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## 11CO<sub>2</sub> Fixation: A Renaissance in PET Radiochemistry

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| <b>Citation</b>          | Rotstein, Benjamin H., Steven H. Liang, Jason P. Holland, Thomas Lee Collier, Jacob M. Hooker, Alan A. Wilson, and Neil Vasdev. 2013. 11CO <sub>2</sub> fixation: a renaissance in PET radiochemistry. <i>Chemical Communications</i> 49(50): 5621.  |
| <b>Published Version</b> | <a href="https://doi.org/10.1039/c3cc42236d">doi:10.1039/c3cc42236d</a>  |
| <b>Accessed</b>          | February 19, 2015 3:35:38 PM EST   |
| <b>Citable Link</b>      | <a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:12111436">http://nrs.harvard.edu/urn-3:HUL.InstRepos:12111436</a>  |
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# ChemComm

Chemical Communications

[www.rsc.org/chemcomm](http://www.rsc.org/chemcomm)

Volume 49 | Number 50 | 25 June 2013 | Pages 5607–5706



ISSN 1359-7345

RSC Publishing

**FEATURE ARTICLE**

Neil Vasdev *et al.*

$^{11}\text{CO}_2$  fixation: a renaissance in PET radiochemistry

## FEATURE ARTICLE

 $^{11}\text{C}$  fixation: a renaissance in PET radiochemistry

Cite this: *Chem. Commun.*, 2013, **49**, 5621

Benjamin H. Rotstein,<sup>a</sup> Steven H. Liang,<sup>a</sup> Jason P. Holland,<sup>a</sup> Thomas Lee Collier,<sup>ab</sup> Jacob M. Hooker,<sup>c</sup> Alan A. Wilson<sup>d</sup> and Neil Vasdev<sup>\*a</sup>

Received 27th March 2013,  
Accepted 1st May 2013

DOI: 10.1039/c3cc42236d

[www.rsc.org/chemcomm](http://www.rsc.org/chemcomm)

Carbon-11 labelled carbon dioxide is the cyclotron-generated feedstock reagent for most positron emission tomography (PET) tracers using this radionuclide. Most carbon-11 labels, however, are installed using derivative reagents generated from  $[^{11}\text{C}]\text{CO}_2$ . In recent years,  $[^{11}\text{C}]\text{CO}_2$  has seen a revival in applications for the direct incorporation of carbon-11 into functional groups such as ureas, carbamates, oxazolidinones, carboxylic acids, esters, and amides. This review summarizes classical  $[^{11}\text{C}]\text{CO}_2$  fixation strategies using organometallic reagents and then focuses on newly developed methods that employ strong organic bases to reversibly capture  $[^{11}\text{C}]\text{CO}_2$  into solution, thereby enabling highly functionalized labelled compounds to be prepared. Labelled compounds and radiopharmaceuticals that have been translated to the clinic are highlighted.

<sup>a</sup> Division of Nuclear Medicine and Molecular Imaging, Massachusetts General Hospital, and Department of Radiology, Harvard Medical School, 55 Fruit Street, Boston, MA 02114, USA. E-mail: [vasdev.neil@mgh.harvard.edu](mailto:vasdev.neil@mgh.harvard.edu); Fax: +1 617-726-6165; Tel: +1 617-643-4736

<sup>b</sup> Advion Inc., 10 Brown Road, Ithaca, NY 14850, USA

<sup>c</sup> Athinoula A. Martinos Center for Biomedical Imaging, Department of Radiology, Massachusetts General Hospital, Harvard Medical School, Charlestown, MA 02129, USA

<sup>d</sup> Research Imaging Centre, Centre for Addiction and Mental Health and Department of Psychiatry, University of Toronto, 250 College Street, Toronto, ON, Canada M5T 1R8

## 1. Introduction

Carbon nuclei make up all organic entities and are the most versatile sites for labelling biological molecules of interest without altering their chemical and/or biological profiles. Carbon-11, a nearly pure positron emitter ( $t_{1/2} = 20.38$  min,  $E_{\text{avg}}(\beta^+) = 0.39$  MeV) is used extensively for developing radiotracers for positron emission tomography (PET), a non-invasive molecular imaging technique. The development of PET radiopharmaceuticals may provide an ideal methodology to enable diagnosis, monitor disease progression, and evaluate drug



**Benjamin H. Rotstein**

Benjamin Rotstein studied chemistry (BSch 2007) at Dalhousie University and University of King's College (Halifax, Nova Scotia). He moved to University of Toronto to continue his education under the supervision of Prof. Andrei Yudin, earning a PhD in 2012. During this time, his research focused on multicomponent reactions and peptide chemistry, and included medicinal chemistry research at GlaxoSmithKline

(Durham, North Carolina). In 2012, he began a postdoctoral position with Dr Neil Vasdev at Harvard Medical School and Massachusetts General Hospital, where he is studying carbon-11 and fluorine-18 radiochemistry and PET imaging.



**Neil Vasdev**

Neil Vasdev obtained a PhD in Radiochemistry from McMaster University (Canada) with Prof. Raman Chirakal and Prof. Gary Schrobilgen in 2003, followed by a postdoctoral fellowship with Prof. Henry VanBrocklin at the E.O. Lawrence Berkeley National Laboratories (USA). In 2004, he started his independent research career at the University of Toronto and CAMH (Canada). In 2011, he was recruited as an Associate Professor at Harvard

Medical School and is the Director of Radiochemistry at Massachusetts General Hospital (USA). He has developed a radiopharmaceutical chemistry research programme aimed at synthesizing new  $^{11}\text{C}$  and  $^{18}\text{F}$ -labelled probes and translating these for human imaging studies.

therapies *in vivo* without eliciting a pharmacological response.<sup>1</sup> In addition to serving as a clinical tool for disease diagnosis, PET is increasingly relevant for drug development as it can provide quantitative pharmacokinetic, biodistribution, and receptor occupancy data for a drug candidate. While the longer half-life of fluorine-18 ( $t_{1/2} = 109.77$  min,  $E_{\text{avg}}(\beta^+) = 0.25$  MeV) offers advantages for synthesis, longer imaging times and multi-centre trials, carbon-11 can more frequently be substituted into biological molecules without causing chemical alterations that could influence the outcomes of imaging studies. Furthermore, repeat imaging studies in the same subject over a short duration are possible with carbon-11 due to its shorter half-life.

Dozens of reagents have been employed for incorporation of no-carrier-added carbon-11 ( $^{11}\text{C}$ ) nuclei in radiotracers.<sup>2</sup> Still, the most common strategy used is methylation using  $^{11}\text{C}$  methyl iodide or  $^{11}\text{C}$  methyl triflate.<sup>3,4</sup> Carbon-11 labelled carbon dioxide is the product of proton bombardment of nitrogen-14 in the presence of small amounts of oxygen by the  $^{14}\text{N}(p,\alpha)^{11}\text{C}$  nuclear reaction. Most often,  $^{11}\text{C}$ CO<sub>2</sub> is transformed into more reactive species, such as the aforementioned methylating reagents, to facilitate radiolabelling. However,  $^{11}\text{C}$ CO<sub>2</sub> itself is an attractive starting material for radiochemists as it is produced directly from the cyclotron in high specific activity. Its use also promises access to high oxidation state functional groups such as carboxylic acids, amides, ureas, carbamates, oxazolidinones and their derivatives without resorting to redox manipulations during radiotracer synthesis. Alternatively,  $^{11}\text{C}$ phosgene (produced from  $^{11}\text{C}$ CO<sub>2</sub> (ref. 5 and 6) or  $^{11}\text{C}$ methane<sup>7–10</sup>) has been used to prepare such functional groups, though it has seen limited adoption due to technical challenges required for its routine production and use.<sup>11</sup> As described in a recent review, the high reactivity of  $^{11}\text{C}$ COCl<sub>2</sub> has been exploited to generate  $^{11}\text{C}$ -labelled intermediates such as isocyanates and carbamoyl chlorides from amines, and chloroformates from alcohols, en route to  $^{11}\text{C}$ ureas,  $^{11}\text{C}$ carbamates and alkyl  $^{11}\text{C}$ carbonates.<sup>12</sup> Carbon-11 labelled carbon monoxide, produced by reduction of  $^{11}\text{C}$ CO<sub>2</sub>,<sup>13</sup> has been more widely employed for preparation of amides, esters, ureas, carbamates and acids using transition metal or selenium-mediated reactions or photoinitiated radical methods.<sup>11</sup> Research has focused on overcoming the low solubility of  $^{11}\text{C}$ CO by employing micro-autoclaves,<sup>14</sup> sequestration reagents such as borane,<sup>15</sup> microfluidics,<sup>16–18</sup> or soluble Xe<sub>(g)</sub> carrier.<sup>19</sup> This topic has been reviewed elsewhere.<sup>2,11,20</sup>

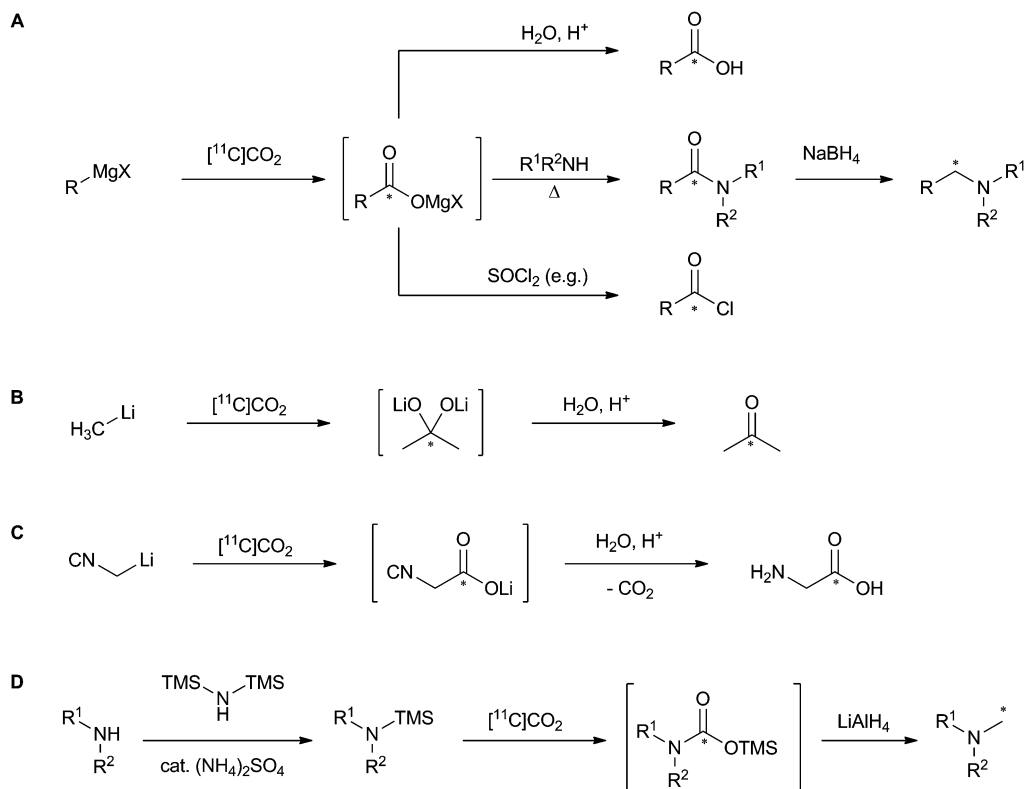
The low chemical reactivity of carbon dioxide poses a challenge for direct incorporation into organic molecules. Carbon dioxide generally requires highly reactive nucleophiles or catalysts to effect covalent bond formation and typically large excesses of CO<sub>2</sub> are used in industrial scale processes with this feedstock.<sup>21</sup> In contrast,  $^{11}\text{C}$ CO<sub>2</sub> is the limiting reagent (10–100 fold excess of precursor) when used in radiochemical transformations. The low amounts of  $^{11}\text{C}$ CO<sub>2</sub> in nitrogen carrier gas obtained from the cyclotron target (typically ~100 nmol) necessitate an efficient CO<sub>2</sub>-trapping solution. Carbon-11 CO<sub>2</sub> reactions are further complicated by the presence of oxygen and byproduct nitric oxides in the target gas mixture that often

require gas purification steps, particularly if transition metals or sensitive catalysts are to be used. Two predominant strategies exist for purification of  $^{11}\text{C}$ CO<sub>2</sub> from target gas. Cryogenic purification consists of depositing the mixture in a vessel cooled by liquid nitrogen. Non-condensable gases are thereby removed, while condensable impurities (such as NO<sub>x</sub> species) can be removed using chemical traps.<sup>22</sup> A second approach is to immobilize the carbon dioxide on a solid material (*e.g.* molecular sieves such as CarboSpheres<sup>23</sup>). After stripping off undesired impurities, the CO<sub>2</sub> is thermally released from the trap into the reactor. In contrast to  $^{11}\text{C}$ COCl<sub>2</sub> and  $^{11}\text{C}$ CO, no additional chemical steps involving elaborate apparatus are required for preparation of  $^{11}\text{C}$ CO<sub>2</sub>. However, as with any gaseous reagents with short half-lives (*e.g.*,  $^{11}\text{C}$ CO<sub>2</sub>,  $^{11}\text{C}$ CO,  $^{11}\text{C}$ COCl<sub>2</sub>,  $^{11}\text{C}$ HCN,  $^{11}\text{C}$ CH<sub>3</sub>I,  $^{11}\text{C}$ CH<sub>3</sub>OTf, and  $^{18}\text{F}$ F<sub>2</sub>) apparatus must be well designed and constructed to allow efficient handling, and be leak-proof. Flow rates must also be appropriate for rapid gas trapping in small volume solutions. Any traps, additives or fixation bases should avoid contamination of the isotope with impurities that could jeopardize the incorporation reaction. Many CO<sub>2</sub>-fixations proceed rapidly at room temperature and ambient pressure. The reaction set-ups are generally very simple, which contributes to good reproducibility of syntheses and high radiochemical yields relative to starting  $^{11}\text{C}$ CO<sub>2</sub> activity are achievable with high specific activity (3–6 Ci μmol<sup>-1</sup>). CO<sub>2</sub> is amenable to transformations into commonly found functional groups, and compounds have been recently advanced for first-in-human trials using  $^{11}\text{C}$ CO<sub>2</sub> fixation (*vide infra*). The purpose of this review is to highlight prominent examples of  $^{11}\text{C}$ CO<sub>2</sub> fixation that have been used to expand the chemical scope of labelled compounds and radiopharmaceuticals.

## 2. $^{11}\text{C}$ CO<sub>2</sub> fixation by basic organometallic reagents

### 2.1 Grignard reagents

Carbon-11 labelled CO<sub>2</sub> has been in use for preparation of labelled carboxylic acids since at least the 1940s (Scheme 1A).<sup>24</sup> Radiolabelled amides have also been synthesized by  $^{11}\text{C}$ CO<sub>2</sub> fixation with Grignard reagents. Heating magnesium carboxylate intermediates in the presence of primary or secondary amines has been reported to produce the corresponding  $^{11}\text{C}$ carboxyamides.<sup>25</sup> The product amides have also been subsequently reduced in the presence of sodium borohydride, giving access to  $^{11}\text{C}$ tertiary amines.<sup>26</sup> More indirectly, carboxylation can be followed by activation using reagents such as thionyl chloride or phthaloyl dichloride to prepare active acylation intermediates such as  $^{11}\text{C}$ acetyl and  $^{11}\text{C}$ propionyl chloride.<sup>27–30</sup> While Grignard reagents have proven useful in the synthesis of simple  $^{11}\text{C}$ carboxylic acids and a selection of their derivatives, their high reactivity inherently limits the potential scope of their applications and enforces requirements for careful handling procedures. Since Grignard reagents readily absorb CO<sub>2</sub> from the atmosphere and are moisture-sensitive they are ideally prepared fresh and manipulated under an inert atmosphere



**Scheme 1**  $^{11}\text{C}$ -fixations using strongly basic organometallic reagents such as Grignard reagents (A); organolithium reagents (B and C); and silanamines (D). TMS: trimethylsilyl; asterisk denotes  $^{11}\text{C}$ .

using anhydrous solvents to obtain high specific activities and reproducibility. The propensity for magnesium salts to precipitate from solution poses challenges for automated synthesis and can necessitate delicate and time consuming filtration steps.

## 2.2 Organolithiums

In the presence of excess methyl lithium  $^{11}\text{C}$  is readily transformed to the dilithium salt of acetone acetal. Hydrolysis leads to  $[2-^{11}\text{C}]$ acetone, which is a useful intermediate for radiochemistry (Scheme 1B).<sup>31–33</sup> Reports using other organolithiums to trap  $^{11}\text{C}$  have been relatively scarce.  $^{11}\text{C}$  pyruvic acid was prepared by tandem methylation–carboxylation of an isocyanide in the presence of methyl lithium and  $^{11}\text{C}$   $\text{CO}_2$ , followed by hydrolysis.<sup>34</sup> Also of note are the syntheses of radiolabelled glycine and two derivative dipeptides using the lithium salt of methylisocyanide. After  $^{11}\text{C}$   $\text{CO}_2$ -fixation the isocyanide is hydrolyzed through the formamide to the amine, yielding  $[1-^{11}\text{C}]$ glycine (Scheme 1C).<sup>35</sup> This species could be coupled to an intramolecularly activated amino acid equivalent under basic conditions.

## 2.3 Silanamines

Silylated amines were developed in the 1980s as fixing agents for  $^{11}\text{C}$   $\text{CO}_2$ . This work was performed by Ram and co-workers and was applied to various targets including imipramine,<sup>36</sup> chlorpromazine,<sup>37</sup> SCH-23390,<sup>38</sup> and tamoxifen.<sup>39</sup> Silanamines were prepared up to one week in advance of radiosynthesis by

refluxing a secondary amine with hexamethyldisilazane (HMDS), often in the presence of catalytic quantities of ammonium sulfate. When exposed to  $^{11}\text{C}$   $\text{CO}_2$  at elevated temperatures, O-silyl carbamates were produced, which were subsequently reduced *in situ* by  $\text{LiAlH}_4$  to produce labelled tertiary methylamines (Scheme 1D).<sup>40</sup> Since silyl groups are electron-donating, the silanamine is rendered more nucleophilic, despite the added steric bulk and weak N–Si bond strength.

The above methods all require the use of an unstable organometallic/organosilicon species for  $^{11}\text{C}$   $\text{CO}_2$  fixation. These reagents are often quite reactive and pose obstacles to reproducibility and automation in a radiochemical setting. Naturally, they also present major chemoselectivity issues during the preparation of PET radiotracers with complex functionalities, often limiting their utility to synthesis of simple prosthetic groups or very small molecules with restricted functionality. These approaches have been previously reviewed<sup>41</sup> and stand in contrast to the recent developments described below.

## 3. Recently developed methods of $^{11}\text{C}$ $\text{CO}_2$ fixation

Over the past 20 years, and under the impetus of “green” chemistry, approaches to  $\text{CO}_2$  fixation have expanded tremendously.<sup>42</sup> Among the most commonly used fixation agents are guanidines,<sup>43</sup> amidines,<sup>44,45</sup> and alkali carbonates,<sup>46</sup> the last of which are not suitable for  $^{11}\text{C}$   $\text{CO}_2$  fixation. Though each of these bases facilitates transamidation of an added amine, the nature of

products of CO<sub>2</sub> trapped by guanidines or amidines have been the subject of some debate. Initial spectroscopic and crystallographic studies suggested that bicarbonate salts are formed in the presence of adventitious water, perhaps by hydrolysis of carbamic intermediates.<sup>43,47–49</sup> More recently, the carbamate anion has been confirmed crystallographically in the context of a bicyclic guanidine.<sup>50</sup> Practically, Hooker *et al.* demonstrated that an amidine, diazabicyclo[5.4.0]undec-7-ene (DBU), was highly efficient at trapping cyclotron-produced [<sup>11</sup>C]CO<sub>2</sub> at ambient temperature and pressure and practical flow rates.<sup>51</sup> Wilson *et al.* later showed that 2-*tert*-butylimino-2-diethylamino-1,3-dimethyl-perhydro-1,3,2-diazaphosphorine (BEMP) was even more effective in this regard.<sup>52</sup> This strategy succeeds by decoupling the [<sup>11</sup>C]CO<sub>2</sub> capture in solution from subsequent covalent bond formation with substrate.

Transition metals have been extensively used for both CO<sub>2</sub> trapping in solution and as fixation catalysts for carbon–carbon bond formation.<sup>53,54</sup> The mode and efficiency of trapping are likely to be dependent on the nature of the metal centre, as well as the ligand and solvent systems. These topics have been the subjects of reviews in recent years.<sup>55,56</sup>

## 4. Functional groups prepared by [<sup>11</sup>C]CO<sub>2</sub> fixation

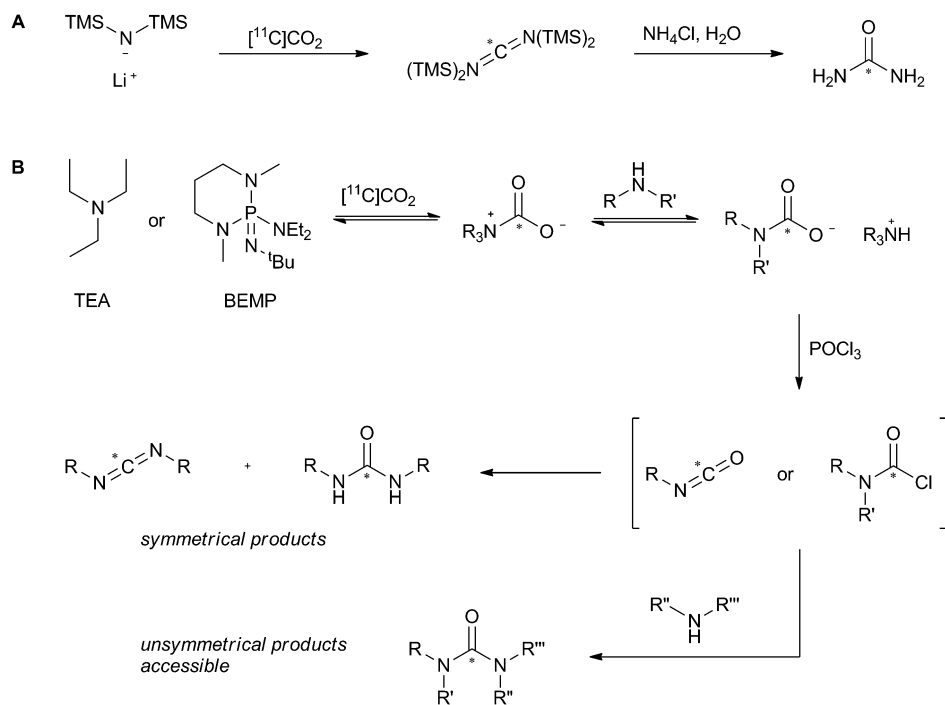
### 4.1 Urea

**4.1.1 Symmetrical ureas.** While early examples of [<sup>11</sup>C]CO<sub>2</sub> fixation centred around formation of carboxylic acids and amides, higher oxidation state functional groups such as ureas

and carbamates have become research targets in the past 20 years. Chakraborty *et al.* reported the first synthesis of [<sup>11</sup>C]urea by bubbling [<sup>11</sup>C]CO<sub>2</sub> through a THF solution of LHMDS, followed by hydrolysis using aqueous ammonium chloride (Scheme 2A).<sup>57</sup> [<sup>11</sup>C]urea could then be condensed with diethyl malate in sulfuric acid to yield [<sup>11</sup>C]uracil.

The synthesis of both ureas and carbamates from carbon dioxide often requires CO<sub>2</sub> to react twice as an electrophile. While we have seen above that in the presence of strong nucleophiles carbon dioxide will react as an electrophile, the resulting carboxylate or carbamate is not electrophilic. It is for this reason that [<sup>11</sup>C]phosgene has previously been used to prepare labelled ureas.<sup>12</sup> An alternative approach is to utilize activating reagents such as phosphoryl chloride or thionyl chloride to generate an acid chloride or isocyanate, which are both highly electrophilic. In its earliest iterations, this strategy was applied to prepare [<sup>11</sup>C-carbonyl]phenyl isocyanate, which promptly reacted further to form both [<sup>11</sup>C]diphenyl urea and [<sup>11</sup>C]diphenyl carbodiimide (Scheme 2B).<sup>58</sup> While observation of these products is clear evidence of the desired reactivity, they also illustrate the practical challenges in harnessing it for preparation of unsymmetrical ureas in high radiochemical yield.

**4.1.2 Unsymmetrical ureas.** By using dilute and carefully balanced solutions of trapping amine, POCl<sub>3</sub>, and nucleophile, unsymmetrical [<sup>11</sup>C]ureas can be produced with high selectivity (Scheme 2B).<sup>59</sup> Simply using a large excess of POCl<sub>3</sub> was found to suppress attack of remaining trapping amine on the <sup>11</sup>C-isocyanate generated *in situ*, but also required even larger excess of the nucleophile, which is also consumed. This results



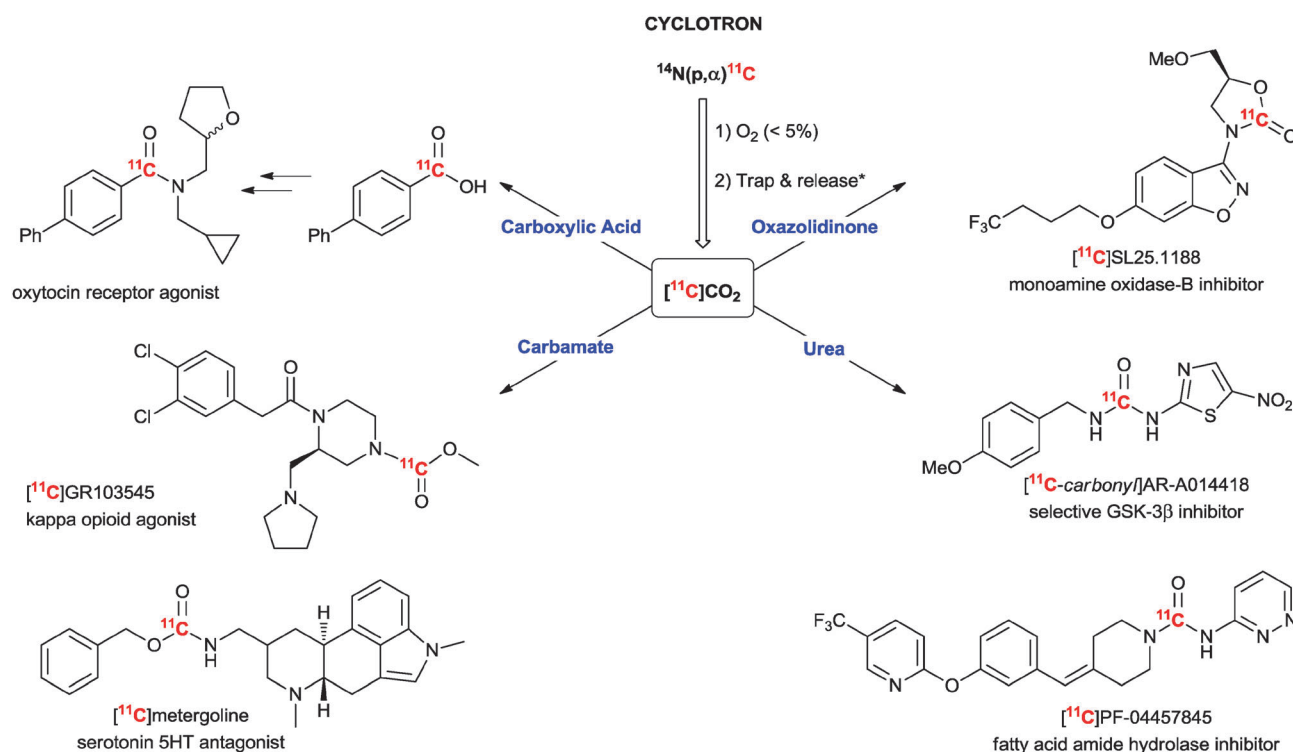
**Scheme 2** Synthesis of [<sup>11</sup>C]ureas by [<sup>11</sup>C]CO<sub>2</sub>-fixation. (A) Synthesis of parent [<sup>11</sup>C]urea using LHMDS and aqueous ammonium chloride. (B) Synthesis of symmetrical and unsymmetrical substituted [<sup>11</sup>C]ureas by activation of a carbamate intermediate with POCl<sub>3</sub>. The utility of this reaction was greatly expanded by judicious control of reaction conditions to favour unsymmetrical products. TEA: triethylamine; TMS: trimethylsilyl; asterisk denotes <sup>11</sup>C.

in a high concentration reaction mixture and consequent challenging purification. Fortunately, it was found that the concentration of trapping amine could be reduced without negatively impacting the yield or prolonging the reaction time beyond two minutes. Aliphatic primary amines are ideal substrates for isocyanate formation, while aniline reacts sluggishly.<sup>59</sup> Cyclic secondary amines are also well tolerated, presumably through an [ $^{11}\text{C}$ ]carbamoyl chloride intermediate, though more hindered secondary amines must be used in higher concentrations to achieve useful conversions. The scope of amine nucleophiles for attack on [ $^{11}\text{C}$ ]isocyanates or [ $^{11}\text{C}$ ]carbamoyl chlorides was limited primarily to dimethylamine for the discovery investigations, but high yields were also obtained with 4-(2-methoxyphenyl)piperazine.<sup>59</sup>

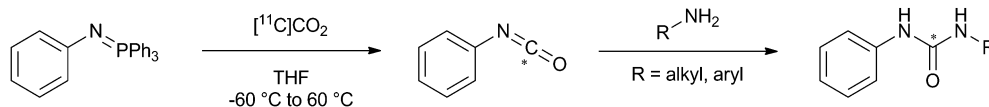
Inhibitors of glycogen synthase kinase 3 $\beta$  (GSK-3 $\beta$ ) are currently under exploration for diverse cancers and neurological illnesses, with the “GSK-3 hypothesis of Alzheimer’s disease” sparking further medicinal chemistry efforts in this area. Vasdev *et al.* prepared the first PET imaging agent for this target, [ $^{11}\text{C}$ -methoxy]AR-A014418.<sup>60</sup> The room temperature synthesis of an isotopologue of this unsymmetrical urea, [ $^{11}\text{C}$ -carbonyl]AR-A014418, demonstrates a [ $^{11}\text{C}$ ]CO $_2$  fixation approach to this tracer.<sup>61</sup> In this case the terminal nucleophile was the aromatic 2-amino-5-nitrothiazole. This is the first successful example of aromatic amines being employed in this role. The total time for synthesis and formulation was 28 minutes, and 70  $\pm$  33 mCi of the tracer could be prepared in 8.3  $\pm$  3.9% uncorrected radiochemical yield (RCY) relative to starting [ $^{11}\text{C}$ ]CO $_2$  with high (4.0  $\pm$  1.1 Ci  $\mu\text{mol}^{-1}$ ) specific

activity at the end of synthesis (Fig. 1). Though AR-A014418 was found to be less potent than initially reported, this methodology affords a general approach to synthesize arrays of  $^{11}\text{C}$ -labelled urea-based GSK-3 $\beta$  inhibitors, which would not be possible using standard  $^{11}\text{C}$ -methylation strategies.

Pfizer’s potent and irreversible fatty acid amide hydrolase (FAAH) inhibitor, PF-04457845, has advanced to clinical trials and has generated recent interest as a scaffold for PET radiotracer development.<sup>62</sup> Whereas a [ $^{18}\text{F}$ ]fluoroethyl derivative, [ $^{18}\text{F}$ ]PF-9811, was recently reported,<sup>63</sup> the isotopologue [ $^{11}\text{C}$ -carbonyl]PF-04457845 was prepared by [ $^{11}\text{C}$ ]CO $_2$  fixation (Fig. 1). The precursors for [ $^{11}\text{C}$ ]CO $_2$  fixation are a non-nucleophilic primary aromatic amine and a cyclic secondary aliphatic amine. To compensate for the higher reactivity of the latter compound, the aromatic amine was employed in 20-fold excess relative to the secondary amine. [ $^{11}\text{C}$ ]CO $_2$  was bubbled into the vial containing the precursors and BEMP in anhydrous CH $_3$ CN. POCl $_3$  was later added, followed by an aqueous quench and purification to provide [ $^{11}\text{C}$ -carbonyl]PF-04457845 in 4.5  $\pm$  1.3% uncorrected radiochemical yield (Fig. 1).<sup>64</sup> The total synthesis time was 25  $\pm$  2 min and the product had a specific activity of 2.0  $\pm$  0.2 Ci  $\mu\text{mol}^{-1}$ . All operations were performed at room temperature. In contrast to the preparation of [ $^{11}\text{C}$ ]AR-A014418, both amines were present in the reaction mixture during [ $^{11}\text{C}$ ]CO $_2$  fixation. Presumably, the reactive intermediate is a [ $^{11}\text{C}$ ]carbamoyl chloride or a mixed phosphate [ $^{11}\text{C}$ ]anhydride. Promising preclinical results, coupled with the known pharmacology and toxicology of PF-04457845, should facilitate clinical translation of this radiotracer.



**Fig. 1** Cyclotron produced [ $^{11}\text{C}$ ]CO $_2$  has been recently applied to radiolabelling of structurally complex carboxylic acids and amides, carbamates, oxazolidinones, and ureas. \*Trapping is typically done with molecular sieves or liquid nitrogen and release is achieved by heating.



**Scheme 3** Synthesis of [ $^{11}\text{C}$ ]phenylisocyanate and [ $^{11}\text{C}$ ]ureas by fixation with *N*-phenyl(triphenylphosphin)imine. Asterisk denotes  $^{11}\text{C}$ .

An alternative approach to [ $^{11}\text{C}$ ]isocyanates is by the condensation of phosphinimines with [ $^{11}\text{C}$ ]CO<sub>2</sub>. Phosphinimines can be prepared from azides or primary amines, and have varying degrees of stability. Commercially available phenyltriphenylphosphinimine was used for [ $^{11}\text{C}$ ]urea preparation with aliphatic and aromatic amines.<sup>65</sup> The radionuclide was trapped in a solution of THF at  $-60\text{ }^\circ\text{C}$  in the presence of the phosphinimine and amine. The reaction was heated to  $60\text{ }^\circ\text{C}$  for 6 minutes to complete the synthesis (Scheme 3). By radio-HPLC, aliphatic amines gave decay corrected unpurified radiochemical conversion yields ranging from 45–49%, while aniline unsurprisingly gave a lower yield of  $8 \pm 1\%$ .

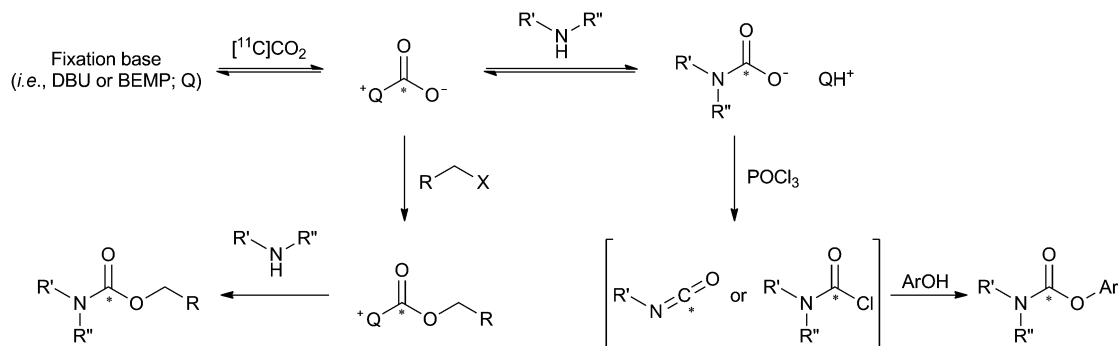
## 4.2 Carbamate

**4.2.1 Syntheses using alkyl electrophiles.** Similar to ureas, carbamates are an attractive functional group for radiolabelling due to their stability *in vivo* and role as a versatile linker for ligand fragments. This functional group has previously been labelled using [ $^{11}\text{C}$ ]phosgene and [ $^{11}\text{C}$ ]carbon monoxide,<sup>11</sup> so a synthetic strategy based around [ $^{11}\text{C}$ ]CO<sub>2</sub> would have extensive applications for the development of PET radiopharmaceuticals. In contrast to ureas, syntheses of *O*-alkyl carbamates do not necessarily require activation of the carbamate salt intermediate if reactive electrophiles are used. This strategy was exemplified by employing benzyl chloride in a solution of DBU, benzylamine, DMF and bubbled [ $^{11}\text{C}$ ]CO<sub>2</sub> (Scheme 4).<sup>51</sup> The total reaction time was 10 minutes at  $75\text{ }^\circ\text{C}$  and achieved a radiochemical yield of 85%. Similar to some of the urea results discussed above, reducing the concentration of benzylamine in this reaction was better tolerated than reducing the concentrations of other reagents. Secondary and sterically hindered amines gave good yields, while anilines were much less reactive. The best electrophiles were benzyl and allyl chlorides since analogous bromides displayed undesired reactivity with amines and DBU. *n*-Pentyl bromide showed desired reactivity,

while more hindered aliphatic electrophiles were troublesome. This technology was used to prepare [ $^{11}\text{C}$ ]metergoline in 32% decay corrected RCY in high specific activity (up to  $5\text{ Ci }\mu\text{mol}^{-1}$ ) in a single step. [ $^{11}\text{C}$ ]MS-275, an isotopologue of the histone deacetylase inhibitor entinostat, was also prepared in this way and showed poor blood–brain barrier penetration in non-human primates.<sup>66</sup> The tracer was synthesized in 25% decay corrected RCY with specific activities of  $2.7\text{--}6.2\text{ Ci }\mu\text{mol}^{-1}$ . Synthesis and purification required 50 min and afforded  $>20\text{ mCi}$  per synthesis.

Methyl [ $^{11}\text{C}$ -carbonyl]carbamates have also been prepared following a similar strategy with methylating agents such as dimethylsulfate (DMS), methyl iodide (CH<sub>3</sub>I), or methyl tosylate (CH<sub>3</sub>OTs) (Scheme 4).<sup>52</sup> In this case, BEMP in DMF was found to be the best trapping solution for [ $^{11}\text{C}$ ]CO<sub>2</sub>. The reactions proceeded with primary or secondary amines. Electron rich 4-methoxyaniline was also tolerated, while acceptable yields of electron deficient 4-nitroaniline could be achieved with increased amine concentration. The reactions also proceeded very rapidly, with maximum yields being reached using one minute of mixing prior to addition of the methylating agent and only 10 seconds afterwards. Increasing the concentration of methylating agent had a negative effect on radiochemical yield. Reversing the order of addition (*i.e.*, first DMS, followed by amine) reportedly produced very low RCYs of product, suggesting the intermediacy of the carbamate salt.

The utility of this process was demonstrated by labelling GR103545, a selective high affinity agonist for the  $\kappa$ -opioid receptor, by [ $^{11}\text{C}$ ]CO<sub>2</sub> fixation. The methyl carbamate has previously been labelled using [ $^{11}\text{C}$ ]methylchloroformate,<sup>67</sup> phosgene with [ $^{11}\text{C}$ ]CH<sub>3</sub>OH,<sup>68</sup> and either [ $^{11}\text{C}$ ]CH<sub>3</sub>I<sup>69</sup> or [ $^{11}\text{C}$ ]CH<sub>3</sub>OTf<sup>70</sup> with cold CO<sub>2</sub> fixation. In this iteration, the “loop” method was employed by lining the inside of a narrow-bore steel tube with a solution of the precursor and BEMP in DMF. Without requiring heating, 70–103 mCi of  $2.9\text{--}4.4\text{ Ci }\mu\text{mol}^{-1}$  [ $^{11}\text{C}$ -carbonyl]GR103545



**Scheme 4** [ $^{11}\text{C}$ -carbonyl]carbamates can be synthesized by nucleophilic alkylation of the trapped [ $^{11}\text{C}$ ]CO<sub>2</sub> followed by amine substitution (left), or through activation of an intermediate carbamate salt using POCl<sub>3</sub> (right). DBU: 1,8-diazabicyclo[5.4.0]undec-7-ene; Q: fixation base; R: H, phenyl, vinyl, alkyl; X: Cl, Br, I, OTs, sulfate; asterisk denotes  $^{11}\text{C}$ .



(Fig. 1) was produced using 0.1 mg of precursor, an uncorrected 13% RCY at end-of-synthesis (EOS).<sup>52</sup> The total synthesis time was 23 minutes.

**4.2.2 Syntheses via isocyanates.** [<sup>11</sup>C-*carbonyl*]carbamates have also been produced through [<sup>11</sup>C]isocyanate intermediates.<sup>59</sup> Again, appropriate stoichiometry must be selected to prevent symmetrical [<sup>11</sup>C]urea formation. The isocyanate is quenched with an alcohol to affix the desired *O*-substituent. While methanol and phenols were originally deployed, in our most recent work we have also successfully used ethanol, isopropanol, and even *tert*-butanol and hexafluoroisopropanol. It is possible that the reaction is assisted by the strongly basic conditions established by the presence of BEMP.

The synthesis and function of [<sup>11</sup>C]CURB (Scheme 5A) is illustrative of the importance of this labelling strategy. FAAH is a serine hydrolase that regulates levels of anandamide, an endocannabinoid, in the central nervous system. FAAH is found heterogeneously throughout the brain, and is an attractive target in studies of addiction, obesity, pain, anxiety, and eating disorders.<sup>71–73</sup> The *O*-aryl carbamate scaffold has shown to be an effective one for design of irreversible inhibitors of FAAH, as the serine-241 residue in the active site has been shown to attack the carbonyl of the carbamate, releasing a phenolic fragment (Scheme 5B). With the potential for structure–activity relationship (SAR) studies in mind, it can be appreciated that the conserved carbamate carbonyl is an ideal position for placing a radiolabel.

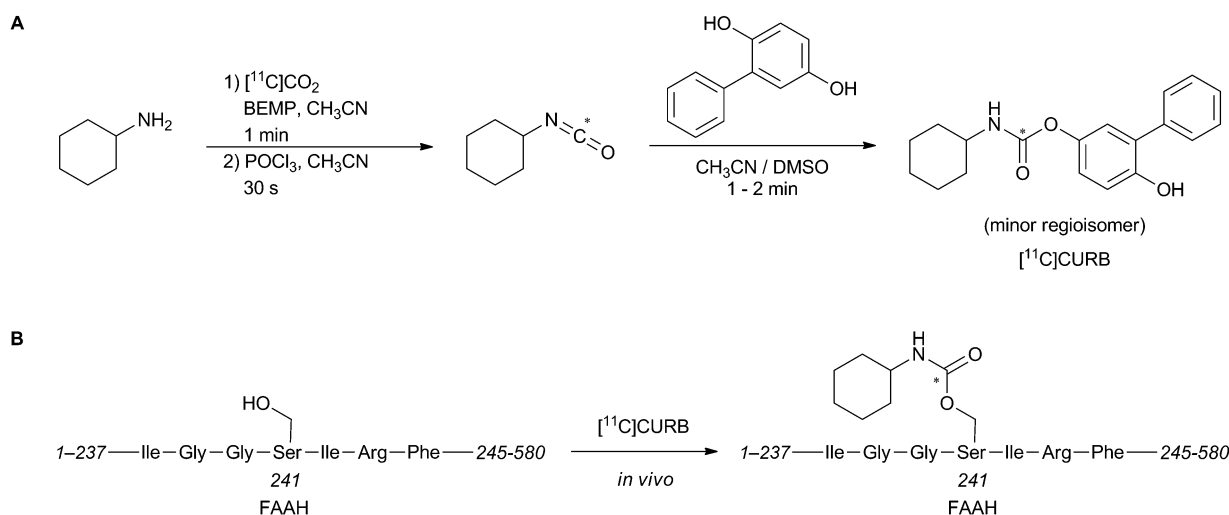
[<sup>11</sup>C]CURB was prepared by [<sup>11</sup>C]CO<sub>2</sub> fixation using cyclohexylamine and BEMP, followed by [<sup>11</sup>C]isocyanate formation with POCl<sub>3</sub>.<sup>59</sup> 2-Phenyldihydroquinone was used to quench the isocyanate, forming a mixture of regioisomers which are separated by HPLC. The radiotracer was isolated in 8% uncorrected RCY with a specific activity of 2.5 Ci μmol<sup>-1</sup> in 27 minutes from end-of-bombardment (EOB). [<sup>11</sup>C]CURB shows high brain penetration in rats, and selectivity was ascertained by blocking studies with a known FAAH inhibitor.<sup>74</sup> The radiotracer has been translated to human use.<sup>75</sup> To optimize radiotracer pharmacokinetics,

an SAR study was performed using a small library of *O*-aryl carbamates. Eight [<sup>11</sup>C]carbamates were prepared in a manner analogous to [<sup>11</sup>C]CURB.<sup>76</sup> Each of the radiotracers demonstrated brain uptake and specificity for FAAH in conscious rodents. Kinetic analysis in rats showed that [<sup>11</sup>C]dihydrooxazole carbamates had greater brain uptake, lower non-specific binding, and faster binding to FAAH than the [<sup>11</sup>C]biphenyl carbamates, such as [<sup>11</sup>C]CURB. The results of the SAR study and the human kinetic studies using [<sup>11</sup>C]CURB will allow for design and selection of the optimal FAAH radiotracer.

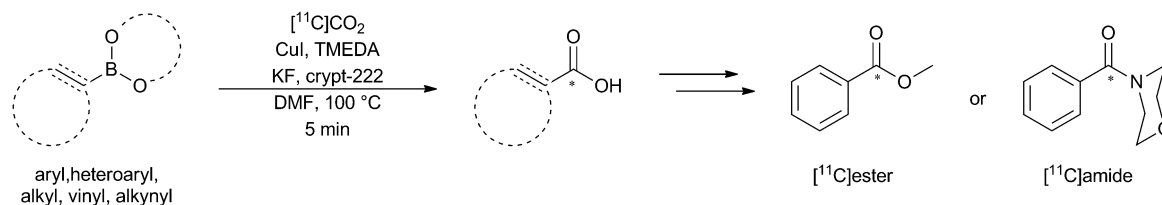
A camptothecin derivative that displays potent antitumor activity, irinotecan, was radiolabelled with carbon-11.<sup>77</sup> Kawamura *et al.* prepared [<sup>11</sup>C]irinotecan both by direct [<sup>11</sup>C]CO<sub>2</sub> fixation and using [<sup>11</sup>C]COCl<sub>2</sub> derived from [<sup>11</sup>C]CO<sub>2</sub> by way of [<sup>11</sup>C]CH<sub>4</sub> and [<sup>11</sup>C]CCl<sub>4</sub>. Using [<sup>11</sup>C]CO<sub>2</sub> directly, the decay corrected RCY was 16.9 ± 2.9% with specific activity 35 min from EOB of 2.1–3.7 Ci μmol<sup>-1</sup>. Using [<sup>11</sup>C]COCl<sub>2</sub>, the decay corrected RCY was 8.8 ± 2.0% with specific activity 35 min from EOB of 2.1–5.3 Ci μmol<sup>-1</sup>. The tracer was used to perform metabolite analysis of the drug in mice.

### 4.3 Oxazolidinone

[<sup>11</sup>C]SL25.1188 (Fig. 1) was developed as a reversibly binding radiotracer for monoamine oxidase-B (MAO-B). The radiolabel is placed as a [<sup>11</sup>C-*carbonyl*]oxazolidinone. The original synthesis employed [<sup>11</sup>C]phosgene, and suffered from a low 3.5–7% decay-corrected RCY with 1.4–1.9 Ci μmol<sup>-1</sup> specific activity after a 30–32 minutes.<sup>78</sup> The application of the amino-alcohol precursor for [<sup>11</sup>C]CO<sub>2</sub> fixation followed by dehydration and intramolecular cyclization improves the radiosynthesis.<sup>79</sup> After optimization of the fixation base, dehydrating agent, and component concentrations, conditions were developed to perform the labelling at ambient temperature using BEMP and POCl<sub>3</sub>. The uncorrected RCY was 11.5 ± 0.9% to prepare 98 ± 8 mCi of the radiotracer with 1.0 ± 0.05 Ci μmol<sup>-1</sup> specific activity after a 30 min synthesis. This tracer has since been successfully translated for human use and specific



**Scheme 5** (A) Radiosynthesis of [<sup>11</sup>C]CURB; (B) To effectively measure brain levels of FAAH using [<sup>11</sup>C]CURB, the radiolabel must be located on the carbonyl or *N*-alkyl fragment of the tracer. Asterisk denotes <sup>11</sup>C.



**Scheme 6** Cu(I) catalyzes  $^{11}\text{C}$ -carboxylation of boronic acid esters. The products can be elaborated to  $^{11}\text{C}$ -esters and  $^{11}\text{C}$ -amides. TMEDA: *N,N,N',N'*-tetramethylethylenediamine; asterisk denotes  $^{11}\text{C}$ .

activities of  $3.7 \pm 0.8 \text{ Ci } \mu\text{mol}^{-1}$  have been achieved by us (unpublished work).

#### 4.4 Carboxylic acid

Transition metal-mediated carboxylation of organic compounds has seen significant development in part due to the reputation of  $\text{CO}_2$  as an environmentally benign feedstock. In particular, catalytic and even metal-free conditions using prefunctionalized organoboron and organozinc reagents have been the focus of much attention.<sup>80–82</sup> The advantages these reagents hold over Grignard and organolithium compounds are their functional group tolerance and relative stability to storage under ambient conditions. Boronic esters have recently been applied for  $^{11}\text{C}$ CO<sub>2</sub> fixation using a copper catalyst.<sup>83</sup> Due to low concentration of  $^{11}\text{C}$ CO<sub>2</sub> available in the reaction mixture, which is in sharp contrast to “cold” (non-radioactive) CO<sub>2</sub>-fixation conditions, the reaction parameters required significant optimization efforts. Since alkoxide bases bound  $^{11}\text{C}$ CO<sub>2</sub> tightly, TMEDA was employed as both fixation base and presumably a ligand for the copper catalyst. A soluble fluoride additive was also found to dramatically improve RCY. With optimized conditions at 90–100 °C, various arylboronic esters were efficiently converted to  $^{11}\text{C}$ carboxylic acids (Scheme 6). Halide, formyl, cyano, and nitro substituents were well tolerated, while protic substituents such as hydroxyl and amino groups, and electron poor heterocycles were challenging substrates. Examples of alkyl, vinyl and alkynyl substrates undergoing  $^{11}\text{C}$ carboxylation were also reported.

The  $^{11}\text{C}$ benzoic acids were also elaborated to a methyl ester (using iodomethane), an amide (*via* an acid chloride), and a succinic ester (*via* carbodiimide activation).<sup>83</sup> An  $^{11}\text{C}$ amide-labelled oxytocin receptor ligand<sup>84</sup> was also reported in 20% decay-corrected RCY with  $1.5 \text{ Ci } \mu\text{mol}^{-1}$  specific activity over 43 minutes synthesis time. Access to various  $^{11}\text{C}$ carboxylic acid derivatives through regiospecific metal catalyzed carboxylation is sure to be widely exploited.

## 5. Conclusion

The last few years have seen considerable enthusiasm for radiosynthesis using  $^{11}\text{C}$ CO<sub>2</sub>. Whereas classical  $^{11}\text{C}$ CO<sub>2</sub> incorporation has relied on highly basic organometallic reagents, modern strategies employ more functional group tolerant reagents and catalysts for activation. This has enabled the synthesis of structurally complex radiopharmaceuticals and precise placement of the carbon-11 label within functional

groups such as ureas, carbamates, and oxazolidinones under very mild conditions, as well as carboxylic acids, esters and amides. Most recently, the carbamate radiotracer  $^{11}\text{C}$ CURB and the oxazolidinone,  $^{11}\text{C}$ SL25.1188 have advanced for human neuroimaging studies of FAAH and MAO-B, respectively. We anticipate that future developments in this field will establish mild and robust strategies for labelling a greater number of functional positions, including amides. Enabling technologies such as microfluidics and automated apparatus should prove to be valuable for rapid method optimization and widespread adoption of  $^{11}\text{C}$ CO<sub>2</sub> fixation for clinical translation.

## Acknowledgements

We thank Hogger & Co. for graphical abstract design.

## References

- 1 S. M. Ametamey, M. Horner and P. A. Schubiger, *Chem. Rev.*, 2008, **108**, 1501.
- 2 R. A. Ferrieri, in *Handbook of Radiopharmaceuticals*, ed. M. J. Welch and C. S. Redvanly, John Wiley & Sons, Ltd., Chichester, 2003, pp. 229–282.
- 3 R. Bolton, *J. Labelled Compd. Radiopharm.*, 2001, **44**, 701.
- 4 M. Allard, E. Fouquet, D. James and M. Szlosek-Pinaud, *Curr. Med. Chem.*, 2008, **15**, 235.
- 5 D. Roeda, C. Crouzel and B. van Zanten, *Radiochem. Radioanal. Lett.*, 1978, **33**, 175.
- 6 C. Crouzel and D. Roeda, *Int. J. Appl. Radiat. Isot.*, 1983, **34**, 1558.
- 7 P. Landais and C. Crouzel, *Appl. Radiat. Isot.*, 1987, **38**, 297.
- 8 K. Nishijima, Y. Kuge, K. Seki, K. Ohkura, N. Motoki, K. Nagatsu, A. Tanaka, E. Tsukamoto and N. Tamaki, *Nucl. Med. Biol.*, 2002, **29**, 345.
- 9 M. Ogawa, Y. Takada, H. Suzuki, K. Nemoto and T. Fukumura, *Nucl. Med. Biol.*, 2010, **37**, 73.
- 10 Y. Branmoullé, D. Roeda and F. Dollé, *Tetrahedron Lett.*, 2010, **51**, 313.
- 11 P. W. Miller, N. J. Long, R. Vilar and A. D. Gee, *Angew. Chem., Int. Ed.*, 2008, **47**, 8998.
- 12 D. Roeda and F. Dollé, *Curr. Top. Med. Chem.*, 2010, **10**, 1680.
- 13 S. K. Zeisler, M. Nader, A. Theobald and F. Oberdorfer, *Appl. Radiat. Isot.*, 1997, **48**, 1091.
- 14 J. Eriksson, O. Åberg and B. Långström, *Eur. J. Org. Chem.*, 2007, 455.
- 15 H. Audrain, L. Martarello, A. Gee and D. Bender, *Chem. Commun.*, 2004, 558.
- 16 P. W. Miller, N. J. Long, A. J. de Mello, R. Vilar, H. Audrain, D. Bender, J. Passchier and A. Gee, *Angew. Chem., Int. Ed.*, 2007, **46**, 2875.
- 17 P. W. Miller, H. Audrain, D. Bender, A. J. de Mello, A. D. Gee, N. J. Long and R. Vilar, *Chem.–Eur. J.*, 2011, **17**, 460.
- 18 S. Kealey, C. Plisson, T. L. Collier, N. J. Long, S. M. Husbands, L. Martarello and A. D. Gee, *Org. Biomol. Chem.*, 2011, **9**, 3313.
- 19 J. Eriksson, J. van den Hoek and A. D. Windhorst, *J. Labelled Compd. Radiopharm.*, 2012, **55**, 223.
- 20 B. Långström, O. Isenko and O. Rahman, *J. Labelled Compd. Radiopharm.*, 2007, **50**, 794.
- 21 I. Omae, *Coord. Chem. Rev.*, 2012, **256**, 1384.

- 22 T. J. Tewson, W. Banks, M. Franceschini and J. Hoffpauir, *Appl. Radiat. Isot.*, 1989, **40**, 765.
- 23 B. H. Mock, M. T. Vavrek and G. K. Mulholland, *Nucl. Med. Biol.*, 1995, **22**, 667.
- 24 J. M. Buchanan, A. B. Hastings and F. B. Nesbett, *J. Biol. Chem.*, 1943, **150**, 413.
- 25 C. Aubert, C. Huard-Perrio and M.-C. Lasne, *J. Chem. Soc., Perkin Trans. 1*, 1997, 2837.
- 26 C. Perrio-Huard, C. Aubert and M.-C. Lasne, *J. Chem. Soc., Perkin Trans. 1*, 2000, 311.
- 27 S. K. Luthra, V. W. Pike and F. Brady, *Chem. Commun.*, 1985, 1423.
- 28 J. McCarron, D. R. Turton, V. W. Pike and K. G. Poole, *J. Labelled Compd. Radiopharm.*, 1996, **38**, 941.
- 29 D.-R. Hwang, N. R. Simpson, J. Montoya, J. J. Mann and M. Laurelle, *Nucl. Med. Biol.*, 1999, **26**, 815.
- 30 P. Mäding, J. Zessin, U. Pleiß, F. Füchtner and F. Wüst, *J. Labelled Compd. Radiopharm.*, 2006, **49**, 357.
- 31 C. Prenant, J. Sastre, C. Crouzel and A. Syrota, *J. Labelled Compd. Radiopharm.*, 1987, **24**, 227.
- 32 A. R. Studenov, M. S. Berridge, D. V. Soloviev, M. Matarrese and S. Todde, *Nucl. Med. Biol.*, 1999, **26**, 431.
- 33 M. van der Meij, N. I. Carruthers, J. D. M. Herscheid, J. A. Jablownowski, J. E. Leysen and A. D. Windhorst, *J. Labelled Compd. Radiopharm.*, 2003, **46**, 1075.
- 34 M. R. Kilbourn and M. J. Welch, *Int. J. Appl. Radiat. Isot.*, 1982, **33**, 359.
- 35 J. M. Bolster, W. Vaalburg, Ph. H. Elsinga, M. G. Woldring and H. Wynberg, *Appl. Radiat. Isot.*, 1986, **37**, 985.
- 36 S. Ram, R. E. Ehrenkaufner and D. M. Jewett, *Appl. Radiat. Isot.*, 1986, **37**, 391.
- 37 S. Ram and L. D. Spicer, *Appl. Radiat. Isot.*, 1989, **40**, 413.
- 38 S. Ram, R. E. Ehrenkaufner and L. D. Spicer, *Appl. Radiat. Isot.*, 1989, **40**, 425.
- 39 S. Ram and L. D. Spicer, *J. Labelled Compd. Radiopharm.*, 1989, **27**, 662.
- 40 S. Ram and R. E. Ehrenkaufner, *Tetrahedron Lett.*, 1985, **26**, 5367.
- 41 S. Ram and R. L. Ehrenkaufner, *Nucl. Med. Biol.*, 1988, **15**, 345.
- 42 T. Sakakura, J.-C. Choi and H. Yasuda, *Chem. Rev.*, 2007, **107**, 2365.
- 43 F. S. Pereira, E. R. de Azevedo, E. F. da Silva, T. J. Bonagamba, D. L. da Silva Agostini, A. Magalhães, A. E. Job and E. R. P. González, *Tetrahedron*, 2008, **64**, 10097.
- 44 M. Yoshida, Y. Komatsuzaki and M. Ihara, *Org. Lett.*, 2008, **10**, 2083.
- 45 E. R. Pérez, M. O. da Silva, V. C. Costa, U. P. Rodrigues-Filho and D. W. Franco, *Tetrahedron Lett.*, 2002, **43**, 4091.
- 46 R. N. Salvatore, S. I. Shin, A. S. Nagle and K. W. Jung, *J. Org. Chem.*, 2001, **66**, 1035.
- 47 F. S. Pereira, D. L. da Silva Agostini, R. D. do Espírito Santo, E. R. de Azevedo, T. J. Bonagamba, A. E. Job and E. R. P. González, *Green Chem.*, 2011, **13**, 2146.
- 48 E. R. Pérez, R. H. A. Santos, M. T. P. Gambardella, L. G. M. de Macedo, U. P. Rodrigues-Filho, J.-C. Launay and D. W. Franco, *J. Org. Chem.*, 2004, **69**, 8005.
- 49 D. J. Heldebrant, P. G. Jessop, C. A. Thomas, C. A. Eckert and C. L. Liotta, *J. Org. Chem.*, 2005, **70**, 5335.
- 50 C. Villiers, J.-P. Dognon, R. Pollet, P. Thuéry and M. Ephritikhine, *Angew. Chem., Int. Ed.*, 2010, **49**, 3465.
- 51 J. M. Hooker, A. T. Reibel, S. M. Hill, M. J. Schueller and J. S. Fowler, *Angew. Chem., Int. Ed.*, 2009, **48**, 3482.
- 52 A. A. Wilson, A. Garcia, S. Houle and N. Vasdev, *Org. Biomol. Chem.*, 2010, **8**, 428.
- 53 Y. Tsuji and T. Fujihara, *Chem. Commun.*, 2012, **48**, 9956.
- 54 S. N. Riduan and Y. Zhang, *Dalton Trans.*, 2010, **39**, 3347.
- 55 K. Huang, C.-L. Sun and Z.-J. Shi, *Chem. Soc. Rev.*, 2011, **40**, 2435.
- 56 T. Fan, X. Chen and Z. Lin, *Chem. Commun.*, 2012, **48**, 10808.
- 57 P. K. Chakraborty, T. J. Mangner and H. T. Chugani, *Appl. Radiat. Isot.*, 1997, **48**, 619.
- 58 A. Schirbel, M. H. Holschbach and H. H. Cohen, *J. Labelled Compd. Radiopharm.*, 1999, **42**, 537.
- 59 A. A. Wilson, A. Garcia, S. Houle, O. Sadvovski and N. Vasdev, *Chem.-Eur. J.*, 2011, **17**, 259.
- 60 N. Vasdev, A. Garcia, W. T. Stableford, A. B. Young, J. H. Meyer, S. Houle and A. A. Wilson, *Bioorg. Med. Chem. Lett.*, 2005, **15**, 5270.
- 61 J. W. Hicks, A. A. Wilson, E. A. Rubie, J. R. Woodgett, S. Houle and N. Vasdev, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 2099.
- 62 D. S. Johnson, C. Stiff, S. E. Lazerwith, S. R. Kesten, L. K. Fay, M. Morris, D. Beidler, M. B. Liimatta, S. E. Smith, D. T. Dudley, N. Sadagopan, S. N. Bhattachar, S. J. Kesten, T. K. Nomanbhoy, B. F. Cravatt and K. Ahn, *ACS Med. Chem. Lett.*, 2011, **2**, 91.
- 63 M. B. Skaddan, L. Zhang, D. S. Johnson, A. Zhu, K. R. Zasadny, R. V. Coelho, K. Kuszpit, G. Currier, K.-H. Fan, E. M. Beck, L. Chen, S. E. Drozda, G. Balan, M. Niphakis, B. F. Cravatt, K. Ahn, T. Bocan and A. Villalobos, *Nucl. Med. Biol.*, 2012, **39**, 1058.
- 64 J. W. Hicks, J. Parkes, O. Sadvovski, J. Tong, S. Houle, N. Vasdev and A. A. Wilson, *Nucl. Med. Biol.*, in press.
- 65 E. W. van Tilburg, A. D. Windhorst, M. van der Mey and J. D. M. Herscheid, *J. Labelled Compd. Radiopharm.*, 2006, **49**, 321.
- 66 J. M. Hooker, S. W. Kim, D. Alexoff, Y. Xu, C. Shea, A. Reid, N. Volkow and J. S. Fowler, *ACS Chem. Neurosci.*, 2010, **1**, 65.
- 67 H. T. Ravert, U. Scheffel, W. B. Mathews, J. L. Musachio and R. F. Dannals, *Nucl. Med. Biol.*, 2002, **29**, 47.
- 68 P. S. Talbot, R. Narendran, E. R. Butelman, Y. Huang, K. Ngo, M. Slifstein, D. Martinez, M. Laruelle and D.-R. Hwang, *J. Nucl. Med.*, 2005, **46**, 484.
- 69 B. W. Shoultz, E. Årstad, J. Marton, F. Willoch, A. Drzezga, H.-J. Wester and G. Henriksen, *Open Med. Chem. J.*, 2008, **2**, 72.
- 70 N. B. Nabulsi, M.-Q. Zheng, J. Ropchan, D. Labaree, Y.-S. Ding, L. Blumberg and Y. Huang, *Nucl. Med. Biol.*, 2011, **38**, 215.
- 71 M. K. McKinney and B. F. Cravatt, *Annu. Rev. Biochem.*, 2005, **74**, 411.
- 72 C. J. Fowler, *Fundam. Clin. Pharmacol.*, 2006, **20**, 549.
- 73 J. E. Schlosburg, S. G. Kinsey and A. H. Lichtman, *AAPS J.*, 2009, **11**, 39.
- 74 A. A. Wilson, A. Garcia, J. Parkes, S. Houle, J. Tong and N. Vasdev, *Nucl. Med. Biol.*, 2011, **38**, 247.
- 75 P. Rusjan, A. A. Wilson, R. Mizrahi, I. Boileau, S. E. Chavez, N. J. Lobaugh, S. J. Kish, S. Houle and J. Tong, *J. Cereb. Blood Flow Metab.*, 2012, **1**.
- 76 A. A. Wilson, J. W. Hicks, O. Sadvovski, J. Parkes, J. Tong, S. Houle, C. J. Fowler and N. Vasdev, *J. Med. Chem.*, 2013, **56**, 201.
- 77 K. Kawamura, H. Hashimoto, M. Ogawa, J. Yui, H. Wakizaka, T. Yamasaki, A. Hatori, L. Xie, K. Kumata, M. Fujinaga and M.-R. Zhang, *Nucl. Med. Biol.*, 2013, DOI: 10.1016/j.nucmedbio.2013.03.004, in press.
- 78 Y. Bramoullé, F. Puech, W. Saba, H. Valette, M. Bottlaender, P. George and F. Dollé, *J. Labelled Compd. Radiopharm.*, 2008, **51**, 153.
- 79 N. Vasdev, O. Sadvovski, A. Garcia, F. Dollé, J. H. Meyer, S. Houle and A. A. Wilson, *J. Labelled Compd. Radiopharm.*, 2011, **54**, 678.
- 80 J. Takaya, S. Tadami, K. Ukai and N. Iwasawa, *Org. Lett.*, 2008, **10**, 2697.
- 81 K. Kobayashi and Y. Kondo, *Org. Lett.*, 2009, **11**, 2035.
- 82 L. Zhang, J. Cheng, T. Ohishi and Z. Hou, *Angew. Chem., Int. Ed.*, 2010, **49**, 8670.
- 83 P. J. Riss, S. Lu, S. Telu, F. I. Aigbirio and V. W. Pike, *Angew. Chem., Int. Ed.*, 2012, **51**, 2698.
- 84 B. R. Bellenie, N. P. Barton, A. J. Emmons, J. P. Heer and C. Salvagno, *Bioorg. Med. Chem. Lett.*, 2009, **19**, 990.