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Research paper

11 β -Hydroxysteroid dehydrogenase type 1 contributes to the regulation of 7-oxysterol levels in the arterial wall through the inter-conversion of 7-ketocholesterol and 7 β -hydroxycholesterol

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ABSTRACT

The atherogenic 7-oxysterols, 7-ketocholesterol (7-KC) and 7 β -hydroxycholesterol (7 β OHC), can directly impair arterial function. Inter-conversion of 7-KC and 7 β OHC has recently been shown as a novel role for the glucocorticoid-metabolizing enzyme 11 β -hydroxysteroid dehydrogenase type 1 (11 β -HSD1). Since this enzyme is expressed in vascular smooth muscle cells, we addressed the hypothesis that inter-conversion of 7-KC and 7 β OHC by 11 β -HSD1 may contribute to regulation of arterial function.

Incubation (4–24 h) of aortic rings with either 7-KC (25 μ M) or 7 β OHC (20 μ M) had no effect on endothelium-dependent (acetylcholine) or -independent (sodium nitroprusside) relaxation. In contrast, exposure to 7-KC (but not to 7 β OHC) attenuated noradrenaline-induced contraction (E_{max}) after 4 h (0.78 ± 0.28 vs 0.40 ± 0.08 mN/mm; $p < 0.05$) and 24 h (2.28 ± 0.34 vs 1.56 ± 0.48 mN/mm; $p < 0.05$). Both 7-oxysterols were detected by GCMS in the aortic wall of chow-fed C57Bl6/J mice, with concentrations of 7-KC (1.41 ± 0.81 ng/mg) higher ($p = 0.05$) than 7 β OHC (0.16 ± 0.06 ng/mg). In isolated mouse aortic rings 11 β -HSD1 was shown to act as an oxo-reductase, inter-converting 7-KC and 7 β OHC. This activity was lost in aorta from 11 β -HSD1^{-/-} mice, which had low oxysterol levels. Renal homogenates from 11 β -HSD1^{-/-} mice were used to confirm that the type 2 isozyme of 11 β -HSD does not inter-convert 7-KC and 7 β OHC.

These results demonstrate that 7-KC has greater effects than 7 β OHC on vascular function, and that 11 β -HSD1 can inter-convert 7-KC and 7 β OHC in the arterial wall, contributing to the regulation of 7-oxysterol levels and potentially influencing vascular function. This mechanism may be important in the cardioprotective effects of 11 β -HSD1 inhibitors.

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1. Introduction

Pro-atherogenic 7-oxysterols form a large component (40%) of oxidized LDL (oxLDL), of which 7-ketocholesterol (7-KC) contributes ~30% [1]. 7-KC is toxic to cells in the vessel wall, and can impair arterial function *ex vivo* [2]. Indeed, 7-KC and its metabolite

Abbreviations: BHT, butylated hydroxytoluene; DMEM, Dulbecco's modified Eagle medium; EDTA, ethylenediaminetetraacetic acid; 11 β -HSD1, 11 β -hydroxysteroid dehydrogenase type 1; 11 β -HSD2, 11 β -hydroxysteroid dehydrogenase type 2; 7 α OHC, 7 α -hydroxycholesterol; 7 β OHC, 7 β -hydroxycholesterol; 7-KC, 7-ketocholesterol.

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7 β -hydroxycholesterol (7 β OHC) inhibited endothelium-dependent, acetylcholine-induced relaxation of rabbit aortic rings *in vitro* [2]. In human umbilical vein endothelial cells (HUVECs), 7 β OHC and 7-KC reduced the histamine-activated release of nitric oxide [3]. This inhibition of endothelial function by 7-oxysterols appears to be independent of their cytotoxic effects [4], but its mechanism is unclear. Importantly, 7-KC and 7 β OHC differ in their pro-atherogenic potential, with 7-KC implicated as the major pro-inflammatory and cytotoxic oxysterol [5]. However, any differences between the functional effects of 7-KC and 7 β OHC in the vasculature have not been addressed.

The balance between 7-KC and 7 β OHC in tissues may be actively modulated. Recently, a novel route of metabolism of 7-oxysterols has been described, involving the enzyme 11 β -hydroxysteroid dehydrogenase (11 β -HSD) type 1. The primary role of 11 β -HSD1 is to catalyse the pre-receptor generation of glucocorticoids, allowing tissue-specific amplification of glucocorticoid receptor activation

[6]. Inactivation of glucocorticoids is catalysed by the type 2 isozyme of 11 β -HSD (11 β -HSD2) [7]. It is becoming increasingly apparent that 7-oxysterols are alternative substrates for 11 β -HSD1 [8,9], and inhibition of the enzyme can result in accumulation of 7-KC [10]. Since both isozymes of 11 β -HSD are present in the arterial wall [11–14], where they are able to inter-convert glucocorticoids [15], it is conceivable that inter-conversion of 7-oxysterols by these enzymes has a role in modulating vascular function.

We used mice with targeted disruption of the 11 β -HSD1 gene (*Hsd11b1*) to investigate the hypothesis that 11 β -HSD1 metabolises 7-oxysterols in the arterial wall, thus influencing 7-KC- and 7 β OHC-mediated modulation of arterial function.

2. Methods

2.1. Chemicals and stock solutions

All solvents were HPLC grade (Fisher, Hemel Hempstead, UK) and were prepared containing an anti-oxidant (0.01% w/v butylated hydroxytoluene (BHT)) to prevent oxidative degradation of the lipids [3]. Steroids and oxysterols were from Steraloids (Newport, Rhode Island, USA), derivatization reagents from Fluka (Buchs, Switzerland), tissue culture reagents from Lonza (Reading, UK) and other chemicals from Sigma-Aldrich (Poole, Dorset, UK). Deuterium-labelled internal standards for GCMS were obtained from CDN Isotopes (Qmx Laboratories, Essex, UK). Stock solutions (30 mg/ml in ethanol with 250 μ g/ml BHT) of 7-KC, 7 β OHC and 7 α OHC (an optical isomer of 7 β OHC) were freshly prepared as required. All steroids were prepared in 100% ethanol. Working solutions for tissue culture were prepared in standard Dulbecco's modified Eagle's medium (DMEM). Working solutions (25 μ M 7-KC; 20 μ M 7 β OHC) for myography were prepared by diluting the appropriate stock solution in DMEM without L-Arginine (Arg) or phenol red, but containing 1% charcoal-stripped foetal calf serum. These were the maximum concentrations of 7-oxysterols that could be achieved without sample precipitation. The final concentration of vehicle (ethanol with 250 μ g/ml BHT) was <0.2%.

2.2. Animals

Male mice (age 8–16 weeks) homozygous for disrupted alleles of 11 β -HSD1 (*Hsd11b1*^{-/-}) [16] or 11 β -HSD2 (*Hsd11b2*^{-/-}) [7], on a C57Bl6/J background [12] were bred in-house. Controls were age- and sex-matched C57Bl6/J mice bred in-house [7,16]. Mice were maintained on standard chow diet and tap water *ad libitum*, under a 16 h/8 h light/dark cycle at 21–24 °C. All procedures were performed under UK Home Office guidelines of humane care and [17,18] animals were culled by cervical dislocation at 10.00 h. Plasma (1 ml) was collected from 2–3 mice in EDTA-coated (1.6 mg/ml) vials (Sarstedt, Monovette) and separated by centrifugation (2000 \times g, 4 °C, 15 min), and an aliquot of BHT was added (50 μ g/5 μ l). Tissues were snap-frozen and stored at -80 °C until use. Aortae for functional investigation were removed from mice, placed in PBS (37 °C), cleaned of peri-adventitial fat and used for myography. Aortae for oxysterol analysis were processed as described below.

2.3. Functional effects of 7-oxysterols on isolated mouse aortic rings

Thoracic aortae were isolated from male C57Bl6/J mice (age 8–10 weeks, $n = 12$) and cut into four rings (2 mm in length). These were either used immediately for short-term (4 h), or incubated in a 24 well plate for extended (24 h), exposure to 7-oxysterols. For short-term exposures, aortic rings were mounted on intra-luminal wires in a small vessel wire myograph [19,20] containing DMEM

without L-Arg (37 °C, continuously perfused with 95%O₂: 5% CO₂) [14]. After the vessels had been equilibrated at their optimum resting force they were contracted with KCl (125 mM, 3 times) to confirm viability and then incubated in the presence of: (1) 7-KC (25 μ M in DMEM without L-Arg), (2) 7 β OHC (20 μ M in DMEM without L-Arg) or (3) vehicle alone (ethanol, with 50 μ g/ml BHT in DMEM without L-Arg), for 4 h (2 rings/treatment/mouse). The incubating medium was replaced every 60 min. After 4 h cumulative concentration–response curves were obtained for 5-hydroxytryptamine (5-HT; 1×10^{-9} – 1×10^{-4} M) and noradrenaline (NA; 1×10^{-9} – 1×10^{-4} M). In addition, cumulative concentration–response curves were obtained for the vasodilators, acetylcholine (ACh; 1×10^{-9} – 1×10^{-4} M, endothelium-dependent) and sodium nitroprusside (SNP; 1×10^{-9} – 1×10^{-4} M, endothelium-independent), following contraction with a sub-maximal concentration of 5-HT (3×10^{-7} – 1×10^{-6} M). Contractile responses are expressed as force per unit length (mN/mm). Relaxations were expressed as a percentage of the contraction in response to the EC₈₀ of 5-HT (% 5-HT).

For extended exposures [14], aortic rings were placed in a 24 well plate and immersed in 1 ml DMEM (without L-Arg) containing either 7-KC (25 μ M), 7 β OHC (20 μ M), or vehicle (ethanol with 50 μ g/ml BHT) and incubated overnight in a humidified incubator (37 °C; 5% CO₂). These vessels were then mounted in a myograph and functional studies performed, as described above, in the continued presence of the appropriate 7-oxysterol or vehicle.

2.4. Determination of plasma and aortic levels of 7-oxysterols and cholesterol

Concentrations of cholesterol and 7-oxysterols in the plasma and aortae were quantified by GCMS. Aortae from *Hsd11b1*^{-/-} or C57Bl6/J mice were pooled from two animals, washed in PBS containing EDTA (0.5 mM), crushed under liquid nitrogen and homogenized. Protein concentration was determined using Bradford assay (Biorad, UK). Deuterium-labelled (²H, d₇) internal standards (IS) were added (50 μ l) and lipids were extracted into chloroform/methanol (2:1, 8 ml) [21,22]. Samples were purified using Bond Elute Diol columns (100 mg, 1 ml; Varian, UK) [23] and hydrolysed following mild saponification [24,25]. Oxidized lipids and cholesterol were extracted from neutralized samples (0.35 ml, 20% acetic acid) into diethyl ether (4 ml, 0.01% BHT) and evaporated to dryness under argon. Total cholesterol and 7-oxysterol concentrations were measured by GCMS and corrected for aortic protein levels.

2.5. Metabolism of 7-oxysterols by 11 β -HSD1 and 11 β -HSD2 *in vitro*

2.5.1. In the mouse aorta

Rings (2 mm long) of aortae from C57Bl6/J and *Hsd11b1*^{-/-} mice ($n = 8$ /group) were placed in a 24 well plate (1/well, in duplicate) and immersed in 1 ml DMEM (without L-Arg) containing 7-KC (25 μ M), 7 β OHC (20 μ M), 7 α OHC (20 μ M) or vehicle (ethanol, with 50 μ g/ml BHT). Samples were incubated overnight in a humidified incubator (37 °C; 5% CO₂) then blotted dry on tissue paper and weighed to allow calculation of conversion velocity (pmol/mg/day). Medium was removed and deuterium-labelled internal standards (IS; [²H], d₇-7-KC (40 ng), [²H], d₇-7 β OHC (10 ng) and [²H], d₇-cholesterol (10 μ g)) added in a single aliquot (50 μ l). [²H], d₇-7 β OHC was used as an internal standard for quantitation of both 7 α OHC and 7 β OHC. Argon gas was flushed through all samples and oxysterols were extracted (8 ml, 100 \times g, 15 min) from media with a mixture of hexane:2-propanol (60:40) [26]. The organic phases were combined, evaporated under

a stream of argon and residues dissolved in chloroform:methanol (2:1, 350 μ l) before storing at -20°C for analysis by GCMS. Results were subsequently corrected for aortic ring weight. In all assays appropriate positive controls were included, with aortic rings incubated with [^3H], d_4 -corticosterone or [^3H], d_4 -11-dehydrocorticosterone (30 nM) to verify the activity of 11 β -HSD isozymes. Samples were processed for analysis as before [27].

2.5.2. In kidney

Murine kidneys contain both isoforms of 11 β -HSD. Homogenates of kidneys from *Hsd11b1*^{-/-} mice (which lack 11 β -HSD1) were used as a source of murine 11 β -HSD2, with kidneys from C57Bl6/J mice as controls. Kidneys were homogenized in phosphate buffer as detailed [10]. Homogenates (400 μ g/ml) were incubated with 7-oxysterols (20 μ M) and the appropriate cofactor (2 mM): NAD⁺ or NADP⁺ for dehydrogenase reactions; NADH or NADPH for reductase reactions. In all assays conversion of dexamethasone (Dex) and 11-dehydrodexamethasone (11-DHDex; 40 μ M) was used as a positive control for confirmation of 11 β -HSD isozyme activity [28].

2.6. Chromatographic analyses

2.6.1. Analysis of 7-oxysterols by gas chromatography/mass spectrometry (GCMS)

7-Oxysterols and cholesterol were converted to trimethylsilyl ether derivatives using a pyridine:hexamethyldisilazan:trimethylchlorosilane mixture (350 μ l, 3:2:1, v/v/v) [29,30]. The derivatized cholesterol metabolites were dissolved in 2% *N,O*-Bis(trimethylsilyl) trifluoroacetamide (BSTFA) in decane (80 μ l) and eluted as follows: initial temperature 180 $^{\circ}\text{C}$ (1 min), increased by 35 $^{\circ}\text{C}/\text{min}$ until 270 $^{\circ}\text{C}$ was achieved (1 min) and then increased by 4 $^{\circ}\text{C}/\text{min}$ to 300 $^{\circ}\text{C}$ (12 min). The oven was then cooled by $-10^{\circ}\text{C}/\text{min}$ to 250 $^{\circ}\text{C}$ (1 min). The injection temperature was 270 $^{\circ}\text{C}$.

A capillary gas chromatograph (Trace GC, Thermo) was coupled to an ion-trap, Polaris Q (Thermo, Hemel Hempstead, UK) mass spectrometer (MS) and equipped with a BPX5 capillary column (25 m, 0.32 mm internal diameter and 0.25 μ m film thickness; SGE, Alva, UK) and operated in SIM mode with electron impact (70 eV), ion source, transfer line and interface temperatures of 200 $^{\circ}\text{C}$, 220 $^{\circ}\text{C}$ and 250 $^{\circ}\text{C}$ respectively. Derivatives of 7-oxysterols and cholesterol were quantified by monitoring the following ions (*m/z*): 7-KC (472, 16.5 min), 7 α / β OH (456, 12.3 & 14.1 min), d_7 -7-KC (479, 16.35 min), d_7 -7 β OH (463, 13.8 min) and d_7 -cholesterol (375, 12.9 min). Limits of detection were assigned as 3:1 signal to noise ratio. Compounds were quantified by the ratio of area under peak of interest to area under peak of internal standard against a standard curve.

2.6.2. Quantitation of steroids by high pressure liquid chromatography

Radio-labelled glucocorticoids were separated by reverse phase HPLC (Symmetry C8 column maintained at 35 $^{\circ}\text{C}$; column length, 15 cm, internal diameter 4.6 mm, pore size 5 μ m, Waters, Edinburgh, UK) and quantified by on-line liquid scintillation counting (2 ml/min; GoldFlow, Meridian, Surrey, UK). Total run time was 35 min (elution times of epi-cortisol, 11-dehydrocorticosterone and corticosterone were typically 12 min, 21 min and 31 min, respectively, with mobile phase of water:acetonitrile:methanol (60:15:25) at 1 ml/min). Dex and 11-DHDex were separated using a mobile phase of water:acetonitrile:methanol (55:20:25) at 1 ml/min with typical retention times for epi-cortisol (10 min), 11-DHDex (12 min) and Dex (16 min). UV detection of all steroids was achieved at 240 nm and epi-cortisol was used as an internal standard. Steroids were quantified by the ratio of area under peak of interest to area under peak of internal standard against a standard curve.

2.7. Statistical analysis

All data are mean \pm standard error of the mean (SEM) where *n* indicates the number of different animals. Values were compared using unpaired Student's *t*-tests or 1-way ANOVA with Dunnett's multiple comparison post-tests, as appropriate. All analyses were performed using Graph Pad Prism v5.0 (GraphPad Software Inc. San Diego, USA). Statistical significance was assumed when *p* < 0.05.

3. Results

3.1. 7-KC, but not 7 β OH, alters vascular function in vitro

Short-term exposure (4 h) of aortae from C57Bl6/J mice to 7-KC (25 μ M), but not 7 β OH (20 μ M), produced a small reduction (*p* = 0.049) in NA-induced maximum contraction (E_{max}), but had no effect on the sensitivity (pD_2) of this response (Fig. 1A, B; Table 1A). 5-HT-mediated contraction was unaltered by exposure to either oxysterol (Table 1A). Pre-treatment of vessels with either 7-KC or 7 β OH did not alter endothelium-dependent relaxation to ACh (Fig. 1C, D). An apparent increase in maximal response to endothelium-independent, SNP-mediated vasorelaxation after incubation with either 7-KC (*p* = 0.05) or 7 β OH (*p* = 0.08) was of borderline statistical significance (Fig. 1E, F).

Long-term (24 h; Fig. 2) exposure of aortae from C57Bl6/J mice to 7-KC (25 μ M), but not 7 β OH (20 μ M), produced a reduced maximum contraction (E_{max} , *p* = 0.049), but no change in sensitivity (pD_2) to NA (Fig. 2A, B; Table 1 B). Prolonged incubation with either 7-oxysterol had no effect on 5-HT-mediated contraction or endothelium-dependent (Fig. 2C, D) or -independent (Fig. 2E, F) relaxation (Table 1 B).

3.2. 7-Oxysterols are present in the mouse aortic wall and altered by deletion of 11 β -HSD1

7-KC (3.52 ± 2.85 nmol/g tissue) and 7 β OH (0.40 ± 0.15 nmol/g) were both detected in the mouse thoracic aortae with levels of 7-KC significantly higher than 7 β OH (*p* = 0.05; *n* = 12). In aortae from *Hsd11b1*^{-/-} mice, 7-KC was only present in levels above the limit of detection in 3 (of 8) samples and 7 β OH was also low (0.12 ± 0.02 nmol/mg). Plasma levels of 7-oxysterols were not different in *Hsd11b1*^{-/-} mice compared with C57Bl6/J mice (7-KC; 0.133 ± 0.016 versus 0.091 ± 0.022 μ M; 7 β OH 0.024 ± 0.002 versus 0.023 ± 0.005 μ M, respectively) although total plasma cholesterol was lower (0.45 ± 0.14 versus 1.03 ± 0.19 μ M, *p* < 0.05).

3.3. 7-Oxysterols are metabolized by 11 β -HSD1 but not by 11 β -HSD2

As expected [15], glucocorticoids were inter-converted by incubation with intact mouse aortic rings. The velocity of reduction of 11-dehydrocorticosterone to corticosterone (Fig. 3A) proceeded considerably ($\sim 10\times$) faster than the dehydrogenation of corticosterone to 11-dehydrocorticosterone. Reduction of 11-dehydrocorticosterone was attenuated in mice lacking 11 β -HSD1, whereas deletion of this enzyme produced only a small (though significant) increase in the dehydrogenation of corticosterone (to 11-dehydrocorticosterone) (Fig. 3A). The oxysterols 7-KC and 7 β OH were also inter-converted by incubation with intact mouse aortic rings. In contrast to glucocorticoids, however, the velocities of reduction of 7-KC (to 7 β OH) and of dehydrogenation of 7 β OH (to 7-KC) were similar following incubation with mouse aortic rings (Fig. 3B). Genetic disruption of *Hsd11b1* significantly reduced the velocity of conversion of both 7-KC and 7 β OH (Fig. 3B), with

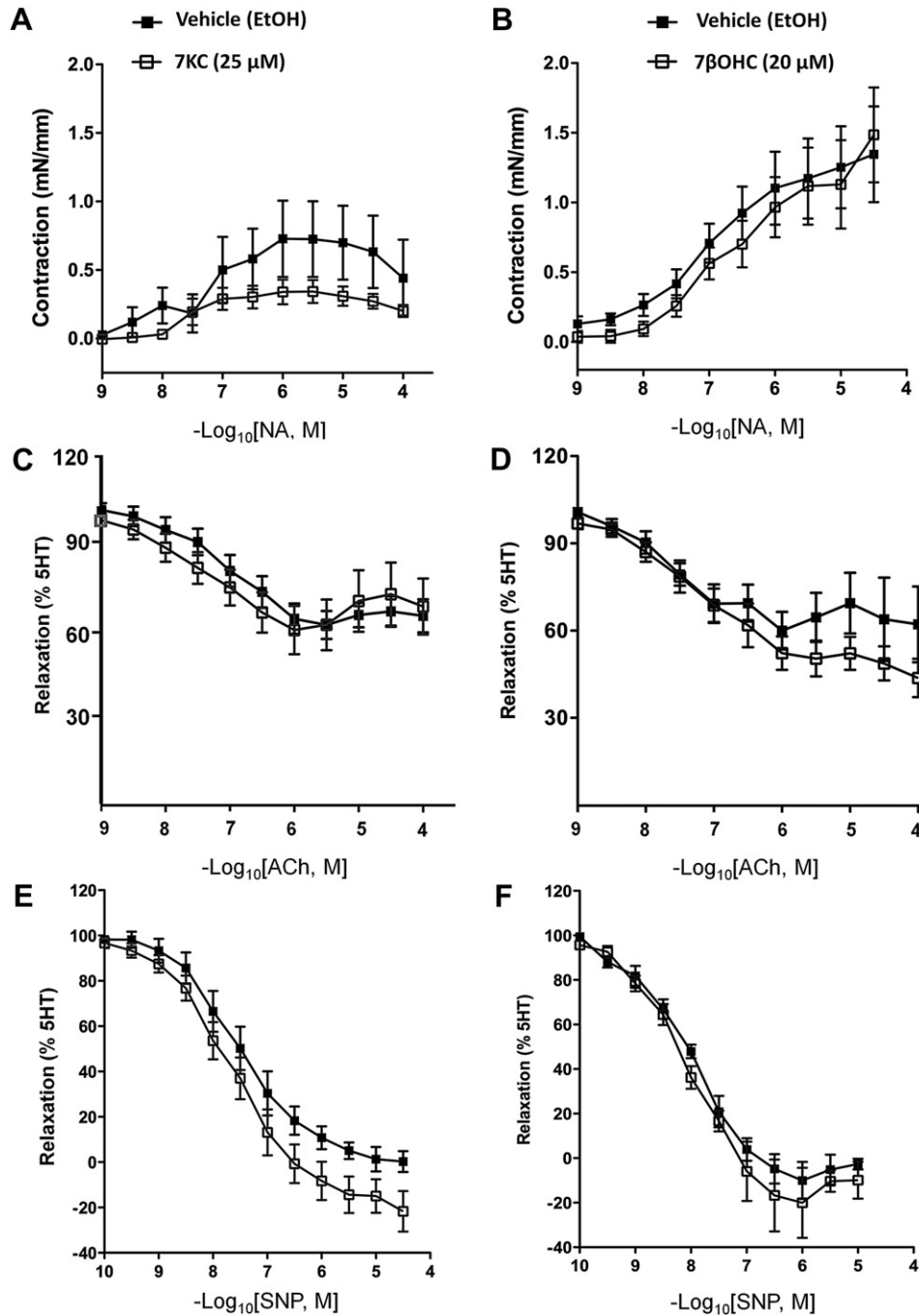


Fig. 1. Short-term (4 h) exposure to 7-ketocholesterol induces agonist-specific functional changes in isolated mouse aorta. Endothelium-intact aortic rings from C57Bl6/J mice were incubated (4 h) with 7-ketocholesterol (7-KC, 25 μM open squares) or 7 β -hydroxycholesterol (7 β OHC, 20 μM , open squares) and compared with vehicle (ethanol containing 50 $\mu\text{g}/\text{ml}$ butylated hydroxytoluene)-treated control (filled squares). Incubation with 7-KC (A), but not 7 β OHC (B), produced a small reduction of noradrenaline (NA)-mediated contraction ($p = 0.04$). Incubations had no effect on acetylcholine (ACh)-mediated relaxation (C, D) whereas 7-KC (E) (but not 7 β OHC (F)), produced a trend towards increased sodium nitroprusside (SNP)-mediated relaxation ($p = 0.054$). Relaxations were expressed on a scale where the response to 5-HT represented 100% and return to baseline was expressed as 0%. All points represent mean \pm SEM, compared by 1-way ANOVA with Tukey's post hoc test, $n = 6-8$.

96 \pm 6% of added substrates being recovered. 7-KC was not inter-converted with 7 α OHC in aortic rings (data not shown).

Hsd11b1^{-/-} mouse kidney homogenates (a rich source of 11 β -HSD2; [31]) were used to determine whether 7-oxysterols are metabolized by this isozyme. As with the aortic rings, conversion of glucocorticoids was used as a positive control for activity of 11 β -HSD2 [14,27]. As anticipated, glucocorticoids were metabolized by mouse renal homogenates with preferential generation of 11-DHDex from Dex (oxidation; not shown). In contrast, renal

homogenates did not inter-convert any of the 7-oxysterols (7 α OHC, 7 β OHC or 7-KC). Unrecovered substrate was \sim 3% or lower for each compound (7 α OHC, 1.9 \pm 0.7%; 7 β OHC, 3.2 \pm 0.3%; KC, 2.1 \pm 0.3%).

4. Discussion

This study shows for the first time that 11 β -HSD1, but not 11 β -HSD2, catalyses the conversion of 7-oxysterols in the vascular wall. Previous work has shown that murine and human 11 β -HSD1

Table 1

Exposure to 7-oxysterols caused an agonist-selective inhibition of contraction, but had no effect on relaxation, of mouse aortic rings *in vitro*.

A) Short (4 h) incubation				
(i) 7-Ketocholesterol (7-KC)				
Agonist	E_{max} (mN/mm or % relaxation)		$pD_2/-logIC_{50}$	
	Vehicle	7-KC	Vehicle	7-KC
NA	0.79 ± 0.28	0.40 ± 0.08*	6.70 ± 0.34	7.92 ± 0.48
5-HT	3.01 ± 0.37	2.47 ± 0.26	6.36 ± 0.08	6.55 ± 0.06
ACh	46.3 ± 5.3	47.1 ± 7.0	6.32 ± 0.09	6.48 ± 0.09
SNP	104.8 ± 3.8	122.0 ± 8.3	7.53 ± 0.22	7.56 ± 0.12
(ii) 7β-Hydroxycholesterol (7βOHC)				
Agonist	E_{max} (mN/mm or % relaxation)		$pD_2/-logIC_{50}$	
	Vehicle	7βOHC	Vehicle	7βOHC
NA	1.60 ± 0.36	1.80 ± 0.61	6.90 ± 0.17	6.80 ± 0.22
5-HT	2.80 ± 0.28	3.30 ± 0.32	6.40 ± 0.04	6.50 ± 0.05
ACh	47.7 ± 6.2	59.1 ± 4.8	7.30 ± 0.32	7.10 ± 0.16
SNP	116.7 ± 10.3	121.7 ± 12.0	7.90 ± 0.11	8.20 ± 0.11
B) Extended (24 h) Incubation				
(i) 7-Ketocholesterol (7-KC)				
Agonist	E_{max} (mN/mm or % relaxation)		$pD_2/-logIC_{50}$	
	Vehicle	7-KC	Vehicle	7-KC
NA	2.28 ± 0.34	1.56 ± 0.48*	7.79 ± 0.16	7.94 ± 0.33
5-HT	4.03 ± 0.24	3.63 ± 0.33	6.61 ± 0.05	6.60 ± 0.12
ACh	66.9 ± 4.5	75.4 ± 6.4	7.47 ± 0.13	7.35 ± 0.21
SNP	108.6 ± 5.0	106.0 ± 1.5	7.04 ± 0.17	6.95 ± 0.19
(ii) 7β-Hydroxycholesterol (7βOHC)				
Agonist	E_{max} (mN/mm or % relaxation)		$pD_2/-logIC_{50}$	
	Vehicle	7βOHC	Vehicle	7βOHC
NA	1.21 ± 0.14	1.16 ± 0.12	6.55 ± 0.06	6.24 ± 0.03
5-HT	2.43 ± 0.14	2.88 ± 0.15	6.51 ± 0.15	6.90 ± 0.32
ACh	46.4 ± 5.6	60.3 ± 5.2	7.17 ± 0.15	6.97 ± 0.31
SNP	104.6 ± 3.46	109.5 ± 1.94	6.84 ± 0.16	6.72 ± 0.19

All values represent mean ± SEM, compared by unpaired Student's *t*-test (vehicle vs 7-oxysterol), *n* = 4–8, **p* < 0.05. NA, noradrenaline; 5-HT, 5-hydroxytryptamine; ACh, acetylcholine; SNP, sodium nitroprusside.

Bold represents significant differences in the data.

converts 7-KC to 7βOHC in the liver and in cultured adipocytes [8,9]. We provide evidence that murine 11β-HSD1 reduces 7-KC to 7βOHC in the vessel wall but, furthermore, that it also oxidizes 7βOHC to 7-KC. Use of *Hsd11b1*^{-/-} mice confirmed that 11β-HSD1 was the sole enzyme responsible for metabolism of 7-KC and 7βOHC in the aortic wall and that deletion of 11β-HSD1 alters vascular 7-oxysterol concentrations. Functional investigations showed differential effects of 7-KC and 7βOHC on vascular function, suggesting that this 11β-HSD1-mediated inter-conversion of 7-oxysterols may influence 7-KC-mediated inhibition of arterial contraction.

7-KC and 7βOHC have both been shown previously to inhibit endothelium-dependent vasorelaxation [32], cause endothelial cell death, and induce production of radical oxygen species [17,33]. This is consistent with the ability of oxidized lipids to impair the endothelium-dependent relaxation of aortic segments from hyperlipidaemic mice [34]. The lack of impact of exposure to oxysterols on endothelium-dependent relaxation was surprising given the previous indications that both 7-KC and 7βOHC inhibit endothelial function [2,3,32] *ex vivo*. One possible explanation for lack of effect on vasorelaxation is the use of a low concentration of 7-oxysterol (20–25 μM) compared with previous studies (180–270 μM; [2,3,32]). The concentrations used for our investigations were the highest we could achieve without precipitation and are consistent with that used (25 μM) to show 7-oxysterol-mediated smooth muscle apoptosis *in vitro* [35]. Furthermore, a recent investigation using high concentrations of 7-KC (205 μM) found no effect of *ex vivo* incubation on ACh-mediated relaxation of mouse aorta [36].

Intriguingly at the concentrations used in this investigation, there was an inhibition of smooth muscle cell contraction by 7-KC

that was not observed with 7βOHC. The mechanism involved is unclear but the effect was selective for noradrenaline, suggesting an impairment in the α₁-adrenoceptor signalling pathway. Impaired contractility is consistent with 7-KC at this concentration having detrimental effects on vascular smooth muscle cells [35]. These results suggest, therefore, that the balance of 7-KC and 7βOHC may have functional and structural implications in the arterial wall.

The concentrations of 7-oxysterols in the vessels of C57Bl6/J mice are consistent with those reported previously in human plasma and vessels [1,24]. Since circulating 7-oxysterols can be sequestered by cells in the vessel wall [37], we assessed the potential of vascular 11β-HSD1 to inter-convert 7-oxysterols in this tissue. Plasma 7-oxysterol levels were not altered in *Hsd11b1*^{-/-} mice although total plasma cholesterol was substantially lower. Consistent with previous reports of reduced intra-vascular cholesterol accumulation with inhibition of 11β-HSD1 [38], we found lower levels of all 7-oxysterols in the aortae of *Hsd11b1*^{-/-} mice. It was, therefore, difficult to assess intra-vascular 7-KC:7βOHC ratios, since 7-KC levels in particular were near to the detection limit, but the data suggest that 7-KC levels are disproportionately reduced in *Hsd11b1*^{-/-} mice, consistent with the enzyme acting predominantly as an oxidase (converting 7βOHC to 7-KC) *in vivo*.

The *ex vivo* incubation of aortic rings described here has not previously been used to assess inter-conversion of 7-oxysterols. This approach confirmed that the stability of 7-oxysterols can be preserved during incubation, as both 7-KC and 7βOHC were successfully recovered from DMEM. It had been postulated that 7-oxysterols may be taken up by the vessels during incubation but the percentage recovery of 7-oxysterols from reaction mixtures did not support this. Preparation of concentrated stock solutions of the 7-oxysterols proved unexpectedly difficult, despite using published methodology [3], with oxysterols precipitating at high concentrations. Based on our own experiences and advice from other groups 7-oxysterol solutions were prepared in DMEM containing FCS containing an antioxidant (BHT; to prevent oxidative degradation of the lipids [3]). It is unlikely that BHT would have a detrimental effect on vascular function as it did not alter histamine-induced NO production in cultured HUVECs [32].

Ex vivo assays clearly demonstrated that incubation of 7-oxysterols with mouse aortic rings results in the conversion of 7βOHC to 7-KC and 7-KC to 7βOHC, but not inter-conversion of 7αOHC and 7-KC. This is consistent with results generated in rats [9,10] and humans [39] but contrasts with the demonstration that 11β-HSD1 in hamsters can inter-convert 7αOHC and 7-KC [40]. The ability of 11β-HSD1 to inter-convert 7-oxysterols explains why carbenoxolone, a non-selective 11β-HSD inhibitor, attenuates 7-oxysterol metabolism in rat hepatic microsomes [10]. Interestingly, in contrast to the predominant reductase direction (11-dehydrocorticosterone to corticosterone) shown for metabolism of glucocorticoids, murine vascular 11β-HSD1 showed similar activity as both reductase (7βOHC to 7-KC) and dehydrogenase (7-KC to 7βOHC) for inter-conversion of oxysterols, consistent with previous reports in liver [9,39]. Under these assay conditions, the reaction velocity for inter-conversion of oxysterols was considerably (approximately 10-fold) higher than for reduction of 11-dehydrocorticosterone. This contrasts with the demonstration of similar reaction velocities observed in other studies [9,40] and is likely to be a consequence of study design as substrate concentrations were higher (~800×) for the oxysterols than for the glucocorticoids.

Residual metabolism of glucocorticoids in aortae from *Hsd11b1*^{-/-} mice is consistent with vascular 11β-HSD2 expression [14,20]. Virtually no residual inter-conversion of 7βOHC and 7-KC

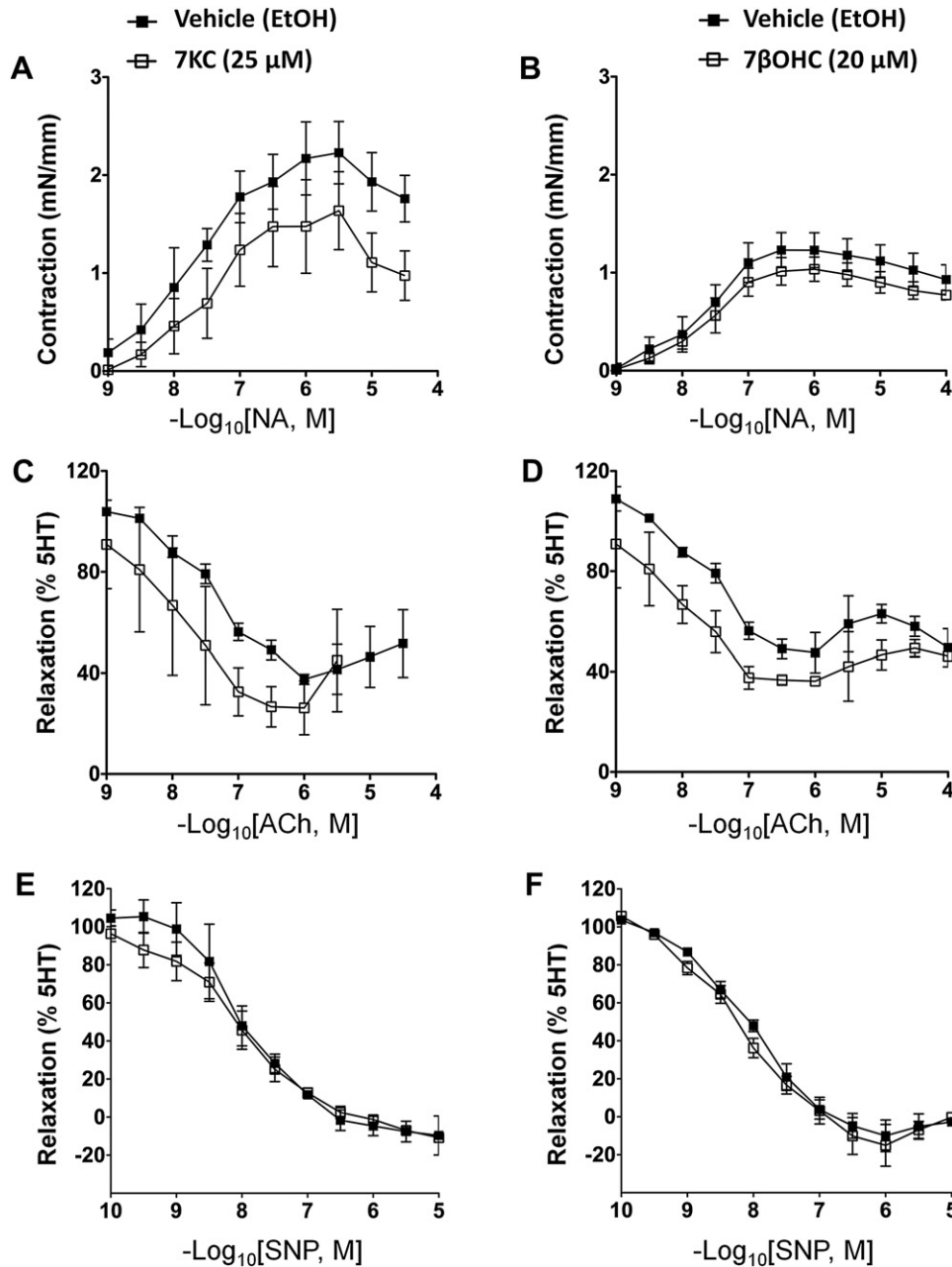


Fig. 2. Long-term (24 h) exposure to 7-ketocholesterol induces agonist selective functional changes in isolated mouse aorta. Endothelium-intact aortic rings from C57Bl6/J mice were incubated (24 h) with 7-ketocholesterol (7-KC, 25 μM open squares) or 7 β -hydroxycholesterol (7 β OHC, 20 μM , open squares) and compared with vehicle (ethanol containing 50 $\mu\text{g}/\text{ml}$ butylated hydroxytoluene)-treated control (filled squares). Incubation with 7-KC (A), but not 7 β OHC (B), produced a small reduction of noradrenaline (NA)-mediated contraction ($p = 0.05$). Incubations had no effect on acetylcholine (ACh)-mediated (C, D) or sodium nitroprusside (SNP)-mediated (E, F) relaxation. All points represent mean \pm SEM, compared by 1-way ANOVA with Tukey's post hoc test, $n = 6-8$.

was observed in aortae from mice lacking 11 β -HSD1. Lack of 7-oxysterol metabolism by 11 β -HSD2 was confirmed using kidney homogenates (since the kidney is rich in 11 β -HSD2 [15]; using kidneys from *Hsd11b1*^{-/-} mice ensured that there was no interference from this isozyme). This finding is consistent with the previous attribution of 7-oxysterol metabolism solely to the action of 11 β -HSD1 in hamster [40], rat [9,10], guinea pig [9,41] and human [39]. There was, however, a notable loss of substrate in the reaction mixtures; suggesting incomplete recovery of substrate, non-enzymatic degradation, or formation of alternative products [42]. There was no loss of substrate in blank samples (containing buffer but no tissue homogenate), confirming chemical stability of 7-oxysterols during the incubation.

Direct action on the cells of the arterial wall may not present the only mechanisms through which oxysterols can influence regulation of arterial function and structure. Previous work in our group [43] has indicated that the ability of oxysterols to act as substrates for 11 β -HSD1 also makes them potential competitive inhibitors of glucocorticoid metabolism. This presents the possibility that endogenous 7-oxysterols contribute to regulation of 11 β -HSD1-dependent glucocorticoid generation. Glucocorticoids can interact directly with the arterial wall to enhance vasoconstriction [44], impair endothelium-dependent relaxation [45], inhibit angiogenesis [27] and reduce vascular lesion formation. There is increasing evidence that these interactions are regulated by the activity of 11 β -HSD1 [27,38]. However, it is notable that no systematic

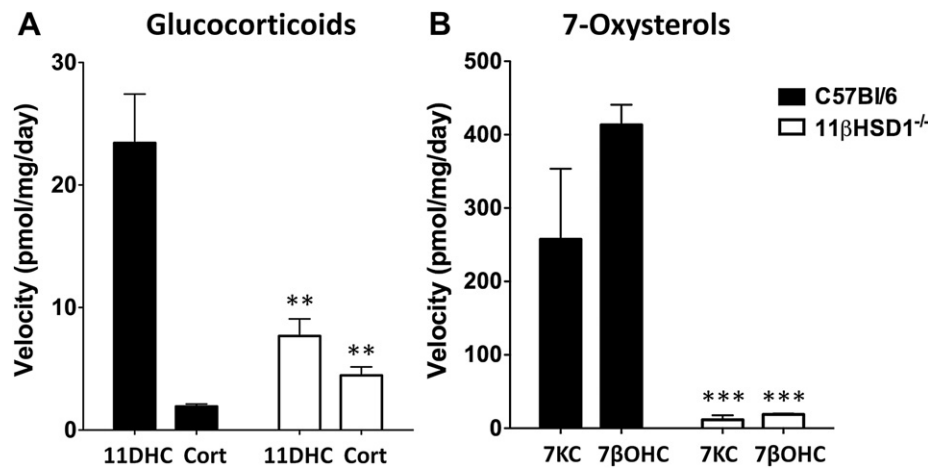


Fig. 3. 11β-HSD1 catalyses reduction of 7-Ketocholesterol (7-KC) and dehydrogenation of 7β-Hydroxycholesterol (7βOHC) in isolated mouse aorta. Incubation with mouse aortic rings (24 h; 37 °C; 5% CO₂) resulted in (A) metabolism of glucocorticoids ($n = 10$); reduction of 11-dehydrocorticosterone (11DHC; 30 nM) to form corticosterone (Cort) was reduced, but not abolished, in aortae from 11β-HSD1^{-/-} mice. Low levels of dehydrogenation of Cort (30 nM; to form 11DHC) were detected in mouse aorta. Surprisingly this was slightly (but significantly) increased in the absence of 11β-HSD1. (B) Both 7-oxysterols (1 μM) were metabolized following exposure to mouse aortic rings ($n = 6$) but, in contrast to glucocorticoids, both dehydrogenation (conversion of 7βOHC to 7-KC) and reduction (7-KC to 7βOHC) reactions were virtually abolished in arteries lacking 11β-HSD1 (11β-HSD1^{-/-}). Data are mean ± SEM, and were compared using unpaired Student's *t*-test, ** $p < 0.01$, *** $p < 0.001$ vs velocity of the same reaction in tissues from C57Bl/6 mice.

difference in vascular function has been observed in vessels from *Hsd11b1*^{-/-} mice [20], so whether alterations in either 7-oxysterol or glucocorticoids influences physiological vascular function remains uncertain. Perhaps interactions of oxysterols with 11β-HSD1 are more important in pathology. In healthy individuals, the maximum concentrations of 7-oxysterols [46,47] are lower than those in patients with atherosclerosis who may have levels of 7-oxysterols in the micromolar range [1]. It is plausible that inhibition of 11β-HSD1-mediated glucocorticoid generation in conditions of 7-oxysterol excess may have an indirect impact on arterial function and remodelling.

Metabolism of 7-oxysterols by 11β-HSD1 may also have implications for the development of new therapies. Selective 11β-HSD1 inhibition prevents atherosclerosis [38] and is being developed for treatment of cardiovascular risk factors [48], but the mechanisms responsible for this atheroprotective effect have not been demonstrated. It is conceivable that the beneficial effects of 11β-HSD1 inhibition are a consequence of prevention of 7-oxysterol interconversion as well as glucocorticoid metabolism.

5. Conclusions

11β-HSD1 influences 7-oxysterol concentrations within the arterial wall. By altering the balance of 7-ketocholesterol and 7β-hydroxycholesterol, 11β-HSD1 may modulate their specific effects on vascular function, especially in disease states in which oxysterol levels are increased.

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References

[1] A.J. Brown, W. Jessup, Oxysterols: sources, cellular storage and metabolism, and new insights into their roles in cholesterol homeostasis, *Molecular Aspects of Medicine* 30 (2009) 111–122.

[2] V. Deckert, L. Persegol, L. Viens, G. Lizard, A. Athias, C. Lallemand, P. Gambert, L. Lagrost, Inhibitors of arterial relaxation among components of human oxidized low-density lipoproteins: cholesterol derivatives oxidized in position 7 are potent inhibitors of endothelium-dependent relaxation, *Circulation* 95 (1997) 723–731.

[3] V. Deckert, A. Brunet, F. Lantoin, G. Lizard, E.M.-v. Brussel, S. Monier, L. Lagrost, M. vid-Duflho, P. Gambert, M.A. Devynck, Inhibition by cholesterol oxides of NO release from human vascular endothelial cells, *Arteriosclerosis, Thrombosis, and Vascular Biology* 18 (1998) 1054–1060.

[4] G. Lizard, S. Guedry, V. Deckert, P. Gambert, L. Lagrost, Evaluation of the cytotoxic effects of some oxysterols and of cholesterol on endothelial cell growth: methodological aspects, *Pathologie Biologie* 45 (1997) 281–290.

[5] A. Vejux, E. Kahn, F. Menetrier, T. Montange, J. Lherminier, J.M. Riedinger, G. Lizard, Cytotoxic oxysterols induce caspase-independent myelin figure formation and caspase-dependent polar lipid accumulation, *Histochemistry and Cell Biology* 127 (2007) 609–624.

[6] J.R. Seckl, B.R. Walker, Minireview: 11β-hydroxysteroid dehydrogenase type 1 – a tissue-specific amplifier of glucocorticoid action, *Endocrinology* 142 (2001) 1371–1376.

[7] Y. Kotelevtsev, R.W. Brown, S. Fleming, C. Kenyon, C.R.W. Edwards, J.R. Seckl, J.J. Mullins, Hypertension in mice lacking 11β-hydroxysteroid dehydrogenase type 2, *Journal of Clinical Investigation* 103 (1999) 683–689.

[8] M. Wamil, R. Andrew, K.E. Chapman, J. Street, N.M. Morton, J.R. Seckl, 7-Oxysterols modulate glucocorticoid activity in adipocytes through competition for 11β-hydroxysteroid dehydrogenase type 1, *Endocrinology* 149 (2008) 5909–5918.

[9] M. Hult, B. Elleby, N. Shafiqat, S. Svensson, A. Rane, H. Jirnvall, L. Abrahamson, U. Oppermann, Human and rodent type 1 11β-hydroxysteroid dehydrogenases are 7β-hydroxycholesterol dehydrogenases involved in oxysterol metabolism, *Cellular and Molecular Life Sciences* 61 (2004) 992–999.

[10] R.A.S. Schweizer, Z. Marcel, Z. Balazs, D. Bernhard, A. Odermatt, Rapid hepatic metabolism of 7-ketocholesterol by 11β-hydroxysteroid dehydrogenase type 1: species-specific differences between the rat, human, and hamster enzyme, *Journal of Biological Chemistry* 279 (2004) 18415–18424.

[11] B.R. Walker, J.L. Yau, L.P. Brett, J.R. Seckl, C. Monder, B.C. Williams, C.R. Edwards, 11β-Hydroxysteroid dehydrogenase in vascular smooth muscle and heart: implications for cardiovascular responses to glucocorticoids, *Endocrinology* 129 (1991) 3305–3312.

[12] N.M. Morton, J.M. Paterson, H. Masuzaki, M.C. Holmes, B. Staels, C. Fievet, B.R. Walker, J.S. Flier, J.J. Mullins, J.R. Seckl, Novel adipose tissue-mediated resistance to diet-induced visceral obesity in 11β-hydroxysteroid dehydrogenase type 1-deficient mice, *Diabetes* 53 (2004) 931–938.

[13] Z. Krozowski, Z. Chai, The role of 11β-hydroxysteroid dehydrogenases in the cardiovascular system, *Endocrine Journal* 50 (2003) 485–489.

[14] C. Christy, P.W.F. Hadoke, J.M. Paterson, J.J. Mullins, J.R. Seckl, B.R. Walker, 11β-Hydroxysteroid dehydrogenase type 2 in mouse aorta: localization and influence on response to glucocorticoids, *Hypertension* 42 (2003) 580–587.

[15] A.R. Dover, P.W.F. Hadoke, L.J. Macdonald, E. Miller, D.E. Newby, B.R. Walker, Intra-vascular glucocorticoid metabolism during inflammation and injury in mice, *Endocrinology* 148 (2006) 166–172.

[16] Y. Kotelevtsev, M.C. Holmes, A. Burchell, P.M. Houston, D. Schmolli, P. Jamieson, R. Best, R. Brown, C.R. Edwards, J.R. Seckl, J.J. Mullins, 11β-Hydroxysteroid dehydrogenase type 1 knockout mice show attenuated glucocorticoid-

- inducible responses and resist hyperglycemia on obesity or stress, *Proceedings of the National Academy of Sciences* 94 (1997) 14924–14929.
- [17] G. Lizard, S. Monier, C. Cordelet, L. Gesquiere, V. Deckert, S. Gueldry, L. Lagrost, P. Gambert, Characterization and comparison of the mode of cell death, apoptosis versus necrosis, induced by 7 β -hydroxycholesterol and 7-ketocholesterol in the cells of the vascular wall, *Arteriosclerosis, Thrombosis, and Vascular Biology* 19 (1999) 1190–1200.
- [18] C.I. Seye, M.W.M. Knaapen, D. Daret, C. Desgranges, A.G. Herman, M.M. Kockx, H. Bult, 7-Ketocholesterol induces reversible cytochrome c release in smooth muscle cells in absence of mitochondrial swelling, *Cardiovascular Research* 64 (2004) 144–153.
- [19] R. Hynynen, M. Suchanek, J. Spandl, N. Back, C. Thiele, V.M. Olkkonen, OSBP-related protein 2 (ORP2) is a sterol receptor on lipid droplets that regulates the metabolism of neutral lipids, *Journal of Lipid Research* 50 (2009) 1305–1315.
- [20] P.W.F. Hadoke, C. Christy, Y.V. Kotelevtsev, B.C. Williams, C.J. Kenyon, J.R. Seckl, J.J. Mullins, B.R. Walker, Endothelial cell dysfunction in mice after transgenic knockout of type 2, but not type 1, 11 β -hydroxysteroid dehydrogenase, *Circulation* 104 (2001) 2832–2837.
- [21] K. Endo, T. Oyama, A. Saiki, N. Ban, M. Ohira, N. Koide, T. Murano, H. Watanabe, M. Nishii, M. Miura, K. Sekine, Y. Miyashita, K. Shirai, Determination of serum 7-ketocholesterol concentrations and their relationships with coronary multiple risks in diabetes mellitus, *Diabetes Research and Clinical Practice* 80 (2008) 63–68.
- [22] H.N. Hodis, S. Hashimoto, W.J. Mack, A. Sevanian, Probuco reduces oxysterol formation in hypertensive rabbits, *Hypertension* 36 (2000) 436–441.
- [23] W. Siems, S. Quast, D. Peter, W. Augustin, F. Carluccio, T. Grune, A. Sevanian, H. Hampl, I. Wiswedel, Oxysterols are increased in plasma of end-stage renal disease patients, *Kidney and Blood Pressure Research* 28 (2005) 302–306.
- [24] A.J. Brown, S. Leong, R.T. Dean, W. Jessup, 7-Hydroxycholesterol and its products in oxidized low density lipoprotein and human atherosclerotic plaque, *Journal of Lipid Research* 38 (1997) 1730–1745.
- [25] H.N. Hodis, D.W. Crawford, A. Sevanian, Cholesterol feeding increases plasma and aortic tissue cholesterol oxide levels in parallel: further evidence for the role of cholesterol oxidation in atherosclerosis, *Atherosclerosis* 89 (1991) 117–126.
- [26] N. Freeman, A. Rusinol, M. Linton, D. Hachey, S. Fazio, M. Sinensky, D. Thewke, Acyl-coenzyme A:cholesterol acyltransferase promotes oxidized LDL/oxysterol-induced apoptosis in macrophages, *Journal of Lipid Research* 46 (2005) 1933–1943.
- [27] G.R. Small, P.W.F. Hadoke, I. Sharif, A.R. Dover, D. Armour, C.J. Kenyon, G.A. Gray, B.R. Walker, Preventing local regeneration of glucocorticoids by 11 β -hydroxysteroid dehydrogenase type 1 enhances angiogenesis, *Proceedings of the National Academy of Sciences* 102 (2005) 12165–12170.
- [28] R. Best, S.M. Nelson, B.R. Walker, Dexamethasone and 11-dehydrodexamethasone as tools to investigate the isozymes of 11 β -hydroxysteroid dehydrogenase in vitro and in vivo, *Journal of Endocrinology* 153 (1997) 41–48.
- [29] S. Dzeletovic, A. Babiker, E. Lund, U. Diczfalussy, Time course of oxysterol formation during in vitro oxidation of low density lipoprotein, *Chemistry and Physics of Lipids* 78 (1995) 119–128.
- [30] S. Dzeletovic, O. Breuer, E. Lund, U. Diczfalussy, Determination of cholesterol oxidation products in human plasma by isotope dilution-mass spectrometry, *Analytical Biochemistry* 225 (1995) 73–80.
- [31] Z. Krozowski, S. Stuchberry, P.C. White, C. Monder, J.W. Funder, Characterisation of 11 β -hydroxysteroid dehydrogenase gene expression: identification of multiple unique forms of messenger ribonucleic acid in the rat kidney, *Endocrinology* 127 (1990) 3009–3013.
- [32] V. Deckert, L. Duverneuil, S. Poupon, S. Monier, N. Le Guern, G. Lizard, D. Masson, L. Lagrost, The impairment of endothelium-dependent arterial relaxation by 7-ketocholesterol is associated with an early activation of protein kinase C, *British Journal of Pharmacology* 137 (2002) 655–662.
- [33] G. Lizard, M. Moisan, C. Cordelet, S. Monier, P. Gambert, L. Lagrost, Induction of similar features of apoptosis in human and bovine vascular endothelial cells treated by 7-ketocholesterol, *The Journal of Pathology* 183 (2006) 330–338.
- [34] V. Deckert, Impairment of endothelium-dependent arterial relaxation by high-fat feeding in ApoE-deficient mice, *Circulation* 100 (1999) 1236–1235.
- [35] W. Martinet, D.M. Schrijvers, J.-P. Timmermans, H. Bult, Interactions between cell death induced by statins and 7-ketocholesterol in rabbit aorta smooth muscle cells, *British Journal of Pharmacology* 154 (2008) 1236–1246.
- [36] L. Kiss, M. Chen, D. Gero, K. Modis, Z. Lacza, C. Szabo, Effects of 7-ketocholesterol on the activity of endothelial poly(ADP-ribose) polymerase and on endothelium-dependent relaxant function, *International Journal of Molecular Medicine* 18 (2006) 1113–1117.
- [37] W. Jessup, P. Wilson, K. Gaus, L. Kritharides, Oxidized lipoproteins and macrophages, *Vascular Pharmacology* 38 (2002) 239–248.
- [38] A. Hermanowski-Vosatka, J.M. Balkovec, K. Cheng, H.Y. Chen, M. Hernandez, G.C. Koo, C.B. Le Grand, Z. Li, J.M. Metzger, S.S. Mundt, H. Noonan, C.N. Nunes, S.H. Olson, B. Pikounis, N. Ren, N. Robertson, J.M. Schaeffer, K. Shah, M.S. Springer, A.M. Strack, M. Strowski, K. Wu, T. Wu, J. Xiao, B.B. Zhang, S.D. Wright, R. Thieringer, 11 β -HSD1 inhibition ameliorates metabolic syndrome and prevents progression of atherosclerosis in mice, *The Journal of Experimental Medicine* 202 (2005) 517–527.
- [39] H. Larsson, Y. Bottiger, L. Iuliano, U. Diczfalussy, In vivo interconversion of 7 β -hydroxycholesterol and 7-ketocholesterol, potential surrogate markers for oxidative stress, *Free Radical Biology and Medicine* 43 (2007) 695–701.
- [40] W. Song, J. Chen, W.L. Dean, R.N. Redinger, R.A. Prough, Purification and characterization of hamster liver microsomal 7 α -hydroxycholesterol dehydrogenase, similarity to type I 11 β -hydroxysteroid dehydrogenase (11 β HSD1), *Journal of Biological Chemistry* 273 (1998) 16223–16228.
- [41] Y. Maeda, H. Nagatomo, F. Uchiyama, J. Nagatomo, M. Yamada, H. Shiotsuki, Y. Ohta, S. Sato, M.H. Kai, K.H. Kondo, A comparative study of the conversion of 7-hydroxycholesterol in rabbit, guinea pig, rat, hamster, and chicken, *Steroids* 67 (2002) 703–708.
- [42] C.D. DuSell, M. Umetani, P.W. Shaul, D.J. Mangelsdorf, D.P. McDonnell, 27-Hydroxycholesterol is an endogenous selective estrogen receptor modulator, *Molecular Endocrinology* 22 (2008) 65–77.
- [43] T. Mitić, S. Shave, S.P. Webster, I. McNae, D.F. Cobice, N. Semjonow, R.E. Dakin, K.E. Chapman, G.G. Lavery, P.W.F. Hadoke, B.R. Walker, R. Andrew, The role of 7-oxysterols to inhibit dehydrogenation of glucocorticoids by 11 β -hydroxysteroid dehydrogenase type 1, submitted for publication.
- [44] M.E. Ullian, *Cardiovascular Research* 41 (1999) 55–64.
- [45] D.G. Johns, A.M. Dorrance, N.L. Tramontini, R.C. Webb, Glucocorticoids inhibit tetrahydrobiopterin-dependent endothelial function, *Experimental Biology and Medicine* 226 (2001) 27–31.
- [46] M. Noshiro, E. Usui, T. Kawamoto, H. Kubo, K. Fujimoto, M. Furukawa, S. Honma, M. Makishima, K.-i. Honma, Y. Kato, Multiple mechanisms regulate circadian expression of the gene for cholesterol 7 α -hydroxylase (cyp7a), a key enzyme in hepatic bile acid biosynthesis, *Journal of Biological Rhythms* 22 (2007) 299–311.
- [47] D.F. Reilly, E.J. Westgate, G.A. FitzGerald, Peripheral circadian clocks in the vasculature, *Arteriosclerosis, Thrombosis, and Vascular Biology* 27 (2007) 1694–1705.
- [48] J. Rosenstock, S.E. Inzucchi, J. Seufert, P.R. Fleck, C.A. Wilson, Q. Mekki, Initial combination therapy with alogliptin and pioglitazone in drug-naïve patients with type 2 diabetes, *Diabetes Care* 33 (2010) 2406–2408.