

VOJNOSANITETSKI PREGLED VOJNOMEDICINSKA AKADEMIJA Crnotravska 17, 11 000 **Beograd, Srbija** Tel/faks: +381 11 2669689 vsp@vma.mod.gov.rs

## **ACCEPTED MANUSCRIPT**

Accepted manuscripts are the articles in press that have been peer reviewed and accepted for publication by the Editorial Board of the *Vojnosanitetski Pregled*. They have not yet been copy edited and/or formatted in the publication house style, and the text could still be changed before final publication.

Although accepted manuscripts do not yet have all bibliographic details available, they can already be cited using the year of online publication and the DOI, as follows: article title, the author(s), publication (year), the DOI.

Please cite this article: ANTIOXIDANT STATUS AND CLINICOPATHOLOGICAL PARAMETERS IN PATIENTS WITH PARKINSON'S DISEASE

## ANTIOKSIDATIVNI STATUS I KLINIČKO-PATOLOŠKI PARAMETARI KOD PACIJENATA OBOLELIH OD PARKINSONOVE BOLESTI

Authors Jadranka Miletić Vukajlović, Snežana Pejić, Ana Todorović, Ana Valenta Šobot, Dunja Drakulić, Ivan Pavlović, Aleksandra Stefanović, Milica Prostran, Tihomir V. Ilić, Marina Stojanov, Vojnosanitetski pregled (2018); Online First September, 2018.

UDC:

DOI: https://doi.org/10.2298/VSP180718148M

When the final article is assigned to volumes/issues of the Journal, the Article in Press version will be removed and the final version appear in the associated published volumes/issues of the Journal. The date the article was made available online first will be carried over.

# ANTIOXIDANT STATUS AND CLINICOPATHOLOGICAL PARAMETERS IN PATIENTS WITH PARKINSON'S DISEASE

## ANTIOKSIDATIVNI STATUS I KLINIČKO-PATOLOŠKI PARAMETARI KOD PACIJENATA OBOLELIH OD PARKINSONOVE BOLESTI

Jadranka Miletić Vukajlović<sup>\*</sup>, Snežana Pejić<sup>†</sup>, Ana Todorović<sup>†</sup>, Ana Valenta Šobot<sup>\*</sup>, Dunja Drakulić<sup>†</sup>, Ivan Pavlović<sup>†</sup>, Aleksandra Stefanović<sup>†</sup>, Milica Prostran<sup>\*</sup>, Tihomir V. Ilić<sup>†</sup>, Marina Stojanov<sup>†</sup>

<sup>•</sup>Department of Physical Chemistry, VINCA Institute of Nuclear Sciences, University of Belgrade, Belgrade, Republic of Serbia

<sup>\*</sup>Department of Molecular Biology and Endocrinology, VINCA Institute of Nuclear Sciences, University of Belgrade, Belgrade, Republic of Serbia

Department of Medical Biochemistry, Faculty of Pharmacy, University of Belgrade, Belgrade, Republic of Serbia

<sup>s</sup>Department of Pharmacology, Clinical Pharmacology and Toxicology, School of Medicine – University of Belgrade, Belgrade, Republic of Serbia

<sup>1</sup> Faculty of Medical Military Academy, Clinic of Neurology, University of Defense, Belgrade, Republic of Serbia

Corresponding author: Jadranka Miletić Vukajlović Department of Physical chemistry VINČA Institute of Nuclear Sciences University of Belgrade P.O.Box 522 11001 Belgrade Republic of Serbia Phone: +381 11 3408 293 Fax: +381 11 8066 434 e-mail: jadranka@vin.bg.ac.rs

Antioxidant status in Parkinson's disease

## **Authors' Contribution**

JM performed experiments, acquisited data, and drafted the manuscript. DD and AT participated in the interpretation of obtained results and writing the manuscript whereas, SP contributed to analysis of data and drafting of the manuscript. AT, AVS and IP performed biochemical measurements. MP, TVI, and MS designed and supervised experimental setup as well as edited the manuscript. TVI also selected the patients for the study. All authors approved the final version of the manuscript.

## Abstract

**Backgroun / Aim.** Constant production of free radicals and antioxidants (AO) in the cell is a part of normal cellular function. Their imbalance might take a part in pathophysiology of many diseases, including Parkinson's disease (PD). Evaluation of the disease status, prooxidant-antioxidant balance (PAB) and antioxidants are being widely estimated. The aim of this study was to examine potential interaction between several AO variables (GSH, SOD, CAT and PAB) and clinicopathological features of patients with PD, particularly Hoehn and Yahr (H&Y) stage.

**Methods**. A multivariate analysis of variance (MANOVA) was conducted to test the hypothesis of the mean differences between clinicopathological characteristics (gender, age at examination, duration of the disease, and H&Y stage) and AO variables, compared with age/sex matched healthy controls. The study included 91 patients with idiopatic PD patients and 20 healthy controls. **Results.** The multivariate effect size was estimated at 0.269, p < 0.001, implying that 27.0% of the variance of the dependent variables was accounted for H&Y stage. Univariate tests showed that there were significant differences (p < 0.001) across the H&Y stage on all AO variables. The H&Y stage remained significant predictor after controlling for the second variable, the disease duration (p < 0.001,  $\eta^2 = 0.249$ ), and there were still significant differences across the H&Y stage on all variables, with effect size ( $\eta^2$ ) ranging from 0.132, p =0.011 (lnGSH) to the still high values of 0.535 (lnPAB), 0.627 (lnSOD) and 0.964 (lnCAT).

**Conclusion**. The results indicate that higher level of oxidative stress in blood of PD patients is possibly related to PD stage. Along with reduction of SOD and GSH level, CAT activity was elevated in comparison to healthy subjects. Furthermore, PAB was shifted toward oxidative stress.

#### Keywords: p

parkinson disease - superoxide dismutase - catalase – glutathione - prooxidantantioxidant balance

#### Apstrakt

Uvod/Cilj. Ćelijska homeostaza zasniva se na konstantnoj produkciji slobodnih radikala i antioksidanasa (AO). Svako narušavanje njihove ravnoteže može dovesti ili učestvovati u patofiziološkim promenama mnogih bolesti, uključujući i Parkinsonovu bolest (PB). Kako bi se pratio status bolesti, koristi se veliki broj parametara, uključujući i prooksidativniantioksidativni balans (PAB) i AO, koji ujedno predstavljaju i fokus ispitivanja ove studije. Stoga, cilj ove studije je ispitivanje potencijalne interakcije između AO varijabli (GSH, SOD, CAT i PAB) i kliničko-patoloških osobina PB pacijenta, najviše Hoehn i Yahr (H&Y) stepena bolesti. Metode. Multivarijantna analiza varijanse (MANOVA) je korišćena za analizu hipoteze međusobnih razlika između kliničko-patoloških karakteristika (pola, starosti, dužine trajanja bolesti i H&Y stepena bolesti) i AO varijabli sa zdravim kontrolama. Studija je uključivala ukupno 111 ispitanika (91 pacijenata kojima je dijagnostifikovana idiopatska PB i 20 zdravih kontrola). Rezultati. Multivarijantni efekat je procenjen na 0,269, p < 0,000, što implicira da se 27.0% varijanse zavisne varijable odnosi na H&Y stepen bolesti. Univarijantni test je pokazao da postoji statistički značajna razlika (p <0,001) kroz H&Y stepen bolesti svih AO varijabli. H&Y stepen bolesti je ostao značajan indikator i nakon uvođenja druge varijable, dužine trajanja bolesti (p <0,001,  $\eta^2 = 0,249$ ). Pokazano je da je ostala značajna razlika kroz H&Y stepen bolesti za sve varijable, tako da je jačina odnosa dve varijable od 0,132 (lnGSH) do i dalje visokih vrednosti 0,535 (lnPAB), 0,627 (lnSOD) i 0,964 (lnCAT). **Zaključak.** Rezultati ukazuju da je visoki nivo oksidativnog stresa u krvi pacijenata obolelih od PB verovatno povezan sa stepenom bolesti. Zajedno sa smanjenjem aktivnosti SOD i nivoa GSH, aktivnost CAT se povećava u poređenju sa zdravim kontrolama. Pored toga, PAB ukazuje na povećani oksidativni stres kod pacijenata obolelih od PB.

#### Ključne reči:

## parkinsonova bolest – superoksid dizmutaza – glutation – prooksidans-antioksidans balans

#### Introduction

Parkinson's disease (PD) is the second most common neurodegenerative disorder after Alzheimer's disease, histologically characterized by progressive loss of dopaminergic neurons in *substantia nigra pars compacta* (*SNpc*) and formation of Lewy bodies<sup>1</sup>. It is manifested by cardinal features such as bradykinesia, rigidity, tremor and postural instability, and good response to levodopa (L-dopa) is often used to support the diagnosis of PD<sup>2</sup>. Although the exact mechanism of PD pathogenesis still remains unclear, studies have indicated that oxidative stress (OS), inflammation, mitochondrial dysfunction and proteasomal inhibition are the major factors that accelerate dopaminergic neurodegeneration<sup>3</sup>.

Oxidative stress is defined as an imbalance between the production of reactive oxygen species (ROS) and antioxidant (AO) defense capacity. ROS are generally short-lived and highly reactive molecules derived from oxygen<sup>4</sup>, varying in their site of formation, physiological function, reactivity and biological half-life. They include free radicals, such as hydroxyl and superoxide radicals, and non-radicals including hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and singlet oxygen<sup>5</sup>. Maintenance of the physiological level of ROS is basically regulated by antioxidant enzymes (AOE) and small antioxidant molecules<sup>6</sup>.

Antioxidant enzymes include superoxide dismutases (SODs), catalase (CAT), glutathione peroxidases (GPxs), glutathione reductases (GRs) and glutathione-S-transferases (GSTs), while non-enzymatic antioxidants are represented by glutathione (GSH), ascorbic acid (vitamin C),  $\alpha$ -tocopherol (Vitamin E), flavonoids, etc.<sup>7</sup>. The main function of SOD is catalyzing the breakdown of highly reactive superoxide anion into oxygen and the less reactive H<sub>2</sub>O<sub>2</sub>, which is further decomposed to water and oxygen by CAT or GPx<sup>8</sup>. Disturbance of AOE activity is strongly implicated in a variety of age-related brain disorders<sup>9</sup>.

Glutathione is the major small AO molecule<sup>6</sup>, with the concentration of 1–3 mM in the brain cells<sup>10</sup>. It is highly abundant in the cytosol (1-11mM), nucleus (3-15 mM) and mitochondria (5-11 mM)<sup>11</sup>. In some studies, a much lower concentration of 2  $\mu$ M was found in blood plasma<sup>10</sup>. GSH can reduce superoxide radicals, hydroxyl radicals, and peroxynitrites, reacting alone or with other enzymes, such as GPx or GST<sup>12</sup>.

Other than individual molecules, one of the important parameters for oxidative stress evaluation is a prooxidant-antioxidant balance (PAB), which determines a state of dynamic balance between free radicals that are produced and those utilized (scavenged)<sup>13</sup>.

Similar to other diseases, a disturbed AO balance renders PD patients more vulnerable to OS. Thus, to further evaluate its degree, the present study investigated PAB and AO enzymes (SOD, CAT), as the first line of defense against ROS and GSH level in the blood of PD patients, compared to healthy subjects. Furthermore, the relation of AO parameters with clinicopathological features such as gender, age, duration of the disease, and Hoehn and Yahr (H&Y) staging was estimated.

#### Methods

#### **Participants**

The study comprised 91 patients with idiopathic PD, and 20 healthy controls, originated in the Republic of Serbia. All blood samples were collected at the Neurology Clinic, Clinical Center of Serbia, University of Belgrade. The study was performed in compliance with the ethical principles of the Declaration of Helsinki and all applicable national laws and regulations. The study protocol was approved by the Ethical Committee of Clinical Centre of Serbia, Belgrade, and written informed consent was obtained from each patient prior to study engagement. All patients had idiopathic PD diagnosed in accordance with UK brain bank criteria<sup>14</sup>. Inclusion criteria were disease duration (up to 25 years), age (30 – 75 years), H&Y stage (I – IV), receiving symptomatic PD therapy and a stable dose of L-dopa medications for longer than 3 months. Patients with current evidence of a recent diagnosis of malignancy marked autonomic disturbances, a renal insufficiency or failure, hepatitis, serious and or unstable gastrointestinal, hematologic or other medical disorders, as well as subjects using antipsychotics were excluded from the study. The clinicopathological features of patients included age, gender, disease duration and H&Y stage of disease (Table 1).

#### Blood sampling and biochemical measurements

Venous blood samples were collected from each patient using conventional techniques into Vacutainer (BD Diagnostics, Plymouth, UK) tubes with K<sub>2</sub>EDTA as an anticoagulant. For PAB measurement, one batch was centrifuged at 1500 g, for 10 min, at 4 °C, within 30 min of collection. Plasma was carefully separated and stored at -80 °C until further processing.

For enzyme activity measurements, the second batch of unfrozen blood was used. All blood samples were diluted with cold  $dH_20$  1:3 (v/v), vortexed and centrifuged for 1 min (10000 g, 15 min, 4°C). Supernatants were collected and kept at -80°C till the assay.

For GSH measurement the blood was prepared as recommended by the kit producer (BIOXYTECH<sup>®</sup> GSH-420<sup>™</sup>, OXIS International Inc., Foster City, CA, USA).

#### Assays

Total SOD activity was measured using Superoxide Dismutase Assay Kit (Cayman Chemical Company, Ann Arbor, MI, USA). The reaction between superoxide radicals ( $O_2$ ) and tetrazolium salt, generated by xanthine oxidase, results in the development of formazan dye, with max absorbance on 450nm. SOD inhibits this reaction by dismutation of  $O_2$  and one unit of SOD is defined as the amount of enzyme needed to exhibit 50 % dismutation of

superoxide radical. Measurements were performed in a microplate reader (Wallac 1420 Victor<sup>2</sup>, Perkin Elmer Inc., Waltham, MA, USA).

Total GSH concentration was determined by the BIOXYTECH<sup>®</sup> GSH-420<sup>TM</sup> Assay (OXIS International, Inc., Foster City, CA, USA). The measurement of total GSH concentration was performed in three colorimetric reaction steps. Tris (2-carboxyethyl) phosphine (TCEP) as a reducing agent, reduces all oxidized glutathione present in the sample. During the second step, chromogen (4-chloro-1-methyl-7-trifluoromethylquinolinium methylsulfate) reacts with thiols in the sample and forms thioethers. Addition of base (NaOH) raises reaction mixture pH over 13 and chromophoric thione is formed as a result of  $\beta$ -elimination specific to the GSH-thioether. GSH concentration is directly proportional to the absorbance at 420 nm.

Catalase activity measurement was performed according to the method by Beutler<sup>15</sup>. The reaction mixture was prepared from 50  $\mu$ l of a Tris-HCl buffer (1 M Tris-HCl, 5 mM EDTA, pH 8.0), 900  $\mu$ l of a substrate (10 mM H<sub>2</sub>O<sub>2</sub>), 30  $\mu$ l of dH<sub>2</sub>O, and 20  $\mu$ l of the sample. Decomposition of H<sub>2</sub>O<sub>2</sub> was monitored spectrophotometrically (UV Line 9400, SI Analytics GmbH, Mainz, Germany) at 230 nm, 3 min at 37°C. One unit of CAT activity is defined as the amount of enzyme which degrades 1  $\mu$ mol of H<sub>2</sub>O<sub>2</sub> per min under the assay conditions. The extinction coefficient for H<sub>2</sub>O<sub>2</sub> is 0.071 mM<sup>4</sup>cm<sup>4</sup>.

#### Prooxidant-antioxidant balance

Evaluation of PAB was performed as described previously<sup>16</sup>. Following the incubation for 2 min at room temperature in dark, 200  $\mu$ l of working solution (1 ml TMB cation solution with 10 ml TMB solution) was added to a 96-well microtiter plate and mixed with 10  $\mu$ l of plasma sample, standard or blank (dH<sub>2</sub>O). The mixture was incubated in a dark place for 12 min, at 37°C and the reaction was stopped by adding 100  $\mu$ l of 2 N HCl. The values of PAB in plasma samples were determined at 450 nm, with a reference wavelength of 620 or 570 nm, by comparing optical density (OD) of a sample to the standard curve. PAB values are expressed in arbitrary units (HK).

#### Statistical analysis

The statistical analyses were performed by the GraphPad Prism and SPSS 18.0 for Windows (SPSS Inc., Chicago, IL, USA). Data are expressed as mean  $\pm$  SD. General linear model (GLM) was used to test the differences between AO and clinicopathological variables, followed by Dunnett and Scheffe *post hoc* tests. Since examined variables had not passed the normality of the distribution (Shapiro-Wilks test), data were previously log-transformed. Pearson correlation analysis was performed to test the correlation between AO/clinicopathological variables. The p-value < 0.05 was considered statistically significant.

#### Results

The average age of healthy controls was  $57.5 \pm 8.5$  years, and for PD patients it was  $62.7 \pm 9.7$  years, with a predominance of males (65.9 %). The H&Y stage 1 was the least one represented (9.9%), (Table 1). The activity of AO enzymes (SOD, CAT), the GSH level and PAB are shown in Figure 1.

A multivariate analysis of variance (MANOVA) was conducted to test the hypothesis of the mean differences between the H&Y stage and AO variables.Prior to conducting the analysis, the Pearson correlation was performed between the dependent variables in order to test the correlation assumption (Table 2) and significant pattern of correlations was observed amongst all of the dependent variables. Since the Box's M value of 110.06 (p<0.001) indicated significant difference between the covariance matrices, the Pillais' Trace test was used. The MANOVA effect (Pillais' Trace = 1.07, F = 9.103, p < 0.001), showed significant differences among the H&Y stage groups on the linear combination of the dependent variables. The multivariate effect size was estimated at 0.269, implying that 27.0 % of the variance of the examined AO parameters was accounted for the H&Y stage.

The homogeneity of variance assumption was tested for the AO variables and two (lnGSH and lnPAB) of the four Levene's F tests were statistically significant (p < 0.05). Prior to conducting a series of follow-up ANOVAs, the Bonferroni procedure was used to protect against Type I error, adjusting the alpha level to p < 0.001. Univariate tests showed that there were significant differences (p < 0.001) across the H&Y stage on all AO variables, with effect size ( $\eta^2$ ) ranging from 0.365 (lnGSH) to the extremely high values of 0.744 (lnPAB), 0.861 (lnSOD) and 0.988 (lnCAT) (Table 3a).

Finally, the series of post-hoc analyses (Dunnett and Scheffe test) were performed to examine individual mean difference comparisons across all H&Y stage and all four AO variables. The results revealed that high effect size observed by univariate analysis is the consequence of the mean differences in AO values between H&Y stage and control values (Dunnett test, p < 0.001). Scheffe test did not reveal a significant mean difference in AO values between any of H&Y stage.

In the next step, to test whether H&Y stageremains significant after controlling for the next clinical variable, the disease duration was added as a covariate to the model. The MANCOVA analysis of the effect of the H&Y stage on all AO parameters was still significant (Pillais' Trace = 0.998, F = 7.560, p <0.000),  $\eta^2$  0.249. Univariate tests showed that there were still significant differences across the H&Y stage on all AO variables, with effect size ( $\eta^2$ ) ranging from 0.132, p = 0.011 (lnGSH) to the still high values of 0.535 (lnPAB), 0.627 (lnSOD) and 0.964 (lnCAT) (Table 3b).

There was no significant association between AO parameters and gender (Pillais' Trace = 0.033, F = 0.713, p = 0.585,  $\eta^2 = 0.033$ ) or age (Pillais' Trace = 0.70, F = 1.558, p = 0.193,  $\eta^2 = 0.070$ ).

#### Discussion

Oxidative stress has long been implicated in pathophysiological mechanisms underlying various neurodegenerative diseases, including PD. Investigation of different oxidant/AO parameters have yielded inconsistent results and it is still challenging to assess these parameters peripheral blood of patients with PD. The current study is focused on the association of specific AO variables (GSH, SOD, CAT, and PAB) and clinicopathological features of patients with PD, particularly H&Y stage.

Among all ROS-scavenging enzymes, SOD is often regarded as the first line of defense and there is sufficient evidence relating superoxide anion to human diseases, such as PD<sup>17</sup>. The results of our study have shown decreased SOD activities in PD patients

compared to healthy subjects, which is in accordance with the findings of some authors<sup>18-21</sup> while the others<sup>22-27</sup>reported increased SOD activity or no significant change at all<sup>26-27</sup>. It is known that AO enzymes are regulated through the AO system to cope with acute or mild OS; however, severe or prolonged OS may induce consumption and decrease of enzyme activity. The decrease of SOD observed in our study might involve inactivation of SOD by ROS or some posttranslational modifications<sup>28</sup>. This observation is comparable with the fact that reduced activity of blood SOD is detected in many chronic diseases such as obstructive pulmonary disease<sup>29</sup>, renal failure<sup>30</sup>, as well as in some neurological disorders<sup>31</sup>. Chronic OS has already been speculated to cause antioxidant consumption and thus a decline in antioxidant levels<sup>32</sup>. Another possible reason for decreased SOD level could be in mutations that not only provoke a decline in its activity but also induce self-aggregation of mutated SOD proteins - an initial cause of neuron malfunction leading to disease, as already shown in a cell culture model of amyotrophic lateral sclerosis<sup>33</sup>. The confirmation of such assumptions requires more extensive research in the field of molecular events related to this disease.

The term OS describes the condition where free radicals production exceeds a capacity of AO system. Studies indicated different findings of erythrocyte CAT activity in PD patients in which no significant changes<sup>27,34</sup> or deficit<sup>18,21</sup> of CAT were recorded in comparison with healthy subjects. PD patients involved in the present study had elevated CAT activity compared to healthy controls, and there were no differences between H&Y stages and disease duration. Similar results were obtained in the research of Younes-Mhenniet et al.<sup>22</sup>, who have not observed the correlation between the duration of illness and CAT activity.

Several studies have shown contrasting results; Sudha et al.<sup>35</sup> observed no significant changes of erythrocyte antioxidants in PD patients while Abraham and coworkers<sup>21</sup> reported decreased AO enzymes activity in PD patients compared to controls. Considering that CAT is crucial in removing  $H_2O_2$  at higher concentrations<sup>36</sup> (GPx are predominant at physiologically low levels of  $H_2O_2^{37}$ ), elevation of CAT activity in the blood of PD patients confirms the general conclusion of this study that PD patients are exposed to chronic oxidative stress<sup>38</sup>.

It is hypothesized that the adjustment of the AO system is based on shifts in AO activities rather than on the formation of new AO resources. Thus, for some aspects of the issue, it may be more useful to study whole groups of radical scavengers rather than focusing on individual molecule species<sup>39</sup>. PAB can be considered as a measure of an imbalance between oxidants (H<sub>2</sub>O<sub>2</sub>, tert-butylhydroperoxide, chloramine T and HClO) and antioxidants (vitamin C, trolox, GSH, uric acid, bilirubin, albumin, and ceruloplasmin)<sup>40</sup>. In our study, PAB has shifted forward the OS, indicating that PD patients had an elevated level of OS compared with healthy subjects, regardless of the H&Y stage.

The physiological roles played by the GSH include maintenance of thiol redox potential, clearing metabolic waste, and as a reservoir for amino acids<sup>41</sup>. Since GSH is involved in antioxidant defense, and regulation of cellular metabolic functions ranging from gene expression, DNA, and protein synthesis to signal transduction, cell proliferation and apoptosis<sup>42</sup>, its depletion might have a wide impact on many physiological and pathological processes. For instance, GSH deficiency has long been implicated in PD degeneration<sup>43</sup>. A recent report even suggests that whole blood GSH may have the utility as a biomarker in PD progression as it was statistically associated with PD status<sup>44</sup>. Accordingly, in our study, a blood concentration of GSH in PD patients was significantly decreased compared to healthy controls, and such tendency was more pronounced through H&Y stages. These findings are important as the changes in the level of GSH have consequences to numerous molecular processes as well as the progression of the disease. Furthermore, it should be emphasized that the exact cause of GSH reduction has not been fully clarified, however, it is known that the most common ways for reducing GSH involve its consumption by GPx, conjugation reaction with proteins<sup>45</sup> and 4-hydroxynonenal (4-HNE)<sup>46</sup> and translocation of GSH/GSSG across the plasma membrane<sup>47</sup>. In order to compensate for this decrease, the possible ways of therapeutic compensation of GSH treatments are investigated. They include intranasal<sup>48</sup>, intravenous, and liposomal<sup>49</sup> GSH augmentation, and some of them showed a promising effect in the treatment of PD disease<sup>50</sup>.

## Conclusions

Obtained results show that some of the examined AO parameters in blood of PD patients are possibly related to PD stage. We observed a correlation of H&Y stage with PAB and AO parameters. The reduction of GSH level was related with higher H&Y stage while PAB, SOD and CAT activity changed regardless of the H&Y score. **Disclosure statement** 

The authors declare that there is no conflict of interests regarding the publication of this paper.

#### Acknowledgments

All authors are grateful to Prof Dr.Marina Svetel for selecting the patients for the study. This study was supported by Ministry of Education, Science and Technological Development, Republic of Serbia, grants 175023, 173044 and 41014.

## REFERENCES

- 1. Jinsmaa Y, Florang VR, Rees JN, Mexas LM, Eckert LL, Allen EM, et al.Dopamine-derived biological reactive intermediates and protein modifications: Implications for Parkinson's disease. Chem Biol Interact 2011; 192(1-2): 118-21.
- 2. Kalia LV, Lng AE. Parkinson's disease. Lancet 2015; 386(9996): 896-912.
- 3. Blesa J, Trigo-Damas I, Quiroga-Varela A, Jackson-Lewis VR. Oxidative stress and Parkinson's disease. Front Neuroanat 2015; 9: 91.
- 4. *Bolisetty S, Jaimes EA*. Mitochondria and reactive oxygen species: physiology and pathophysiology. Int J Mol Sci 2013; 14(3): 6306-44.
- 5. *Dröge W*. Free radicals in the physiological control of cell function. Physiol Rev 2002; 82(1): 47-95.
- 6. *Gandhi S, Abramov AY*. Mechanism of oxidative stress in neurodegeneration. Oxid Med Cell Longev 2012; 2012: 428010.
- 7. Carocho M, Ferreira IC. A review on antioxidants, prooxidants and related controversy: natural and synthetic compounds, screening and analysis

methodologies and future perspectives. Food Chem Toxicol 2013; 51:15-25.

- 8. *Fridovich I.* Superoxide radical and superoxide dismutases. Annu Rev Biochem 1995; 64(1): 97–112.
- 9. *Dasuri K, Zhang L, Keller JN*. Oxidative stress, neurodegeneration, and the balance of protein degradation and protein synthesis. Free Radical Bio Med 2013; 62: 170-85.
- 10.*Schafer FQ, Buettner GR.* Redox environment of the cell as viewed through the redox state of the glutathione disulfide/glutathione couple. Free Radical Bio Med 2001; 30(11): 1191-212.
- 11.Valko M, Leibfritz D, Moncol J, Cronin MT, Mazur M, Telser J. Free radicals and antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol 2007; 39(1): 44-84.
- 12.*Masella R, Di Benedetto R, Varì R, Filesi C, Giovannini C*. Novel mechanisms of natural antioxidant compounds in biological systems: involvement of glutathione and glutathione-related enzymes. J Nutr Biochem 2005; 16(10): 577-86.
- 13.Sahebkar A, Mohammadi A, Atabati A, Rahiman S, Tavallaie S, Iranshahi M, et al. Curcuminoids Modulate Pro-Oxidant–Antioxidant Balance but not the Immune Response to Heat Shock Protein 27 and Oxidized LDL in Obese Individuals. Phytother Res 2013; 27(12): 1883–88.
- 14.*Hughes AJ, Daniel SE, Kilford L, Lees AJ.* Accuracy of clinical diagnosis of idiopathic Parkinson's disease: a clinico-pathological study of 100 cases. J Neurol Neurosurg Psychiatry 1992;55(3):181–4.
- 15.*Beutler E.* Catalase. In: Beutler E, editor. Red cell metabolism: A manual of biochemical methods. 3rd ed. Orlando: Grune and Stratton; 1984: p. 105-6.
- 16.Miletić J, Drakulić D, Pejić S, Petković M, Ilić TV, Miljković M, et al. Prooxidant–antioxidant balance, advanced oxidation protein products and lipid peroxidation in Serbian patients with Parkinson's disease. Int J Neurosci 2018; 128:(7) 600-7.
- 17. Hayyan M, Hashim MA, AlNashef IM. Superoxide ion: generation and chemical implications. Chem Rev 2016; 116(5): 3029-85.
- 18.de la Torre MR, Casado A, López-Fernández ME, Carrascosa D, Casado MC, Venarucci D, et al. Human aging brain disorders: role of antioxidant enzymes. Neurochem Res 1996; 21(8): 885–8.
- 19.Bostantjopoulou S, Kyriazis G, Katsarou Z, Kiosseoglou G, Kazis A, Mentenopoulos G. Superoxide dismutase activity in early and advanced Parkinson's disease. Funct Neurol 1997; 12(2): 63–8.
- 20.*Ihara Y, Chuda M, Kuroda S, Hayabara T*. Hydroxyl radical and superoxide dismutase in blood of patients with Parkinson's isease:relationship to clinical data. J Neurol Sci 1999; 170(2): 75–6.
- 21. Abraham S, Souandarajan CC, Vivekanandhan S, Behari M: Erythrocyte antioxidant enzymes in Parkinson's disease. Indian J Med Res 2005; 121(2): 111–15.
- 22. Younes-Mhenni S, Frih-Ayed M, Kerkeni A, Bost M, Chazot G. Peripheral blood markers of oxidative stress in Parkinson's disease. Eur Neurol 2007;58(2): 78-83.

- 23.Kalra J, Rajput AH, Mantha SV, Prasad K. Serum antioxidant enzyme activity in Parkinson's disease. Mol Cell Biochem 1992; 110(2):165–8.
- 24.*Kocaturk PA*, *Akbostanci MC*, *Tan F*, *Kavas GO*. Superoxide dismutase activity and zinc and copper concentrations in Parkinson's disease. Pathophysiology 2000; 7(1): 63–7.
- 25.Serra JA, Dominguez RO, De Lustig ES, Guareschi EM, Famulari AL et al.Parkinson's disease is associated with oxidative stress: comparison of peripheral antioxidant profiles in living Parkinson's, Alzheimer's and vascular dementia patients. J Neural Transm 2001; 108(10): 1135–48.
- 26.Barthwal MK, Srivastava N, Shukla R, Nag D, Seth PK, Srirnal RC, et al. Polymorphonuclear leukocyte nitrite content and antioxidant enzymes in Parkinson's disease patients. Acta Neurol Scand 1999; 100(5): 300–4.
- 27.*Sudha K, Rao A, Rao S, Rao A*. Free radical toxicity and antioxidants in Parkinson's disease. Neurol India 2003; 51(1): 60–2.
- 28.*Hu N, Ren J.* Reactive Oxygen Species Regulate Myocardial Mitochondria through Post-Translational Modification. ROS 2016; 2(4): 264-71.
- 29.*Ahmad A, Shameem M, Husain Q*. Altered oxidant-antioxidant levels in the disease prognosis of chronic obstructive pulmonary disease. Int J Tuberc Lung Dis 2013; 17(8): 1104-9.
- 30.*Aziz MA, Majeed GH, Diab KS, Al-Tamimi RJ.* The association of oxidantantioxidant status in patients with chronic renal failure. Ren Fail 2016; 38(1): 20-6.
- 31.*Liu Z, Zhou T, Ziegler AC, Dimitrion P, Zuo L*. Oxidative stress in neurodegenerative diseases: from molecular mechanisms to clinical applications. Oxid Med Cell Longev 2017; 2017: 2525967.
- 32.*Polidori MC, Stahl W, Eichler O, Niestroj I, Sies H*. Profiles of antioxidants in human plasma. Free Radic Bio Med 2001; 229-35.
- 33.*Durham HD, Roy J, Dong L, Figlewicz DA*. Aggregation of mutant Cu/Zn superoxide dismutase proteins in a culture model of ALS. J Neuropathol Exp Neurol 1997; 56(5): 523-30.
- 34.*Kilinç A, Yalçin AS, Yalçin D, Taga Y, Emerk K*. Increased erythrocyte susceptibility to lipid peroxidation in human Parkinson's disease.Neurosci Lett 1988: 87(3): 307–10.
- 35.*Sudha K, Rao A, Rao S, Rao A*. Free radical toxicity and antioxidants in Parkinson's disease. Neurol India 2003;51(1): 60-2.
- 36.*Makino N, Mochizuki Y, Bannai S, Sugita Y*. Kinetic studies on the removal of extracellular hydrogen peroxide by cultured fibroblasts. J Biol Chem 1994; 269(2): 1020-5.
- 37.*Flohé L, Loschen G, Günzler WA, Eichele E.* Glutathione peroxidase, V. The kinetic mechanism. Hoppe-Seyler' s Zeitschrift für physiologische Chemie 1972; 353(1): 987-1000.
- 38.*Todorović A, Pejić S, Stojiljković V, Gavrilović L, Popović N, Pavlović I, et al.* Antioxidative enzymes in irradiated rat brain—indicators of different regional radiosensitivity. Childs Nerv Syst 2015; 31(12): 2249-56.
- 39.*Saleh L, Plieth C.* Total low-molecular-weight antioxidants as a summary parameter, quantified in biological samples by a chemiluminescence inhibition assay. Nat Protoc 2010; 5(10):1627-34.

- 40.*Alamdari DH, Paletas K, Pegiou T, Sarigianni M, Befani C, Koliakos G.* A novel assay for the evaluation of the prooxidant-antioxidant balance, before and after antioxidant vitamin administration in type II diabetes patients. Clin Biochem 2007; 40(3-4): 248–54.
- 41.*Zeevalk GD, Razmpour R, Bernard LP*. Glutathione and Parkinson's disease: is this the elephant in the room? Biomed Pharmacother 2008; 62(4): 236-49.
- 42.*Mischley LK, Standish LJ, Weiss NS, Padowski JM, Kavanagh TJ, White CC, et al.* Glutathione as a biomarker in Parkinson's disease: Associations with aging and disease severity. Oxid Med Cell Longev 2016; 2016:9409363.
- 43.Meister A, Anderson ME. Glutathione. Annu Rev Biochem 1983; 52(1): 711-60.
- 44.*Aquilano K, Baldelli S, Ciriolo MR*. Glutathione: new roles in redox signaling for an old antioxidant. Front Pharmacol 2014; 5: 196.
- 45.Lu SC. Regulation of glutathione synthesis. Mol Aspects Med 2009; 30(1-2): 42-59.
- 46.*Malone PE, Hernandez MR*. 4-Hydroxynonenal, a Product of Oxidative Stress, Leads to an Antioxidant Response in Optic Nerve Head Astrocytes. Exp Eye Res 200; 84(3): 444-54.
- 47.*Ballatori N, Krance SM, Marchan R, Hammond CL*. Plasma membrane glutathione transporters and their roles in cell physiology and pathophysiology. Mol Aspects Med 2009; 30(1-2): 13-28.
- 48.*Mischley LK, Lau RC, Shankland EG, Wilbur TK, Padowski JM*. Phase IIb Study of Intranasal Glutathione in Parkinson's Disease. J Parkinsons Dis 2017; 7(2): 289-99.
- 49.*Otto M, Magerus T, Langland J*. The Use of Intravenous Glutathione for Symptom Management of Parkinson's Disease: A Case Report. Altern Ther Health Med 2017; 23(7): 88-92.
- 50.Sechi G, Deledda MG, Bua G, Satta WM, Deiana GA, Pes GM, et al. Reduced intravenous glutathione in the treatment of early Parkinson's disease. Prog Neuropsychopharmacol Biol Psychiatry 1996; 20(7): 1159-70.

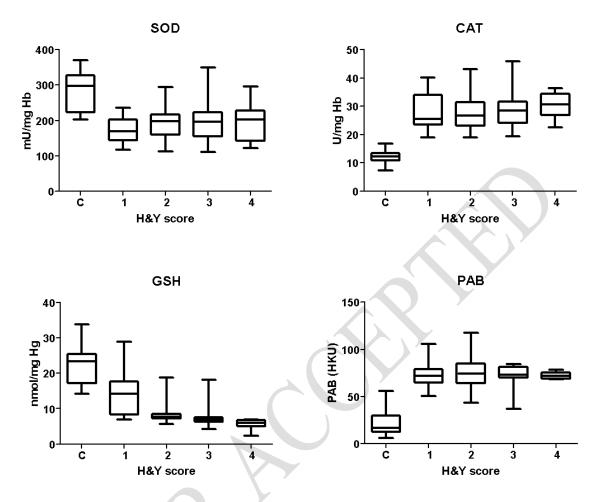


Fig. 1 – Superoxide dismutase (SOD) and catalase (CAT) activity, glutathione concentration (GSH), and prooxidant-antioxidant balance (PAB) in blood of healthy controls (C) and different H&Y score of patients with Parkinson disease. Boxes represent values between 25<sup>th</sup> and 75<sup>th</sup> percentiles. Medians are given inside the boxes. Whiskers extend between min and max values.

Domographic and clinical data of	Table 1
Demographic and clinical data of Characteristic	N (%)
Gender	
Male	60 (65.9)
Female	31 (34.1)
Age at examination (years)	62.7±9.7#
<59	28 (30.8)
59-70	44 (48.3)
>70	19 (20.9)
Age at disease onset (years)	53.8±9.1#
Disease duration (years)	8.8±6.2#
<3	18 (19.8)
3-8	35 (38.5)
>8	38 (41.8)
H & Y stage	
1	9 (9.9)
2	31 (34.1)
3	27 (29.7)
4	24 (26.4)

N -number of patients; (%)-fractions in relation to N; #-Mean values ± SD

Table 2.

## Pearson correlation between AO parameters

## Correlations

	nGSH	nSOD	InCAT	nPAB
nGSH Pearson Correlation	1	·0.498**	).581**	).595**
Sig. (2-tailed)		0.000	0.000	0.000
N	111	111	111	111
InSOD Pearson Correlation		1	-0.922**	-0.793**
Sig. (2-tailed)			0.000	0.000
N		111	111	111
<b>InCAT</b> Pearson Correlation			1	).864**
Sig. (2-tailed)				0.000
N			111	111

\*\*. Correlation is significant at the 0.01 level (2-tailed)

stage and	d AO	paramet	ers after c	controlling	for diseas	e duratio
		lnGSH	lnSOD	lnCAT	lnPAB	
a						
H&Y	F	3.462	38.268	607.374	26.187	
	р	0.011	p<0.001	p<0.001	p<0.001	
	eta <sup>2</sup>	13.2%	62.7%	96.4%	53.5%	
b						
Disease	F	0.042	3.523	0.650	0.790	
duration	р	0.837	0.064	0.422	0.377	
	eta <sup>2</sup>	0.000	3.7%	0.7%	0.9%	

GLM analysis of the associations between a) H&Y stage and AO parameters; b) H&Y
stage and AO parameters after controlling for disease duration

GSH – glutathione, SOD – superoxide dismutase, CAT – catalase, PAB – prooxidant/antioxidant balance, eta – quantified variance components, p – value indicates statistical significance

Received on July 18, 2018. Revised on August 9, 2018. Accepted on September 11, 2018. Online First September, 2018.

Table 3.