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Genes From a Translational Analysis Support a Multifactorial Nature of White Matter Hyperintensities

Lorna M. Lopez, PhD; W. David Hill, MSc; Sarah E. Harris, PhD; Maria Valdes Hernandez, PhD; Susana Munoz Maniega, PhD; Mark E. Bastin, PhD; Emma Bailey, PhD; Colin Smith, MD; Martin McBride, PhD; John McClure, PhD; Delyth Graham, PhD; Anna Dominiczak, MD; Qiong Yang, PhD; Myriam Fornage, PhD; M. Arfan Ikram, MD, PhD; Stephanie Debette, MD, PhD; Lenore Launer, PhD; Joshua C. Bis, PhD; Reinhold Schmidt, MD; Sudha Seshadri, MD; David J. Porteous, PhD; John Starr, MD; Ian J. Deary, PhD; Joanna M. Wardlaw, MD

- *Background and Purpose*—White matter hyperintensities (WMH) of presumed vascular origin increase the risk of stroke and dementia. Despite strong WMH heritability, few gene associations have been identified. Relevant experimental models may be informative.
- *Methods*—We tested the associations between genes that were differentially expressed in brains of young spontaneously hypertensive stroke–prone rats and human WMH (using volume and visual score) in 621 subjects from the Lothian Birth Cohort 1936 (LBC1936). We then attempted replication in 9361 subjects from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE). We also tested the subjects from LBC1936 for previous genome-wide WMH associations found in subjects from CHARGE.
- *Results*—Of 126 spontaneously hypertensive stroke–prone rat genes, 10 were nominally associated with WMH volume or score in subjects from LBC1936, of which 5 (*AFP*, *ALB*, *GNA11*, *RBM8a*, and *MRPL18*) were associated with both WMH volume and score (*P*<0.05); 2 of the 10 (*XPNPEP1*, *P*=6.7×10⁻⁵; *FARP1*, *P*=0.024) plus another spontaneously hypertensive stroke–prone rat gene (*USMG5*, *P*=0.00014), on chromosomes 10, 13, and 10 respectively, were associated with WMH in subjects from CHARGE. Gene set enrichment showed significant associations for downregulated spontaneously hypertensive stroke–prone rat genes with WMH in humans. In subjects from LBC1936, we replicated CHARGE's genome-wide WMH associations on chromosomes 17 (*TRIM65* and *TRIM47*) and, for the first time, 1 (*PMF1*).
- *Conclusions*—Despite not passing multiple testing thresholds individually, these genes collectively are relevant to known WMH associations, proposed WMH mechanisms, or dementia: associations with Alzheimer's disease, late-life depression, ATP production, osmotic regulation, neurodevelopmental abnormalities, and cognitive impairment. If replicated further, they suggest a multifactorial nature for WMH and argue for more consideration of vascular contributions to dementia. (*Stroke*. 2015;46:00-00. DOI: 10.1161/STROKEAHA.114.007649.)

Key Words: genetics ■ humans ■ leukoencephalopathies ■ magnetic resonance imaging

Correspondence to Joanna M. Wardlaw, MD, Brain Research Imaging Centre, Division of Neuroimaging Sciences, University of Edinburgh, Bramwell Dott Bldg, Western General Hospital, Crewe Rd, Edinburgh, EH4 2XU, United Kingdom. E-mail joanna.wardlaw@ed.ac.uk

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From the Centre for Cognitive Ageing and Cognitive Epidemiology (L.M.L., M.V.H., S.M.M., M.E.B., J.S., I.J.D., J.M.W.), Division of Neuroimaging Sciences, Brain Research Imaging Centre, (M.V.H., S.M.M., M.E.B., J.M.W.) and Academic Neuropathology (C.S.), Centre for Clinical Brain Sciences, University of Edinburgh, Edinburgh, United Kingdom; Centre for Cognitive Ageing and Cognitive Epidemiology, Medical Genetics Section, University of Edinburgh Centre for Genomics and Experimental Medicine and MRC Institute of Genetics and Molecular Medicine, University of Edinburgh, United Kingdom; Centre for Cognitive Ageing and Cognitive Epidemiology, Medical Genetics Section, University of Edinburgh, Cunited Kingdom (W.D.H., S.E.H., D.J.P.); Department of Bioengineering, Imperial College London, London, United Kingdom (E.B.); BHF Glasgow Cardiovascular Research Centre, Institute of Cardiovascular and Medical Sciences, University of Glasgow, Glasgow, United Kingdom (M.M., J.M., D.G., A.D.); Department of Biostatistics, Boston University School of Public Health, MA (Q.Y.); The Framingham Heart Study, Boston, MA (Q.Y., S.S.); The Human Genetics Center and Institute of Molecular Medicine, The University of Texas Health Science Center, Houston (M.F.); Departments of Epidemiology, Radiology and Neurology, Erasmus Medical Center, Rotterdam, The Netherlands (M.A.I.); Netherlands Consortium for Healthy Aging, Leiden, The Netherlands (M.A.I.); 12 INSERM U740 (Paris 7 University) and U708 (Bordeaux University), Bordeaux, France (S.D.); Department of Neurology, Boston University School of Medicine, MA (S.D., S.S.); Laboratory of Epidemiology and Population Sciences, Intramural Research Program, National Institute on Aging, National Institutes of Health, Bethesda, MD (L.L.); Cardiovascular Health Research Unit, Department of Medicine, University of Washington, Seattle (J.C.B.); and Clinical Division of Neurogeriatrics, Department of Neurology, Medical University of Graz, Graz, Austria (R.S.).

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White matter hyperintensities (WMH) of presumed vascular origin, a major component of cerebral small vessel disease (SVD), double the risk of stroke and dementia.¹ Despite considerable societal effect, the causes of WMH and SVD are poorly understood.² Conventional vascular risk factors explain little of the WMH variance.³ Family studies,⁴ several rare monogenic SVD disorders,⁵ and epidemiology⁶ suggest that genetic predisposition is important.

Identification of genetic factors for SVD has been challenging. Several replicable single-nucleotide polymorphisms (SNPs) associated with WMH have been identified in 1 locus on chromosome 17q25,^{7.8} although the exact gene(s) and biological pathways to WMH are unclear. Few other replicable genes have been found in genome-wide association studies (GWAS),^{9,10} and little is known of their functional significance.

Experimental SVD models might provide insight into human SVD. The spontaneously hypertensive stroke-prone rat (SHRSP) is a relevant model of spontaneous SVD.¹¹ It was selectively crossbred (1974) from Wistar-Kyoto (WKY) rats via the spontaneously hypertensive rat (SHR, 1963).¹² Hypertension, established in SHRSP rats by 10 weeks of age, is considered to be the main cause of their brain disease. However, differences in protein and gene expression in SHRSP rats versus WKY rats at 5 weeks of age (before measurable blood pressure rises) suggest underlying susceptibilities to SVD.13 Compared with WKY controls, 5-week-old SHRSP rats have reduced claudin 5 (tight junction) and myelin basic protein and increased microglia (IBA1) and glial activation (GFAP)¹³; at 16 and 21 weeks, increase in smooth muscle actin was seen, thought to reflect arteriolar smooth muscle hyperplasia secondary to hypertension. SHRSP gene expression differences at 5 weeks of age were more numerous than at 16 or 21 weeks of age and included downregulation of Mmp14, Mbp, GFAP, AVP, Alb, and Igf2, upregulation of Gucy1A3, Rps9, Fos, and JunB, early-growth response, cell-signaling genes,

and overexpression of genes involved in neurological diseases (stroke, depression, and blood–brain barrier leakage),¹⁴ rather than just hypertension. Recent gene sequencing of SHRSP rats (and 26 other rat models of common human diseases)¹⁵ revealed that genes that were either shared between or uniquely mutated in these rat models were significantly over-represented in human GWAS hits for hypertension or metabolism-related phenotypes, suggesting coevolution of these genes and their role in common diseases in models and humans.¹⁵

In a hypothesis-driven collaborative approach, we tested for associations between genes that were differentially expressed in the brains of 5-week-old SHRSP rats14 and WMH in humans. We used data from 5-week-old rats because gene expression differences were more frequent at that age than at 16 or 21 weeks, and we wanted to minimize the confounding of tissue changes by secondary effects of hypertension and to optimize the chances of detecting genes related to WMH susceptibility. We focused on WMH as the most frequent feature of SVD with the most data available in replication cohorts. We first tested the subjects from Lothian Birth Cohort 1936 (LBC1936)^{16,17} and then attempted replication in subjects from the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium.7 To provide confidence in the relevance of subjects from LBC1936, we also sought CHARGE's⁷ previously reported WMH-gene associations in the subjects from LBC1936.

Subjects

The subjects from LBC1936 are community-dwelling individuals living in South East Scotland who underwent detailed cognitive, biomedical, genetic assessments, and detailed brain MRI at \approx 73 years of age (n=866).^{16,17} The MRI acquisition, methods for assessing WMH burden¹⁷ qualitatively¹⁸ and quantitatively,¹⁹ and proportions with WMH by either method²⁰ have been reported. This study was approved by the Lothian (REC 07/MRE00/58) and Scottish Multicentre

Methods

 Table 1.
 Genes Associated With Cerebral Small Vessel Disease in Rats That Are Associated With WMH in Older Humans: 126

 Differentially Expressed Genes Between Spontaneously Hypertensive Stroke Prone and Wild-Type Rats Were Tested for Association

 With WMH in Subjects From LBC1936 and 10 Genes Were Significantly Associated (P<0.05) With Either WMH Volume or Fazekas Score</td>

				Discovery: LBC1936							Replication: CHARGE	
					Genotyped SNPs			Imputed SNPs	Imputed SNPs			
					WMH Volume	Fazekas Score		WMH Volume	Fazekas Score		WMH Volume	
Chromosome	Gene	Start Position	Stop Position	nSNPs	P Value	P Value	nSNPs	P Value	P Value	nSNPs	P Value	
4	AFP	74 520 796	74 540 356	13	0.0021	0.00090	77	0.0037	0.0037	67	0.841	
4	ALB	74 488 869	74 505 834	11	0.0026	0.0017	61	0.0063	0.0068	53	0.718	
7	GNAI1	79602075	79686661	42	0.034	0.033	181	0.014	0.015	166	0.767	
1	RBM8A	144218994	144 222 801	13	0.038	0.057	26	0.029	0.024	21	0.539	
2	INPP5D	233633279	233 781 288	69	0.041	0.78	198	0.044	0.87	162	0.989	
10	XPNPEP1	111614513	111673192	18	0.042	0.14	130	0.15	0.23	120	6.7×10⁻⁵	
9	NR4A3	101 623 957	101 668 994	13	0.045	0.16	62	0.11	0.25	56	0.484	
13	FARP1	97 593 434	97 900 024	154	0.049	0.25	550	0.18	0.51	468	0.024	
6	MRPL18	160 131 481	160 139 451	24	0.059	0.039	89	0.16	0.048	76	0.224	
1	SIPA1L2	230 600 334	230717866	80	0.087	0.0093	340	0.20	0.018	285	0.885	

nSNPs is the number of SNPs considered in the gene test. CHARGE indicates Cohorts for Heart and Aging Research in Genomic Epidemiology; LBC1936, Lothian Birth Cohort 1936; SNP, single-nucleotide polymorphism; and WMH, white matter hyperintensities.

(MREC/01/0/56) Research Ethics Committees; all subjects gave written informed consent.

The subjects from LBC1936 had genome-wide SNP data on 542050 SNPs,²¹ imputed to 2.5 million SNPs with HapMap2.²² There were 621 participants (392 men) from LBC1936 with both MRI and genetic data (mean age, 72.67 years; SD=0.73 years; Table I and Methods in the online-only Data Supplement). We excluded 48 subjects from LBC1936 with a history of stroke or dementia.

Gene Analysis

In the 5-week-old SHRSP rats, 162 genes were differentially expressed compared with 5-week-old WKY rats in frontal and midcoronal brain sections (Table II in the online-only Data Supplement).¹⁴ We used the following databases to match the SHRSP Illumina IDs to human genes (Materials and Table II in the online-only Data Supplement): Ensembl-http://www.ensembl.org, GeneCardshttp://www.genecards.org, Illumina ID search-http://www.genscript.com, NCBI-http://www.ncbi.nlm.nih.gov, and Rat Genome Database-http://www.rgd.mcw.edu. Of the 162 SHRSP genes, 132 had an equivalent human gene, 8 transcripts were mapped to the same gene, 20 were uncharacterized in humans, and 2 had no human homologue. Of the 132 genes, 126 were available for association testing using the Versatile Gene-based Association Study (VEGAS) test.23 We first performed a genome-wide association analysis on subjects from LBC1936 using PLINK software²⁴ to test the genetic association between 542050 genotyped SNPs and 2 WMH measurements using a linear regression analysis: (1) log transformed WMH volume (mL), with age, sex, intracranial volume, and first 4 multiple dimension scaling components for population stratification as covariates; and (2) summed Fazekas score of periventricular and deep WMH, with age, sex, and the first 4 multiple dimension scaling population stratification components as covariates. We used both WMH volume and Fazekas score20 to increase the reliability of the results. We did not stratify by vascular risk factors because hypertension (although it was the strongest vascular risk factor) explained <2% of WMH variance in subjects from LBC1936.3 The VEGAS software summarized evidence for association with WMH in subjects from LBC1936 per gene by considering the P values of all 543050 SNPs that were located within 17681 unique autosomal genes (including SNPs±50 kb outside of genes to include regulatory regions). For a more direct comparison with CHARGE (which used imputed data), we also performed a gene-based test on LBC1936's 2447 226 HapMap2 derived *P* values (after removing SNPs with a minor allele frequency of <0.01 and imputation quality of <0.3) with VEGAS software as above.

Replication in Subjects From CHARGE

We then tested whether any of the 126 SHRSP genes were also associated with WMH in subjects from CHARGE by using data from CHARGE's published genome-wide meta-analysis of WMH in 9361 stroke-free individuals from 7 community-based cohorts.⁷ We performed a gene-based test using VEGAS software, which summarized the evidence for association with WMH burden on a per gene basis, as above, by considering the associated *P* values of all HapMap2 SNPs located within 17787 autosomal genes (including SNPs±50 kb outside of genes to include regulatory regions).

Gene Set Enrichment

We performed a gene set enrichment analysis²⁵ to investigate the enrichment of the 126 SHRSP genes in the LBC1936 and CHARGE data associated with WMH, accounting for whether these were upregulated or downregulated (online-only Data Supplement),²⁶ corrected for multiple testing using a false discovery rate (FDR) method.²⁷

Replication of Previous CHARGE Findings in Subjects From LBC1936

To demonstrate our ability to detect WMH-gene associations in subjects from LBC1936, we attempted replication of CHARGE's

genome-wide associations with WMH^{7,8} in the subjects from the LBC1936 Cohort in a genome-wide association analysis using the 2534887 SNPs imputed to HapMap2, with WMH (volume and Fazekas score) in Mach2QTL software.²⁸

We applied Bonferroni correction for multiple testing (P=0.05/126 genes=0.0004). We did not include the 2 WMH phenotypes in the Bonferroni correction as they are highly correlated (r^2 =0.77). Because of the overconservative nature of Bonferroni correction for multiple testing,²⁹ a nominal significance threshold of *P* value of <0.05 was required for replication efforts.

Results

SHRSP Genes in Subjects From LBC1936

Of the 126 candidate SHRSP-derived genes, 10 were nominally associated with WMH in subjects from LBC1936 (*P*<0.05; Table 1). Using imputed or genotyped data, 5 genes were associated with WMH volume (*AFP*, *ALB*, *GNA11* [*RBM8A* and *INPP5D*, both borderline]); 3 of these (*AFP*, *ALB*, and *GNA11*) and 2 others (*MRPL18* and *SIPA1L2*) were associated with WMH Fazekas scores. Three other genes were associated with WMH volume using genotyped data only (*XNXPEP1*, *NR4A3*, and *FARP1*). None of these genes individually passed Bonferroni correction in subjects from LBC1936 (all were *P*>0.0004), in part, reflecting the LBC1936 sample size.

SHRSP Genes in Subjects From CHARGE

Two of these 10 genes were also associated with WMH in subjects from CHARGE (*XPNPEP1*, $P=6.7\times10^{-5}$; and *FARP1*, P=0.024; Table 1). Full details of all 126 SHRSP to LBC1936 to CHARGE gene associations are given in Table III in the online-only Data Supplement. Several other of the 126 SHRSP genes (outside the 10/126 described above) showed significance at P<0.05 in subjects from CHARGE (eg, *USMG5*, *MED17*, *ZNF461*, *C20orf7*, *EGR1*, *ARC*, *NUDT14*, and *MMP14*) of which 1 (*USMG5*, P<0.000142) passed Bonferroni correction (P<0.0004).

Gene Set Enrichment

Using gene set enrichment analysis, all 126 SHRSP candidate genes were not enriched in subjects from LBC1936 for association with WMH in the 17681 genes tested here (WMH volume, P=0.34; Fazekas score, P=0.81), but this would not preclude the possibility that in either upregulated or downregulated gene sets, there was an abundance of genes showing an enriched association. We tested the upregulated (n=76) and downregulated (n=50)SHRSP genes separately and found significant enrichment for Fazekas scores in SHRSP downregulated genes (P=0.035; FDR, 0.046) but not SHRSP upregulated genes (P=0.921; FDR, 0.899). WMH volume showed significant enrichment in downregulated (P=0.018; FDR, 0.025) but not upregulated (P=0.802; FDR, 0.780) genes. In the CHARGE consortium, there was no significant enrichment for either the total set of 126 genes (P=0.0514), the upregulated (P=0.109; FDR, 0.266) or the downregulated genes (P=0.173; FDR, 0.149).

Replication of CHARGE's Previous Genome-Wide Association in Subjects From LBC1936

We sought CHARGE's previous genome-wide association results for WMH⁷ in subjects from LBC1936. Of CHARGE's 15 SNPs ($P < 1 \times 10^{-5}$) associated with WMH (Table 2),⁷ 3 SNPs replicated in subjects from LBC1936 with both WMH volume and Fazekas score at P < 0.05 (rs3744028, rs1055129, and rs1052053); rs1052053, a miss-sense variant on chromosome 1 in the polyamine-modulated factor 1 gene (*PMF1*), has not replicated previously.

Discussion

We used a clinically relevant translational approach¹⁵ to identify potential new gene associations for WMH, a common cause of cognitive impairment, stroke, and dementia. We found parallels between differentially expressed genes in a young spontaneous SVD model and WMH-gene associations in older humans. Two novel genes on chromosome 10 derived from SHRSP rats were associated with WMH, XPNPEP1 in both LBC1936 and CHARGE and USMG5 in CHARGE only. Several other genes were nominally associated with WMH in LBC1936 or CHARGE although none passed multiple testing. We replicated 3 of CHARGE's WMH-gene associations in subjects from LBC1936: 2 (rs3744028 and rs1055129) on chromosome 17q25 and 1 previously unreplicated SNP (rs1052053) on chromosome 1, a miss-sense variant in the polyamine-modulated factor 1 gene, PMF1, that has a role in the cell cycle. Jointly, these approaches yielded 6 genes (3 from the SHRSP rats and 3 replicates of a GWAS finding) and 5 further rat-derived genes based on the LBC1936 sample alone, which despite not passing multiple testing thresholds individually, as a group they are notable for their involvement in biological pathways relevant to WMH pathogenesis.²

Of the 2 SHRSP genes found in LBC1936 and CHARGE, *XPNPEP1* is X-prolyl aminopeptidase (aminopeptidase P) 1, soluble, associated with biliary atresia, and located in a region on chromosome 10 that is associated with Alzheimer's disease.³⁰ FARP1 is Pleckstrin domain protein 1, associated with brain volume differences,³¹ and important in synapse development.32 The SHRSP-CHARGE-associated gene USMG5 is upregulated during skeletal muscle growth 5 homolog (also known as diabetes mellitus-associated protein in insulin sensitive tissues, or DAPIT), sits on chromosome 10, and maintains ATP synthase populations in mitochondria.³³ All 5 SHRSP genes associated with both WMH volume and Fazekas score in subjects from LBC1936 (AFP, ALB, GNAI1, RBM8A, and MRPL18) are associated with white matter-relevant diseases in humans. Despite not surviving correction for multiple testing, there was a notable consistency in their association with 2 separate WMH measures. AFP encodes α -fetoprotein, a major plasma protein produced in the yolk sac and liver during fetal life. Abnormally, high amounts of α -fetoprotein are found in ataxia telangiectasia,34 also associated with abnormal white matter.35 ALB encodes albumin, a soluble monomeric protein important for maintaining plasma oncotic pressure found in cerebral WMH,36 and cerebrospinal fluid as blood-brain barrier function deteriorates with ageing and dementia.^{2,37} GNAI1 encodes guanine nucleotide-binding protein (G protein), alpha-inhibiting activity polypeptide 1, implicated with Alzheimer's disease.³⁸ RBM8A is an RNA binding protein that has differential expression in Alzheimer's disease,³⁹ associations with a range of intellectual disabilities in humans and anxiety-related behavior in mice,40 with schizophrenia, several neurodevelopmental intellectual disabilities, anxiety behavior and may target neuronal genes to regulate behaviors. WMH in old age are known associates of late-onset depression,⁴¹ and they are also associated with lower age 11 IQ.42 MRPL18 is the mitochondrial ribosomal protein L18, previously associated

 Table 2.
 Association of SNPs Previously Associated With WMH in CHARGE in Subjects From LBC1936 and the corresponding SNP

 Association Results Are Given for LBC1936 WMH Volume and Fazekas Score

	3 0	JOURNAL		OFTHECHARGEERICA		s H	LBC1936	WMH Volume			Fazekas Score	
						Effect						
SNP	Chromosome	Nearest Gene	Risk Allele	Allele Freq	P Value	Allele	Allele Freq	<i>r</i> ²	β	P Value	β	<i>P</i> Value
rs3744028	17	TRIM65	С	0.18	4.0×10 ⁻⁹	Т	0.81	0.99	-0.217	0.00287	-0.287	0.000511
rs1055129	17	TRIM47	G	0.30	4.1×10 ⁻⁸	G	0.28	0.97	0.286	9.5×10 ⁻⁶	0.305	3.34×10 ⁻⁵
rs7894407	10	PDCD11	Т	0.63	6.1×10 ⁻⁷	Т	0.63	0.99	-0.026	0.662	-0.029	0.665
rs1892525	1	RP11-518D3.1	G	0.69	7.2×10 ⁻⁷	G	0.73	0.99	0.070	0.269	0.107	0.135
rs10814323	9	RECK	А	0.21	1.7×10 ⁻⁶	G	0.77	1.00	0.056	0.390	0.034	0.651
rs6992136	8	RPL32P19	G	0.85	3.2×10 ⁻⁶	G	0.85	0.81	0.101	0.259	0.075	0.458
rs11731436	4	AC097110.1	С	0.64	3.3×10 ⁻⁶	G	0.35	0.91	-0.035	0.565	-0.041	0.549
rs1052053	1	PMF1	А	0.62	5×10 ⁻⁶	G	0.39	1.00	-0.112	0.047	-0.127	0.048
rs2167089	3	AC098970.2	G	0.73	6×10 ⁻⁶	Т	0.26	0.97	0.061	0.342	0.044	0.545
rs10012573	4	COL25A1	А	0.94	6×10 ⁻⁶	С	0.06	0.85	0.097	0.481	0.029	0.855
rs11625623	14	PTGDR	G	0.23	7.7×10 ⁻⁶	G	0.23	1.00	-0.051	0.460	-0.021	0.792
rs16901064	5	RNASEN	С	0.84	7.8×10 ⁻⁶	С	0.85	0.99	0.030	0.695	0.055	0.532
rs6945846	7	FOXP2	С	0.2	7.9×10 ⁻⁶	Т	0.78	0.90	-0.036	0.625	0.113	0.175
rs11629135	14	MTHFD1	G	0.93	8.6×10 ⁻⁶	G	0.92	0.99	0.034	0.749	-0.057	0.641
rs9410016	9	C9orf62	G	0.41	9.7×10 ⁻⁶	G	0.39	0.99	-0.029	0.603	-0.041	0.521

Allele frequency is the frequency of the effect allele. r^2 is a measure of the imputation quality to HapMap2. β is the regression coefficient. CHARGE indicates Cohorts for Heart and Aging Research in Genomic Epidemiology; LBC1936, Lothian Birth Cohort 1936; SNP, single-nucleotide polymorphism; and WMH, white matter hyperintensities.

with multiple sclerosis.⁴³ These 7 SHRSP-derived genes are related to pathologies (ataxia telangiectasia, blood-brain barrier impairment, Alzheimer's disease, multiple sclerosis, depression, developmental intellectual disabilities, and brain size) that display white matter abnormalities or affect intellectual function. Impaired ATP production because of defects in *USMG5*, the gene that replicated from SHRSP to CHARGE, could increase susceptibility to WMH via ischemia.

The genes that were downregulated in the SHRSP were significantly enriched in subjects from LBC1936 for WMH. This may be because, in a complex disease such as SVD/WMH, several individually modest genetic defects in different components of key pathways, when present in combination, increase disease risk. This interpretation is consistent with differential protein expression seen in SHRSP¹³ and the absence, so far, of individual major human gene defects explaining either sporadic WMH or lacunar stroke.⁹

The lack of consistent replication from SHRSP to LBC1936 to CHARGE requires caution. The power and required significance threshold of the LBC1936 was modest for GWAS, hence our hypothesis-driven approach. Genes associated with WMH in subjects from LBC1936 but not CHARGE could be false positives; other factors include greater heterogeneity of WMH assessment and greater age range in subjects from CHARGE. The narrow age range of subjects from LBC1936 minimizes the effect of age, possibly helping to expose relevant genes. CHARGE-contributing studies used several methods of quantifying WMH, different MR scanner field strengths, and generations of technology and sequences. However, WMH volume and visual scores are highly correlated,20 and our replication of 3 findings from CHARGE in subjects from LBC1936 suggests that our approach has some validity. The CHARGE cohorts may have used different imputation platforms or more SNPS may have failed quality assurance in subjects from LBC1936, contributing to differences between the imputation results. There are several limitations to gene-based analysis, including the omission of nonautosomal genes, the effect of noncausal SNPs to dilute association (in particular, in the presence of a strong genetic association with a single locus within or in the regulatory region of a given gene, thus missing important associations), the lack of knowledge on (and overlap of) gene boundaries, the possibility that an SNP variant may influence a gene distal to its site, thus not corresponding to a gene that it is located next to it, and the potential of the genetic data not to tag causative genetic variants. Power may have been limited (despite CHARGE's large sample size) to detect associations with some genes. We did not stratify the human cohorts by risk factors as these explained <2% of WMH variance in subjects from LBC1936,³ and risk-stratified genetic data were unavailable for CHARGE. We did not test gene associations with other SVD features in addition to WMH because a total SVD burden score was not available for CHARGE. Although it is a relevant model of spontaneous SVD^{11,12} and of human hypertension and metabolic disorders,¹⁵ like any model, the SHRSP has translational limitations, arguing for additional studies at different ages and brain regions, with or without environmental stressors.

This work has the following strengths: accurate LBC1936 WMH phenotyping¹⁷ and genetic information in this relatively large narrow age-range older population.¹⁶ The Glasgow SHRSP colony is long established, with carefully controlled environments. The mRNA data were obtained from the same rats that provided protein expression data.¹³ Replication in other SHRSP colonies and examination of related strains (eg, SHR's) may be informative. The genomes of SHRSP and 26 other complex disease phenotype models were recently sequenced,¹⁵ showing associations between genes in rat models of hypertension and human GWAS hits for hypertension phenotypes.¹⁵ This provides support for our reverse-translational discovery approach, suggesting that genes in disease models have coevolved and may contribute to disease-related phenotypes in humans.

Our findings require validation. The selection of candidate genes for investigation could be widened by examining more genes from the 5-week-old SHRSP rats (Table II in the onlineonly Data Supplement), other models,¹⁵ and in larger samples of well-phenotyped humans, such as from METASTROKE and the Wellcome Trust Case-Control Consortium. This translational analysis of experimental models and human disease suggests some aspects of the genetic architecture underlying SVD, stroke, and dementia and argues for greater awareness of vascular contributions to neurodegeneration.

Figure I and Tables IV and V in the online-only Data Supplement provide the top SNP ($P < 1 \times 10^{-5}$) and gene (P < 0.001) associations with WMH variables in subjects from LBC1936 for further reference.

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Disclosures

None.

References

- Debette S, Markus HS. The clinical importance of white matter hyperintensities on brain magnetic resonance imaging: systematic review and meta-analysis. *BMJ*. 2010;341:c3666. doi: 10.1136/bmj.c3666.
- Wardlaw JM, Smith C, Dichgans M. Mechanisms of sporadic cerebral small vessel disease: insights from neuroimaging. *Lancet Neurol.* 2013;12:483–497. doi: 10.1016/S1474-4422(13)70060-7.

- Wardlaw JM, Allerhand M, Doubal FN, Valdes Hernandez M, Morris Z, Gow AJ, et al. Vascular risk factors, large artery atheroma, and brain white matter hyperintensities. *Neurology*. 2014;82:1331–1338. doi: 10.1212/WNL.00000000000312.
- Kochunov P, Glahn D, Winkler A, Duggirala R, Olvera RL, Cole S, et al. Analysis of genetic variability and whole genome linkage of wholebrain, subcortical, and ependymal hyperintense white matter volume. *Stroke*. 2009;40:3685–3690. doi: 10.1161/STROKEAHA.109.565390.
- Yamamoto Y, Craggs L, Baumann M, Kalimo H, Kalaria RN. Review: molecular genetics and pathology of hereditary small vessel diseases of the brain. *Neuropathol Appl Neurobiol*. 2011;37:94–113. doi: 10.1111/j.1365-2990.2010.01147.x.
- Jackson CA, Hutchison A, Dennis MS, Wardlaw JM, Lindgren A, Norrving B, et al. Differing risk factor profiles of ischemic stroke subtypes: evidence for a distinct lacunar arteriopathy? *Stroke*. 2010;41:624– 629. doi: 10.1161/STROKEAHA.109.558809.
- Fornage M, Debette S, Bis JC, Schmidt H, Ikram MA, Dufouil C, et al. Genome-wide association studies of cerebral white matter lesion burden: the CHARGE consortium. *Ann Neurol.* 2011;69:928–939. doi: 10.1002/ ana.22403.
- Verhaaren BF, de Boer R, Vernooij MW, Rivadeneira F, Uitterlinden AG, Hofman A, et al. Replication study of chr17q25 with cerebral white matter lesion volume. *Stroke*. 2011;42:3297–3299. doi: 10.1161/ STROKEAHA.111.623090.
- Traylor M, Farrall M, Holliday EG, Sudlow C, Hopewell JC, Cheng YC, et al; Australian Stroke Genetics Collaborative; Wellcome Trust Case Control Consortium 2 (WTCCC2); International Stroke Genetics Consortium. Genetic risk factors for ischaemic stroke and its subtypes (the METASTROKE collaboration): a meta-analysis of genome-wide association studies. *Lancet Neurol.* 2012;11:951–962. doi: 10.1016/ S1474-4422(12)70234-X.
- Adib-Samii P, Rost N, Traylor M, Devan W, Biffi A, Lanfranconi S, et al; Australian Stroke Genetics Collaborative; Wellcome Trust Case-Control Consortium-2 (WTCCC2); METASTROKE; International Stroke Genetics Consortium. 17q25 Locus is associated with white matter hyperintensity volume in ischemic stroke, but not with lacunar stroke status. *Stroke*. 2013;44:1609–1615. doi: 10.1161/STROKEAHA.113.679936.
- Bailey EL, McCulloch J, Sudlow C, Wardlaw JM. Potential animal models of lacunar stroke: a systematic review. *Stroke*. 2009;40:e451–e458. doi: 10.1161/STROKEAHA.108.528430.
- Bailey EL, Smith C, Sudlow CM, Wardlaw JM. Is the spontaneously hypertensive stroke prone rat a pertinent model of subcortical ischemic stroke? A systematic review. *Int J Stroke*. 2011;6:434–444. doi: 10.1111/j.1747-4949.2011.00659.x.
- Bailey EL, Wardlaw JM, Graham D, Dominiczak AF, Sudlow CL, Smith C. Cerebral small vessel endothelial structural changes predate hypertension in stroke-prone spontaneously hypertensive rats: a blinded, controlled immunohistochemical study of 5- to 21-week old rats. *Neuropathol Appl Neurobiol*. 2011;37:711–726. doi: 10.1111/j.1365-2990.2011.01170.x.
- Bailey EL, McBride MW, Beattie W, McClure JD, Graham D, Dominiczak AF, et al. Differential gene expression in multiple neurological, inflammatory and connective tissue pathways in a spontaneous model of human small vessel stroke. *Neuropathol Appl Neurobiol*. 2014;40:855–872. doi: 10.1111/nan.12116.
- Atanur SS, Diaz AG, Maratou K, Sarkis A, Rotival M, Game L, et al. Genome sequencing reveals loci under artificial selection that underlie disease phenotypes in the laboratory rat. *Cell*. 2013;154:691–703. doi: 10.1016/j.cell.2013.06.040.
- Deary IJ, Gow AJ, Taylor MD, Corley J, Brett C, Wilson V, et al. The Lothian Birth Cohort 1936: a study to examine influences on cognitive ageing from age 11 to age 70 and beyond. *BMC Geriatr*. 2007;7:28. doi: 10.1186/1471-2318-7-28.
- Wardlaw JM, Bastin ME, Valdés Hernández MC, Maniega SM, Royle NA, Morris Z, et al. Brain aging, cognition in youth and old age and vascular disease in the Lothian Birth Cohort 1936: rationale, design and methodology of the imaging protocol. *Int J Stroke*. 2011;6:547–559. doi: 10.1111/j.1747-4949.2011.00683.x.
- Fazekas F, Chawluk JB, Alavi A, Hurtig HI, Zimmerman RA. MR signal abnormalities at 1.5 T in Alzheimer's dementia and normal aging. *AJR Am J Roentgenol*. 1987;149:351–356. doi: 10.2214/ajr.149.2.351.
- Hernández Mdel C, Ferguson KJ, Chappell FM, Wardlaw JM. New multispectral MRI data fusion technique for white matter lesion segmentation: method and comparison with thresholding in FLAIR images. *Eur Radiol.* 2010;20:1684–1691. doi: 10.1007/s00330-010-1718-6.

- Valdés Hernández Mdel C, Morris Z, Dickie DA, Royle NA, Muñoz Maniega S, Aribisala BS, et al. Close correlation between quantitative and qualitative assessments of white matter lesions. *Neuroepidemiology*. 2013;40:13–22. doi: 10.1159/000341859.
- Houlihan LM, Davies G, Tenesa A, Harris SE, Luciano M, Gow AJ, et al. Common variants of large effect in F12, KNG1, and HRG are associated with activated partial thromboplastin time. *Am J Hum Genet*. 2010;86:626–631. doi: 10.1016/j.ajhg.2010.02.016.
- 22. Wain LV, Verwoert GC, O'Reilly PF, Shi G, Johnson T, Johnson AD, et al; LifeLines Cohort Study; EchoGen consortium; AortaGen Consortium; CHARGE Consortium Heart Failure Working Group; KidneyGen consortium; CKDGen consortium; Cardiogenics consortium; CardioGram. Genome-wide association study identifies six new loci influencing pulse pressure and mean arterial pressure. *Nat Genet.* 2011;43:1005–1011. doi: 10.1038/ng.922.
- Liu JZ, McRae AF, Nyholt DR, Medland SE, Wray NR, Brown KM, et al; AMFS Investigators. A versatile gene-based test for genome-wide association studies. *Am J Hum Genet*. 2010;87:139–145. doi: 10.1016/j. ajhg.2010.06.009.
- Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*. 2007;81:559–575. doi: 10.1086/519795.
- Hill WD, Davies G, van de Lagemaat LN, Christoforou A, Marioni RE, Fernandes CP, et al. Human cognitive ability is influenced by genetic variation in components of postsynaptic signalling complexes assembled by NMDA receptors and MAGUK proteins. *Transl Psychiatry*. 2014;4:e341. doi: 10.1038/tp.2013.114.
- Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U* S A. 2005;102:15545–15550. doi: 10.1073/pnas.0506580102.
- Wang K, Li M, Bucan M. Pathway-based approaches for analysis of genomewide association studies. *Am J Hum Genet*. 2007;81:1278–1283. doi: 10.1086/522374.
- Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genet Epidemiol*. 2010;34:816–834. doi: 10.1002/gepi.20533.
- 29. Perneger TV. What's wrong with Bonferroni adjustments. *BMJ*. 1998;316:1236–1238. doi: 10.1136/bmj.316.7139.1236.
- Grupe A, Li Y, Rowland C, Nowotny P, Hinrichs AL, Smemo S, et al. A scan of chromosome 10 identifies a novel locus showing strong association with late-onset Alzheimer disease. *Am J Hum Genet*. 2006;78:78– 88. doi: 10.1086/498851.

 Stein JL, Hua X, Lee S, Ho AJ, Leow AD, Toga AW, et al; Alzheimer's Disease Neuroimaging Initiative. Voxelwise genome-wide association study (vGWAS). *Neuroimage*. 2010;53:1160–1174. doi: 10.1016/j. neuroimage.2010.02.032.

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- Cheadle L, Biederer T. The novel synaptogenic protein Farp1 links postsynaptic cytoskeletal dynamics and transsynaptic organization. J Cell Biol. 2012;199:985–1001. doi: 10.1083/jcb.201205041.
- Ohsakaya S, Fujikawa M, Hisabori T, Yoshida M. Knockdown of DAPIT (diabetes-associated protein in insulin-sensitive tissue) results in loss of ATP synthase in mitochondria. J Biol Chem. 2011;286:20292–20296. doi: 10.1074/jbc.M110.198523.
- Waldmann TA, McIntire KR. Serum-alpha-fetoprotein levels in patients with ataxia-telangiectasia. *Lancet*. 1972;2:1112–1115. doi: 10.1016/ S0140-6736(72)92717-1.
- Ciemins JJ, Horowitz AL. Abnormal white matter signal in ataxia telangiectasia. AJNR Am J Neuroradiol. 2000;21:1483–1485.
- Grinberg LT, Thal DR. Vascular pathology in the aged human brain. Acta Neuropathol. 2010;119:277–290. doi: 10.1007/s00401-010-0652-7.
- Farrall AJ, Wardlaw JM. Blood-brain barrier: ageing and microvascular disease–systematic review and meta-analysis. *Neurobiol Aging*. 2009;30:337–352. doi: 10.1016/j.neurobiolaging.2007.07.015.
- Silver M, Janousova E, Hua X, Thompson PM, Montana G; Alzheimer's Disease Neuroimaging Initiative. Identification of gene pathways implicated in Alzheimer's disease using longitudinal imaging phenotypes with sparse regression. *Neuroimage*. 2012;63:1681–1694. doi: 10.1016/j. neuroimage.2012.08.002.
- Wong J. Altered expression of RNA splicing proteins in Alzheimer's disease patients: evidence from two microarray studies. *Dement Geriatr Cogn Dis Extra*. 2013;3:74–85. doi: 10.1159/000348406.
- Alachkar A, Jiang D, Harrison M, Zhou Y, Chen G, Mao Y. An EJC factor RBM8a regulates anxiety behaviors. *Curr Mol Med.* 2013;13: 887–899. doi: 10.2174/15665240113139990019.
- Herrmann LL, Le Masurier M, Ebmeier KP. White matter hyperintensities in late life depression: a systematic review. *J Neurol Neurosurg Psychiatry*. 2008;79:619–624. doi: 10.1136/jnnp.2007.124651.
- Valdés Hernández Mdel C, Booth T, Murray C, Gow AJ, Penke L, Morris Z, et al. Brain white matter damage in aging and cognitive ability in youth and older age. *Neurobiol Aging*. 2013;34:2740–2747. doi: 10.1016/j.neurobiolaging.2013.05.032.
- 43. Fischer MT, Sharma R, Lim JL, Haider L, Frischer JM, Drexhage J, et al. NADPH oxidase expression in active multiple sclerosis lesions in relation to oxidative tissue damage and mitochondrial injury. *Brain*. 2012;135(pt 3):886–899, doi: 10.1093/brain/aws012.

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Genes From a Translational Analysis Support a Multifactorial Nature of White Matter Hyperintensities

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