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Genome Sequences of *Salmonella enterica* Serovar Typhimurium, Choleraesuis, Dublin, and Gallinarum Strains of Well-Defined Virulence in Food-Producing Animals[∇]

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***Salmonella enterica* is an animal and zoonotic pathogen of worldwide importance and may be classified into serovars differing in virulence and host range. We sequenced and annotated the genomes of serovar Typhimurium, Choleraesuis, Dublin, and Gallinarum strains of defined virulence in each of three food-producing animal hosts. This provides valuable measures of intraserovar diversity and opportunities to formally link genotypes to phenotypes in target animals.**

Salmonella enterica causes salmonellosis in humans and other warm-blooded animals. Over 2,600 serovars have been classified according to the reactivity of antisera to somatic lipopolysaccharide and flagellar antigens and are broadly grouped on the basis of host range and disease presentation. The molecular basis of the differential virulence and tropism of serovars remains ill defined (20). An understanding of such processes is required to develop strategies for disease control and to predict the threat posed by isolates from animals.

The extent to which currently sequenced strains are typical of the wider serovar is open to question. We report the sequencing and annotation of four strains representing serovars that produce significant illness in food-producing animals: *S. Typhimurium* strain ST4/74 (11), originally isolated from a calf with salmonellosis in the United Kingdom (17) and the parent of the widely used mouse virulent *hisG* auxotroph SL1344 (10); *S. Choleraesuis* var. *kunzendorf* strain SCSA50, a field isolate from a case of swine typhoid in the United Kingdom (3); *S. Dublin* strain SD3246, a Vi-negative isolate from a calf with systemic salmonellosis in the United Kingdom (24); and *S. Gallinarum* SG9, first described to cause fowl typhoid in orally dosed chickens by Smith in 1955 (19). Crucially, the virulence of each strain has been reciprocally compared in calves, pigs, and chickens (3, 4, 6, 14, 15, 16, 17, 24, 25, 26, 27), fulfilling Koch's postulates and enabling strain genotypes to be linked to phenotypes in target hosts.

Sequencing and annotation. 36 cycle paired-end sequencing was carried out on an Illumina GAIIx, yielding between 80 and 150X coverage. SOAPdenovo (13) was used to generate *de*

novo contigs, and reads aligned to a reference using Novoalign (Novocraft, Selangor, Malaysia). *S. Typhimurium* 4/74 reads were assembled on the genome and large plasmid of strain SL1344 (<http://www.sanger.ac.uk/Projects/Salmonella/>). *S. Choleraesuis* SCSA50 reads were assembled on the genome of strain SC-B67 (7) and its virulence plasmid (28). *S. Dublin* SD3246 reads were assembled on the genome of strain CT_02021853 (accession no. CP001144). *S. Gallinarum* SG9 reads were assembled on the genome of strain 287/91 (22). The *de novo* and reference contigs were combined using MUMmer (12) and Gap4 (5).

Sequences were annotated using GenoPipe (<http://genopipe.bioinfo-portal.cdac.in/>) and a combination of gene prediction software (1, 8, 18, 21). Manual curation followed to enhance the annotation, including pseudogene prediction and assignment of start sites. Genes with unsuitable names for submission were searched against SwissProt (23), and genes with a large degree of overlap were checked for domains (2, 9) and hits in SwissProt. If no domains or matches were found, the gene was removed from the annotation.

Intraserovar comparisons indicated that the complete *S. Typhimurium* 4/74 genome contained just eight single-nucleotide polymorphisms (SNPs) relative to SL1344, consistent with the shared history of the strains and high-quality sequencing and assembly. The *hisG* allele varied between the strains as expected (10).

Nucleotide accession numbers. Sequences were deposited in GenBank and assigned the following accession numbers; *S. Typhimurium* 4/74 (CP002487-CP002490), *S. Choleraesuis* SCSA50 (CM001062 to CM001063), *S. Dublin* SD3246 (CM001151 to CM001152), and *S. Gallinarum* SG9 (CM001153 to CM001154).

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REFERENCES

1. Altschul, S. F., W. Gish, W. Miller, E. W. Myers, and D. J. Lipman. 1990. Basic local alignment search tool. *J. Mol. Biol.* **215**:403–410.
2. Bateman, A., et al. 2004. The Pfam protein families database. *Nucleic Acids Res.* **32**:D138–D141.
3. Bolton, A. J., G. D. Martin, M. P. Osborne, T. S. Wallis, and J. Stephen. 1999. Invasiveness of Salmonella serotypes Typhimurium, Choleraesuis and Dublin for rabbit terminal ileum in vitro. *J. Med. Microbiol.* **48**:801–810.
4. Bolton, A. J., M. P. Osborne, T. S. Wallis, and J. Stephen. 1999. Interaction of Salmonella choleraesuis, Salmonella dublin and Salmonella typhimurium with porcine and bovine terminal ileum in vivo. *Microbiology* **145 (Pt 9)**: 2431–2441.
5. Bonfield, J. K., K. Smith, and R. Staden. 1995. A new DNA sequence assembly program. *Nucleic Acids Res.* **23**:4992–4999.
6. Chadfield, M. S., D. J. Brown, S. Aabo, J. P. Christensen, and J. E. Olsen. 2003. Comparison of intestinal invasion and macrophage response of Salmonella Gallinarum and other host-adapted Salmonella enterica serovars in the avian host. *Vet. Microbiol.* **92**:49–64.
7. Chiu, C. H., et al. 2005. The genome sequence of Salmonella enterica serovar Choleraesuis, a highly invasive and resistant zoonotic pathogen. *Nucleic Acids Res.* **33**:1690–1698.
8. Delcher, A. L., D. Harmon, S. Kasif, O. White, and S. L. Salzberg. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res.* **27**:4636–4641.
9. Finn, R. D., et al. 2010. The Pfam protein families database. *Nucleic Acids Res.* **38**:D211–D222.
10. Hoiseth, S. K., and B. A. Stocker. 1981. Aromatic-dependent Salmonella typhimurium are non-virulent and effective as live vaccines. *Nature* **291**:238–239.
11. Jones, P. W., P. Collins, and M. M. Aitken. 1988. Passive protection of calves against experimental infection with Salmonella typhimurium. *Vet. Rec.* **123**: 536–541.
12. Kurtz, S., et al. 2004. Versatile and open software for comparing large genomes. *Genome Biol.* **5**:R12.
13. Li, R., et al. 2010. De novo assembly of human genomes with massively parallel short read sequencing. *Genome Res.* **20**:265–272.
14. Paulin, S. M., A. Jagannathan, J. Campbell, T. S. Wallis, and M. P. Stevens. 2007. Net replication of Salmonella enterica serovars Typhimurium and Choleraesuis in porcine intestinal mucosa and nodes is associated with their differential virulence. *Infect. Immun.* **75**:3950–3960.
15. Paulin, S. M., et al. 2002. Analysis of Salmonella enterica serotype-host specificity in calves: avirulence of S. enterica serotype gallinarum correlates with bacterial dissemination from mesenteric lymph nodes and persistence in vivo. *Infect. Immun.* **70**:6788–6797.
16. Pullinger, G. D., et al. 2007. Systemic translocation of Salmonella enterica serovar Dublin in cattle occurs predominantly via efferent lymphatics in a cell-free niche and requires type III secretion system 1 (T3SS-1) but not T3SS-2. *Infect. Immun.* **75**:5191–5199.
17. Rankin, J. D., and R. J. Taylor. 1966. The estimation of doses of Salmonella typhimurium suitable for the experimental production of disease in calves. *Vet. Rec.* **78**:706–707.
18. Schattner, P., A. N. Brooks, and T. M. Lowe. 2005. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. *Nucleic Acids Res.* **33**:W686–W689.
19. Smith, H. W. 1955. Observations on experimental fowl typhoid. *J. Comp. Pathol.* **65**:37–54.
20. Stevens, M. P., T. J. Humphrey, and D. J. Maskell. 2009. Molecular insights into farm animal and zoonotic Salmonella infections. *Philos Trans. R. Soc. Lond. B Biol. Sci.* **364**:2709–2723.
21. Suzek, B. E., M. D. Ermolaeva, M. Schreiber, and S. L. Salzberg. 2001. A probabilistic method for identifying start codons in bacterial genomes. *Bioinformatics* **17**:1123–1130.
22. Thomson, N. R., et al. 2008. Comparative genome analysis of Salmonella Enteritidis PT4 and Salmonella Gallinarum 287/91 provides insights into evolutionary and host adaptation pathways. *Genome Res.* **18**:1624–1637.
23. UniProt Consortium. 2011. Ongoing and future developments at the Universal Protein Resource. *Nucleic Acids Res.* **39**:D214–D219.
24. Wallis, T. S., S. M. Paulin, J. S. Plested, P. R. Watson, and P. W. Jones. 1995. The Salmonella dublin virulence plasmid mediates systemic but not enteric phases of salmonellosis in cattle. *Infect. Immun.* **63**:2755–2761.
25. Watson, P. R., S. M. Paulin, A. P. Bland, P. W. Jones, and T. S. Wallis. 1995. Characterization of intestinal invasion by Salmonella typhimurium and Salmonella dublin and effect of a mutation in the invH gene. *Infect. Immun.* **63**:2743–2754.
26. Watson, P. R., S. M. Paulin, P. W. Jones, and T. S. Wallis. 2000. Interaction of Salmonella serotypes with porcine macrophages in vitro does not correlate with virulence. *Microbiology* **146**:1639–1649.
27. Wray, C., and W. J. Sojka. 1978. Experimental Salmonella typhimurium infection in calves. *Res. Vet. Sci.* **25**:139–143.
28. Yu, H., et al. 2006. Complete nucleotide sequence of pSCV50, the virulence plasmid of Salmonella enterica serovar Choleraesuis SC-B67. *Plasmid* **55**: 145–151.