- 1 A Proof-of-Concept Clinical Trial Using Mesenchymal Stem Cells for the
- 2 Treatment of Corneal Epithelial Stem Cell Deficiency
- 3 Margarita Calonge, MD, PhD, 1,2 Inmaculada Pérez, PhD, 1 Sara Galindo, PhD, 1,2
- 4 Teresa Nieto-Miguel, PhD,<sup>2,1</sup> Marina López-Paniagua, PhD,<sup>1,2</sup> Itziar Fernández,
- 5 PhD,<sup>2,1</sup> Mercedes Alberca, PhD,<sup>3</sup> Javier García-Sancho, MD, PhD,<sup>3</sup> Ana Sánchez,
- 6 MD, PhD<sup>3</sup> José M. Herreras, MD, PhD, 1,2
- <sup>8</sup> IOBA (Institute of Applied Ophthalmobiology), University of Valladolid, Paseo de
- 9 Belén 17, E-47011, Valladolid, Spain
- <sup>2</sup>CIBER-BBN (Biomedical Research Networking Centre in Bioengineering,
- Biomaterials and Nanomedicine), Carlos III National Institute of Health, Spain
- <sup>3</sup>IBGM (Institute of Molecular Biology and Genetics), University of Valladolid and
- National Research Council (CSIC), Calle Sanz y Fores 3, E-47003, Valladolid, Spain
- 15 Author Contributions
- Margarita Calonge (calonge@ioba.med.uva.es): Conception and design, collection
- and assembly of data, data analysis and interpretation, financial, manuscript writing,
- 18 final approval of the manuscript.
- 19 Inmaculada Pérez (maku@ioba.med.uva.es): Collection and assembly of data, data
- 20 analysis and interpretation.
- 21 Sara Galindo (sgalindor@ioba.med.uva.es): Data analysis and interpretation,
- 22 manuscript writing.

- Teresa Nieto-Miguel (tnietom@ioba.med.uva.es): Collection and assembly of data,
- 24 data analysis and interpretation, manuscript writing.
- Marina López-Paniagua (marina@ioba.med.uva.es): Collection and assembly of
- data, data analysis and interpretation, manuscript writing.
- 27 Itziar Fernández (itziar.fernandez@ioba.med.uva.es): Data statistical analysis and
- 28 interpretation, manuscript writing.
- Mercedes Alberca (kikaalberca@hotmail.com): Data analysis and interpretation.
- Javier García-Sancho (jgsancho@ibgm.uva.es): Conception and design, manuscript
- 31 writing.

- Ana Sánchez (asanchez@ibgm.uva.es): Conception and design, manuscript writing.
- José M Herreras (herreras@ioba.med.uva.es): Conception and design, collection
- and assembly of data, data analysis, final approval of the manuscript.
- 36 Corresponding author:
- 37 Margarita Calonge, MD, PhD (corresponding author)
- 38 IOBA, University of Valladolid
- 39 Campus Miguel Delibes, Paseo Belén, 17
- 40 E-47011 Valladolid, Spain
- 41 E-mail: calonge@ioba.med.uva.es
- 42 Phone: +34 983 184750 Fax: +34 983 184762

•	1
7	<

Running head: Mesenchymal stem cells for co	orneal failure
---	----------------

# 46 List of abbreviation

- 47 AEMPS: Spanish Agency of Medicines and Sanitary Products
- 48 CLET: cultivated limbal epithelial transplantation
- 49 hAM: human amniotic membrane
- 50 LSCD: limbal stem cell deficiency
- 51 MSC: mesenchymal stem cells
- 52 MSCT: mesenchymal stem cell transplantation

53

# **ABSTRACT**

Ocular stem cell transplantation derived from either autologous or allogeneic donor
corneoscleral junction is a functional cell therapy to manage extensive and/or severe
limbal stem cell deficiencies that lead to corneal epithelial failure. Mesenchymal stem
cells have been properly tested in animal models of this ophthalmic pathology, but
never in human eyes despite their potential advantages. We conducted a 6- to 12-
month proof-of-concept, randomized, double-masked pilot trial to test whether
allogeneic bone marrow-derived mesenchymal stem cell transplantation (MSCT,
n=17) was as safe and as equally efficient as allogeneic cultivated limbal epithelial
transplantation (CLET, n=11) to improve corneal epithelial damage due to limbal
stem cell deficiency. Primary endpoints demanded combination of symptoms, signs,
and the objective improvement of the epithelial phenotype in central cornea by in-vivo
confocal microscopy. This proof-of-concept trial showed that MSCT was as safe and
efficacious as CLET. Global success at 6-12 months was 72.7%-77.8% for CLET
cases and 76.5%-85.7% for MSCT cases (not significant differences). Central
corneal epithelial phenotype improved in 71.4% and 66.7% of MSCT and CLET
cases, respectively at 12 months (p=1.000). There were no adverse events related to
cell products. This trial suggests first evidence that MSCT facilitated improvement of
a diseased corneal epithelium due to lack of its stem cells as efficiently as CLET.
Consequently, not only CLET but also MSCT deserves more preclinical
investigational resources before the favorable results of this proof-of-concept trial
could be transformed into the larger numbers of the multicenter trials that would
provide stronger evidence. (ClinicalTrials.gov number, NCT01562002.)

- 79 **Key words:** clinical trial, corneal blindness, corneal epithelial stem cells, human
- proof-of-concept, *in vivo* confocal microscopy, limbal stem cell deficiency,
- mesenchymal stem cell, stem cell transplantation

# INTRODUCTION

84

85

86

87

88

89

90

91

92

93

94

95

96

97

98

99

100

101

102

103

104

105

106

107

108

Corneal epithelial failure due to extensive or severe limbal stem cell deficiency (LSCD) is an end-stage pathology resulting from multiple diseases that destroy the corneal epithelium stem cell niche, located at the sclerocorneal limbus. LSCD results in recurrent corneal epithelial ulceration, neovascularization, and opacification because of the inability of the limbal niche to renew the corneal epithelium.<sup>1–3</sup> Corneal transplantation is not a viable primary solution as the donor tissue cannot replace the damaged corneal epithelial stem cells.<sup>4</sup> The first attempts to replace native limbal epithelial cells were to transplant whole limbal tissue from donor eyes.5 More recently, single limbal epithelial transplantation places small pieces of limbal tissue (not isolated cells) from the healthy fellow eye.<sup>6</sup> In 1997, stem cell-based therapies based on cultivated limbal epithelial cells commenced a significant breakthrough in regenerative medicine<sup>7</sup> and are currently an established therapy, both from autologous and allogeneic sources, the latter used when there is no possibility of a healthy donor contralateral eye.<sup>2,4,8–12</sup> Human amniotic membrane (hAM) transplantation, useful in sectoral and mild LSCD and an excellent cell carrier for stem cell growth and transplantation, has not been shown to help LSCD cases that are total and/or severe. 13,14 In this study, we explored, for the first time in human eyes in which medical therapy had failed, the potential capacity of mesenchymal stem cells (MSC) to treat corneal epithelial pathology due to LSCD. 15-17 MSC could have potential advantages over limbal epithelial stem cells for this purpose because they can be easily obtained from many tissue types without dependence of deceased donors. 18 Additionally, they can be cultured in vitro to clinical scales in a short period of time, overcoming the dependence on and the limitations of limbal epithelial cells, which are difficult to obtain, isolate, and culture and have limited availability. 19,20

Moreover, cryopreserved MSC can be transplanted without loss of potency,<sup>21</sup> whereas cryopreserved limbal epithelial stem cells have not been transplanted in humans yet.<sup>22,23</sup> Finally, allogeneic MSC can be transplanted without the need of host immunosuppression,<sup>20,24,25</sup> while allogeneic transplantation of limbal epithelial stem cells requires one year of systemic immunosuppression to avoid immune rejection.<sup>9,11</sup>

We report herein a proof-of-concept clinical trial aimed to evaluate the initial safety and clinical efficacy of MSC *versus* the established therapy with limbal epithelial cells for corneal epithelial pathology due to LSCD. An initial clinical success would warrant the economic expenditure necessary to carry out a more thorough investigation of the mechanism of action of not only limbal stem cells but also MSC before proceeding with larger clinical trials.

# **MATERIAL AND METHODS**

# Study design and patients

This was a Phase I-II randomized, controlled, double-masked, unicenter clinical trial based on the hypothesis that allogeneic bone marrow-derived mesenchymal stem cell transplantation (MSCT) is as safe and effective as allogeneic cultivated limbal epithelial transplantation (CLET) to treat patients with total and/or severe LSCD.

It was designed as a proof-of-concept clinical trial to include only the minimum required number of transplants necessary to prove the hypothesis.

The protocol (EudraCT 2010-023535-42) was approved by the local Ethics

Committee of our institution and the Spanish Agency of Medicines and Sanitary

Products (AEMPS, www.aemps.gob.es). The Clinical Trials.gov Identifier is 132 NCT01562002. 133 All procedures were conducted in accordance with the principles of the Declaration of 134 Helsinki, good manufacturing and clinical practice guidelines, and the European 135 Union Tissues and Cells Directive. All patients gave written informed consent. The 136 trial was sponsored by our institution, Advanced Therapies Unit, University of 137 Valladolid, Spain, under the guidance of the Advanced Therapies National Program 138 (Ministry of Heath, Government of Spain). 139 We enrolled adult patients with bilateral and severe disease, as it is ethically 140 indicated in an exploratory proof-of-concept clinical trial. The following 141 inclusion/exclusion criteria were met: (1) diagnosis of target disease characterized by 142 (a) corneal epithelial failure because of LSCD graded as total and/or severe, 143 meaning that at least three quadrants of the limbus were damaged (as visualized by 144 slit-lamp) and/or that the central cornea was involved<sup>9</sup>; (b) invariably accompanied by 145 146 blindness or low vision due to opacified central cornea; and (c) in which all available medical therapies (i.e., topical medications and hAM transplantations mainly in acute 147 phases of chemical injuries) had failed (Tables 1 and 2 for detailed previous 148 treatment in each patient); (2) no ocular surgeries in the previous 6 months other 149 than another cell transplant within this trial; (3) the affected eye had to have 150 undergone medical therapies to quiet and reverse as much as possible any treatable 151 limbal dysfunction; and (4) no contraindications for immunosuppressive therapies. 152 As the final outcome of cell transplantation strongly depends on the etiology of limbal 153 154 damage, the following three etiological categories were considered<sup>11</sup> at the initial

visit: Chemical injuries; immune-based inflammatory diseases (e.g. Stevens-Johnson

syndrome, mucous membrane pemphygoid, atopic keratoconjunctivitis); and other less inflammatory conditions (e.g., sequelae from multiple surgeries, chronic sequelae from infectious keratitis, congenital aniridia). The allocation to the two treatment groups (see below) was balanced according to these etiologies.

After screening for inclusion/exclusion criteria and balancing the allocation of the

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

156

157

158

159

# Randomization and masking

three prognostic etiologic categories, all scheduled transplants were randomly allocated in a 1:1 ratio to CLET or MSCT. Randomization was balanced by the use of permuted blocks of varying block size with a maximum size of 6. The randomization schedule was computer-generated (R Statistical Software). When both eyes of a patient were to receive transplants, the use of CLET or MSCT was randomly assigned so that both eyes did not necessarily receive the same type of transplant. Some eyes, due to the failure of the transplant, received more than one transplantation. For each repeated surgery, the assignment of CLET or MSCT was random. Thus the new transplants, CLET or MSCT, were not necessarily the same as the preceding one that failed. The Cell Processing Unit staff was aware of group assignment to prepare either cell product. The only difference in the final package that arrived at the Medical Institution the day of surgery was the type of cell cultivated, which was impossible to discern by the naked human eye. Everything else, including the packaging, was identical and followed the good manufacturing procedures. The products were identified by the randomization number, and only the Statistical Unit and the Cell Processing Unit

knew the identity of each product. All attending sanitary personnel and the patients themselves were completely masked as to the type of cells being transplanted.

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

179

180

### **Procedures**

Cells destined for CLET or MSCT (investigational products 09-137 and 10-134, respectively) were cultured on top of hAM at the University of Valladolid Cell Processing Unit, operating under good manufacturing practices and licensed and accredited by the AEMPS. Donor hAM and cadaveric limbal rings (≤60 years of age) came from a registered and accredited tissue bank (Blood-Tissue Community Center, Oviedo, Spain). Bone marrow from iliac crest was collected from allogeneic donors ≤60 years of age who gave written consent and were under other approved trials.<sup>26–28</sup> De-epithelialized hAM (size 2.5x2.5 cm) were prepared using our published standard protocol<sup>11</sup> and served as the substratum for both cell types. For CLET, two 2x2 mm pieces of allogeneic limbal rings were processed and cultured, as described. 11 For MSCT, allogeneic MSC were obtained and characterized as reported, <sup>26–28</sup> analyzing CD90, CD73, CD166, and CD105 as positive markers and CD14, CD34, CD45, and HAD-DR as negative markers in accordance with the International Society for Cellular Therapy (ISCT) position statement.<sup>29</sup> The cell products were harvested when the cultured cells were ~90% confluent (~250,000 cells). For the limbal epithelial cells, this took 3-4 weeks, and for the MSC it took 3-5 days. The quality criteria for the cell products were (1) sterility, (2) hAM integrity, (3) adherence of the hAM to the plate, (4) 80-90% of cell confluence (monolayer) observed under inverted phase contrast microscopy, and (5) cell morphology (polygonal shape for CLET and spindle shape for MSCT) observed under inverted phase contrast microscopy. After

determining that the cultures were negative for aerobes, anaerobes, fungi, and mycobacteria, they were delivered within 4 h of surgery. Some of the cultures that were assigned to patients who cancelled their surgeries (see below) were allowed to finish growing and later processed for immunostaining to test for limbal, mesenchymal, and differentiated corneal epithelial cell markers, as previously published<sup>30,31</sup> (see Supplementary Appendix for more details). To prevent any possible immune allograft rejection, patients receiving CLET underwent a mild immunosuppressive therapy. 11 While patients receiving MSCT would not normally need such therapy due to the absence of immune rejection by allogenic MSC, 15-<sup>17,24,25</sup> oral immunosuppression was instituted to eliminate immune suppression as a variable and to maintain the double masked nature of the study; otherwise, the trial could not have been masked. Thus, all patients were started at the initial visit on 1.5-2.0 g/day of mycophenolate mofetil; 3-5 mg/kg/day of cyclosporine A, or 1-2 mg/kg/day of azathioprine were also permitted if, for any reason, mycophenolate mofetil was not available or the patient was already using one of the other two immunomodulating agents. This treatment was maintained for 12 months after transplantation and discontinued in the next 3 months. We closely monitored potential side effects clinically and by blood/urine work-ups every 1-2 months. No other systemic medications were added. Surgery took place 3-4 weeks after the initial visit, and all were performed using the identical technique by the same experienced surgeon. 11 Briefly, after preparing the recipient corneal-limbal bed (i.e. scraping off the corneal-limbal pannus), hAM with cells for either CLET or MSCT were placed with the membrane facing up and the cells facing down, to facilitate their fast as possible access to the damaged corneal

and limbal bed. In this way, the cells were in direct physical and functional contact

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

with the tissues to be repaired and protected from the external environment by the hAM. The transplant was sutured to the bared sclera and covered with a bandage contact lens for 4 weeks.

Twenty-four hours after surgery, each patient was evaluated and topical treatment with the fixed combination of 1% prednisolone acetate and 0.3% tobramycin (Tobradex®, Alcon Laboratories, FT. Worth, TX, USA) was prescribed 4 times per day until the hAM dissolved. The stitches and the contact lens were also removed between 4 and 6 weeks. Then, 1 mg/ml dexamethasone (Maxidex®, Alcon Laboratories) was instilled 4 times a day and slowly tapered in the next 3 months. Anti-glaucoma medications were the only other topical medication allowed (other than lubricants) and only in those patients who were previously using them, as they had already been diagnosed with glaucoma. The patients were evaluated 24 h, 1 week, and 4 weeks after surgery. Evaluations were then performed every month for the first 6 months, and every two months until the first year. All personnel related to patient care, and the patients themselves, were masked as to the type of cells transplanted.

### Outcomes

Evaluation endpoints were collected at the initial baseline visit and at 6 months and 12 months. Three self-administered questionnaires evaluated symptoms and quality of life: the Single Item Dry Eye Questionnaire (SIDEQ), the Ocular Surface Disease Index (OSDI), and the National Eye Institute 25-item Visual Function Questionnaire (NEI-VFQ25).<sup>32,33</sup> The following clinical signs were evaluated by slit-lamp biomicroscopy: conjunctival redness, central corneal epithelial opacity, corneal

epithelial integrity manifesting as superficial keratitis and persistent epithelial defects, and corneal superficial neovascularization (area/length) (Table 3).

recovery (Table 3).

Visual acuity was determined as mandatory in clinical trials, although improved acuity is never the aim of these kind of trials. Procedures such as stem cell transplantation are intended to promote recovery of the corneal epithelium. Even with successful repair of the corneal epithelium, visual deficiency may continue due to deeper corneal damage (e.g., full thickness corneal destruction in severe chemical burns) or involvement of other parts of the eye (e.g., pre-existing cataract or various anomalies such as glaucoma, retinal pathology, etc.). In fact, we determined the presence of these factors before surgery to inform patients of what to expect in terms of visual

We used *in vivo* confocal microscopy (IVCM) (Heidelberg Retinal Tomograph HRT-3 and Rostock Cornea Module, Heidelberg Engineering GmbH, Heidelberg, Germany) to image the basal epithelium phenotype in the central cornea, as described by us<sup>11,34</sup> and others.<sup>35–37</sup> The confocal images are used to determine the presence of the normal homogenous corneal epithelial cell phenotype as well as conjunctival-like or mixed epithelial cell phenotypes typically present in the damaged central cornea. This provided an objective assessment of the presence of LSCD in central cornea and of the efficacy of restoration therapies (Table 3). See Supplementary Appendix for details on all end-points.

The primary outcomes were as follows: (1) improvement in any of the three questionnaires; (2) improvement by at least one step in at least two of the three following clinical parameters: conjunctival redness, central corneal epithelial opacity, or superficial punctate keratitis; (3) complete absence of persistent epithelial defects;

and (4) presence of a more corneal epithelial-like phenotype in the central cornea. The change in epithelial phenotype could be either a change from a conjunctival-like epithelium to either a corneal-like or a mixed epithelial phenotype or from a mixed epithelial phenotype to a corneal-like epithelium phenotype. Secondary outcomes included (1) amelioration of at least a one-step in superficial corneal peripheral neovessels (area/length) and (2) vision improvement of two lines or more in those cases that had the potential of vision gain with this cell transplantation alone and no additional surgeries (Table 3, column: Visual Prognosis and Potential for Visual Recovery, Grade 1 patients).

The outcome was considered successful when either a complete or a partial success was accomplished. A complete success meant that all four primary outcomes were achieved, and a partial success meant that at least two of the four primary outcomes or one primary and one secondary outcome were achieved. Failure meant that only one or none of the primary outcomes was met.<sup>11</sup>

# Statistical analysis

Statistical analyses were performed by a PhD licensed statistician, who estimated that a sample of 10 transplants per group would give 80% statistical power to detect the non-inferiority of the experimental group. The calculations assumed a success rate of 80% in each group and a non-inferiority margin of 25% at an alpha level of 0.05. Consequently it was considered that a minimum of 10 transplants per group was sufficient for the exploratory nature of this proof-of-concept trial.

Quantitative characteristics were expressed as means ± standard deviations (SD), and qualitative variables were described as percentages. The median and interquartile range (IQR) were used to summarize distributions of ordinal variables. Normality assumptions were checked by the Shapiro-Wilk test. Differences between the means of two independent groups were tested by Student's t-test or the nonparametric alternative, Mann-Whitney U test, if the normality assumption was not valid. Levene's test was used to check homogeneity of variance. Relationships between two qualitative variables were evaluated by chi-square test or Fisher's exact test with small expected cell counts. Analysis of variance (ANOVA) with repeated measures on one factor was utilized to test for mean differences over time. The sphericity assumption was checked by Mauchly's test and, in case of violation of sphericity, the Greenhouse-Geisser correction was used. If there were differences in the repeated ANOVA measures, Bonferroni post hoc testing was used to determine where differences lay in a pairwise analysis. When data had marked deviations from the normality assumption Friedman's test was used, followed by the post hoc analysis based on Wilcoxon-Nemenyi-McDonald-Thompson test. Kaplan-Meier survival analysis was applied to estimate transplant survival. The log-rank test was used to compare the survival curves of each transplant type. The R Statistical Software version 3.1.3 was used (Foundation for Statistical Computing, Vienna, Austria).

318

319

320

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

313

314

315

316

317

#### **RESULTS**

### Clinical trial sample

We initially recruited 27 Caucasian patients (36 eyes, 42 potential transplants) (Fig. 1). Five patients (10 eyes) cancelled surgery due to different personal/logistical reasons. Therefore, the final number of transplant surgeries was 37 (26 eyes, 22 patients). Among them, 16 cases were randomized to CLET and 21 to MSCT. Nine cases (24.3% of the total 37; 95% confidence interval [CI], 12.4 to 41.6), 5 CLET and 4 MSCT, lost their transplants within the first week due to loss of the bandage contact lens. Based on our preclinical data (unpublished), we considered that stem cells may have not completely reached their tissue target in less than 7 days, thus these transplants were excluded. We consequently included in this trial 28 transplant surgeries from 23 eyes (20 patients), 11 cases were randomized to CLET and 17 to MSCT that were fully 

patients), 11 cases were randomized to CLET and 17 to MSCT that were fully assessable at the minimum established period of 6 months (Fig 1). Of those, 23 reached 12 month follow-up and only 5 transplants did not: one (MSCT-2) was withdrawn due to a violent relapse of concomitant atopic dermatitis and its ocular component, atopic keratoconjunctivitis, that ruined his transplant and also worsened considerably his fellow non-transplanted eye. The other 4 cases were successfully re-grafted for the benefit of the patient (2 failed MSCTs; 1 failed CLET, 1 partially successful CLET). See Tables 1 and 2 for more details.

Table 4 shows the summary characteristics of the 28 fully assessable cell transplants; the detailed characteristics at baseline, 6 months, and 12 month of each case are shown in Tables 1 and 2.

Patients with assessable transplants had a mean age of 49.3±10.8 years (range, 28-77 years). Females comprised 42.9% (95% CI, 25 to 62.6) of the transplant recipients and males 57.1% (95% CI, 37.4 to 75) (p=0.253). The assignment of

(Table 4). Time from LSCD to cell transplant was not significantly different between 346 CLET and MSCT (Table 4). 347 The etiology groups leading to the target disease and the severity and extension of 348 the disease were equally distributed between CLET and MSCT patients (Table 4). 349 Consequently the different nature and severity of the background disease had no 350 influence in the results. As this was an initial pilot trial, we decided not to restrict 351 access regardless of the etiology. 352 Although we intended to transplant only one eye in this proof-of-concept trial, 4 cases 353 had transplants in both eyes to attend patient demands, because both eyes were 354 highly symptomatic and had not responded to medical therapy or to previous hAM 355 transplantations (see inclusion criteria). 356 There were no intra- or post-operative complications. No episodes of immune 357 rejection were recorded. Oral immunosuppression was used in all 28 transplant 358 cases. Mycophenolate mofetil was prescribed in 12 cases (3 CLET, 9 MSCT), 359 cyclosporine A in 6 cases (2 CLET, 4 MSCT); and azathioprine in 7 cases (4 CLET, 3 360 361 MSCT). Three patients had two immunosuppressants concomitantly due to their systemic disease. The drugs were well tolerated in all cases, and no discontinuations 362 363 were necessary. Mycophenolate mofetil had to be lowered from 2 g/day to 1.5 or 1 364 g/day in 3 cases due to asthenia. Cyclosporine A was also lowered from 5 to 3 mg/kg/day in two cases due to mild elevation in blood pressure. 365 There were 3 serious adverse events and 21 non-serious adverse events, including 366 10 that were mild and 11 that were moderate to severe (Table 5). All were unrelated 367 to the type of cell transplantation. Most were due to activation or recurrence of 368

patients to the CLET or MSCT groups was statistically independent of age or gender

baseline disease, and some were attributable to the concomitant immunosuppression.

371

372

373

374

375

376

377

378

379

380

381

382

383

384

385

386

387

388

389

390

391

369

370

# Final outcome and survival analysis

MSCT in the same patient and 1 CLET failed).

The overall success for all cell transplants after 6 months was 75% (21 of 28 cases: 13 complete successes, 8 partial successes). After 12 months, the success rate was 82.6% (19 of 23: 15 complete successes, 4 partial successes). Five transplants were evaluated until month 6 (4 regrafts, one withdrawal), accounting for the different percentages. Except for CLET-2 and MSCT-15 that went from partial successes at 6 months to complete successes at 12 months, the final fate of all transplants was already established at 6 months. The percentage of successful cases at 6 and 12 months was slightly higher for MSCT (76.5% and 85.7% respectively) than for CLET (72.7% and 77.8% respectively), but the differences were not statistically significant (Table 4). Thus, the final results were statistically independent of the type of cell transplant. All failures were in eyes with either chemical injuries (4 of 7; 57.1%) or immunemediated inflammatory diseases (3 of 7; 42.9%). None of the five non-inflammatory disease cases failed. Among chemical burns transplants, 75% were successful. Two of the failed cases were CLET, and two were MSCT. For transplants performed in inflammatory immune-based diseases, 57.1% were successful. One CLET transplant failed and two MSCT transplants failed in the same patient. Similarly, 25% of all cell transplants were immune-mediated diseases and of those, 57.1% were successful (2

For primary and secondary evaluation endpoints, there were no significant differences between the two groups except for central corneal opacity, which was slightly more improved in MSCT cases (Table 4). Within each of the two groups, one of the symptom questionnaires showed improvement. Conjunctival redness and central corneal opacity also decreased and superficial corneal integrity, both keratitis and ulceration, improved for both groups (Fig 2).

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

Evaluation of the epithelial phenotype in the central cornea by laser IVCM was the most objective primary endpoint. Based on the established criteria, complete success required that the corneal epithelial phenotype of the transplanted eyes must have improved by at least one step towards the normal corneal epithelial phenotype. At baseline, there was no significant difference in the distribution of the conjunctival-like or the mixed epithelial phenotype in the central cornea (Fisher's exact test, baseline p=0.226). Consequently, the outcome of each type of cell transplant could not have been influenced by a more frequent presence of a more favorable phenotype before cell therapy in any of the three etiologic groups (Table 4). The change in epithelial phenotype at the central cornea (Fig 2) was not significantly different between CLET and MSCT at 6 months (p=0.524) or at 12 months (p=0.5562). After 6 months, 50% of CLET cases (95% CI, 23.7 to 6.3) and 62.5% of MSCT cases (95% CI, 35.9 to 83.7) had improved the epithelial phenotype in the central cornea. At 12 months after surgery, 66.7% of CLET (95% CI, 30.9 to 91) and 71.4% of MSCT (95% CI, 42 to 90.4) had improved the epithelial phenotype in the central cornea. The differences between CLET and MSCT were not significant (p=0.6891 and p=1.000 at 6 and 12 months, respectively).

For CLET, a corneal epithelial phenotype was present in 20.0% (95% CI, 3.5 to 55.8) and 33.3% (95% CI, 9 to 69.1) of the transplants at 6 and 12 months, respectively

(differences were not significant). For MSCT, the corneal epithelial phenotype was present in 43.8% (7 cases, 95% CI, 20.8 to 69.5) at 6 months and 57.1% (8 cases, 95% CI, 29.7-81.2) at 12 months (p=0.0469, and p=0.0234, respectively). The percentage of cases reaching a corneal-like epithelial phenotype was not significantly different between CLET and MSCT at 6 (p=0.4152) or 12 months (p=0.854). Survival curve analysis showed that the differences between CLET and MSCT survival was not significant at either 6 or 12 months (log-rank test, p = 0.817, Fig 2). See Fig 3 as example.

In summary, our hypothesis of non-inferiority for MSCT versus CLET was confirmed at 6 and 12 months (p=0.0446 and p=0.0244, respectively).

# DISCUSSION

This is the first clinical trial showing that allogeneic bone marrow-derived MSC can be safely transplanted to the human ocular surface, as far as we know. By doing that in the context of a controlled double-masked randomized trial, we demonstrated that non-epithelial stem cells are safe and as efficient (MSCT 85.7% success) as corneal epithelial stem cells derived from the limbus (CLET 77.8% success) in restoring the corneal epithelial phenotype damaged due to LSCD. Due to its proof-of-concept design, the number of transplants was small but statistically sufficient to prove our hypothesis. Additionally, through the balanced allocation of different etiologies to either group, we guaranteed no bias in this sense. Finally, as in any initial exploratory trial, only severe LSCD of diverse etiology were included.

hAM was used as a substrate to culture both limbal epithelial cells and MSC in the present clinical trial. hAM itself has re-epithelizing, anti-fibrotic, anti-inflammatory,

anti-angiogenic, and anti-microbial features.<sup>38</sup> In patients with partial and/or mild 441 LSCD, which maintain residual stem cell function, hAM transplantation can improve 442 their clinical situation by supporting regeneration of residual limbal stem cells.<sup>39</sup> 443 However, cell-free hAM transplantation is insufficient for regeneration of the ocular 444 surface in patients with severe and/or total LSCD. 13,14 445 Because this is the first study of its kind, our results cannot be compared with similar 446 studies in humans; nevertheless, they can be compared to other CLET series, 447 including our own in which Ramirez et al. compared the CLET success rates among 448 previous studies and found that ours was similar to or better than the others. 11 449 At present, the only techniques accepted as established therapies to recuperate from 450 corneal epithelial failure due to limbal niche destruction is either full tissue (limbal) 451 transplant<sup>5,6,40</sup> or cell-based therapy.<sup>2,4,8–12,41</sup> While other cell therapies have been 452 proposed and tested in animals, the only established cell therapy approved so far for 453 humans is CLET, which is based on in vitro cultivation of limbal niche cells. At 454 455 present, a cell product (Holoclar®) has just been approved by the European Agency of Medicines based on a previous large study. 9 The authorization includes the 456 performance of a post-authorization multicenter trial. The approved indication for this 457 product is moderate to severe limbal deficiency due to chemical/thermal burns, and 458 the cells must be autologous, thus restricting all other etiologies and all bilateral 459 cases unless a 1-2 mm<sup>2</sup> limbal biopsy can be safely removed from one of the eyes. 460 For bilateral cases or other etiologies, allogeneic CLET has proved to be effective in 461 our hands, 11 confirming previous studies. 8,10,41,42 462 However, CLET has several limitations. The main one is that epithelial stem cells 463 must be extracted from their niches where they are thought to represent less than 464

10% of all cells. 1,3 This means that the procedure is dependent upon healthy donor eyes or cadaveric donations, which may be limited. In practice, the small limbal biopsies can be lost because of contamination and/or lack of adequate growth. Typically, it takes 3-5 weeks to cultivate a sufficient quantity for transplantation. These factors make both autologous and allogenic CLET expensive and timeconsuming. The natural step forward is to turn to the most commonly used adult stem cell in regenerative medicine, MSC. Currently these cells are being used in clinical trials to treat multiple diseases including osteo-articular, liver, kidney, cardiac, hematological (graft-versus-host disease), lung problems, keratoconus, and preclinically in retinal repair. 43-46 There is also abundant literature on the successful use of these cells in animal models of limbal deficiency or corneal burns. 17,47–53 Although it remains unclear if MSC can transdifferentiate into corneal epithelial cells,51-56 strong evidence suggests that other multiple mechanisms contribute simultaneously to the therapeutic action. These cells have the capacity to migrate into injured tissues<sup>53</sup> and exert anti-inflammatory and immunomodulatory properties. They have paracrine activity via the production of multiple trophic and growth factors that reduce tissue injury and protect tissue from further adverse effects while enhancing tissue repair. Finally, they are able to stimulate development of resident stem cells. 16,17,24,25,53-60 However, all of these effects have not been demonstrated in humans yet. In the case of CLET, there are preclinical data indicating that transplanted limbal cells migrate to the damaged limbal and corneal areas and that they repopulate and regenerate them to some extent. 61-63 However, in humans this has never been demonstrated, mainly due to the technical difficulty of tracking in vivo the transplanted cells.

465

466

467

468

469

470

471

472

473

474

475

476

477

478

479

480

481

482

483

484

485

486

487

Most of our failures (57.1%) were chemical injuries, confirming other authors' reports<sup>9</sup> and our own previous results.<sup>11</sup> The second most frequent failures (42.9%) were immune-mediated inflammatory diseases. None of the cases with non-inflammatory diseases failed. The overall effectiveness of the both CLET and MSCT was high considering that 57% of the cases were chemical burns, of which 75% were successful regardless of which transplantation protocol was used. Similarly, 25% of the cases were immune-mediated inflammatory cases, of which 57% were successful.

Although within each transplant type, one of the symptom questionnaires showed improvement, our experience during this trial and the previously published one<sup>11</sup> is that patients had a lot of difficulty in expressing in writing what they were feeling. For instance, many complained about the numerous questions they needed to answer, and at the end they were not sure what to answer. Plus it was very confusing for them to answer when they had a useful remaining eye, especially in the vision-related quality of life questionnaire.

This trial included only severe end-stage LSCD syndromes, as is typical for initial clinical trials. Vision improvement is usually dependent on multiple factors beyond the LSCD, and consequently this was not the goal of either transplant procedure in this trial. Although some patients proceeded with further surgical measures to improve vision after the 12-month mandatory follow-up period, these procedures were beyond the scope of this study. Nevertheless, our encouraging results indicate the possibility of restoring to health less severe limbal disease with these transplants, thus preventing further corneal damage and visual deterioration. Finally, these cell transplant procedures may ultimately improve the health of the limbal niche where stem cells and other important cells normally reside. This will result in greater

success if corneal transplantation from either cadaveric donors or by artificial corneas becomes necessary, making the cell transplantation procedures complementary to the tissue transplantation procedures.

# **CONCLUSIONS**

In summary, we have shown in this proof-of-concept clinical trial that MSC used in MSCT can safely and effectively help treating corneal pathology due to LSCD. Further progress in treating severe and blinding pathology due to LSCD will depend on more research that explores the mechanism by which the transplanted stem cells improve the corneal surface cell phenotype. One of the next steps is to organize multicenter clinical trials of both MSCT and CLET. However before that can occur, it is essential to develop a commonly agreed upon set of diagnostic criteria for LSCD so that the prevalence of this pathology can be determined and that data developed among the participating centers can be effectively compared.

# **Acknowledgements**

We thank Dr. J. Murta, Dr. M. J. Quadrado (Coimbra, Portugal), Dr. T. RodríguezAres (Santiago de Compostela), Dr. J. C. Pastor, Dr. D. Galarreta (Valladolid), Dr. C.
Sánchez-Tomero (Madrid), Dr. J. M. Granados (Albacete), Dr. O. Martín (Albacete),
and Dr. B. Hoyos (Cádiz) for referring patients to this trial. We also thank Dr. B.
Bromberg (Certified Editor in Life Science of Xenofile Editing
(www.xenofileediting.com) for his assistance in the final editing and preparation of
this manuscript. This work was supported by Advanced Therapies Program, Ministry

of Health, Spain [SAS/2481/2009]; Regional Center for Regenerative Medicine and
Cell Therapy, Castile and Leon, Spain [SAN 1178/200]; CIBER-BBN, Spain; Spanish
I Network on Cell Therapy, Spain [TerCel RD12/0019/0036]. The authors have no
commercial or proprietary interest in any concept or product described in this article.
The protocol (EudraCT 2010-023535-42) was approved by the local Ethics
Committee of our institution and the Spanish Agency of Medicines and Sanitary
Products (AEMPS, www.aemps.gob.es). The Clinical Trials.gov Identifier is
NCT01562002. All authors have read the journal's authorship agreement and policy
on disclosure of potential conflicts of interest. Presented in part at the 2017 annual
meeting of the Association for Research in Vision and Ophthalmology (ARVO) as a
paper: Invest Ophthalmol Vis Sci 2017;58:ARVO E-abstract 3372.

### REFERENCES

- 1. Notara M, Alatza A, Gilfillan J, et al. In sickness and in health: Corneal epithelial stem cell biology, pathology and therapy. *Exp Eye Res*.
- 554 2010;90(2):188-195.
- Oie Y, Nishida K. Regenerative medicine for the cornea. *Biomed Res Int.* 2013;2013.
- Dziasko MA, Armer HE, Levis HJ, Shortt AJ, Tuft S, Daniels JT. Localisation of epithelial cells capable of holoclone formation in vitro and direct interaction with stromal cells in the native human limbal crypt. *PLoS One*. 2014;9(4).
- 560 4. Nakamura T, Inatomi T, Sotozono C, Koizumi N, Kinoshita S. Ocular surface 561 reconstruction using stem cell and tissue engineering. *Prog Retin Eye Res.* 562 2016;51:187-207.
- 563 5. Kenyon KR, Tseng SC. Limbal autograft transplantation for ocular surface disorders. *Ophthalmology*. 1989;96(5):709-22-3.
- 565 6. Sangwan VS, Basu S, MacNeil S, Balasubramanian D. Simple limbal epithelial 566 transplantation (SLET): a novel surgical technique for the treatment of 567 unilateral limbal stem cell deficiency. *Br J Ophthalmol*. 2012;96(7):931-934.
- 7. Pellegrini G, Traverso CE, Franzi AT, Zingirian M, Cancedda R, De Luca M.

  Long-term restoration of damaged corneal surfaces with autologous cultivated

  corneal epithelium. *Lancet*. 1997;349(9057):990-993.
- 571 8. Shortt AJ, Secker GA, Rajan MS, et al. Ex Vivo Expansion and Transplantation 572 of Limbal Epithelial Stem Cells. *Ophthalmology*. 2008;115(11):1989-1997.

- 9. Rama P, Matuska S, Paganoni G, Spinelli A, De Luca M, Pellegrini G. Limbal
- Stem-Cell Therapy and Long-Term Corneal Regeneration. *N Engl J Med.*
- 575 2010;363(2):147-155.
- 10. Baylis O, Figueiredo F, Henein C, Lako M, Ahmad S. 13 years of cultured
- limbal epithelial cell therapy: a review of the outcomes. *J Cell Biochem*.
- 578 2011;112(4):993-1002.
- 11. Ramírez BE, Sánchez A, Herreras JM, et al. Stem Cell Therapy for Corneal
- Epithelium Regeneration following Good Manufacturing and Clinical
- Procedures. *Biomed Res Int.* 2015;2015:408495.
- 582 12. Singh V, Shukla S, Ramachandran C, et al. Science and Art of Cell-Based
- Ocular Surface Regeneration. In: ; 2015:45-106.
- 13. Tseng SC, Prabhasawat P, Barton K, Gray T, Meller D. Amniotic membrane
- transplantation with or without limbal allografts for corneal surface
- reconstruction in patients with limbal stem cell deficiency. *Arch Ophthalmol*
- 587 (Chicago, III 1960). 1998;116(4):431-441.
- 588 14. Sabater AL, Perez VL. Amniotic membrane use for management of corneal
- limbal stem cell deficiency. *Curr Opin Ophthalmol.* 2017;28(4):363-369.
- 590 15. Joe AW, Gregory-Evans K. Mesenchymal Stem Cells and Potential
- Applications in Treating Ocular Disease. *Curr Eye Res.* 2010;35(11):941-952.
- 16. Ren G, Chen X, Dong F, et al. Concise review: mesenchymal stem cells and
- translational medicine: emerging issues. Stem Cells Transl Med. 2012;1(1):51-
- 594 58.

- Yao L, Bai H. Review: mesenchymal stem cells and corneal reconstruction. *Mol* Vis. 2013;19(November):2237-2243.
- 18. Rohban R, Pieber TR. Mesenchymal Stem and Progenitor Cells in
   Regeneration: Tissue Specificity and Regenerative Potential. *Stem Cells Int.* 2017;2017:1-16.
- O'Callaghan AR, Daniels JT. Concise review: limbal epithelial stem cell
   therapy: controversies and challenges. *Stem Cells*. 2011;29(12):1923-1932.
- Zhang L, Coulson-Thomas VJ, Ferreira TG, Kao WWY. Mesenchymal stem
   cells for treating ocular surface diseases. *BMC Ophthalmol*. 2015;15(S1):155.
- Luetzkendorf J, Nerger K, Hering J, et al. Cryopreservation does not alter main characteristics of Good Manufacturing Process–grade human multipotent mesenchymal stromal cells including immunomodulating potential and lack of malignant transformation. *Cytotherapy*. 2015;17(2):186-198.
- Lužnik Z, Bertolin M, Breda C, et al. Preservation of Ocular Epithelial Limbal
   Stem Cells: The New Frontier in Regenerative Medicine. *Adv Exp Med Biol.* 2016;951:179-189.
- Osei-Bempong C, Ghareeb AE, Lako M, Figueiredo FC, Armitage WJ. Defining the optimal cryoprotectant and concentration for cryopreservation of limbal stem cells. *Cryobiology*. 2018;84:98-102.
- Ho MSH, Mei SHJ, Stewart DJ. The Immunomodulatory and Therapeutic
   Effects of Mesenchymal Stromal Cells for Acute Lung Injury and Sepsis. *J Cell Physiol.* 2015;230(11):2606-2617.

- Griffin MD, Ritter T, Mahon BP. Immunological Aspects of Allogeneic
   Mesenchymal Stem Cell Therapies. *Hum Gene Ther*. 2010;21(12):1641-1655.
- 619 26. Orozco L, Soler R, Morera C, Alberca M, Sánchez A, García-Sancho J.
- Intervertebral Disc Repair by Autologous Mesenchymal Bone Marrow Cells: A
- 621 Pilot Study. *Transplantation*. 2011;92(7):822-828.
- Orozco L, Munar A, Soler R, et al. Treatment of Knee Osteoarthritis With
   Autologous Mesenchymal Stem Cells. *Transplant J.* 2013;95(12):1535-1541.
- 28. Vega A, Martín-Ferrero MA, Del Canto F, et al. Treatment of Knee
- Osteoarthritis With Allogeneic Bone Marrow Mesenchymal Stem Cells.
- 626 *Transplantation*. 2015;99(8):1681-1690.
- Dominici M, Le Blanc K, Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy
- position statement. *Cytotherapy*. 2006;8(4):315-317.
- 30. Nieto-Miguel T, Calonge M, de la Mata A, et al. A comparison of stem cell-
- related gene expression in the progenitor-rich limbal epithelium and the
- differentiating central corneal epithelium. *Mol Vis.* 2011;17.
- 633 31. López-Paniagua M, Nieto-Miguel T, De La Mata A, et al. Consecutive
- expansion of limbal epithelial stem cells from a single limbal biopsy. Curr Eye
- 635 Res. 2013;38(5).
- 636 32. Schiffman RM, Christianson MD, Jacobsen G, Hirsch JD, Reis BL. Reliability
- and validity of the Ocular Surface Disease Index. Arch Ophthalmol (Chicago, III
- 638 *1960*). 2000;118(5):615-621.

- Mangione CM, Lee PP, Gutierrez PR, et al. Development of the 25-item
  National Eye Institute Visual Function Questionnaire. *Arch Ophthalmol*(Chicago, III 1960). 2001;119(7):1050-1058.
- 642 34. Ramírez BE, Victoria DA, Murillo GM, Herreras JM, Calonge M. In vivo 643 confocal microscopy assessment of the corneoscleral limbal stem cell niche 644 before and after biopsy for cultivated limbal epithelial transplantation to restore 645 corneal epithelium. *Histol Histopathol.* 2015;30(2):183-192.
- Nubile M, Lanzini M, Miri A, et al. In Vivo Confocal Microscopy in Diagnosis of Limbal Stem Cell Deficiency. *Am J Ophthalmol*. 2013;155(2):220-232.
- 36. Araújo AL de, Ricardo JR da S, Sakai VN, Barros JN de, Gomes JÁP.
   Impression cytology and in vivo confocal microscopy in corneas with total
   limbal stem cell deficiency. *Arq Bras Oftalmol*. 2013;76(5):305-308.
- 651 37. Mastropasqua L, Calienno R, Lanzini M, et al. In vivo confocal microscopy of 652 the sclerocorneal limbus after limbal stem cell transplantation: Looking for 653 limbal architecture modifications and cytological phenotype correlations. *Mol* 654 *Vis.* 2016;22:748-760.
- Jirsova K, Jones GLA. Amniotic membrane in ophthalmology: properties,
   preparation, storage and indications for grafting—a review. *Cell Tissue Bank*.
   2017;18(2):193-204.
- 39. Anderson DF. Amniotic membrane transplantation for partial limbal stem cell
   deficiency. *Br J Ophthalmol*. 2001;85(5):567-575.
- 660 40. Miri A, Al-Deiri B, Dua HS. Long-term outcomes of autolimbal and allolimbal

- transplants. *Ophthalmology*. 2010;117(6):1207-1213.
- 41. Zhao Y, Ma L. Systematic review and meta-analysis on transplantation of ex vivo cultivated limbal epithelial stem cell on amniotic membrane in limbal stem cell deficiency. *Cornea*. 2015;34(5):592-600.
- Basu S, Fernandez MM, Das S, Gaddipati S, Vemuganti GK, Sangwan VS.
   Clinical outcomes of xeno-free allogeneic cultivated limbal epithelial
   transplantation for bilateral limbal stem cell deficiency. *Br J Ophthalmol*.
   2012;96(12):1504-1509.
- Squillaro T, Peluso G, Galderisi U. Clinical Trials with Mesenchymal Stem
   Cells: An Update. *Cell Transplant*. 2016;25(5):829-848.
- 44. Sánchez-Guijo F, Caballero-Velázquez T, López-Villar O, et al. Sequential
   third-party mesenchymal stromal cell therapy for refractory acute graft-versus host disease. *Biol Blood Marrow Transplant*. 2014;20(10):1580-1585.
- 45. Alió del Barrio JL, El Zarif M, de Miguel MP, et al. Cellular Therapy With
   Human Autologous Adipose-Derived Adult Stem Cells for Advanced
   Keratoconus. Cornea. 2017;36(8):952-960.
- 46. Alonso-Alonso ML, Srivastava GK. Current focus of stem cell application in retinal repair. *World J Stem Cells*. 2015;7(3):641.
- Ma Y, Xu Y, Xiao Z, et al. Reconstruction of Chemically Burned Rat Corneal
   Surface by Bone Marrow-Derived Human Mesenchymal Stem Cells. Stem
   Cells. 2006;24(2):315-321.
- 48. Gu S, Xing C, Han J, Tso MOM, Hong J. Differentiation of rabbit bone marrow

- 683 mesenchymal stem cells into corneal epithelial cells in vivo and ex vivo. *Mol*684 *Vis.* 2009;15(January):99-107.
- Jiang T-S, Cai L, Ji W-Y, et al. Reconstruction of the corneal epithelium with
   induced marrow mesenchymal stem cells in rats. *Mol Vis.* 2010;16(July):1304 1316.
- 688 50. Reinshagen H, Auw-Haedrich C, Sorg R V., et al. Corneal surface 689 reconstruction using adult mesenchymal stem cells in experimental limbal stem 690 cell deficiency in rabbits. *Acta Ophthalmol.* 2011;89(8):741-748.
- 691 51. Rohaina CM, Then KY, Ng AMH, et al. Reconstruction of limbal stem cell
  692 deficient corneal surface with induced human bone marrow mesenchymal stem
  693 cells on amniotic membrane. *Transl Res.* 2014;163(3):200-210.
- Ahmed SK, Soliman AA, Omar SMM, Mohammed WR. Bone marrow
   mesenchymal stem cell transplantation in a rabbit corneal alkali burn model (a
   histological and immune histo-chemical study). *Int J Stem Cells*. 2015;8(1):69 78.
- Galindo S, Herreras JM, López-Paniagua M, et al. Therapeutic Effect of Human
   Adipose Tissue-Derived Mesenchymal Stem Cells in Experimental Corneal
   Failure Due to Limbal Stem Cell Niche Damage. Stem Cells.
   2017;35(10):2160-2174.
- Nieto-Miguel T, Galindo S, Reinoso R, et al. In vitro simulation of corneal
   epithelium microenvironment induces a corneal epithelial-like cell phenotype
   from human adipose tissue mesenchymal stem cells. *Curr Eye Res*.
   2013;38(9):933-944.

- 55. Sánchez-Abarca LI, Hernández-Galilea E, Lorenzo R, et al. Human Bone
   Marrow Stromal Cells Differentiate Into Corneal Tissue and Prevent Ocular
   Graft-Versus-Host Disease in Mice. Cell Transplant. 2015;24(12):2423-2433.
- Harkin DG, Foyn L, Bray LJ, Sutherland AJ, Li FJ, Cronin BG. Concise reviews:
   can mesenchymal stromal cells differentiate into corneal cells? A systematic
   review of published data. *Stem Cells*. 2015;33(3):785-791.
- Holan V, Trosan P, Cejka C, et al. A Comparative Study of the Therapeutic
   Potential of Mesenchymal Stem Cells and Limbal Epithelial Stem Cells for
   Ocular Surface Reconstruction. Stem Cells Transl Med. 2015;4(9):1052-1063.
- 715 58. Omoto M, Katikireddy KR, Rezazadeh A, Dohlman TH, Chauhan SK.

  716 Mesenchymal stem cells home to inflamed ocular surface and suppress

  717 allosensitization in corneal transplantation. *Invest Ophthalmol Vis Sci.*718 2014;55(10):6631-6638.
- 719 59. Wen L, Zhu M, Madigan MC, et al. Immunomodulatory effects of bone marrow-720 derived mesenchymal stem cells on pro-inflammatory cytokine-stimulated 721 human corneal epithelial cells. Shi X-M, ed. *PLoS One*. 2014;9(7):e101841.
- Hu N, Zhang Y-Y, Gu H-W, Guan H-J. Effects of bone marrow mesenchymal stem cells on cell proliferation and growth factor expression of limbal epithelial cells in vitro. *Ophthalmic Res.* 2012;48(2):82-88.
- Higa K, Shimmura S, Kato N, et al. Proliferation and Differentiation of
   Transplantable Rabbit Epithelial Sheets Engineered with or without an Amniotic
   Membrane Carrier. *Investig Opthalmology Vis Sci.* 2007;48(2):597.

- 728 62. Xu B, Fan T-J, Zhao J, et al. Transplantation of tissue-engineered human 729 corneal epithelium in limbal stem cell deficiency rabbit models. *Int J* 730 *Ophthalmol*. 2012;5(4):424-429.
- 731 63. Brown KD, Low S, Mariappan I, et al. Plasma polymer-coated contact lenses 732 for the culture and transfer of corneal epithelial cells in the treatment of limbal 733 stem cell deficiency. *Tissue Eng Part A*. 2014;20(3-4):646-655.
- 64. Efron N. Grading scales for contact lens complications. *Ophthalmic Physiol* 735 *Opt.* 1998;18(2):182-186.

### FIGURE LEGENDS

Figure 1. Consort flow diagram. Twenty-eight transplants (23 eyes of 20 patients)
were included and fully assessable at the minimum established period of 6 months.

Eleven (9 eyes, 9 patients) were CLET and 17 (16 eyes, 14 patients) were MSCT.

Figure 2. Evaluation Endpoints and Final Outcome for Cultured Limbal

742

743

759

760

761

738

Epithelial Transplantation (CLET, N=11) and Mesenchymal Stem Cell 744 Transplantation (MSCT, N=17). 745 Panel A shows all clinical signs evaluated with anterior segment slit-lamp 746 biomicroscopy: ocular (conjunctival) redness, corneal epithelial integrity, and central 747 corneal epithelial opacity. Boxes extend from the 25th to the 75th percentile, 748 horizontal bars represent the median, and whiskers extend 1.5 times the length of the 749 interquartile range (IQR) above and below the 75th and 25th percentiles, 750 751 respectively. The mean of each group is shown by black diamonds. Individual values 752 for each subject are indicated by filled circles. Conjunctival redness and central corneal epithelial opacity improved significantly from baseline to final evaluation at 12 753 754 months. Superficial keratitis improved significantly by 6 months. Both CLET and MSCT groups improved similarly, except for corneal opacity, which was significantly 755 better for MSCT at 6 and 12 months (see Table 4 for mean numerical values and 756 Tables 1 and 2 for individual values). Panel B shows mosaic plots. The area of each 757 rectangle is proportional to the observed frequency in that cell. Labels show the 758

conditional relative percentages of each possible epithelial phenotype for each

transplant type at baseline, 6, and 12 months. The number of cases is indicated in

brackets. Mosaic plots represent contingency tables as a matrix of rectangles, the

dimensions of which are proportional to the observed frequencies of each cross-classification. Cases with conjunctival-like (Conj), corneal-like (Corn), or mixed phenotypes were divided into the relative proportions across transplant type. CLET and MSCT performed equally well (no significant differences) regarding this main objective evaluation outcome. The conjunctival-like phenotype decreased while the corneal-like phenotype increased over time. Panel C shows the successful outcome for each type of transplant at each visit (left). Partial success rates are represented by the lighter-colored area. There were no significant differences between the percentage of successful cases with CLET or MSCT at 6 months (72.7% vs 76.5%) or 12 months (77.8% vs 85.7%) months. Kaplan-Meier survival curve analysis (right) shows a probability of success after CLET of 0.818 (95% CI, 0.6191 to 1.00) at 6 months and of 0.716 (95% CI, 0.488 to 1.00) at 12 months. MSCT survival probability was 0.812 at 6 months (95% CI, 0.642 to 1.00) and 0.75 at 12 months (95% CI, 0.565 to 0.995). The difference in survival between the two types of cell transplants was not significant at either of the two periods (log-rank test, p = 0.817).

Figure 3. This 44-year-old woman (patient No. 10, see Tables 1 and 2) had bilateral corneal epithelial failure due to 360° limbal stem cell deficiency caused by a 20-year duration of Stevens-Johnson's syndrome. Opacity was restricted to the anterior cornea (epithelium and anterior stroma) and the lens (cataract; not seen in this photograph). Before entering this trial, she was treated aggressively for her extremely severe secondary dry eye (a spot of squamous metaplasia can be seen at the lower inner periphery of her left limbal area) during the course of 6 months. She lost her first bilateral CLET transplants prematurely (48 h after surgery) due to inadequate contact lens fitting. After her second bilateral transplants (right eye, CLET; left eye,

MSCT), her eyelids were maintained closed for 10 days except to quickly deliver eyedrops, thus keeping the transplants in place. Panels A and B show her right eye and left eye, respectively, 5 weeks before (upper), 6 months (middle) and 12 months (lower) after a successful CLET and after a successful MSCT. *In vivo* confocal microscopy images of the basal central corneal epithelium showed a mixed epithelial phenotype before surgery (upper) and corneal-like phenotypes at 6 months (middle) and 12 months (lower) after both CLET (Panel A) and MSCT (Panel B). Baseline visual acuity in her right eye was 0.25 (upper) and improved to 0.32 at 6 months (middle) and 12 months (lower). Baseline visual acuity in her left eye was 0.01 (upper) and improved to 0.25 at 6 months (middle) and to 0.32 after 12 months (lower). The patient did not want to undergo any other surgery as she was able to carry on with her life, and we did not encourage further surgery as any trauma in these patients carries the risk of triggering violent inflammatory relapse.

#### 800 TABLES

Table 1. Baseline data (0) and outcomes at 6 months (6) and 12 months (12) after cultivated limbal epithelial transplantation (CLET) in the 11 assessable cases (9 eyes, 9 patients) suffering from ocular surface failure due to limbal stem cell deficiency syndrome (LSCD).

CLET No./Eye	Patient No. Gender/ Age	LSCD etiology (months elapsed till transplant): Etiology*/Grade† 2 <sup>nd</sup> diagnoses	SIDEQ 0/6/12	OSDI 0/6/12	VEO 25 0/6/12	Visual Potential‡	BCVA (ETDRS) 0/6/12	Conj redness (0-4) 0/6/12	Central cornea epithelial opacity (0-4) 0/6/12	Corneal neovessel area (0-4) 0/6/12	Corneal neovessel length (0-4) 0/6/12	Corneal ( staining (0-4) 0/6/12	Corneal PED (0-4) 0/6/12	Central corneal epithelial phenotype (IVCM) 0/6/12	Days to AM re-absorption from surgery	Final outcome 6 and 12 months	Comments
1/OD	1/F/49	Post-infectious keratitis (240)+2 previous PKP: 3/T Glaucoma	24/9/16	89.6/ 50.0/65.9	45.4/ 57.3/55.0	4	0.025/ 0.001/0.001	1/1/1	3/3/3	2/2/1	2/1/1	4/2/1	0/0/0	Conj/ Conj/Conj	24	Partial success/ Partial success	PKP performed at month 18. Although remaining clear, vision deteriorated due to advanced glaucoma
2/OS	2/F/77	Persistent corneal ulcer (20) 2nd to recurrent ocular surface carcinoma (120): 3/T	17/17/13	87.5/ 50.0/60.0	46.8/ 49.0/35.3	1	0.04/ 0.2/0.25	2/2/1	1/1/0	2/1/1	2/1/1	3/1/0	3/0/0	Conj/ NP/Corn	30	Partial success/ Success	Patient chronically immunosuppressed due to heart transplant
3/OD	3/M/47	Chemical+mechanical injury (24): 1/T Irreversible retinal pathology	8/6/8	57.5/ 25.0/32.5	47.1/ 32.5/34.5	0	0.001/ 0.001/0.001	2/2/1	4/3/2	3/1/1	3/2/1	3/2/1	0/0/0	Conj/ Mix/Mix	30	Success/ Success	-
4/OS	7/M/48	Chemical injury (46)+2 previous AMT: 1/T	14/13/-	62.5/ 83.3/-	50.9/ 43.8/-	4	0.001/ 0.001/-	3/2/-	3/3/-	4/3/-	3/3/-	4/3/-	2/1/-	Conj/ Conj/-	90	Partial Success/-	Regrafted CLET-5, for further potential improvement
5/OS	7/M/48	Chemical injury (54)+2 previous AMT+previous CLET-4: 1/T	9/8/14	83.3/ 62.5/80.6	52.9/ 50.9/55.0	4	0.001/ 0.001/0.001	2/1/1	3/2/2	3/3/3	3/2/2	3/1/0	1/0/0	Conj/ Mix/Mix	90	Success/ Success	-
6/OS	9/M/62	Chemical injury (600)+2 previous AMT+ PKP: 1/T Glaucoma, exotropia Chemical injury (608)+2	13/18/-	61.1/ 85.4/-	53.8/ 57.0/-	4	0.001/ 0.001/-	3/3/-	2/3/-	4/4/-	3/3/-	2/2/-	3/2/-	Conj/ Conj/-	21	Failure/-	Regrafted CLET-7
7/OS	9/M/62	previous AMT+previous PKP+previous CLET-6: 1/T Glaucoma, exotropia	17/22/16	93.8/ 72.7/85.4	53.6/ 53.4/56.0	4	0.001/ 0.01/0.01	3/2/2	3/3/2	4/4/4	3/3/3	2/0/0	2/0/0	Conj/ Conj/Conj	30	Partial success/ Partial Success	-
8/OD	10/F/44	Stevens Johnson+multiple AMT (120): 2/T Cataract	28/20/21	100.0/ 93.8/97.5	47.4/ 49.8/47.3	2	0.25/ 0.32/0.32	3/3/1	3/1/1	3/1/1	3/2/1	2/1/1	0/0/0	Mix/ Corn/Corn	8	Success/ Success	-
9/OD	13/M/48	Chemical injury (84)+3 previous AMT: 1/T	16/11/15	81.8/ 72.9/79.2	30.6/ 35.8/39.8	4	0.0001/ 0.0001/0.0001	2/2/1	4/4/4	4/4/4	4/4/4	0/0/0	0/0/0	Conj/ Mix/Mix	30	Failure/ Failure	-
10/OD	16/M/50	Graft vs host disease (168)+5 previous PKP+conj flap+sclera patch: 2/T	17/22/18	87.5/ 90.0/89.6	56.3/ 59.1/52.5	0	0.001/ 0.001/0.001	4/3/2	4/4/4	4/4/4	4/4/4	0/0/0	0/0/0	Conj/ Conj/Conj	10	Failure/ Failure	Extremely thin cornea under pannus prevented its removal at surgery. Thickness increased so as to proceed with a
11/OS	21/M/41	Chemical injury (36)+5 AMT: 1/T Cataract, glaucoma	10/11/9	62.5/ 64.6/87.5	53.0/ 55.1/48.4	4	0.32/ 0.5/0.4	2/1/1	2/2/2	3/2/1	2/1/1	2/0/0	0/0/0	Mix/ Corn/Corn	34	Success/ Success	keratoprothesis PKP at month 14 +cataract surgery +hard contact lens recovered full vision
Mean (SD)		181.8 (219.2)	15.7 (6.1)/ 14.3 (5.8)/ 14.4 (4.1)-	78.8 (15.1)/ 68.2 (20.7)/ 75.4 (19.8)	489 (7.0)/ 49.4 (8.8)/ 47.1 (8.6)			-	-	-	-	-	-	-	36.1 (27.9)		-
Median (IQR)		-	-	-	-			2 (1)/ 2 (1)/ 1 (0)	3 (1)/3 (1)/2 (1)	3 (1)/ 3 (2.5)/ 1 (3)	3 (0.5)/ 2 (1.5)/ 1 (2)			-			-

Assessable cases were those reaching at least 6 postoperative months; \*1: chemical injuries, 2: immune-based inflammatory diseases, 3: non-inflammatory diseases; †T: total, S: severe; ‡Visual potential: 1, improvement with CLET only (corneal opacity was only superficial); 2, improvement with one surgery different form corneal transplant after CLET (i.e. cataract removal); 3, improvement with subsequent corneal transplant plus another surgery (cataract removal unless otherwise specified) after CLET, and 0: No possibility of improvement (i.e., due to irreversible retinal pathology); BCVA, Best corrected visual acuity; BCVA values 0.01, 0.001, 0.0001, and 0.00001 equivalent to counting fingers, hand motion, light perception, and no light perception respectively; ; ETDRS, Early Treatment Diabetic Retinopathy Study; Conj, conjunctival; Corn, corneal; IQR, interquartile range; IVCM, in vivo confocal microscopy; SIDEQ, Single Item Dry Eye Questionnaire; VFQ25, National Eye Institute 25-item Visual Function Questionnaire (0-100); OSDI, Ocular Surface Disease Index (0-100); PED, persistent epithelial defect; AMT, amniotic membrane transplantation; PKP, penetrating keratoplasty; SD, standard deviation; M, male; F, female; NP: not performed; OS, left eye; OD, right eye.

Table 2. Baseline data (0) and outcomes at 6 months (6) and 12 months (12) after bone marrow-derived mesenchymal stem cell transplantation (MSCT) in the 17 assessable cases (16 eyes, 14 patients) suffering from ocular surface failure due to limbal stem cell deficiency syndrome (LSCD).

MSCT No. /Eye	Patient Nº Gender /Age	LSCD etiology (months elapsed till transplant): Etiology*/Grade†/ 2nd diagnoses	SIDEQ 0/6/12	OSDI 0/6/12	VFQ25 0/6/12	Visual Potential‡	BCVA (ETDRS) 0/6/12	Conj redness e (0-4) 0/6/12	Central corneal pithelial opacity (0- 4) 0/6/12	Corneal neovessel: area (0-4) 0/6/12	Corneal neovessel length (0-4) 0/6/12	Corneal staining (0-4) 0/6/12	Corneal PED (0-4) 0/6/12	Central corneal epithelial phenotype (IVCM) 0/6/12	Days to AM reabsorption from surgery	Final outcome 6/12	Comments
1/OS	4/F/31	Chemical injury (24)+ 2 previous conj resection+AMT: 1/T Cataract	18/15/8	39.6/ 47.9/64.6	22.6/ 43.0/29.4	2	0.125/ 0.158/ 0.125	1/1/1	1/2/3	3/3/3	2/2/3	2/0/2	1/0/0	Mix/ Conj/Conj	10	Failure/ Failure	Poor compliance
2/OS	6/M/53	Atopic kerato conjunctivitis (170): 2/T Post-infectious keratitis Unsuccessful cataract surgery	24/23/-	95.8/ 93.8/-	43.6/ 41.0-	2	0.04/ 0.025/-	3/0/-	2/0/-	3/0/-	2/0/-	1/0/-	0/0/-	Mix/ NP/-	17	Partial success/	Withdrawn at month 7: intense systemic flare-up worsened both eyes, perforating OS cornea; PKP required
3/OD	7/M/48	Chemical injury (46)+multiple AMT: 1/T Glaucoma	12/15/-	77.1/ 85.4/-	55.0/ 56.8/-	4	0.001/ 0.001/-	3/3/-	3/3/-	4/4/-	4/4/-	3/2/-	0/0/-	Conj/ Conj/-	25	Failure/	Regrafted MSCT-4
4/OD	7/M/48	Chemical injury (54)+multiple AMT+previous MSCT-3: 1/T Glaucoma, cataract	9/8/11	83.3/ 62.5/77.8	54.2/ 50.9/57.8	4	0.001/ 0.001/ 0.001	3/2/1	3/2/1	4/4/4	4/3/2	2/2/1	0/0/0	Conj/ Mix/Corn	60	Success/ Success	PKP at month 15
5/OD	8/M/70	Multiple surgeries for proliferative vitreoretinopathy (48): 3/S Irreversible macular pathology	6/3/7	12.5/ 15.6/6.3	54.6/ 63.8/62.4	4	0.001/ 0.001/ 0.001	1/1/0	2/0/1	2/1/1	1/1/1	2/1/1	1/0/0	Mix/ Corn/Corn	10	Success/ Success	PKP at month 30
6/0	10/F/44	Stevens Johnson (120)+multiple AMT: 2/T Cataract	28/23/16	1000/ 93.8/97.5	46.1/ 49.3/41.8	2	0.01/ 0.25/ 0.32	3/3/1	4/3/1	4/3/2	3/2/1	3/2/2	0/0/0	Mix/ Corn/Corn	8	Success/ Success	-
7/OS	11/F/37	Congenital aniridia (132): 3/S Cataract, nystagmus	16/14/12	38.9/ 45.5/63.6	39.3/ 41.6/43.3	4	0.062/ 0.1/0.1	1/0/0	4/3/2	3/2/1	2/2/2	0/0/0	0/0/0	Conj/ Mix/Mix	22	Success/ Success	-
8/OD	12/F/42	Atopic kerato conjunctivitis (132)+2 previous AMT: 2/T Cataract	21/20/-	79.2/ 77.1/-	39.2/ 40.2/-	2	0.031/ 0.025/-	1/2/-	1/1/-	2/2/-	2/2/-	1/1/-	0/0/-	Conj/ Conj/-	15	Failure/	Regrafted MSCT-9 (Table 2)
9/OD	12/F/42	Atopic kerato conjunctivitis (140)+2 previous AMT+previous MSCT- 8: 2/T	17/21/23	66.7/ 66.7/75.0	33.9/ 39.7/27.8	2	0.05/ 0.05/ 0.05	2/2/1	1/1/1	2/2/2	2/2/2	1/1/1	0/0/0	Conj/ Conj/Conj	15	Failure/ Failure	Poor compliance Systemic disease poorly controlled
10/0	14/F/53	Cataract.  Congenital aniridia (160): 3/S Cataract, nystagmus, glaucoma	21/20/21	83.3/ 75.0/85.4	44.7/ 43.8/47.8	2	0.1/ 0.08/ 0.08	1/1/0	2/1/1	2/1/1	2/1/1	3/1/0	0/0/0	Mix/ Corn/Corn	28	Success/ Success	Had a previous CLET (lost transplant No. 7) Cataract surgery at month 20 did not recover vision
11/OD	17/F/28	Chemical injury (72)+previous cadaveric limbal transplant +AMT+PKP: 1/T Severe perennial allergic conjunctivitis, cataract.	19/13/13	64.6/ 33.3/33.3	40.3/ 35.8/46.8	4	0.01/ 0.01/ 0.01	3/2/1	1/1/1	4/4/4	3/2/2	1/1/0	0/1/0	Conj/ Mix/Conj	20	Partial success/ Partial success	PKP at month 13
12/OD	18/M/65	Chemical injury (360)+previous  AMT+PKP+intracorneal rings: 1/T 2  Cataract	24/16/16	81.3/ 70.8/79.2	44.1/ 33.6/37.2	4	0.12 0.5/0.4	2/1/1	1/1/1	3/3/3/	2/2/2	2/0/0	0/0/0	Mix/ Corn/Corn	29	Success/ Success	PKP at month 20
13/OS	19/F/49	Chemical injury (80)+oral mucosal transplant+2 AMT: 1/S	23/19/18	91.7/ 97.9/83.3	37.5/ 48.8/38.6	3	0.01/ 0.2/0.2	1/1/1	2/1/0	2/1/1	2/1/1	1/0/0	0/0/0	Conj/ Corn/Corn	20	Success/ Success	PKP at month 24
14/OS	20/F/44	Stevens Johnson (90)+3 previous	23/16/21	83.3/ 62.5/81.3	46.1/ 47.8/42.5	4	0.158/ 0.5/0.32	3/2/2	2/2/1	2/2/2	1/1/1	4/3/1	0/0/0	Mix/ Corn/Corn	15	Success/ Success	Cataract progression
15/OD	21/M/41	Chemical injury (45)+5 previous AMT+PKP: 1/T Cataract, glaucoma	9/12/7	64.6/ 64.6/70.8	44.0/ 33.6/48.0	4	0.01/ 0.01/ 0.001	4/3/2	4/2/1	4/3/3	3/2/1	4/NP/1	4/2/0	Conj/ Conj/Mix	85	Partial success/ Success	Initially performed for impending perforation
16/OD	22/M/54	Cataract	20/18/7	70.8/ 66.7/89.6	40.6/ 21.7/36.3	3	0.25/ 0.25/ 0.25	2/1/1	1/0/0	1/1/1	2/1/1	0/0/0	0/0/0	Mix/ Corn/Corn	30	Success/ Success	-
17/OS	22/M/54	Chemical injury (95)+3 previous AMT: 1/T Cataract	28/22/19	77.1/ 70.8/56.3	37.0/ 34.4/33.6	4	0.001/ 0.001/ 0.001	4/2/1	4/4/3	4/4/4	3/3/2	3/1/0	3/0/0	Conj/ Conj/Conj	60	Partial success/ Partial success	-
Mean (SD) Median		108 9 (77 8)	18.7 (6.5)/ 16.4 (5.4)/ 14.2 (5.7)	71.2 (22.5)/ 66.5 [21.8]/ 68.9 (24.0)	42.5 (8.0)/ 42.7 (9.9)/ 42.4 (9.8)		0.001	- 2 (2)/ 2 (1)/ 1	- 2 /21/4 /41/4	- 3 (2)/ 2 (2)/	- 2 (1)/ 2 (1)/				27.6 (21.1)		
(IQR)			-	-	-	-	-	2 (2)/ 2 (1)/ 1 (0)	2 (2)/ 1 (1)/ 1 (0)	3 (2)/ 2 (2)/ 2 (2)	2 (1)/ 2 (1)/ 1.5 (1)			-		-	

Assessable cases were those reaching at least 6 postoperative months; \*1: chemical injuries, 2: immune-based inflammatory diseases, 3: non-inflammatory diseases; †T: total, S: severe; ‡Visual potential: 1, improvement with CLET only (corneal opacity was only superficial); 2, improvement with one surgery different form corneal transplant after CLET (i.e. cataract removal); 3, improvement with subsequent corneal transplant plus another surgery (cataract removal unless otherwise specified) after CLET, and 0: No possibility of improvement (i.e. due to irreversible retinal pathology); BCVA, Best corrected visual acuity; BCVA values 0.01, 0.001, 0.0001, and 0.00001 equivalent to counting fingers, hand motion, light perception, and no light perception respectively; Conj, conjunctival; Corn, corneal; IQR, interquartile range; IVCM, in vivo confocal microscopy; VFQ25, National Eye Institute 25 Visual Function Questionnaire (0-100); OSDI, Ocular Surface Disease Index (0-100); SIDEQ, Single Item Score Dry Eye questionnaire (0-28); AMT, amniotic membrane transplantation; PED, persistent epithelial defect; PKP, penetrating keratoplasty; SD, standard deviation; M, male; F, female; NP, not performed; OS, left eye; OD, right eye.

Table 3. Ocular Surface Clinical Signs Evaluated to Define and Grade Limbal Stem Cell Deficiency and Score the Evaluation End-Points.

			Corneal Epithe	elial Integrity		Visual Prognosis and Poten	tial for Visual Recovery	
	Conjunctival Redness*+	Central Corneal Epithelial Opacity*	Superficial Punctate Keratitis*	Epithelial Ulceration Area*	Corneal Superficial Neo Vascularization Area/Length*	Previous Ocular Media Opacity	Surgeries Judged Necessary to Recover Full Potential Vision	Central Corneal Phenotype (In <i>Vivo</i> Confocal Microscopy)†
Grade 0	White conjunctiva	None		None	None /None	Any grade of corneal opacity plus non-corneal irreversible visual loss (e.g., irreversible retinal pathology, advanced glaucoma)	No potential for gain: Stem cell transplant performed to deal with pain and avoid globe removal	CORNEAL:  Regular, hexagonal cells with a cell diameter  <20 µm; dark cytoplasm, dark nucleus; hyper- reflective, bright well-defined cell margins.
Grade 1	Widening of the vessels	Mild: clearly visible pupil		≤1/4	≤¹/₄ / 1mm	Corneal opacity restricted to anterior cornea (and anterior stroma)	One surgical procedure: stem cell transplant only	CONJUNCTIVAL: Closely packed round or irregularly shaped cells; cell diameter of >20 µm (irregular size); large nucleus/cytoplasm ratios; dark cytoplasm and bright, hyperreflective nucleus with ill- defined cell margins. Occasional goblet cells
Grade 2	Mild redness	Moderate: hazily visible pupil		>¼ and ≤½	>¼ and ≤½ / 2-3 mm	Corneal opacity restricted to anterior cornea (as Grade 1) plus another non-corneal reason for visual loss (e.g., cataract)	Two surgical procedures: stem cell transplant + non- corneal surgery (e.g., cataract removal)	MIXED: Both corneal and conjunctival phenotypes are present
Grade 3	Moderate redness	Severe: faintly visible pupil		>½ and ≤¾	>½ and ≤¾ / 4-5 mm	Full thickness corneal opacity	Two surgical procedures: stem cell transplant + corneal transplant	
Grade 4	Intense redness	Severe: no visible pupil		>¾	>¾ / ≥6 mm	Full thickness corneal opacity plus another non-corneal reason for visual loss (e.g., cataract)	Three surgical procedures: stem cell transplant + corneal transplant + non-corneal surgery (e.g. cataract removal)	

<sup>\*</sup>Evaluated by slit-lamp biomicroscopy.

806

807

†Evaluated with the Heidelberg Retinal Tomograph HRT-3 and Rostock Cornea Module (HRT3, Heidelberg Engineering GmbH, Heidelberg, Germany).

<sup>+</sup> Evaluated following the Efron Scale for conjunctival redness.<sup>64</sup>

Table 4. Characteristics, Endpoint Values, and Outcome of the 28 Assessable Cases (23 eyes from 20 patients) of Corneal Epithelial Failure Due to Limbal Epithelial Stem Cell Deficiency Randomized to Cultivated Limbal Epithelial Transplantation (CLET) or Mesenchymal Stem Cell Transplantation (MSCT).

CHARACTERISTIC / ENDPOINT	CLET (N = 11)	MSCT (N = 17)	p- values*
Females/Males — no. (%); 95% confidence interval (CI)	3 (27.3); 7.3 to 60.7/ 8 (72.7); 39.1 to 92.7	9 (52.9); 28.5 to 76.1/ 8 (47.1); 23.9 to 71.5	0.2530
Age — years (mean±SD)	52.4±10.5	47.2±10.8	0.3448
Limbal Stem Cell Deficiency			
Grade: Total/ Severe — no. (%), 95% CI	11 (100) /0 (0)	13 (76.5); 49.8 to 92.2/ 4 (23.5); 7.8 to 50.2	0.1324
Etiology — no. (%); 95% CI			
Chemical Burns —16 (57.1)	7 (63.6); 31.6-87.6	9 (52.9); 28.5 to 76.1	0.8669
Immune-based Inflammatory Diseases — 7 (25.0)	2 (18.2); 3.2 to 52.3	5 (29.4); 11.4 to 56	0.8232
Non-inflammatory Diseases, Other — 5 (17.9)	2 (18.2); 3.2 to 52.3	3 (17.7); 4.7 to 44.2	1
Months from Disease Onset to Cell Transplant — mean±SD	181.8±219.2	108.9±77.8	0.9437
Days from Cell Transplant to Supporting Amniotic Membrane Reabsorption — mean±SD	36.1±27.9	27.6±21	0.1634
Primary Evaluation End-Points Baseline/ 6 months/ 12 months			
Symptoms/Quality of Life* Questionnaires (range) — mean±SD			
Single Item Dry Eye Questionnaire SIDEQ (0-28)	15.7±6.1/ 14.3±5.8/ 14.4±4.1	18.7±6.5/ 16.4±5.4/ 14.2±5.7 <b>p=0.0237</b> and <b>p=0.0336</b> between baseline and 6 or 12 months	0.2379/ 0.3408/ 0.9178
Ocular Surface Disease Index OSDI (0-100; severe>32)	78.8±15.1/ 68.2±20.7/ 75.4±19.8 p=0.0318 and p=0.0072 between baseline and 6 or 12 months	71.2±22.5/ 66.5±21.8/ 68.9±24	0.4795/ 0.8352/ 0.4306
National Eye Institute 25-item Visual Function Questionnaire NEI-VFQ25 (0-100)	48.9±7/ 49.4±8.8/ 47.1±8.6	42.5±8/ 42.7±9.9/ 42.4±9.8	0.0239/ 0.0774/ 0.2524
Clinical signs (range) — median (interquartile range [IQR])			
Conjunctival redness (0-4)	2 (1)/ 2(1)/ 1 (0) p=0.0012 between baseline and 12 months	2 (2)/ 2(1)/ 1 (0) p<0.0001 between baseline and 12 months	0.6057/ 0.2638/ 0.2438
Central corneal epithelial opacity (0-4)	3 (1)/ 3 (1)/ 2 (1) p=0.0129 between baseline and 12 months	2 (2)/ 1 (1)/ 1 (0) p=0.0023 between baseline and 12 months	0.1323/ 0.0275/ 0.04
Corneal epithelial integrity: superficial punctate keratitis (0-4)	2 (1)/ 1 (2)/ 0 (1) p=0.0428 and p=0.0012 between baseline and 6 or 12 months	2 (2)/ 1 (1.25)/ 0.5 (1) p=0.0263 and p=0.0006 between baseline and 6 or 12 months	0.4693/ 0.7356/ 0.3532
Corneal epithelial integrity: persistent epithelial defect or ulceration (0-4)	0 (2)/ 0 (0)/ 0 (0)	0 (0)/ 0 (0)/ 0 (0)	0.2558/ 0.3422/ 1
Epithelial phenotype in central cornea ( <i>in vivo</i> confocal microscopy) — no. (%); 95%			,

CI			
Conjunctival phenotype	9 (81.8); 47.8 to 96.8/	9 (52.9); 28.5 to 76.1/	0.24
18 (64.3)/ 11 (42.3)/ 7 (30.4)	5 (50); 23.7 to 76.3/	6 (37.5); 16.3 to 64.1/	1.
	3 (33.3); 9 to 69.1	4 (28.6); 9.6 to 58	0.24
Mixed phenotype	2 (18.2); 3.2 to 52.3 /	8 (47.1); 23.9 to 71.5/	1.
10 (35.7)/ 6 (23.1)/ 5 (21.7)	3 (30); 8.1 to 64.6/	3 (18.8); 5 to 46.3/	0.49
	3 (33.3); 9 to 69.1	2 (14.3); 2.5 to 43.9	0.57
Corneal phenotype	0 (0); 0 to 32.2/	0 (0); 0 to 22.9/	0.82
0 (0)/ 9 (34.6)/ 11 (47.8)	2 (20); 3.5 to 55.8/	7 (43.8); 20.8 to 69.5%)/	0.41
	3 (33.3); 9 to 69.1	8 (57.1); 29.7 to 81.2	0.8
Secondary Evaluation End-Points —	, ,	, ,	
baseline/ 6 months/ 12 months			
Corneal neovessels: area (0-4) — median	3 (1)/ 3 (2.5)/ 1 (3)	3 (2)/ 2 (2)/ 2 (2)	0.31
(IQR)	p=0.0129 between baseline and 12	p=0.0307 and p=0.0045 between	0.5
. ,	months	baseline and 6 or 12 months	0.84
Corneal neovessels: length (0-4) —	3 (0.5)/ 2 (1.5)/ 1 (2)	2 (1)/ 2 (1)/ 1.5 (1)	0.0
median (IQR)	p=0.0307 and p=0.0044 between	p=0.0055 between baseline and 12	0.22
, ,	baseline and 6 or 12 months	months	0.70
Best-corrected visual acuity: all cases† —	$0.06\pm0.11/0.09\pm0.17/0.11\pm0.16$	0.06±0.07/ 0.13±0.16/ 0.13±0.14	0.14
mean±SD			0.11
			0.36
Best-corrected visual acuity: successful	$0.08\pm0.13/0.13\pm0.19/0.14\pm0.18$	$0.06\pm0.08/\ 0.15\pm0.18/\ 0.14\pm0.15$	0.68
cases† — mean±SD			0.45
OLIO OFFICE IN CLIT COME	0 (70 7) 00 0 1 00 7/	10 (7) (7) 10 0 1 00 0	0.93
SUCCESSFUL OUTCOME	8 (72.7); 39.3 to 92.7/	13 (76.5); 49.8 to 92.2/	0.82
6 months/ 12 months — no. (%); 95% CI	7 (77.8); 40.2 to 96.1	12 (85.7); 56.2 to 97.5	0.62

\* Between CLET and MSCT groups. Significant p-values are highlighted in bold characters.
Only significant P-values are shown in the 2nd (CLET) and the 3rd (MSCT) columns.

†Only one case, CLET n°2 (Table 1), had a grade 1 potential to recover visual acuity, meaning that it was previously considered that her damage was restricted to the corneal epithelium. Her visual acuities were 0.04, 0.2 and 0.25 at baseline, 6, and 12 months, respectively.

Table 5. Serious and Non-serious Adverse Events Encountered in All Cell Transplants
Performed (N = 37), Including 9 Transplants that Did Not Reach the Minimum
Established 6 Months of Follow Up (5 CLET and 4 MSCT) Plus the 28 That Did (11
CLET and 17 MSCT)\*.

EVENT		CLET (N = 16)		N	/ISCT (N = 21)	
	No. of Events (%)/ Transplant-Patient No./ Baseline Disease/ Immuno- suppressants	Relation to Study medication (Cell Transplant)/ Severity/ Attributable Relation	Final Outcome/ Comments	No. of Events (%)*/ Transplant-Patient No./ Baseline Disease/ Immuno- suppressants	Relation to Study Medication (Cell Transplant)/ Severity/ Attributable Relation	Final Outcome/ Comments
Serious adverse events						
Herpes simplex keratitis				1 (2.7)/ 8-15/ Persistent corneal ulcer due to herpes simplex keratitis/ Mycophenolate mofetyl	Unrelated/ Moderate/ Recurrence of baseline disease, facilitated by surgical trauma and/or immuno suppression used	Solved with sequelae/ Penetrating corneal transplant performed due to risk of perforation
Corneal perforation				1 (2.7)/ 2-6/ Atopic keratoconjunctivitis secondary to severe atopic dermatitis/ Cyclosporine+ azathioprine	Unrelated/ Severe/ Intense relapse of baseline disease, already poorly controlled systemically	Solved with sequelae/ Tectonic corneal transplant
Ocular surface neoplasia	1 (2.7)/ 2-2/ Moderate/ Persistent corneal ulcer due to ocular surface neoplasia and its required treatments/ Cyclosporine+ azathioprine	Unrelated/ Moderate/ Recurrence of baseline disease at same rate as before cell transplant (patient long-term immuno suppressed due to previous heart transplant)	Solved (excisional surgery) with no sequelae/ Cell transplant action was not interrupted as persistent corneal wound healed			
Non-serious adverse events†		а апорыну				
Loss of transplant within 24 hr after surgery (Not assessable at first evaluation)	1 (2.7)/ 4-5/ Severe/ Chemical injury/ mycophenolate mofetil	Unrelated/ Baseline disease? (same fate for all previous ammiotic membranes grafts)	Unsolved with no sequelae/ No further actions taken; patient was withdrawn	3 (8.1)/ 1,2,3-5/ Severe/ Chemical injury/ mycophenolate mofetil	Unrelated/ Severe/ Baseline disease (same fate for all previous amniotic membrane grafts)	Unsolved with no sequelae/ No further actions taken; patient was withdrawn
Loss of transplant within 48 hr after	4 (10.8)/4, 6-14; 7- 14; 9-19/ Severe/ Stevens-Johnson (No. 5, 6);	Unrelated/ Severe/ Bandage contact lens displacement?	Solved with o sequelae (all regrafted successfully)			

surgery	congenital aniridia (No. 7); chemical injury (No. 9)/ Cyclosporine (No.5, 6); azathioprine (No. 7); mycophenolate mofetil (No. 9)				
Corneal			2 (5.4)/ 14-20/	Unrelated/ Mild/	Solved with
erosion			Moderate/ Stevens-	Misdirected	no sequelae/
			Johnson's	lashes rubbing	Lashes were
			syndrome/	cornea due to	removed
			Micophenolate	underlying	permanently
			mofetil	disease	
Flu episode			1 (2.7)/ 10-14/	Unrelated/ Mild/	Solved with
			Moderate/	Patient was not	no
			Congenital aniridia/	vaccinated	sequelae/
			azathioprine		no effect in
			-		transplant

\* CLET, cultivated limbal epithelial transplantation; MSCT, mesenchymal stem cell transplantation.

† Non-serious adverse events shown are those that were moderate or severe. The remaining non-serious adverse events were mild, easily solved and all were considered to be unrelated with cell transplant: nausea, vomiting, pharyngitis, twisted ankle, and reconstruction of anophthalmic socket to improve existent cosmetic prosthesis in the contralateral eye. Three patients complained of mild asthenia and their mycophenolate mofetil was lowered to 1.5 d/day in one patient and to 1 mg/kg in 2 more patients. Two patients had transient blood pressure mild elevation that was brought under control by lowering their cyclosporine dose from 5 to 3 mg/kg/day.

Figure 1
Click here to download high resolution image

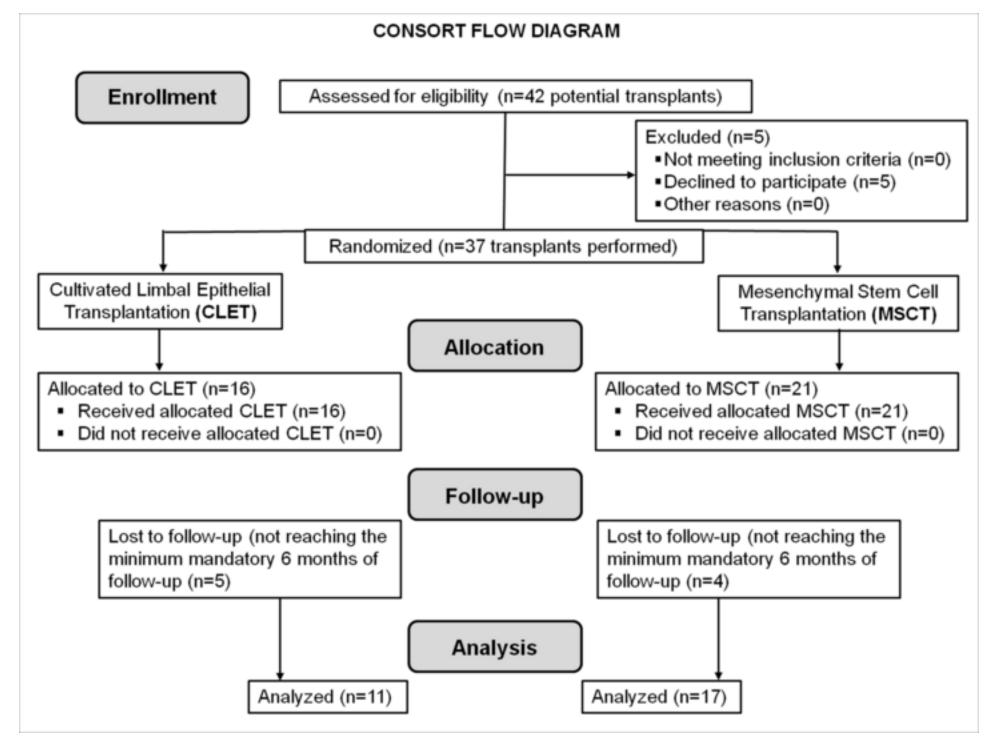


Figure 2
Click here to download high resolution image

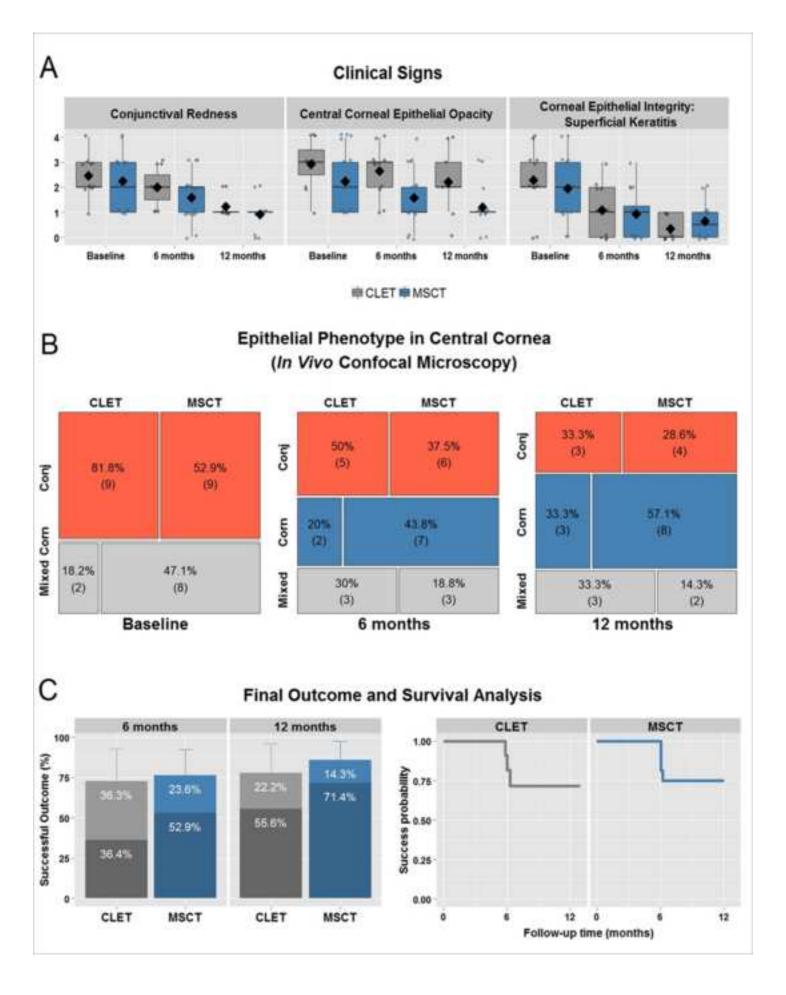
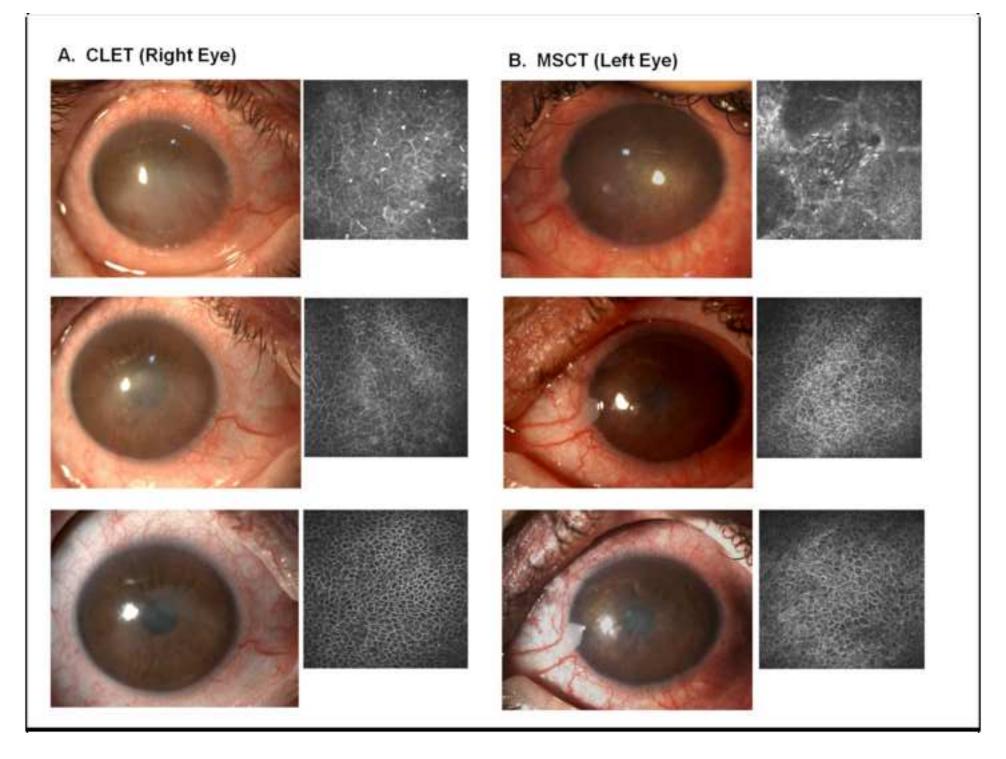


Figure 3 Click here to download high resolution image



# **Supplementary Appendix**

This appendix has been provided by the authors to give readers additional information about this study.

Supplement to: Calonge M, Pérez I, Galindo S, Nieto-Miguel T, López-Paniagua M, Fernández I, Alberca M, García-Sancho J, Sánchez A, Herreras JM. A Proof-of-Concept Clinical Trial Using Mesenchymal Stem Cells for the Treatment of Corneal Epithelial Stem Cell Deficiency

#### **Table of Contents**

Supplemental Methods	2
Preparation of amniotic membrane	2
Preparation of cultivated limbal epithelial cells for transplantation (CLET)	2
Preparation of bone marrow-derived mesenchymal stem cells for transplantation (MSCT)	3
Immunofluorescence microscopy	4
Patients	5
Evaluation end-points	
Supplemental Results	8
Characterization of bone marrow-derived mesenchymal stem cells	8
Immunofluorescence microscopy	8
Supplementary Tables	
Supplementary Table S1	10
Supplementary Table S2	10
Supplementary Figure	11
Supplementary Figure S1	11
Supplemental Discussion of Immunofluorescence Results	12
Supplemental References	14

### **Supplemental Methods**

### Preparation of amniotic membrane

Human amniotic membranes (2.5x2.5 cm) were used as carriers for cells, and they were prepared using our previously described method. Briefly, the membranes were stored at -80°C upon arrival. Immediately before use, they were thawed, washed with phosphate buffered saline (PBS, Life Technologies-Gibco, Carlsbad, CA, USA), treated with trypsin for 15 min at room temperature, and gently scraped to remove the epithelial cells from the underlying basement membrane. Afterwards, the samples were washed twice in PBS to remove cellular debris to obtain the de-epithelialized amniotic membranes. Each amniotic membrane was attached to the bottom of a 35-mm cell culture dish with the basement membrane side up. The quality criteria for the amniotic membrane were as follows: (1) minimum size to almost cover a 35-mm diameter culture plate, (2) tissue sterility, (3) tissue integrity, and (4) adherence to the plate.

### Preparation of cultivated limbal epithelial cells for transplantation (CLET)

Cadaveric limbal rings were preserved within 7 days from donor death, and they were processed during the 4 h after arrival following a modification of our previously reported protocols.<sup>1,2</sup> Briefly, two 2x2 mm pieces of limbal tissue (limbal explants) were extracted from limbal ring, and both were plated onto the de-epithelialized amniotic membranes (2.5x2.5 cm). The limbal explants were maintained initially under a drop of fetal bovine serum (FBS) (Life Technologies-Gibco) in standard conditions of 37°C, 95% humidified air, and 5% CO<sub>2</sub> gas mixture. After 24 h, 3 ml of the following culture medium were added: DMEM/F12 media (1:1 mixture) (Life Technologies-Gibco), 5% FBS (Life Technologies-Gibco), 50 µg/ml hydrocortisone (Sigma Aldrich, St. Louis, MO, USA), 0.5 ng/ml

cholera toxin (Gentaur, Kampenhout, Belgium), 5 ng/ml insulin-transferrin-selenium (ITS) (Sigma Aldrich), 0.5% dimethylsulfoxide (Sigma Aldrich), 2.5 ng/ml human epidermal growth factor (Life Technologies-Gibco), and 0.5 mg/ml gentamicin (Life Technologies-Gibco). The limbal explants were kept in culture until a cellular outgrowth front of approximately 2 mm was present, and then they were removed to allow further cell proliferation until cells reached 90% confluence (3-4 weeks). The mean time for explant removal was 12.18 ± 1.17 days (mean ± standard error of the mean, SEM). The culture medium was changed every 3 days. The quality criteria for the limbal explants were as follows: (1) 2x2 mm size, (2) tissue sterility, (3) adherence to the amniotic membrane, and (4) appearance of cell outgrowth in less than 15 days.

# Preparation of bone marrow-derived mesenchymal stem cells for transplantation (MSCT)

Bone marrow was processed as we previously reported,<sup>3,4</sup> obtaining 20-200x10<sup>6</sup> of mesenchymal stem cells from every donor after 2 weeks of culture. Subsequently, the mesenchymal stem cells were characterized following the International Society for Cellular Therapy (ISCT) position statement.<sup>3,5,6</sup> Expression of the positive markers CD73, CD90, CD105, and CD166 and the negative markers CD14, CD34, CD45, and HAD-DR was analyzed by flow cytometry. In addition, cell viability was studied by trypan blue staining. Data were reported as means ± SEMs. For MSCT use, 100,000 fresh cells (passage 2) were seeded in a drop of FBS onto a 2.5x2.5 cm piece of de-epithelized amniotic membrane. After 2 h, 2 ml of DMEM medium (High glucose; Life Technologies-Gibco) containing 20% FBS (Life Technologies-Gibco), and 0.5 mg/ml gentamicin (Life Technologies-Gibco) were added, and incubation continued at 37°C, 95% humidified air, and 5% CO<sub>2</sub> gas mixture until cells achieved 90% confluence (3-5 days) The culture medium was changed every 3 days.

#### Immunofluorescence microscopy

Amniotic membrane-cell grafts, with either cultivated epithelial limbal cells or mesenchymal stem cells, were monitored under a phase contrast microscope (Eclipse TS100, Nikon, Tokyo, Japan) and fixed with 4% formaldehyde (Panreac, Barcelone, Spain). Before immunofluorescence assays, each amniotic membrane-cell graft was cut into 5 pieces of about 1 cm<sup>2</sup> each. Immunofluorescence assays were performed following a previously reported protocol.<sup>2</sup> The samples were permeabilized for 10 min with 0.3% Triton X-100 (Sigma-Aldrich), blocked with 5% donkey serum (Sigma-Aldrich) for 1 h at room temperature, and incubated overnight at 4°C with specific primary antibodies (Table S1). Subsequently, samples were incubated 1 h at room temperature with the corresponding secondary antibody (Alexa Fluor® 488 donkey anti-mouse 1:200 or donkey anti-rabbit 1:300; Life Technologies). Cell nuclei were counterstained with propidium iodide (1:6,000; Life Technologies). Each piece obtained from a single amniotic membrane-cell graft was incubated with one primary antibody. The marker analyzed in each piece was randomly assigned. Regarding the total number of experiments performed, the same marker was analyzed in different areas of the total amniotic membrane-cell graft surface (near the explant or at the graft edge). Images were acquired with an inverted fluorescence microscope (DM4000B, Leica, Wetzlar, Germany). The percentage of positive cells was estimated for each marker. Negative controls included the omission of primary antibodies. All antibodies were previously validated in different positive controls by our research group.<sup>2,7</sup> At least four samples from different cell donor were analyzed for each condition (n=4).

#### **Patients**

Prior to the initiation of these procedures, all patients and all allogeneic tissue donors underwent mandatory screening for the following transmittable diseases: human immunodeficiency virus, human T-cell leukemia-lymphoma virus, syphilis, and hepatitis B-C.

For transplantation surgery, retrobulbar anesthesia was achieved with 3 cc of 5% lidocaine (Lidocaine Braun®, Braun Medical SA, Mensungen, Germany). First, a conjunctival peritomy was performed and tissues were recessed, leaving the sclera bare. Fibrovascular pannus, if present, was scraped and removed from the recipient cornea extending to the limbal area, allowing a gentle 360° limbal peritomy to be performed. The scraped surface was polished with a diamond bur, and bleeding vessels were cauterized. Then the CLET or MSCT graft was carefully lifted from the culture dish and placed with the cells facing the recipient ocular surface. The graft was then sutured to the perilimbal episclera, 2-4 mm posterior to the limbus, with 8 interrupted 10-0 nylon stitches. Topical eyedrops (see below) were then applied, and an 18-22 mm diameter bandage contact lens was set in place, and the eye was patched for 24 h.

Twenty-four hours after surgery, each patient was evaluated and topical treatment with the fixed combination of 1% prednisolone acetate and 0.3% tobramycin (Tobradex®, Alcon Laboratories, FT. Worth, TX, USA) was prescribed 4 times per day until the amniotic membrane dissolved. The stitches and the contact lens were also removed between 4 and 6 weeks. Then, 1 mg/ml dexamethasone (Maxidex®, Alcon Laboratories) was instilled 4 times a day and slowly tapered in the next 3 months.

#### **Evaluation end-points**

Limbal stem cell deficiency-related symptoms and their impact on daily life activities were evaluated with three self-administered questionnaires. The Single Item Dry Eye Questionnaire (SIDEQ) gives a 0-4 score to each of 5 different questions about the presence of dryness, foreign body sensation, burning/stinging, pain, itching, sensitivity to light, and blurred vision (maximum score: 28); the Ocular Surface Disease Index (OSDI) also evaluates ocular surface symptoms with 12 questions, and scores >12 indicate abnormal symptomatology, and >32 means severe symptoms (maximum score 100).8 The visual function-related aspects of the quality of life were evaluated with The National Eye Institute 25-item Visual Function Questionnaire (NEI-VFQ25), where higher scores on a 0 to 100 scale indicate better function.9

Right after the questionnaires were administered, best corrected visual acuity was measured using the standard Early Treatment Diabetic Retinopathy Study (ETDRS), as is mandatory in clinical trials. It is crucial to note that vision improvement is never the primary goal of this kind of cell transplantation because this technique intends only to reconstruct the corneal epithelium. It will not affect deeper corneal opacification, cataract, glaucoma that often accompanies these pathologies, or other potential causes of diminished vision such as concomitant retinal pathology in post-multiple surgery cases, nystagmus in congenital aniridia, and others. For these conditions, if they are not irreversible, other visual rehabilitation techniques might be needed after CLET or MSCT. To avoid misinterpretation by the patient, the potential dependence of the visual prognosis on the surgical procedures judged to be necessary to restore vision after cell transplantation was explained at the initial visit and given a grade as shown in Table 1.

After determination of the best corrected visual acuity, the ocular surface clinical status was evaluated as routinely done by anterior segment biomicroscopy using a slit lamp and taking photographs

(IMAGENet program Fuji Fujifilm Finepix S1 Pro. Fuji Photo Film Co., LTD., Tokyo, Japan; Slit lamp Topcon SL-8Z, Topcon Corp., Hasunuma-cho, Habasi-ku, Tokyo, Japan) at each visit. All evaluated parameters at the initial visit and at 6 and 12 months after transplantation and associated scales are shown in Table 1. Ocular redness was evaluated in the bulbar conjunctiva, proximal to the cornea. Nasal and temporal areas were assessed independently based on the Efron scale (score 0-4), <sup>10</sup> and the final score was obtained after averaging both values. Corneal epithelial integrity was evaluated with the vital stain sodium fluorescein using a commercial strip previously wetted and applied to the inferior fornix. After 2 min, the degree of staining was recorded using a cobalt blue filter (Topcon Corp., Tokyo, Japan) over the light source of the slit-lamp biomicroscope and a yellow Wratten #12 filter (Eastman Kodak, Rochester, NY, USA). Both superficial punctuate keratitis (Oxford scheme, 0-5 score) <sup>11</sup> as well as the potential presence of persistent epithelial defect were recorded.

In vivo laser confocal microscopy was the last-performed evaluation end-point (always by same coauthor IP). We used the Heidelberg Retinal Tomograph HRT-3 and Rostock Cornea Module (HRT3, Heidelberg Engineering GmbH, Heidelberg, Germany) and followed the protocol as previously described. Topical anesthesia was achieved with 0.1% tetracaine chlorhydrate and 0.4% oxibuprocaine chlorhydrate solution (Colircusí Anestésico Doble®, Alcon Laboratories) Optical sections from the central cornea were taken at all layers of the epithelium, and the basal layers were then evaluated for the defined phenotypes, as explained in Table 1.

Several other tests were performed at the initial visit and at 6 and 12 months that were not related to outcomes but are part of any routine ophthalmic evaluation. Schirmer test without topical anesthesia evaluated tear production. One Schirmer sterile strip (Tearflo; HUB Pharmaceuticals LLC, Rancho Cucamonga, CA, USA), was placed in the lateral canthus of the inferior lid margin. The length of wetting was measured after 5 min, with eyes closed. Intraocular pressure was evaluated using a Perkins tonometer (Perkins MK 2; HS Clemens Clarke International, Essex, United Kingdom). Fundus

evaluation was by indirect ophthalmoscopy under pharmacologic pupil dilation. When media opacity prevented visualization of intraocular structures by slit-lamp examination or funduscopy, anterior segment optical coherence tomography (OCT) and posterior segment echography (ultrasound) were routinely performed.

During the course of all visits, patients were carefully questioned for potential medication side-effects or any other possible adverse event by two clinicians (co-authors MC and JMH) who also evaluated clinical parameters independently. In case of disagreement, the average score was recorded.

### **Supplemental Results**

### Characterization of bone marrow-derived mesenchymal stem cells

Bone marrow-derived mesenchymal stem cells had the phenotype defined by the ISCT. The positive markers CD73, CD90, CD105 and CD166 were expressed by  $99.7 \pm 0.1\%$ ,  $99.9 \pm 0.02\%$ ,  $97.8 \pm 0.5\%$ , and  $98.98 \pm 0.31\%$  of the cells, respectively. Negative markers CD14, CD34, CD45 and HLA-DR were expressed by  $0.1 \pm 0.04\%$ ,  $0.03 \pm 0.02\%$ ,  $0.09 \pm 0.03\%$ , and  $0.05 \pm 0.02\%$  of the cells, respectively. The viability of the bone marrow-derived mesenchymal stem cells was  $98.6 \pm 0.002\%$ .

#### Immunofluorescence microscopy

Protein markers K15 and p63alpha for limbal epithelial cells and K3 for differentiated corneal epithelial cells were analyzed in both types of cells cultured on amniotic membrane. The percentage of limbal epithelial cells positive for limbal stem cell markers K15 and p63alpha was 90% and 70% respectively (Figure S1 and Table S2). The corneal differentiated epithelial protein K3 was expressed

by 80% of limbal cells (Figure S1 and Table S2). These markers K15, p63alpha, and K3 were expressed by 90% of mesenchymal stem cells cultured on amniotic membrane (Figure S1 and Table S2).

# **Supplemental Tables**

Antibody	Specificity	Category	Clone	Source	Working dilution
Keratin 3 (K3)*	Differentiated corneal epithelial cells	Mouse monoclonal	AE5	Mp Biomedical (Illkirch, France)	1:50
Keratin 15 (K15)	Limbal epithelial stem cells	Mouse monoclonal	LHK15	Millipore (Billerica, MA, USA)	1:50
Alpha isoform of uclear protein 63 (p63alpha)	Limbal epithelial stem cells	Rabbit polyclonal	-	Cell Signaling (Danvers, MA, USA)	1:50

<sup>\*</sup>K3 and K12 are the most specific markers for the corneal epithelium, and they are not expressed in limbal epithelial stem cells.

Either of these markers can be used to determine the corneal epithelial phenotype.

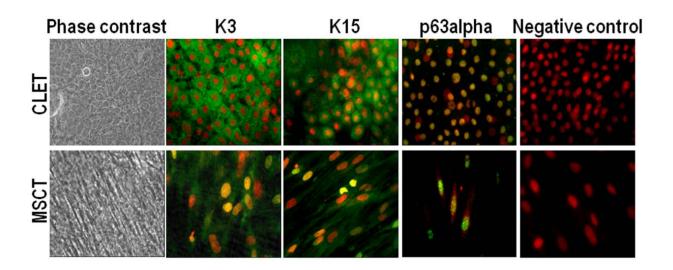
Table S2. Characterization of cultivated limbal epithelial cells and bone marrow-derived mesenchymal stem cells for transplantation (CLET and MSCT, respectively).

Sample	Mean time ofculture on amniotic membrane	Mean % of K3	Mean % of K15	Mean % of p63alpha	Cell morphology	Cell stratification
CLET	3 – 4 weeks	80	90	70	Cuboidal	No
MSCT	3 – 5 days	90	90	90	Elongated	No

K: Keratin; p63alpha: Alpha isoform of nuclear protein 63.

# **Supplemental Figure**

Figure S1. Characterization of cultivated limbal epithelial cells and bone marrow-derived mesenchymal stem cells for transplantation (CLET and MSCT, respectively). Representative images captured by phase contrast (20X magnification) and immunofluorescence microscopes (40X magnification), n=4. Corneal epithelial cell (K3) and limbal epithelial stem cell (K15 and p63alpha) markers were analyzed. Green, K3, K15, and p63alpha marker expression; red, nuclei counterstained with propidium iodide.



### **Supplemental Discussion of Immunofluorescence Results**

The quality of the transplanted amniotic membrane-containing cell grafts (CLET or MSCT) were characterized in parallel. We observed that around 80% of limbal epithelial cells cultured on amniotic membranes expressed the limbal epithelial stem cell markers K15 and p63alpha. This was consistent with the results previously reported by Zakaria et al., <sup>12</sup> although the data are not directly comparable due to different culture media and scaffolds that were used by these authors. In their amniotic membrane-limbal epithelial cell grafts, the predominant phenotype (>50%) consisted of cells that expressed ABCG2, ΔNp63, and K14 markers. Moreover, these authors reported negative expression for the corneal proteins K3/12 and desmoglein. 12 In contrast, we found that protein K3 was also expressed by about 80% of limbal cells on grafts, showing that at least some of these cells expressed limbal epithelial stem cell markers and corneal markers at the same time. These results could suggest the presence of a high percentage of transient amplifying cells (K15<sup>+</sup>, p63alpha<sup>+</sup>, and K3<sup>+</sup>) in amniotic membrane-limbal epithelial cell grafts. These data agree with the fact that human limbal epithelium contains mainly transient amplifying cells and that limbal epithelial stem cells represent less than 10% of the total limbal basal cell population. 13–15 Therefore, our grafts would be suitable for ocular surface treatment. This is consistent with the finding of Rama et al., 16 who reported that cultures containing more than 3% p63 positive cells have a high probability of leading to successful corneal epithelial regeneration. In contrast, cultures with 3% or less p63 positive cells have a lower probability for successful corneal regeneration. Their data are not directly comparable to ours because of the different culture conditions. On the other hand, it is not possible to rule out the potential migration of limbal MSC from the limbal explant stroma to the amniotic membrane under culture conditions. In fact, this could be an explanation for the high expression of K3 and p63alpha observed in CLET cultures, as different authors have reported K3 and p63alpha expression in MSC

cultured on amniotic membrane.<sup>17–22</sup> However, the morphology observed by both phase contrast and immunofluorescence microscopy in the LESC cultures was polygonal, more similar to epithelial-like cells, suggesting that it is very unlikely that MSC from limbal stroma were contaminating the CLET cultures. Mesenchymal stem cells cultivated on amniotic membranes were positive for K3 and K15 proteins. These data agree with previous studies in which different cytokeratins (K3, K12, K18) were expressed by cells obtained from bone marrow or adipose tissues.<sup>17–21</sup> In addition, we detected the p63alpha marker in cells cultivated on the amniotic membranes, in accordance with results reported by other investigators.<sup>22–24</sup> However, the expression of this marker by mesenchymal stem cells is currently controversial because several groups showed that MSC did not express p63 protein.<sup>18,20</sup>

### **Supplemental References**

- Ramírez BE, Sánchez A, Herreras JM, et al. Stem Cell Therapy for Corneal Epithelium Regeneration following Good Manufacturing and Clinical Procedures. *Biomed Res Int.* 2015;2015:408495.
- 2. López-Paniagua M, Nieto-Miguel T, De La Mata A, et al. Consecutive expansion of limbal epithelial stem cells from a single limbal biopsy. *Curr Eye Res.* 2013;38(5).
- Orozco L, Soler R, Morera C, Alberca M, Sánchez A, García-Sancho J. Intervertebral Disc Repair by Autologous Mesenchymal Bone Marrow Cells: A Pilot Study. *Transplantation*. 2011;92(7):822-828.
- Vega A, Martín-Ferrero MA, Del Canto F, et al. Treatment of Knee Osteoarthritis With Allogeneic Bone Marrow Mesenchymal Stem Cells. *Transplantation*. 2015;99(8):1681-1690.
- Orozco L, Munar A, Soler R, et al. Treatment of Knee Osteoarthritis With Autologous Mesenchymal Stem Cells. *Transplant J.* 2013;95(12):1535-1541.
- 6. Dominici M, Le Blanc K, Mueller I, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy*. 2006;8(4):315-317.
- 7. Nieto-Miguel T, Calonge M, de la Mata A, et al. A comparison of stem cell-related gene expression in the progenitor-rich limbal epithelium and the differentiating central corneal epithelium. *Mol Vis.* 2011;17.
- 8. Schiffman RM, Christianson MD, Jacobsen G, Hirsch JD, Reis BL. Reliability and validity of the Ocular Surface Disease Index. *Arch Ophthalmol (Chicago, Ill 1960)*. 2000;118(5):615-621.
- 9. Mangione CM, Lee PP, Gutierrez PR, et al. Development of the 25-item National Eye Institute Visual Function Questionnaire. *Arch Ophthalmol (Chicago, III 1960)*. 2001;119(7):1050-1058.
- 10. Efron N. Grading scales for contact lens complications. *Ophthalmic Physiol Opt*.

- 1998;18(2):182-186.
- 11. Bron AJ, Evans VE, Smith JA. Grading of corneal and conjunctival staining in the context of other dry eye tests. *Cornea*. 2003;22(7):640-650.
- 12. Zakaria N, Possemiers T, Dhubhghaill SN, et al. Results of a phase I/II clinical trial: standardized, non-xenogenic, cultivated limbal stem cell transplantation. *J Transl Med*. 2014;12(1):58.
- Lavker RM, Dong G, Cheng SZ, Kudoh K, Cotsarelis G, Sun TT. Relative proliferative rates of limbal and corneal epithelia. Implications of corneal epithelial migration, circadian rhythm, and suprabasally located DNA-synthesizing keratinocytes. *Invest Ophthalmol Vis Sci*. 1991;32(6):1864-1875.
- 14. Cotsarelis G, Cheng SZ, Dong G, Sun TT, Lavker RM. Existence of slow-cycling limbal epithelial basal cells that can be preferentially stimulated to proliferate: implications on epithelial stem cells. *Cell.* 1989;57(2):201-209.
- 15. Schlötzer-Schrehardt U, Kruse FE. Identification and characterization of limbal stem cells. *Exp Eye Res.* 2005;81(3):247-264.
- 16. Rama P, Matuska S, Paganoni G, Spinelli A, De Luca M, Pellegrini G. Limbal Stem-Cell Therapy and Long-Term Corneal Regeneration. *N Engl J Med*. 2010;363(2):147-155.
- 17. Brzoska M, Geiger H, Gauer S, Baer P. Epithelial differentiation of human adipose tissuederived adult stem cells. *Biochem Biophys Res Commun.* 2005;330(1):142-150.
- 18. Reinshagen H, Auw-Haedrich C, Sorg R V., et al. Corneal surface reconstruction using adult mesenchymal stem cells in experimental limbal stem cell deficiency in rabbits. *Acta Ophthalmol.* 2011;89(8):741-748.
- 19. Vossmerbaeumer U, Ohnesorge S, Kuehl S, et al. Retinal pigment epithelial phenotype induced in human adipose tissue-derived mesenchymal stromal cells. *Cytotherapy*. 2009;11(2):177-188.

- Martínez-Conesa EM, Espel E, Reina M, Casaroli-Marano RP. Characterization of ocular surface epithelial and progenitor cell markers in human adipose stromal cells derived from lipoaspirates. *Invest Ophthalmol Vis Sci.* 2012;53(1):513-520.
- 21. Nieto-Miguel T, Galindo S, Reinoso R, et al. In vitro simulation of corneal epithelium microenvironment induces a corneal epithelial-like cell phenotype from human adipose tissue mesenchymal stem cells. *Curr Eye Res.* 2013;38(9):933-944.
- 22. Shaharuddin B, Osei-Bempong C, Ahmad S, et al. Human limbal mesenchymal stem cells express *ABCB5* and can grow on amniotic membrane. *Regen Med.* 2016;11(3):273-286.
- 23. Reza HM, Ng B-Y, Phan TT, Tan DTH, Beuerman RW, Ang LP-K. Characterization of a novel umbilical cord lining cell with CD227 positivity and unique pattern of P63 expression and function. *Stem Cell Rev.* 2011;7(3):624-638.
- 24. Curtis KM, Aenlle KK, Frisch RN, Howard GA. TAp63γ and ΔNp63β promote osteoblastic differentiation of human mesenchymal stem cells: regulation by vitamin D3 Metabolites. Wijnen A van, ed. *PLoS One*. 2015;10(4):e0123642.