been used previously to study cancer progression and reversal (Parikh et al., 2014), to prioritize rare mutations in protein-coding and non-coding genomic regions (Fredriksson et al., 2014; Zhang et al., 2018; Reyna et al., 2019), to study how *PIK3CA* mutations interact with others components of luminal-breast cancer cell signaling network and predict clinical outcomes (McGee et al., 2017) among others. Most network approaches to study cancer use protein-protein interactions where the nodes are proteins and the links between them represent physical protein-protein interactions (Ozturk et al., 2018). In signaling networks, nodes are also proteins but the links represent signaling relations between them (McGee

### **REFERENCES**

- An, J., Sun, Y., and Rettig, M. B. (2004). Transcriptional coactivation of c-Jun by the KSHV-encoded LANA. *Blood* 103, 222–228. doi:10.1182/blood-2003-05-1538
- Ayllón, N., Villar, V., Galindo, R. C., Kocan, K. M., Šíma, R., López, J. A., et al. (2015). Systems biology of tissue-specific response to *Anaplasma phagocytophilum* reveals differentiated apoptosis in the tick vector *Ixodes scapularis*. *PLoS Genet*. 11:e10 05120. doi: 10.1371/journal.pgen.1005120
- Berger, S. L., Kouzarides, T., Shiekhattar, R., and Shilatifard, A. (2009). An operational definition of epigenetics. *Genes. Dev.* 23, 781–783. doi:10.1101/gad.1787609
- Borjesson, D. L., Kobayashi, S. D., Whitney, A. R., Voyich, J. M., Argue, C. M., and Deleo, F. R. (2005). Insights into pathogen immune evasion mechanisms: *Anaplasma phagocytophilum* fails to induce an apoptosis differentiation program in human neutrophils. *J. Immunol.* 174, 6364–6372. doi: 10.4049/jimmunol.174.10.6364
- Brites-Neto, J., Duarte, K. M., and Martins, T. F. (2015). Tick-borne infections in human and animal population worldwide. Vet. World. 8, 301–315. doi:10.14202/vetworld.2015.301-315
- Brown, J. M., and Attardi, L. D. (2005). The role of apoptosis in cancer development and treatment response. *Nat. Rev. Cancer.* 5, 231–237. doi:10.1038/nrc1560
- Cabezas-Cruz, A., Espinosa, P., Alberdi, P., and de la Fuente, J. (2019). Tick-pathogen interactions: the metabolic perspective. *Trends Parasitol*. 35, 316–328. doi: 10.1016/j.pt.2019.01.006
- Cheeseman, K., and Weitzman, J. B. (2015). Host-parasite interactions: an intimate epigenetic relationship. *Cell Microbiol.* 17, 1121–1132. doi: 10.1111/cmi.12471
- Clarke, P., Meintzer, S. M., Widmann, C., Johnson, G. L., and Tyler, K. L. (2001). Reovirus infection activates JNK and the JNK-dependent transcription factor c-Jun. J. Virol. 75, 11275–11283. doi: 10.1128/JVI.75.23.11275-11283.2001

et al., 2017). Other approaches explicitly violate or relax rules of gene and/or protein interactions and allows for biological noise and uncertainty that are expected to occur in tumor cells (Creixell et al., 2015). Our approach is different to those previously reported in two fundamental ways: (i) semantic networks (Estrada-Peña et al., 2018) connect nodes using GO terms which are broader in scope than pathways (Creixell et al., 2015) or protein-protein interactions, and (ii) the links between nodes are weighted based in experimentally-determined protein levels. Thus, semantic networks have the potential to identify not only key BPs, but also those proteins with the higher contribution to that BP in response to the selected stimuli. These two properties, protein-BP connectivity and weighted contribution of proteins result in Emerging Biological Pathways unique to the stimuli in question (e.g., Theileria spp., A. phagocytophilum and carcinogens). The characterization of key proteins and BPs may lead to the identification of fundamental processes involved in carcinogenesis, with possible implication in disease prevention and control.

### **AUTHOR CONTRIBUTIONS**

AC-C, AE-P, and JdF: conceived the idea, drafted the manuscript, reviewed and accepted the manuscript in its current form.

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- Creixell, P., Reimand, J., Haider, S., Wu, G., Shibata, T., Vazquez, M., et al. (2015).
  Pathway and network analysis of cancer genomes. *Nat. Methods* 12, 615–621.
  doi: 10.1038/nmeth.3440
- Dasgupta, A., Nomura, M., Shuck, R., and Yustein, J. (2016). Cancer's Achilles' Heel: Apoptosis and Necroptosis to the Rescue. *Int. J. Mol. Sci.* 18:E23. doi: 10.3390/ijms18010023
- de la Fuente, J., Antunes, S., Bonnet, S., Cabezas-Cruz, A., Domingos, A. G., Estrada-Peña, A., et al. (2017). Tick-pathogen interactions and vector competence: identification of molecular drivers for tick-borne diseases. Front. Cell. Infect. Microbiol. 7:114. doi: 10.3389/fcimb.2017.00114
- de la Fuente, J., Ayoubi, P., Blouin, E. F., Almazán, C., Naranjo, V., and Kocan, K. M. (2005). Gene expression profiling of human promyelocytic cells in response to infection with *Anaplasma phagocytophilum*. *Cell. Microbiol*. 7, 549–559. doi: 10.1111/j.1462-5822.2004.00485.x
- Dunne, J. A., Lafferty, K. D., Dobson, A. P., Hechinger, R. F., Kuris, A. M., Martinez, N. D., et al. (2013). Parasites affect food web structure primarily through increased diversity and complexity. *PLoS Biol.* 11:e1001579. doi: 10.1371/journal.pbio.1001579
- Estrada-Peña, A., de la Fuente, J., Ostfeld, R. S., and Cabezas-Cruz, A. (2015). Interactions between tick and transmitted pathogens evolved to minimise competition through nested and coherent networks. *Sci. Rep.* 5:10361. doi: 10.1038/srep10361
- Estrada-Peña, A., Villar, M., Artigas-Jerónimo, S., López, V., Alberdi, P., Cabezas-Cruz, A., et al. (2018). Use of graph theory to characterize human and arthropod vector cell protein response to infection with *Anaplasma phagocytophilum*. Front. Cell. Infect. Microbiol. 8:265. doi: 10.3389/fcimb.2018. 00265
- Fernald, K., and Kurokawa, M. (2013). Evading apoptosis in cancer. *Trends Cell Biol.* 23, 620–633. doi: 10.1016/j.tcb.2013.07.006
- Fernandes, R., Ferreira, S., and Botelho, M. C. (2018). Commentary: theileria parasites secrete a prolyl isomerase to maintain host leukocyte transformation. Front. Med. 5:120. doi: 10.3389/fmed.2018.00120

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Fredriksson, N. J., Ny, L., Nilsson, J. A., and Larsson, E. (2014). Systematic analysis of non-coding somatic mutations and gene expression alterations across 14 tumor types. *Nat. Genet.* 46, 1258–1263. doi: 10.1038/ng.3141

- González-Herrero, I., Rodríguez-Hernández, G., Luengas-Martínez, A., Isidro-Hernández, M., Jiménez, R., García-Cenador, M. B., et al. (2018). The making of leukemia. *Int. J. Mol. Sci.* 19:E1494. doi: 10.3390/ijms190 51494
- Guo, Y., Nie, Q., MacLean, A. L., Li, Y., Lei, J., and Li, S. (2017). Multiscale modeling of inflammation-induced tumorigenesis reveals competing oncogenic and oncoprotective roles for inflammation. *Cancer Res.* 77, 6429–6441. doi: 10.1158/0008-5472.CAN-17-1662
- Hamza, M. S., Reyes, R. A., Izumiya, Y., Wisdom, R., Kung, H. J., and Luciw, P. A. (2004). ORF36 protein kinase of Kaposi's sarcoma herpesvirus activates the c-Jun N-terminal kinase signaling pathway. J. Biol. Chem. 279, 38325–38330. doi: 10.1074/jbc.M400964200
- Hanahan, D., and Weinberg, R. A. (2000). The hallmarks of cancer. Cell 100, 57–70. doi: 10.1016/S0092-8674(00)81683-9
- Hanahan, D., and Weinberg, R. A. (2011). Hallmarks of cancer: the next generation. *Cell* 144, 646–674 doi: 10.1016/j.cell.2011.02.013
- Harrington, K. J., Vile, R. G., Melcher, A., Chester, J., and Pandha, H. S. (2010). Clinical trials with oncolytic reovirus: moving beyond phase I into combinations with standard therapeutics. *Cytokine Growth Factor Rev.* 21, 91–98. doi: 10.1016/j.cytogfr.2010.02.006
- Hayashida, K., Hattori, M., Nakao, R., Tanaka, Y., Kim, J. Y., Inoue, N., et al. (2010). A schizont-derived protein, TpSCOP, is involved in the activation of NF-kappaB in Theileria parva-infected lymphocytes. *Mol. Biochem. Parasitol*. 174, 8–17. doi: 10.1016/j.molbiopara.2010.06.005
- IJdo, J. W., and Mueller, A. C. (2004). Neutrophil NADPH oxidase is reduced at the *Anaplasma phagocytophilum* phagosome. *Infect Immun.* 72, 5392–5401. doi: 10.1128/IAI.72.9.5392-5401.2004
- Ji, X., Zhang, Q., Du, Y., Liu, W., Li, Z., Hou, X., et al. (2014). Somatic mutations, viral integration and epigenetic modification in the evolution of hepatitis B virus-induced hepatocellular carcinoma. *Curr. Genomic* 15, 469–480. doi: 10.2174/1389202915666141114213833
- Karki, R., Man, S. M., and Kanneganti, T. D. (2017). Inflammasomes and Cancer. Cancer Immunol. Res. 5, 94–99. doi: 10.1158/2326-6066.CIR-16-0269
- Kinnaird, J. H., Weir, W., Durrani, Z., Pillai, S. S., Baird, M., and Shiels, B. R. (2013).
  A bovine lymphosarcoma cell line infected with theileria annulata exhibits an irreversible reconfiguration of host cell gene expression. *PLoS ONE*. 8:e66833. doi: 10.1371/journal.pone.0066833
- Lafferty, K. D., Dobson, A. P., and Kuris, A. M. (2006). Parasites dominate food web links. *Proc. Natl. Acad. Sci. U.S.A.* 103, 11211–11216. doi:10.1073/pnas.0604755103
- Lee, H. C., Kioi, M., Han, J., Puri, R. K., and Goodman, J. L. (2008).
  Anaplasma phagocytophilum-induced gene expression in both human neutrophils and HL-60 cells. Genomics 92, 144–151. doi: 10.1016/j.ygeno.2008. 05.005
- Marsolier, J., Perichon, M., DeBarry, J. D., Villoutreix, B. O., Chluba, J., Lopez, T., et al. (2015). *Theileria* parasites secrete a prolyl isomerase to maintain host leukocyte transformation. *Nature* 520, 378–382. doi: 10.1038/nature 14044
- Masui, K., Onizuka, H., Cavenee, W. K., Mischel, P. S., and Shibata, N. (2019). Metabolic reprogramming in the pathogenesis of glioma: update. Neuropathology 39, 3–13. doi: 10.1111/neup.12535
- McGee, S. R., Tibiche, C., Trifiro, M., and Wang, E. (2017). Network analysis reveals a signaling regulatory loop in the PIK3CA-mutated breast cancer predicting survival outcome. *Genom. Proteom. Bioinforma*. 15, 121–129. doi:10.1016/j.gpb.2017.02.002
- Medjkane, S., Perichon, M., Marsolier, J., Dairou, J., and Weitzman, J. B. (2014). Theileria induces oxidative stress and HIF1α activation that are essential for host leukocyte transformation. Oncogene 33, 1809–1817. doi:10.1038/onc.2013.134
- Medjkane, S., and Weitzman, J. B. (2013). A reversible Warburg effect is induced by Theileria parasites to transform host leukocytes. *Cell Cycle*. 12, 2167–2168. doi: 10.4161/cc.25540
- Moyano, A. J., Racca, A. C., Soria, G., Saka, H. A., Andreoli, V., Smania, A. M., et al. (2018). c-Jun Proto-oncoprotein plays a protective role in lung epithelial cells exposed to staphylococcal α- and Toxin. *Front. Cell. Infect. Microbiol.* 8:170. doi: 10.3389/fcimb.2018.00170

Oh, J. K., and Weiderpass, E. (2014). Infection and cancer: global distribution and burden of diseases. *Ann. Glob. Health* 80, 384–392. doi: 10.1016/j.aogh.2014.09.013

- Otranto, D., Dantas-Torres, F., Brianti, E., Traversa, D., Petrić, D., Genchi, C., et al. (2013). Vector-borne helminths of dogs and humans in Europe. *Parasit Vectors* 6:16. doi: 10.1186/1756-3305-6-16
- Ozturk, K., Dow, M., Carlin, D. E., Bejar, R., and Carter, H. (2018). The emerging potential for network analysis to inform precision cancer medicine. *J. Mol. Biol.* 430, 2875–2899. doi: 10.1016/j.jmb.2018.06.016
- Parfett, C. L., and Desaulniers, D. (2017). A Tox21 approach to altered epigenetic landscapes: assessing epigenetic toxicity pathways leading to altered gene expression and oncogenic transformation in vitro. Int. J. Mol. Sci. 18:E1179. doi: 10.3390/ijms18061179
- Parikh, A. P., Curtis, R. E., Kuhn, I., Becker-Weimann, S., Bissell, M., Xing, E. P., et al. (2014). Network analysis of breast cancer progression and reversal using a tree-evolving network algorithm. *PLoS Comput. Biol.* 10:e1003713. doi: 10.1371/journal.pcbi.1003713
- Parkin, D. M. (2006). The global health burden of infection-associated cancers in the year 2002. Int. J. Cancer 118, 3030–3044. doi: 10.1002/ijc.21731
- Reyna, M. A., Haan, D., Paczkowska, M., Verbeke, L. P. C., Vazquez, M., Kahraman, A., et al. (2019). PCAWG drivers and functional annotation group, ICGC/TCGA pan-cancer analysis of whole genomes. pathway and network analysis of more than 2,500 whole cancer genomes. bioRxiv 2019:385294. doi: 10.1101/385294
- Sinclair, S. H., Rennoll-Bankert, K. E., and Dumler, J. S. (2014). Effector bottleneck: microbial reprogramming of parasitized host cell transcription by epigenetic remodeling of chromatin structure. Front. Genet. 5:274. doi: 10.3389/fgene.2014.00274
- Takaki, A., Kawano, S., Uchida, D., Takahara, M., Hiraoka, S., and Okada, H. (2019). Paradoxical roles of oxidative stress response in the digestive system before and after carcinogenesis. *Cancers* 11:E213. doi: 10.3390/cancers11020213
- Tretina, K., Gotia, H. T., Mann, D. J., and Silva, J. C. (2015). Theileria-transformed bovine leukocytes have cancer hallmarks. Trends Parasitol. 31, 306–314. doi: 10.1016/j.pt.2015.04.001
- Villar, M., Popara, M., Ayllón, N., Fernández de Mera, I. G., Mateos-Hernández, L., Galindo, R. C., et al. (2014). A systems biology approach to the characterization of stress response in *Dermacentor reticulatus* tick unfed larvae. *PLoS ONE* 9:e89564. doi: 10.1371/journal.pone.0089564
- Vogelstein, B., Papadopoulos, N., Velculescu, V. E., Zhou, S., Diaz, L. A., and Kinzler, K. W. (2013). Cancer genome landscapes. *Science* 339, 1546–1558 doi: 10.1126/science.1235122
- Wulf, G. M., Ryo, A., Wulf, G. G., Lee, S. W., Niu, T., Petkova, V., et al. (2001).
  Pin1 is overexpressed in breast cancer and cooperates with Ras signaling in increasing the transcriptional activity of c-Jun towards cyclin D1. EMBO J. 20, 3459–3472. doi: 10.1093/emboj/20.13.3459
- Xie, J., Pan, H., Yoo, S., and Gao, S. J. (2005). Kaposi's sarcoma-associated herpesvirus induction of AP-1 and interleukin 6 during primary infection mediated by multiple mitogen-activated protein kinase pathways. J. Virol. 79, 15027–15037. doi: 10.1128/JVI.79.24.15027-15037.2005
- Yu, X., Ma, R., Wu, Y., Zhai, Y., and Li, S. (2018). Reciprocal regulation of metabolic reprogramming and epigenetic modifications in cancer. *Front. Genet.* 9:394. doi: 10.3389/fgene.2018.00394
- Zhang, W., Bojorquez-Gomez, A., Velez, D. O., Xu, G., Sanchez, K. S., Shen, J. P., et al. (2018). A global transcriptional network connecting non-coding mutations to changes in tumor gene expression. *Nat. Genet.* 50, 613–620. doi: 10.1038/s41588-018-0091-2
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