

AGEING AND RETAIL DISPLAY TIME IN RAW BEEF ODOUR ACCORDING TO THE DEGREE OF LIPID OXIDATION

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ABSTRACT

This study aims to assess the changes in beef aroma over time when steaks are stored in retail display under high oxygen conditions from pre-aged knuckles for 15 or 22 days in vacuum conditions. Odorous volatile compounds were analysed by solid phase microextraction (SPME) / gas chromatography mass spectrometry (GC-MS). Results were grouped as *low*, *medium* and *high* oxidative groups according to thiobarbituric acid reacting substances values after 9 days of display. The intensity of off-odours in the raw meat increased with ageing and display time and oxidative groups. Based in correlations between the variables and regressions of the compounds through display, eight compounds were proposed as odour shelf-life markers. Among them, five were

most sensitive and precise in all oxidative groups: 1-hexanol in meat aged for 15 days, ethyl- octanoate and 2-pentylfuran in meat aged for 22 days, and penta and hexanoic acids in the two studied ageing times.

KEYWORDS

SPME-GCMS, volatile compounds, meat aroma, modified atmosphere packaging, meat quality

1. INTRODUCTION

Beef is usually aged for 7 to 21 days in order to improve tenderness and homogeneity, being the optimum period dependent on several factors, including the muscle type and the breed under consideration (Monsón, Sañudo & Sierra, 2005; Thompson, 2002). In addition, ageing could affect other quality parameters, such as odour, taste and flavour in the cooked meat due to the change in the concentration of precursors (Koutsidis, Elmore, Oruna-Concha, Campo, Wood & Mottram, 2008; Meinert et al., 2009), but also to the development of odorous compounds in the raw product coming from chemical and enzymatic reactions and bacterial action (Casaburi, Piombino, Nychas, Villani & Ercolini, 2015; Estévez, Morcuende, Ventanas & Cava, 2003; Insausti, Beriain, Gorraiz & Purroy, 2002). The mentioned processes occur even when ageing is conducted at refrigeration, vacuum and dark conditions. Nevertheless, after cooking the meat could reach high flavour acceptability scores (Colle et al., 2015; Pérez-Juan et al., 2014; Smith et al., 2008). In fact, ageing up to 21 days improves the flavour in some beef breeds (Monsón et al., 2005).

Commonly, after ageing the meat is packed and preserved in retail display during some days until purchasing. It has been shown that steaks displayed without oxygen improve the eating quality (Aaslyng, Tørngren & Madsen, 2010), but at commercial level steaks are still mostly sold in oxygen-permeable overwrap or in packs with high oxygen levels due to colour issues. Previous studies found unacceptable rancid flavour developed in beef *Longissimus dorsi* displayed for around nine days in high oxygen packs (Campo, Nute, Hughes, Enser, Wood & Richardson, 2006; Resconi, Escudero, Beltrán, Olleta, Sañudo & Campo, 2012), but shorter or extended periods of odour shelf-life would occur depending on numerous factors such as the antioxidant status of the meat. In a study with lambs, for example, dietary supplementation of vitamin E led to lower levels

of lipid oxidation derived compounds, such as 2-heptanone and 1-penten-3-ol in the cooked meat (Rivas-Cañedo et al., 2013), which may affect the odour perception of the meat.

The effect of time when ageing the meat or when stored in retail display under high oxygen conditions in the beef aroma development has been previously studied using cooked samples (Campo et al., 2006; Gorraiz, Beriain, Chasco & Insausti, 2002; Ma, Hamid, Bekhit, Robertson & Law, 2012; O'Quinn et al., 2016; Resconi et al., 2012; Stetzer, Cadwallader, Singh, McKeith & Brewer, 2008; Watanabe, Kamada, Imanari, Shiba, Yonai & Muramoto, 2015), but less has been published regarding the raw meat (Insausti et al., 2002; La Storia et al., 2012; Saraiva, Oliveira, Silva, Martins, Ventanas & García, 2015). Most studies have used cooked meat matching the frequent way of consumption, but also because a richer variety and intensity of aromas is developed (Resconi, Escudero & Campo, 2013). The odour of the raw meat in the store while buying is becoming less important since the meat is usually sold packaged. However, when opening the package at home, the odour should be agreeable for the consumer (Schindler, Krings, Berger & Orlie, 2010). Furthermore, some compounds already present or developed in the raw meat, remain after cooking and could affect the flavour perception (Insausti et al., 2002; Rota & Schieberle, 2005; Schindler et al., 2010).

The aim of this study was to evaluate the effect of display time on the odour characteristics and the volatile compounds involved in raw knuckle beef steaks pre-aged under vacuum conditions during 15 or 22 days, according to the extent of lipid oxidation. Furthermore, it also aims to identify shelf-life markers between the volatile compounds analysed. It is already known that lipid oxidation affects odour development; therefore samples will be grouped by their oxidative potential, since

different key compounds may affect the change in the aroma characteristics through display.

2. MATERIAL AND METHODS

2.1 Samples, ageing and display conditions

This study used 48 crossbred entire young bulls 12.8-13.9 mo with a cold carcass weight of 231-340 kg and an intramuscular fat content in *longissimus thoracis* muscle of 1.5-2.5 %. The animals were raised on the same farm and fed concentrates (based on maize, barley and soya) and cereal straw *ad libitum*. After slaughtering in a commercial abattoir following standard procedures meeting welfare regulations, both knuckles from each animal were obtained, vacuum packaged and aged for either 15 days (left side) or 22 days (right side), in the dark at 3 ± 1 °C. Then, 0.6-cm thick steaks were obtained and randomly allocated for each display time. Samples from each animal were distributed in all ageing and display times. The samples from day 0 of display were analysed within the day of sampling, whereas the rest of the steaks were placed individually in polyethylene commercial trays, flushed with a food-grade gas mixture: 80% of oxygen and 20% of carbon dioxide, as commonly used at commercial level, and sealed with a polyethylene and polyamide laminate film (50 µm, water vapour transmission rate of $5 \text{ g/m}^2\cdot\text{day}$ at 23 °C and 85% RH, an O₂ transmission rate of $5 \text{ cm}^3/\text{m}^2\cdot\text{day}$ at 23 °C and 0% RH and a CO₂ transmission rate of $25 \text{ cm}^3/\text{m}^2\cdot\text{day}$ at 23 °C and 0% RH; Linpac Packaging S.L., Spain). Then, trays were placed in simulated retail display (Koxka, V1VI1-5; Pamplona, Spain), under light (cool white fluorescent illumination, 1200 lux, 16 h on, 8 off, Mazdafluor Aviva TF/36w; Philips, Eindhoven, Holland), at $4 \text{ °C}\pm 1 \text{ °C}$ for 5, 7 or 9 days. Lipid oxidation, odour and volatile compounds analyses were

conducted in the *rectus femoris* muscle from the same steak. Such muscle was minced and aliquots for each analysis were immediately taken.

2.2 Lipid oxidation and oxidative groups

Lipid oxidation was measured with the thiobarbituric acid reactive substances (TBARS) method of Pfalzgraf, Frigg and Steinhart (1995). Meat samples of 10 g were homogenized with 20 mL of trichloroacetic acid (10%, VWR) using an Ultra-Turrax T25 (Janke & Kunkel, Staufen, Germany). Samples were centrifuged (Gyrozen 1248R, Daejeon, Korea) at 4000 rpm for 30 min at 4 °C and the supernatants filtered through qualitative paper (F1093 grade, Chmlab, Barcelona, Spain). Two millilitres of the filtrates were taken in duplicates and mixed with 2 mL of thiobarbituric acid (20 mM, Sigma, Aldrich), homogenized and incubated for 20 min in a water bath (Grant W14, Cambridge, UK) at 97 °C. Absorbance was measured at 532 nm (spectrophotometer Unicam 5625 UV/VIS, Cambridge, UK). TBARS values were calculated from a daily standard curve of 1,1,3,3-tetramethoxypropane (TMP, Sigma Aldrich), and expressed as mg malondialdehyde (MDA)/kg sample. Based on the results obtained at day 9 of display in pre-aged knuckles for 15 days, samples were grouped according to the extent of lipid oxidation in low oxidative ($n=16$ animals/ageing), when less than 1 mg MDA/kg was obtained, medium ($n=15$ animals/ageing), when TBARS values between 1-2 mg MDA/kg were found and high oxidative ($n=17$ animals/ageing), when TBARS value exceeded 2 (Campo et al., 2006). Thus, each animal was allocated in a single level of oxidation.

2.3 Volatile compounds analysis

2.3.1 Chemicals

Dipropylene glycol 99% was supplied by Alfa Aesar (Karlsruhe, Germany); (E,E)-2,4-decadienal $\geq 89\%$ by Lancaster Synthesis (Eastgate, UK); butanoic acid 99.5%, pentanoic acid 98%, octanoic acid 98%, ethyl hexanoate 99% and 1-octen-3-one $>99\%$ by Fluka (Madrid Spain); hexanoic acid 99.5%, heptanoic acid 99%, nonanoic acid $\geq 96\%$, decanoic acid $\geq 98\%$, hexanal 98%, nonanal 95%, (E)-2-heptenal $\geq 95\%$, (E)-2-octenal 94%, (E)-2-nonenal 97%, (E,E)-2,4-nonadienal $\geq 85\%$, (E)-2-undecenal $\geq 95\%$, phenylacetaldehyde $>95\%$, 2-butanone $>99\%$, 2-heptanone $\geq 98\%$, 3-octanone $\geq 98\%$, 2-octanone $\geq 98\%$, 2-nonanone $\geq 99\%$, 3-octen-2-one $\geq 98\%$, 3-nonen-2-one 95%, 1-hexanol 98%, 1-octen-3-ol 98%, ethyl octanoate $\geq 98\%$, methyl 2-methylbutanoate $\geq 98\%$ and 2-pentylfuran ≥ 98 by Aldrich (Madrid, Spain). The compounds (Z)-2-octenal and (E,Z)-2,4-decadienal were found in commercial (E)-2-octenal and (E,E)-2,4-decadienal, respectively.

2.3.2 Sample preparation and extraction by HS-SPME

For each steak, 4 g (± 0.001) of minced meat was collected in the bottom of 20 mL SPME vial. The vial was purged for 20 seconds with nitrogen and immediately crimp capped to avoid the presence of oxygen in the headspace. Samples were kept at 4 °C until analysis.

The vial was then equilibrated at 37 °C for 10 min in the automated sample preparation unit. After that, a polydimethylsiloxane/divinylbenzene (PDMS/DVB) 65 μm film thickness fibre (Supelco-Spain, Madrid, Spain) was exposed to the headspace of the unstirred sample for 40 min at 37 °C and then desorbed directly in the injection port of the chromatographic system in the conditions detailed below. After desorption, the fibre

was cleaned for 10 min at 250 °C in the back out unit. Total automation of the extraction procedure was achieved using a CTC CombiPAL autosampler (Zwingen, Switzerland), which was programmed using the CycleComposer with macroeditor software and equipped with sample trays and temperature controlled incubator tray.

2.3.3 Gas chromatography-mass spectrometry conditions

The instrument was a CP-3800 chromatograph coupled to a Saturn 2200 ion trap mass spectrometric detection system supplied by Varian (Sunnyvale, CA, USA). The fibre was desorbed directly in the injection port of the GC-MS in splitless mode for 5 min at 250 °C and a pressure pulse of 30 psi during this period of time (the column flow during this period of time was 2 mL/min). Helium was the carrier gas at a flow rate of 1 mL/min.

A DB-WAXETR capillary column (J&W Scientific, Folsom, CA, USA) of 60 m × 0.25 mm I.D., a film thickness of 0.25 µm, and preceded by a 3 m × 0.25 mm uncoated (deactivate, intermediate polarity) precolumn from Supelco-Spain was used. The oven temperature was initially 40 °C during 10 min, then raised by 4 °C/min to 140 °C, followed by a rate of 10 °C/min to 220 °C and finally held at this temperature for 5 min. The MS parameters were a transfer line temperature of 230 °C and a trap temperature of 170 °C with emission current of 80 µA. The global run time was recorded in full scan mode (33–300 m/z mass range). The chromatographic data were analysed by Varian Saturn GC-MS Workstation Version 5.52 software. The identity of 32 odorants was determined through the mass spectra and the linear retention indices and confirmed by the injection of the pure reference standards when available.

Given that some of the selected analytes did not give a signal in TIC or else appeared together with peaks of interference that make it difficult to integrate, it was decided to consider selective mass.

The quantitation of the compounds using selective mass was carried out applying a response factor obtained calculated with a dipropylene glycol control solution analysed by GC-MS every 16 meat samples. This control solution contained a compound of each family to which its response factor was calculated. This response factor was subsequently applied to all compounds of the same family: butanoic acid (acids), 1-hexanol (alcohols), nonanal (saturated aldehydes), (E)-2-nonenal (alkenals), methyl 2-methylbutanoate (esters), 3-octanone (saturated ketones), 1-octen-3-one (unsaturated ketones) and 2-pentylfuran.

2.4 Odour characterisation

The sensory analysis was conducted by six panellists, one male and five females, from 30 to 58 years old, experienced in the sensory analysis of meat and meat products. Descriptors were developed by consulting bibliography (Schevey, Toshkov & Brewer, 2013) and by agreement of the members in a session where two samples of raw beef (a vacuum packed sample and a sample stored seven days in high oxygen pack) were presented for training. Panellists rated the intensity of the odour, using a lineal scale from 0 (no detected) to 10 (very intense odour). The seven descriptors were: raw meat odour (odour associate with raw minced beef), rancid odour (odour associated with oxidized fat), metallic odour (odour associated with a ferrous sulphate solution), sweet odour (odour associated with sugar), musty odour (odour associated with a wet

cardboard/highly humid room), cooked meat odour (odour associated with a stewed beef) and sulphur odour (odour associated with a cooked yolk of an egg).

Similarly and simultaneously than for the volatile analysis, after opening the package at each day of display, steaks were minced and 4 g were transferred to a snap clap glass vials (20 mL) and kept at 4 ± 2 °C until the evaluation within three hours of preparation. The evaluation was performed in a room with controlled temperature (20 °C). The samples were tempered at room temperature for 30 min before analysed. Panellists sniffed the same vial on intervals of at least 15 min to allow the aroma compounds to reach the balance in the headspace.

2.5 Data analysis

A General Linear Model (GLM) was applied with oxidative group (<1 (low), 1-2 (medium), >2 mg (high) MDA/kg) ageing (15 and 22 days) and display time (0, 5, 7, 9 days) as fixed effects. Since interactions were significant ($P\leq 0.05$), data were analysed within ageing and oxidative group with display time as fixed effect and within ageing and display time with oxidative group as fixed effect. A Tukey test was performed to show differences between display days and oxidative groups. Linear and exponential regressions for the volatile compounds through display were conducted. Finally, Pearson correlations between the variables studied were performed. All the statistical analyses were carried out with SPSS (v22.0).

3. RESULTS AND DISCUSSION

The experiment was performed to evaluate the behaviour of odorous volatile compounds and odour characteristics from raw knuckle beef steaks during display in high oxygen modified atmosphere packing (MAP) and pre-aged during 15 or 22 days, according to their lipid oxidation potential. The large number of samples used (at least 15 samples per oxidative group) makes the data reliable. The target aroma compounds were selected having regard to the scientific literature of raw beef (Casaburi et al., 2015; Insausti et al., 2002; Panseri et al., 2015; Pérez, Rojo, González & Lorenzo, 2008; Saraiva et al., 2015; Schindler et al., 2010). In Table 1 the list of variables, the main effects studied and their interactions are presented. Since many interactions were found, Table 2 shows the mean concentration of the volatile compounds within lipid oxidation group (low, medium and high) and display time for the meat pre-aged for 15 days, whereas for 22 days they are presented in Table 3. Similarly, the results of the sensory analysis and TBARS values are presented in Tables 4 and 5. Also, the R^2 of linear and exponential regressions are shown when significant ($P \leq 0.05$). In Table 6 the Pearson correlations between volatile compounds, odours and TBARS values are shown. Finally, based in all the results, a proposal of eight volatile compounds as shelf-life markers in raw meat is discussed.

3.1 Acids

Carboxylic saturated fatty acids from C4-C10 were quantified and all of them, except decanoic acid, were affected by display in high oxygen MAP (Table 1). Three of them, pentanoic, hexanoic and heptanoic acids, increased more clearly with display time either exponentially and/or linearly (Tables 2 and 3). Furthermore, those three compounds were generally affected by the oxidative group and were highly correlated with TBARS

analysis and rancid odour intensity (Tables 1-3 and 6). Hexanoic acid, together with butanoic and nonanoic acids, are usually raised in spoiled raw meat stored, imparting fatty, gammy, cheesy and dairy odours (Casaburi et al., 2015). However, in other studies using non-inoculated meat, but related to the effect of ageing/storage in raw/cooked meat, the results are controversial and sometimes acids have not even been reported (Gorraiz et al., 2002; Insausti et al., 2002; Ma et al., 2012; Meinert, Andersen, Bredie, Bjergegaard & Aaslyng, 2007; Resconi et al., 2012). In one study, hexanoic acid was found in raw beef stored in MAP but not in vacuum conditions, and no statistical effect of storage time (10 vs 21 days) was observed (Saraiva et al., 2015). On the other hand, in a study with pork meat, butanoic and nonanoic acids decreased in raw meat refrigerated in oxygen permeable films for 10 days and hexanoic acid was only found in the cooked and refrigerated meat but not in the raw meat (Estévez et al., 2003).

With respect to pre-ageing 15 vs 22 days in vacuum packs, in our study the seven acids quantified had shown a direct and/or interactions effects, being the effect of ageing of higher importance compared to display time for nonanoic and decanoic acids (lower *P*, higher *F* values, Table 1). These two volatiles were generally higher in the samples aged for 15 days (Tables 2-3). The behaviour of the rest of acids was more erratic. Nevertheless, Watanabe *et al.* (2015) found no effect of ageing until 30 days in volatile fatty acids from cooked *M. biceps femoris* in beef.

Differences between studies could be related with different methodology approaches, i.e. SPME extractions are more sensitive to acids compared to dynamic headspace extraction (DHE) (Rivas-Cañedo, Juez-Ojeda, Nuñez & Fernández-García, 2011), but also due to the complexity in the development of these compounds, which explains the high variability usually found. The acids in the meat can derive from several pathways,

including enzymatic and chemical reactions and bacterial action, being the source lipids, aminoacids or carbohydrates (Resconi et al., 2013; Whitfield, 1998). At the same time, acids could be further degraded to other aroma compounds (Casaburi et al., 2015).

3.2 Aldehydes and ketones

The effect of display found in aldehydes and ketones is not surprising since they are major lipid oxidation products (Table 1) and MAP with high concentration of oxygen promotes oxidation. However, a high variability was generally found, as reported previously (Estévez et al., 2003) and this may explain the low R^2 generally found when regression of carbonyl compounds throughout time of display were projected (Table 2). Saturated aldehydes have been previously described as affected by storage in raw beef packed with high oxygen, but alkenals and alkadienals had not been reported (Insausti et al., 2002). In our study three saturated aldehydes, seven alkenals and alkadienals and phenylacetaldehyde were quantified, and among them (*E*)-2-undecenal and 2-heptanone could be potential predictors of odour shelf-life as will be explained in section 3.6. Specifically, 2-heptanone has been already proposed as a candidate for the aroma differences between less or more oxidized cooked beef (Resconi et al., 2012) and had also shown an increment with storage time in raw meat (Estévez et al., 2003; Insausti et al., 2002), although the increase of 3-hydroxy-2-butanone was more pronounced in pork meat (Estévez et al., 2003).

Some aldehydes such as (*E,E*)-2,4-nonadienal, (*E*)-2-heptenal and (*E*)-2-nonenal were also influenced by ageing for 15 or 22 days (Table 1). Other authors have not found significant differences between day 2 to 30 in cooked beef due to a high variability of the data (Watanabe et al., 2015).

3.3 Esters

Ethyl- hexanoate and ethyl- octanoate increased with display time in MAP and oxidative group (Tables 1-3) and they are between the esters of greater odour impact in raw meat, giving fruity off-odours (Casaburi et al., 2015). In general, esters derived from the esterification of alcohols and carboxylic acids are found in meat after microbial enzyme activity (Casaburi et al., 2015).

3.4 Alcohols and 2-pentylfuran

Alkylfurans and alcohols are mainly formed when aminoacids and ribose interacted in the lipid oxidation pathway; 2-pentylfuran, 1-octen-3-ol and 1-hexanol, for example, arise mainly from linoleic acid (Elmore, Campo, Enser & Mottram, 2002; Van Ba, Amna & Hwang, 2013). Since ageing increases the concentration of free aminoacids and sugars (Koutsidis et al., 2008), while lipid oxidation in the meat intensifies through the days in display in high oxygen packs, the rise of 2-pentylfuran and 1-hexanol caused by display time, oxidative group and ageing seems reasonable (Tables 1-3). Although this furan is higher when meat is cooked at high temperatures, it has been also reported in raw meat, but not affected by storage time (Estévez et al., 2003; Meinert et al., 2007; Soncin, Chiesa, Cantoni & Biondi, 2007). In cooked meat, its concentration was reduced with the use of antioxidants and was related with off-flavour notes (Morán et al., 2013; Stetzer et al., 2008), but again no significant effect of storage has been observed (Estévez et al., 2003; Ma et al., 2012; Meinert et al., 2007; Stetzer et al., 2008; Watanabe et al., 2015).

The alcohol, 1-hexanol, followed a similar behaviour than pentylfuran, although its rate of formation was higher and in general more abundant (Tables 2-3). The alcohol can

also be increased in stored meat due to the growth of natural spoilage bacteria (La Storia et al., 2012), giving pungent, ethereal, fuel oil, fruity, alcoholic and sweet with a green top notes (Casaburi et al., 2015). In other studies, 1-hexanol and 1-octen-3-ol seem to increase with storage time, but the effect is not clear enough (Estévez et al., 2003; Ma et al., 2012; Watanabe et al., 2015). In the present study, the behaviour of 1-octen-3-ol was not clear according to the factors studied.

3.5 Odour characterisation

Several interactions between the factors studied were found (ageing, display, oxidative group), thus, odours were analysed within ageing time and within oxidative group or display days (Tables 1, 4 and 5). In general, the intensity of the typical odour of raw beef decreased with ageing and display time (Tables 1, 4 and 5). The antioxidant capacity of the meat also showed a significant effect, since samples from the high oxidative group presented lower intensities of raw meat odour compared to the low and medium oxidative groups, especially in meat aged for 15 days. On the other hand, rancid odour intensity increased with the days of display, ageing time and oxidative group (Tables 1, 4 and 5). In the high oxidative samples, pre-aged for 15 days, from the 7th day, the rancid odour is strong (the mean value exceed 5 out of 10, Table 4), as it happened in the medium oxidative samples aged for 22 days (Table 5). As expected, comparing all the descriptors used in this study, rancid odour was the best indicator of the odour changes through display time (generally regressions with higher R^2 , Tables 4-5) both. In another study assessing off-odour development in unaged raw meat (*longissimus dorsi*) displayed in MAP (60% O₂, 30% CO₂ and 10% N₂), the odour

shelf-life was around seven days for a rustic breed (Retinta) and 10-12 days in other five local Spanish beef breeds (Insausti, Beriain, Purroy, Alberti, Gorraiz & Alzueta, 2001).

In the less oxidized samples (display 0, aged 15 days, low and medium oxidative groups), after raw meat odour, sweet odour was the second most intense odour, but when odour is deteriorating other odours such as rancid, musty, sulphury and even cooked meat odour gained positions (Table 4). Dairy, sour-acid, old were other attributes found in oxidized samples by panellists. Metallic odour was in general a low intense odour, but also showed a diminution with the time on display, especially for the meat aged during 22 days (Table 5). In individual samples, other sweet/fruity intense odours were also described in a late stage probably due to esters caused by microbial spoilage (Casaburi et al., 2015). According to Schevey *et al.* (2013) fresh meat odour gives rise to sweetness, and finally rancidity takes over storage.

Regardless the ageing method, the intensity of off-odours (musty, sour, yeasty, putrid) in loins have been previously described to increase linearly from 14 to 49 days of ageing, but differences were substantially reduced when trimming discoloured, dehydrated or putrefied areas (Lepper-Blilie, Berg, Buchanan & Berg, 2016). When cooked, differences in aged flavour were perceived with the period of ageing but did not necessarily indicated a loss of quality (Lepper-Blilie et al., 2016). Values higher than 2 mg malonaldehyde/kg muscle were obtained in the medium oxidative samples aged for 22 days, at day 9 of display and at day 7 in the high oxidative groups previously aged for 15 or 22 days. Therefore, unpleasant cooked flavour would be expected, as this value has been proposed as the limiting threshold for beef eating acceptability (Campo et al., 2006).

3.6 Potential shelf-life markers

Eight volatile compounds (penta/hexa/heptanoic acids, 1-hexanol, (*E*)-2-undecenal, ethyl- octanoate, 2-heptanone and 2-pentylfuran) of raw meat in high oxygen MAP increased during display and were highly correlated with rancid odours and TBARS (Table 6). Therefore, they are proposed as shelf-life markers. Off-odour in the raw meat stored in high oxygen packs has been previously related with lipid oxidation (Insausti et al., 2001), but branched chain amino acids and pyruvate catabolism were more important than lipid oxidation in raw meat refrigerated in film (Estévez et al., 2003).

When regressions of the eight compounds throughout display time are compared, penta and hexanoic acids showed high R^2 , especially in the exponential regressions from all oxidative and ageing groups. Whereas 1-hexanol it is also an outstanding predictor of time in display for the 15 days aged meat, ethyl- octanoate and 2-pentylfuran are for the 22 days aged meat, considering all oxidative groups, which may represent better commercial situations. The five mentioned compounds were also abundant and with higher rates of increase with display time. In refrigerated cooked meat, mainly hexanal, but also other saturated aldehydes have been found to be the odorous compounds more abundant and with higher rate of increase (Drumm & Spanier, 1991) and therefore indicators of warmed over flavour. Finally, comparing the oxidative groups, no particular compound/s seems to predict changes over display only in one of the three groups studied, which means that the differences between the groups are mainly quantitative.

In Figures 1-5 of the supplementary material the increase of these five compounds through display are represented in box plots. It is possible to visualize the high variability of the compounds in the headspace of raw meat, which was higher in meat

exposed to MAP compared to meat vacuum packed. High variation in the generation of volatile compounds from meat have been also found previously (Estévez et al., 2003; Watanabe et al., 2015), and reasons are several, due to the complexity in the formation of the compounds and the interactions that could take place.

ASSOCIATED CONTENT

Supplementary material

Box plots for five compounds proposed as shelf-life markers of knuckle steaks.

CONCLUSIONS

In the raw knuckle, around five days of odour shelf-life is expected in high oxidative samples displayed in high oxygen atmosphere packaging and previously aged during 22 days. On the other hand, low oxidative meat aged for 15 days could be displayed during nine days, without severe alteration of the odour. Those results indicate the high importance of controlling the balance between anti- and pro-oxidants in the meat.

The odour shelf-life of the meat could be predicted by the analysis of volatile compounds thorough SPME-GCMS analysis. Results have shown that the most sensitive and precise predictors of display in all oxidative groups were 1-hexanol for meat pre-aged during 15 days, ethyl- octanoate and 2-pentylfuran for meat pre-aged 22 days, and penta/ and hexanoic acids for both ageing times.

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Table 1. F value and significance of ageing (15 or 22 d), display time (0, 5, 7 or 9 d) and oxidative group¹ (low, medium or high) effects and their interactions in volatile compounds analysed by SPME-GCMS, odour intensity and TBARS value from raw knuckle steaks

	LRI	Quan ion	Ageing	Display	Oxidative Group	A x D	A x OG	D x OG	A x D x OG
Acids									
<i>Butanoic acid</i>	1632	60	ns	5.84 ***	ns	5.29 ***	3.47 *	3.75 ***	ns
<i>Pentanoic acid</i>	1744	60	34.3 ***	124 ***	45.2 ***	5.27 ***	ns	4.15 ***	2.93 **
<i>Hexanoic acid</i>	1851	60	4.34 *	155 ***	35.0 ***	ns	4.39 *	7.02 ***	3.42 **
<i>Heptanoic acid</i>	1975	60	6.46 *	85.7 ***	7.76 ***	5.84 ***	3.69 *	2.17 *	3.16 **
<i>Octanoic acid</i>	2051	60	ns	11.2 ***	ns	11.4 ***	ns	ns	2.51 *
<i>Nonanoic acid</i>	2116	60	17.0 ***	4.87 **	ns	4.62 **	ns	ns	3.16 **
<i>Decanoic acid</i>	2200	60	28.0 ***	ns	ns	ns	3.88 *	ns	ns
Alcohols									
<i>1-Hexanol</i>	1359	69	10.9 ***	84.1 ***	81.8 ***	ns	3.45 *	8.35 ***	3.08 **
<i>1-Octen-3-ol</i>	1449	57	ns	15.1 ***	5.13 **	15.0 ***	ns	ns	3.72 ***
Aldehydes									
<i>Hexanal</i>	1052	57	ns	ns	ns	4.74 **	ns	ns	ns
<i>Heptanal</i>	1188	71	ns	6.85 ***	6.31 **	ns	9.26 ***	3.03 **	ns

<i>Nonanal</i>	1402	98+120	ns	2.98 *	ns	ns	ns	ns	ns
<i>(E)-2-Heptenal</i>	1344	97	9.98 **	10.3 ***	7.71 ***	ns	ns	ns	2.44 *
<i>(Z)-2-Octenal</i>	1420	83	4.37 *	21.8 ***	13.9 ***	ns	ns	ns	2.14 *
<i>(E)-2-Nonenal</i>	1562	93	9.64 **	10.6 ***	7.06 ***	2.80 *	ns	ns	2.94 **
<i>(E,E)-2,4-Nonadienal</i>	1741	81	7.11 **	3.50 *	4.02 *	ns	ns	ns	ns
<i>(E)-2-Undecenal</i>	1781	83	ns	49.5 ***	45.8 ***	ns	4.10 *	3.91 ***	ns
<i>(E,Z)-2,4-Decadienal</i>	1799	81	ns	15.8 ***	ns	ns	ns	ns	2.58 *
<i>(E,E)-2,4-Decadienal</i>	1854	81	5.37 *	21.1 ***	10.4 ***	ns	ns	ns	2.61 *
<i>Phenylacetaldehyde</i>	1681	91+120	3.88 *	3.36 *	ns	ns	ns	ns	ns
Esters									
<i>Ethyl- hexanoate</i>	1236	88	ns	44.5 ***	15.0 ***	3.44 *	9.68 ***	2.72 *	6.37 ***
<i>Ethyl- octanoate</i>	1438	88	ns	115 ***	36.5 ***	ns	23.6 ***	2.17 *	2.62 *
Ketones									
<i>2-Butanone</i>	668	72	ns	3.51 *	ns	ns	ns	ns	ns
<i>2-Heptanone</i>	1192	99	3.93 *	58.4 ***	10.4 ***	ns	ns	ns	ns
<i>3-Octanone</i>	1265	99	ns	20.3 ***	3.30 *	ns	6.76 ***	2.96 **	ns

<i>2-Octanone</i>	1293	58	ns	35.8 ***	5.55 **	ns	7.63 **	ns	ns
<i>2-Nonanone</i>	1400	142	ns	16.3 ***	ns	3.49 *	3.28 *	ns	2.88 **
<i>1-Octen-3-one</i>	1311	70	ns	20.7 ***	7.85 ***	4.59 **	ns	ns	2.52 *
<i>3-Octen-2-one</i>	1427	111	ns	8.09 ***	3.06 *	2.96 *	ns	ns	ns
<i>3-Nonen-2-one</i>	1535	125	ns	4.77 **	ns	ns	ns	ns	ns
<i>2,3-Octanedione</i>	1332	99	ns	5.33 ***	ns	5.62 ***	ns	ns	ns
Others									
<i>2-Pentylfuran</i>	1235	138	11.0 **	38.7 ***	33.5 ***	ns	ns	4.32 ***	ns
Odour intensity									
<i>Raw meat odour</i>			84.7 ***	57.9 ***	22.9 ***	3.24 *	5.02 **	ns	3.19 **
<i>Rancid odour</i>			54.4 ***	184 ***	70.4 ***	ns	ns	5.29 ***	ns
<i>Metallic odour</i>			ns	17.8 ***	4.60 *	3.18 *	ns	ns	2.96 **
<i>Sweet odour</i>			23.1 ***	14.4 ***	ns	2.87 *	ns	ns	2.54 *
<i>Musty odour</i>			ns	39.2 ***	14.0 ***	12.6 ***	13.4 ***	ns	3.54 **
<i>Cooked meat odour</i>			4.56 *	51.1 ***	ns	6.53 ***	3.77 *	3.31 **	2.13 *
<i>Sulphury odour</i>			11.6 ***	18.8 ***	7.37 ***	ns	3.66 *	ns	2.62 *

TBARS value	20.5 ***	210 ***	146 ***	4.46 **	ns	20.5 ***	2.23 *
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¹ Low, medium and high oxidative groups, when reached < 1, 1-2 or > 2 mg MDA/kg muscle (TBARS values), respectively, on day 9 of display in samples aged 15 days. LRI: linear retention index calculated for DB-WAXETR capillary column. A: aging. D: display. OG: oxidative group. Quan ion: m/z used for quantification. ns: $P > 0.05$, *: $P \leq 0.05$, **: $P \leq 0.01$, *** $P \leq 0.001$.

Table 2. Mean and standard deviation for volatile compounds ($\mu\text{g/g}$) of knuckle steaks aged for 15 days under vacuum conditions and displayed in modified atmosphere packaging (0, 5, 7 or 9 days), according oxidative group¹.

Compound	Oxidative group	Display (days)				<i>P</i> display	Regressions ³	
		0	5	7	9		Linear	Exponential
<i>Butanoic acid</i>	Low	17.9±14.8	18.6±17.6	26.5±10.5 ^X	25.5±12.1	0.193		0.093
	Medium	9.76±7.68 ^b	10.9±12.9 ^b	22.9±9.3 ^{aXY}	24.1±9.2 ^a	<0.001	0.240	0.216
	High	18.4±10.1	16.7±19.8	17.7±5.0 ^Y	20.9±6.7	0.772		
<i>P</i> oxidative group		0.067	0.437	0.018	0.363			
<i>Pentanoic acid</i>	Low	1.31±0.95 ^c	2.28±1.37 ^{bc}	3.42±1.49 ^{abY}	4.44±1.69 ^{aY}	<0.001	0.398	0.462
	Medium	0.825±0.514	2.22±1.93 ^b	5.54±1.39 ^{aX}	6.53±1.84 ^{aY}	<0.001	0.628	0.633
	High	^b 1.57±0.97 ^c	4.82±5.00 ^{bc}	6.64±2.04 ^{bX}	12.2±4.8 ^{aX}	<0.001	0.467	0.646
<i>P</i> oxidative group		0.052	0.042	<0.001	<0.001			
<i>Hexanoic acid</i>	Low	14.0±10.3 ^c	45.3±24.3 ^{bY}	71.4±40.1 ^{abY}	89.6±35.3 ^{aZ}	<0.001	0.481	0.619
	Medium	9.42±6.28 ^c	56.3±39.9 ^{bY}	133±24 ^{aX}	144±24 ^{aY}	<0.001	0.753	0.802
	High	19.7±17.2 ^b	125±121 ^{aX}	142±35 ^{aX}	177±36 ^{aX}	<0.001	0.447	0.750
<i>P</i> oxidative group		0.073	0.009	<0.001	<0.001			
<i>Heptanoic acid</i>	Low	1.33±0.53 ^b	1.98±1.14 ^{bXY}	2.04±1.02 ^{bY}	3.29±1.49 ^{aY}	<0.001	0.241	0.225
	Medium	1.21±0.91 ^b	1.73±0.88 ^{bY}	3.29±1.14 ^{aX}	3.89±2.00 ^{aY}	<0.001	0.366	
	High	1.05±0.81 ^c	3.26±2.27 ^{bX}	3.29±1.17 ^{bX}	5.76±2.02 ^{aX}	<0.001	0.455	0.642
<i>P</i> oxidative group		0.575	0.018	0.003	0.001			
<i>Octanoic acid</i>	Low	4.87±2.28 ^{ab}	4.63±2.57 ^{ab}	3.93±1.97 ^{bY}	6.77±3.26 ^a	0.019		
	Medium	4.43±2.83	3.68±1.82	6.19±2.80 ^X	6.59±4.93	0.055		0.081
	High	3.49±2.15 ^b	5.04±2.65 ^{ab}	4.18±2.01 ^{abY}	5.69±2.56 ^a	0.044	0.078	0.135
<i>P</i> oxidative group		0.255	0.271	0.016	0.662			
<i>Nonanoic acid</i>	Low	13.9±6.2	16.2±15.2	10.1±9.1	17.6±11.1	0.242		
	Medium	16.4±11.5	11.3±6.2	15.9±10.7	17.2±21.0	0.635		
	High	8.83±7.92	16.3±10.5	11.1±8.8	8.71±7.65	0.049		

	<i>P</i> oxidative group		0.053	0.390	0.203	0.137			
<i>Decanoic acid</i>	Low		0.733±0.257	0.880±0.498	0.661±0.276	0.758±0.234	0.318		
	Medium		0.784±0.521	0.643±0.327	0.736±0.179	0.710±0.443	0.794		
	High		0.605±0.250	0.777±0.540	0.593±0.187	0.622±0.231	0.340		
			0.350	0.376	0.194	0.453			
<i>1-Hexanol</i>	Low		2.60±3.90 ^{by}	24.36±23.00	99.5±102.3 ^{az}	149±114 ^{ay}	<0.001	0.314	0.725
	Medium		6.39±12.84 ^{by}	^{by} 85.9±87.6	332±121 ^{ay}	440±258 ^{ay}	<0.001	0.510	0.775
	High		63.8±107.1 ^{cx}	^{bxy} 439±699 ^{bcx}	841±365 ^{bx}	1328±552 ^{ax}	<0.001	0.457	0.674
	<i>P</i> oxidative group		0.014	0.014 0.169	<0.001	<0.001			
<i>1-Octen-3-ol</i>	Low		0.120±0.134 ^{by}	±0.153 ^b	0.672±0.391 ^a	0.329±0.410 ^{by}	<0.001	0.129	
	Medium		0.346±0.443 ^{bxy}	0.331±0.422	0.944±0.640 ^a	0.508±0.338 ^{abxy}	0.002		
	High		0.455±0.324 ^x	^b 0.424±0.348	0.672±0.481	0.815±0.605 ^x	0.045	0.077	
	<i>P</i> oxidative group		0.015	0.088	0.244	0.017			
<i>Hexanal</i>	Low		2.69±4.18	9.41±15.79 ^{xy}	3.85±6.27	33.8±89.9	0.196		
	Medium		36.2±133.6	5.32±7.30 ^y	22.5±29.8	30.5±33.4	0.654		0.261
	High		25.4±61.8	20.2±17.2 ^x	28.7±45.0	49.9±92.5	0.506		
	<i>P</i> oxidative group		0.521	0.014	0.079	0.751			
<i>Heptanal</i>	Low		5.29±4.81	9.44±13.63 ^y	8.92±9.89 ^y	11.6±13.2	0.439		
	Medium		5.21±4.37 ^b	29.0±44.5 ^{abxy}	50.7±36.7 ^{ax}	28.7±35.5 ^{ab}	0.007	0.110	
	High		19.8±31.9 ^b	60.3±52.5 ^{ax}	33.2±33.2 ^{abxy}	30.8±33.7 ^{ab}	0.024		
	<i>P</i> oxidative group		0.055	0.003	0.001	0.135			
<i>Nonanal</i>	Low		1.85±3.52	2.24±1.56	0.565±0.573	5.31±8.97	0.054		
	Medium		3.57±12.33	2.55±3.76	7.93±21.98	5.61±8.72	0.705		
	High		5.07±12.42	4.80±4.56	19.4±48.4	4.36±5.50	0.242		
	<i>P</i> oxidative group		0.670 0.012	0.089	0.231	0.894			
<i>(E)-2-Heptenal</i>	Low		±0.025 ^b	0.339±0.514 ^a	0.054±0.081 ^{by}	0.188±0.182 ^{aby}	0.006		
	Medium		0.077±0.250	0.220±0.244	0.273±0.318 ^x	0.240±0.160 ^y	0.155	0.075	
	High		0.138±0.291 ^b	0.795±1.029 ^a	0.409±0.276 ^{abx}	0.831±0.427 ^{ax}	0.002	0.115	
	<i>P</i> oxidative group		0.281	0.054	0.001	<0.001			

<i>(Z)</i> -2-Octenal	Low	0.014±0.032	0.166±0.293	0.081±0.119 ^Y	0.099±0.085 ^Y	0.087	
	Medium	0.032±0.088 ^b	0.137±0.075 ^{ab}	0.206±0.127 ^{aX}	0.265±0.207 ^{aX}	<0.001	0.305
	High	0.061±0.096 ^b	0.437±0.521 ^a	0.223±0.121 ^{abX}	0.411±0.229 ^{aX}	0.001	0.120
<i>P</i> oxidative group		0.233	0.037	0.003	<0.001		
<i>(E)</i> -2-Nonenal	Low	0.040±0.057 ^b	0.208±0.254 ^{aXY}	0.076±0.127 ^{ab}	0.114±0.110 ^{abY}	0.021	
	Medium	0.098±0.281	0.105±0.101 ^Y	0.198±0.247	0.120±0.120 ^Y	0.514	
	High	0.095±0.169 ^b	0.358±0.389 ^{aX}	0.168±0.140 ^{ab}	0.376±0.274 ^{aX}	0.004	0.088
<i>P</i> oxidative group		0.629	0.045	0.145	<0.001		
<i>(E,E)</i> -2,4-Nonadienal	Low	0.011±0.046	0.339±0.647 ^{XY}	0.055±0.064 ^Y	0.477±1.01 ^X	0.094	
	Medium	0.267±1.034	0.143±0.168 ^Y	0.344±0.410 ^Y	0.439±0.335 ^X	0.567	
	High	0.066±0.196 ^b	0.864±0.968 ^{bX}	0.709±0.496 ^{bX}	1.95±1.62 ^{aY}	<0.001	0.263
<i>P</i> oxidative group		0.454	0.014	<0.001	<0.001		
<i>(E)</i> -2-Undecenal	Low	0.105±0.161 ^{bY}	1.44±1.21 ^{aY}	1.07±0.66 ^{aZ}	1.20±0.50 ^{aZ}	<0.001	0.217
	Medium	0.286±0.573 ^{bXY}	1.79±1.08 ^{aXY}	2.21±0.59 ^{aY}	2.43±0.82 ^{aY}	<0.001	0.528
	High	0.771±1.019 ^{bX}	4.70±5.49 ^{aX}	3.80±1.36 ^{aX}	4.31±1.31 ^{aX}	0.001	0.163
<i>P</i> oxidative group		0.023	0.015	<0.001	<0.001		
<i>(E,Z)</i> -2,4-Decadienal	Low	0.029±0.070	2.06±5.02	0.089±0.100 ^Y	0.535±0.786	0.095	
	Medium	0.330±0.954	0.555±0.726	0.550±0.641 ^X	0.842±1.056	0.451	
	High	0.201±0.401 ^b	2.05±1.93 ^a	0.508±0.252 ^{bX}	0.890±1.082 ^b	<0.001	
<i>P</i> oxidative group		0.364	0.318	0.003	0.544		
<i>(E,E)</i> -2,4-Decadienal	Low	0.049±0.156	6.21±14.80	0.992±1.189 ^Z	2.85±2.42 ^Y	0.112	
	Medium	0.879±2.921 ^c	2.40±2.65 ^{bc}	4.40±2.65 ^{abY}	5.74±3.51 ^{aXY}	<0.001	0.277
	High	1.15±2.18 ^b	10.4±12.4 ^a	7.29±3.26 ^{abX}	8.77±4.34 ^{aX}	0.001	0.135
<i>P</i> oxidative group		0.301	0.149	<0.001	<0.001		
Phenylacetaldehyde	Low	0.740±1.723	0.299±0.833	0.206±0.578	0.170±0.462	0.375	
	Medium	1.29±2.80	0.309±0.665	0.067±0.177	0.699±1.865	0.236	
	High	1.50±2.39 ^a	0.233±0.676 ^b	n.d. ^b	0.065±0.268 ^b	0.002	0.178
<i>P</i> oxidative group		0.637	0.949	0.237	0.229		
Ethyl- hexanoate	Low	0.012±0.019 ^{bY}	0.058±0.081 ^{bY}	0.562±0.693 ^{aY}	0.268±0.345 ^{abY}	0.001	0.108

	Medium	0.047±0.071 ^{bXY}	0.364±0.745 ^{bXY}	1.25±1.12 ^{aXY}	1.20±0.99 ^{aY}	<0.001	0.244	
	High	0.060±0.048 ^{cX}	0.734±0.702 ^{bcX}	1.56±1.32 ^{abX}	2.69±2.05 ^{aX}	<0.001	0.343	
	<i>P</i> oxidative group	0.025	0.008	0.033	<0.001			
<i>Ethyl- octanoate</i>	Low	1.38±1.95 ^{bY}	15.9±12.3 ^{aY}	18.0±14.5 ^{aY}	22.2±15.0 ^{aZ}	<0.001	0.300	
	Medium	3.91±8.79 ^{cXY}	27.2±12.6 ^{bY}	44.4±12.7 ^{aX}	45.6±11.3 ^{aY}	<0.001	0.673	
	High	13.8±20.7 ^{bX}	49.3±14.9 ^{aX}	55.3±17.1 ^{aX}	59.1±18.1 ^{aX}	<0.001	0.489	0.550
	<i>P</i> oxidative group	0.024	<0.001	<0.001	<0.001			
<i>2-Butanone</i>	Low	0.070±0.078	0.089±0.068	0.062±0.073	0.073±0.084	0.780		
	Medium	0.080±0.070	0.048±0.038	0.049±0.059	0.068±0.117	0.629		
	High	0.101±0.137	0.074±0.098	0.061±0.040	0.133±0.189	0.376		
	<i>P</i> oxidative group	0.664	0.305	0.813	0.340			
<i>2-Heptanone</i>	Low	0.020±0.008 ^{bY}	0.265±0.150 ^a	0.212±0.164 ^{aY}	0.356±0.257 ^a	<0.001	0.312	
	Medium	0.051±0.103 ^{bXY}	0.351±0.219 ^a	0.344±0.185 ^{aXY}	0.520±0.299 ^a	<0.001	0.380	
	High	0.094±0.100 ^{bX}	0.461±0.471 ^a	0.468±0.195 ^{aX}	0.717±0.623 ^a	<0.001	0.230	0.520
	<i>P</i> oxidative group	0.047	0.218	0.001	0.067			
<i>3-Octanone</i>	Low	0.038±0.026 ^Y	0.565±0.643 ^Y	0.251±0.285 ^Y	0.591±1.129	0.065	0.064	0.264
	Medium	0.104±0.201 ^{bXY}	1.10±0.94 ^{aXY}	0.739±0.399 ^{aX}	0.917±0.632 ^a	<0.001	0.176	0.537
	High	0.399±0.657 ^{bX}	2.00±1.70 ^{aX}	0.889±0.437 ^{bX}	0.877±0.539 ^b	<0.001		0.265
	<i>P</i> oxidative group	0.033	0.005	<0.001	0.470			
<i>2-Octanone</i>	Low	0.050±0.069 ^{bY}	0.643±0.633 ^a	0.465±0.352 ^{abY}	0.519±0.549 ^{aY}	0.003	0.125	
	Medium	0.041±0.119 ^{bY}	0.809±0.557 ^a	0.712±0.393 ^{aXY}	1.03±0.63 ^{aXY}	<0.001	0.359	
	High	0.125±0.168 ^{bX}	1.12±0.96 ^a	1.02±0.59 ^{aX}	1.51±1.53 ^{aX}	0.001	0.212	
	<i>P</i> oxidative group	0.127	0.187	0.006	0.030			
<i>2-Nonanone</i>	Low	0.020±0.010	0.028±0.016	0.023±0.010	0.025±0.015	0.412		
	Medium	0.025±0.012	0.030±0.015	0.028±0.014	0.039±0.023	0.128		
	High	0.022±0.007	0.042±0.039	0.028±0.017	0.044±0.039	0.078		0.070
	<i>P</i> oxidative group	0.393	0.256	0.458	0.130			
<i>1-Octen-3-one</i>	Low	0.018±0.027 ^b	0.145±0.172 ^{aY}	0.022±0.020 ^{bY}	0.081±0.091 ^{ab}	0.001		
	Medium	0.041±0.095 ^b	0.226±0.182 ^{aXY}	0.076±0.042 ^{bX}	0.141±0.124 ^{ab}	0.001		

	High	0.110±0.223 ^b	0.392±0.358 ^{aX}	0.092±0.061 ^{bX}	0.112±0.093 ^b	<0.001	
	<i>P</i> oxidative group	0.172	0.026	<0.001	0.276		
<i>3-Octen-2-one</i>	Low	0.001±0.003 ^b	0.056±0.060 ^{abY}	0.035±0.029 ^{bY}	0.136±0.196 ^a	0.004	0.136
	Medium	0.010±0.037 ^b	0.059±0.037 ^{bY}	0.118±0.083 ^{bxY}	0.253±0.216 ^a	<0.001	0.309
	High	0.002±0.005 ^b	0.156±0.146 ^{bX}	0.187±0.169 ^{abX}	0.418±0.492 ^a	<0.001	0.210
	<i>P</i> oxidative group	0.466	0.006	0.001	0.064		
<i>3-Nonen-2-one</i>	Low	n.d.	0.003±0.007	0.001±0.002 ^Y	0.005±0.007 ^Y	0.031	0.072
	Medium	n.d. ^b	0.002±0.003 ^b	0.014±0.018 ^{abXY}	0.022±0.026 ^{aXY}	0.001	0.193
	High	0.001±0.002	0.033±0.052	0.018±0.025 ^X	0.033±0.041 ^X	0.031	0.082
	<i>P</i> oxidative group	0.156	0.010	0.031	0.027		
<i>2,3-Octanedione</i>	Low	0.714±1.551	4.43±7.58	0.936±2.048 ^Y	20.9±63.4	0.238	
	Medium	3.94±14.37	4.41±5.29	5.47±5.98 ^{XY}	10.1±15.9	0.438	0.392
	High	1.10±2.50	8.95±12.39	10.4±16.9 ^X	11.3±23.2	0.222	0.063
	<i>P</i> oxidative group	0.496	0.265	0.049	0.709		
<i>2-Pentylfuran</i>	Low	0.063±0.080 ^b	1.71±1.58 ^{abY}	1.87±2.73 ^{abZ}	2.94±3.34 ^{aY}	0.008	0.170
	Medium	0.709±2.329 ^c	4.13±3.82 ^{bcXY}	5.69±2.71 ^{abY}	8.43±5.93 ^{aY}	<0.001	0.339
	High	0.720±1.177 ^c	9.20±11.92 ^{bX}	11.6±5.0 ^{abX}	18.6±12.8 ^{aX}	<0.001	0.330
	<i>P</i> oxidative group	0.362	0.019	<0.001	<0.001		

¹ Oxidative groups, according TBARS values on day 9 of display from samples aged 15 days: low (< 1 mg MDA/kg, *n*=16), medium (1-2 mg MDA/kg, *n*=15), and high (> 2 mg MDA/kg, *n*=17). ²: Root mean squared error. ³: the R² of linear and exponential regressions from each compound through time of display are shown when significant (*P*≤0.05). n.d.: not detected. a-c: different superscripts within a row denote statistical differences, according the Tukey test (*P*≤0.05) between display time within oxidative group for a compound. X-Z: different superscripts in the same column denote statistical differences (*P*≤0.05) between oxidative groups within day on display, for a compound.

Table 3. Mean and standard deviation for volatile compounds ($\mu\text{g/g}$) of knuckle steaks aged for 22 days under vacuum conditions and displayed in modified atmosphere packaging (0, 5, 7 or 9 days), according oxidative group¹.

Compound	Oxidative group	Display (days)				<i>P</i> display	Regressions ³	
		0	5	7	9		Linear	Exponential
<i>Butanoic acid</i>	Low	26.6±21.0 ^a	9.59±7.58 ^{bY}	20.1±8.9 ^{ab}	21.6±8.5 ^{ab}	0.011		
	Medium	19.7±10.4 ^{ab}	14.0±9.2 ^{bY}	21.9±9.9 ^{ab}	25.3±9.8 ^a	0.031		
	High	30.3±13.2 ^a	25.2±15.4 ^{abX}	17.8±5.6 ^b	21.6±6.0 ^{ab}	0.014	0.121	0.080
<i>P</i> oxidative group		0.165	0.001	0.385	0.478			
<i>Pentanoic acid</i>	Low	1.26±0.70 ^{cY}	1.68±0.82 ^{cY}	5.27±3.28 ^{bY}	9.28±4.13 ^a	<0.001	0.400	0.483
	Medium	1.39±0.72 ^{cY}	4.33±3.93 ^{bcXY}	6.34±2.20 ^{bXY}	12.3±4.0 ^a	<0.001	0.532	0.681
	High	2.27±0.89 ^{cX}	6.92±3.45 ^{bX}	8.58±4.27 ^{bX}	12.9±4.5 ^a	<0.001	0.514	0.651
<i>P</i> oxidative group		0.001	<0.001	0.026	0.239			
<i>Hexanoic acid</i>	Low	10.0±5.6 ^{cY}	40.0±18.1 ^{cZ}	97.6±57.2 ^b	157±63 ^a	<0.001	0.530	0.777
	Medium	15.3±6.6 ^{cXY}	93.9±61.0 ^{bY}	124±38 ^b	173±36 ^a	<0.001	0.670	0.829
	High	22.8±15.2 ^{bX}	143±59 ^{aX}	119±17 ^a	144±25 ^a	<0.001	0.551	0.679
<i>P</i> oxidative group		0.004	<0.001	0.163	0.194			
<i>Heptanoic acid</i>	Low	0.504±0.254 ^c	2.80±1.48 ^b	4.51±1.62 ^a	3.73±1.93 ^{ab}	<0.001	0.542	
	Medium	0.881±0.709 ^c	3.24±1.87 ^b	3.50±0.98 ^{ab}	4.98±2.05 ^a	<0.001	0.491	
	High	0.812±0.464 ^b	3.68±1.62 ^a	3.72±1.82 ^a	4.44±1.88 ^a	<0.001	0.438	
<i>P</i> oxidative group		0.093	0.339	0.176	0.446			
<i>Octanoic acid</i>	Low	1.79±1.10 ^b	6.03±2.45 ^a	7.15±2.65 ^{aX}	3.54±0.98 ^b	<0.001	0.310	0.440
	Medium	3.10±1.80 ^b	6.18±3.40 ^a	5.75±2.47 ^{aXY}	4.93±2.13 ^{ab}	0.009	0.098	0.172
	High	3.12±2.09 ^b	6.31±3.56 ^a	4.49±3.19 ^{abY}	4.11±1.06 ^{ab}	0.012		0.092
<i>P</i> oxidative group		0.054	0.970	0.036	0.188			
<i>Nonanoic acid</i>	Low	4.22±4.12 ^b	14.3±4.7 ^a	16.0±7.7 ^{aX}	2.05±0.85 ^b	<0.001	0.125	0.144
	Medium	7.66±6.68 ^b	15.6±8.5 ^a	12.2±7.5 ^{abXY}	5.99±6.38 ^b	0.005		
	High	8.57±10.66	13.0±7.8	6.98±7.66 ^Y	6.87±3.71	0.110		
<i>P</i> oxidative group		0.252	0.620	0.006	0.126			

<i>Decanoic acid</i>	Low	0.402±0.217	0.397±0.104 ^Y	0.522±0.148 ^X	0.510±0.221	0.121	
	Medium	0.611±0.404	0.570±0.287 ^{XY}	0.609±0.171 ^X	0.580±0.232	0.974	
	High	0.568±0.226 ^{ab}	0.730±0.330 ^{ax}	0.379±0.118 ^{by}	0.565±0.184 ^{ab}	0.001	
<i>P</i> oxidative group		0.114	0.004	<0.001	0.800		
<i>1-Hexanol</i>	Low	9.18±19.91 ^{by}	43.9±103.5 ^{by}	297±440 ^{by}	628±374 ^a	<0.001	0.248
	Medium	15.6±22.8 ^{cY}	167±179 ^{bcY}	451±328 ^{by}	915±526 ^a	<0.001	0.436
	High	114±129 ^{bx}	461±277 ^{bx}	975±447 ^{ax}	1082±593 ^a	<0.001	0.480
<i>P</i> oxidative group		<0.001	<0.001	<0.001	0.241		
<i>1-Octen-3-ol</i>	Low	0.534±0.470 ^{ab}	0.146±0.441 ^b	0.210±0.268 ^b	0.983±1.011 ^a	0.004	
	Medium	0.763±0.696 ^a	0.095±0.164 ^b	0.565±0.690 ^{ab}	1.07±0.70 ^a	0.001	
	High	1.01±0.61 ^a	0.255±0.254 ^b	0.445±0.309 ^b	0.387±0.380 ^b	<0.001	0.234
<i>P</i> oxidative group		0.086	0.325	0.108	0.037		
<i>Hexanal</i>	Low	0.477±0.722 ^b	20.6±40.3 ^{ab}	49.3±65.1 ^a	20.4±24.7 ^{ab}	0.022	0.113
	Medium	1.12±1.27 ^b	39.8±47.9 ^a	37.4±56.7 ^{ab}	13.3±9.0 ^{ab}	0.020	
	High	14.1±31.6	39.7±50.4	65.8±134.3	14.8±7.5	0.179	0.155
<i>P</i> oxidative group		0.075	0.429	0.696	0.545		
<i>Heptanal</i>	Low	3.34±2.71 ^b	14.8±16.8 ^b	19.2±22.8 ^{ab}	37.2±32.0 ^a	0.003	0.214
	Medium	16.1±39.0	14.0±11.5	25.6±25.4	39.7±52.4	0.220	
	High	10.2±7.7	20.3±29.5	16.6±31.3	8.44±5.74	0.432	
<i>P</i> oxidative group		0.289	0.653	0.636	0.081		
<i>Nonanal</i>	Low	0.215±0.282 ^Y	3.52±4.71	3.42±5.64	1.28±0.81	0.075	
	Medium	0.425±0.392 ^{XY}	7.43±10.33	11.1±18.0	2.44±1.91	0.040	
	High	2.60±4.42 ^X	4.25±8.03	4.53±8.13	1.21±0.96	0.457	
<i>P</i> oxidative group		0.023	0.371	0.162	0.079		
<i>(E)-2-Heptenal</i>	Low	0.029±0.040	0.189±0.300	0.486±0.932	0.422±0.346	0.112	0.097
	Medium	0.014±0.028	1.37±2.56	0.515±0.493	0.967±0.846	0.062	
	High	0.040±0.042 ^b	0.724±0.702 ^{ab}	1.17±1.65 ^a	0.783±0.516 ^{ab}	0.011	0.126
<i>P</i> oxidative group		0.159	0.114	0.179	0.257		
<i>(Z)-2-Octenal</i>	Low	0.001±0.006 ^{by}	0.102±0.151 ^{abY}	0.202±0.219 ^a	0.225±0.146 ^a	0.002	0.253

	Medium	0.036±0.069 ^{bXY}	0.437±0.512 ^{aX}	0.228±0.194 ^{ab}	0.455±0.502 ^a	0.010	0.116
	High	0.061±0.085 ^{bX}	0.449±0.239 ^{aX}	0.325±0.380 ^a	0.265±0.235 ^{ab}	0.001	0.101
	<i>P</i> oxidative group	0.034	0.008	0.427	0.314		
<i>(E)</i> -2-Nonenal	Low	0.044±0.045	0.124±0.172	0.332±0.586	0.122±0.070	0.114	
	Medium	0.034±0.037 ^b	0.558±0.730 ^a	0.284±0.246 ^{ab}	0.412±0.388 ^{ab}	0.014	0.081
	High	0.068±0.116 ^b	0.360±0.371 ^{ab}	0.495±0.545 ^a	0.334±0.268 ^{ab}	0.011	0.117
	<i>P</i> oxidative group	0.428	0.055	0.440	0.173		
<i>(E,E)</i> -2,4-Nonadienal	Low	n.d.	0.291±0.440	1.08±2.15	0.534±0.440	0.093	0.077
	Medium	n.d.	3.55±7.65	0.919±1.500	1.35±1.31	0.115	
	High	0.130±0.304	1.52±1.73	4.31±9.69	1.45±0.87	0.121	
	<i>P</i> oxidative group	0.068	0.140	0.205	0.186		
<i>(E)</i> -2-Undecenal	Low	0.114±0.127 ^{bY}	1.18±0.80 ^{abY}	1.89±1.54 ^{aY}	2.21±0.54 ^a	<0.001	0.418
	Medium	0.258±0.236 ^{bXY}	2.83±2.43 ^{aX}	2.54±1.10 ^{aXY}	3.33±1.30 ^a	<0.001	0.354
	High	0.444±0.487 ^{bX}	4.06±1.57 ^{aX}	3.36±1.15 ^{aX}	3.64±1.39 ^a	<0.001	0.441
	<i>P</i> oxidative group	0.021	<0.001	0.008	0.080		
<i>(E,Z)</i> -2,4-Decadienal	Low	0.016±0.045 ^Y	0.989±1.47	1.07±1.30	0.232±0.150	0.023	0.077
	Medium	0.023±0.038 ^{bXY}	3.03±3.17 ^a	0.843±0.930 ^b	0.560±0.475 ^b	<0.001	
	High	0.163±0.277 ^X	2.23±3.12	1.83±3.15	0.524±0.260	0.033	
	<i>P</i> oxidative group	0.026	0.130	0.385	0.163		
<i>(E,E)</i> -2,4-Decadienal	Low	n.d. ^b	3.28±4.97 ^{abY}	7.26±8.94 ^a	4.21±1.95 ^{ab}	0.008	0.164
	Medium	0.068±0.186 ^b	13.3±15.2 ^{aX}	6.49±5.14 ^{ab}	7.04±3.67 ^{ab}	0.001	0.081
	High	0.428±0.822 ^b	12.1±11.0 ^{aXY}	10.9±11.6 ^a	6.63±2.20 ^{ab}	0.001	0.116
	<i>P</i> oxidative group	0.039	0.035	0.341	0.138		
Phenylacetaldehyde	Low	2.16±4.86	0.647±0.966	0.814±2.006	3.27±8.01	0.430	
	Medium	8.74±18.47	0.560±0.828	0.976±2.113	1.36±4.73	0.083	0.090
	High	17.8±55.0	1.04±3.45	0.287±1.182	n.d.	0.212	0.062
	<i>P</i> oxidative group	0.431	0.806	0.524	0.345		
Ethyl- hexanoate	Low	0.084±0.114 ^{bY}	0.246±0.854 ^b	0.285±0.473 ^b	1.39±0.71 ^a	<0.001	0.150
	Medium	0.147±0.149 ^{bXY}	0.145±0.185 ^b	0.944±1.283 ^b	2.93±2.13 ^a	<0.001	0.274

	High	0.307±0.262 ^{bx}	0.439±0.404 ^{ab}	0.875±0.706 ^{ab}	1.15±1.39 ^a	0.012	0.135	
	<i>P</i> oxidative group	0.004	0.313	0.088	0.030			
<i>Ethyl- octanoate</i>	Low	0.945±0.995 ^{by}	26.9±24.7 ^a	33.3±27.1 ^a	48.0±11.3 ^a	<0.001	0.403	0.736
	Medium	4.26±5.55 ^{bxY}	37.4±19.6 ^a	43.5±19.1 ^a	50.6±15.2 ^a	<0.001	0.557	0.682
	High	9.11±10.70 ^{bx}	37.4±10.0 ^a	38.6±12.9 ^a	38.2±14.0 ^a	<0.001	0.469	0.579
	<i>P</i> oxidative group	0.007	0.214	0.394	0.091			
<i>2-Butanone</i>	Low	0.051±0.051	0.061±0.057	0.065±0.068	0.096±0.048 ^x	0.462		
	Medium	0.313±0.706	0.061±0.061	0.079±0.038	0.071±0.048 ^{xy}	0.193		
	High	0.203±0.333	0.074±0.078	0.051±0.031	0.046±0.024 ^y	0.045	0.115	
	<i>P</i> oxidative group	0.263	0.824	0.261	0.047			
<i>2-Heptanone</i>	Low	0.026±0.026 ^c	0.212±0.141 ^{bc}	0.314±0.259 ^b	0.742±0.225 ^a	<0.001	0.480	
	Medium	0.067±0.077 ^b	0.411±0.431 ^a	0.469±0.270 ^a	0.613±0.379 ^a	<0.001	0.295	0.520
	High	0.067±0.056 ^b	0.511±0.374 ^a	0.477±0.180 ^a	0.634±0.149 ^a	<0.001	0.443	0.635
	<i>P</i> oxidative group	0.067	0.053	0.113	0.630			
<i>3-Octanone</i>	Low	0.028±0.027 ^y	0.406±0.367 ^y	1.13±2.44	1.32±1.09	0.089	0.110	0.612
	Medium	0.121±0.167 ^{bxY}	1.56±1.71 ^{ax}	0.838±0.668 ^{ab}	0.874±0.482 ^{ab}	0.002	0.075	0.431
	High	0.169±0.218 ^{cx}	0.966±0.486 ^{axy}	0.772±0.599 ^{ab}	0.473±0.272 ^{bc}	<0.001	0.106	0.325
	<i>P</i> oxidative group	0.048	0.014	0.774	0.019			
<i>2-Octanone</i>	Low	0.031±0.049 ^b	0.457±0.259 ^{ab}	0.828±0.970 ^a	1.04±0.71 ^a	0.001	0.284	
	Medium	0.096±0.129 ^b	1.13±1.18 ^a	1.01±0.57 ^a	1.18±0.80 ^a	0.001	0.224	
	High	0.099±0.125 ^b	0.809±0.566 ^a	0.736±0.416 ^a	0.655±0.527 ^a	<0.001	0.213	
	<i>P</i> oxidative group	0.138	0.064	0.520	0.159			
<i>2-Nonanone</i>	Low	0.010±0.007 ^b	0.016±0.007 ^b	0.021±0.015 ^b	0.068±0.077 ^a	<0.001	0.151	
	Medium	0.015±0.007	0.038±0.051	0.031±0.017	0.045±0.026	0.065	0.101	
	High	0.013±0.006 ^b	0.028±0.022 ^a	0.027±0.017 ^{ab}	0.031±0.012 ^a	0.007	0.164	
	<i>P</i> oxidative group	0.138	0.167	0.232	0.146			
<i>1-Octen-3-one</i>	Low	0.005±0.009 ^{by}	0.102±0.123 ^{aby}	0.138±0.136 ^a	0.068±0.042 ^{ab}	0.004	0.163	
	Medium	0.017±0.025 ^{bxY}	0.311±0.310 ^{ax}	0.134±0.123 ^b	0.098±0.055 ^b	<0.001		
	High	0.092±0.157 ^x	0.147±0.141 ^{xy}	0.196±0.255	0.074±0.045	0.194		0.093

<i>P</i> oxidative group		0.023	0.020	0.569	0.359		
<i>3-Octen-2-one</i>	Low	n.d. ^Y	0.060±0.048	0.291±0.561	0.095±0.081	0.064	
	Medium	0.001±0.001 ^{XY}	0.445±1.090	0.221±0.213	0.267±0.342	0.245	
	High	0.002±0.004 ^X	0.177±0.120	0.387±0.755	0.174±0.125	0.057	0.063
<i>P</i> oxidative group		0.038	0.228	0.710	0.320		
<i>3-Nonen-2-one</i>	Low	n.d.	0.005±0.009	0.071±0.208	0.006±0.007	0.281	
	Medium	n.d.	0.039±0.059	0.029±0.034	0.026±0.042	0.063	0.071
	High	n.d. ^b	0.033±0.034 ^a	0.025±0.036 ^{ab}	0.016±0.017 ^{ab}	0.005	0.073
<i>P</i> oxidative group		0.340	0.050	0.504	0.357		
<i>2,3-Octanedione</i>	Low	0.058±0.038	23.4±43.1	9.10±10.00	3.94±7.08	0.062	0.465
	Medium	0.124±0.124 ^b	26.2±31.7 ^a	4.75±6.22 ^b	1.80±1.07 ^b	<0.001	0.294
	High	2.78±7.17	9.83±14.03	9.23±17.46	2.44±2.19	0.174	0.142
<i>P</i> oxidative group		0.126	0.291	0.530	0.458		
<i>2-Pentylfuran</i>	Low	0.065±0.077 ^{bY}	1.54±1.98 ^b	6.25±10.75 ^{abY}	8.91±6.72 ^a	0.007	0.176
	Medium	0.345±0.440 ^{bXY}	8.82±16.89 ^{ab}	7.36±5.75 ^{abY}	17.0±11.7 ^a	0.002	0.198
	High	0.567±0.766 ^{cX}	9.56±5.55 ^b	17.1±12.2 ^{abX}	20.2±12.9 ^a	<0.001	0.415
<i>P</i> oxidative group		0.029	0.069	0.006	0.161		

¹ Oxidative groups, according TBARS values on day 9 of display from samples aged 15 days: low (< 1 mg MDA/kg, *n*=16), medium (1-2 mg MDA/kg, *n*=15), and high (> 2 mg MDA/kg, *n*=17). ²: Root mean squared error. ³: the R² of linear and exponential regressions from each compound through time of display are shown when significant (*P*≤0.05). n.d.: not detected. a-c: different superscripts within a row denote statistical differences, according the Tukey test (*P*≤0.05) between display time within oxidative group for a compound. X-Z: different superscripts in the same column denote statistical differences (*P*≤0.05) between oxidative groups within day on display, for a compound.

Table 4. Mean and standard deviation for intensity¹ of odours and TBARS (mg MDA/kg muscle) of raw knuckle steaks aged for 15 days under vacuum conditions and displayed in modified atmosphere packaging (0, 5, 7 or 9 days) according oxidative group².

Attribute	Oxidative group	Display (days)				<i>P</i> display	Regressions ³	
		0	5	7	9		Linear	Exponential
<i>Raw meat odour</i>	Low	4.15±2.26 ^{aXY}	3.35±1.26 ^{abX}	2.39±0.64 ^{bcX}	1.98±0.60 ^{cx}	<0.001	0.375	0.252
	Medium	4.66±2.25 ^{aX}	3.23±1.21 ^{bX}	2.35±0.70 ^{bX}	2.04±0.49 ^{bX}	<0.001	0.375	0.323
	High	2.82±1.12 ^{aY}	2.26±0.86 ^{abY}	1.70±0.45 ^{bcY}	1.29±0.57 ^{cY}	<0.001	0.342	0.437
<i>P</i> oxidative group		0.026	0.014	0.003	<0.001			
<i>Rancid odour</i>	Low	1.92±0.94 ^{bXY}	3.20±0.66 ^{aY}	3.43±0.69 ^{aY}	3.61±0.73 ^{az}	<0.001	0.429	0.396
	Medium	1.46±0.65 ^{cY}	3.55±0.77 ^{bXY}	3.89±1.11 ^{bY}	5.09±1.17 ^{aY}	<0.001	0.659	0.691
	High	2.35±0.73 ^{dX}	4.21±0.89 ^{cX}	5.30±0.73 ^{bX}	6.25±0.98 ^{aX}	<0.001	0.755	0.710
<i>P</i> oxidative group		0.009	0.002	<0.001	<0.001			
<i>Metallic odour</i>	Low	0.271±0.374	0.254±0.213	0.422±0.437	0.354±0.374	0.526		
	Medium	0.455±0.464	0.151±0.146	0.202±0.383	0.188±0.207	0.053	0.092	
	High	0.527±0.292 ^a	0.329±0.273 ^{ab}	0.147±0.153 ^b	0.371±0.126 ^a	<0.001	0.124	
<i>P</i> oxidative group		0.150	0.085	0.064	0.101			
<i>Sweet odour</i>	Low	2.20±1.66	1.31±1.00	2.01±0.60 ^X	1.77±0.55	0.107		
	Medium	2.56±1.03 ^a	1.29±0.66 ^b	1.41±0.34 ^{bY}	1.39±0.77 ^b	<0.001	0.263	0.160

	High	1.98±0.80 ^a	1.59±0.68 ^{ab}	1.38±0.51 ^{by}	1.27±0.49 ^b	0.009	0.162	
	<i>P</i> oxidative group	0.408	0.495	0.001	0.063			
<i>Musty odour</i>	Low	0.546±0.434 ^{xy}	0.551±0.420	0.554±0.384 ^y	0.500±0.322 ^y	0.977		
	Medium	0.338±0.356 ^y	0.690±0.623	0.837±0.719 ^{xy}	0.777±0.466 ^y	0.078	0.101	
	High	0.780±0.489 ^{bx}	0.994±0.627 ^b	1.21±0.82 ^{bx}	1.95±1.07 ^{ax}	<0.001	0.187	
	<i>P</i> oxidative group	0.022	0.081	0.027	<0.001			
<i>Cooked meat odour</i>	Low	0.455±0.602	0.689±0.288	0.467±0.438 ^{xy}	0.791±0.383 ^{xy}	0.091		
	Medium	0.177±0.337 ^b	0.953±0.662 ^a	0.389±0.472 ^{by}	1.13±0.65 ^{ax}	<0.001	0.183	
	High	0.196±0.344 ^b	0.664±0.451 ^a	0.823±0.365 ^{ax}	0.735±0.295 ^{ay}	<0.001	0.270	
	<i>P</i> oxidative group	0.156	0.196	0.012	0.040			
<i>Sulphury odour</i>	Low	1.31±1.32	0.729±0.520 ^y	1.07±0.60	0.948±0.363	0.224		
	Medium	1.89±1.91	0.836±0.744 ^y	1.20±0.80	1.31±0.52	0.092		
	High	2.34±1.50 ^a	1.44±0.69 ^{bx}	1.49±0.69 ^b	1.32±0.58 ^b	0.009	0.145	
	<i>P</i> oxidative group	0.183	0.007	0.231	0.063			
<i>TBARS value</i>	Low	0.067±0.012 ^b	0.356±0.177 ^{aby}	0.661±0.650 ^{ay}	0.629±0.260 ^{az}	<0.001	0.294	0.735
	Medium	0.070±0.015 ^d	0.626±0.148 ^{cy}	1.00±0.33 ^{by}	1.43±0.22 ^{ay}	<0.001	0.832	0.931
	High	0.074±0.013 ^d	1.36±0.54 ^{cx}	1.99±0.87 ^{bx}	3.07±0.63 ^{ax}	<0.001	0.751	0.893
	<i>P</i> oxidative group	0.260	<0.001	<0.001	<0.001			

¹ 0 (no detected) – 10 (very intense). ² Oxidative groups, according TBARS values on day 9 of display from samples aged 15 days: low (< 1 mg MDA/kg), medium (1-2 mg MDA/kg), and high (> 2 mg MDA/kg). ³: the R² of linear and exponential regressions from each compound through time of display are shown when significant ($P \leq 0.05$). RMSE: Root mean squared error. a-d: different superscripts within a row denote statistical differences, according the Tukey test ($P \leq 0.05$), between display time within oxidative group for a compound. X-Z: different superscripts in the same column denote statistical differences ($P \leq 0.05$) between oxidative groups within day on display, for a compound.

Table 5. Mean and standard deviation for intensity¹ of odours and TBARS (mg MDA/kg muscle) of raw knuckle steaks aged for 22 days under vacuum conditions and displayed in modified atmosphere packaging (0, 5, 7 or 9 days) according oxidative group².

Attribute	Oxidative group	Display (days)				<i>P</i> display	Regressions ³	
		0	5	7	9		Linear	Exponential
<i>Raw meat odour</i>	Low	2.75±0.67 ^a	1.72±0.41 ^{bc}	2.14±0.46 ^{bx}	1.44±0.39 ^{cx}	<0.001	0.324	0.288
	Medium	2.33±0.82 ^a	1.84±0.39 ^{ab}	1.64±0.72 ^{bx}	1.26±0.68 ^{bx}	0.002	0.244	0.231
	High	2.72±0.63 ^a	1.70±0.40 ^b	0.989±0.583 ^{cy}	0.498±0.312 ^{dy}	<0.001	0.727	0.675
<i>P</i> oxidative group		0.194	0.583	<0.001	<0.001			
<i>Rancid odour</i>	Low	2.24±0.76 ^c	3.29±1.13 ^{by}	4.08±0.67 ^{by}	5.20±0.91 ^{ay}	<0.001	0.532	0.499
	Medium	2.45±0.53 ^c	4.51±0.72 ^{bx}	4.88±1.40 ^{by}	6.06±1.53 ^{axy}	<0.001	0.583	0.636
	High	2.58±0.87 ^c	5.25±1.14 ^{bx}	5.82±1.26 ^{bx}	7.13±1.64 ^{ax}	<0.001	0.648	0.672
<i>P</i> oxidative group		0.417	<0.001	<0.001	0.034			
<i>Metallic odour</i>	Low	0.638±0.391 ^a	0.270±0.240 ^b	0.133±0.155 ^b	0.277±0.171 ^b	<0.001	0.317	
	Medium	0.394±0.314 ^a	0.325±0.147 ^{ab}	0.178±0.146 ^b	0.331±0.123 ^{ab}	0.034		
	High	0.770±0.575 ^a	0.317±0.116 ^b	0.225±0.165 ^b	0.436±0.199 ^b	<0.001	0.194	
<i>P</i> oxidative group		0.067	0.645	0.255	0.124			
<i>Sweet odour</i>	Low	1.63±0.38	1.53±0.81	1.37±0.59	1.20±0.49	0.406		
	Medium	1.45±0.67	1.39±0.73	1.30±0.80	0.791±0.370	0.065	0.074	
	High	1.62±0.60 ^a	0.957±0.569 ^b	1.42±0.48 ^{ab}	0.999±0.565 ^b	0.002	0.094	

<i>P</i> oxidative group		0.607	0.062	0.863	0.243			
<i>Musty odour</i>	Low	0.327±0.148 ^{by}	0.575±0.372 ^b	0.632±0.507 ^b	2.03±0.59 ^a	<0.001	0.312	
	Medium	0.763±0.550 ^{bx}	0.807±0.344 ^b	0.587±0.495 ^b	1.62±0.86 ^a	<0.001	0.079	
	High	0.568±0.405 ^{bxY}	0.852±0.407 ^b	0.450±0.376 ^b	1.81±0.71 ^a	<0.001	0.174	
<i>P</i> oxidative group		0.015	0.101	0.505	0.561			
<i>Cooked meat odour</i>	Low	0.321±0.390 ^b	0.400±0.519 ^{by}	0.555±0.465 ^b	1.50±0.33 ^a	<0.001	0.192	
	Medium	0.185±0.245 ^c	0.698±0.510 ^{bxY}	0.433±0.371 ^{bc}	1.14±0.43 ^a	<0.001	0.288	
	High	0.318±0.383 ^c	0.897±0.506 ^{bx}	0.774±0.357 ^b	1.47±0.58 ^a	<0.001	0.359	
<i>P</i> oxidative group		0.467	0.030	0.059	0.174			
<i>Sulphury odour</i>	Low	2.24±0.85 ^{axY}	0.833±0.576 ^{cY}	0.999±0.572 ^{bcY}	1.70±0.97 ^{ab}	<0.001	0.218	
	Medium	2.55±1.26 ^{ax}	1.43±0.61 ^{bx}	1.39±0.62 ^{bxY}	2.00±0.67 ^{ab}	0.001	0.107	
	High	1.50±0.64 ^{by}	1.32±0.51 ^{bxY}	1.56±0.69 ^{bx}	2.13±0.52 ^a	0.004	0.065	0.049
<i>P</i> oxidative group		0.008	0.014	0.049	0.439			
<i>TBARS value</i>	Low	0.071±0.011 ^{cY}	0.506±0.391 ^{bcY}	0.796±0.545 ^{by}	1.60±0.56 ^{aY}	<0.001	0.498	0.792
	Medium	0.080±0.014 ^{cY}	0.832±0.394 ^{by}	1.20±0.48 ^{by}	2.19±0.85 ^{axY}	<0.001	0.632	0.872
	High	0.104±0.021 ^{cX}	1.39±0.55 ^{bx}	2.47±1.05 ^{ax}	3.08±1.02 ^{ax}	<0.001	0.678	0.899
<i>P</i> oxidative group		<0.001	<0.001	<0.001	0.005			

¹ 0 (no detected) – 10 (very intense). ² Oxidative groups, according TBARS values on day 9 of display from samples aged 15 days: low (< 1 mg MDA/kg), medium (1-2 mg MDA/kg), and high (> 2 mg MDA/kg). RMSE: Root mean squared error. ³: the R² of linear and exponential regressions from each compound through time of display are

shown when significant ($P \leq 0.05$). a-d: different superscripts within a row denote statistical differences, according the Tukey test ($P \leq 0.05$), between display time within oxidative group for a compound. X,Y: different superscripts in the same column denote statistical differences ($P \leq 0.05$) between oxidative groups within day on display, for a compound.

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2 **Table 6.** Pearson correlations coefficients between the volatile compounds, TBARS and odour attributes of knuckle steaks aged for 15 or 22 days under
 3 vacuum conditions and displayed in modified atmosphere packaging (0, 5, 7 or 9 days).

	<i>TBARS</i>	<i>Raw meat</i>	<i>Rancid</i>	<i>Mettalic</i>	<i>Sweet</i>	<i>Moisty</i>	<i>Cooked meat</i>	<i>Sulphury</i>
<i>Butanoic acid</i>	ns	-0.257***	ns	0.208***	-0.280***	ns	ns	ns
<i>Pentanoic acid</i>	0.814***	-0.503***	0.755***	ns	-0.364***	0.447***	0.429***	ns
<i>Hexanoic acid</i>	0.698***	-0.446***	0.688***	-0.170***	-0.366***	0.389***	0.422***	ns
<i>Heptanoic acid</i>	0.637***	-0.350***	0.629***	-0.230***	-0.242***	0.347***	0.353***	ns
<i>Octanoic acid</i>	0.106*	ns	0.184***	-0.287***	ns	ns	0.108*	-0.191***
<i>Nonanoic acid</i>	ns	0.171***	ns	-0.228***	ns	ns	ns	-0.222***
<i>Decanoic acid</i>	ns	0.108*	ns	-0.135**	ns	ns	ns	ns
<i>1-Hexanol</i>	0.822***	-0.459***	0.706***	ns	-0.289***	0.403***	0.368***	ns
<i>1-Octen-3-ol</i>	0.134**	-0.221***	ns	0.105*	-0.112*	0.147**	ns	ns
<i>Hexanal</i>	0.135**	ns	0.177***	-0.136**	ns	ns	0.176***	ns
<i>Heptanal</i>	0.175***	ns	0.158**	-0.119*	-0.107*	0.150**	ns	ns
<i>Nonanal</i>	ns	ns	ns	-0.106*	ns	ns	ns	ns
<i>(E)-2-Heptenal</i>	0.349***	-0.191***	0.374***	ns	-0.186***	0.133*	0.266***	ns
<i>(Z)-2-Octenal</i>	0.430***	-0.185***	0.438***	-0.123*	-0.212***	0.199***	0.331***	ns
<i>(E)-2-Nonenal</i>	0.369***	-0.175***	0.392***	ns	-0.167***	0.112*	0.288***	ns
<i>(E,E)-2,4-Nonadienal</i>	0.225***	-0.125*	0.239***	ns	ns	ns	0.169***	ns
<i>(E)-2-Undecenal</i>	0.654***	-0.335***	0.617***	-0.119*	-0.270***	0.289***	0.369***	ns

<i>(E,Z)</i> -2,4-Decadienal	0.120*	ns	0.159**	-0.108*	ns	ns	0.178***	ns
<i>(E,E)</i> -2,4-Decadienal	0.378***	-0.150**	0.396***	-0.134**	-0.169***	0.116*	0.293***	ns
<i>Phenylacetaldehyde</i>	ns	ns	ns	0.225***	ns	ns	ns	ns
<i>Ethyl- hexanoate</i>	0.542***	-0.284***	0.426***	ns	-0.230***	0.369***	0.194***	ns
<i>Ethyl- octanoate</i>	0.635***	-0.355***	0.636***	-0.237***	-0.260***	0.406***	0.374***	ns
<i>2-Butanone</i>	ns	ns	ns	ns	ns	ns	ns	ns
<i>2-Heptanone</i>	0.579***	-0.337***	0.602***	-0.130*	-0.265***	0.331***	0.390***	ns
<i>3-Octanone</i>	0.231***	ns	0.267***	-0.143**	-0.137**	0.103*	0.276***	ns
<i>2-Octanone</i>	0.426***	-0.209***	0.459***	-0.149**	-0.186***	0.225***	0.275***	ns
<i>2-Nonanone</i>	0.279***	-0.146**	0.302***	-0.104*	-0.142**	0.191***	0.265***	ns
<i>1-Octen-3-one</i>	0.144**	ns	0.198***	-0.183***	ns	ns	0.195***	ns
<i>3-Octen-2-one</i>	0.300***	-0.157**	0.319***	-0.123*	-0.147**	0.121*	0.216***	ns
<i>3-Nonen-2-one</i>	0.206***	-0.123*	0.227***	ns	-0.136**	ns	0.164**	ns
<i>2,3-Octanedione</i>	ns	ns	0.105*	-0.105*	ns	ns	0.177***	ns
<i>2-Pentylfuran</i>	0.714***	-0.382***	0.662***	ns	-0.281***	0.323***	0.379***	ns
<i>TBARS</i>	1	-0.524***	0.819***	ns	-0.282***	0.463***	0.438***	ns
<i>Raw meat</i>		1	-0.656***	ns	0.543***	-0.382***	-0.272***	-0.251***
<i>Rancid</i>			1	-0.147**	-0.455***	0.450***	0.503***	ns
<i>Mettalic</i>				1	ns	ns	ns	0.190***
<i>Sweet</i>					1	-0.288***	-0.322***	-0.107*
<i>Moisty</i>						1	0.269***	ns
<i>Cooked meat</i>							1	ns

4 ns: $P > 0.05$, *: $P \leq 0.05$, **: $P \leq 0.01$, *** $P \leq 0.001$.