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Citation	Albertine, Jennifer M., William J. Manning, Michelle DaCosta, Kristina A. Stinson, Michael L. Muilenberg, and Christine A. Rogers. 2014. "Projected Carbon Dioxide to Increase Grass Pollen and Allergen Exposure Despite Higher Ozone Levels." PLoS ONE 9 (11): e111712. doi:10.1371/journal.pone.0111712. http://dx.doi.org/10.1371/journal.pone.0111712.
Published Version	doi:10.1371/journal.pone.0111712
Accessed	February 17, 2015 5:51:27 AM EST
Citable Link	http://nrs.harvard.edu/urn-3:HUL.InstRepos:13454693
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Projected Carbon Dioxide to Increase Grass Pollen and Allergen Exposure Despite Higher Ozone Levels



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Abstract

One expected effect of climate change on human health is increasing allergic and asthmatic symptoms through changes in pollen biology. Allergic diseases have a large impact on human health globally, with 10-30% of the population affected by allergic rhinitis and more than 300 million affected by asthma. Pollen from grass species, which are highly allergenic and occur worldwide, elicits allergic responses in 20% of the general population and 40% of atopic individuals. Here we examine the effects of elevated levels of two greenhouse gases, carbon dioxide (CO₂), a growth and reproductive stimulator of plants, and ozone (O₃), a repressor, on pollen and allergen production in Timothy grass (Phleum pratense L.). We conducted a fully factorial experiment in which plants were grown at ambient and/or elevated levels of O₃ and CO₂, to simulate present and projected levels of both gases and their potential interactive effects. We captured and counted pollen from flowers in each treatment and assayed for concentrations of the allergen protein, Phl p 5. We found that elevated levels of CO₂ increased the amount of grass pollen produced by \sim 50% per flower, regardless of O_3 levels. Elevated O_3 significantly reduced the PhI p 5 content of the pollen but the net effect of rising pollen numbers with elevated CO₂ indicate increased allergen exposure under elevated levels of both greenhouse gases. Using quantitative estimates of increased pollen production and number of flowering plants per treatment, we estimated that airborne grass pollen concentrations will increase in the future up to \sim 200%. Due to the widespread existence of grasses and the particular importance of *P. pratense* in eliciting allergic responses, our findings provide evidence for significant impacts on human health worldwide as a result of future climate change.

Citation: Albertine JM, Manning WJ, DaCosta M, Stinson KA, Muilenberg ML, et al. (2014) Projected Carbon Dioxide to Increase Grass Pollen and Allergen Exposure Despite Higher Ozone Levels. PLoS ONE 9(11): e111712. doi:10.1371/journal.pone.0111712

Editor: Tai Wang, Institute of Botany, Chinese Academy of Sciences, China

Received June 4, 2014; Accepted September 24, 2014; Published November 5, 2014

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Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the Supporting Information files.

Funding: The authors have no support or funding to report.

Competing Interests: The authors have declared that no competing interests exist.

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Introduction

One expected effect of climate change on human health is increasing allergic and asthmatic symptoms through changes in pollen biology [1–5]. Allergic diseases have a large impact on human health globally, with 10–30% of the population affected by allergic rhinitis and more than 300 million affected by asthma [6]. Pollen from grass species, which are highly allergenic and occur worldwide, elicits allergic responses in 20% of the general population and 40% of atopic individuals [7].

Climate change is noticeably affecting plant, animal and human systems and is anticipated to have large impacts on human health [4], [5], [8], [9]. A major concern is that wind-borne pollen, a primary cause of allergic rhinitis, may change in timing, amount, and allergenicity with future climate change, and may increase both the symptom severity and number of people affected [1], [3], [5], [9], [10], [11]. This concern is particularly relevant for grasses, which are widely distributed over the globe and affect a large number of sensitized individuals [1], [4], [12]. In fact, peaks in atmospheric grass pollen have been directly correlated to

ambulance calls by patients under respiratory stress [13] and ER visits for asthma and wheeze [14].

Here we examine the effects of elevated levels of two greenhouse gases, carbon dioxide (CO_2), a growth and reproductive stimulator of plants, and ozone (O_3), a repressor, on pollen and allergen production in Timothy grass (*Phleum pratense* L.).

Atmospheric CO_2 levels resulting from anthropogenic sources, a major driver of climate change, are expected to rise from around 400 ppm currently to 730–1020 ppm by the year 2100 [15]. Likewise, tropospheric O_3 has been predicted to rise from current background levels of about 30–40 ppb to 42–84 ppb by 2100 [15], [16]. These greenhouse gases have been shown to affect plant growth in contrasting ways. Carbon dioxide has been well characterized as stimulating plant growth through increased photosynthetic carbon assimilation [17]. Ozone has been shown to decrease growth due to oxidative damage of photosynthetic components [18]. Together these two gases act antagonistically on plants, the degree to which depends highly on plant species and other environmental factors [18].

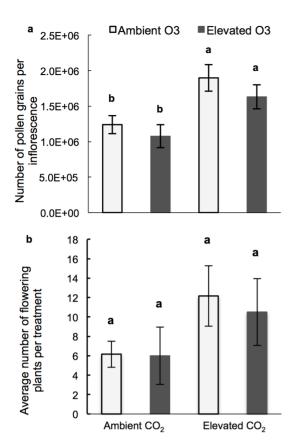


Figure 1. Effects on flowering and pollen production. a) Average pollen number per inflorescence. **b**) Average number of flowering plants per treatment. Ambient $O_3 = 30$ ppb ozone; Elevated $O_3 = 80$ ppb ozone; Ambient $CO_2 = 400$ ppm carbon dioxide; Elevated $CO_2 = 800$ ppm carbon dioxide. Significant differences (p<0.05) denoted by letters above bars determined by the Tukey-Kramer test. doi:10.1371/journal.pone.0111712.g001

The effects of elevated CO₂ on the production and allergen content of grass pollen have not been examined to date [4], although increases in pollen production 55–90% have been reported in other allergenic plants, such as ragweed (Ambrosia artemisiifolia L.), at elevated CO₂ [9], [19], [20]. Elevated CO₂ has also been shown to increase the reproductive output of ragweed populations through increasing the reproduction of subordinate plants in a competitive stand [21]. In a meta-analysis, C_3 and C_4 grass species demonstrated 44% and 33% growth stimulation, respectively, in response to elevated CO₂ [22]. It is therefore likely that elevated CO2 could also increase pollen output in grasses. In vivo exposure to O₃ has been shown to reduce amount of viable pollen by preventing pollen maturation through reduced anther starch content in perennial rye grass (Lolium perenne L.) and increase the group 5 allergen content of the pollen [23], [24]. Ozone has also been shown to decrease pollen viability of in vitro exposed Timothy grass pollen through disruption of the cell membrane; however, this disruption could increase allergen exposure by releasing cytoplasmic allergencontaining granules from within pollen [25], [26]. Finally, allergenicity has been found to decrease in Timothy grass pollen from reduced IgE recognition due to mechanical damage and post translational modifications when pollen was exposed in vitro to the mixture of air pollutants: ozone, sulfur dioxide, and nitrogen dioxide [26]. While these studies suggest that there could be a reduction in pollen viability and change in allergen quality at

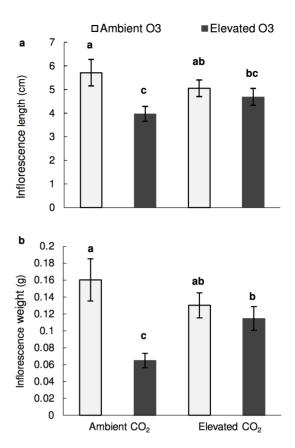


Figure 2. Effect on inflorescence size. a) inflorescence length (cm) **b)** Inflorescence weight (g) Ambient $O_3 = 30$ ppb ozone; Elevated $O_3 = 80$ ppb ozone; Ambient $CO_2 = 400$ ppm carbon dioxide; Elevated $CO_2 = 800$ ppm carbon dioxide. Significant differences (p<0.05) denoted by letters above bars were determined using the Tukey-Kramer test. doi:10.1371/journal.pone.0111712.g002

projected O_3 levels, it is not clear how these effects would interact with expected stimulatory effects of CO_2 on growth and reproduction in grasses.

Timothy grass (*Phleum pratense* L.) is a widespread perennial C₃ grass species used in agriculture and found growing naturally throughout temperate zones of North America and Europe. It produces a single inflorescence per plant with abundant amounts of easily aerosolized pollen, and is a major cause of early summer allergies. The allergens in Timothy grass pollen are similar to, and cross-reactive with, allergens from many other grass taxa; hence Timothy grass pollen extracts are often used in skin testing for broadly diagnosing grass allergy [7], [12]. The processes that affect these proteins in Timothy grass may also be representative of responses in other grass taxa.

In this study we determined the interactive effects of CO_2 and O_3 at current levels (400 ppm CO_2 , 30 ppb O_3) and at projected elevated levels (800 ppm CO_2 , 80 ppb O_3) in a full factorial design, on both the amount of pollen produced and the concentration of Phl p 5 protein, the major allergen in Timothy grass pollen. Plant exposure to gas treatments was entirely *in vivo* in order to assess whole plant responses. On each plant we measured: number of pollen grains per inflorescence, concentration of Phl p 5 per inflorescence and per pollen grain, inflorescence weight, inflorescence length, and number of inflorescences produced. Using generalized linear model approaches, we examined the effects of the factorial treatments on each parameter, and on the relationships between parameters.

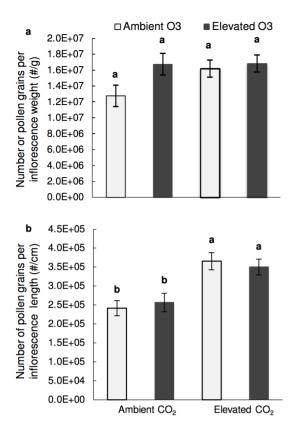


Figure 3. Relationship between pollen amount and inflorescence size. a) Pollen Number per length of inflorescence b) Pollen number per gram of inflorescence. Ambient $O_3 = 30$ ppb ozone; Elevated $O_3 = 80$ ppb ozone; Ambient $CO_2 = 400$ ppm carbon dioxide; Elevated $CO_2 = 800$ ppm carbon dioxide. Significant differences (p< 0.05) denoted by letters above bars were determined using the Tukey-Kramer test.

doi:10.1371/journal.pone.0111712.g003

Methods

Seeds of Phleum pratense L. var. CLIMAX were sown in 10.5 cm×10.5 cm×35 cm pots (Treepots, Hummert International, Missouri, USA) at the rate of 20 seeds per pot using MM 200 growing medium (SunGro, Washington, USA). At seeding, equal numbers of pots were placed per treatment in continuously-stirred tank reactor (CSTRs) chambers [27] and plants were allowed to germinate and grow through maturity. Grass was fertilized weekly with a dilute concentration of Peat Light Special (1/3 concentration; 15-16-17; Peter's Professional; Scotts, Ohio USA) and watered as needed. Plants were exposed to natural light since chambers were located in a single greenhouse, however light was also supplemented with 400W metal Halide lights (Metal Arc, Sylvania, Massachusetts, USA) which were on 12- hours/day for the first five weeks, then on for 16-hours/day for the remainder of the experiment. Greenhouse temperatures ranged from 15.5°C-26°C. Chambers were monitored for temperature and relative humidity levels using Hoboware data loggers (Onset Computer Corp; Massachusetts, USA).

Eight CSTRs [27] chambers were assigned to factorial treatments with either 30 ppb or 80 ppb ozone and either 400 ppm or 800 ppm carbon dioxide, to simulate present and future projected levels of both gases, respectively. Carbon dioxide treatments were administered continuously while ozone treatments were applied 9:00–16:00 daily to better simulate natural exposure

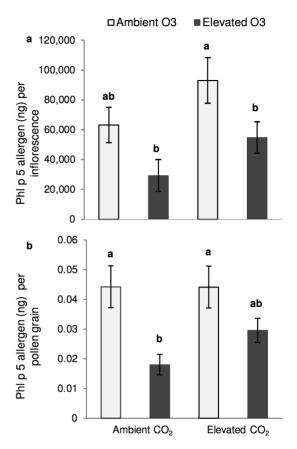


Figure 4. Phl p 5 allergen content. a) Average Phl p 5 concentration per inflorescence **b**) Average Phl p 5 concentration per pollen Ambient $O_3 = 30$ ppb ozone; Elevated $O_3 = 80$ ppb ozone; Ambient $CO_2 = 400$ ppm carbon dioxide; Elevated $CO_2 = 800$ ppm carbon dioxide. Significant differences (p<0.05) denoted by letters above bars determined by the Tukey-Kramer test. doi:10.1371/journal.pone.0111712.g004

cycles. Experiments were repeated three times for a total of six replications (two replications per experiment); each replication was randomized in chambers and blocked by experiment (two block per experiment) in analysis to account for changes in environmental variables (light, temperature) across repeated experiments.

To capture pollen, polyethylene bags were placed over flowers upon emergence and held open. Bag and flower spike were removed following complete dehiscence of pollen and stored at - $20^{\circ}\mathrm{C}$ until analysis.

Bagged flowers were washed three times with PBS-T (Phosphate Buffered Saline with 0.05% Tween 20 pH 7.4) to remove pollen; the amount of PBS-T was determined based on flower length and was measured precisely using a micro-pipette. Protein was extracted from the pollen by incubating at room temperature in the PBS-T solution for 2 hours. Pollen was separated by centrifugation at RCF 2,200×g. The extracted protein supernatant was stored at -20° C until allergen analysis. The pelleted pollen was suspended in a precisely measured volume of PBS-T solution and counted using a hemocytometer (3 times and averaged).

The Phl p 5 allergen content was determined using a monoclonal sandwich enzyme-linked immunosorbent assay (ELISA) (Indoor Biotechnologies, Inc., Charlottesville, VA, USA) using a mouse monoclonal IgG1 primary antibody to Phl p 5a&b, a 1% bovine serum albumin in PBS-T blocking agent, and a biotinolated mouse IgG1 secondary detection antibody to Phl p

5a&b. Concentrations were determined calorimetrically using streptavidin-peroxidase and substrate 1 mM ABTS (2,2'-azino-di-(3 ethylbenzthiazoline sulfonic acid)) in 70 mM citrate- phosphate buffer (pH 4.2). Serial dilutions of protein extracts were compared to a standard curve to quantify Phl p 5 allergen in each sample.

Analysis of variance of experimental data was used to account for treatment differences and was performed using a generalized linear model in SAS 9.3 (SAS Institute Inc, North Carolina, USA). Differences between treatment means were determined using Tukey- Kramer post-hoc test to account for uneven replications in response to treatments. Linear regressions were performed using analysis of covariance (ANCOVA) to evaluate relationship between co-factors.

Results

Elevated CO_2 (800 ppm) increased the amount of pollen produced by individual inflorescences by $\sim 53\%$ whereas elevated O_3 had no effect on the amount of pollen produced (Figure 1a; File S1). Since each plant only produces one inflorescence, the pollen output per plant was increased. The number of plants flowering per chamber was increased by elevated CO_2 , with and without elevated O_3 , during the experiment but was not statistically significant (Figure 1b).

Elevated CO_2 did not stimulate an increase in inflorescence length or weight as has been found in ragweed (Figure 2a) [19]. However, our findings suggest that stimulation by CO_2 offsets ozone damage and partially ameliorates O_3 —induced reductions in inflorescence size (Figure 2a, 2b). Furthermore, plants grown at elevated CO_2 and O_3 did not differ in the amount of pollen per weight (gram) of inflorescence than the control (Figure 3a). However, inflorescences of plants grown at elevated CO_2 regardless of O_3 level produced significantly more pollen per length of inflorescence (Figure 3b).

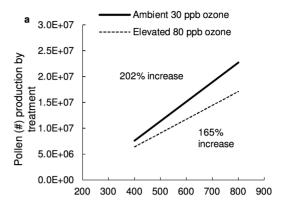
Concentrations of Phl p 5 allergen per inflorescence and per pollen grain were not affected by CO_2 levels but were reduced by elevated O_3 . (Figure 4a, 4b). Again, elevated CO_2 partially ameliorated this reduction by elevated O_3 (Figure 4b).

Finally, to determine the overall changes in pollen and allergen production between current and projected CO_2 and O_3 , we multiplied the average number of inflorescences produced per treatment by the number of pollen grains produced per inflorescence in that treatment (Figure 5a). Linear regression analysis determined a significant increase in pollen production of approximately 200% of current levels of CO_2 to future predicted levels for both O_3 treatments. The increase was only slightly affected by elevated O_3 (202% at ambient O_3 , 165% at elevated O_3), indicating that we can expect a large increase in pollen production due to increased CO_2 regardless of future ozone levels.

Likewise, for Phl p 5 allergen levels, we multiplied the concentration of Phl p 5 per inflorescence by the average number of inflorescences per treatment. Linear regression analysis indicated a significant allergen increase at high $\rm CO_2$ concentrations with a larger 190% increase in Phl p 5 allergen production at ambient $\rm O_3$ than the 48% increase at elevated $\rm O_3$ (Figure 5b). Although the rate of increase of Phl p 5 concentration in response to elevated $\rm CO_2$ was slowed by elevated $\rm O_3$, the strong $\rm CO_2$ -stimuation of pollen production suggests increased exposure to Timothy grass allergen overall.

Discussion

We provide the first evidence that pollen production is significantly stimulated by elevated carbon dioxide in grasses, a finding that expands earlier work on the annual forb, common ragweed [9], [19] to another important allergenic plant taxonomic



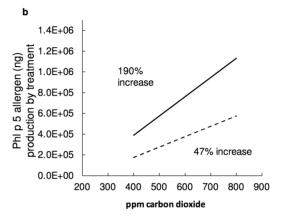


Figure 5. Predicted increases in pollen and allergen. a) Predicted increase in pollen production calculated by multiplying the average number of inflorescences in each treatment by the number of pollen produced per inflorescence. CO_2 by O_3 interaction is not significant indicating equal rates of increase with increasing CO_2 **b)** Predicted increase in PhI p 5 allergen production calculated by multiplying the average number of inflorescences per treatment by the concentration of PhI p 5 per Inflorescence. The CO_2 by O_3 interaction was significant (ANCOVA p<0.05) indicating that elevated O_3 reduced the rate of increase of PhI p 5 in response to elevated CO_2 . Solid line (–) determines trend at current ambient level of ozone (30 ppb). Dashed line (– --) is future elevated level of ozone (80 ppb). Percentage number on graph indicates the percent increase in production with increasing CO_2 levels.

doi:10.1371/journal.pone.0111712.g005

group. The inflorescences produced more pollen without increasing inflorescence size, indicating a greater number of male flowers per inflorescence and/or that male flowers in the inflorescence produced more pollen per anther.

The number of plants flowering per chamber was increased by elevated O_3 or elevated CO_2 during the experiment but was not statistically significant (Figure 1b). However, others have found an increased number of plants flowering in Timothy grass in response to elevated CO_2 , while elevated O_3 had no impact on flowering [28]. In our study, the trend toward more plants flowering at elevated CO_2 in Figure 1b indicates, however, that stimulation of flowering in Timothy grass warrants further research. Moreover, rising temperatures related to elevated CO_2 may lengthen the flowering season thereby further increasing atmospheric pollen loads [19], [29], [30].

Biomass allocation in response to elevated carbon dioxide may be an important factor in pollen production [9], [20], [21]. In a concurrent study, biomass allocation was analyzed at different growth stages over the Timothy grass life cycle [31]. It was found that the highest level of biomass stimulation, particularly in the photosynthetic leaf tissue, occurred in the tillering stages leading up to the flowering stage, which could have contributed to increased energy available for pollen production during flowering.

In a previous study, ragweed increased Amb a 1 production, the major allergen in the pollen coat, in response to elevated levels of CO_2 [10]. Ragweed differs in physiology from grasses; for instance, it is an annual plant, a dicot, and can be self-compatible, while grasses are perennial, monocots, and self-incompatible. The physiological importance of these two allergens may differ across these species [32]. Another explanation for lack of stimulation on Phl p 5 production could be that our work was conducted using moderate levels of nitrogen (\sim 5% N applied in solution once a week) and Phl p 5 protein production was potentially limited by nitrogen availability [17], [33].

Our experiment differed from other work on ozone in that we exposed our plants from seeding through dehiscence as would occur naturally with background O3 levels, while other experiments used more artificial exposures and durations [23–26]. In our study, the number of pollen grains produced per inflorescence was unaffected by O₃; however, we did not test the viability of the pollen nor quantitate the maturity of the pollen, so we do not know the impacts on its function. There is a chance that our study underestimates the treatment effects on allergen concentration due to a change in recognition of the allergen by the monoclonal antibodies in our ELISA assay. Since ozone is a strong oxidant known to quickly degrade fatty acids and double bonds found in proteins [34], oxidative damage could have altered the Phl p 5 protein resulting in reduced recognition by the mouse IgG antibodies used in ELISA. Rogerieux et al. [26] came to similar conclusions, using IgE antibodies, when looking at multiple individual allergens found in Timothy grass. This degradation of allergen by ozone would also happen in the natural environment impacting recognition of allergens by human immune systems and hence is not exclusively an artifact of the experiment.

The implications of increasing CO_2 for human health are clear. Stimulation of pollen production will increase airborne concentrations and increase exposure and suffering in pollen allergic individuals. The additional effects of increasing ozone are more complex. Ozone alone impacts human health negatively and has been shown to exacerbate the allergic airway response through irritation of the mucus membrane in respiratory pathways [35].

References

- Beggs PJ (2004) Impacts of climate change on aeroallergens: past and future. Clim Exp Allergy 34: 1507–1513.
- Beggs JP, Bambrick HJ (2005) Is the global rise of asthma an early impact of anthropogenic climate change? Environ Health Perspect 113: 915–919.
- Cecchi L, D'Amato G, Ayres JG, Galan C, Forastiere F, et al. (2010) Projections on the effects of climate change on allergenic asthma: the contributions of aerobiology. Allergy 651: 1073–1081.
- Gamble JL, Reid CE, Post E, Sacks J (2008) A Review of the Impacts of Climate Variability and Change on Aeroallergens and their Associated Effects-Final Report of the Global Change Research Program: US EPA.
- Ziska LH, Epstein PR, Rogers CA (2008) Climate change, aerobiology, and public health in the Northeast United States. Mitig Adapt Strat Glob Change 13: 607–613.
- Pawankar R, Canonica GW, Holgate ST, Lockey RF (2011) World Allergy Organization White Book on Allergy 2011–2012: Executive Summary. World Allergy Organization.
- Andersson K, Lidholm J (2003) Characteristics and Immunobiology of Grass Pollen Allergens. Int Arch Allergy Immunol 130: 87–107.
- Shea KM, Truckner RT, Weber RW, Peden DB (2008) Climate Change and allergic disease. Clin Rev Allergy Immunol 122: 443

 –451.
- Ziska LH, Caulfield FA (2000) Rising CO₂ and pollen production of common ragweed (Ambrosia artemisiifolia L.), a known allergy- inducing species; implications for public health. Aust J Plant Physiol 27: 893–898.

The projected O_3 levels are nearing the current US National Ambient Air Quality standard (80 ppb pre-2008 and 75 ppb post-2008) used to protect human health [36]. Hence elevated levels would likely elicit negative respiratory health effects independent of any additional indirect health effects as a result of elevated CO_2 . The IPCC AR5 predicts lower O_3 levels in some areas in the future due to air pollution mitigation strategies to reduce precursors to O_3 and also climate feedbacks that affect O_3 formation [37]. While increasing levels would increase airway irritation by ozone, lower levels of O_3 would mean sustained levels of Phl p 5 allergen with higher Timothy grass pollen production resulting in increased allergen exposure and the potential for further worsening of allergic symptoms.

Conclusion

Taken together, our results strongly suggest significant increases in grass pollen production and allergen exposure under future predicted levels of CO_2 and O_3 of 165–202%. Due to the widespread existence of grasses and their importance in eliciting allergic responses, these results indicate there will be a significant impact on human health worldwide as a result of future climate change.

Supporting Information

File S1 Data File. Raw experimental data of flower number, flower weight, flower length, pollen count and ELISA Phl p 5 assay results used to produce this manuscript. (PDF)

Acknowledgments

A special thanks to Gaurav Dhawan, MPH and undergraduates Matt Seeley and Erin Stockman who helped with the tedious process of collecting, preparing, and counting millions of pollen grains.

Author Contributions

Conceived and designed the experiments: JMA WJM MD MLM CAR. Performed the experiments: JMA. Analyzed the data: JMA KAS CAR. Contributed reagents/materials/analysis tools: CAR WJM KAS. Contributed to the writing of the manuscript: JMA CAR KAS MLM MD WJM.

- Singer BD, Ziska LH, Frenz DA, Gebhard DE (2005) Increasing Amb a 1 content in common ragweed (Ambrosia artemisiifolia L.) as a function of rising atmospheric CO₂. Funct Plant Biol 32: 667–670.
- Ziello C, Sparks TH, Estrella N, Belmonte J, Bergmann KC, et al. (2012).
 Changes to airborne pollen counts across Europe. PLoS ONE 7(4): e34076.
- White JF, Bernstein DI (2003) Key pollen allergens in North America. Ann Allergy Asthma Immunol 91: 425–435.
- Héguy L, Garneau M, Goldberg MS, Raphoz M, Guay F, et al. (2008) Associations between grass and weed pollen and emergency department visits for asthma among children in Montreal. Environmental Research 106: 203–211.
- Darrow LA, Hess J, Rogers CA, Tolbert PE, Klein M, et al. (2012) Ambient pollen concentrations and emergency department visits for asthma and wheeze. J Allergy Clin Immunol 130: 630–638.
- 15. Meehl GA, Stocker TF, Collins WD, Friedlingstein P, Gaye AT, et al. 2007. Global climate projections. In: Solomon S, Qin C, Manning M, Chen Z, Marquis M, Averyt KB, et al., Editors. Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. New York: Cambridge University Press, pp. 47–847.
- Vingarzan R (2004) A review of surface ozone background levels and trends. Atmos Environ 38: 3431–3442.
- 17. Leakey AD, Ainsworth EA, Bernacchi CJ, Rogers A, Long SP, et al. (2009) Elevated $\rm CO_2$ effects on plant carbon, nitrogen and water relations: six important lessons form FACE. J Exp Bot 60: 2859–2876.

- 18. Ainsworth EA, Yendrek CR, Sitch S, Collins WJ, Emberson LD (2012) The effects of tropospheric ozone on net primary productivity and implications for climate change. Annu Review of Plant Biol 63: 637-661
- 19. Rogers CA, Wayne PM, Macklin EA, Muilenberg ML, Wagner CJ, et al. (2006) Interaction of the onset of spring and elevated atmospheric CO2 on ragweed (Ambrosia artemisiifolia L.) pollen production. Environ Health Perspect 114:
- 20. Wayne P, Foster S, Connolly J, Bazzaz F, Epstein P (2002) Production of allergenic pollen by ragweed (Ambrosia artemisiifolia L.) in increased in CO2enriched atmospheres. Ann Allergy Asthma Immunol 88: 279-280.
- 21. Stinson KA, Bazzaz FA (2006) CO₂ enrichment reduces reproductive dominance in compering stands of Ambrosia artemisiifolia L. (common ragweed). Oecologia 147: 155-163.
- 22. Wand SJE, Midgley GF, Jones MH, Curtis PS (1999) Responses of wild C4 and C3 grass (Poaceae) species to elevated atmospheric CO2 concentration: a metaanalytic test of current theories and perceptions. Glob Change Biol 5: 723-741.
- 23. Masuch G, Franz J-Th, Shcoene K, Musken H, Bergmann K-Ch (1997) Ozone increases group 5 allergen content of *Lolium perenne*. Allergy 52: 874–875.

 24. Schoene K, Franz J-Th, Masuch G (2004) The effect of ozone on pollen
- development in Lolium perenne L. Environ Pollut 131: 347-354.
- 25. Motta AC, Marliere M, Peltre G, Sterenberg PA, Lacroix G (2006) Trafficrelated air pollutants induce the release of allergen- containing cytoplasmic granules from grass pollen. Int Arch Allergy Immunol 139: 294–298.
- 26. Rogerieux F, Godfrin D, Senechal H, Motta AC, Marliere M, et al. (2007) Modifications of *Phleum pratense* grass pollen allergens following artificial exposure to gaseous air pollutants (O₃, NO₂, SO₂). Int Arch Allergy Immunol 143: 127-134
- 27. Manning WJ, Krupa SV (1992) Experimental methodology for studying the effects of ambient ozone on crops and trees. In: Lefohn AS, editor. Surface Level Ozone Exposures and their Effects on Vegetation. Michigan: Lewis Publishing, pp. 111–119.
- 28. Johnson BG, Hale BA, Ormrod DP (1996) Carbon dioxide and ozone effects on growth in a legume- grass mixture. J Environ Qual 25: 908-916.

- 29. Frei T (1998) The effects of climate change in Switzerland 1969-1996 on airborne pollen quantities from hazel, birch, and grass. Grana 37: 172-179.
- Ziska LH, Knowlton K, Rogers CA, Dalan D, Tierney N, et al. (2011) Recent warming by latitude associated with increase of ragweed pollen in central North America. Proceedings of the National Academy of Sciences 108: 4248-4251.
- 31. Albertine JM (2013) Understanding the links between Human Health and Climate Change: Agricultural productivity and Allergenic Pollen Production of Timothy Grass (Phleum pratense L.) under future predicted levels of Carbon Dioxide and Ozone. PhD Dissertation, The University of Massachusetts-Amherst. Available: http://scholarworks.umass.edu/cgi/viewcontent. cgi?article=1782&context=open_access_dissertations. Accessed15 September
- 32. Knox RB, Suphioglu C (1996) Pollen allergens: development and function. Sex Plant Reprod 9: 318-323.
- 33. Townsend AR, Howarth RW, Bazzaz FA, Booth MS, Cleveland CC et al. (2003) Human health effects of a changing global nitrogen cycle. Front Ecol Environ 1: 240-246.
- 34. Roshchina VV, Roshchina VD (2003) Ozone and Plant Cell. Boston: Kluwer Academic Press, pp. 55-102.
- 35. Peden DB, Woodrow Setzer R, Devlin RB (1995) Ozone exposure has both a priming effect on allergen-induced responses and an intrinsic inflammatory action in the nasal airways on perennially allergic asthmatics. Am J Respir Crit Care Med 151: 1336-1345.
- 36. EPA NAAQS (Environmental Protection Agency National Ambient Air Quality Standards). 2014. NAAQS homepage. Available: http://www.epa.gov/air/ criteria.html. Accessed 1 May 2014.
- 37. Kirtman B, Power SB, Adedoyin JA, Boer GJ, Bojariu R, et al. (2013) Near-term Climate Change: Projections and Predictability. In: Stocker TF, Qin D, Plattner G-K, Tignor M, Allen SK, Boschung J, et al., editors. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change, New York: Cambridge University Press, pp. 953-1028.