



DIGITAL ACCESS TO SCHOLARSHIP AT HARVARD

The emerging role of metabolomics in the development of biomarkers for pulmonary hypertension and other cardiovascular diseases (2013 Grover Conference series)

The Harvard community has made this article openly available.
[Please share](#) how this access benefits you. Your story matters.

Citation	Lewis, Gregory D. 2014. "The emerging role of metabolomics in the development of biomarkers for pulmonary hypertension and other cardiovascular diseases (2013 Grover Conference series)." <i>Pulmonary Circulation</i> 4 (3): 417-423. doi:10.1086/677369. http://dx.doi.org/10.1086/677369 .
Published Version	doi:10.1086/677369
Accessed	February 17, 2015 12:29:59 PM EST
Citable Link	http://nrs.harvard.edu/urn-3:HUL.InstRepos:13890784
Terms of Use	This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

(Article begins on next page)

The emerging role of metabolomics in the development of biomarkers for pulmonary hypertension and other cardiovascular diseases (2013 Grover Conference series)

Gregory D. Lewis

Cardiology Division and Pulmonary and Critical Care Unit, Department of Medicine, Massachusetts General Hospital, Boston, Massachusetts, USA; and Harvard Medical School, Boston, Massachusetts, USA

Abstract: The functional and prognostic significance of pulmonary hypertension (PH) is well established, yet our understanding of circulating peptides and metabolites that might mark or contribute to various forms of PH remains limited. Metabolites are the end result of all the regulatory complexity present in a cell, tissue, or organism and therefore serve as the most proximal reporters of the body's response to a disease process or drug therapy. This review presents the rationale, methodology, and preliminary findings from studies that apply comprehensive metabolite profiling to gain knowledge of new circulating markers of PH.

Keywords: metabolite, pulmonary hypertension, exercise.

Pulm Circ 2014;4(3):417-423. DOI: 10.1086/677369.

The development of elevated pulmonary arterial pressure (PAP), either in unselected populations or in individuals with a variety of cardiopulmonary diseases, is associated with poor prognosis.¹⁻⁵ Pulmonary hypertension (PH, i.e., mean PAP > 25 mmHg) leads to right ventricular (RV) dysfunction that is closely associated with impaired exercise capacity⁶ and renal and hepatic dysfunction⁷ as well as morbidity⁸ and mortality.^{3,9} Despite the functional and prognostic significance of right ventricular–pulmonary vascular (RV-PV) dysfunction, peptides and metabolites that might mark or contribute to RV-PV dysfunction remain incompletely defined. Knowledge of new circulating markers of RV-PV dysfunction or susceptibility to RV-PV dysfunction may provide more precise estimates of risk while also defining the pathways perturbed in individual patients, revealing new targets for intervention and ultimately enabling an individualized approach to care.^{10,11}

To address unmet needs in disease biomarkers, investigators have turned to comprehensive profiling of large numbers of endogenous metabolites and proteins in biofluids, which has been termed “metabolomics” and “proteomics,” respectively (Figure 1). Metabolites are the end result of all the regulatory complexity present in a cell, tissue,

or organism, including transcriptional and translational regulation as well as posttranslational modifications. Therefore, metabolites are the most proximal reporters of the body's response to a disease process or drug therapy (Figure 2). As described below, emerging metabolomics techniques will allow us to “overlay” new biomarkers and multi-marker scores on existing cardiovascular disease diagnostic tests.¹² It is anticipated that some new markers will be uncorrelated with or “orthogonal” to existing markers, thus providing additional pathobiological insights and information for cardiovascular disease management.¹³ In addition to serving as disease biomarkers, circulating metabolites may themselves participate in previously unanticipated roles as regulatory signals with hormone-like functions.¹⁴

METHODS FOR METABOLIC PROFILING

Present estimates suggest that the human metabolome consists of approximately 3,000–5,000 endogenous small molecules (i.e., <1 kDa in size), as described in the Human Metabolome Project (<http://www.hmdb.ca>), and ~40,000 total metabolites, including exogenous compounds originating from nutrients, microbiota, drugs, and other sources.^{15,16} The metabolome spans a variety of chemical compound

Address correspondence to Dr. Gregory D. Lewis, Heart Failure and Cardiac Transplantation Unit, Massachusetts General Hospital, Bigelow 800, 55 Fruit Street, Boston, MA 02114, USA. E-mail: glewis@partners.org.

Submitted January 12, 2014; Accepted April 30, 2014; Electronically published August 1, 2014.

© 2014 by the Pulmonary Vascular Research Institute. All rights reserved. 2045-8932/2014/0403-0008. \$15.00.

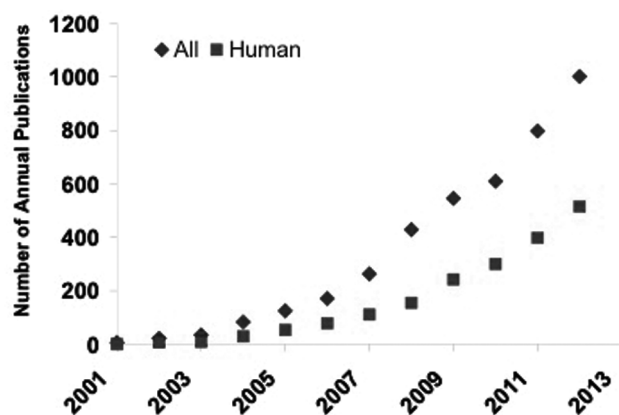


Figure 1. Rapid expansion of proteomics and metabolomics in biomedical research: the number of annual publications in metabolomics from 2001 to 2012. Criteria for inclusion of publications included the terms “metabolome” and “metabolomics.” A color version of this figure is available online.

classes, including those that are anionic versus cationic or lipophilic versus hydrophilic. Therefore, no single analytical method is capable of analyzing all metabolites.

Two core technologies are utilized to perform metabolic profiling: nuclear magnetic resonance (NMR) and tandem mass spectrometry (MS/MS). For recent reviews of metabolomics technologies, please see articles by Dunn,¹⁷ Kuehnbaum,¹⁸ and Ramautar.¹⁹ NMR requires relatively lit-

tle sample preparation and is nondestructive, allowing for subsequent structural analyses. However, the method tends to have low sensitivity and can detect only highly abundant analytes. MS/MS coupled with liquid chromatography (LC), on the other hand, has higher sensitivity for small molecules and is also applicable to a wide range of biological fluids (including serum, plasma, and urine). Recent advances in MS technology now enable researchers to determine analyte masses with such high precision and accuracy that metabolites can be identified unambiguously even in complex fluids.¹⁹

These technologies can be used to characterize biological samples either in a targeted manner or in a pattern discovery manner. In the former, the investigator targets a predefined set of metabolites for analysis. In the latter, the investigator is faced with a complex pattern of peaks, many of which are anonymous: the molecular identities of the species giving rise to the peaks are generally not known. While the targeted approach is more limited, the analysis is more straightforward, as the biochemicals giving rise to the signals have already been identified. An example of metabolite coverage by a targeted platform that is being actively applied to metabolic profiling of cardiovascular diseases²⁰⁻²³ is shown in Table 1. This platform can acquire the data for 290 metabolites in ~60 minutes from less than 200 μ L of plasma. As can be seen in Table 1, the platform

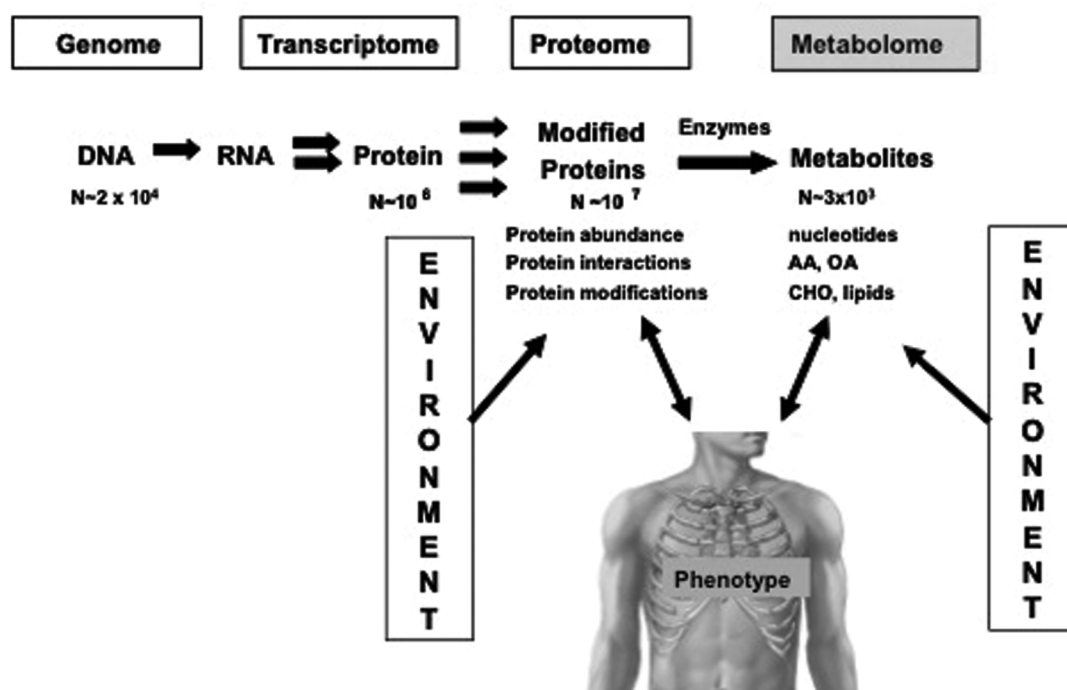


Figure 2. Integration of “omics” approaches and their relationship to phenotype. AA: amino acids; OA: organic acids; CHO: carbohydrates. A color version of this figure is available online.

Table 1. Approximate coverage of KEGG metabolism areas

Metabolism area	Human metabolites	
	Number	Coverage, %
Amino acid metabolism	370	21
Nucleotide metabolism	109	46
Carbohydrate metabolism	236	20
Energy metabolism	80	44
Metabolism of cofactors and vitamins	148	22
Metabolism of other amino acids	105	30
Biosynthesis of secondary metabolites	114	20
Lipid metabolism	284	15
Biodegradation of xenobiotics	180	6
Biosynthesis of polyketides and nonribosomal peptides	18	22
Glycan biosynthesis and metabolism	90	4

Note: KEGG: Kyoto encyclopedia of genes and genomes.¹⁶

provides nearly uniform coverage of all pathways in the KEGG database. Selection of metabolites for incorporation into our platform is based on (1) their biological diversity (i.e., “sentinels” in as many of the known metabolic pathways as possible), (2) their biological relevance (i.e., arginine-nitric oxide [NO] signaling pathway intermediates), and (3) their ability to be detected by LC-MS methods. The pattern discovery, or fingerprint, approach is inherently less biased, but the need for subsequent unambiguous identification of the peaks can be difficult.

STATISTICAL APPROACHES AND DATA REDUCTION FOR “OMICS” APPROACHES

Although high-throughput metabolomics approaches to biomarker discovery bring many advantages, they also bring the danger of generating false-positive associations as a result of multiple testing and overfitting of data. Replication is imperative to minimize overfitting of data.²⁴ Application of traditional statistical approaches (e.g., Bonferroni correction) in this setting tends to levy an insurmountable statistical penalty that can obscure biologically relevant associations. Newer statistical techniques,²⁵ such as advanced resampling methods or control of the false-discovery rate,²⁶ coupled with functional trend analysis in which changes in constituents of common pathways are analyzed together, can aid in detecting subtle but important changes in multiple variables identified in “omics” approaches.²⁷⁻²⁹ Several data reduction strategies can be used to construct multivariate metabolite biomarker profiles,³⁰ including discriminant

analysis,³¹ partial least squares, principal components analysis,³² and artificial neural networks.³³

APPLICATION OF METABOLOMICS FOR CARDIOVASCULAR BIOMARKER DISCOVERY

A very limited number of metabolites, such as glucose, lipids, creatinine, urea, and uric acid, have been used for decades to assess an individual’s (pre-)disease condition. In 1971, Arthur Robinson and Linus Pauling³⁴ conceived the core idea that information-rich data reflecting the functional status of a complex biological system resides in the quantitative and qualitative pattern of metabolites in body fluids. Currently, in the field of inborn errors of metabolism, an extensive repertoire of metabolites is used as biomarkers for diagnosis, progression, and response to treatment.³⁵ As might be expected, the application of metabolomics to complex cardiovascular diseases with multiple potential clinical confounders (i.e., medication exposure, age, gender, comorbidities) has been more difficult than that to Mendelian disorders.³⁶ Interindividual variability and adequacy of signal-to-noise intensity for certain metabolites represent additional hurdles in metabolomics studies.

To help circumvent these problems, investigators have applied these emerging technologies to unique clinical scenarios where serial sampling can be performed in patients both before and after a controlled perturbation, thereby allowing each patient to serve as his or her own biological control.^{20,21,29} As proof of principle, our group has applied targeted MS-based metabolomics to patients undergoing “planned myocardial ischemia,”^{21,22,29} elicited by exercise, and “planned myocardial infarction,”²⁰ induced by alcohol septal ablation in patients with hypertrophic obstructive cardiomyopathy. In both cases, robust metabolic changes were evident within minutes of the physiologic perturbation, when traditional markers of myocardial injury such as troponin remained unchanged.

A global metabolomics analysis of plasma revealed a pathway in both humans and mice linking microbiota metabolism of dietary choline and phosphatidylcholine to cardiovascular disease pathogenesis.³⁷ Metabolic profiling has also demonstrated distinct patterns of myocardial substrate utilization in individuals undergoing surgical coronary revascularization. Turer et al.³⁸ found that patients with left ventricular dysfunction demonstrated global suppression of metabolic fuel uptake and limited myocardial metabolic reserve and flexibility after global ischemia/reperfusion stress in the setting of cardiac surgery. Shah and colleagues³⁹ used a targeted MS/MS-based platform to profile 69 metabolites so as to evaluate suspected ischemic heart disease in more than 600 subjects from the Duke CATHGEN

biorepository who underwent cardiac catheterization. They demonstrated that peripheral blood metabolite profiles, one enriched for branched-chain amino acid metabolites and another comprising urea cycle metabolites, add to the discriminative capability for coronary artery disease, compared with models containing only clinical variables.

METABOLOMICS IN PH

The profiling of metabolites, including lipids, sugars, amino acids, and amines, is particularly relevant to RV-PV dysfunction. Some circulating metabolites (i.e., cyclic guanosine monophosphate [cGMP] and other NO pathway intermediates such as arginine, asymmetric dimethyl-arginine [ADMA], ornithine, citrulline, tryptophan hydroxylase metabolites, and catecholamines) have already been implicated in mediating pulmonary vascular tone, as described below. Altered metabolic substrate utilization, in the form of a mitochondrial metabolic switch from glucose oxidation to glycolysis, has been observed in the setting of PV remodeling,^{40,41} RV hypertrophy,⁴² and RV dysfunction,⁴³ offering further incentives to apply metabolite profiling to PH in left ventricular dysfunction. Through metabolomic profiling studies in PH, it will be important to ascertain the extent to which plasma metabolite levels reflect alterations in RV-PV metabolism while also evaluating small molecules with known vasoactive properties, as well as metabolites not previously associated with PH, as potential biomarkers of RV-PV dysfunction.

There is an emerging paradigm of PH as a systemic, multiorgan disease characterized by mitochondrial dysfunction.⁴⁴ Mitochondria integrate inputs regarding fuel supply (oxygen, glucose, lipids) and have recently been recognized to secrete mitokines that act on distant organs. This exciting work suggests that when mitochondria from one organ become dysfunctional, they can signal mitochondria from other tissues by releasing diffusible factors termed mitokines.⁴⁵ PH patients demonstrate insulin resistance in skeletal muscle analogous to observations in type 2 diabetes. This is particularly relevant because circulating metabolite clusters consisting of elevated levels of branched-chain and aromatic amino acids have been shown to independently predict future onset of insulin resistance and diabetes more than a decade before overt disease onset.²³

Recently, Fessel and colleagues⁴⁶ reported findings from metabolomic analyses of human pulmonary microvascular endothelial cells expressing two different disease-causing mutations in the bone morphogenetic protein receptor type 2 gene (*BMPR2*). This study confirmed previously described increases in glycolysis but also found significant upregulation of the pentose phosphate pathway, increases

in nucleotide salvage and polyamine biosynthesis pathways, decreases in fatty acid oxidation pathways, and impairment of tricarboxylic acid (TCA) cycle anaplerosis. The study went on to confirm increased isocitrate dehydrogenase activity in the setting of elevated isocitrate levels in *BMPR2*-mutant cells. This proof-of-principle study provides insight into potential metabolic pathways altered in RV-PV dysfunction beyond the primary focus to date on altered glucose homeostasis in PH.

Plasma-based metabolomics studies in PH are just beginning to emerge. One recent study in a porcine model of pulmonary embolism examined pre- versus postembolism plasma metabolite levels and found alterations in metabolites related to energy balance in hypoxic conditions, including glycolysis-derived metabolites, ketone bodies, and TCA cycle intermediates.⁴⁷ The extent to which easily measured circulating metabolite levels will reflect metabolic changes in experimental models requires further investigation. For example, physiologic perturbations such as hypoxia and exercise can result in increases in circulating TCA cycle intermediates,²¹ which may or may not reflect intracellular TCA cycle activity.

METABOLOMICS IN WHO GROUP 2 PH

There may be a particularly important role for application of metabolite profiling in furthering understanding of RV-PV dysfunction in the setting of left-heart disease (i.e., WHO group 2 PH). Heart failure (HF) due to left ventricular systolic dysfunction (LVSD) is commonly associated with the development of PH.⁶ However, the ability of the RV to respond to increased PAP at rest and during exercise is highly variable in HF and remains poorly understood.⁴⁸ Studies in both experimental models and patients have suggested a maladaptive imbalance between small-molecule pulmonary vasodilators and vasoconstrictors in HF. Altered NO signaling, including decreased NO synthase (NOS) bioavailability and increased production of endogenous small-molecule NOS inhibitors (i.e., ADMA), may contribute to the pathophysiology of PH in HF.⁴⁹ Like impaired NO responsiveness, decreased production of prostacyclin (PGI₂) and dysregulation of PGI₂ metabolism may also contribute to PH in LVSD.⁵⁰ Furthermore, insulin resistance with decreased peroxisome proliferator-activated receptor γ activation can accelerate adverse remodeling of the pulmonary vasculature in HF.⁵¹

Shao et al.⁵² recently reported that arginine-NO pathway intermediates can distinguish patients with differential burdens of WHO group 2 PH. Melenovsky et al.⁵³ found that impaired transpulmonary cGMP release relative to transpulmonary B-type natriuretic peptide uptake

was present in HF patients with high pulmonary vascular resistance and that phosphodiesterase-5 inhibition restored cGMP responsiveness. That study is an example of careful physiologic phenotyping with multisite sampling that can serve as a basis for potentially inferring local metabolism in the pulmonary circulation via measurement of circulating small molecules such as cGMP. While these studies provide promise that circulating markers of arginine-NO pathway intermediates associate with hemodynamic profiles, further investigations are needed to determine whether circulating metabolite levels from metabolic profiling will predict responses to therapies and prognosis in RV-PV dysfunction.

CONCLUSION

Although metabolomics technologies are still under development, they complement other functional genomic approaches, such as high-throughput genome sequencing and RNA expression analysis.⁵⁴ Together, they hold great promise to transform our ability to profile samples and discover multiple new biomarkers for RV-PV dysfunction. Novel multimarker approaches may also permit identification of alterations in specific pathways to inform the best preventative and therapeutic interventions for each individual, by restoring the specific perturbed pathways to their normal function.

Source of Support: Hassenfeld Clinical Scholar Award and grant R01HL119154 from the National Institutes of Health.

Conflict of Interest: None declared.

REFERENCES

- Di Salvo TG, Mathier M, Semigran MJ, Dec GW. Preserved right ventricular ejection fraction predicts exercise capacity and survival in advanced heart failure. *J Am Coll Cardiol* 1995;25(5):1143–1153.
- Lam CS, Roger VL, Rodeheffer RJ, Borlaug BA, Enders FT, Redfield MM. Pulmonary hypertension in heart failure with preserved ejection fraction: a community-based study. *J Am Coll Cardiol* 2009;53(13):1119–1126.
- Ghio S, Gavazzi A, Campana C, Inserra C, Klersy C, Sebastiani R, Arbustini E, Recusani F, Tavazzi L. Independent and additive prognostic value of right ventricular systolic function and pulmonary artery pressure in patients with chronic heart failure. *J Am Coll Cardiol* 2001;37(1):183–188.
- Lam CS, Borlaug BA, Kane GC, Enders FT, Rodeheffer RJ, Redfield MM. Age-associated increases in pulmonary artery systolic pressure in the general population. *Circulation* 2009;119(20):2663–2670.
- Bursi F, McNallan SM, Redfield MM, Nkomo VT, Lam CS, Weston SA, Jiang R, Roger VL. Pulmonary pressures and death in heart failure: a community study. *J Am Coll Cardiol* 2012;59(3):222–231.
- Butler J, Chomsky DB, Wilson JR. Pulmonary hypertension and exercise intolerance in patients with heart failure. *J Am Coll Cardiol* 1999;34(6):1802–1806.
- Korr KS, Gandsman EJ, Winkler ML, Shulman RS, Bough EW. Hemodynamic correlates of right ventricular ejection fraction measured with gated radionuclide angiography. *Am J Cardiol* 1982;49(1):71–77.
- Meluzin J, Špinarová L, Hude P, Krejčí J, Dušek L, Vitovec J, Panovsky R. Combined right ventricular systolic and diastolic dysfunction represents a strong determinant of poor prognosis in patients with symptomatic heart failure. *Int J Cardiol* 2005;105(2):164–173.
- de Groote P, Millaire A, Foucher-Hossein C, Nague O, Marchandise X, Ducloux G, Lablanche JM. Right ventricular ejection fraction is an independent predictor of survival in patients with moderate heart failure. *J Am Coll Cardiol* 1998;32(4):948–954.
- Cortese DA. A vision of individualized medicine in the context of global health. *Clin Pharmacol Ther* 2007;82(5):491–493.
- Braunwald E. Cardiology: the past, the present, and the future. *J Am Coll Cardiol* 2003;42(12):2031–2041.
- Zethelius B, Berglund L, Sundstrom J, Ingelsson E, Basu S, Larsson A, Venge P, Arnlov J. Use of multiple biomarkers to improve the prediction of death from cardiovascular causes. *N Engl J Med* 2008;358(20):2107–2116.
- Sabatine MS, Morrow DA, de Lemos JA, Gibson CM, Murphy SA, Rifai N, McCabe C, Antman EM, Cannon CP, Braunwald E. Multimarker approach to risk stratification in non-ST elevation acute coronary syndromes: simultaneous assessment of troponin I, C-reactive protein, and B-type natriuretic peptide. *Circulation* 2002;105(15):1760–1763.
- He W, Miao FJ, Lin DC, Schwandner RT, Wang Z, Gao J, Chen JL, Tian H, Ling L. Citric acid cycle intermediates as ligands for orphan G-protein-coupled receptors. *Nature* 2004;429(6988):188–193.
- Wishart DS, Jewison T, Guo AC, Wilson M, Knox C, Liu Y, Djoumbou Y, et al. HMDB 3.0—the Human Metabolome Database in 2013. *Nucleic Acids Res* 2013;41(D1):D801–D807.
- Kanehisa Laboratories. Kyoto encyclopedia of genes and genomes. 2010. <http://www.genome.jp/kegg/>.
- Dunn WB, Broadhurst DI, Atherton HJ, Goodacre R, Griffin JL. Systems level studies of mammalian metabolomes: the roles of mass spectrometry and nuclear magnetic resonance spectroscopy. *Chem Soc Rev* 2011;40(1):387–426.
- Kuehnbaum NL, Britz-McKibbin P. New advances in separation science for metabolomics: resolving chemical diversity in a post-genomic era. *Chem Rev* 2013;113(4):2437–2468.
- Ramautar R, Berger R, van der Greef J, Hankemeier T. Human metabolomics: strategies to understand biology. *Curr Opin Chem Biol* 2013;17(5):841–846.
- Lewis GD, Wei R, Liu E, Yang E, Shi X, Martinovic M, Farrell L, et al. Metabolite profiling of blood from individuals undergoing planned myocardial infarction reveals early markers of myocardial injury. *J Clin Invest* 2008;118(10):3503–3512.

21. Lewis GD, Farrell L, Wood MJ, Martinovic M, Arany Z, Rowe GC, Souza A, et al. Metabolic signatures of exercise in human plasma. *Sci Transl Med* 2010;2(33):33ra37.
22. Magnusson M, Lewis GD, Ericson U, Orho-Melander M, Hedblad B, Engström G, Östling G, et al. A diabetes-predictive amino acid score and future cardiovascular disease. *Eur Heart J* 2013;34(26):1982–1989.
23. Wang TJ, Larson MG, Vasani RS, Cheng S, Rhee EP, McCabe E, Lewis GD, et al. Metabolite profiles and the risk of developing diabetes. *Nat Med* 2011;17(4):448–453.
24. Ransohoff DF. Rules of evidence for cancer molecular-marker discovery and validation. *Nat Rev Cancer* 2004;4(4):309–314.
25. De Livera AM, Olshansky M, Speed TP. Statistical analysis of metabolomics data. In: Roessner U, Dias DA, eds. *Metabolomics: tools for natural product discovery*. *Methods Mol Biol* 2013;1055:291–307.
26. van der Greef J, Stroobant P, van der Heijden R. The role of analytical sciences in medical systems biology. *Curr Opin Chem Biol* 2004;8(5):559–565.
27. Mootha VK, Lindgren CM, Eriksson KF, Subramanian A, Sihag S, Lehar J, Puigserver P, et al. PGC-1 α -responsive genes involved in oxidative phosphorylation are coordinately down-regulated in human diabetes. *Nat Genet* 2003;34(3):267–273.
28. Berriz GF, King OD, Bryant B, Sander C, Roth FP. Characterizing gene sets with funcAssociate. *Bioinformatics* 2003;19(18):2502–2504.
29. Sabatine MS, Liu E, Morrow DA, Heller E, McCarroll R, Wiegand R, Berriz GF, Roth FP, Gerszten RE. Metabolomic identification of novel biomarkers of myocardial ischemia. *Circulation* 2005;112(25):3868–3875.
30. Villas-Bôas SG, Moxley JF, Åkesson M, Stephanopoulos G, Nielsen J. High-throughput metabolic state analysis: the missing link in integrated functional genomics of yeasts. *Biochem J* 2005;388(2):669–677.
31. Manly BFJ. *Multivariate statistical methods: a primer*. 3rd ed. Boca Raton, FL: Chapman & Hall/CRC, 2005.
32. Raamsdonk LM, Teusink B, Broadhurst D, Zhang N, Hayes A, Walsh MC, Berden JA, et al. A functional genomics strategy that uses metabolome data to reveal the phenotype of silent mutations. *Nat Biotechnol* 2001;19(1):45–50.
33. Bishop C. *Neural networks for pattern recognition*. Oxford: Clarendon, 1995.
34. Pauling L, Robinson AB, Teranishi R, Cary P. Quantitative analysis of urine vapor and breath by gas-liquid partition chromatography. *Proc Natl Acad Sci USA* 1971;68(10):2374–2376.
35. Wikoff WR, Gangoiti JA, Barshop BA, Siuzdak G. Metabolomics identifies perturbations in human disorders of propionate metabolism. *Clin Chem* 2007;53(12):2169–2176.
36. Kirschenlohr HL, Griffin JL, Clarke SC, Rhydwen R, Grace AA, Schofield PM, Brindle KM, Metcalfe JC. Proton NMR analysis of plasma is a weak predictor of coronary artery disease. *Nat Med* 2006;12(6):705–710.
37. Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, Dugar B, Feldstein AE, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* 2011;472(7341):57–63.
38. Turer AT, Stevens RD, Bain JR, Muehlbauer MJ, van der Westhuizen J, Mathew JP, Schwinn DA, Glower DD, Newgard CB, Podgoreanu MV. Metabolomic profiling reveals distinct patterns of myocardial substrate use in humans with coronary artery disease or left ventricular dysfunction during surgical ischemia/reperfusion. *Circulation* 2009;119(13):1736–1746.
39. Shah SH, Bain JR, Muehlbauer MJ, Stevens RD, Crosslin DR, Haynes C, Dungan J, et al. Association of a peripheral blood metabolic profile with coronary artery disease and risk of subsequent cardiovascular events. *Circ Cardiovasc Genet* 2010;3(2):207–214.
40. McMurtry MS, Bonnet S, Wu X, Dyck JR, Haromy A, Hashimoto K, Michelakis ED. Dichloroacetate prevents and reverses pulmonary hypertension by inducing pulmonary artery smooth muscle cell apoptosis. *Circ Res* 2004;95(8):830–840.
41. Xu W, Koeck T, Lara AR, Neumann D, DiFilippo FP, Koo M, Janocha AJ, et al. Alterations of cellular bioenergetics in pulmonary artery endothelial cells. *Proc Natl Acad Sci USA* 2007;104(4):1342–1347.
42. Takeyama D, Kagaya Y, Yamane Y, Shiba N, Chida M, Takahashi T, Ido T, Ishide N, Takishima T. Effects of chronic right ventricular pressure overload on myocardial glucose and free fatty acid metabolism in the conscious rat. *Cardiovasc Res* 1995;29(6):763–767.
43. Piao L, Fang YH, Cadete VJ, Wietholt C, Urboniene D, Toth PT, Marsboom G, et al. The inhibition of pyruvate dehydrogenase kinase improves impaired cardiac function and electrical remodeling in two models of right ventricular hypertrophy: resuscitating the hibernating right ventricle. *J Mol Med* 2010;88(1):47–60.
44. Sutendra G, Michelakis ED. The metabolic basis of pulmonary arterial hypertension. *Cell Metab* 2014;19(4):558–573.
45. Durieux J, Wolff S, Dillin A. The cell-non-autonomous nature of electron transport chain-mediated longevity. *Cell* 2011;144(1):79–91.
46. Fessel JP, Hamid R, Wittmann BM, Robinson LJ, Blackwell T, Tada Y, Tanabe N, Tatsumi K, Hemnes AR, West JD. Metabolomic analysis of bone morphogenetic protein receptor type 2 mutations in human pulmonary endothelium reveals widespread metabolic reprogramming. *Pulm Circ* 2012;2(2):201–213.
47. Bujak R, García-Álvarez A, Rupérez FJ, Nuño-Ayala M, García A, Ruiz-Cabello J, Fuster V, Ibáñez B, Barbas C. Metabolomics reveals metabolite changes in acute pulmonary embolism. *J Proteome Res* 2014;13(2):805–816.
48. Lewis GD, Murphy RM, Shah RV, Pappagianopoulos PP, Malhotra R, Bloch KD, Systrom DM, Semigran MJ. Pulmonary vascular response patterns during exercise in left ventricular systolic dysfunction predict exercise capacity and outcomes. *Circ Heart Fail* 2011;4(3):276–285.
49. Pullamsetti S, Kiss L, Ghofrani HA, Voswinkel R, Haredza P, Klepetko W, Aigner C, et al. Increased levels and reduced catabolism of asymmetric and symmetric dimethylarginines in pulmonary hypertension. *FASEB J* 2005;19(9):1175–1177.
50. Galiè N, Manes A, Branzi A. Prostanoids for pulmonary arterial hypertension. *Am J Respir Med* 2003;2(2):123–137.

51. Rabinovitch M. PPAR γ and the pathobiology of pulmonary arterial hypertension. In: Yuan JXJ, Ward JPT, eds. Membrane receptors, channels and transporters in pulmonary circulation. *Adv Exp Med Biol* 2010;661:447–458.
52. Shao Z, Wang Z, Shrestha K, Thakur A, Borowski AG, Sweet W, Thomas JD, Moravec CS, Hazen SL, Tang WH. Pulmonary hypertension associated with advanced systolic heart failure: dysregulated arginine metabolism and importance of compensatory dimethylarginine dimethylaminohydrolase-1. *J Am Coll Cardiol* 2012;59(13):1150–1158.
53. Melenovsky V, Al-Hiti H, Kazdova L, Jabor A, Syrovatka P, Malek I, Kettner J, Kautzner J. Transpulmonary B-type natriuretic peptide uptake and cyclic guanosine monophosphate release in heart failure and pulmonary hypertension: the effects of sildenafil. *J Am Coll Cardiol* 54(7):595–600.
54. Shah SH, Hauser ER, Bain JR, Muehlbauer MJ, Haynes C, Stevens RD, Wenner BR, et al. High heritability of metabolomic profiles in families burdened with premature cardiovascular disease. *Mol Syst Biol* 2009;5:258.