



# DIGITAL ACCESS TO SCHOLARSHIP AT HARVARD

## Genetic Structure of Chimpanzee Populations

The Harvard community has made this article openly available.  
[Please share](#) how this access benefits you. Your story matters.

<b>Citation</b>	Becquet, Celine, Nick Patterson, Anne C. Stone, Molly Przeworski, and David Reich. 2007. Genetic structure of chimpanzee populations. PLoS Genetics 3(4): e66.
<b>Published Version</b>	<a href="https://doi.org/10.1371/journal.pgen.0030066">doi://10.1371/journal.pgen.0030066</a>
<b>Accessed</b>	February 19, 2015 7:35:58 AM EST
<b>Citable Link</b>	<a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:10178303">http://nrs.harvard.edu/urn-3:HUL.InstRepos:10178303</a>
<b>Terms of Use</b>	This article was downloaded from Harvard University's DASH repository, and is made available under the terms and conditions applicable to Other Posted Material, as set forth at <a href="http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA">http://nrs.harvard.edu/urn-3:HUL.InstRepos:dash.current.terms-of-use#LAA</a>

*(Article begins on next page)*

# Genetic Structure of Chimpanzee Populations

Celine Becquet<sup>1</sup>, Nick Patterson<sup>2</sup>, Anne C. Stone<sup>3</sup>, Molly Przeworski<sup>1\*</sup>, David Reich<sup>2,4\*</sup>

**1** Department of Human Genetics, University of Chicago, Chicago, Illinois, United States of America, **2** Broad Institute of Harvard and MIT, Cambridge, Massachusetts, United States of America, **3** School of Human Evolution and Social Change, Arizona State University, Tempe, Arizona, United States of America, **4** Department of Genetics, Harvard Medical School, Boston, Massachusetts, United States of America

**Little is known about the history and population structure of our closest living relatives, the chimpanzees, in part because of an extremely poor fossil record. To address this, we report the largest genetic study of the chimpanzees to date, examining 310 microsatellites in 84 common chimpanzees and bonobos. We infer three common chimpanzee populations, which correspond to the previously defined labels of “western,” “central,” and “eastern,” and find little evidence of gene flow between them. There is tentative evidence for structure within western chimpanzees, but we do not detect distinct additional populations. The data also provide historical insights, demonstrating that the western chimpanzee population diverged first, and that the eastern and central populations are more closely related in time.**

Citation: Becquet C, Patterson N, Stone A, Przeworski M, Reich D (2007) Genetic structure of chimpanzee populations. *PLoS Genet* 3(4): e66. doi:10.1371/journal.pgen.0030066

## Introduction

Standard taxonomies recognize two species of chimpanzees: bonobos (*Pan paniscus*) and common chimpanzees (*P. troglodytes*), whose current ranges in Africa do not overlap. Common chimpanzees have been classified further into three populations or subspecies based on their separation by geographic barriers (generally rivers): western (*P. troglodytes verus*), central (*P. t. troglodytes*), and eastern (*P. t. schweinfurthii*) [1,2]. While there are no or only slight morphological or behavioral differences among the common chimpanzees [3–5], genetic studies of mitochondrial DNA (mtDNA) [6,7] and the Y chromosome [8] have supported the geography-based population designations [6,8], and mtDNA studies have led to the proposal of a fourth common chimpanzee subspecies, *P. t. vellerosus*, around the Sanaga river in Cameroon [9,10]. However, studies of single loci provide at best partial information about history and population subdivision [11]; for example, analyses of X and Y chromosome datasets [12] suggest that genetic diversity is highest in central and lowest in western chimpanzees, while mtDNA suggests a different pattern [8]. Resequencing and microsatellite-based datasets have also provided inconsistent evidence about whether eastern chimpanzees are more diverse than bonobos [5,14]. To obtain a clear picture of chimpanzee population structure, a large number of independently evolving regions should be studied simultaneously.

The most comprehensive study of chimpanzees to date—including multiple loci and samples from western, central, and eastern chimpanzees and bonobos—found few fixed genetic differences among chimpanzee populations and estimated autosomal  $F_{ST}$  values between populations of 0.09–0.32, overlapping the range of differentiation seen in humans. Fischer et al. argued from these results that there are no chimpanzee subspecies and suggested instead that chimpanzee variation might be characterized by continuous gradients of gene frequencies, with ongoing gene flow across groups [5]. This and the other multilocus datasets that have been published to date [13–15] are small compared with recent genetic assessments of human structure [16], however, and have not yet provided a clear picture. For example, mtDNA and Y chromosome data have been interpreted as

showing discontinuity among chimpanzee populations [6–10], potentially at odds with the model proposed by Fischer et al. [5].

An accurate picture of chimpanzee population structure is also crucial for understanding their history. For example, Won and Hey estimated that common chimpanzees and bonobos split ~0.9 million years ago (Mya), and western and central chimpanzees split ~0.42 Mya, with low levels of migration from western to central since that time [17]. This analysis, which assumed that the populations split from a common ancestor, would need to be reevaluated if the data were better described by a model of stable isolation by distance [5].

To clarify chimpanzee population structure, we gathered an order-of-magnitude larger dataset than has previously been available. This allowed us to test whether genetic data alone can be used to assign chimpanzees to the categories of western, central, and eastern chimpanzees, whether there is evidence for substantial admixture between groups, and whether there is unrecognized substructure among the chimpanzees [18].

## Results/Discussion

We analyzed data from 310 polymorphic microsatellites in 84 individuals: 78 common chimpanzees and six bonobos. These samples were chosen to include multiple representatives of each putative population. Of the common chimpan-

**Editor:** Gilean A. T. McVean, University of Oxford, United Kingdom

**Received** January 8, 2007; **Accepted** March 13, 2007; **Published** April 20, 2007

A previous version of this article appeared as an Early Online Release on March 13, 2007 (doi:10.1371/journal.pgen.0030066.eor).

**Copyright:** © 2007 Becquet et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Abbreviations:** ASD, average squared distance; LR, likelihood ratio; mtDNA, mitochondrial DNA; Mya, million years ago; PCA, principal components analysis; SMM, stepwise mutation model;  $t_{MRC A}$ , time since the most recent common ancestor

\* To whom correspondence should be addressed. E-mail: mfp@uchicago.edu (MP); reich@genetics.med.harvard.edu (DR)

## Author Summary

Common chimpanzees have been traditionally classified into three populations: western, central, and eastern. While the morphological or behavioral differences are very small, genetic studies of mitochondrial DNA and the Y chromosome have supported the geography-based designations. To obtain a crisp picture of chimpanzee population structure, we gather far more data than previously available: 310 microsatellite markers genotyped in 78 common chimpanzees and six bonobos, allowing a high resolution genetic analysis of chimpanzee population structure analogous to recent studies that have elucidated human structure. We show that the traditional chimpanzee population designations—western, central, and eastern—accurately label groups of individuals that can be defined from the genetic data without any prior knowledge about where the samples were collected. The populations appear to be discontinuous, and we find little evidence for gradients of variation reflecting hybridization among chimpanzee populations. Regarding chimpanzee history, we demonstrate that central and eastern chimpanzees are more closely related to each other in time than either is to western chimpanzees.

zees, 41 were reported as western, 16 as central, seven as eastern, three as hybrids, and 11 did not have a reported subpopulation (Table 1). This dataset was designed to include a similar number of genetic markers (and in fact included many of the same markers) as the dataset analyzed by Rosenberg et al. to elucidate human population structure [16]. Because of high mutation rates, microsatellite alleles often have arisen multiple times, and hence it is difficult to resolve the genealogy at any locus. A benefit of the high mutation rate, however, is that microsatellites provide more information about recent historical events per locus compared with resequencing data [19].

### Cluster Analysis

To explore the genetic evidence for subdivision among chimpanzees, we first applied the program STRUCTURE to the dataset (Materials and Methods) [20]. Each STRUCTURE analysis requires a hypothesized number of populations and assigns individuals to these populations—without using any pre-assigned population labels—in a way that minimizes the amount of Hardy-Weinberg disequilibrium and linkage disequilibrium across widely separated markers. The analysis strongly supports the division of the samples of common chimpanzees and bonobos into at least four discontinuous subpopulations. Although the software does not provide a formal statistical procedure for choosing the number of clusters, Pritchard et al. [20] suggest using the model with the highest likelihood. When we ran the software assuming models of one to six clusters (averaging results for three random number seeds for each model), the likelihood of the data for four clusters was higher than for any other model. The inferred clusters correspond remarkably well to the reported labels of western, central, eastern, and bonobo, and also agree well with the assignments based on mtDNA or Y chromosome haplotypes (Figure 1; Table 1).

The multilocus dataset also provides power to identify individuals with multiple ancestries and to assess their ancestry proportions. This cannot be done reliably using studies of single loci such as the Y chromosome or

mtDNA, because individuals can in fact be descendants of multiple ancestral populations without carrying DNA from some of the populations at the locus being studied. The STRUCTURE analysis identified nine individuals as having more than 5% genetic ancestry from two clusters (Table 2).

Of the individuals identified by STRUCTURE as likely hybrids, seven were born in captivity, and just two were wild-caught, consistent with what would be expected if there were low rates of migration between central and western chimpanzees in the wild (Table 2) [17]. Interestingly, individual number 54, one of two wild-caught individuals identified as a hybrid by this analysis, has an mtDNA haplotype hypothesized to correspond to *P. t. vellorosos* [9]. The two captive-born chimpanzees with the putative *P. t. vellorosos* haplotype, however, have markedly different estimates of ancestry proportions, and thus there is no evidence from the STRUCTURE analysis that these individuals form a distinct population: the population ancestry estimates are 45% central and 55% western for number 54; 84% central and 16% western for number 78; and 100% western for number 67.

We also used STRUCTURE to validate a minimal set of markers that could be useful for classifying chimpanzees in conservation studies (Table S1). The top 30 markers (ranked by informativeness for assigning individuals to populations [19]) provide excellent power for classification. Of 75 chimpanzees estimated as having 100% ancestry in one group by all markers, we found that 71 were classified identically by the top 30 markers (by the criterion that at least 90% of the ancestry is assigned to the same group). Of nine individuals identified as hybrids with all the markers, six were also detected as hybrids with the reduced set. In addition to quantitative precision, the microsatellite panel also has a qualitative advantage over single marker studies in classifying chimpanzee hybrids: mtDNA and Y chromosome analyses cannot detect first generation female hybrids (Table 1) or reliably classify hybrids of the second or higher generation.

### Principal Components Analysis

We next carried out principal components analysis (PCA). This approach has been shown to have similar power to capture population structure as STRUCTURE, but also provides a formal way of assigning statistical significance to population subdivision [21]. When the PCA is applied to the chimpanzee data, the results support four discontinuous populations into which almost all chimpanzees and bonobos can be classified. The first three principal components (eigenvectors) are all highly statistically significant ( $p < 10^{-12}$ ) and nearly perfectly separate western, central, and eastern chimpanzees, and bonobos (Figure 2). Only six chimpanzees fall visually outside of the clusters, a subset of the nine identified by STRUCTURE as having at least 5% genetic contribution from more than one population (Table 2). The fourth eigenvector ( $p = 0.011$ ) is also significant, and the fifth is not significant ( $p = 0.44$ ).

The eigenvectors are strongly correlated to the population labels. We used nonparametric analysis (Kruskal-Wallis tests) to explore whether the values of each sample along the four significant eigenvectors were significantly correlated to the four pre-existing population labels. The overall statistic is

**Table 1.** Details of the 84 Samples in This Study

ID	Other Identifier(s)	Sex	Sample Source	Reported Category	After Genetic Analysis	Reported Birthplace	Classification Based on mtDNA/Y Chromosome Genotype
1	Amelie	f	Leipzig	Central	Central	Haut-Ogooué	
2	Chiquita	f	Leipzig	Central	Central	Haut-Ogooué	
3	Berthe	f	Leipzig	Central	Central	Captive born	
4	Bakoumba	m	Leipzig	Central	Central	Haut-Ogooué	Y, central
5	Noemie	f	Leipzig	Central	Central	Estuaire	
6	Clara	f	Leipzig	Central	Central	Gabon	
7	Minkebe	m	Leipzig	Central	Central	Captive born	
8	Masuku	f	Leipzig	Central	Central	Captive born	
9	Gemini	f	Leipzig	Central	Central	Estuaire	
10	Henri	m	Leipzig	Central	Central	Nyanga	Y, central
11	Ivindo	m	Leipzig	Central	Central	Ogooué-Ivindo	Y, central
12	Moanda	m	Leipzig	Central	Central	Haut-Ogooué	Y, central
13	Lalala	f	Leipzig	Central	Central	Estuaire	
14	Makata	m	Leipzig	Central	Central	Haut-Ogooué/Ogooué-Ivindo	Y, central
15	Makokou	f	Leipzig	Central	Central	Captive born	
16	Pt 197, stud number 277, IPBIR 496	m	Arizona	Central	Central	Wild caught, origin unknown	mtDNA, central
17	Akila	f	Leipzig	Eastern	Mostly or all eastern	Burundi	
18	Alley	f	Leipzig	Eastern	Eastern	Southeast Congo	
19	Amizero	f	Leipzig	Eastern	Eastern	Burundi	
20	Annie	f	Leipzig	Eastern	Eastern	Northeast Congo	
21	Judy	f	Leipzig	Eastern	Eastern	Southeast Congo	
22	Mimi	f	Leipzig	Eastern	Eastern	Northeast Congo	
23	Pt 169, ISIS number 3850	f	Arizona	Eastern	Western/eastern	Captive born	mtDNA, eastern
24	Annaclara	f	Leipzig	Western	Western	Captive born	
25	Frits	m	Leipzig	Western	Western	Sierra Leone	
26	Hilko	m	Leipzig	Western	Western	Captive born	
27	Lisbeth	f	Leipzig	Western	Western	Sierra Leone	
28	Louise	f	Leipzig	Western	Western	Captive born	
29	Marco	m	Leipzig	Western	Western	Sierra Leone	
30	Oscar	m	Leipzig	Western	Western	Captive born	
31	Regina	f	Leipzig	Western	Western	Sierra Leone	
32	Socrates	m	Leipzig	Western	Western	Captive born	
33	Sonja	f	Leipzig	Western	Western	Sierra Leone	
34	Yoran	m	Leipzig	Western	Western	Captive born	
35	Yvonne	f	Leipzig	Western	Western	Sierra Leone	
36	Pt 81, studbook number 380	f	Arizona	Western	Western	Sierra Leone	mtDNA, western; Y, western
37	Pt 82, studbook number 341	m	Arizona	Western	Western	Sierra Leone	mtDNA, western; Y, western
38	Pt 83, studbook number 459	f	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western
39	Pt 87, ISIS number 1149	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
40	Pt 88, ISIS number 1144	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
41	Pt 90, ISIS number 1339	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
42	Pt 97, ISIS number 2036	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
43	Pt 98, ISIS number 2724	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
44	Pt 99, studbook number 561	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
45	Pt 100, ISIS number 3000	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
46	Pt 101, ISIS number 3214	m	Arizona	Western	Western	Wild-caught, origin unknown	mtDNA, western; Y, western
47	Pt 102, ISIS number 1068	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
48	Pt 103, ISIS number 3340	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
49	Pt 104, ISIS number 3339	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
50	Pt 105, ISIS number 2435	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
51	Pt 106, stud number 430, ISIS 2377	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
52	Pt 107, stud number 142, ISIS 2474	f	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western
53	Pt 112, stud number 314	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
54	Pt 114, ISIS number 2412	m	Arizona	Western	Western/central	Wild caught, origin unknown	mtDNA, Nigerian; Y, western
55	Pt 115, ISIS number 2738	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
56	Pt 117, ISIS number 1641	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
57	Pt 120, ISIS number 2216	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
58	Pt 121, ISIS number 2549	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
59	Pt 122, ISIS number 2417	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
60	Pt 124, ISIS number 2404	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
61	Pt 125, ISIS number 2554	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
62	Pt 126, ISIS number 1818	m	Arizona	Western	Western	Wild caught, origin unknown	mtDNA, western; Y, western
63	Coriell NA03448	m	Coriell/IPBIR	Western	Western	Captive born	mtDNA, western; Y, western
64	Coriell NA03450	m	Coriell/IPBIR	Western	Western	Captive born	mtDNA, western; Y, western
65	Marilyne (Coriell NS03612)	f	Coriell/IPBIR	Unreported	Western/central	Captive born	mtDNA, western
66	Kipper (Coriell NS03629)	m	Coriell/IPBIR	Unreported	Western	Captive born	mtDNA, western; Y, western

Table 1. Continued.

ID	Other Identifier(s)	Sex	Sample Source	Reported Category	After Genetic Analysis	Reported Birthplace	Classification Based on mtDNA/Y Chromosome Genotype
67	Gay (Coriell NS03639)	f	Coriell/IPBIR	Unreported	Western	Captive born	mtDNA, Nigerian
68	Juan (Coriell NS03641)	m	Coriell/IPBIR	Unreported	Western/central	Captive born	Y, western
69	Lizzie (Coriell NS03646)	f	Coriell/IPBIR	Unreported	Mostly or all western	Captive born	mtDNA, western
70	Sheena (Coriell NS03650)	f	Coriell/IPBIR	Unreported	Western	Captive born	mtDNA, western
71	Jimoh (Coriell NS03657)	m	Coriell/IPBIR	Unreported	Western	Captive born	mtDNA, western; Y, western
72	Alicia (Coriell NS03659)	f	Coriell/IPBIR	Unreported	Western	Captive born	
73	Garbo (Coriell NS03660)	f	Coriell/IPBIR	Unreported	Western	Captive born	
74	Tank (Coriell NS03623)	m	Coriell/IPBIR	Unreported	Western	Captive born	mtDNA, western; Y, western
75	Kasey (Coriell NS03656)	f	Coriell/IPBIR	Unreported	Western	Captive born	mtDNA, western
76	Pt 13	m	Arizona	Hybrid	Mostly or all central	Captive born	mtDNA, central; Y, eastern
77	Pt 113, stud number 721	m	Arizona	Hybrid	Western/central	Captive born	mtDNA, central; Y, western
78	Pt 123, stud number 662	m	Arizona	Hybrid	Mostly central	Captive born	mtDNA, Nigerian; Y, central
79	Ulindi	f	Leipzig	Bonobo	Bonobo	Captive born	
80	Yasa	f	Leipzig	Bonobo	Bonobo	Captive born	
81	IPBIR number 092	f	Coriell/IPBIR	Bonobo	Bonobo	Captive born	
82	IPBIR number 251	m	Coriell/IPBIR	Bonobo	Bonobo	Captive born	
83	IPBIR number 367	f	Coriell/IPBIR	Bonobo	Bonobo	Captive born	
84	IPBIR number 661	m	Coriell/IPBIR	Bonobo	Bonobo	Captive born	

F, female; M, male; IPBIR, Integrated Primate Biomaterials and Information Resource; ISIS, International Species Identification System; Pt, *P. troglodytes*. doi:10.1371/journal.pgen.0030066.t001

highly significant ( $p < 10^{-10}$ ) for the first three eigenvectors but insignificant for the fourth ( $p = 0.97$ ), indicating that this eigenvector is capturing population subdivision that is different from the traditional western/central/eastern/bonobo designations.

To explore whether the fourth eigenvector might reflect an as-yet-undefined chimpanzee population, we carried out analyses separately on the western chimpanzee ( $n = 49$ ), central chimpanzee ( $n = 16$ ), eastern chimpanzee ( $n = 6$ ), and bonobo ( $n = 6$ ) samples (including all individuals that were clearly classified by both PCA and STRUCTURE). Western chimpanzees are the only population with evidence for internal substructure ( $p = 5.5 \times 10^{-5}$ ). The first eigenvector obtained when western chimpanzees are analyzed by themselves strongly correlates to the fourth eigenvector in the main analysis ( $r^2 = 0.92$ ;  $p < 10^{-12}$ ) (Figure S1), indicating that the fourth eigenvector describes subdivision within western chimpanzees.

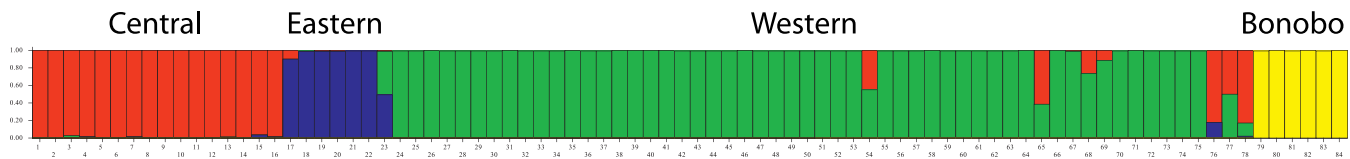
Although the fourth eigenvector seems to be detecting real structure, it does not mark out discontinuous subpopulations of western chimpanzees (Figure S1). The failure to reveal the details of the structure is evident not only in the PCA, but also in an application of STRUCTURE to the western chimpanzees only, in which a model of only one cluster is most likely. There is also no pattern to the classification of western chimpanzees even when we consider a model of two clusters (unpublished data). The most likely explanation is that there is not enough data to assign individuals to different ancestries. Understanding of the fourth eigenvector in the PCA will require more genetic data and better information about geographic origin. In particular, we note that the only wild caught western samples for which we had geographic information are from one location (Sierra Leone), thus we could not perform a test for correlation with geography.

### Testing for Additional Populations

Could there be additional population structure [18] among the chimpanzees that we have not yet detected? A particular concern is that our sample size is limited, decreasing our power to detect further structure especially among non-western chimpanzees.

To place an upper bound on further structure especially among the central chimpanzees, we considered the possibility that, among the 16 central chimpanzees, a subset is from a different population. We performed PCA on 10 central/6 eastern, 11 central/5 eastern, 12 central/4 eastern, 13 central/3 eastern and 14 central/2 eastern chimpanzees and assessed what fraction of 1,000 random resamplings of central and eastern chimpanzees showed evidence of structure ( $p < 0.05$ ). This allowed us to assess power to detect an additional population as diverged as the eastern chimpanzees.

The resampling analysis found that 6, 5, 4, 3, and 2 eastern chimpanzees could be detected from amidst the central chimpanzees with 100%, 100%, 99%, 54%, and 7% probability, respectively. Since the  $F_{ST}$  between central and eastern chimpanzees is 0.05–0.09 (Table 2) [5], this allowed us to place an upper bound on the undetected structure that might exist among central chimpanzees given that we did not detect further structure. If the three samples with the *P. t. vellorossus* mtDNA haplotype in our study constitute members of a distinct population, their differentiation from central chimpanzees is likely to be  $F_{ST} \leq 0.09$ , lower than those observed between some pairs of human populations [22]. An important caveat is that we have no power to detect population structure for chimpanzees missed by our sampling (we also have little power if there are fewer than three individuals from a population). Thus, a more geographically systematic survey, including more animals from a denser grid in Africa, may detect further structure.



**Figure 1.** STRUCTURE Analysis, Blinded to Population Labels, Recapitulates the Reported Population Structure of the Chimpanzees  
Individuals 76–78 are reported hybrids. Only two individuals with a >5% proportion of ancestry in more than one inferred cluster are wild born: number 54 and number 17. Red, central; blue, eastern; green, western; yellow, bonobo.  
doi:10.1371/journal.pgen.0030066.g001

### Evidence for Inbreeding

To test for inbreeding among the chimpanzees, we examined whether heterozygosity within individuals was significantly lower than would be expected from random mating in the population (Materials and Methods). Western and central chimpanzees both show evidence for a reduced number of heterozygous genotypes ( $p < 0.05$ ) (Protocol S1) (we had too few eastern and bonobo samples to perform an informative test). A caveat is that misscoring of heterozygous genotypes, or the presence of polymorphisms under the primers used for genotyping, could both result in an artifactual excess of homozygotes. To follow up this initial evidence of inbreeding among chimpanzees, further analyses could search for multimegabase contiguous stretches of homozygosity [23].

### First and Second Generation Hybrids

To test for first and second generation hybrids, we calculated the likelihood of the data under the hypothesis that an individual is an  $F_1$  hybrid, compared with the alternative hypothesis of an older 50%–50% mixture of the ancestral populations. To test whether the individual is an  $F_2$ /backcross—a mixture of an  $F_1$  with an unadmixed individual—we compared the likelihood of this model compared with the alternative hypothesis of an older 75%–25% mixture of the two ancestral populations (Materials and Methods).

Of the nine putative hybrids identified by STRUCTURE, the  $F_1$  hybrid test identifies captive-born individual number 23 (an approximately 50%–50% eastern-western hybrid by STRUCTURE analysis) as an  $F_1$ , with a likelihood ratio (LR) of

~24,000,000:1. The  $F_2$ /backcross test identifies the captive-born individual number 68 (a 74%–26% western-central hybrid by STRUCTURE analysis) as an  $F_2$ /backcross, with an LR of ~37:1 (the evidence is weaker because the signal of an  $F_2$ /backcross is more subtle). There are no other hybrids identified by either the  $F_1$  or  $F_2$ /backcross test, suggesting that the other animals in Table 2 could descend from third generation or older admixture events, or be members of as-yet unidentified populations.

The  $F_1$  test produced a particularly intriguing pattern in number 54, a wild-caught individual with mtDNA that has been hypothesized to be diagnostic of *P. t. vellorosos* origin [9,10]. Individual number 54 is estimated to be a 55%–45% western-central mixture (Table 2) and shows an LR of 7:1 in favor of being an old mixture, compared with the alternative of a first generation  $F_1$  hybrid. However, a careful examination shows that the pattern of variation at number 54 fits neither the hypothesis of a first generation hybrid or an older mixture. To demonstrate this, we simulated 100 different western-central  $F_1$  hybrids and 100 older western-central mixtures by random sampling from the population allele frequencies. Simulated older mixtures always generated an LR of >100,000:1 relative to the alternative hypothesis of an  $F_1$ . Simulated  $F_1$  hybrids always gave an LR < 1:2. The LR for individual number 54 of 7:1 falls outside of either expectation. This individual fits neither model, suggesting ancestry from an as-yet undetermined population.

### Allele Frequency Differentiation

To estimate the degree of allele frequency differentiation between chimpanzee groups, we computed the  $R_{ST}$  statistic, a

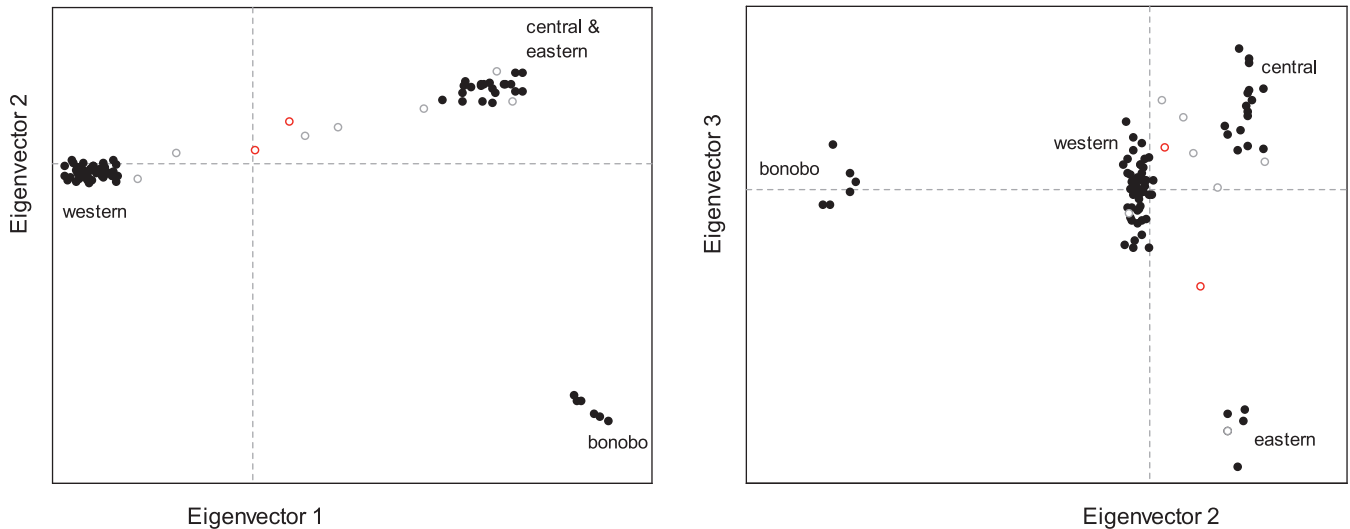
**Table 2.** Individuals with >5% Ancestry from More than One Cluster

ID	Sex	Reported Status	STRUCTURE Analysis			PCA (Estimate of Percentage Is Qualitative)	Other Genetic Information from mtDNA and Y Chromosome
			W	C	E		
17	F	Eastern		9	91	All eastern	
23	F	Eastern	49	1	50	~50% eastern, ~50% western	mtDNA, eastern
54	M	Western	55	45		~50% western, ~50% central	mtDNA, vellorosos
65	F	Unknown	39	61		~50% western, ~50% central	mtDNA, western
68	M	Unknown	74	26		~65% western, ~35% central	Y, western
69	F	Unknown	89	11		All western	mtDNA, western
76	M	Hybrid		81	19	All central	mtDNA, central; Y, eastern
77	M	Hybrid	50	50		~50% western, ~50% central	mtDNA, western; Y, central
78	M	Hybrid	15	82	3	~15% western, ~85% central	mtDNA, vellorosos; Y central

All nine individuals in this table are indicated by STRUCTURE to have >5% ancestry from at least two populations. Of these, two are wild born: number 17 and number 54. PCA confirms the mixed ancestry of six individuals (number 23, number 54, number 65, number 68, number 77, and number 78) (compare Figures 1 and 2).

F, female; M, male.

doi:10.1371/journal.pgen.0030066.t002



**Figure 2.** PCA, Without Using Population Labels, Divides the 84 Chimpanzees into Four Apparently Discontinuous Populations of Western, Central, Eastern, and Bonobo

Plots of eigenvectors 1 versus 2, and eigenvectors 2 versus 3, show clustering into populations, with the expected assignments for the 75 individuals identified as all of one ancestry by STRUCTURE (solid circles). The nine individuals identified by STRUCTURE as hybrids (open circles) are for the most part identified as hybrids by PCA as well. There are two individuals (red open circles) reported as being of a particular population but that in fact appear to be hybrids: number 23, reported as eastern but in fact a western-eastern hybrid, and number 54, a wild-born individual reported as western but in fact a western-central hybrid.

doi:10.1371/journal.pgen.0030066.g002

microsatellite-based estimator of  $F_{ST}$  [24,25].  $R_{ST}$  assumes the stepwise mutation model (SMM), in which the number of repeats changes by one or two or more units with an equal probability of increasing or decreasing. A goodness of fit test suggests that this simple model provides a reasonable match to our data (Figure S2). Encouragingly, the  $R_{ST}$  estimates of population differentiation, obtained based on assuming this model, are very similar to estimates of  $F_{ST}$  from a smaller multilocus dataset based on resequencing (Table 3) [5].

A particularly intriguing feature of the allele frequency differentiation results is that the allele frequency differentiation between bonobos and western chimpanzees is higher than that between bonobos and central or eastern chimpanzees (Table 3). This likely reflects greater genetic drift in the western lineage since divergence, as has also been suggested by an analysis of resequencing data [17].

**Table 3.** Genetic Differentiation among Populations

Location	Eastern	Central	Bonobo
Western	0.31 (0.32)	0.25 (0.29)	0.68 (0.68)
Eastern	—	0.05 (0.09)	0.57 (0.54)
Central	—	—	0.51 (0.49)

Pairwise  $R_{ST}$  (versus  $F_{ST}$  from [5]) is shown.  $R_{ST}$  (a microsatellite-based estimator of  $F_{ST}$  [24]) is calculated using the Arlequin software [25] comparing 49 western, six eastern, and 16 central chimpanzees, and six bonobos. Analysis is restricted to autosomal loci with <5% missing data (leaving  $\geq 220$  markers in all cases). For the 49 western chimpanzees, we used the 48 individuals identified by western by STRUCTURE plus individual number 54, which was born in the western range. All  $R_{ST}$  values are significantly different from zero ( $p < 0.002$ ), as determined by 10,000 permutations. The values in parentheses are quoted from the SNP-based study of Fischer et al. [5]. Our study has less sampling error but relies on imperfect assumptions about the microsatellite mutation process, and so is more subject to systematic error. The close agreement between the two studies is encouraging. doi:10.1371/journal.pgen.0030066.t003

### Central and Eastern Chimpanzees Are Most Closely Related in Time

The high frequency differentiation of western chimpanzees compared with other groups (Table 3) is consistent with them having been the first population to diverge, but does not prove it. An alternative explanation for the data is that there has been a smaller effective population size on the western lineage since their divergence, resulting in high genetic drift in this population. We therefore applied a formal test to assess which pair of populations is most closely related.

We approached the problem by testing whether three unrooted phylogenetic trees are consistent with the data for chimpanzees: (1) western-central and bonobo-eastern forming clades, (2) western-eastern and bonobo-central forming clades, and (3) eastern-central and bonobo-western forming clades. If a tree provides a good description of the history of the population, then the allele frequency differences between two populations should only reflect changes since they split. For example, the difference in allele frequency between central and eastern chimpanzees should have arisen entirely since their divergence from a common ancestral population and so should be uncorrelated to the allele frequency differences between western chimpanzees and bonobos.

To implement this idea, we calculated the difference in frequency within clades for all alleles and then tested for a correlation across clades. When we carry out this analysis for the first and second hypothesized trees, a correlation is observed, rejecting these trees at a significance of  $p = 0.00025$  and  $p = 0.0027$ , respectively (Table 4). The hypothesized central-eastern/bonobo-western clade is the only one consistent with the data ( $p = 0.37$ ). Thus, our analysis does not find any evidence for gene flow between western and central chimpanzees since their initial split, as has previously been hypothesized [17]. If gene flow did occur, it would have had to

**Table 4.** Eastern and Central Chimpanzees Phylogenetically Most Closely Related

Clade 1	Clade 2	Correlation between Allele Frequency Differences in Each Clade	p-Value (Two-Sided)
Central-western	Eastern-bonobo	0.090	0.00025
Eastern-western	Central-bonobo	0.065	0.0027
Central-eastern	Western-bonobo	-0.013	0.37

There are three possible unrooted trees relating to the four populations. If the clades into which the trees are partitioned correctly capture the population relationships, the allele frequencies should be uncorrelated when comparing clade 1 and clade 2. We observe significant correlation across clades for all phylogenetic trees other than the one in which central and eastern chimpanzees cluster. To correct for the nonindependence of microsatellite alleles, we calculated significance by a weighted jackknife analysis removing each marker in turn to generate normally distributed Z-scores; these were then converted to *p*-values.  
doi:10.1371/journal.pgen.0030066.t004

be sufficiently low to fall below the threshold of detection with our present data size and the test we applied. We conclude that eastern and central are more closely related in time than either population is to western chimpanzees.

### Population Separation Times

To estimate the times of genetic divergence among chimpanzees, we used the average squared distance (ASD) statistic [26]. For microsatellites evolving under the SMM, the expected time since the most recent common ancestor ( $t_{\text{MRCA}}$ ) of two chromosomes is expected to be  $\text{ASD}/2\mu$ , where  $\mu$  is the average mutation rate per year per locus, averaged across loci. Because allele lengths change according to a random walk, the ASD between allele lengths in two chromosomes is expected to increase linearly with time and is thus expected to act like heterozygosity in sequence comparisons. By averaging ASD over pairs of chromosomes within and across populations, we can estimate the average  $t_{\text{MRCA}}$  within and across populations.

The results confirm that genetic diversity (heterozygosity) is least for western chimpanzees and bonobos, higher for eastern chimpanzees, and highest for central chimpanzees, consistent with results obtained from a nucleotide resequencing

dataset [5]. To estimate the absolute  $t_{\text{MRCA}}$  within and across populations, we used two estimates of the microsatellite mutation rate. The first,  $\mu = 6.57 \times 10^{-3}$  per year, relies on a 7 Mya estimated average  $t_{\text{MRCA}}$  between humans and chimpanzees and the observation that two western chimpanzees are  $\sim 14.8$  times less genetically diverged than humans and chimpanzees [27]. We also obtained a second estimate,  $4.71 \times 10^{-5}$  per year, based on estimated rates of microsatellite mutation in humans, and assuming 15 years per generation (Table 5).

The absolute values of the time estimates for these  $t_{\text{MRCA}}$ s should be treated with caution because of uncertainty about the microsatellite mutation rate process and the calibrations used to obtain absolute dates. Nevertheless, the  $t_{\text{MRCA}}$  estimates are consistent with previous results from smaller resequencing based datasets [8]. We note that the central-eastern, central-western, and central-central  $t_{\text{MRCA}}$ s are all similar, which appears at first to contradict the claim that the populations split at different times. However, most of the genetic divergence reflects ancestral diversity, which is shared among all the chimpanzees (explaining why the  $t_{\text{MRCA}}$  estimates are substantially older than estimates of population splitting times [14,17]). More refined analyses are needed,

**Table 5.** Estimates of Divergence Time from ASD

Time Since the Most Recent Common Genetic Ancestor ( $t_{\text{MRCA}}$ )	$t_{\text{MRCA}}$ in Mya, Assuming 7 Mya for Human-Chimp Genetic Divergence (Calibration Time) <sup>a</sup>	$t_{\text{MRCA}}$ in Mya (Using Microsatellite Mutation Rate Estimates from Humans) <sup>a</sup>
Within-western	0.47	0.71 (0.62–0.81)
Within-central	0.85 (0.75–0.98)	1.29 (1.15–1.45)
Within-eastern	0.73 (0.61–0.86)	1.09 (0.93–1.28)
Within-bonobo	0.63 (0.53–0.76)	0.95 (0.81–1.10)
Central-eastern	0.89 (0.77–1.03)	1.35 (1.20–1.53)
Central/eastern-western	0.84 (0.73–0.97)	1.30 (1.16–1.43)
Central/eastern/western-bonobo	1.56 (1.29–1.90)	2.36 (1.97–2.79)

$t_{\text{MRCA}}$  represents the average time to the most recent common ancestor of a pair of chromosomes sampled in the same or different populations. It can be substantially older than the split time, as it also reflects differences accumulated in the ancestral population (90% confidence intervals from 10,000 bootstraps).

<sup>a</sup>An absolute  $t_{\text{MRCA}}$  for western chimpanzees is obtained by assuming that the human-chimpanzee  $t_{\text{MRCA}}$  is  $\sim 7$  Mya. We calibrate the  $t_{\text{MRCA}}$  for western chimpanzees at 0.47 Mya, since human-chimpanzee sequence divergence is estimated to be 14.8 times higher than western-western divergence [27]. An alternative estimate of the absolute dates comes from setting the mutation rate for microsatellites for humans to be  $7.06 \times 10^{-4}$  per generation and assuming 15 years per generation. This is obtained from direct estimates of mutation rates in humans for tetra-, tri-, and dinucleotides [37], adjusting for the relative proportions of each type of marker in our dataset: 222 tetranucleotide (including marker D125297, which was observed to have an unusually high mutation rate), 62 trinucleotide, and 11 dinucleotide.

doi:10.1371/journal.pgen.0030066.t005



such as the allele frequency correlation analysis presented in Table 4, or model-based approaches, to detect the subtle patterns that indicate the splitting order of the chimpanzee populations.

## Conclusions

We have carried out the largest analysis of chimpanzee genetic variation to date, which shows that the western, central, and eastern chimpanzee subspecies designations correspond to clusters of individuals with similar allele frequencies that can be defined from the genetic data without regard to the population labels. Moreover, we find little evidence for admixture between groups in the wild. Our analysis also provides the first formal test showing that the central and eastern chimpanzee populations are more closely related to each other in time than either is to western chimpanzees.

PCA also further suggests population structure within western chimpanzees. However, more data—more samples, genetic markers, and information about geographic origin—would be needed to understand this structure. We find no support for the proposed fourth population of common chimpanzees *P. t. vellerosus*. However, the failure to detect a distinct population cluster for these individuals could simply reflect a lack of power. Our analysis does allow us to state that even if *P. t. vellerosus* is a distinct population, its level of allele differentiation from either western, central, or eastern chimpanzees is not likely to exceed  $F_{ST} = 0.09$ .

We finally emphasize that although we attempted to include chimpanzees from as wide a range of sites in Africa as possible, the geographic sampling of the chimpanzees that we studied was likely nonrandom. The fact that our study did not include chimpanzees from some regions—including where chimpanzees are now extinct—could create the appearance of too much discontinuity [28]. Future studies including chimpanzees across a denser grid of populations within Africa could, in principle, identify intermediate populations of chimpanzees and demonstrate more graded patterns of variation.

## Materials and Methods

**Data collection.** The samples for this study were assembled from four sources: DNA collections at the Max Planck Institute (Leipzig, Germany), Anne Stone's laboratory at Arizona State University [8], the Coriell Cell Repositories (Camden, New Jersey, United States) [29], and the Integrated Primate Biomaterial and Information Resource (Camden, New Jersey, United States) [30]. A total of 91 samples were genotyped, and 84 were included in analysis after filtering (below). We had information about the approximate geographic origin of 25 wild-caught chimpanzees (Table 1). For most remaining samples, we had a population designation, and sometimes Y chromosome and mtDNA genotypes (A. Stone, unpublished data). As far as possible, we attempted to use pedigree information to remove related individuals from the captive chimpanzees.

We assembled all the DNA samples at a single site (the Broad Institute) and carried out whole-genome DNA amplification (WGA) for all samples to generate a quantity sufficient for analysis [31]. The WGA samples were shipped to a company that specializes in genotyping microsatellite markers for human disease gene mapping studies (PreventionGenetics, <http://www.preventiongenetics.com>). The microsatellite markers all contain tandem repeats of two, three, or four nucleotides that vary in repeat number across individuals. For example, at a single marker, different individuals might have between three and 11 contiguous repeats of a GATA sequence. The assays used for genotyping were designed for humans. However, we hypothesized that many of them would work for chimpanzees because of the ~98.8% sequence similarity [32].

Assays were attempted for 470 microsatellites. Most came from the Marshfield Screening Set 13 (designed for linkage screens in humans [33,34]) and were supplemented with markers from a separate mapping study. Genotyping quality was assessed by specialists at PreventionGenetics using standard measures of genotyping performance. A score of one to four was given to each marker (with one being the best and four the worst). Markers were scored as >2 because of uncertainty in allele assignment, or an excessive number of missing genotypes, or an excess in the numbers of homozygotes or noninteger alleles (defined as alleles with noninteger length differences compared with frequent alleles). We used the 310 markers that were designated as of highest or second-highest quality because the two sets produced indistinguishable inferences on our data (unpublished). For all analyses other than the use of STRUCTURE, we considered only autosomal or pseudoautosomal markers, since these could be treated uniformly. This resulted in 295 markers; we also excluded two additional pseudoautosomal markers for the PCA and  $F_1/F_2$  hybrid analyses. Genotypes for all markers are available in Dataset S1.

A subset of 84 of the 91 genotyped samples were chosen for further study after removing two due to a high missing data rate, one due to evidence for contamination (more than two genotypes at many loci), and four due to evidence of genetic relatedness: two accidental duplicates, and two apparent first degree relatives. For each pair of related individuals, we dropped the one with the lower success rate. The duplicate individuals allowed us to assess genotyping quality. For the two individuals studied in duplicate, 1.18% of genotyping calls differed, suggesting an error rate per genotype of ~0.59% (i.e., we estimate that on average approximately two loci were mistyped per individual).

**Data analysis.** We applied two complementary methods to characterize population structure in chimpanzees. First, we used the software STRUCTURE in the “admixture” mode, so that individuals were allowed to have ancestry from multiple populations. We used a model of correlated allele frequencies, a “burn-in” of 100,000 Markov Chain Monte Carlo (MCMC) iterations, and 1,000,000 follow-on MCMC iterations.

We also analyzed the data using a new implementation of PCA [21] available online in the EIGENSOFT software package [35]. Briefly, the analysis begins with a rectangular matrix, with the 84 rows corresponding to the individuals, and the columns corresponding to the alleles (the cells give the number of copies of each allele for each individual: zero, one, or two). To analyze the data, we perform a singular value decomposition, a procedure that produces eigenvectors and eigenvalues. The first eigenvector separates the samples in a way that explains the largest amount of variability, while the second and subsequent ones explain lesser amounts of variability. Thus, the first eigenvector distinguishes individuals from the population that is most differentiated, and each subsequent eigenvector separates the next most differentiated population. Eigenvalues above a significance cutoff represent significant population structure for the associated eigenvector [21].

To examine evidence for inbreeding in these samples, we used the output of STRUCTURE to assign individuals to populations, excluding individuals with >5% ancestry from more than one population and focusing our study on the population samples with more than ten individuals (treating captive and wild-caught individuals separately). Thus, we analyzed 48 western chimpanzees, including 34 wild-caught and 14 captive-born individuals. We also analyzed 13 wild-caught central chimpanzees (Table 1). We considered two statistics:  $H_w$ , the average heterozygosity within an individual's two chromosomes, and  $R_w$ , calculated as  $\frac{\sum_m (m_1 - m_2)^2}{M}$ ,

where  $m_1$  and  $m_2$  are the alleles' number of repeat units at marker  $m$  within an individual, and  $M$  is the total number of markers considered. We used the average value of  $H_w$  and  $R_w$  over individuals in a population to test the hypothesis of random mating and assessed significance by a permutation test. Specifically, we generated 1,000 samples of  $n$  individuals, randomly assigning to each of them two alleles from the pool, then counted how often  $H_w$  or  $R_w$  was as small or smaller than observed.  $H_w$  was significantly lower than expected for all samples considered (at the  $p < 0.05$  level), while  $R_w$  was significantly lower in wild-caught western chimpanzees (Protocol S1).

For other analyses in which we were interested in studying only individuals identified unambiguously as being from one population, we excluded the captive-born individuals defined as putative hybrids by STRUCTURE.

**Application of the stepwise mutation model.** We examined whether our data fit the SMM in the 49 individuals that are identified as western chimpanzees by both STRUCTURE and PCA. We focused on western chimpanzees because they have the largest sample size and

hence provide us with the most power to detect a departure from the SMM. Under the one-step SMM,  $\sigma_i^2$ , the variance of an allele with  $i$  repeats, is an estimator of  $2N\mu$ , the population mutation parameter [36]. For each marker, we calculated  $E(\sigma_i^2)$ , the expectation over all alleles of a given marker, thus obtaining an estimate of  $2N\mu$  for the 221 tetra-, 62 tri-, and 11 dinucleotides included in other analyses. Averaging this estimate over all markers of the same type and dividing by  $N = 10,000$  [17], we obtain an estimate of  $\mu$  for each type of marker,  $\hat{\mu} = 3.77 \times 10^{-3}$ ,  $1.91 \times 10^{-3}$ , and  $2.17 \times 10^{-3}$ , respectively. These estimates are roughly similar to independent estimates [37] based on microsatellites in humans:  $6.40 \times 10^{-4}$ ,  $7.10 \times 10^{-4}$ , and  $1.51 \times 10^{-3}$ , respectively.

To assess the goodness of fit of the SMM, we compared the observed distribution of  $\sigma_i^2$  to the expected distribution, obtained by using the coalescent simulator SIMCOAL2 [38]. We generated 500 independent replicates for each type of marker under a standard neutral equilibrium model, with an effective population size [17] of 10,000, a sample size of 98 chromosomes, and a mutation rate per generation set to  $\hat{\mu}$ . The range constraints for the number of repeat units were set to be equal to the maximum repeat observed in the sample for each type of marker. We tested whether there was a significant difference in the distributions by an asymptotic Wilcoxon rank sum test, carrying out the test separately for each type of marker. The observed distributions do not differ significantly from the expectation under the SMM (Figure S2).

**Tests for  $F_1$  and  $F_2$ /backcross hybrids.** We calculated a log Bayes-factor to test the hypothesis that a chimpanzee is an  $F_1$  hybrid of two known populations versus the alternative that it is a 50%–50% mixture (i.e., it is an older hybrid). For a given autosomal marker, one can compute a log-factor under the assumption that the allele frequencies are known; these log-factors can then be summed across all markers. In practice, our population samples are small, and so allele frequency estimates are imprecise. To account for uncertainty in the allele frequencies, we used a hierarchical Bayesian model for the unknown frequencies, with a Dirichlet prior distribution for the frequencies (the details of this calculation are similar to those described by Lockwood et al. [39]). We verified the performance of the test by simulation (see text).

## Supporting Information

**Dataset S1.** Raw Genotype Data in a Format Appropriate for STRUCTURE Analysis

Found at doi:10.1371/journal.pgen.0030066.sd001 (305 KB TXT).

**Figure S1.** The Significant Fourth Eigenvector from the Analysis of All 84 Chimpanzees Is Correlated to the First Eigenvector from Analysis of Western Chimpanzees Only ( $r^2 = 0.92$ )

Here, we present the correlation for the 49 individuals that are clearly identified as western chimpanzees by both STRUCTURE and

PCA, demonstrating that these eigenvectors are revealing the same population structure.

Found at doi:10.1371/journal.pgen.0030066.sg001 (11 KB PDF).

**Figure S2.** Goodness of Fit of the SMM

Distributions of the expected  $\sigma_i^2$  over markers for observed and simulated datasets. The distributions of  $E(\sigma_i^2)$  were computed separately for (a) 221 tetra-, (b) 62 tri-, and (c) 11 dinucleotides genotyped in the 49 western samples (shown in blue). In red are the results of 500 simulations for each class of microsatellites. In all three cases, the observed distribution is not significantly different from the expected distribution, as assessed by a permutation test (see Materials and Methods for more details).

Found at doi:10.1371/journal.pgen.0030066.sg002 (51 KB PDF).

**Protocol S1.** Testing for Inbreeding

Found at doi:10.1371/journal.pgen.0030066.sd002 (51 KB PDF).

**Table S1.** Information on Microsatellites Used for This Study

Found at doi:10.1371/journal.pgen.0030066.st001 (22 KB PDF).

## Acknowledgments

We thank the Albuquerque Biological Park, Detroit Zoo, Lincoln Park Zoo, Riverside Zoo, Sunset Zoo, the Primate Foundation of Arizona, New Iberia Research Center, and the Southwest Foundation for Biomedical Research for sharing chimpanzee samples. We thank Gavin McDonald for sample processing and Jim Weber at PreventionGenetics for personally ensuring that the genotypes for the chimpanzees were carefully scored. We thank Jean Wickings, Svante Pääbo, Philip Morin, Anne Fischer, and four anonymous reviewers, for sharing their samples and/or comments on earlier versions of the manuscript. We are also grateful to Jennifer Caswell, Graham Coop, Sridar Kudaravalli, Daniel Lieberman, Simon Myers, John Novembre, David Pilbeam, Alkes Price, and Maryellen Ruvolo for useful discussions.

**Author contributions.** MP and DR conceived and designed the experiments. CB, MP, and DR performed the experiments and wrote the paper. CB, NP, MP, and DR analyzed the data. CB, NP, ACS, MP, and DR contributed reagents/materials/analysis tools. MP and DR cosupervised the work.

**Funding.** This work was supported by a National Institutes of Health K-01 career transition award to NP, an Alfred P. Sloan Fellowship to MP, and a Burroughs Wellcome Career Development Award to DR. Chimpanzee-sampling and subspecies identification analyses were supported by a grant from the National Science Foundation (BCS-0073871) to AS.

**Competing interests.** The authors have declared that no competing interests exist.

## References

- Hill WCO (1969) The chimpanzee; a series of volumes on the chimpanzee. In: Bourne GH, editor. Volume 1. Basel, New York: S. Karger AG. pp. 22–49.
- Groves CP (2001) Primate taxonomy. Washington (D. C.): Smithsonian Institution Press. 350 p.
- Albrecht GH, Miller JMA (1993) Geographic variation in primates. A review with implications for interpreting fossils. In: Species, species concepts, and primate evolution. In: Kimbel WH, Mar LB, editors. New York: Plenum Press. pp. 123–161.
- Shea BT, Leigh SR, Groves CP (1993) Multivariate craniometric variation in chimpanzees: implications for species identification in paleoanthropology. In: Species, species concepts, and primate evolution. In: Kimbel WH, Mar LB, editors. New York: Plenum Press. pp. 265–296.
- Fischer A, Pollack J, Thalmann O, Nickel B, Paabo S (2006) Demographic history and genetic differentiation in apes. *Curr Biol* 16: 1133–1138.
- Morin PA, Moore JJ, Chakraborty R, Jin L, Goodall J, et al. (1994) . Kin selection, social structure, gene flow, and the evolution of chimpanzees *Science* 265: 1193–1201.
- D'Andrade RG, Morin PA (1996) A principle components and individual-by-site analysis of chimpanzee and human mitochondrial DNA. *Am Anthropol* 98: 352–370.
- Stone AC, Griffiths RC, Zegura SL, Hammer MF (2002) High levels of Y-chromosome nucleotide diversity in the genus *Pan*. *Proc Natl Acad Sci U S A* 99: 43–48.
- Gonder MK, Oates JF, Disotell TR, Forstner MR, Morales JC, et al. (1997) A new west African chimpanzee subspecies? *Nature* 388: 337.
- Gonder MK, Disotell TR, Oates JF (2006) New genetic evidence on the evolution of chimpanzee populations and implications for taxonomy. *Int J Primatol* 27: 1103–1127.
- Hudson RR, Coyne JA (2002) Mathematical consequences of the genealogical species concept. *Evolution Int J Org Evolution* 56: 1557–1565.
- Kaessmann H, Wiebe V, Paabo S (1999) Extensive nuclear DNA sequence diversity among chimpanzees. *Science* 286: 1159–1162.
- Yu N, Jensen-Seaman MI, Chennick L, Kidd JR, Deinard AS, et al. (2003) Low nucleotide diversity in chimpanzees and bonobos. *Genetics* 164: 1511–1518.
- Reinartz GE, Karron JD, Phillips RB, Weber JL (2000) Patterns of microsatellite polymorphism in the range-restricted bonobo (*Pan paniscus*): Considerations for interspecific comparison with chimpanzees (*P. troglodytes*). *Mol Ecol* 9: 315–328.
- Fischer A, Wiebe V, Paabo S, Przeworski M (2004) Evidence for a complex demographic history of chimpanzees. *Mol Biol Evol* 21: 799–808.
- Rosenberg NA, Pritchard JK, Weber JL, Cann HM, Kidd KK, et al. (2002) Genetic structure of human populations. *Science* 298: 2381–2385.
- Won YJ, Hey J (2002) Divergence population genetics of chimpanzees. *Mol Biol Evol* 22: 297–307.
- Gagneux P (2002) The genus *Pan*: Population genetics of an endangered outgroup. *Trends in Genet* 18: 327–330.
- Rosenberg NA, Li LM, Ward R, Pritchard JK (2003) Informativeness of genetic markers for inference of ancestry. *Am J Hum Genet* 73: 1402–1422.
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* 155: 945–959.
- Patterson N, Price AL, Reich D (2006) Population structure and eigenanalysis. *PLoS Genet* 2: e190. doi:10.1371/journal.pgen.0020190
- Cavalli-Sforza L, Menozzi P, Piazza A (1994) The history and geography of human genes. Princeton (N.J.): Princeton University Press. 518 p.

23. Carothers AD, Rudan I, Kolcic I, Polasek O, Hayward C, et al. (2006) Estimating human inbreeding coefficients: Comparison of genealogical and marker heterozygosity approaches *Ann Hum Genet* 70: 666–676.
24. Slatkin M (1995) A measure of population subdivision based on microsatellite allele frequencies. *Genetics* 139: 457–462.
25. Schneider S, Roessli D, Excoffier L (2000) Arlequin: A software for population genetics data analysis, version 2000 [computer program]. Geneva: Department of Anthropology, University of Geneva.
26. Goldstein DB, Pollock DD (1997) Launching microsatellites: A review of mutation processes and methods of phylogenetic interference *J Hered* 88: 335–342.
27. Patterson N, Richter DJ, Gnerre S, Lander ES, Reich D (2006) Genetic evidence for complex speciation of humans and chimpanzees. *Nature* 441: 1103–1108.
28. Kittles RA, Weiss KM (2003) Race, ancestry and genes: Implications for defining disease risk. *Annu Rev Genomics Hum Genet* 4: 33–67.
29. Coriell Cell Repositories [Internet]. Camden (NJ): Coriell Institute for Medical Research. Available: [http://locus.umdnj.edu/primates/species\\_summ.html](http://locus.umdnj.edu/primates/species_summ.html). Accessed 20 March 2007.
30. Integrated Primate Biomaterials and Information Resource [Internet]. Individual Samples Currently Available. Camden (NJ): Coriell Institute for Medical Research. Available: [http://www.ipbir.org/ipbir\\_\\_cgi/tax.cgi?mode=8&id=init&lvl=0](http://www.ipbir.org/ipbir__cgi/tax.cgi?mode=8&id=init&lvl=0). Accessed 20 March 2007.
31. Hosono S, Faruqi AF, Dean FB, Du Y, Sun Z, et al. (2003) Unbiased whole-genome amplification directly from clinical samples. *Genome Res* 13: 954–964.
32. Chimpanzee Sequencing and Analysis Consortium (2005) Initial sequence of the chimpanzee genome and comparison with the human genome. *Nature* 437: 69–87.
33. Mammalian Genotyping Service [Internet]. Marshfield (WI): National Heart, Lung, and Blood Institute. Available: <http://research.marshfieldclinic.org/genetics>. Accessed 20 March 2007.
34. Ghebranious N, Vaske D, Yu A, Zhao C, Marth G, et al. (2003) STRP screening sets for the human genome at 5 cM density. *BMC Genomics* 24: 6.
35. EIGENSOFT software package [Internet]. Boston: Reich Laboratory, Harvard Medical School, Department of Genetics. Available: <http://genepath.med.harvard.edu/~reich/Software.htm>. Accessed 20 March 2007.
36. Valdes AM, Slatkin M, Freimer NB (1993) Allele frequencies at microsatellite loci: The stepwise mutation model revisited. *Genetics* 133: 737–749.
37. Zhivotovsky LA, Rosenberg NA, Feldman MW (2003) Features of evolution and expansion of modern humans, inferred from genomewide microsatellite markers. *Am J Hum Genet* 72: 1171–1186.
38. Excoffier L, Novembre J, Schneider S (2000) SIMCOAL: A general coalescent program for the simulation of molecular data in interconnected populations with arbitrary demography. *J Hered* 91: 506–509.
39. Lockwood JR, Roeder K, Devlin B (2001) A Bayesian hierarchical model for allele frequencies. *Genet Epidemiol* 20: 17–33.