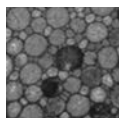


Palaeogeographical patterns in Late Ordovician bryozoan morphology as proxies for temperature

ANDREA JIMÉNEZ-SÁNCHEZ, PAUL D. TAYLOR & JAVIER B. GÓMEZ



Several studies have revealed temperature-related patterns in recent bryozoans, both in the chemical composition of the skeleton and in the morphological characters of the colonies, but comparable studies on Palaeozoic bryozoans are lacking. In this paper a statistical analysis of the morphological differences is undertaken between congeneric species of some Ordovician bryozoans from warm- and cold-water settings. For this study ten eurythermic cosmopolitan bryozoan genera from the Upper Ordovician were selected from the Mediterranean, Avalonia, Baltic and Laurentia-Siberian provinces. These genera are: *Ceramopora* and *Ceramoporella* (Cystoporata); *Diplotrypa*, *Eridotrypa*, *Hallopora*, *Heterotrypa*, *Monticulipora* and *Trematopora* (Trepotomata); *Graptodictya* (Cryptostomata); and *Kukersella* (Cyclostomata). The study involved 154 samples belonging to 104 different species. Twenty-eight morphological characters were measured, although only 21 were used in the final statistical analysis. Univariate (t, F, Kolmogorov-Smirnov and Mann-Whitney tests), multivariate discriminant and multivariate ordination (Principal Coordinates, Principal Components, Correspondence, and Detrended Correspondence) analyses were performed on the data. For the univariate and multivariate discriminant analyses, the total set of samples was divided *a priori* into cold- and warm-water subsets based on palaeolatitude: samples from the Mediterranean province were attributed to the cold-water subset, whereas samples from Avalonia, Baltic and the Laurentian-Siberian provinces were included in the warm-water subset. For the multivariate ordination analysis no *a priori* grouping by water temperature was imposed, and the aim of these analyses was to test whether different samples were correctly arranged along a water temperature gradient. The univariate statistical analysis showed that there are clear morphological differences between cold- and warm-water species in six of the ten Late Ordovician bryozoan genera analysed in this study, although these differences are only evident for some of the characters used, and only when the analysis is performed on individual genera. The best characters to differentiate species by water temperature are those related to the size of the zooidal polymorphs, especially the diameters of the autozoocia, mesozooecia and exilazooecia. With the exception of one genus (*Trematopora*), cold-water species have larger zooids. The discriminant analysis was able to classify correctly as warm- or cold-water 100% of the samples for two genera, slightly below 95% for two other genera, and between 67% and 90% for the remaining six genera. Finally, the multivariate ordination analysis was able to separate species by palaeogeographical province in some genera, but these provinces were not correctly arranged along a palaeolatitudinal gradient using any of the methods used. • Key words: bryozoans, cosmopolitan genera, latitudinal adaptations, univariate and multivariate statistical analysis.

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The Bryozoa is a phylum of mostly marine invertebrates containing species that are adapted to living in waters of a wide range of temperatures, from the warmth of the tropics to the cold of the Arctic and Antarctic oceans. However, their temperature-related distribution has changed since the Palaeozoic, when they were most abundant on tropical platforms, compared with today, where their abundance is greater in higher latitudes (Taylor & Allison 1998, Taylor & Sendino 2010). Notwithstanding the prevalence of bryo-

zoans on tropical platforms in the Palaeozoic, during the Late Ordovician (455–445 Ma), they were one of the most abundant invertebrate groups, together with brachiopods and echinoderms, on the North Gondwana platforms, located during this time at high latitudes, between 40° and 60° S (Jiménez-Sánchez & Villas 2010).

During the Late Ordovician a total of 68 bryozoan genera are known to have colonised the cold carbonate platforms of the Mediterranean province (Cocks & Torsvik

2006, Jiménez-Sánchez & Villas 2010); 22 of these genera were endemic to this province while 46 were also present on tropical and equatorial carbonate platforms (Jiménez-Sánchez & Villas 2010). Study of the cosmopolitan genera offers the possibility of ascertaining if and how these genera were able to adapt morphologically, chemically and mineralogically to life across such a wide range of temperatures.

Published studies carried out on Recent bryozoans belonging to the orders Cheilostomata and Cyclostomata point to some common temperature-related patterns. Carbonate skeletons of cheilostome bryozoans living in warm waters comprise calcite, aragonite or are bimineralic (with calcite overlain by aragonite), and in those employing calcite the percentage of Mg is often high (Smith *et al.* 2006, Taylor *et al.* 2009). On the other hand, in cold-water, cheilostome species with aragonitic and bimineralic skeletons are rare and the Mg content in the calcite is typically lower (Kuklinski & Taylor 2008, 2009; Loxton *et al.* 2012). These mineralogical and chemical differences in bryozoan skeletons have been linked to environmental factors – mainly temperature – because they have also been found to occur within congeneric species that inhabit different temperature habitats. In addition, Schäfer & Bader (2008) showed that the Mg content of bryozoan calcite could vary in a single skeleton depending on the season when the calcite was secreted, its content being lower in cold than warm water. We here place the limit between cold- and warm-water at around 18 °C, the temperature beneath which tropical corals cannot live at the present-day (*e.g.*, Schlager 2005, fig. 2.8).

In addition to these mineralogical and chemical variations, cold- and warm-water species may show differences in morphology, including the development of polymorphic zooids and other features linked to environmental variations in seasonality, predator pressure, availability of food etc. For example, according to Hughes & Jackson (1990) and Kuklinski & Taylor (2008), avicularia, generally regarded as having a defensive role, are smaller and fewer in number in high latitude congeneric species, suggesting that predator pressure is relatively less important than physical stresses in these environments. In addition, there is an inverse intraspecific relationship between the size of the zooids in living cheilostome bryozoan colonies and the ambient temperature at the time of budding (see Okamura *et al.* 2011). Limited evidence suggests that this relationship is also true between congeneric species: Kuklinski & Taylor (2008) found that the size of the autozoecia in cheilostome species belonging to six of the eight genera they studied was significantly greater in the Arctic than in congeneric species from lower latitude, warmer water sites.

The aim of this paper is to test statistically for morphological differences in representative Upper Ordovician eurythermic bryozoan genera that inhabited the equatorial and tropical latitudes of Avalonia and Baltic and Laurentia-

Siberia provinces, compared with bryozoans from the almost polar latitudes of the Mediterranean province (Carnic Alps, Montagne Noire, Iberian Chains and Libya). As we are considering palaeolatitude as a proxy for water temperature, the prediction is that related (*e.g.* congeneric) species from different palaeolatitudinal settings will show consistent morphological differences, similar to those found among Recent bryozoans from waters of different temperature.

Studied material

In this study the term ‘sample’ is used for species from a single locality. Thus, if the same species has been described in four different localities, it is counted here as four different ‘samples’ of the same species. A sample can be a single colony, or several colonies or fragments of colonies, provided they come from the same locality and have been identified as conspecific.

A total of ten genera were studied, all present at both high and low palaeolatitudes. These genera are: *Ceramopora* (Cystoporata; 9 samples belonging to 6 species), *Ceramoporella* (Cystoporata; 15 samples, 9 species), *Diplotrypa* (Trepotomata; 17 samples, 14 species), *Eridotrypa* (Trepotomata; 22 samples, 14 species), *Graptodictya* (Cryptostomata; 9 samples, 6 species), *Hallopora* (Trepotomata; 32 samples, 23 species), *Heterotrypa* (Trepotomata; 5 samples, 3 species), *Kukersella* (Cyclostomata; 12 samples, 1 species), *Monticulipora* (Trepotomata; 21 samples, 17 species) and *Trematopora* (Trepotomata; 12 samples, 11 species). One-hundred and seven of these samples came from the low and middle-low latitude provinces of Avalonia, Baltic and Laurentia-Siberia, and the rest (45 samples) from the high and middle-high latitude of Mediterranean province. We have also included a small number of samples belonging to the genera *Diplotrypa*, *Eridotrypa*, *Graptodictya* and *Monticulipora* from the late Middle Ordovician in order to enlarge the sample size for each genus in all studied regions. The species *Hallopora elegantula*, *Kukersella borealis* and *Monticulipora kolaluensis* have been recorded from low and middle-low latitudes as well as high- and middle-high latitudes.

Most of the material used is stored in the Natural History Museum, London (NHMUK) and in the Paleontological Museum of the University of Zaragoza (MPUZ), Spain. The rest of the samples are stored in other European and American institutions. A list of samples, localities and institutions where the samples are stored is provided as online supplementary material (www.geology.cz).

Methodology

Morphological study of the bryozoan genera listed above was undertaken by measuring several characteristics of the

Table 1. Morphological characters used in the statistical analysis.

Character	Transcription	Genus									
		<i>Ceramopora</i>	<i>Ceramoporella</i>	<i>Diplotrypa</i>	<i>Eridotrypa</i>	<i>Graptodictya</i>	<i>Haltopora</i>	<i>Heterotrypa</i>	<i>Kukersella</i>	<i>Monticulipora</i>	<i>Trematopora</i>
AAZS	Autozooeccial intersection angle with colony surface						X	X			
AD	Autozooeccial diameter	X	X	X	X		X	X	X	X	X
ADalWTh	Autozooeccial distal wall thickness								X		
AD _{large}	Autozooeccial large diameter					X					
AltPxWTh	Autozooeccial lateral and proximal wall thickness								X		
AthD	Acanthostyle diameter							X			X
AWThEx	Autozooeccial wall thickness in exozone	X	X		X		X	X			X
BchD	Branch diameter						X	X	X		X
BchD _{small}	Branch small diameter					X					
EndozoneD	Endozone diameter							X	X		
EndozoneTh	Endozone thickness					X					
ExiD	Exilazooeccial diameter	X	X								
ExozoneTh	Exozone thickness							X	X		
MD	Mesozooeccial diameter				X		X	X		X	X
N°A1mm	Number of autozooeccia per one millimetre				X			X		X	X
N°A1mm ²	Number of autozooeccia per one square millimetre						X	X		X	X
N°Ath1mm ²	Number of acanthostyle per one square millimetre										X
N°M1mm	Number of mesozooeccia per one millimetre									X	
N°M1mm ²	Number of mesozooeccia per one square millimetre									X	
N°MDph1mm	Number of mesozooeccial diaphragms per one millimetre						X				
ZWTh	Zooeccial wall thickness				X					X	

colonies and of component parts such as individual polymorph zooids and acanthostyles (Fig. 1). The resulting quantitative data was analysed with univariate and multivariate statistical techniques using the PAST v. 2.15 palaeontological software (Hammer *et al.* 2008).

A total of 28 different morphological characters were measured, both from the colony (height and diameter of branches in most cases) and from individual elements of the colony (autozooeccia, mesozooeccia, exilazooeccia, acanthostyles *etc.*). Measurements were taken either directly from thin sections of the samples using a microscope graticule, or from scaled photographs. Not all of the measurable morphological characters could be taken from each sample due to the lack of correctly oriented thin sections. Only those characters that were measured in at least 80% of the samples of particular genera were included in the final statistical analysis. Missing values in the remaining characters for a genus were computed as the arithmetic mean of the character in the samples belonging to the same temperature range (*i.e.* cold- or warm-water). Table 1 lists the characters used in the statistical analyses (21 out of the initial 28), together with the genera in which the characters were measured.

In order to check whether a specific character could be used to separate cold- and warm-water taxa, several uni-

variate and multivariate statistical tests were performed.

First, using the *a priori* knowledge of the palaeogeographical positions of each palaeocontinent in the Late Ordovician (Fortey & Cocks 2003, Jiménez-Sánchez & Villas 2010), the total sample was divided into two subsamples: a cold-water subsample (Mediterranean province) and a warm-water subsample (Avalonia region and Baltic and Laurentia-Siberian provinces). It is important to remember here that all of the genera selected for this study contain both cold- and warm-water species.

The first statistical test checked whether the two subsamples (for a specific genus) belonged to the same underlying population or to two different populations. Differences in mean, variance and distribution were tested.

The type of test performed depended on the probability distribution of the sub-samples for each genus. If both sub-samples of a genus were Shapiro-Wilk normally distributed, then the ‘t’ test of equality of means and the ‘F’ test of equality of variances were performed. On the other hand, if either of the two sub-samples of the genus was not Shapiro-Wilk normally distributed, then two non-parametric tests were performed: the Mann-Whitney test for the equality of means, and the Kolmogorov-Smirnov test for the equality of distributions. Normality was tested for a level of significance of 5% (*i.e.*, the null hypothesis of an

underlying normal distribution was rejected if the p-value is smaller than 0.05).

The outcome of the previous univariate analysis determines whether a specific character could be used to split, in a statistically significant sense, cold- from warm-water species belonging to the same genus. But this statistical significance can be marginal, in which case the differences cannot be used reliably to classify species of the same genus as either cold- or warm-water. In order to assess the reliability of the assignation of a new taxon to one of the two subsamples (cold- or warm-water), a multivariate discriminant analysis was performed. Discriminant analysis is a statistical technique allowing the study of differences between two or more groups of objects with respect to several variables simultaneously. In this sense it can be understood as a sort of multivariate version of the univariate analysis explained above. The aim of the method is to find a function (the discriminant function) that best separates one group from the other. Once the discriminant function is known, any new species from the studied genera can be assigned to one or the other of the two groups. A discriminant function able to classify at least 95% of the species in the correct (*a priori*) group is considered here to be good.

In the previous statistical analyses taxa were separated on the basis of their palaeogeographical occurrence into two water temperature regimes: cold- and warm-water. However, water temperature is not a cold/warm binary variable. There are obvious temperature gradients from the cold waters of the near-polar location of some parts of the Mediterranean province, to the warm waters of the equatorial regions of Laurentia. Consequently, in order to check for environmental temperature gradients and their impact on morphological features, several ordination and data clustering multivariate analyses were carried out, comprising Principal Coordinates, Principal Components, Correspondence, Detrended Correspondence, and Discriminant analyses. The Euclidean distance was used in all multivariate tests requiring a similarity measure.

Results

Univariate analysis

Table 2 shows the results of the univariate tests performed. The first column gives the genus and the analysed character. The second column states the number of studied samples per genus, divided into subsets from cold- and warm-water. In the third column the results of the normality tests for both populations, expressed in terms of p-values, are given. A significance level of 5% was chosen, so p-values less than 0.05 indicate that the sample did not pass this test, *i.e.*, the distribution of the subjacent population is not normal; in this column numbers in boldface indi-

cate that both subsamples (cold- and warm-water) passed the normality test, whereas numbers in red indicate that at least one subsample did not pass this test for a specific character. In the fourth and fifth columns, respectively, are given the results of the 'F' and 't' tests; these results are only shown when both subsamples passed the normality test (crossed-out figures mean that the normality test was not passed). The sixth and seventh columns show the results of the Kolmogorov-Smirnov (K-S) and Mann-Whitney (M-N) tests, respectively; these tests were used only when subsamples did not pass the normality test. The last column gives the arithmetic mean of each analysed character in the cold- and warm-water subsamples; the higher value is marked in boldface.

The results summarized in Table 2 show that not all characters were able to discriminate between cold- and warm-water subsamples, some characters being much more useful than others. The best characters to differentiate the subsamples are autozooeical diameter (AD) and the diameters of the different polymorphic mesozooecia and exilazooecia (MD and ExiD, respectively). In all genera these diameters are larger in cold-water species than they are in warm-water species (Fig. 2), with the exception of the genus *Trematopora* in which the diameters of autozooeecia and mesozooecia are smaller in species from cold- than warm-water, although it is still possible to differentiate the subsamples using the Mann-Whitney test for the equality of means.

None of the characters analysed in the genera *Ceramopora*, *Graptodictya*, *Heterotrypa* and *Kukersella* were able to separate the cold- from warm-water subsamples. Genera belonging to the order Trepostomata seem to show the greatest morphological differences between cold- and warm-water species; however, as pointed out above, there are exceptions (*e.g.*, *Heterotrypa*). The genus *Ceramoporella* (order Cystoporata) is also notable because the two subsamples are clearly distinguishable using two of the three analysed characters (AD and ExiD).

The character 'autozooeical diameter (AD)' is the only one common to all ten studied genera. This fact allowed an additional univariate analysis to test whether cold- and warm-water species could be separated using this character in all genera at the same time. As can be seen from the last row of Table 2, the two subsamples do not pass the test for normality, and cold- and warm-water species can be only marginally separated using the Kolmogorov-Smirnov test (p-value = 0.045).

Discriminant analysis

For this analysis the same *a priori* knowledge of palaeolatitude (as a proxy for water temperature) is used to classify each species into one of two mutually exclusive groups: cold-water

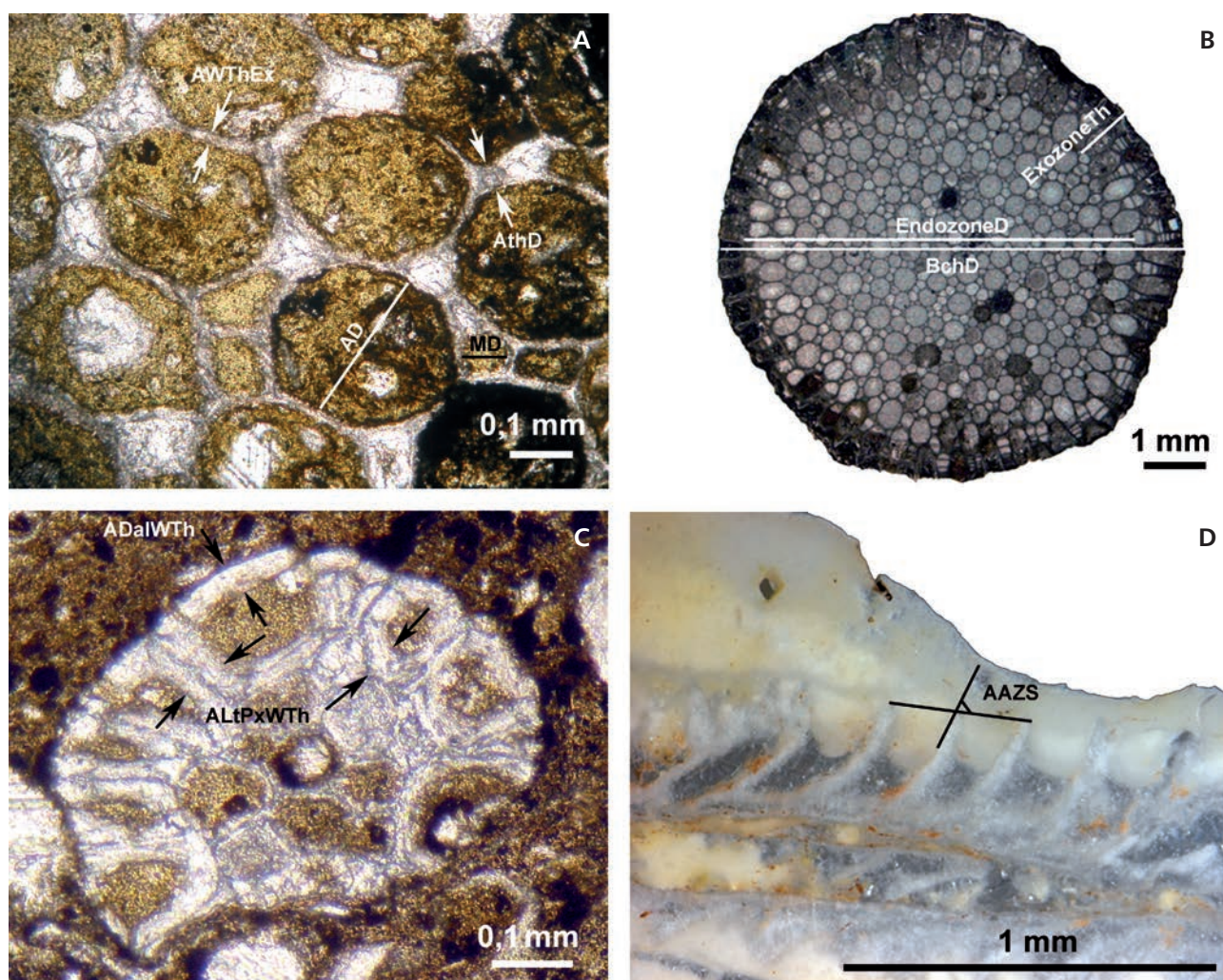


Figure 1. A – tangential section of *Monticulipora cystiphragmata* Jiménez-Sánchez, 2010 showing three types of measures: AD – autozooeical diameter; AthD – acanthostyle diameter; AWThEx – autozooeical wall thickness in exozone; and MD – mesozoeical diameter. The larger autozooeical diameter (AD_{large}) was measured in genera with oval autozooeical apertures. Zooeical wall thickness (ZWTh) was measured when the colony had a high number of mesozoeocia. Exilazooeical diameter (ExiD) was measured in tangential sections of the genera *Ceramopora* and *Ceramoporella*. $N^{\circ}A1mm$, $N^{\circ}A1mm^2$, $N^{\circ}Ath1mm^2$, $N^{\circ}M1mm$ and $N^{\circ}M1mm^2$ have been also measured in tangential sections. • B – transverse section of *Hallopora peculiaris* (described by Buttler 1991) showing characters branch diameter (BchD), endozone diameter (EndozoneD) and exozone thickness (ExozoneTh). In the genus *Graptiodictya*, which has an ellipsoidal cross section, the endozone thickness (EndozoneTh) and the branch small diameter ($BchD_{small}$) were measured instead of endozone and branch diameter. • C – transverse section of *Kukersella borealis* (described by Jiménez-Sánchez 2009) showing the three types of walls measured in *Kukersella*. • D – longitudinal section of *Ceramopora invenustra* Bassler, 1911 showing how the autozooeical angle with the zoarial surface (AAZS) was measured. Number of mesozoeical diaphragms per one millimetre ($N^{\circ}MDph1mm$) was also measured in this type of section.

species and warm-water species. A discriminant analysis was carried out for each of the 10 genera in the dataset.

The discriminant function for the genera *Heterotrypa* and *Kukersella* (Fig. 3) correctly classified all of the species (100% success rate). For the genera *Ceramoporella* and *Diplotrypa* the success rate was only slightly below 95%, although both genera gave a success rate of 100% when one species was excluded from the analysis (*C. inclinata* Jiménez-Sánchez, 2009, from the Mediterranean Province, and *D. moniliformis* Bassler, 1911, from the Baltic Province; see Fig. 4). The remaining genera have discriminant functions with success rates between 67% and 90% (Fig. 5).

Multivariate ordination analysis

The multivariate ordination analyses, whose aim is to discover if samples belonging to different provinces can be ordered according to a temperature (palaeolatitude) gradient, gave variable results. Using Principal Components, Principal Coordinates, Correspondence and Detrended Correspondence analyses, species belonging to the genera *Ceramopora*, *Eridotrypa*, *Hallopora*, *Heterotrypa*, *Kukersella* and *Monticulipora* did not follow patterns consistent with the province they belonged to: the convex hulls for the species from each province overlap each other and thus there is

Table 2. Results of the univariate statistical analysis.

Genus	No. of samples		Normality test		F test	t test	K-S test	M-W test	Mean	
	Cold	Warm	Cold	Warm					Cold	Warm
<i>Ceramopora</i> (AD)	4	5	0.07759	0.9907	0.05718	0.56718	0.4772	1	0.425	0.376
<i>Ceramopora</i> (AWThEx)	4	5	0.1612	0.03865	0.25194	0.26861	0.4772	0.3175	0.07	0.06
<i>Ceramopora</i> (ExiD)	4	5	0.5774	0.4211	0.35677	0.31117	0.7543	0.373	0.14	0.116
<i>Ceramoporella</i> (AD)	4	11	0.9033	0.492	3.6434E-06	0.0003591	0.005224	0.001465	0.5125	0.21091
<i>Ceramoporella</i> (AWThEx)	4	11	0.272	0.005451	0.020658	0.81268	0.3294	0.9201	0.0325	0.034
<i>Ceramoporella</i> (ExiD)	4	11	0.2353	0.04557	0.18313	0.0002392	0.005224	0.001465	0.19	0.09
<i>Diplotrypa</i> (AD)	4	13	0.9109	0.1276	0.062265	0.0078819	0.009553	0.01975	0.5425	0.40385
<i>Diplotrypa</i> (MD)	4	13	0.1612	0.06886	0.39698	0.0003457	0.003459	0.001681	0.18	0.12
<i>Diplotrypa</i> (N°A1mm)	4	13	0.5706	0.3415	0.27607	0.2519	0.4044	0.2811	1.6125	1.9638
<i>Diplotrypa</i> (ZWTh)	4	13	0.6283	0.00596	0.12871	0.49542	0.8819	0.6092	0.0225	0.01869
<i>Eridotrypa</i> (AD)	5	17	0.05023	0.0002997	0.70558	0.25228	0.1073	0.08107	0.16	0.21
<i>Eridotrypa</i> (AWThEx)	5	17	0.9683	0.006238	0.85275	0.2559	0.5753	0.3284	0.0662	0.087
<i>Eridotrypa</i> (BchD)	5	17	0.6611	0.01403	0.23458	0.02719	0.01296	0.00824	1.938	4.194
<i>Graptodictya</i> (AD _{large})	5	4	0.04354	0.4064	0.11042	0.4841	0.3572	0.5159	0.114	0.1275
<i>Graptodictya</i> (BchD _{small})	5	4	0.001942	0.3478	0.3883	0.78045	0.8778	0.9048	0.8	0.7125
<i>Graptodictya</i> (EndozoneTh)	5	4	0.7399	0.006192	0.0006888	0.75131	0.08215	0.254	0.194	0.23
<i>Hallopora</i> (AAZS)	9	23	0.04143	0.001587	0.055239	0.30267	0.8257	0.5973	76.463	81.269
<i>Hallopora</i> (AD)	9	23	0.5126	0.2652	0.24297	0.007662	0.01339	0.01329	0.35556	0.2556
<i>Hallopora</i> (AWThEx)	9	23	0.1864	0.179	0.031724	0.086003	0.0296	0.1648	0.05989	0.04217
<i>Hallopora</i> (BchD)	9	23	0.9721	0.8158	0.28221	0.93246	0.9849	0.8998	5.6756	5.7557
<i>Hallopora</i> (MD)	9	23	0.9041	0.05718	0.47178	0.0018407	0.01339	0.002971	0.13778	0.0813
<i>Hallopora</i> (N°A1mm ²)	9	23	0.02763	3.529E-05	2.8904E-05	0.025826	5.665E-05	0.0003645	4.4311	12.17
<i>Hallopora</i> (N°MDph1mm)	9	23	0.1983	0.001328	0.61405	0.35048	0.02743	0.2144	14.909	16.849
<i>Heterotrypa</i> (AAZS)	2	3	1	0.6369	0.18996	0.63328	0.7796	1	83.5	80.667
<i>Heterotrypa</i> (AD)	2	3	1	0	0.69069	0.78878	0.7796	0.7	0.255	0.25
<i>Heterotrypa</i> (AthD)	2	3	1	0.6369	0.21773	0.24009	0.4249	0.4	0.08	0.0433
<i>Heterotrypa</i> (AWThEx)	2	3	1	1	0.14305	0.11073	0.06267	0.2	0.075	0.03
<i>Heterotrypa</i> (BchD)	2	3	1	0.6369	0.13524	0.45034	0.7796	0.8	4.94	6.3333
<i>Heterotrypa</i> (EndozoneD)	2	3	1	1	0.069273	0.15405	0.06267	0.2	2.395	3.75
<i>Heterotrypa</i> (ExozoneTh)	2	3	1	1	0.0005946	0.55962	0.7796	1	1.52	1.12
<i>Heterotrypa</i> (MD)	2	3	1	0.2983	0.031105	0.41095	0.4249	0.5	0.1005	0.1233
<i>Heterotrypa</i> (N°A1mm ²)	2	3	1	1	0.14019	0.19187	0.06267	0.2	9.15	20.75
<i>Heterotrypa</i> (N°A1mm)	2	3	1	0.9921	0.57848	0.072393	0.06267	0.2	2.41	3.8833

no way to separate the provinces. The genus *Hallopora* shows the highest degree of overlap (Fig. 6), an unexpected result since *Hallopora* is one of the genera whose subsamples are most easily separable using the univariate analysis (see Table 2). In order to see if the results for *Hallopora* could be improved, we repeated the multivariate analysis using only those characters that gave positive results in the univariate analysis (five of the seven characters: Table 2). The results did not change and the convex hulls of the provinces were still superimposed.

On the other hand, the multivariate analyses gave better results for *Ceramoporella*, *Diplotrypa*, *Graptodictya* and *Trematopora*. Species of the genus *Ceramoporella* (Fig. 7) are clearly clustered by palaeogeographical province, and the convex hull of each province does not overlap with

other hulls in any of the multivariate techniques used. The species *C. grandis* (Ernst & Key, 2007) from the Mediterranean Province and *C. interporosa* (Ulrich, 1893) from the Laurentian-Siberian Province are the most distant from the centres of their respective provinces. Species of the genus *Diplotrypa* (Fig. 8) are also clustered by province, but in this case with a slight overlap between the Baltic and Laurentian-Siberian provinces (*D. moniliformis*, defined by Bassler, 1911 from the Baltic Province, plots inside the convex hull of the Laurentian-Siberian species); also *Diplotrypa* cf. *westoni* (Maw *et al.*, 1976) from Burma (whose inclusion in the Mediterranean Province is uncertain) plots very close to the Baltic Province polygon. The positions of different species of *Diplotrypa* seem to have no correlation with temperature gradients.

Table 2. continued

Genus	No. of samples		Normality test		F test	t test	K-S test	M-W test	Mean	
	Cold	Warm	Cold	Warm					Cold	Warm
<i>Kukersella</i> (AD)	2	10	1	0.03247	0.97811	0.84363	0.3959	0.6667	0.13	0.136
<i>Kukersella</i> (ADalWTh)	2	10	1	0.05002	0.23652	0.0092321	0.05321	0.09091	0.021	0.0411
<i>Kukersella</i> (AltPxWTh)	2	10	1	0.08975	0.48578	0.013811	0.05321	0.0303	0.0105	0.036
<i>Kukersella</i> (BchD)	2	10	1	0.8052	0.22428	0.13932	0.1139	0.2727	0.545	0.833
<i>Kukersella</i> (EndozoneD)	2	10	1	0.4946	0.0071644	0.16992	0.1139	0.2576	0.2005	0.367
<i>Kukersella</i> (ExozoneTh)	2	10	1	0.0276	0.019733	0.3122	0.2227	0.4091	0.1605	0.204
<i>Monticulipora</i> (AD)	2	19	1	0.001698	0.21754	0.0017515	0.02726	0.009524	0.355	0.2207
<i>Monticulipora</i> (MD)	2	19	1	0.1032	0.81122	0.032632	0.06714	0.06667	0.125	0.0826
<i>Monticulipora</i> (N°A1mm ²)	2	19	1	0.9662	0.226	0.090286	0.04335	0.05714	8.35	19.126
<i>Monticulipora</i> (N°A1mm)	2	19	1	0.0003672	0.264	0.045328	0.01669	0.009524	2.75	4.702
<i>Monticulipora</i> (N°M1mm ²)	2	19	1	0.0004178	0.37746	0.41612	0.7899	0.5143	4.6	13.039
<i>Monticulipora</i> (N°M1mm)	2	19	1	3.399E-05	0.12149	0.78268	0.3988	0.6762	1.05	1.235
<i>Monticulipora</i> (ZWTh)	2	19	1	0.01334	0.95755	0.41605	0.2954	0.5905	0.015	0.0215
<i>Trematopora</i> (AD)	8	4	0.005044	0.4043	0.65231	0.032954	0.1502	0.04848	0.11125	0.1725
<i>Trematopora</i> (AthD)	8	4	0.6257	0.4877	0.49354	0.057387	0.3788	0.1212	0.05125	0.025
<i>Trematopora</i> (AWThEx)	8	4	0.004482	0.6806	0.0054533	0.15	0.04809	0.02222	0.116	0.02225
<i>Trematopora</i> (BchD)	8	4	0.6373	0.7679	0.27576	0.79224	0.7399	0.6828	2.3075	2.4675
<i>Trematopora</i> (MD)	8	4	0.425	0.85	0.91122	0.42582	0.9857	0.5333	0.06875	0.0775
<i>Trematopora</i> (N°A1mm ²)	8	4	0.0319	0.3207	0.031865	0.68906	0.3788	0.3394	12.582	11.54
<i>Trematopora</i> (N°A1mm)	8	4	0.3947	0.8862	0.13548	0.36096	0.3788	0.6828	3.8687	2.675
<i>Trematopora</i> (N°Ath1mm ²)	8	4	0.00955	0.008004	0.46253	0.45525	0.04809	0.1414	32.683	19.375
All genera (AD)	45	109	0.0007871	1.835E-05	4.3255E-06	0.062164	0.0449	0.6518	0.28578	0.24233

The nine species of the genus *Graptodictya* (Fig. 9) cluster around two clearly differentiated points: one group includes Laurentian-Siberian species and the other the Mediterranean Province, although in this group the species *G. vinassae*, described by Conti (1990) from Sardinia, plots a large distance from the other Mediterranean Province species. The only *Graptodictya* species from the Baltic Province plots far from both the Mediterranean and Laurentia-Siberia provinces, precluding this genus from defining a clear temperature gradient.

Lastly, species of the genus *Trematopora* (Fig. 10) form two clusters, the larger of which is composed by the species from the Mediterranean Province, with *T. acanthostylita* Jiménez-Sánchez, 2009 being the species most distant from the centre of the cluster. The smaller cluster groups species from the Baltic Province. In two of the multivariate analyses (Correspondence and Detrended Correspondence), *Trematopora? promigenia* (Ulrich, 1893) from the Laurentian-Siberian Province plots inside the convex hull of the Mediterranean Province. As in previous analyses, no clear temperature gradient is apparent.

Conclusions

Ten different eurythermic bryozoan genera from the Upper Ordovician have been studied in order to test whether mor-

phological differences exist between cold- and warm-water species. The study was carried out on 154 samples belonging to 104 different species. Twenty-eight morphological characters were measured, although only 21 were used in the final statistical analysis. For the univariate and discriminant analysis, the total sample of species was divided into cold- and warm-water subsamples: species from Mediterranean Province were attributed to the cold-water subsample, whereas species from Avalonia, Baltic and Laurentian-Siberian provinces were included in the warm-water subsample. No *a priori* grouping by water temperature was imposed in the multivariate ordination analysis performed subsequently, where the aim was to test whether different species, colour coded by palaeogeographical province, were correctly arranged along a water temperature gradient.

Statistical analyses allowed the following conclusions to be drawn:

1. The univariate statistical analysis showed that there are clear morphological differences between cold- and warm-water species (*i.e.*, from high and low latitudes, respectively) in more than half of the Upper Ordovician bryozoan genera analysed in this study (*Ceramoporella*, *Diplotrypa*, *Eridotrypa*, *Hallopora*, *Monticulipora* and *Trematopora*), although these differences are only evident for some of the 21 characters used, and only when the analysis is

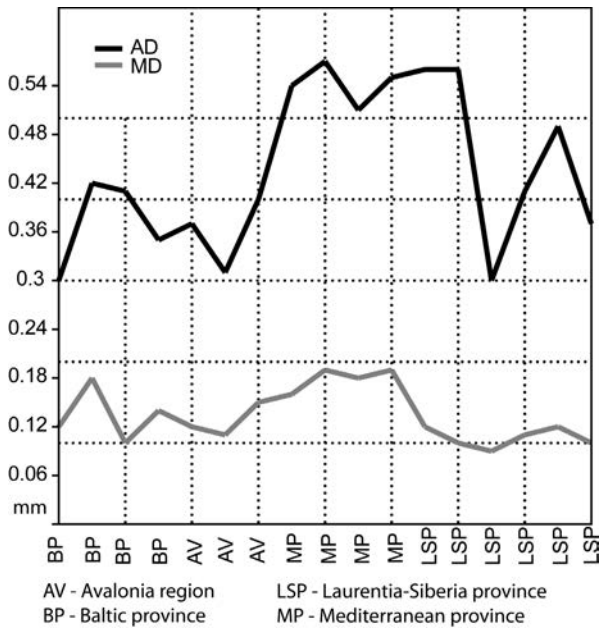


Figure 2. Relationship between autozooeical diameter (AD) and mesozooeical diameter (MD) in the genus *Diplotrypa*. It can be observed that larger diameters, both in autozooeica and mesozooeica, belong to species coming from the Mediterranean Province (proxy for cold-water).

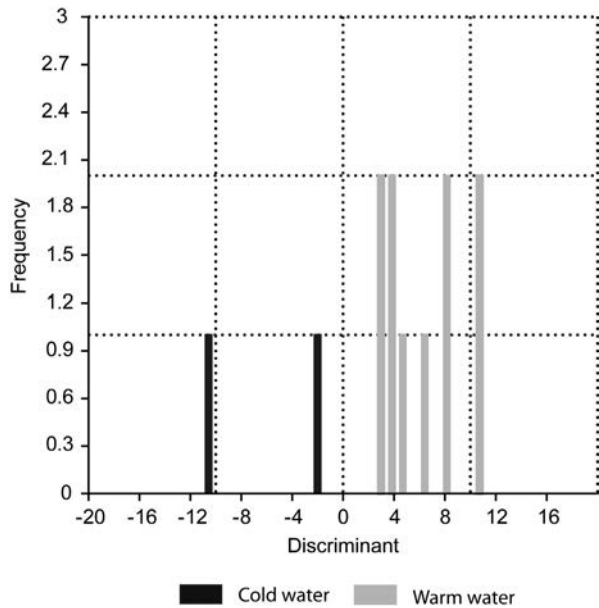


Figure 3. Discriminant analysis of the genus *Kukersella* with the subsets from cold- and warm-water clearly separated. The discriminant function separates correctly 100% of the samples. The discriminant function is: $-88.594 \times AD + 245.49 \times ALtPxWTh + 148.1 \times ADalWTh - 3.8485 \times EndozoneD + 64.107 \times ExozoneTh + 27.09 \times BchD$.

performed on individual genera. When all 10 genera were pooled together no statistically significant differences between warm- and cold-water species could be found.

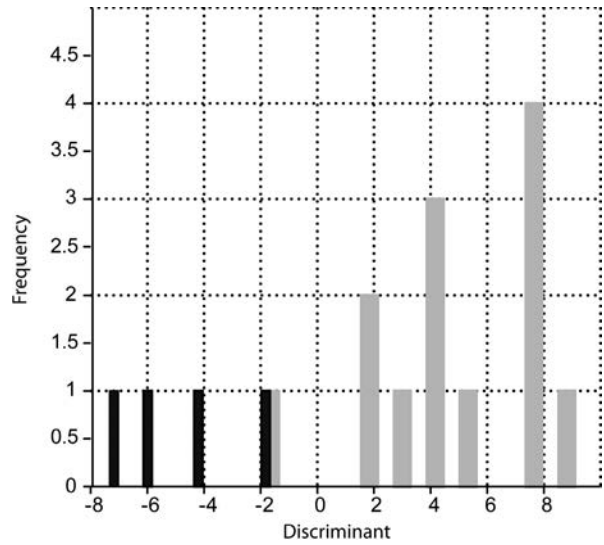


Figure 4. Discriminant analysis of the samples belonging to the genus *Diplotrypa*. In this case the percentage of samples correctly discriminated is slightly less than 95%; one sample of *D. moniliformis* Bassler, 1911 was placed in the field of cold-water samples.

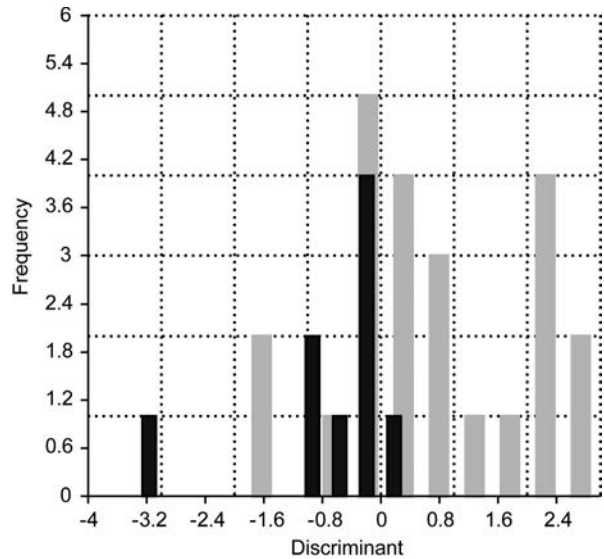


Figure 5. Discriminant analysis of the genus *Hallopora* showing a case of poor discrimination between cold- and warm-water subsets.

2. Genera from the order Trepotomata show the greatest temperature-related morphological differences.

3. The best characters to differentiate species by water temperature are those related to the size of the zooidal polymorphs, especially the diameters of the autozooeica, mesozooeica and exilazooeica. With the exception of one genus (*Trematopora*), cold-water species have larger zooids. This finding is consistent with the inverse correlation between temperature and zooid size among Recent cheilostome bryozoans, although this relationship has been best established between zooids

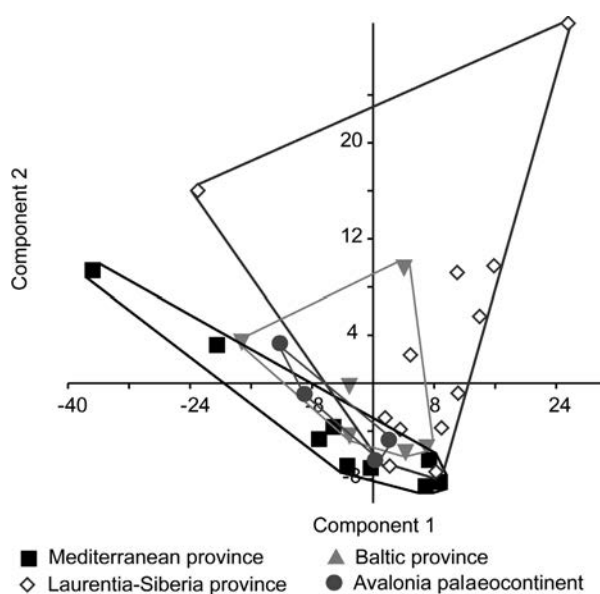


Figure 6. Principal Component analysis of the samples belonging to the genus *Hallopora*. The high degree of overlap between provinces does not allow differentiation of the bryozoan provinces present in the Upper Ordovician.

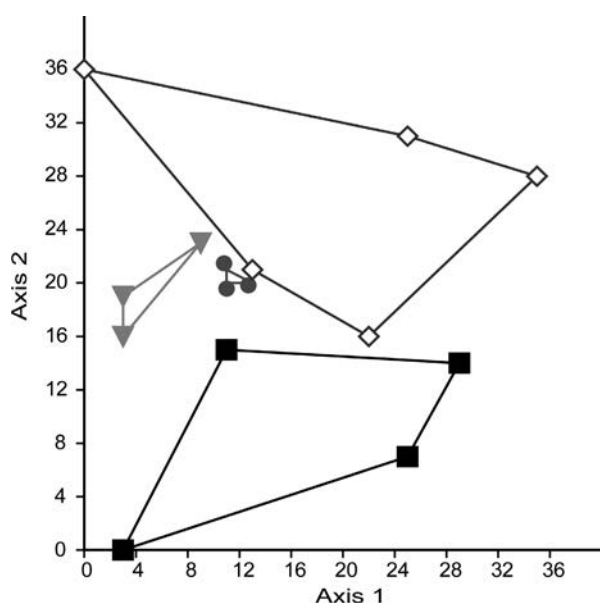


Figure 7. Detrended Correspondence analysis of the species belonging to the genus *Ceramoporella*. For this genus the different provinces are clearly differentiated, with no overlap between provinces.

within colonies and between colonies of the same species rather than between species within genera as here (Kuklinski & Taylor 2008).

4. The discriminant analysis showed the success rate of the discriminant function to be 100% for two genera (*Heterotrypa* and *Kukersella*), slightly below 95% for two other genera, and between 67% and 90% for the remaining

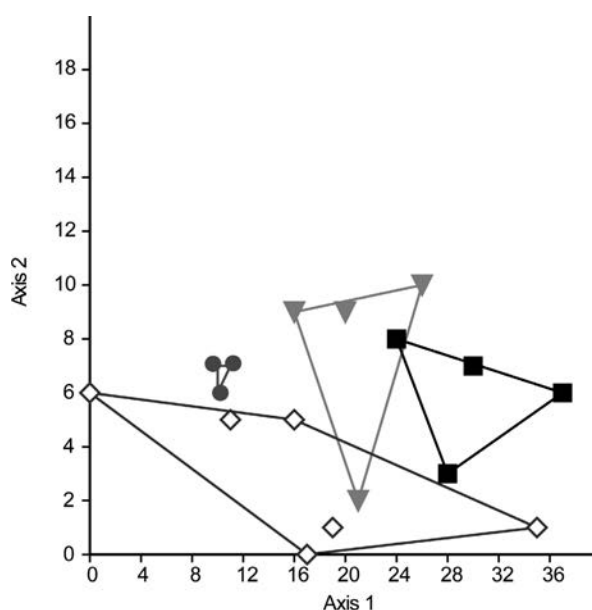


Figure 8. Detrended Correspondence analysis for species of the genus *Diplotrypa*. The samples belonging to this genus are well separated according to province, but there is a slight overlap between three provinces.

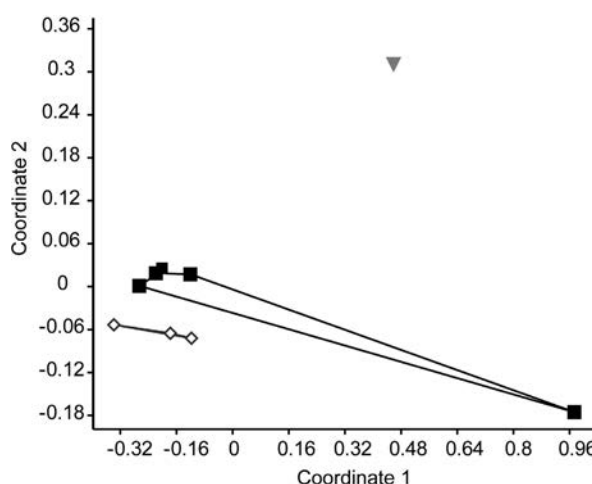


Figure 9. Principal Coordinate analysis of the genus *Craptodiptya*. The few samples analysed of this genus are correctly arranged by provinces, but the sample belonging to *G. vinassae* plots far away from the rest of the Mediterranean samples and the only species of the Baltic Province plots a large distance from both the Mediterranean and Laurentia-Siberia provinces.

six genera. This means that a robust classification of a new species of a specific genus as warm- or cold-water is not possible in most cases.

5. The multivariate ordination analysis was able to separate species by palaeogeographical province in some genera, but these provinces were not arranged along a correct temperature gradient using any of the methods tested (Principal Coordinates, Principal Components, Correspondence, and Detrended Correspondence analyses). This lack of correlation between temperature gradient and bryozoans provinces may

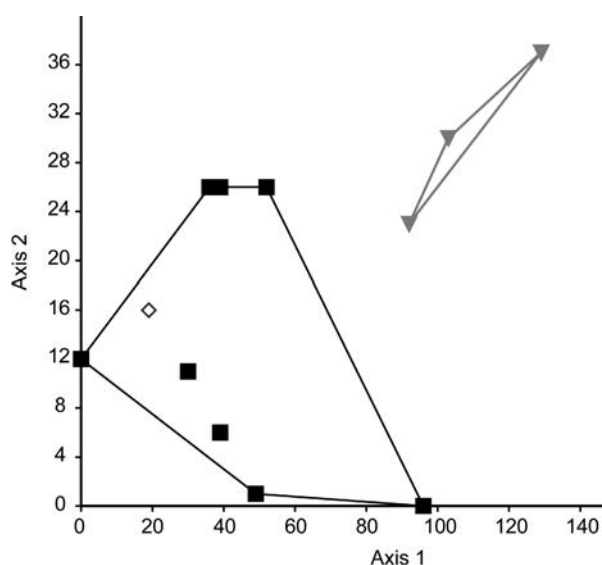


Figure 10. Detrended Correspondence analysis of the genus *Trematopora*. There is a good separation between the Baltic and the Mediterranean provinces. The only species belonging to the Laurentia-Siberian Province plots inside the convex hull of the Mediterranean Province.

be due to the increase in water temperature (Boda event) prior to the Hirnantian glaciation (Fortey & Cocks 2005).

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Bibliography

BASSLER, R.S. 1911. The early Paleozoic Bryozoa of the Baltic provinces. *United States National Museum Bulletin* 77, 1–382.

BUTTLE, C.J. 1991. A new upper Ordovician bryozoan fauna from the Slade and Redhill Beds, South Wales. *Palaeontology* 34, 77–108.

COCKS, L.R.M. & TORSVIK, T.H. 2006. European geography in a global context from the Vendiant to the end of the Palaeozoic. *Journal of the Geological Society of London* 32, 83–95. DOI 10.1144/GSL.MEM.2006.032.01.05

CONTI, S. 1990. Upper Ordovician Bryozoa from Sardinia. *Palaeontographia Italica* 77, 85–165.

ERNST, A. & KEY, M. 2007. Upper Ordovician Bryozoa from the Montagne de Noire, Southern France. *Journal of Systematic Palaeontology* 5, 359–428. DOI 10.1017/S1477201907002155

FORTEY, R.A. & COCKS, L.R.M. 2003. Palaeontological evidence bearing on global Ordovician-Silurian continental reconstruc-

tions. *Earth-Science Reviews* 61, 245–307. DOI 10.1016/S0012-8252(02)00115-0

FORTEY, R.A. & COCKS, L.R.M. 2005. Late Ordovician global warming – the Boda event. *Geology* 33, 405–408. DOI 10.1130/G21180.1

HAMMER, Ø., HARPER, D.A.T. & RYAN, P.D. 2008. *PAST v 2.15*. Paleontological Statistics Software: Package for Education and Data analysis.

HUGHES, D.J. & JACKSON, J.B.C. 1990. Do constant environments promote complexity of form?: The distribution of bryozoan polymorphism as a test of hypotheses. *Evolution* 44, 889–905. DOI 10.2307/2409553

JIMÉNEZ-SÁNCHEZ, A. 2009. The upper Katian (Ordovician) bryozoans from the Eastern Iberian Chains (NE Spain). *Bulletin of Geosciences* 84, 687–738. DOI 10.3140/bull.geosci.1156

JIMÉNEZ-SÁNCHEZ, A. 2010. New Monticuliporidae (Trepotomata) from the Cystoid Limestone Formation (Upper Ordovician) of the Iberian Chains (NE Spain). *Geodiversitas* 32(2), 177–199. DOI 10.5252/g2010n2a1

JIMÉNEZ-SÁNCHEZ, A. & VILLAS, E. 2010. The bryozoan dispersion into the Mediterranean margin of Gondwana during the pre-glacial Late Ordovician. *Palaeogeography, Palaeoclimatology, Palaeoecology* 294, 220–231. DOI 10.1016/j.palaeo.2009.11.027

KUKLINSKI, P. & TAYLOR, P.D. 2008. Are bryozoans adapted for living in the Arctic? *Virginia Museum of Natural History, Special Publication* 15, 101–110.

LOXTON, J., KUKLINSKI, P., MAIR, J.M., JONES, M.S. & PORTER, J.S. 2012. Patterns of magnesium-calcite distribution in the skeleton of some polar bryozoan species, 169–185. In ERNST, A., SCHÄFER, P. & SCHOLZ, J. (eds) *Bryozoan Studies*. Springer, Heidelberg.

MAW, U.B., SAN, U.B. ROSS, J.R.P. & CIOCHON, R.L. 1976. The Ordovician bryozoan (Ectoproct) *Diplotrypa* from Central Burma. *Geological Magazine* 113, 515–518. DOI 10.1017/S001675680004125X

OKAMURA, B., O’DEA, A. & KNOWLES, T. 2011. Bryozoan growth and environmental reconstruction by zooid size variation. *Marine Ecology Progress Series* 430, 133–146. DOI 10.3354/meps08965

SCHÄFER, P. & BADER, B. 2008. Geochemical composition and variability in the skeleton of the bryozoan *Cellaria sinuosa* (Hassall): biological versus environmental control. *Virginia Museum of Natural History, Special Publication* 15, 269–278.

SCHLAGER, W. 2005. *Carbonate sedimentology and sequence stratigraphy*. 200 pp. SEPM, Tulsa. DOI 10.2110/csp.05.08

SMITH, A.M., KEY, M.M. JR. & GORDON, D.P. 2006. Skeletal mineralogy of bryozoans: taxonomic and temporal patterns. *Earth-Science Reviews* 78, 287–306. DOI 10.1016/j.earscirev.2006.06.001

TAYLOR, P.D. & ALLISON, P.A. 1998. Bryozoan carbonates through time and space. *Geology* 26, 459–462. DOI 10.1130/0091-7613(1998)026<0459:BCTTAS>2.3.CO;2

TAYLOR, P.D., JAMES, N.P., BONE, Y., KUKLINSKI, P. & KYSER, T.K. 2009. Evolving mineralogy of cheilostome bryozoans. *Palaios* 24, 440–452. DOI 10.2110/palo.2008.p08-124r

TAYLOR, P.D. & SENDINO, C. 2010. Latitudinal distribution of bryozoan-rich sediments in the Ordovician. *Bulletin of Geosciences* 85, 565–572. DOI 10.3140/bull.geosci.1177

ULRICH, E.O. 1893. On Lower Silurian Bryozoa of Minnesota. *Minnesota Geology and Natural History Survey, Final Report* 3(1), 96–332.