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SNP rs403212791 in exon 2 of the *MTNR1A* gene is associated with reproductive seasonality in the Rasa aragonesa sheep breed

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3	reproductive seasonality in the Rasa aragonesa sheep breed.
5	Running title: MTNR1A in Reproductive Seasonality
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27 Abstract

28 The aim of this study was to characterize and identify causative SNPs in the 29 MTNR1A gene responsible for the reproductive seasonality traits in the Rasa 30 aragonesa sheep breed. A total of 290 ewes (155, 84 and 51 mature, young 31 and ewe lambs, respectively) from one flock were controlled from January to August. The following three reproductive seasonality traits were considered: the 32 33 total days of anoestrus (TDA) and the progesterone cycling months (P4CM); 34 both ovarian function seasonality traits based on blood progesterone levels; and 35 the oestrus cycling months (OCM) based on oestrous detection, which indicate 36 behavioural signs of oestrous. We have sequenced the total coding region plus 37 733 and 251 bp from the promoter and 3'-UTR regions, respectively, from the 38 gene in 268 ewes. We found 9 and 4 SNPs associated with seasonality traits in 39 the promoter (for TDA and P4CM) and exon 2 (for the three traits), respectively. 40 The SNPs located in the gene promoter modify the putative binding sites for various trans-acting factors. In exon 2, two synonymous SNPs affect RFLP 41 42 sites, rs406779174/Rsal (for the three traits) and rs430181568/MnII (for OCM), 43 and they have been related with seasonal reproductive activity in previous association studies with other breeds. SNP rs400830807, which is located in 44 45 the 3'-UTR, was associated with the three traits, but this did not modify the putative target sites for ovine miRNAs according to *in silico* predictions. Finally, 46 47 the SNP rs403212791 (NW_014639035.1: g.15099004G>A), which is also 48 associated with the three seasonality phenotypes, was the most significant SNP 49 detected in this study and was a non-synonymous polymorphism, leading a

change from an Arginine to a Cysteine (R336C). Haplotype analyses confirmed
the association results and showed that the effects found for the seasonality
traits were caused by the SNPs located in exon 2. We have demonstrated that
the T allele in the SNP rs403212791 in the *MNTR1A* gene is associated with a
lower TDA and higher P4CM and OCM values in the Rasa Aragonesa breed. **Keywords:** sheep, oestrous behaviour, ovulatory activity, *MTNR1A*, SNP

57

58 **1. Introduction**

59 Rasa Aragonesa is an autochthonous Mediterranean sheep breed from northeastern Spain that is mainly reared in extensive or semi-extensive farming 60 61 systems and oriented for meat production. Improvements in efficiency on farms 62 are made possible by exploiting changes in genetics, nutrition and the 63 management of an ewe flock. In this sense, the Cooperative Oviaragon-Grupo 64 Pastores carries out a selection programme for prolificacy in Rasa Aragonesa sheep that began in 1994, currently includes 490,337 ewes because the 65 number of lambs born per ewe plays a key role in the efficiency and viability of 66 these farms [1]. The annual lambing rate can also be increased by getting a 67 68 higher proportion of ewes to breed out-of-season. Sheep breeds from the Mediterranean area have a marked seasonality for breeding activities, showing 69 70 seasonal oestrous behaviours and ovulation patterns. The maximal 71 reproductive activity is associated with short days, with the highest percentage 72 of ewes exhibiting ovulatory activities from August to March. This reproductive 73 seasonality causes a seasonal fluctuation in lamb market prices, with the lowest

74 prices being when the lamb supply is the highest (late spring to early fall). 75 Several methods to control the reproduction of sheep have been used such as environmental manipulation (light control), the sudden introduction of rams, 76 77 which induces oestrus in ewes (ram effect), or other methods based on the 78 administration of exogenous hormones. However, the increasing demand for hormone-free products and the evolution of European rules and directives 79 80 towards a reduction in or even complete cessation of the use of exogenous 81 hormones has led to the search for alternative methods, such as light control, the ram effect or the use of genetic markers. Spring ovulatory activities have 82 83 heritability and repeatability values of 0.20 and 0.30, respectively [2], but they 84 are only measured in females, are exhibited relatively late in an ewe's life and 85 are only present in some management systems. Thus, the use of genetic 86 markers would be a powerful tool in selection programmes. Only two candidate 87 genes related to reproductive seasonality traits have been successfully 88 identified, including melatonin receptor subtype 1A (MTNR1A) [3-13], and the 89 arylalkylamine N-acetyltransferase (AANAT)[14]. Arylalkylamine Nacetyltransferase is involved in the biosynthesis of melatonin and controls daily 90 91 changes in melatonin production. Melatonin acts through high-affinity G-protein 92 coupled receptors, one of which is melatonin receptor 1A encoded by the 93 MTNR1A gene. MTNR1A has been characterized in several breeds as a 94 candidate gene and appears to play a key role in the control of photoperiod-95 induced seasonality mediated by circadian melatonin concentrations [15-16]. 96 However, MTNR1A genotypes did not show an association with reproductive 97 seasonality in Ile de France and Cornell flocks (composed of East Friesian 98 Cross, Dorset, Finnsheep × Dorset, and Finnsheep ewes) [17-18], suggesting

that the effects of MTNR1A polymorphisms may depend on the genetic 99 100 background and/or environmental conditions. Furthermore, previous studies 101 have associated two polymorphic RFLP sites (606/Rsal and 612/MnII) within the 102 MTNR1A coding sequence with reproductive activity. However, since these 103 SNPs are synonymous, they are not causative mutations, indicating that other 104 polymorphisms in linkage disequilibrium or regulatory sequences in the 105 *MNTR1A* gene could be influencing the ability to breed out of season. Genetic 106 mapping of quantitative trait loci (QTL) and a genome-wide association study 107 (GWAS) for aseasonal reproduction in sheep using microsatellites and SNPs, 108 respectively, revealed several chromosomes and SNPs that could be implicated 109 in this trait [19-20].

Therefore, the main objective of this study was to identify causative SNPs responsible for reproductive seasonality traits in the Rasa aragonesa sheep breed. Polymorphisms were detected and characterized from the entire *MTNR1A* gene coding region and promoter. Then, an association study was performed between all the polymorphisms detected and the three reproductive seasonality traits.

116

117 2. Material and Methods

118 2.1 Ethics statement

All experimental procedures were performed in accordance with the guidelines
of the European Union (2003/65/CE) and Spanish regulations (RD 1201/2005,
BOE 252/34367-91) for the use and care of animals in research. No hormonal
treatments were applied to ewes during the study.

123 2.2 Animal resources

124 Phenotypic seasonality data were obtained from a Rasa aragonesa sheep flock 125 managed at an experimental farm ("Pardina de Ayés") owned by Oviaragón 126 S.C.L., located in the Pre-Pyrenees (Ayés, Sabiñánigo, Huesca; North-Eastern 127 Spain, 42° 29' 48.55" N 0° 23' 37.54", 790 m above sea level) and described in 128 Martinez-Royo et al. [20]. The experimental period lasted from January to 129 August in 2012. The flock was composed of 290 ewes in the following three age 130 groups: mature (5.2-7.2 y, n=155; 5.5 \pm 0.5; mean \pm SD), young (all: 1.9 y, 131 n=84) and, ewe lambs (all: 0.94 y; n=51) at the beginning of the experiment. 132 Their individual live weight (LW) and body condition score (BCS) on a 1 to 5 133 scale [21] were assessed every three weeks. Mean LW and BCS values were 134 similar in mature and young ewes age groups. Pooled overall means and 135 standard deviations for the entire experimental period were 52.5 \pm 7.7 kg and 136 2.9 ± 0.3 for LW and BCS, respectively, for mature and young ewes. However, 137 ewe lambs had an LW and BCS of 40.6 ± 3.8 kg and 2.8 ± 0.1 , respectively. 138 Management of the ewes is described in Martinez-Royo et al. [20]. The ewes were handled in a single lot and subjected to the same management, nutrition 139 140 and environmental conditions.

141 2.3 Measurement of reproductive seasonality traits

Three reproductive seasonality traits were considered and described in Martinez-Royo et al. [20]. Briefly, the first was the total days of anoestrus (TDA) based on weekly individual plasma progesterone levels. TDA was the sum of the days in anoestrus, considering anoestrus as periods with three or more consecutive progesterone concentrations lower than 0.5 ng/ml. Likewise, ewes were not considered for this study if they were not cycling in the preceding

breeding season (based on three samples one week apart taken in October),
had progesterone levels below the threshold in all samples taken in January,
and had more than 4 consecutive samples higher than or equal to the threshold
(possible pathological ewes).

The second reproductive seasonality trait was the progesterone cycling months (P4CM), which was defined for each ewe as the rate of cycling months based on progesterone determination. When progesterone levels were higher than or equal to 0.5 ng/ml in at least one blood sample in the month, an ewe was considered cyclic.

Finally, the third reproductive seasonality trait considered was the oestrus cycling months (OCM), which is defined as the rate of months cycling based on daily oestrous records for each ewe. Eight vasectomised rams fitted with harnesses and marking crayons were mixed with the ewes, and daily oestrous detection was examined [22]. Thus, after natural mating, oestrus was recorded as a coloured mark on the ewes' rump.

163 2.3 Sampling and genotyping

164 Genomic DNA was extracted from blood samples from 268 ewes (138, 79 and 165 51 mature, young and ewe lambs, respectively) from the total ewes in the flock 166 (n=290) using the FlavorPrep Genomic DNA mini kit (Flavorgen, Ibian, 167 Zaragoza, Spain). Twenty-two ewes were not considered because of missing 168 data for some variables. Direct Sanger sequencing of the PCR products from all 169 the ewes (n=268) was used to search for genotype polymorphisms in the 170 experimental population. Primers were used to amplify the total coding, 5'-UTR, 171 partial 3'-UTR, and promoter genomic regions in the MNTR1A gene using three 172 different PCR reactions (Supplementary Table 1). Genomic DNA (50 ng) was

173 amplified in a final PCR volume of 25 µl containing 5 pmol of each primer, 200 nM dNTPs, 2.0 mM MgCl2, 50 mM KCl, 10 mM Tris-HCl, 0.1% Triton X-100 174 175 and 1 U Tag polymerase (Biotools, Madrid, Spain). The following cycling 176 conditions were used: an initial denaturation step at 94 °C for 3 min; 35 cycles of 94 °C for 1 min, the annealing temperature for 1 min, and 72 °C for 1 min and 177 30 s for fragments 1-3 and 2, respectively; and a final extension step at 72 °C 178 179 for 10 min. The annealing temperatures for each fragment are indicated in 180 Supplementary Table 1. All PCR amplifications of genomic DNA were 181 performed in a MyCycler thermal cycler (BioRad, Madrid, Spain). The PCR 182 products were purified using the FlavorPrep Gel/PCR purification mini kit Ibian, Zaragoza, Spain), according to the manufacturer's 183 (Flavorgen, 184 instructions. The PCR products were sequenced in both directions by STAB 185 Vida Lda. (Caparica, Portugal) using an ABI 3730XL sequencer (Applied 186 Biosystems, CA, USA).

187 The homology searches were performed using BLAST (National Centre for 188 Biotechnology Information: https://blast.ncbi.nlm.nih.gov/Blast.cgi). To align the 189 sequences, the CLUSTAL Omega (http://www.ebi.ac.uk/Tools/msa/clustalo/) 190 used. The Variant Effect Predictor software was software (VEP: 191 http://www.ensembl.org/Ovis aries/Tools/VEP?db=core), which predicts the 192 possible impact of an amino acid substitution on the structure and function of a 193 protein, was used. The degree of the amino acid change was also assessed 194 the using BLOSUM 62 scoring matrix (http://www.ncbi.nlm.nih. 195 Gov/Class/FieldGuide/BLOSUM62.txt), where low values indicate more drastic 196 changes. Antisense matches for individual miRNAs in the 3'-UTR sequences of 197 the isolated ovine MTNR1A gene were determined using MirTarget

198 (http://mirdb.org/) (http://www.targetscan.org/vert_71/) and TargetScan 199 programs. The locations of the SNPs and gene clusters were identified based 200 the latest sheep genome version for Ovis aries (Oar v4.0, on 201 https://www.ncbi.nlm.nih.gov/genome/?term=ovis+aries).

- 202
- 203 2.4 Linkage disequilibrium (LD)

The gametic LD among SNPs (D' and r^2) within the *MTNR1A* gene was calculated and visualized using the Haploview v4.2 program [23]. LD blocks were defined based on the four-gamete rule algorithm [24].

207 2.5 Statistical analysis

208 2.5.1. SNP association studies

The Hardy–Weinberg equilibrium exact test values and observed and expected heterozygosities and minor allele frequency (MAF) for each SNP were calculated using the PLINK 1.9 software [25].

212 The associations between the *MTNR1A* gene polymorphisms and reproductive seasonality traits (TDA, P4CM, and OCM) were determined by fitting a Linear 213 214 Mixed Model using the Mixed procedure (MIXED) in the SAS statistical package. The model included the genotype of the SNPs (S), the age (mature, 215 216 young and ewe lambs) (A), and the interaction between the age x genotype of 217 the SNPs $(A \times S)$ as fixed effects; the live weight (LW) and body condition score 218 (BCS) as covariates; and the animal (A) and residual I as random effects. 219 Homogeneous variance was considered for the residual (e $\sim N$ (0, $\sigma 2$)). The 220 equation for the model was as follows:

221
$$Y = \mu + S + A + (A \times S) + LW + BCS + A + e$$

To test the differences between genotypes for each breed, the least square means (LSMs) for each pair-wise comparison were estimated. A Bonferroni correction was fitted to take into account the multiple tests. All SNPs were independently analysed with the same statistical model.

226

227 2.5.2. Haplotype association studies

228 SNPs were phased with PLINK1.9 [23] considering the blocks defined by 229 Haploview and using the expectation–maximization (E–M) algorithm to assign 230 individual haplotypes. We considered diplotypes with a posterior probability 231 higher than 0.8.

Associations between the haplotypes and reproductive seasonality traits were performed using a Linear Mixed Model using the Mixed procedure (MIXED) in SAS. The fitted model was similar to that used for the SNP association studies but included the haplotype (H) effect and the interaction between the age × haplotype (A × H). Haplotypes for each individual were codified as 0, 1 or 2, indicating the number of copies of each haplotype. The equation for the model was as follows:

239

$$Y = \mu + H + A + (A \times H) + LW + BCS + A + e$$

Only haplotypes with a frequency greater than or equal to 1% were considered.
To test the differences between the haplotypes, the LSMs for each pair-wise
comparison were estimated. The Bonferroni correction was applied to take into
account multiple tests.

244

245 **3. Results and discussion.**

246 3.1. Structural characterization and linkage disequilibrium

247

248 All of exon 1, 1120 bp from exon 2 (869 bp from the protein coding region plus 249 251 bp from the 3'-UTR) and 733 bp from the promoter region were sequenced. 250 Exons were identified by comparison with ovine sequences (GenBank 251 sequence NM 001009725 and AF078545). Sequences from the total population 252 (n=268) revealed 35 polymorphisms, including 13 SNPs in the promoter region, 253 1 SNP in exon 1 and 21 SNPs in exon 2 (17 and 4 SNPs in the coding and 3'-254 UTR regions, respectively) (Table 1). Table 1 shows the location, alias 255 (nomenclature in this manuscript), dbSNP identifiers, amino acid substitution 256 effect, and genotypic and allelic frequencies of the SNPs. MTNR1A is in reverse 257 orientation on the genome, and SNPs are ordered according to their position in 258 the latest genome version (Oar4.0: GenBank acc. number NW_014639035.1). 259 All SNPs were in Hardy-Weinberg equilibrium, though exon 1 (snp_22) showed 260 a reduced number of heterozygous animals. In the promoter region, we 261 sequenced the same fragment as in Martinez-Royo et al. [12], which is located 262 536 bp upstream from the transcription start site (TSS). We did not find any new 263 SNPs in this region. As described in Martinez-Royo et al. [12], six of these 264 polymorphisms modify putative binding sites for various trans-acting factors, 265 including snp_26 (Brn-2 and Oct-1 consensus sites), snp_27 (SRY), snp_31 266 (SRY), snp_32 (Nkx-2), snp_33 (SRY), and snp_34 (EF2). In exon 1, a 267 synonymous polymorphism was detected; in contrast, 9 non-synonymous SNPs 268 were found in exon 2. Four of the SNPs had not been previously described in 269 other sheep breeds, with two of these being non-synonymous polymorphisms, 270 one that changes from a Lysine to Threonine (K335T) (snp 9) and the other a 271 non-conservative change from a Glycine to Valine (G123V) (snp_20), and both

substitutions were predicted as deleterious (with a SIFT value of 0.04) by the VEP software. However, we did not find homozygous animals for the predicted deleterious allele (G and A for snp_9 and snp_20, respectively), and only one heterozygous animal for each SNP was found in two different animals. None of the other non-synonymous substitutions were predicted as deleterious; all were considered tolerated with SIFT values ranging from 0.37 (snp_10) to 0.05 (snp_12).

279 Fig. 1 shows the linkage disequilibrium plot among the SNPs in MTNR1A. Four 280 LD blocks were predicted (Fig. 1). One block included all the promoter SNPs except snp_23, snp_24, snp_25 and snp_27 (Block 4). These two last SNPs 281 282 (snp_23 and snp_24) constituted Block 3. Finally, two different blocks were 283 predicted in exon 2. Polymorphisms in the promoter and exon 2 showed low linkage disequilibrium. The maximum value of D' and r² between the promoter 284 and exon 2 SNPs were 0.82 and 0.14 (Figure 1), respectively, being separated 285 286 by approximately 24 Kb.

287

288 3.2. SNP association studies

289 The median values and 5th–95th percentiles for the TDA, P4CM and OCM traits 290 for the population are shown in Supplementary Table 2. Thirty-seven of the 291 ewes (29, 6 and 2 mature, young and ewe lambs, respectively) did not present 292 anoestrus during the experiment (TDA=0). Similarly, 87 (60, 17 and 10 mature, 293 young and ewe lambs, respectively) and 9 (7 and 2 mature and young ewes, 294 respectively) ewes were cycling throughout the experiment when considering 295 the P4CM and OCM traits, respectively. TDA and P4CM had a negative 296 correlation because P4CM was a less strict trait for ovarian function than TDA in

the three age groups studied. The phenotypic correlations among the threetraits are shown in Supplementary Table 3.

299

300 3.2.1. Promoter region

301 Results from the association studies are shown in Supplementary Table 4. 302 SNPs snp 9 and snp 20 were not considered in the association analysis 303 because only one heterozygous animal was found (MAF<0.01). The interaction 304 between the SNP and age affected the TDA and P4CM traits in the promoter 305 region (Table 2 and Supplementary Table 4). The two blocks associated with these SNPs were in complete linkage disequilibrium (D'=1 and $r^2=1$) ([snp 29, 306 snp_30 and snp_34] and [snp_24, snp_26, snp_32 and snp_33]) (Figure 1). 307 308 After Bonferroni correction, only the TDA phenotype differed among the SNPs 309 (P<0.05) at ewe lamb age (SNP x age fixed effect), showing significant 310 differences between the homozygous alternative genotypes. However, this 311 association may be spurious as it relies upon an unbalanced distribution of 312 genotypes and a small number of ewe lambs (n = 51) for each SNP. In this 313 sense, only 3 homozygous animals for the MAF allele were found for snp_24, 314 snp 26, snp 29, snp 30, snp 32, snp 33 and snp 34 among the ewe lambs. 315 Considering all ages (SNP fixed effect), the SNPs were significantly associated 316 with the TDA and P4MC traits, showing a balanced distribution of genotypes 317 (Table 2 and Supplementary Table 4), with a homozygous animal frequency for 318 the MAF allele of approximately 17% (Table 1). The present results are in 319 agreement with those described by Martinez-Royo et al. [12] in that SNPs in the 320 MNTR1A gene promoter are associated with reproductive seasonality. 321 However, our results are slightly different because only the OCM trait was

322 examined in that study. Martinez-Royo et al. [12] found 5 associated SNPs, two 323 of which were also associated with TDA and P4CM in the present study, 324 snp 28 (called 677 in Martinez-Royo et al. [12]) and snp 34 (called 422 in 325 Martinez-Royo et al. [12]). The C (snp_26), A (snp_32), A (snp_33), and A 326 (snp_34) alleles were associated with lower TDA and higher P4CM values and 327 created putative binding sites for various trans-acting factors, such as EF-2, 328 SRY, Nkx-2 and Brn-2, respectively. These SNPs showed strong linkage 329 disequilibrium (Fig. 1). These allelic variants may play a role in gene regulation 330 by increasing the MTNR1A gene expression level (mRNA) and thus the final 331 protein contents. Therefore, these variations may enhance the ability of 332 MNTR1A to mediate the physiological function of melatonin, decreasing TDA 333 and increasing P4CM.

334

335 3.2.2. Coding region

336 The SNP found in exon 1 was synonymous and not associated with 337 reproductive seasonality traits. Furthermore, this SNP was not in Hardy-338 Weinberg equilibrium. For any SNP, the interaction between the SNP and age 339 was not significantly associated with reproductive seasonality traits, though four 340 SNP effect associations were found (Table 3 and Supplementary Table 4). Two 341 of them were at the snp_17 (612/MnII) and snp_18 (606/RsaI) polymorphic 342 RFLP sites that have been previously associated with seasonality traits in Rasa 343 aragonesa and other breeds [3-13]. These results are in agreement with those 344 from Martinez-Royo et al. [12], in which snp 18 was associated with the OCM 345 trait in the Rasa aragonesa sheep breed. In that study, snp 17 was not 346 associated, but here, we found a significant association with the OCM trait.

347 However, these SNPs are synonymous polymorphisms and therefore not 348 causative mutations. One of the other two significant SNPs is a non-349 synonymous polymorphism (snp_8), which led to a change of an Arginine to a 350 Cysteine (R336C); the other one is located in the 3'-UTR region (snp_2). Snp_8 351 was the most significant SNP and was predicted as tolerated (with a SIFT value 352 of 0.22) by the VEP software. The GG genotype had greater value than the AA 353 genotype, whereas the AG genotype presented intermediate values for the TDA 354 (73.08±7.25, 53.57±7.36 and 20.85±13.13 for the GG, AG and AA genotypes, respectively) and P4CM (0.80±0.03, 0.86±0.03 and 0.94±0.05 for the GG, AG 355 and AA genotypes, respectively) traits. However, only significant differences 356 357 were observed between the GG and AA genotypes for the OCM trait 358 (0.49±0.03, 0.54±0.03 and 0.64±0.06 for the GG, AG and AA genotypes, 359 respectively). This SNP has been previously described in two populations from Iran and Morocco (NEXTGEN project, http://nextgen.epfl.ch/). Similar results 360 361 were also found for snp_2, as the two SNPs showed high linkage disequilibrium (D'=1, r²=0.94). Snp_2 was located in the 3'-UTR region; this SNP could modify 362 363 putative ovine miRNA target sites. However, this SNP did not modify a miRNA 364 target sequence according to the MirTarget and Targetscan software 365 predictions. Snp_8 could be the causative mutation for the observed effects in 366 the reproductive seasonality traits because it was the most significant SNP and 367 produces an amino acid change. According to the BLOSUM 62 scoring matrix, 368 the substitution generated by snp_8 (arginine to cysteine) is rated -3, with the 369 most severe rating (i.e., introduction of a premature stop codon) being -4. 370 Arginine is a positively charged amino acid, whereas cysteine is polar in nature. 371 Moreover, cysteine is important for the generation of cysteine knots and

disulfide linkages between subunits in some proteins and could potentially affectthe binding of melatonin to its receptor.

374

375 3.3. Haplotype association studies

Haplotype association studies were performed considering the four LD blocks predicted with Haploview (Figure 1 and Supplementary Table 5) and a block containing all the significant SNPs (Block 5; Supplementary Table 5). Totals of 5, 22, 3, 6 and 21 haplotypes were found for blocks 1, 2, 3, 4 and 5, respectively. Because we only considered diplotypes with a posterior probability higher than 0.8, we performed the haplotype analysis with 245, 243, 268, 268 and 233 ewes for blocks 1, 2, 3, 4 and 5, respectively.

383 3.3.1. Promoter region.

384 The statistically significant SNP associations were confirmed by MTNR1A-385 specific haplotype analyses. The haplotype and haplotype x age effects affected 386 the TDA and P4CM traits (Table 4 and Supplementary Table 5). Haplotypes 3 387 and 1 for blocks 3 and 4, respectively, showed significant associations with the 388 TDA and P4CM traits. In this sense, homozygous h3/h3 animals (n= 102) had 389 higher and lower values for TDA and P4CM, respectively, than animals with one 390 copy (n= 119) or no h3 copies (n= 47) for block 3. However, only significant 391 differences were found between having 0 and 2 copies of the haplotype. For 392 this haplotype, only snp 24 was associated with TDA and P4CM, as having the 393 G allele showed higher and lower values for these traits (Table 2), respectively, 394 but without modifying any trans-acting factor putative binding sites. Thus, the 395 SNPs in block 3 could not be the responsible for the observed effects. Similar 396 results were found for haplotype 1 in block 4 because the of the high degree of

397 LD between the two blocks (D'=1), as homozygous h1/h1 animals (n= 103) had 398 higher values than animals with one copy (n = 117) or no h1 copies (n = 48) for 399 block 4 (Table 4). For this haplotype, the T, C, A, C, G, G and G alleles for 400 SNPs snp_26, snp_28, snp_29, snp_30, snp_32, snp_33 and snp_34, 401 respectively, were also associated with higher and lower TDA and P4CM 402 values, respectively, in the SNP association studies (Table 2). In silico analysis 403 showed that the T (snp_26), G (snp_32), G (snp_33), and G (snp_34) alleles 404 were associated with higher and lower TDA and P4CM, respectively, and could 405 destroy the putative binding sites for various trans-acting factors (EF-2, SRY, 406 Nkx-2 and Brn-2, respectively), demonstrating that the SNPs may play a role 407 in regulating gene expression. However, the LSMs for the haplotype x age 408 effect were only significant for the TDA trait in the SNP association studies, 409 while for P4CM, a trend was only observed between having 0 or 2 copies for 410 haplotypes at an ewe lamb age after Bonferroni correction. In this sense, only 411 three ewe lambs had 0 copies of the h3 (block 3) and h1 (block 4) haplotypes 412 for the TDA trait, and even then, these associations were not very confident.

413 3.3.2. Coding region.

414 In exon 2, the haplotype affected the TDA, P4CM and OCM traits (Table 5 and 415 Supplementary Table 5), but the interaction between the haplotype and the age 416 was not significant. Homozygous h3/h3 animals (n= 15) had lower values for 417 TDA than animals with one copy (n = 97) or no h3 copies (n = 133) for block 1. 418 For the P4CM and OCM traits, homozygous h3/h3 animals had higher values 419 than animals with one copy or no h3 copies. Significant differences were found 420 between animals carrying 0 copies and 1 or 2 copies of the haplotype for the 421 three traits. A trend was also observed between 1 and 2 copies (P=0.064) for

422 the TDA trait. Regarding block 2, results similar to those found for block 1 were 423 observed due to the high degree of LD between the two blocks (D'=0.97). 424 Significant differences were found between the three diplotypes for haplotype 425 h2, showing that homozygous h2/h2 animals (n= 13) had lower values than 426 animals with no h2 copies (n= 131). An intermediate TDA value was observed 427 for the heterozygous diplotype (n= 99). For the P4CM and OCM traits, 428 homozygous h2/h2 animals had higher values than animals with one copy or no 429 h2 copies, showing the significant differences between having 0 and 2 copies 430 for the haplotype for both traits.

For these haplotypes, the A, A, C and A alleles in snp_2 (block 1), snp_8 (block 431 432 2), snp_17/MnII (block 2) and snp_18/RsaI (block 2), respectively, were 433 associated with lower TDA and higher P4CM and OCM values in the SNP 434 associations studies (Table 3). Only snp_8 produces a putative functional effect (a change of an Arginine to a Cysteine) (see section 3.2.2). The association 435 436 results obtained for snp_2, snp_17 and snp_18 could be due to the high linkage 437 disequilibrium between these SNPs and snp_8, as the causative mutation of the 438 traits studied in this work.

439 3.3.3. Block with significant SNPs.

To determine whether the phenotype effects observed were caused by snp_8 in exon 2 or SNPs in the promoter, we created another haplotype block with all the significant SNPs associated with the seasonality traits (block 5; Supplementary Table 5). We found 3 haplotypes associated with TDA and P4CM and two associated with OCM (Table 6 and Supplementary Table 5). One copy of the h4 (AACAACCTCTAAA) and h13 (AACAGTTCACGGG) haplotypes showed lower TDA values than having no copies of this haplotype (alleles significantly

447 associated with lower TDA and higher P4CM and OCM values for each SNP in 448 each haplotype according to the SNP association results are shown in bold). 449 These haplotypes had the alleles from exon 2 (A, A, and A for snp_2, snp_8, 450 and snp_18/Rsal, respectively), but not those from the promoter region 451 associated with lower TDA values in the SNP association studies (Tables 2 and 452 3). In the same way, the h12 haplotype (GGCGATCTCTAAA), which was 453 associated with higher TDA values, showed that the alleles from the SNPs in 454 exon 2 were associated with an increase in this trait (G, G, and G for snp 2, 455 snp 8, and snp 18/Rsal, respectively). However, SNPs in the promoter region were associated with lower instead of higher TDA values (the A, T, C, T, C, T, 456 A, A, and A alleles for snp_24, snp_25, snp_26, snp_28, snp_29, snp_30, 457 458 snp_32, snp_33 and snp_34, respectively). Similar results were found for P4CM 459 (the same significant haplotypes were found in the TDA phenotype). For the OCM trait, the two haplotypes associated with higher OCM values (h10, 460 461 AACAGCTCACGGG and h13, AACAGTTCACGGG) had the alleles from exon 462 2 (A, A, and A for snp_2, snp_8, and snp_18/Rsal, respectively), but not those 463 from the promoter region associated with higher OCM values in the SNP association studies (Tables 2 and 3). These results indicated that the effect is 464 465 due to SNPs in exon 2 and not from those in the promoter region. As discussed 466 in sections 3.2.2 and 3.3.2, snp_8 located in exon 2 might be the causative 467 mutation of the effect found in this study because this SNP leads to a change in 468 an amino acid. Despite the long distance between the SNPs located in the 469 promoter and exon 2 (approximately 24 kb) and the low linkage disequilibrium found between them (the maximum values of r² and D' between snp 8 and the 470 471 significant SNPs in promoter region for snp_29 snp_30, and snp_34 were 0.06

and 0.33, respectively), it appears that the effects observed for the SNPs in thepromoter region are due to the linkage disequilibrium with snp_8.

474

475 **4. Conclusions**

476 We have sequenced the entire coding region plus 733 and 251 bp from the 477 promoter and 3'-UTR regions, respectively, in the MTNR1A gene in 268 Rasa 478 aragonesa sheep breed ewes, finding 13 SNPs associated with seasonality 479 traits in the promoter and exon 2 regions. Haplotype analyses confirmed these 480 results and allowed an assessment of the impact of the SNPs located in the 481 promoter and exon 2 regions on the effects observed for the seasonality values. 482 In this sense, alleles associated with lower TDA values (or higher values for 483 P4CM) in the promoter are in linkage disequilibrium with those in exon 2 484 responsible for the effects found in this study, and they can be segregated 485 together. Thus, the observed effects could be due to SNPs in exon 2 but not the 486 SNPs in the promoter. We have demonstrated that the snp_8 T allele in the 487 MNTR1A gene is associated with lower and higher TDA and P4CM values, respectively, both of which are seasonality traits for ovarian function based on 488 blood progesterone levels; this allele is also associated with higher OCM 489 490 values, which indicates behavioural signs of oestrous in the Rasa Aragonesa 491 breed. This SNP is in linkage disequilibrium with snp_17 (612/MnII) and snp_18 492 (606/Rsal), which have been related with seasonal reproductive activities in 493 several association studies with other breeds. However, these SNPs are 494 synonymous polymorphisms and do not appear to be the causative mutations. 495 In this sense, snp 8 could be the causative mutation for the observed effects in

496 the reproductive seasonality traits because it produces an amino acid change497 from an Arginine to a Cysteine (R336C).

498

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602 Figure captions

Fig. 1. Schematic representation of ovine MTNR1A and the linkage disequilibrium plot for the SNPs in *MTNR1A* for the Rasa aragonesa population using Haploview. MTNR1A is in a reverse orientation on the genome, and the SNPs are ordered according to their position in the latest genome version (Oar4.0: GenBank acc. number NW_014639035.1). The linkage disequilibrium colour scheme corresponds with the D' parameter, while the linkage disequilibrium values correspond to the r^2 parameter. Haplotype blocks are indicated by dark lines. A strong LD (D' = 1, LOD \geq 2) is indicated by red, and lighter shades of pink indicate varying degrees of LD, with lighter shades displaying less than darker shades (D' < 1, LOD \geq 2). White indicates low LD (D' < 1, LOD < 2).

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King the second **Table 1.** Information about the location and amino acid substitution effect of the identified SNPs in the *MTNR1A* gene and their genotypic and allelic frequencies. *MTNR1A* is in a reverse orientation on the genome, and the SNPs are ordered according to their positions in the latest genome version (Oar4.0: GenBank acc. number NW_014639035.1).

Location	Alias ¹	dbSNPs	Oar 4.0 (NW_014639035.1)	Amino acid substitution	Genotype	Genotype frequencies	Allele	Allele frequencies
exon 2	snp_1	rs420016236	g.15098857C>T	3' UTR	CC 🔨	0.597	С	0.78
				variant	СТ	0.366	Т	0.22
					TT	0.037		
	snp_2	rs400830807	g.15098860G>A	3' UTR	GG	0.544	G	0.742
				variant	GA	0.392	А	0.258
					AA	0.064		
	snp_3	rs414185743	g.15098868T>C	3' UTR	TT	0.322	Т	0.568
				variant	TC	0.493	С	0.432
					CC	0.185		
	snp_4	rs423194759	g.15098916T>C	3' UTR	TT	0.322	Т	0.568
				variant	TC	0.493	С	0.432
					CC	0.185		
	snp_5	rs403826495	g.15098968T>C	I348V	TT	0.322	Т	0.568
				(ATA/GTA)	TC	0.493	С	0.432
					CC	0.185		
	snp_6	rs413084140	g.15098976T>C	H345R	TT	0.322	Т	0.568
				(CAT/CGT)	TC	0.493	С	0.432
					CC	0.185		
	snp_7	rs426523476	g.15098996A>G	P338P	AA	0.182	G	0.57
				(CCC/CCT)	AG	0.496	А	0.43
					GG	0.322		
	snp_8	rs403212791	g.15099004G>A	R336C	GG	0.511	G	0.724
				(CGC/TGC)	GA	0.425	А	0.276
					AA	0.064		
	snp_9	ss2137144054	g.15099006T>G	K335T	TT	0.996	Т	0.998
				(AAA/ACA)	GT	0.004	G	0.002
					GG	0		

			ACCEDTEL	NANUCODDT				
sr	np_10	rs416266900	g.15099204G>T	A269D	GG	0.613	G	0.786
				(GCC/GAC)	GT	0.346	Т	0.214
					TT	0.041		
sr	np_11	rs429718221	g.15099206G>A	P268P	GG	0.333	G	0.576
				(CCC/CCT)	GA	0.486	А	0.424
					AA	0.181		
sr	np_12	rs404378206	g.15099223C>T	V263I	CC	0.909	С	0.955
				(GTT/ATT)	СТ	0.091	Т	0.045
					TT 🔨	0		
sr	np_13	rs417800445	g.15099296C>T	R238R	CC	0.624	С	0.796
				(AGG/AGA)	СТ	0.342	Т	0.204
					T	0.034		
sr	np_14	rs427019119	g.15099314C>T	L232L	CC	0.617	С	0.792
				(CTG/CTA)	СТ	0.35	Т	0.208
					TT	0.033		
sr	np_15	rs407388227	g.15099391C>T	V207I	CC	0.634	С	0.794
				(GTC/ATC)	СТ	0.321	Т	0.206
					TT	0.045		
sr	np_16	rs420819884	g.15099422C>T	V196V	CC	0.918	С	0.955
				(GTG/GTA)	СТ	0.074	Т	0.045
					TT	0.008		
sr	np_17	rs430181568	g.15099485C>T	P175P	CC	0.634	С	0.792
				(CCG/CCA)	СТ	0.317	Т	0.208
					TT	0.049		
sr	np_18	rs406779174	g.15099491G>A	Y173Y	GG	0.28	G	0.525
				(TAC/TAT)	GA	0.49	А	0.475
					AA	0.23		
sr	np_19	ss2137144055	g.15099575C>T	T145T	CC	0.922	С	0.959
			7	(ACG/ACA)	СТ	0.074	Т	0.041
					TT	0.004		
sr	np_20	ss2137144056	g.15099642C>A	G123V	CC	0.996	С	0.998
				(GGA/GTA)	CA	0.004	А	0.002
					AA	0		
sr	np_21	rs419680097	g.15099644C>A	T122T	CC	0.638	С	0.79
				(ACG/ACT)	CA	0.305	А	0.21

			ACCEPTEI) MANUSCRIPT		0.057		
					AA	0.057	-	
exon 1	snp_22	ss2137144057	g.15121956G>A	N16N	GG	0.696	G	0.761
				(AAT/AAC)	GA	0.13	A	0.239
					AA	0.174		
Promoter	snp_23	rs406334919	g.15122684T>C		TT	0.699	Т	0.831
					тс	0.265	С	0.169
					CC	0.036		
	snp_24	rs419743392	g.15122766G>A		GG	0.379	G	0.601
					GA	0.445	А	0.399
					AA	0.176		
	snp_25	rs400561563	g.15122788C>T		CC	0.544	С	0.735
					СТ	0.382	Т	0.265
					TT	0.074		
	snp_26	rs399461430	g.15122829T>C		TT	0.379	Т	0.601
					тс	0.445	С	0.399
					CC	0.176		
	snp_27	rs412826644	g.15122893G>A		GG	0.915	G	0.956
					GA	0.081	А	0.044
					AA	0.004		
	snp_28	rs426266687	g.15122902C>T		CC	0.382	С	0.605
					СТ	0.445	Т	0.395
					TT	0.173		
	snp_29	rs402949406	g.15122931C>A		CC	0.386	С	0.607
					CA	0.441	А	0.393
					AA	0.173		
	snp_30	rs411931887	g.15122934C>T		CC	0.387	С	0.607
					СТ	0.441	Т	0.393
					TT	0.173		
	snp_31	rs406184829	g.15123052T>C		TT	0.695	Т	0.829
					тс	0.268	С	0.171
					CC	0.037		
	snp_32	rs415456480	g.15123097G>A		GG	0.379	G	0.601
					GA	0.445	А	0.399
					AA	0.176		
	snp_33	rs428880789	g.15123143G>A		GG	0.379	G	0.601

			C CD IDT			
		ACCEPTED MANUS	GA	0.445	A	0.399
			AA	0.176		
snp_34	rs405080439	g.15123157G>A	GG	0.386	G	0.607
		-	GA	0.441	А	0.393
			AA	0.173		
snp_35	rs429917252	g.15123238C>T	CC	0.699	С	0.831
			СТ	0.265	Т	0.169
			ТТ	0.036		
neclature used for each SNP in this	work.					

¹Nomeclature used for each SNP in this work.

Table 2. Type III test for the SNP and SNP x age effects for the *MTNR1A* polymorphisms using the seasonality phenotype data from Rasa aragonesa ewes. The least square means and standard errors of the *MTNR1A* polymorphisms in the seasonality phenotype data in Rasa aragonesa ewes are also shown. Only significant SNPs after Bonferroni correction are shown. M=mature; Y=young; L=ewe lambs. snp_29-snp_30-snp_34 and snp_24-snp_26-snp_32-snp_33 were linked (r^2 =1). Different letters indicate significant differences: a, b: P<0.05; c, d: P<0.01; and e, f: P<0.001.

		P-value				P-value				
SNP	Trait	SNP		LSmeans SNP		SNPxAge	Age		LSmeans SNPxA	ge
			AA/CC/GG	AC/CT/AG	CC/TT/AA			AA/CC/GG	AC/CT/AG	CC/TT/AA
snp_29	TDA	0.01	68.72 <u>+</u> 7.33c	58.84 <u>+</u> 7.13cd	37.05 <u>+</u> 11.29d	0.012	Μ	54.74 <u>+</u> 9.96	46.61 <u>+</u> 9.05	61.56 <u>+</u> 11.44
snp_30							Y	63.45 <u>+</u> 10.55	63.67 <u>+</u> 10.13	49.51 <u>+</u> 14.24
snp_34							L	87.96 <u>+</u> 10.96a	66.23 <u>+</u> 10.94ab	0.09 <u>+</u> 25.91b
	P4CM	0.015	0.81 <u>+</u> 0.03a	0.85 <u>+</u> 0.03ab	0.93 <u>+</u> 0.04b	0.03	Μ	0.85 <u>+</u> 0.03	0.88 <u>+</u> 0.03	0.83 <u>+</u> 0.04
							Y	0.82 <u>+</u> 0.04	0.82 <u>+</u> 0.04	0.88 <u>+</u> 0.05
_							L	0.76 <u>+</u> 0.04	0.83 <u>+</u> 0.04	1.08 <u>+</u> 0.1
			GG/TT/GG/GG	AG/AG/CT/AG	AA/CC/AA/AA			GG/TT/GG/GG	AG/AG/CT/AG	AA/CC/AA/AA
snp_24	TDA	0.008	68.86+7.32c	58.48+7.15cd	36.58+11.26d	0.013	Μ	55.83 <u>+</u> 10.04	46.21 <u>+</u> 9.03	90.81 <u>+</u> 11.37
snp_26							Y	63.16 <u>+</u> 10.55	63.37 <u>+</u> 10.13	49.19 <u>+</u> 14.23
snp_32							L	87.58 <u>+</u> 10.78a	65.84 <u>+</u> 10.76ab	-0.25 <u>+</u> 25.11b
snp_33	P4CM	0.013	0.81+0.03a	0.85+0.03ab	0.93+0.04b	0.033	Μ	0.85 <u>+</u> 0.04	0.89 <u>+</u> 0.03	0.84 <u>+</u> 0.04
							Y	0.82 <u>+</u> 0.04	0.83 <u>+</u> 0.04	0.88 <u>+</u> 0.04
							L	0.73 <u>+</u> 0.04	0.86 <u>+</u> 0.04	1.08 <u>+</u> 0.1
			TT	СТ	CC			TT	СТ	CC
snp_28	TDA	0.008	37.02+11.27c	58.50+7.13cd	69.10+7.31c	0.012	Μ	61.60 <u>+</u> 11.42	45.77 <u>+</u> 9.06	56.00 <u>+</u> 9.93
							Y	49.44 <u>+</u> 14.22	63.61 <u>+</u> 10.12	63.41 <u>+</u> 10.54
							L	0.00 <u>+</u> 25.59a	66.13 <u>+</u> 10.74ab	87.87 <u>+</u> 10.77b
	P4CM	0.013	0.93+0.04a	0.85+0.03ab	0.81+0.03b	0.03	Μ	0.83 <u>+</u> 0.04	0.89 <u>+</u> 0.03	0.85 <u>+</u> 0.04
							Y	0.88 <u>+</u> 0.05	0.83 <u>+</u> 0.04	0.82 <u>+</u> 0.04
							L	1.08 <u>+</u> 0.04	0.82 <u>+</u> 0.04	0.76 <u>+</u> 0.1
			CC	СТ	TT			CC	СТ	TT
snp_25	TDA	0.025	52.40+7.15b	65.52+7.54a	68.99+11.93ab	0.104	Μ	47.50 <u>+</u> 9.49	52.83 <u>+</u> 9.35	46.69 <u>+</u> 18.14
							Y	56.84 <u>+</u> 9.58	66.05 <u>+</u> 10.86	54.57 <u>+</u> 22.70
							L	52.85 <u>+</u> 10.60	86.66 <u>+</u> 12.09	105.7 <u>+</u> 16.09
	P4CM	0.005	0.88+0.03d	0.80+0.03c	0.80+0.04cd	0.089	Μ	0.88 <u>+</u> 0.04	0.86 <u>+</u> 0.04	0.88 <u>+</u> 0.07
							Y	0.85 <u>+</u> 0.04	0.82 <u>+</u> 0.04	0.83 <u>+</u> 0.09
							L	0.90 <u>+</u> 0.04	0.73 <u>+</u> 0.05	0.71 <u>+</u> 0.6

Table 3. Type III test for the SNP effects on the *MTNR1A* gene using the seasonality phenotype data from Rasa aragonesa ewes. The least square means and standard errors for the SNP effects on the *MTNR1A* polymorphisms are also shown. Only significant SNPs after Bonferroni correction are shown. Different letters indicate significant differences: a, b: P<0.05; c, d: P<0.01; and e, f: P<0.001.

		P-value			
SNP	Trait	SNP	I	Smeans SNP	
snp_2			GG	GG GA	
	TDA	0.0001	66.62 <u>+</u> 7.91a,c	48.79 <u>+</u> 7.88b	16.25 <u>+</u> 15.15d
	P4CM	0.003	0.81 <u>+</u> 0.03a	0.87 <u>+</u> 0.03b	0.96 <u>+</u> 0.06b
	OCM	0.015	0.49 <u>+</u> 0.03a	0.55 <u>+</u> 0.03ab	0.64 <u>+</u> 0.07b
snp_8			GG	GA	AA
	TDA	<0.0001	73.08 <u>+</u> 7.25c,e	53.57 <u>+</u> 7.36a,d	20.85 <u>+</u> 13.13b,f
	P4CM	0.001	0.80 <u>+</u> 0.03a,c	0.86 <u>+</u> 0.03b	0.94 <u>+</u> 0.05b,d
	OCM	0.022	0.49 <u>+</u> 0.03a 🗸	0.54 <u>+</u> 0.03ab	0.64 <u>+</u> 0.06b
snp_17			TT	TC	CC
	TDA	0.037	78.41 <u>+</u> 14.37	59.38 <u>+</u> 8.40	50.71 <u>+</u> 7.94
	P4CM	0.073	0.76 <u>+</u> 0.06	0.83 <u>+</u> 0.03	0.86 <u>+</u> 0.03
	OCM	0.018	0.39 <u>+</u> 0.06a	0.50 <u>+</u> 0.04ab	0.55 <u>+</u> 0.03b
snp_18		0.0002	72.25 <u>+</u> 9.01e	58.05 <u>+</u> 7.78c	33.28 <u>+</u> 9.69f,d
	TDA	0.003	0.79 <u>+</u> 0.03c	0.84 <u>+</u> 0.03a	0.92 <u>+</u> 0.04b,d
	P4CM	0.02	0.46 <u>+</u> 0.04a	0.50 <u>+</u> 0.03ab	0.57 <u>+</u> 0.04b
	OCM	0.0002	72.25 <u>+</u> 9.01e	58.05 <u>+</u> 7.78c	33.28 <u>+</u> 9.69f,d

Table 4. Type III test for the haplotype and haplotype x age effects for blocks 3 and 4 on the *MTNR1A* gene promoter region using the seasonality phenotype data from Rasa aragonesa ewes. The least square means and standard errors for the haplotype and haplotype x age effects for the *MTNR1A* haplotypes are also shown. Only significant SNPs after Bonferroni correction are shown. Different letters indicate significant differences: a, b: P<0.05; c, d: P<0.01; and e, f: P<0.001.

					SNP effect ²					SN	P x age effect ²	
Trait	Block ¹	Haplotype	Freq.	P-value	0 сору	1 сору	2 copies	P-value	Age	0 сору	1 copy	2 copies
TDA	3	h3 (TG)	0.61	0.010	35.52+11.35c	57.61+7.29cd	67.32+7.43d	0.013	М	58.98+11.51	44.94+9.23	52.56+10.22
					—	—			Υ	48.2 <u>+</u> 14.35	62.36 <u>+</u> 10.26	62.15 <u>+</u> 10.67
									L	-0.61 <u>+</u> 25.7a	65.53 <u>+</u> 10.87ab	87.27 <u>+</u> 10.89b
	4	h1 (TCACTGGGC)	0.61	0.011	35.1 <u>+</u> 11.32c	57.64 <u>+</u> 7.29cd	66.94 <u>+</u> 7.41d	0.016	М	58.10 <u>+</u> 11.34	44.38 <u>+</u> 9.35	52.86 <u>+</u> 10.23
									Y	47.96+14.32	62.13 +10.23	61.91+10.64
									L	-0.76 <u>+</u> 25.7a	66.40 <u>+</u> 10.78ab	86.94 <u>+</u> 10.89b
P4CM	3	h3 (TG)	0.61	0.016	0.93 <u>+</u> 0.04a	0.85 <u>+</u> 0.03ab	0.082 <u>+</u> 0.03b	0.036	М	0.84 <u>+</u> 0.04	0.89 <u>+</u> 0.03	0.86 <u>+</u> 0.04
					_		_		Y	0.88 <u>+</u> 0.05	0.83 <u>+</u> 0.04	0.83 <u>+</u> 0.04
									L	1.08 <u>+</u> 0.10	0.83 <u>+</u> 0.04	0.76 <u>+</u> 0.04
	4	h1 (TCACTGGGC)	0.61	0.015	0.93 <u>+</u> 0.04a	0.85 <u>+</u> 0.03ab	0.82 <u>+</u> 0.03b	0.031	М	0.84 <u>+</u> 0.04	0.89 <u>+</u> 0.04	0.86 <u>+</u> 0.04
					—		_		Y	0.88 <u>+</u> 0.05	0.83 <u>+</u> 0.04	0.83 <u>+</u> 0.05
						× [▼]			L	1.08 <u>+</u> 0.10	0.83 <u>+</u> 0.04	0.69 <u>+</u> 0.07

¹ Block 3: snp_23 - snp_24 and Block4: snp_26 - snp_28 - snp_29 - snp_30 - snp_31 - snp_32 - snp_33 - snp_34 - snp_35.

² M=mature; Y=young; L=ewe lambs. 0 copy: LSmeans and SE for 0 copy of the haplotype; 1 copy: LSmeans and SE for 1 copy of the haplotype; and 2 copies: LSmeans and SE for 2 copies of the haplotype.

Table 5. Type III test for the haplotype effects of blocks 1 and 2 in exon 2 on the *MTNR1A* gene using the seasonality phenotype data from Rasa aragonesa ewes. The least square means and standard errors for the haplotype effect for the *MTNR1A* gene are also shown. Only significant SNPs after Bonferroni correction are shown. Different letters indicate significant differences: a, b: P<0.05; c, d: P<0.01; and e, f: P<0.001

					SNP effect ²		
Trait	Block ¹	Haplotype	Freq.	P-value	0 сору	1 copy	2 copies
TDA	1	h3 (CATCTTG)	0.26	0.0001	66.62 <u>+</u> 7.91a,e	48.79 <u>+</u> 7.88b	16.25 <u>+</u> 15.15b,f
	2	h2 (AGGCCCCAC)	0.26	0.0004	65.13 <u>+</u> 7.85a,e	50.33 <u>+</u> 7.96b	13.78 <u>+</u> 15.42a,f
P4CM	1	h3 (CATCTTG)	0.26	0.0029	0.81 <u>+</u> 0.03a	0.87 <u>+</u> 0.03b	0.96 <u>+</u> 0.06b
	2	h2 (AGGCCCCAC)	0.26	0.014	0.82 <u>+</u> 0.03a	0.86 <u>+</u> 0.03ab	0.97 <u>+</u> 0.06b
OMC	1	h3 (CATCTTG)	0.26	0.015	0.49 <u>+</u> 0.03a	055 <u>+</u> 0.03b	0.64 <u>+</u> 0.06b
	2	h2 (AGGCCCCAC)	0.26	0.023	0.49 <u>+</u> 0.03a	0.54 <u>+</u> 0.03ab	0.64 <u>+</u> 0.07b

¹Block1: snp_1 - snp_2 - snp_3 - snp_4 - snp_5 - snp_6 - snp_7; Block2: snp_8 - snp_10 - snp_11 - snp_13 - snp_14 - snp_15 - snp_17 - snp_18 - snp_21

² 0 copy: LSmeans and SE for 0 copy of the haplotype; 1 copy: LSmeans and SE for 1 copy of the haplotype; and 2 copies: LSmeans and SE for 2 copies of the haplotype.

Table 6. Type III test for the haplotype effects of block 5 on the *MTNR1A* gene using the seasonality phenotype data from Rasa aragonesa ewes. The least square means and standard errors for the haplotype effect on the *MTNR1A* gene are also shown. Only significant haplotypes after Bonferroni correction are shown. Different letters indicate significant differences: a, b: P<0.05; c, d: P<0.01; and e, f: P<0.001. Alleles associated with lower TDA and higher P4CM and OMC values for each SNP according to SNP association results are shown in bold.

				SNP effect ²		
Trait	Haplotype	Freq.	P-value	0 сору	1 copy	2 copies
TDA	h4(AACAACCTCTAAA)	0.17	0.0387	63.17 <u>+</u> 7.88a	48.77 <u>+</u> 8.44b	37.25 <u>+</u> 21.15ab
	h12 (GG C GAT CTCTAAA)	0.02	0.029	56.83 <u>+</u> 7.42a	98.48 <u>+1</u> 7.77b	
	h13 (AACA GTTCACGGG)	0.05	0.0014	59.20 <u>+</u> 7.45c	30.74 <u>+</u> 11.94d	
P4CM	h4(AACAACCTCTAAA)	0.17	0.014	0.81 <u>+</u> 0.03a	0.87 <u>+</u> 0.03b	0.97 <u>+</u> 0.08b
	h12 (GG C G A T CTCTAAA)	0.02	0.040	0.83 <u>+</u> 0.03a	0.71 <u>+</u> 0.07b	
	h13 (AACA GTTCACGGG)	0.05	0.026	0.83 <u>+</u> 0.03a	0.92 <u>+</u> 0.04b	
OMC	h10 (AACAGCTCACGGG)	0.003	0.048	0.50 <u>+</u> 0.03a	0.61 <u>+</u> 0.03b	
	h13 (AACA GTTCACGGG)	0.05	0.0014	0.50 <u>+</u> 0.03c	0.65 <u>+</u> 0.05ad	

¹ Block 5: snp_2 - snp_8 - snp_17 - snp_18 - snp_24 - snp_25 - snp_26 - snp_28 - snp_29 - snp_30 - snp_32 - snp_33 - snp_34

² 0 copy: LSmeans and SE for 0 copy of the haplotype; 1 copy: LSmeans and SE for 1 copy of the haplotype; and 2 copies: LSmeans and SE for 2 copies of the haplotype.



Highlights

- Significant SNPs in promoter and exon 2 of the MTNR1A were found.
- Haplotype analyses showed that the effects found were caused by the SNPs in exon 2.
- SNP rs403212791 in exon 2 gene is associated to reproductive seasonality
- The SNP rs403212791 leads a change of an Arginine to Cysteine (R336C).
- The SNP rs403212791 was not previously reported associated for reproductive seasonality.