



DIGITAL ACCESS TO SCHOLARSHIP AT HARVARD

Nutrition, Epigenetics, and Diseases

The Harvard community has made this article openly available.
[Please share](#) how this access benefits you. Your story matters.

Citation	Jang, Hyeran, and Carlo Serra. 2014. "Nutrition, Epigenetics, and Diseases." <i>Clinical Nutrition Research</i> 3 (1): 1-8. doi:10.7762/cnr.2014.3.1.1. http://dx.doi.org/10.7762/cnr.2014.3.1.1 .
Published Version	doi:10.7762/cnr.2014.3.1.1
Accessed	April 17, 2018 4:46:18 PM EDT
Citable Link	http://nrs.harvard.edu/urn-3:HUL.InstRepos:11879849
Terms of Use	This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA

(Article begins on next page)



Nutrition, Epigenetics, and Diseases

Hyeran Jang^{1,2*}, Carlo Serra^{1,2}

¹Division of Endocrinology, Diabetes and Hypertension, Brigham and Women's Hospital, Boston, MA 02115, USA

²Harvard Medical School, Boston, MA 02115, USA

Increasing epidemiological evidence suggests that maternal nutrition and environmental exposure early in development play an important role in susceptibility to disease in later life. In addition, these disease outcomes seem to pass through subsequent generations. Epigenetic modifications provide a potential link between the nutrition status during critical periods in development and changes in gene expression that may lead to disease phenotypes. An increasing body of evidence from experimental animal studies supports the role of epigenetics in disease susceptibility during critical developmental periods, including peri-conceptual period, gestation, and early postnatal period. The rapid improvements in genetic and epigenetic technologies will allow comprehensive investigations of the relevance of these epigenetic phenomena in human diseases.

Key Words: Nutrition, Reprogramming, Development, Epigenetic, Disease

***Corresponding author** Hyeran Jang

Address Division of Endocrinology, Diabetes and Hypertension
Brigham and Women's Hospital/HMS 221 Longwood ave. EBRC-212
Boston, MA 02115, USA

Tel +1-617-525-9054 **Fax** +1-617-582-6193

E-mail hjang1@partners.org

Received November 28, 2013

Revised December 21, 2013

Accepted December 21, 2013

Introduction

Many human diseases that appear in adulthood are related to growth patterns during early life. It is now recognized that early-life nutrition and other environmental factors play key roles in the pathogenesis and in the predisposition of specific human diseases. In this regard, in 1995 Dr. Barker wrote: "The fetal origins hypothesis states that fetal undernutrition in middle to late gestation, which leads to disproportionate fetal growth, programs later coronary heart disease." [1]. The word "program" illustrates the idea that the environmental stimuli received during critical periods of early fetal development can generate permanent changes in body structure and function that affect the homeostasis of specific organs in the adult life [2]. The responsiveness of a growing body to external cues is defined as developmental plasticity. Developmental plasticity derives from the ability of our genes to organize different ranges of physiological or morphological states in response to environmental conditions during fetal development. In this review we focus on particular situations in human and experimental animal models to underline the link between nutritional and environmental stimuli during critical periods of the early development and the susceptibility to disease development in the adult life.

© 2014 The Korean Society of Clinical Nutrition

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dutch Famine Birth Cohort

Insights in the importance of nutritional supply during a critical period on long-term disease outcome were gained from the Dutch Famine cohort. The Dutch Famine cohort is one of the most well-known cohorts that have been used to investigate the effects of prenatal undernutrition in humans. During the Nazi occupation from November 1944 to May 1945, food supply was extremely limited in some parts of the Netherlands. Daily rations during this famine period started at less than 1,000 kcal in November 1944, and decreased to about 500 kcal by April 1945. This six month food shortage had a major impact on particular situations, such as ongoing pregnancies. From 1998, the Dutch Famine Birth Cohort Study began reporting the outcome of pregnancies that occurred during this famine period and the consequences of massive maternal undernutrition on the offspring on a long-term scale [3-5]. Results showed an increased risk of cardiovascular diseases 40-50 years later in those children born to mothers who experienced extremely severe undernutrition during the first trimester of pregnancy [6]. The incidence of cardiovascular diseases was 2 times higher in this group compared to the control cohort [6]. Also, the serious nutritional deprivation increased the risk of metabolic disorders and breast cancer decades later in this cohort [7-9]. Notably, depending on the period of starvation (early versus late pregnancy, and pre-conceptional versus postnatal undernutrition) marked differences in disease outcomes were observed, indicating that the first trimester of pregnancy is particularly vulnerable to disease outcome in adulthood.

Why Your DNA Isn't Your Destiny

Another interesting epidemiological study pointed out the importance of nutritional status during puberty and its impact on the offspring's health, by investigating if the availability of food to one generation affected longevity and health of the descendants. This study conducted by Dr. Bygren, investigated the long-term effects of feast and famine on children growing up in the isolated Swedish county of Norrbotten, with the goal to unravel whether "Parents' experiences early in their lives change the traits they passed to their offspring" [10]. In the 19th century, Norrbotten was so isolated that if the harvest failed, people starved. Harvest failures occurred erratically, causing famines in 1800, 1812, 1821, 1836 and 1856. Bygren and colleagues found that if an individual went through a famine as a teenager, his or her grandchildren would have

shown higher mortality risk ratio, but only if the grandchildren were the same sex as the grandparent who starved [10,11]. Starvation of a male only changed his grandson's mortality risk and starvation of a female only changed her granddaughter's mortality risk [10,11].

Early nutrition and onset of metabolic phenotype in adult

The three main insights of a number of studies from the Dutch and Norrbotten's cohorts are: 1) there may be critical developmental time windows where overnutrition or undernutrition may promote or impede disease development; 2) epigenetic factors can be modified by hunger exposure during these critical time windows; and 3) acquired epigenetic alterations are passed on to offspring. To better address these points, various animal models have been developed to explore whether nutritional or environmental modifications over critical developmental windows affect lifelong disease risk. It has been shown, for example, that overfeeding dams during gestation and lactation with high-caloric diet or reducing the number of pups in a litter could indirectly overnourish pups. Table 1 and Table 2 summarize some of the results observed in the offspring's phenotype induced by specific maternal dietary effects in different developmental time windows and in different experimental models. From these animal studies we can conclude that maternal overnutrition or undernutrition may act as an early mechanism of programming of the developing embryo/fetus, and may instruct a status of 'metabolic stress', restricting early embryonic cell proliferation and the generation of appropriately sized stem-cell pools. These observed phenotypic changes are dictated by the epigenetic-based molecular mechanisms that govern stability, transfer and expression of the eukaryotic genome.

Epigenetics

Since the Human Genome Project completed sequencing the 3 billion chemical base pairs that make up human DNA in April 2003, it became clear not only that we are able to read nature's complete genetic blueprint, but also that the information stored in the sequence of the DNA is not enough *per se* to completely explain human development, physiology and disease. The field dedicated to decipher the heritable features that complements the genetic information stored in the DNA sequence is termed "epigenetics". The prefix *epi-* is derived from the Greek preposition *ἐπί*, meaning above, on or over. Formally, epigenetics refers to the study of heritable changes in gene ex-

Table 1. Experimental animal models with maternal dietary modifications during development or early life, and their impact on phenotype of offspring

Species	Dietary modification	Time window of dietary modification	Observations in offspring	Reference
Mouse	Maternal low protein diet (9% vs 19% control)	One ovulatory cycle (3.5 days) prior to mating	Abnormal anxiety-related behavior in offspring; elevated systolic blood pressure	[50]
Rat	Maternal low protein diet (9% vs 19% control)	Preimplantation period (0–4.25 day postcoitum)	Low birth weight; altered postnatal growth; hypertension	[51]
Mouse	Maternal obesogenic diet (OD, 10% simple sugars, 20% animal lard, 28% polysaccharide, 23% protein, 4.5 kcal/g) vs standard chow diet (SD, 7% simple sugars, 3% fat, 50% polysaccharide, 15% protein, 3.5 kcal/g)(52)	6 weeks prior to mating – gestation – lactation 1. OD-OD-OD 2. SD-SD-SD 3. OD-OD-SD 4. SD-SD-OD	Dysmetabolic and non-alcoholic fatty liver disease (NAFLD) phenotype in offspring of postnatal OD	[52] [53]
	Maternal hypercaloric diet (OD, 16% fat, 33% sugar) vs standard chow (SD, 3% fat, 7% sugar)(53)			
Rat	Maternal high fat diet (HF diet: 60% kcal from fat vs CHOW diet: 17% kcal from fat)	Gestation and lactation 1.CHOW-CHOW 2.HF-HF	Increase in susceptibility to diet-induced obesity	[54]
Rat	Maternal high-fat diet (HF diet: 60% kcal from fat vs CHOW diet: 13.5%)	Gestation and lactation 1.CHOW-CHOW 2.CHOW-HF 3.HF-CHOW 4.HF-HF	Increase in adiposity, leptin and impaired glucose intolerance in postnatal HF offspring	[55]

OD: obesogenic diet, SD: standard chow diet, NAFLD: non-alcoholic fatty liver disease, HF: high fat diet, CHOW: chow diet.

Table 2. Rodent small litter models of postnatal overnutrition, and their impact on metabolic phenotype

Species	Litter size	Observations in pups from small litter	Reference
Mouse	3:9 (small:control)	Increase in body weight; impairment in glucose tolerance; decrease in insulin sensitivity in the heart	[56]
Mouse	4:9 (small:control)	increase in body weight and fat mass; decrease in physical activity and energy expenditure in females only	[43]
Mouse	4:9:20 (small:control:large)	Increase in body weight, adiposity (number and size) and blood insulin level; impaired glucose tolerance	[57]
Rat	4:10:16 (small:control:large)	Increase in body weight and body fat weight	[58]
Rat	3:10 (small:control)	Increase in body weight and body fat at postnatal day 21 and 180; central leptin resistance in adult life (postnatal day 180)	[59]
Rat	3:10:19 (small:control:large)	Early postnatal hyperinsulinemia; decreased glucose tolerance; increased susceptibility to streptozotocin-induced diabetes in adulthood	[60]
Rat	4:12:18 (small:control:large)	Increase in body weight; reduction in pancreatic glucose responsiveness; differential expression of pancreatic islets genes (13 genes) at postnatal day 26, 10 of them were maintained by postnatal day 110	[61]

pression that are not caused by changes in the DNA sequence [12]. In a more practical sense, epigenetics includes the study of DNA modifications, the post-translational changes of the

protein constituents of the chromatin, namely the histones, the chromatin modifications that appear to define biologic states in local regions of chromosomes, and the interaction of microR-

NAs with the genome [13]. Recent epigenetic studies have, for example, highlighted the mechanistic nature of the interactions between DNA and the enzymes that perform DNA replication, transcription, recombination, and repair [14].

DNA methylation

DNA methylation is a common modification in mammalian genomes and is considered a stable epigenetic mark transmitted through DNA replication and cell division [15]. Methylation is a covalent binding of a chemical methyl group to the cytosine (C) base in DNA sequence, specifically at CpG (cytosine-phosphate-guanine) dinucleotides. Methylation of C is typically associated with the 5' end of gene sequences, where CpG dinucleotides islands are particularly dense. Hypermethylation of the CpG islands is associated with transcriptional repression, while their hypomethylation is associated with the transcriptional activation of surrounding genes [16]. DNA methylation established during development and early postnatal life plays critical roles to regulate cell- and tissue-specific gene expression and genomic imprinting. Genomic imprinting is an epigenetic phenomenon in which the expression of a gene copy depends on its parent of origin [15,17], the significance of DNA methylation in genomic imprinting was highlighted in CpG islands of imprinted genes such as IGF2 and H19 [18]. In normal cells, the paternal IGF2 and maternal H19 gene are expressed while maternal IGF2 and paternal H19 are silenced by DNA methylation [19]. *De novo* DNA methylation is catalyzed by the DNA methyltransferases (DNMT) 3A and 3B, whereas DNA methylation is maintained after the replication of the DNA by DNMT1, which methylates hemi-methylated DNA [17]. Once established, DNA methylation is essentially maintained throughout life; however, a gradual hypomethylation occurs during aging, and it has been linked with some types of cancer [20].

Histone modification

The DNA in the cells is packaged as chromatin. The basic unit of chromatin is a nucleosome, which comprises 147 bp of DNA wrapped around a core of 8 histone proteins (two copies of histone H2A, H2B, H3 and H4) [21]. The N-terminal tails of the histones are subject to post-translational modifications including acetylation, methylation, ubiquitination, sumoylation, and phosphorylation [22]. Histone modifications lead to structural changes of the chromatin, and to the recruitment of effector proteins, for example transcription factors, which in turn bring about specific cellular processes by modulating target gene expression. Histone acetylation is exclusively associated

with active and open chromatin states, while the methylation of lysine residues in the N-terminal of histones can either be an active or repressive mark depending on the specific lysine involved [23]. Many families of histone-modifying enzymes have been identified, including histone acetyl transferases, deacetylases, methyltransferases, and demethylases [24].

Non-coding RNAs

Non-coding RNAs (ncRNAs) have been implicated in the epigenetic regulation of gene expression by gene silencing or target mRNA degradation. Recent studies have shown that human miRNAs can induce chromatin remodeling [25,26], suggesting that DNA methylation, histone modification and miRNAs may work in concert to regulate gene expression.

Developmental plasticity

During the development of multicellular organisms, different cells and tissues acquire different programs of gene expression. It is well recognized that a series of precisely timed and regulated epigenetic changes are required to ensure the proper development of complex organisms like humans [15,27]. At the morula stage, an early step of embryonic development, DNA demethylation occurs to erase all of the parent-of-origin methylation marks, except those of the imprinted genes. This allows for the inheritance of parental-specific monoallelic expression in somatic tissues throughout adulthood. This demethylation phase is followed by the *de novo* DNA methylation of the genome to establish the proper methylation patterns of the growing organism [28]. In such a way, as the embryo grows the offspring acquires their appropriate epigenetic features. In parallel to these changes occurring in somatic cells, another set of genomic reprogramming takes place in the cells of the germ line during gametogenesis [29]. In fact, during gonadal sex determination, primordial germ cells undergo genome-wide demethylation, which erases previous parental-specific methylation marks. Afterward, for example in the male germ line, paternal methylation marks occur in specific genes in the gonocytes that subsequently will develop into spermatogonia. Conversely, the female germ line establishes maternal methylation marks of imprinted genes at a later stage. In addition to DNA methylation, histone modifications are thought to play a role in the establishment of both sex-specific and non sex-specific marks because an extensive loss of histone methylation and acetylation occurs along with the loss of DNA methylation at the morula stage [30]. Epigenetic marks assure the proper expression of the imprinted

genes throughout the embryonic development. For example, genomic imprinting results in the monoallelic, parent-of-origin dependent expression of genes specifically required for key developmental steps. It is predictable from this scenario that the aberrant methylation of imprinted genes, mostly loss of imprinting (LOI) in the early stages of development, can alter the expression of critical genes, and then may bring out birth defects and adulthood diseases such as cancer [31]. In this regard, given the nature of the monoallelic expression, imprinted genes are particularly susceptible to the effect of epigenetic aberrations. In addition, for the strict dependence of these early steps of development to nutritional support, inherited and acquired epigenetic marks are particularly vulnerable to the interference coming from environmental stimuli [32].

Early life nutrition and altered epigenetic regulation

Due to the dynamic changes of the epigenetic regulation in development, particularly during gametogenesis and early embryogenesis, the epigenome displays labile nature, which allows it to respond and adapt to environmental stressors, including nutritional modification. For instance, periconceptional supplementation or restriction of the maternal diet with betaine, choline, folic acid, methionine, or vitamin B-12 in experimental models have been shown to affect the establishment of DNA methylation patterns, altering the gene expression and phenotype of the offspring [33]. Three main experimental approaches are in use to better understand the underlying mechanism by which nutritional modifications affect the epigenetic profile during critical developmental windows: 1) the study of epigenetically labile

genes, such as the insulin-like growth factor 2 (IGF2) gene; 2) the use of specific, natural animal models, such as the *agouti* mice; and 3) the targeting of a specific organ that is central for energy metabolism using dietary modification.

Insulin-like growth factor II (IGF2)

IGF2 is a key protein in human growth and development [34]. The *IGF2* gene is maternally imprinted and is one of the best-characterized epigenetically regulated loci [32]. The imprinting of this locus is maintained through the methylation of a DNA sequence named the differentially methylated region (DMR), the hypomethylation of which leads to bi-allelic expression of the *IGF2* gene. As illustrated by retrospective studies from the Dutch famine cohort [35] as well as in experimental animal studies [36–38], the *IGF2* gene has been shown to be characterized by a labile methylation pattern depending on the nutritional or environmental stimuli received by the growing organism during the early life. Table 3 summarizes some data that support the observed developmental plasticity of the *IGF2* locus. Notably, post-weaning mice fed a methyl-deficient diet exhibit permanent LOI and dysregulated expression of the *IGF2* gene [37], suggesting that both childhood and maternal diet contribute to the LOI in the *IGF2* locus in humans.

Agouti model

The *agouti* mouse has been extensively used to investigate the phenotypic impact of the nutritional modification during critical developmental periods, and the environmental influence on the fetal epigenome [39]. The *agouti viable yellow* (*A^{vy}*) locus regulates mouse coat color; more importantly, the

Table 3. Effect of maternal dietary modifications on imprinted gene, *IGF2*

Species	Modification during development	Time window for the modification	Observations in offspring	Reference
Human	Famine (Dutch Famine cohort)	Periconceptional period	Decrease in methylation of CpG dinucleotides in the <i>IGF2</i> nearly 60 years after	[35]
Human	Supplementary folic acid use	Periconceptional period	Higher methylation of the <i>IGF2</i> differentially methylated region (DMR)	[62]
Rat	Maternal low protein diet (8% vs 20 %) and high fat diet (45% vs 10%) after weaning	2 weeks prior to mating – gestation – lactation	Increase in adipose tissue <i>Igf2</i> mRNAs by the low protein prenatal diet	[38]
Rat	Maternal low protein diet (9% vs 19% control)	Preimplantation period	Decrease in <i>H19</i> imprinted gene expression in male blastocysts; reduction in <i>H19</i> and <i>Igf2</i> expression in male fetal liver at day 20 of gestation	[36]
Mouse	Methyl deficiency (methionine, choline, folic acid and vitamin B12)	60-day post-weaning	Loss of imprinting of <i>Igf2</i>	[37]

Table 4. Effects of maternal diet or environmental factors on yellow agouti offspring

Dietary or environmental modification	Observations	Reference
<i>a/a</i> females mated with the same strain of mottled yellow or pseudoagouti <i>A^{vy}/a</i> males; methyl-supplemented diets (folic acid, vitamin B12, betaine, and choline) to pregnant dams	Increase in <i>agouti</i> /black mottling in the direction of the pseudo-agouti phenotype	[40]
<i>A^{vy}/a</i> females mated with <i>a/a</i> males; methyl-supplemented diets (2 different diets) to pregnant dams	Increase in DNA methylation in the <i>agouti</i> LTR (long terminal repeat); change the phenotype of offspring in the healthy, longer-lived direction	[63]
F0 <i>A^{vy}/a</i> dams mated with <i>a/a</i> males; F1 <i>A^{vy}/a</i> female mated to <i>a/a</i> brother; F2 <i>A^{vy}/a</i> female mated to <i>a/a</i> brother; methyl-supplemented diets (folic acid, vitamin B12, betaine, and choline) to pregnant dams	Average <i>A^{vy}/a</i> coat color was darker in the methyl-supplemented diet group; epigenetic phenotypes are maternally heritable, but no cumulative effect of supplementation across successive generations on coat color; prevention of transgenerational amplification of obesity by methyl-supplementation	[64,65]
<i>a/a</i> females mated with <i>A^{vy}/a</i> males; 250 mg/kg diet of genistein for 2 weeks prior to mating, gestation and lactation	Coat color of <i>A^{vy}/a</i> offspring shifted toward pseudo-agouti phenotype	[66]
<i>a/a</i> females mated with <i>A^{vy}/a</i> males; 50 mg/kg diet of bisphenol A (BPA) for 2 weeks prior to mating, gestation and lactation	Coat color of <i>A^{vy}/a</i> offspring shifted toward yellow by decreasing CpG methylation in an intracisternal <i>A</i> particle retrotransposon upstream of the <i>Agouti</i> gene	[67]

product of the *agouti* gene interferes with the regulation of body weight at the level of the hypothalamus. DNA hypomethylation of the *agouti* gene promoter results in the accumulation of the *agouti* protein; as consequence, the mouse develops a yellow coat color as well as obesity. Conversely, hypermethylation of the promoter reduces the level of the *agouti* protein, and this, consequently, results in mice with lean phenotype and brown coat color. Interestingly, feeding *agouti* female pregnant mice with a diet enriched with methyl donor supplementation such as folic acid, vitamin B12, choline and betaine has been shown to modify the phenotype of their heterozygote offspring [40]. This effect was confirmed to be mediated by the hypermethylation of the *A^{vy}* promoter in the offspring of supplemented dams [41]. Table 4 provides additional evidences that specific maternal dietary treatments or environmental factors affect the phenotype of the *agouti* offspring through epigenetic mechanism.

Hypothalamic DNA methylation

The central brain region integrates various peripheral signals to regulate energy balance [42]. The hypothalamus plays a key role in the regulation of food intake and energy expenditure. For these reasons, the hypothalamus is an excellent organ candidate to study the epigenetic changes that mediate the developmental reprogramming of energy metabolism. Not surprisingly, maternal overnutrition [43–46] and undernutrition [47,48] have been shown to alter hypothalamic DNA methylation.

These changes in DNA methylation have been reported to persist from early life to adulthood [43,46], affecting the overall metabolism in the adult.

Conclusion

Developing organisms seem to have a wide range of susceptibility to epigenetic changes [49]. Appropriate dynamics in epigenetic modifications are essential for embryogenesis, early fetal development and early postnatal growth. Consequently, the inadequate establishment of epigenetic modifications during critical developmental periods due to changes in the maternal diet or other environmental factors may induce pediatric developmental diseases and even affect health in adulthood. Since much of the reprogramming that occurs during early life may go unrecognized until adulthood, a better understanding of the interplay between genetic and epigenetic interaction in critical time windows of development would improve our ability to determine individual susceptibility to a wide range of diseases.

Conflict of interest

We declare that we have no conflict of interest.

Reference

- Barker DJ. Fetal origins of coronary heart disease. *BMJ* 1995;311:171–4.
- Byrne CD, Phillips DI. Fetal origins of adult disease: epidemiology and mechanisms. *J Clin Pathol* 2000;53:822–8.

3. Roseboom TJ, Painter RC, van Abeelen AF, Veenendaal MV, de Rooij SR. Hungry in the womb: what are the consequences? Lessons from the Dutch famine. *Maturitas* 2011;70:141-5.
4. Painter RC, Osmond C, Gluckman P, Hanson M, Phillips DI, Roseboom TJ. Transgenerational effects of prenatal exposure to the Dutch famine on neonatal adiposity and health in later life. *BJOG* 2008;115:1243-9.
5. Roseboom T, de Rooij S, Painter R. The Dutch famine and its long-term consequences for adult health. *Early Hum Dev* 2006;82:485-91.
6. Painter RC, de Rooij SR, Bossuyt PM, Simmers TA, Osmond C, Barker DJ, Bleker OP, Roseboom TJ. Early onset of coronary artery disease after prenatal exposure to the Dutch famine. *Am J Clin Nutr* 2006;84:322-7.
7. de Rooij SR, Painter RC, Roseboom TJ, Phillips DI, Osmond C, Barker DJ, Tanck MW, Michels RP, Bossuyt PM, Bleker OP. Glucose tolerance at age 58 and the decline of glucose tolerance in comparison with age 50 in people prenatally exposed to the Dutch famine. *Diabetologia* 2006;49:637-43.
8. Painter RC, De Rooij SR, Bossuyt PM, Osmond C, Barker DJ, Bleker OP, Roseboom TJ. A possible link between prenatal exposure to famine and breast cancer: a preliminary study. *Am J Hum Biol* 2006;18:853-6.
9. de Rooij SR, Painter RC, Holleman F, Bossuyt PM, Roseboom TJ. The metabolic syndrome in adults prenatally exposed to the Dutch famine. *Am J Clin Nutr* 2007;86:1219-24.
10. Bygren LO, Kaati G, Edvinsson S. Longevity determined by paternal ancestors' nutrition during their slow growth period. *Acta Biotheor* 2001;49:53-9.
11. Pembrey ME, Bygren LO, Kaati G, Edvinsson S, Northstone K, Sjöström M, Golding J; ALSPAC Study Team. Sex-specific, male-line transgenerational responses in humans. *Eur J Hum Genet* 2006;14:159-66.
12. Bird A. Perceptions of epigenetics. *Nature* 2007;447:396-8.
13. Wade PA, Archer TK. Epigenetics: environmental instructions for the genome. *Environ Health Perspect* 2006;114:A140-1.
14. Musselman CA, Lalonde ME, Côté J, Kutateladze TG. Perceiving the epigenetic landscape through histone readers. *Nat Struct Mol Biol* 2012;19:1218-27.
15. Bird A. DNA methylation patterns and epigenetic memory. *Genes Dev* 2002;16:6-21.
16. Bird AP. CpG-rich islands and the function of DNA methylation. *Nature* 1986;321:209-13.
17. Reik W, Dean W, Walter J. Epigenetic reprogramming in mammalian development. *Science* 2001;293:1089-93.
18. Li E, Beard C, Jaenisch R. Role for DNA methylation in genomic imprinting. *Nature* 1993;366:362-5.
19. Rainier S, Johnson LA, Dobry CJ, Ping AJ, Grundy PE, Feinberg AP. Relaxation of imprinted genes in human cancer. *Nature* 1993;362:747-9.
20. Gonzalo S, Jaco I, Fraga MF, Chen T, Li E, Esteller M, Blasco MA. DNA methyltransferases control telomere length and telomere recombination in mammalian cells. *Nat Cell Biol* 2006;8:416-24.
21. Luger K, Mäder AW, Richmond RK, Sargent DF, Richmond TJ. Crystal structure of the nucleosome core particle at 2.8 Å resolution. *Nature* 1997;389:251-60.
22. Berger SL. An embarrassment of niches: the many covalent modifications of histones in transcriptional regulation. *Oncogene* 2001;20:3007-13.
23. Jenuwein T, Allis CD. Translating the histone code. *Science* 2001;293:1074-80.
24. Cole PA. Chemical probes for histone-modifying enzymes. *Nat Chem Biol* 2008;4:590-7.
25. Kim DH, Saetrom P, Snøve O Jr, Rossi JJ. MicroRNA-directed transcriptional gene silencing in mammalian cells. *Proc Natl Acad Sci U S A* 2008;105:16230-5.
26. Szenthe K, Koroknai A, Banati F, Bathori Z, Lozsa R, Burgyan J, Wolf H, Salamon D, Nagy K, Niller HH, Minarovits J. The 5' regulatory sequences of active miR-146a promoters are hypomethylated and associated with euchromatic histone modification marks in B lymphoid cells. *Biochem Biophys Res Commun* 2013;433:489-95.
27. Li E. Chromatin modification and epigenetic reprogramming in mammalian development. *Nat Rev Genet* 2002;3:662-73.
28. Bernal AJ, Jirtle RL. Epigenomic disruption: the effects of early developmental exposures. *Birth Defects Res A Clin Mol Teratol* 2010;88:938-44.
29. Jirtle RL, Skinner MK. Environmental epigenomics and disease susceptibility. *Nat Rev Genet* 2007;8:253-62.
30. Weaver JR, Susiarjo M, Bartolomei MS. Imprinting and epigenetic changes in the early embryo. *Mamm Genome* 2009;20:532-43.
31. DeBaun MR, Niemitz EL, McNeil DE, Brandenburg SA, Lee MP, Feinberg AP. Epigenetic alterations of H19 and LIT1 distinguish patients with Beckwith-Wiedemann syndrome with cancer and birth defects. *Am J Hum Genet* 2002;70:604-11.
32. Murphy SK, Jirtle RL. Imprinting evolution and the price of silence. *Bioessays* 2003;25:577-88.
33. Sinclair KD, Allegrucci C, Singh R, Gardner DS, Sebastian S, Bispham J, Thurston A, Huntley JF, Rees WD, Maloney CA, Lea RG, Craigen J, McEvoy TG, Young LE. DNA methylation, insulin resistance, and blood pressure in offspring determined by maternal periconceptional B vitamin and methionine status. *Proc Natl Acad Sci U S A* 2007;104:19351-6.
34. Reik W, Constancia M, Dean W, Davies K, Bowden L, Murrell A, Feil R, Walter J, Kelsey G. Igf2 imprinting in development and disease. *Int J Dev Biol* 2000;44:145-50.
35. Heijmans BT, Tobi EW, Stein AD, Putter H, Blauw GJ, Susser ES, Slagboom PE, Lumey LH. Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Proc Natl Acad Sci U S A* 2008;105:17046-9.
36. Kwong WY, Miller DJ, Ursell E, Wild AE, Wilkins AP, Osmond C, Anthony FW, Fleming TP. Imprinted gene expression in the rat embryo-fetal axis is altered in response to periconceptional maternal low protein diet. *Reproduction* 2006;132:265-77.
37. Waterland RA, Lin JR, Smith CA, Jirtle RL. Post-weaning diet affects genomic imprinting at the insulin-like growth factor 2 (Igf2) locus. *Hum Mol Genet* 2006;15:705-16.
38. Claycombe KJ, Uthus EO, Roemmich JN, Johnson LK, Johnson WT. Prenatal low-protein and postnatal high-fat diets induce rapid adipose tissue growth by inducing Igf2 expression in Sprague Dawley rat offspring. *J Nutr* 2013;143:1533-9.
39. Dolinoy DC. The agouti mouse model: an epigenetic biosensor for nutritional and environmental alterations on the fetal epigenome. *Nutr Rev* 2008;66 Suppl 1:S7-11.
40. Wolff GL, Kodell RL, Moore SR, Cooney CA. Maternal epigenetics and methyl supplements affect agouti gene expression in Avy/a mice. *FASEB J* 1998;12:949-57.
41. Waterland RA, Jirtle RL. Early nutrition, epigenetic changes at transposons and imprinted genes, and enhanced susceptibility to adult chronic diseases. *Nutrition* 2004;20:63-8.
42. Flier JS. Obesity wars: molecular progress confronts an expanding epidemic. *Cell* 2004;116:337-50.
43. Li G, Kohorst JJ, Zhang W, Laritsky E, Kunde-Ramamoorthy G, Baker MS, Fiorotto ML, Waterland RA. Early postnatal nutrition determines adult physical activity and energy expenditure in female mice. *Diabetes* 2013;62:2773-83.
44. Plagemann A, Harder T, Brunn M, Harder A, Roepke K, Wittrock-Staar M, Ziska T, Schellong K, Rodekamp E, Melchior K, Dudenhausen JW. Hypothalamic proopiomelanocortin promoter methylation becomes altered by early overfeeding: an epigenetic model of obesity and the metabolic syndrome. *J Physiol* 2009;587:4963-76.
45. Plagemann A, Roepke K, Harder T, Brunn M, Harder A, Wittrock-Staar M, Ziska T, Schellong K, Rodekamp E, Melchior K, Dudenhausen JW. Epigenetic malprogramming of the insulin receptor promoter due to

- developmental overfeeding. *J Perinat Med* 2010;38:393-400.
46. Vucetic Z, Kimmel J, Totoki K, Hollenbeck E, Reyes TM. Maternal high-fat diet alters methylation and gene expression of dopamine and opioid-related genes. *Endocrinology* 2010;151:4756-64.
 47. Begum G, Stevens A, Smith EB, Connor K, Challis JR, Bloomfield F, White A. Epigenetic changes in fetal hypothalamic energy regulating pathways are associated with maternal undernutrition and twinning. *FASEB J* 2012;26:1694-703.
 48. Coupé B, Amarger V, Grit I, Benani A, Parnet P. Nutritional programming affects hypothalamic organization and early response to leptin. *Endocrinology* 2010;151:702-13.
 49. Gluckman PD, Hanson MA, Buklijas T, Low FM, Beedle AS. Epigenetic mechanisms that underpin metabolic and cardiovascular diseases. *Nat Rev Endocrinol* 2009;5:401-8.
 50. Watkins AJ, Wilkins A, Cunningham C, Perry VH, Seet MJ, Osmond C, Eckert JJ, Torrens C, Cagampang FR, Cleal J, Gray WP, Hanson MA, Fleming TP. Low protein diet fed exclusively during mouse oocyte maturation leads to behavioural and cardiovascular abnormalities in offspring. *J Physiol* 2008;586:2231-44.
 51. Kwong WY, Wild AE, Roberts P, Willis AC, Fleming TP. Maternal undernutrition during the preimplantation period of rat development causes blastocyst abnormalities and programming of postnatal hypertension. *Development* 2000;127:4195-202.
 52. Oben JA, Mouralidarane A, Samuelsson AM, Matthews PJ, Morgan ML, McKee C, Soeda J, Fernandez-Twinn DS, Martin-Gronert MS, Ozanne SE, Sigala B, Novelli M, Poston L, Taylor PD. Maternal obesity during pregnancy and lactation programs the development of offspring non-alcoholic fatty liver disease in mice. *J Hepatol* 2010;52:913-20.
 53. Oben JA, Patel T, Mouralidarane A, Samuelsson AM, Matthews P, Pombo J, Morgan M, McKee C, Soeda J, Novelli M, Poston L, Taylor P. Maternal obesity programmes offspring development of non-alcoholic fatty pancreas disease. *Biochem Biophys Res Commun* 2010;394:24-8.
 54. Tamashiro KL, Terrillion CE, Hyun J, Koenig JI, Moran TH. Prenatal stress or high-fat diet increases susceptibility to diet-induced obesity in rat offspring. *Diabetes* 2009;58:1116-25.
 55. Sun B, Purcell RH, Terrillion CE, Yan J, Moran TH, Tamashiro KL. Maternal high-fat diet during gestation or suckling differentially affects offspring leptin sensitivity and obesity. *Diabetes* 2012;61:2833-41.
 56. Martins MR, Vieira AK, de Souza EP, Moura AS. Early overnutrition impairs insulin signaling in the heart of adult Swiss mice. *J Endocrinol* 2008;198:591-8.
 57. Aubert R, Suquet JP, Lemonnier D. Long-term morphological and metabolic effects of early under- and over-nutrition in mice. *J Nutr* 1980;110:649-61.
 58. Fiorotto ML, Burrin DG, Perez M, Reeds PJ. Intake and use of milk nutrients by rat pups suckled in small, medium, or large litters. *Am J Physiol* 1991;260:R1104-13.
 59. Rodrigues AL, de Moura EG, Passos MC, Trevenzoli IH, da Conceição EP, Bonono IT, Neto JF, Lisboa PC. Postnatal early overfeeding induces hypothalamic higher SOCS3 expression and lower STAT3 activity in adult rats. *J Nutr Biochem* 2011;22:109-17.
 60. You S, Götz F, Rohde W, Dörner G. Early postnatal overfeeding and diabetes susceptibility. *Exp Clin Endocrinol* 1990;96:301-6.
 61. Waterland RA, Garza C. Early postnatal nutrition determines adult pancreatic glucose-responsive insulin secretion and islet gene expression in rats. *J Nutr* 2002;132:357-64.
 62. Steegers-Theunissen RP, Obermann-Borst SA, Kremer D, Lindemans J, Siebel C, Steegers EA, Slagboom PE, Heijmans BT. Periconceptional maternal folic acid use of 400 microg per day is related to increased methylation of the IGF2 gene in the very young child. *PLoS One* 2009;4:e7845.
 63. Cooney CA, Dave AA, Wolff GL. Maternal methyl supplements in mice affect epigenetic variation and DNA methylation of offspring. *J Nutr* 2002;132:2393S-400S.
 64. Waterland RA, Travisano M, Tahiliani KG. Diet-induced hypermethylation at agouti viable yellow is not inherited transgenerationally through the female. *FASEB J* 2007;21:3380-5.
 65. Waterland RA, Travisano M, Tahiliani KG, Rached MT, Mirza S. Methyl donor supplementation prevents transgenerational amplification of obesity. *Int J Obes (Lond)* 2008;32:1373-9.
 66. Dolinoy DC, Weidman JR, Waterland RA, Jirtle RL. Maternal genistein alters coat color and protects Avy mouse offspring from obesity by modifying the fetal epigenome. *Environ Health Perspect* 2006;114:567-72.
 67. Dolinoy DC, Huang D, Jirtle RL. Maternal nutrient supplementation counteracts bisphenol A-induced DNA hypomethylation in early development. *Proc Natl Acad Sci U S A* 2007;104:13056-61.