

1 **What are the main factors influencing the presence of faecal bacteria pollution in groundwater**
2 **systems in developing countries?**

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8

9 **Abstract**

10 Groundwater is the major source of drinking water in most rural areas in developing countries. This
11 resource is threatened by the potential presence of faecal bacteria coming from a variety of sources and
12 pollution paths, the former including septic tanks, landfills, and crop irrigation with untreated, or
13 insufficiently treated, sewage effluent. Accurately assessing the microbiological safety of water resources
14 is essential to reduce diseases caused by waterborne faecal exposure. The objective of this study is to
15 discern which are the most significant sanitary, hydrogeological, geochemical, and physical variables
16 influencing the presence of faecal bacterial pollution in groundwater by means of statistical multivariate
17 analyses. The concentration of *Escherichia coli* was measured in a number of waterpoints of different
18 types in a rural area located in the coast of Kenya, assessing both a dry and a wet season. The results from
19 the analyses reaffirm that the design of the well and their maintenance, the distance to latrines, and the
20 geological structure of the waterpoints are the most significant variables affecting the presence of *E. coli*.
21 Most notably, the presence of faecal bacteria in the study area correlates negatively with the
22 concentration of ion Na⁺ (being an indirect indicator of fast recharge in the study site), and also negatively
23 with the length of the water column inside the well.

24 Keywords: *E. coli*, groundwater, Kenia, multivariate statistics, principal component analysis

25

26 **1. Introduction**

27 Worldwide, human populations rely heavily on groundwater as a drinking water source. This situation
28 is even more significant in Asia and Africa, where groundwater is the major source of drinking water and
29 has an important role in improving health and sustaining urban livelihoods (Adelana and MacDonald,
30 2008; MacDonald et al., 2012). Although groundwater is typically assumed to be free of bacterial
31 pathogens, surveys carried out during the last decades indicate that a significant fraction of groundwater

32 supply sources are responsible for water-borne diseases outbreaks around the world (e.g., Bhattacharjee
33 et al., 2002). Globally, 25% of people lack access to water free from microbial contamination (Nowicki et
34 al., 2019). In Africa, this figure doubles, to a value above 50% (Bain et al., 2014), far from compliance with
35 the Sustainable Development Goal number 6 of the United Nations.

36 Hand-pumped tube-wells, being low-cost and low-tech efficient solutions, offer affordable access to
37 shallow aquifers in many developing countries across Africa, Asia and the Pacific. These type of wells,
38 most generally operated by families or small rural communities, are a valid alternative to private or
39 governmentally-operated deep boreholes (Ferguson et al., 2012a). However, they are susceptible to
40 faecal contamination, arising from a variety of sources, such as septic tanks or latrines infiltration,
41 improper disposal of solid urban wastes, leachate from landfills, anthropogenic controlled water
42 recharge, or crop excess irrigation with untreated or insufficiently treated sewage effluent (Charles et al.,
43 2008; Goyal et al., 1984; Matthess et al., 1988; Oteng-Pepurah et al., 2018; Yates et al., 1985).

44 Once bacteria reach the groundwater, and under very favourable conditions with respect to flow,
45 geochemistry and lack of competing indigenous biomass, bacterial pathogens can eventually travel
46 considerably long distances (Sharma and Srivastava, 2011). Groundwater transport in shallow aquifers
47 is primarily a function of the hydrogeological setting and climate conditions (Macler and Merkle, 2000).
48 It is known that the transport, rate of survival, and fate of microbes in the subsurface environment are
49 directly influenced by the microbial population (both diversity and individual characteristics and
50 concentrations, e.g., Barba et al., 2019a), the microbes physical state (dead or alive), the type and
51 characteristics of the subsurface soil and aquifer sediment, and the hydrological conditions, such as water
52 temperature and quality (Rao et al., 1986; Perujo et al., 2017). Therefore, in order to protect drinking
53 water supply wells against microbial contamination, it is essential to establish safe setback distances
54 between wastewater disposal services and water wells (Blaschke et al., 2016). Following numerous
55 laboratory and field-based studies, these safe setback distances should be defined as a function of local
56 soil parameters (e.g., grain sizes, Knappett et al., 2012a, and angularity, Saiers and Ryan, 2005), and
57 general hydrogeology conditions (e.g., Knappett et al., 2008, Pang 2009).

58 Understanding the mechanisms of bacterial fate and transport in the subsurface is of great importance
59 to control soil and groundwater pollution (Carles-Brangari et al., 2017,2018; Sepehrnia et al., 2018a).
60 Recent studies focus in understanding the role of the vadose zone in the flow and transport of *Escherichia*
61 *coli* (*E. coli*) through the soil until reaching the shallow water table in unconfined aquifers (Sepehrnia et
62 al., 2018b; Weldeyohannes et al., 2018). Yet, this should be completed with the detailed analysis of the
63 impact of design, construction, and maintenance of individual wells. As an example, Kilungo et al., (2018)
64 compares the water quality of samples from wells of different designs, in order to help guiding future
65 efforts in providing affordable and sustainable interventions to improve access to clean and safe water
66 in rural communities without centralized supply and sewage networks. Other authors (e.g., Olajuyigbe et

67 al., 2017) examine some relevant socio-economic characteristics of population, such as gender, age,
68 household size, family size, employment, and average income, in order to capture information about the
69 exposure of hand-dug wells to pollution and contamination. Devane et al. (2018) reviewed different
70 tracking tools to recommend the suitable method to determine faecal sources in rural areas. Finally, some
71 studies try to correlate the temporal variation in *E. coli* concentrations as a function of seasonal rainfall
72 characteristics (Elangovan et al., 2018; Guy Howard et al., 2003; Kayembe et al., 2018), well depths,
73 distance to a septic tank, and population density (Dayanti et al., 2018; Martínez-Santos et al., 2017;
74 Rohmah et al., 2018).

75 Some authors investigated the correlation between the presence of faecal bacteria and diverse sanitary
76 risk factors, with the objective of assessing the microbiological risk posed by groundwater sources
77 (Ercumen et al., 2017; Godfrey et al., 2006; Lin et al., 2018). Combining together hydrogeological and
78 non-hydrogeological variables within the same study is quite rare (Ferguson et al., 2012b; Knappett et
79 al., 2012b; Leber et al., 2011; van Geen et al., 2011), and to our knowledge, there is no study that combines
80 different type of variables with the goal of screening the variables that are actually correlated and ranked
81 to eventually predict faecal pollution concentrations. Therefore, the main goal of this paper is to discern
82 what are the hydrological, geochemical, physical, and sanitary variables potentially influencing the
83 presence of faecal bacterial pollution in groundwater sources in rural areas. The method proposed is
84 based on performing a number of multivariate statistics evaluations, being tested in the coastal aquifer
85 located in Kwale (Kenia), one of the multiple zones along the African continent heavily affected by
86 bacterial pollution (Mzuga et al., 1998; Nowicki et al., 2019; Tole, 1997). The analyses involved shallow
87 aquifers of very different geologies and hydrochemical facies, as well as different types of waterpoints in
88 terms of construction and maintenance. Understanding which variables are affecting, and to what degree,
89 the presence of *E. coli* in the groundwater sources, could provide significant knowledge for an accurate
90 management of land uses and water resources to avoid faecal contamination to population. Faecal
91 pollution is the source of a combination of sanitary and educational problems that are perpetuating
92 gender inequality and poverty in rural areas in developing countries.

93

94 2. Methods

95 2.1 Study area

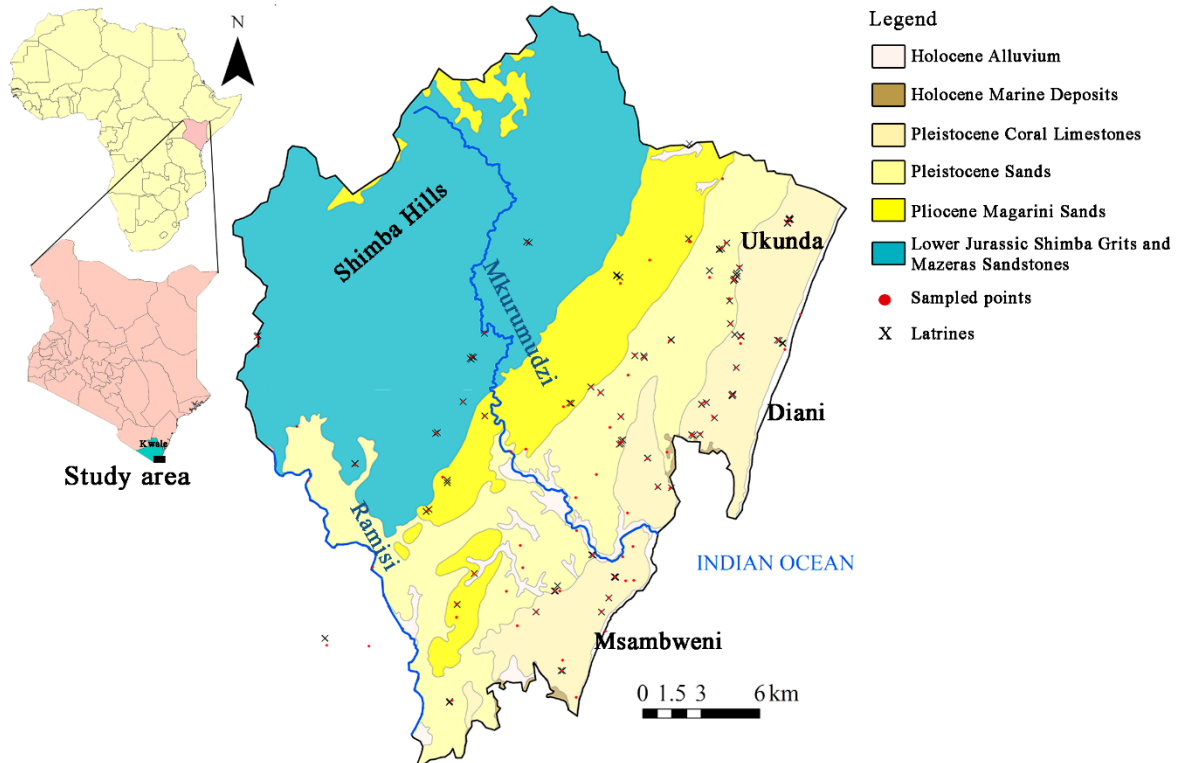
96 The study area is located in Kwale County, a rural coastal area in South-East Kenya, near the border with
97 Tanzania (Fig. 1). The area is populated by small communities spread from the Indian Ocean coast to the
98 Shimba Hills range. The economy of these communities is mainly based in self-consumption livestock.
99 There is no wastewater treatment, and the basic sanitation facilities in the area are pit latrines. The
100 communities are supplied by diverse type of groundwater points (WP) that can be classified in four
101 groups: (1) hand-dug wells (large-diameter wells, less than 30 meter deep, and frequently uncovered),

102 (2) hand-dug wells equipped with handpumps (also large-diameter wells, less than 30 meter deep), (3)
 103 handpump boreholes (small diameter, less than 30 meter deep, with a concrete cover on the surface),
 104 and (4) deep boreholes (small diameter, with depths exceeding 30 m).

105 The study area spans three geological units and two hydrological systems: a deep aquifer composed by
 106 quartz-feldspar sandstones with fossil wood horizons in the lower section constituting the Mazeras
 107 Formation, and a shallow aquifer composed by young geological materials, these including Pliocene
 108 Magarini sands (dominantly quartzitic, and hosting the heavy mineral-rich sands), Kilindini sands (mainly
 109 composed of limestone), and coral reef of the Pleistocene karstified limestone. The hydrochemical facies
 110 and the water isotopic composition indicate hydraulic connectivity across the materials that comprise
 111 the shallow aquifer (Ferrer et al., 2019). The same data show that the Mazeras sandstones in the Shimba
 112 Hills are hydraulically connected with the deep aquifer. Because of this inhomogeneous geological setup,
 113 the hydrochemical composition of the groundwater sampled at the wells has a distinctive signature
 114 depending on the corresponding geological formation.

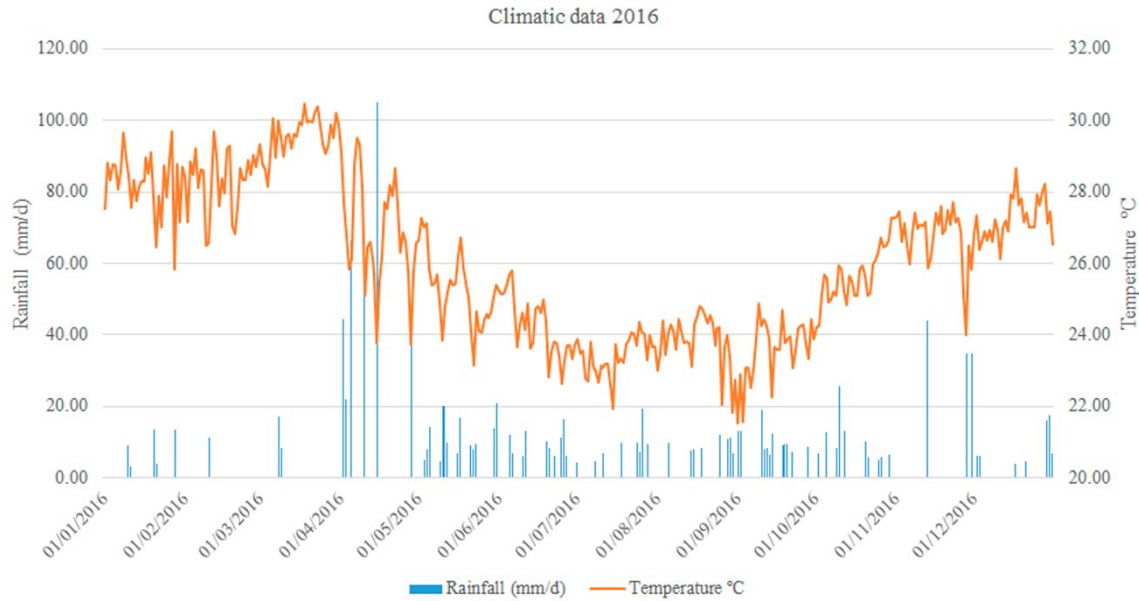
115 The area is characterised by a bimodal rainfall pattern. In Kenya, the “long rains” generally fall from April
 116 to June, whilst the “short rains” occur between October and December (CWSB, 2013). The driest months
 117 are from January to March (Fig. 2).

118



119

120 *Figure 1. Plain view of the study area with the geological units outcropping. The location of the sampled points (red circles)*
 121 *and those of the pit latrines (black crosses) are displayed.*



122

123 *Figure 2. Rainfall and Average daily temperature data in 2016 from a weather station located in the Shimba Hills .*

124 Despite the population density has been reported as a significant variable for faecal bacteria spatial
 125 characterization in other parts of the world (Knappett et al., 2011; van Geen et al., 2011), the information
 126 about the density of each community in the study area is missing.

127 **2.2. Water sampling**

128 Two sampling campaigns were carried out in March 2016 (end of the dry season) and June 2016 (end of
 129 the wet season) to measure several hydrochemical and bacteriological parameters under two very
 130 different climate conditions. During the field surveys, the number of sampling points were 78 (March)
 131 and 77 (June), here including waterpoints from all four groups presented before. In particular, all deep
 132 borehole sampled were on the range of 30 to 80 m depth. In addition, the main rivers in the study area,
 133 Mkurumudzi and Ramisi (Fig. 1), were also sampled in both campaigns.

134 Samples for hydrochemical analysis were taken from wells used daily by the population. The sampling
 135 protocol differed depending on the water point characteristics. In the boreholes or wells equipped with
 136 a handpump, a plastic tube was connected to the outlet, and the internal mechanism was flooded before
 137 sampling to avoid air contact. We ensured that at least three casing volumes of groundwater were
 138 removed by the handpump before sampling. Hand-dug wells were sampled using an electrical pump
 139 whenever the height of the water column allowed it, or using a plastic bucket as the last option. In
 140 completely closed deep boreholes connected to a tank, a pipe was connected to the flow cell and to the
 141 tank entry.

142 Bacteriological samples were taken using the same methodology just explained, except in those points in
 143 which a bucket was needed. In those cases, a stainless steel bucket previously sterilized with ethanol was

144 used. In the waterpoints with handpumps, samples were taken at the outlet point, cleaned with ethanol
 145 before sampling was performed. All the points eventually used in the statistical analyses were sampled
 146 from 8h to 15h, the time rate that the communities in the area pump water, using an integrated sample
 147 by getting some volumes at different times of the day. Furthermore, in 4 specific points we analyzed the
 148 faecal bacteria evolution through a day, by sampling the same well at different hours; however, we could
 149 not establish a clear temporal pattern, since some points showed more pollution at early hours and other
 150 at the latest hours of the day, and so the daily evolution was not eventually explored.

151 **2.3. Physicochemical parameters and ion analyses**

152 The methodology to measure the physicochemical parameters and the diverse ion analyses is described
 153 in detail in Ferrer et al. (2019).

154 **2.4. Bacteria concentration determination**

155 Concentrations of *E. coli* were determined using Aquagenx Compartment Bag Test (CBT) (Aquagenx,
 156 2015). CBTs allow for a quantitative assessment of *E. coli* concentration based on a most probable
 157 number (MPN) along with an upper 95% confidence interval (Foster and Willetts, 2018; Gronewold et
 158 al., 2017; Stauber et al., 2014). MPN testing involves multiple presence/absence tests on different
 159 volumes of the same sample. Samples were collected in sterile purpose-made bags, stored in a fridge
 160 during their transport, and processed within 24h, 30h or 48h after collection, depending on temperature
 161 recommendations by the manufacturer (Stauber et al., 2014). MPN was calculated with data supplied by
 162 the manufacturer, here enclosed as Table 1, and based on the World Health Organization “Guidelines for
 163 Drinking Water Quality” 4th Edition, assigning risk categories of drinking water to *E. coli* level ranges.

164 *Table 1. E. coli risk categories of drinking water (modified from Aquagenx, 2015), and values assigned for the statistical*
 165 *analyses.*

Sampled volume with colour changed	Risk categories	Value assigned for the statistical analyses
0/100 ml	Safe	0
1-10/100ml	Intermediate risk	1
11-100/100 ml	High risk	2
>100/100 ml	Very High risk/Unsafe	3

166

167 **2.5. Sanitary risk inspections**

168 A questionnaire on sanitary risk factors was carried out based on Wright et al. (2013) at each
 169 groundwater point. It comprised 13 questions (see Table 2) that could be answered as Y/N. The first 10
 170 questions were answered for all points, and the last 3 for hand-dug wells only.

171 Table 2. Questions related to value the sanitary risk factors according to Wright et al. 2013

Question 1	Does the cement floor extend more than 1.5 m from the well?
Question 2	Is there any ponding of water on the cement floor?
Question 3	Are there cracks in the cement floor which could permit water to enter the well?
Question 4	Is the pump loose where attached to the base, allowing water to enter the casing?
Question 5	Is the drainage channel cracked, broken or in need of cleaning?
Question 6	Do animals have access to within 10 m of the well?
Question 7	Are there any latrines within 10 m of the well?
Question 8	Are there any additional latrines within 30 m of the well?
Question 9	Are there any open water sources within 20 m of the borehole?
Question 10	Are there any uncapped wells within 30 m of the borehole?
Question 11	Is there any scattered waste within 30 m of the well?
Question 12	Is the cover of the well unsanitary?
Question 13	Is there any scattered waste inside the well?

172

173 **2.6. Multivariate statistics analysis**

174 Multivariate statistics is a suitable technique to treat big datasets involving different sorts of variables,
 175 from quantitative to categorical, and thus amenable to be used to combine biochemical, hydraulic,
 176 geological and external conditions (such as design, drilling characteristics. and maintenance) of water
 177 points (Barba et al., 2019b). Principal Component Analysis (PCA) is a multivariate statistics method
 178 which involves the analysis of a number of parameters or variables, revealing associations between them,
 179 known as (vario)factors or components. Analyses were performed using the IBM-SPSS software.

180 The PCA analyses were subjected to Orthogonal Varimax rotation (Thompson, 2004). This implies the
 181 rotation of the original system into the directions of largest variance in the dataset. Prior to the extraction
 182 of the factors, the Kaiser-Meyer-Olkin (KMO) and the Bartlett sphericity tests were conducted to assess
 183 the suitability of the existing data for factor analysis. KMO returns values between 0 and 1, and values
 184 >0.50 are considered suitable for factor analysis (Hair et al., 1995; Tabachnik and Fidell, 2007). The
 185 Bartlett sphericity test checks if the observed correlation matrix diverges significantly from the identity
 186 matrix. It should be significant ($p < 0.05$) for factor analysis to be suitable.

187 **2.7. Selecting variables for the statistical analyses**

188 Since the objective was to establish the variables that best could explain the *E. Coli* concentration
 189 distribution, arising from a large number of variables, we used a methodology divided in two steps. First,
 190 we removed the main variable of interest, *E. Coli* concentration, from the set, in order to reduce the
 191 number of active variables that could be used later in the final analyses; this way, information
 192 redundancy is eliminated, and the most significant variables or parameters can be elucidated. The second
 193 step involved the introduction of the variable *E. Coli* concentration into the statistical analyses.

194 Statistical parametric methods (such as PCA) perform best when data follows a unimodal symmetric
 195 distribution (Paliy and Shankar, 2016). For this reason, some variables from the initial dataset were
 196 grouped, transformed and/or eliminated. Following Barba et al., (2019b), non-Gaussian hydrochemical
 197 variables were transformed to log concentrations, these being Alkalinity, Eh (a proxy for redox
 198 conditions), and the concentrations of SO_4^{2-} , Na^+ , Cl^- , and SiO_2 . On the other hand, TOC (Total Organic
 199 Carbon), DO (Dissolved Oxygen), and the concentrations of NO_3^- and NH_4^+ , were added to the analysis as
 200 raw data without any transformation (mostly based on a trial and error basis). The most redundant, and
 201 therefore less informative geochemical variables, such as Mg^{2+} , Ca^{2+} and K^+ , were disregarded due to the
 202 strong correlation with other hydrochemical elements. The discrete (also termed categorical) variables
 203 were transformed to continuous ones based on a logical structure, as indicated in Table 3. Due to the zero
 204 variability in the response in the questions 4 and 5 of the questionnaire (Table 2), these two questions
 205 were not included in the analysis, being statistically insignificant.

206 *Table 3. Assigning categorical data to quantitative values to be included in the statistical analysis.*

Variable	Weights	Value assigned	Justification
Geology	Pliocene sands	1	According to the aquifer units composition based on the conceptual model described in Ferrer et al., 2019b.
	Pleistocene sands	2	
	Pleistocene sands /corals	3	
	Pleistocene corals	4	
	Sandstones.	5	
Aquifer unit	Shallow aquifer	0	
	Deep aquifer	1	
Type of well	Hand-dug well	1	Increasing from the simplest structure to the most complex one.
	Hand-dug wells w/handpump	2	
	Handpump borehole	3	
	Deep borehole	4	
Sanitary risk factors (questionnaire. Table 2)	No	0	Binary answer for each one of the questions
	Yes	1	

207

208 **2.8. The use of generalized mixed models**

209 To assess the variables that most significantly influence the presence of *E. coli*, generalized mixed models
 210 with Poisson error distribution (Bates et al., 2015) were used. First, an additional PCA was performed,
 211 including only the variables found significant in the final PCA, but removing the *E.coli* concentration
 212 variable. Then, the scores for the main variofactors were extracted for each observation. Finally, a
 213 generalized mixed models analysis was performed, where concentration of *E. coli* was included as a
 214 dependent variable, and the covariates included correspond to the main principal variofactors of the PCA
 215 runs. As repeated measures were taken at each waterpoint, "Sample ID" was modelled as a random factor.
 216 For all tests, the significance level was set at $\alpha = 0.05$ (two-tailed test). Overdispersion was tested and
 217 eventually corrected by including the number of observation as a random factor (Broström and

218 Holmberg, 2011). The output of the generalized mixed models correspond to the significant correlation
219 between *E.coli* and the variofactor. All runs were performed using R 3.5.1 (Team, 2018).

220

221 3. Results

222 3.1. *E. coli* quantification

223 33 of the 78 waterpoints sampled in March 2016 showed low-risk, meaning no *E. coli* colonies were
224 detected; 5 waterpoints were classified as intermediate-risk, 12 as high-risk, and 28 were in the range
225 very high-risk/unsafe. Samples from surface bodies (rivers) were classified as very high risk. In the June
226 2016 campaign, *E. coli* risk was measured in 77 waterpoints; 34 showed low-risk, 3 intermediate-risk,
227 15 high-risk, and 25 very high-risk/unsafe (see the Supplementary material).

228 From the 72 waterpoints sampled in both campaigns, in 9 (13%) *E. coli* risk reduced from the March to
229 the June campaign; contrarily, in 5 (7%) of the points, the risk increased in that same period.

230 3.2. PCA results

231 Five different PCAs were carried out to evaluate all the information available and considering all type of
232 wells/boreholes present in the study area (Table 4). The variables included in the first sets of PCAs to
233 value to correlation of *E. coli* with all type of water points were: geology; aquifer unit, type of well,
234 sanitary risk factors (Questions 1, 2, 6, 7, 8, 9, 10), field parameters (conductivity, pH, TOC, alkalinity, DO,
235 Eh), hydrochemical parameters (NH₄, Cl, SO₄, NO₃, Na, Si), and seasonality (March or June campaigns).
236 The closest latrine was considered for every waterpoint, even if several were found nearby. Surface water
237 samples were not included in the analyses.

238 All the analyses in this first set are compiled in Table 4. Each analysis involves a different number of
239 variables, as this was a consequence of several trials to find the number of variables that were significant,
240 yet leading to a large value of the KMO test, meaning that the results were statistically significant. For
241 each one of the final PCAs presented in Table 4, we indicate the extracted components, the variables
242 involved in each component, the proportion of variance represented by each component, and the
243 measure of sampling adequacy (KMO test value). A brief explanation about what means each group of
244 correlated variables that form each component is also included for later discussion.

245 *Table 4. Components extracted from the first set of PCA analyses (bold indicates negative correlation); -log indicates*
246 *that log transformation was performed (geochemical variables); Q stands for "question" (from Table 2). For all PCA's*
247 *the Bartlett sphericity test was significant (p<0.001). The variables involved in each component, the proportion of*
248 *variance represented by each component, and the measure of sampling adequacy (KMO test value) are included.*

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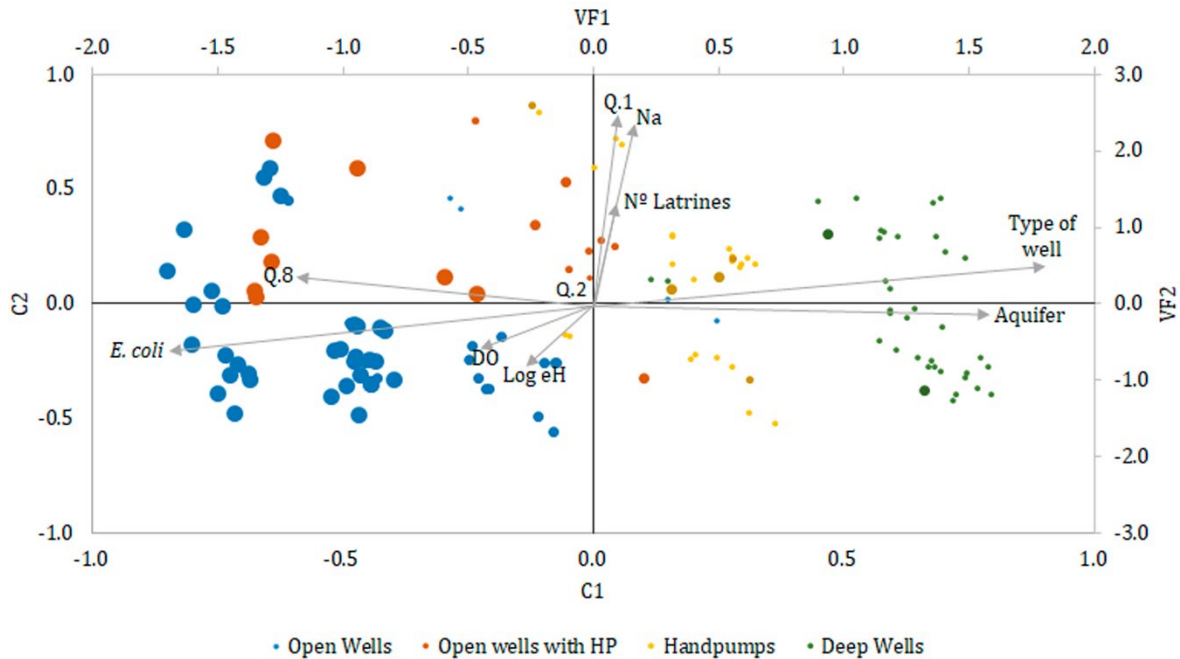
Type of variables	# of variables	PCA number	Extracted components	% of variance	Total of variance	KMO Test Value	Indication of each component
Physicochemical parameters	14	PCA ₁	C1: Geology, Log Cl, Log EC, Log Na, Log SO ₄	25.65	72.74	0.69	Major ions and geological setup
			C2: Aquifer unit, Geology, Log Si	15.18			Aquifer unit
			C3: NH ₄ , Log Eh , Log Alkalinity	11.38			Redox state
			C4: Date, DO	11.07			Oxygen as function of seasonality
			C5: NO ₃ , TOC	9.46			Nitrate correlated with TOC
Sanitary risk factor+latrine data	12	PCA ₂	C1: Q6, Type of well	17.41	63.62	0.51	Deep boreholes mainly from the industries have a fence
			C2: Q1, Q9, Q10	13.34			Unknown explanation
			C3: Num. Latrines , distance latrines	12.44			Presence and distance from latrines
			C4: Q7	10.49			Isolated variable representing a statistical component
			C5: Q2	9.94			Isolated variable representing a statistical component
Physicochemical + <i>E. coli</i>	7	PCA ₃	C1: Aquifer unit , <i>E. coli</i>	21.32	62.44	0.50	Highest presence of <i>E. coli</i> in the shallow aquifer
			C2: Date, DO	21.18			Oxygen as function of seasonality
			C3: log Eh , Na	19.94			Redox state
Sanitary risk + <i>E. coli</i> +latrine data	8	PCA ₄	C1: Q1, Q8 , Type of well, <i>E. coli</i>	26.31	59.55	0.60	<i>E. coli</i> concentration increases with presence of latrines nearby and poor well construction
			C2: Q10, Distance latrines	17.5			Unknown explanation
			C3: Q2, Q7	15.74			Unknown explanation
Physicochemical data + <i>E. coli</i> + Sanitary risk factor +latrine data	10	PCA ₅	C1: Type of well, Aquifer unit, Q8 , <i>E. coli</i>	25.39	69.05	0.63	Main variables related to presence of <i>E. coli</i>
			C2: Log Na, Q1	15.74			Waterpoints located in the coastline have cemented floor
			C3: DO, log Eh, Num. Latrines	15.08			Eh partially depends on DO content
			C4: Q2	12.84			Isolated variable representing a statistical component
PCA ₅ without <i>E. coli</i>	9	PCA _{5.1}	C1: Type of well, Aquifer unit, Q8	23.32	69.99	0.61	Main variables related to presence of <i>E. coli</i>
			C2: Log Na, Q1	17.49			Waterpoints located in the coastline have cemented floor
			C3: DO, log Eh, Num. Latrines	15.26			Eh partially depends on DO content
			C4: Q2	13.92			Isolated variable representing a statistical component

251 ***All type of groundwater points (wells/boreholes)***

252 A first analysis, PCA₁ (see Table 4), was conducted in order to observe which physicochemical variables
253 displayed high correlation and to exclude those which would make the subsequent PCAs (2 to 5)
254 redundant or masked (thus reducing reliability). A significant result from this analysis is that dissolved
255 oxygen changes with seasonality (component #4); this could be attributed to an increment in recharge
256 of oxygenated water, with high DO values, during the wet season. PCA₂ was conducted in order to exclude
257 the sanitary risk factors (from Table 2) that add no significant information in the subsequent analyses.

258 Once the two firsts PCAs were conducted, the most redundant variables were detected, then removed
259 from the list of variables, and additional PCAs were performed adding the variable representing the
260 concentration of *E. coli*. PCA₃ and PCA₄ (Table 4) thus contain the most relevant hydrochemical variables
261 extracted respectively from PCA₁ and PCA₂, plus *E. coli* concentrations. In PCA₄, the first component
262 indicates that *E. coli* concentrations correlate positively with the presence of latrines nearby (Q8), but
263 negatively with the presence of cement floor (Q1, indicating improper construction). Notice that type of
264 well and presence of cement floor were positively correlated, as virtually all handpumps are cemented.

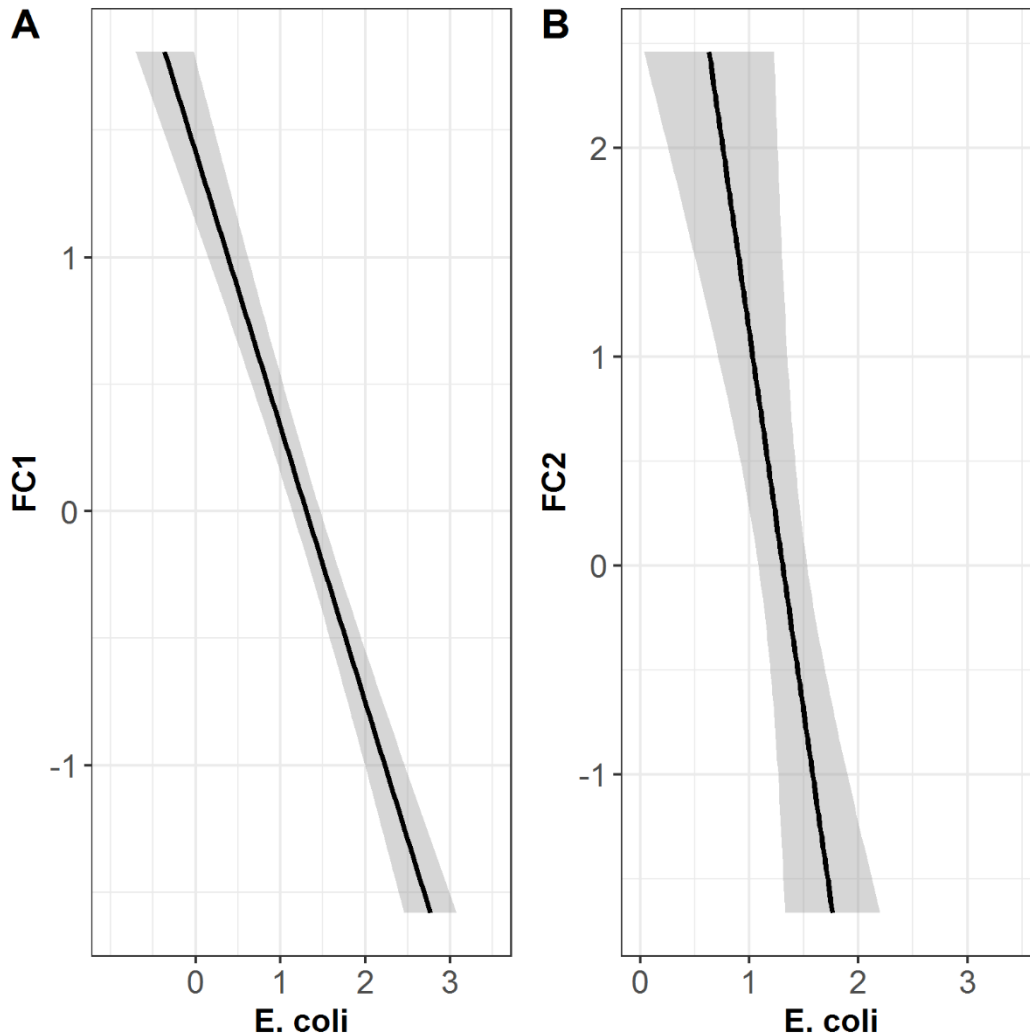
265 The variables most correlated with the presence of *E. coli*, after PCA₃ and PCA₄, were selected to conduct
266 PCA₅. It includes hydrochemical parameters, sanitary risk factors, latrine data and *E. coli* quantification,
267 for a total of ten variables. Results indicate that all deep boreholes have pumps, and that the probability
268 of faecal bacterial pollution increased with the presence of nearby latrines and with uncapped wells.
269 Component #2 merges Na concentration and the presence of cemented floor around the well, as the latter
270 is common in waterpoints situated near the coast line, with sea water intrusion influence. PCA₅ is
271 represented in Figure 3 in a projection on the plane corresponding to variofactors 1 and 2 allowing a
272 graphical depiction of some of the variables.



273

274 *Figure 3. Graphical depiction of the results of PCA₅ projected in variofactor space (VF1 and VF2 axes). Position of*
 275 *samples is scaled for visualization purposes. Size of the points increase with E. coli measurements. Grey arrows*
 276 *represent the contribution of each variable projected into the variofactor plane, so that components can be easily*
 277 *identified.*

278 Once the final correlation between hydrogeological and non-hydrogeological parameters with the
 279 presence of *E. coli* was obtained, a new and final PCA_{5.1} (see Table 4) was performed including the same
 280 variables as PCA₅, except for *E. coli* concentration, that was removed from the set. The main goal was to
 281 assess the variables that most significantly influence the presence of *E. coli*. Afterwards, a generalised
 282 mixed model with Poisson error distribution was performed, including principal component variofactors
 283 as covariates. The covariates affecting significantly the presence of *E. coli* were only C1 (Figure 4a; $\chi^2_1=$
 284 63.379; $p < 0.001$; $\beta = -1.04$) and C2 (Figure 4b; $\chi^2_1= 3.852$; $p = 0.049$; $\beta = -0.19$), while C3 ($\chi^2_1= 2.655$; $p =$
 285 0.103) and C4 ($\chi^2_1= 0.199$; $p = 0.655$) were not significant (p values exceeded 0.05), thus making the
 286 conclusions of PCA₅ even more robust.



287

288 *Figure 4. Significant relation between E. coli and variofactor 1 (A) and variofactor 2 (B) from PCA_{5.1} considering all*
 289 *types of wells. E. coli ranges from safe (0) to unsafe (3) (Table 1). Regarding the PCA_{5.1} results: FC1 is positively*
 290 *correlated with Type of well and Aquifer unit, and negatively with the distance to latrines within 30 m (Q8). FC2 is*
 291 *positively correlated with Na concentration and the extension of the cement floor (Q1).*

292

293 ***Hand-dug wells and Hand-dug wells with handpumps***

294 Another way of reading Figure 3a is by noticing that hand-dug wells (regardless whether they are
 295 equipped with handpumps) are the well types with the highest presence of *E. coli*. This is in agreement
 296 with previous studies (Dayanti et al., 2018; Kilungo et al., 2018; Mzuga et al., 1998; Ugochukwu and Ojike,
 297 2019). In order to find which variables are affecting the presence of *E. coli* in these most polluted points,
 298 five new PCAs were performed now only including data from hand-dug wells. Therefore, we included
 299 here the variables such as groundwater depth (GWL), groundwater column height within the well, and
 300 some specific sanitary risk factors related only to this type of waterpoints. Like in the previous sets of
 301 PCAs, both sampling surveys were included (Table 5).

302
303

Table 5. Components extracted from the second set of PCA analyses (bold indicates negative correlation). For all PCAs the Bartlett sphericity test was significant ($p < 0.001$). The variables involved in each component, the proportion of variance represented by each component, and the measure of sampling adequacy (KMO test value) are included

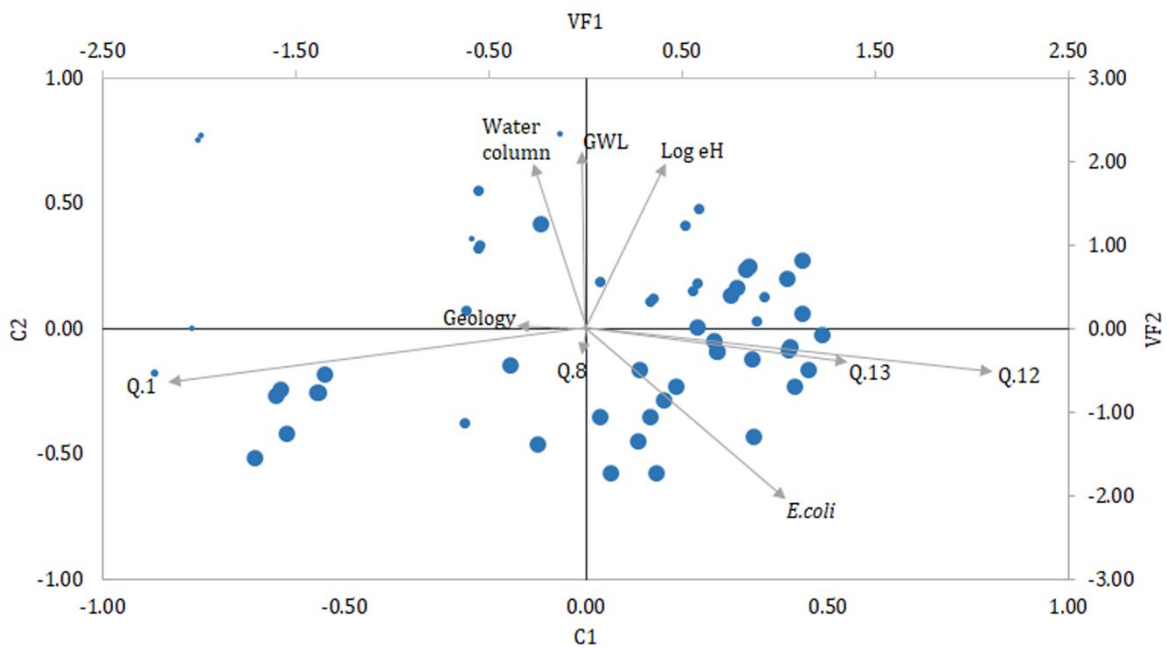
Type of variables	# of variables	PCA number	Extracted components	% of variance	Total variance	KMO Test Value	Indication of each component
Physicochemical parameters	15	PCA _a	C1: Geology, Log EC, Log Na, Log Cl, Log SO ₄	23.02	86.72	0.540	Major ions and geological setup
			C2: Geology, Log EC, Log Alkalinity, GWL	15.32			GWL related to geology and chemical properties
			C3: Date, DO	11.57			Oxygen as function of seasonality
			C4: NO₃ , Log Si	10.42			Relevant minor geochemical species
			C5: GWL, Water Column	9.86			Water levels
			C6: Log Eh, NH₄	8.96			Redox state
			C7: TOC	7.57			Isolated variable representing a statistical component
Sanitary risk data	12	PCA _b	C1: Q6, Num. Latrines	23.02	71.13	0.564	Latrines located inside the main villages in the coast, where animals have no physical access
			C2: Q1, Q3, Q10	15.32			Well construction and maintenance parameters
			C3: Q8, Q12, Q13	11.57			Pollution and sanitary conditions from presence of latrines
			C4: Q11	11.04			Isolated variable representing a statistical component
			C5: Q2, Distance Latrines	10.18			Unknown explanation
Physicochemical + <i>E. coli</i>	8	PCA _c	C1: Log Eh, <i>E. coli</i> , Water Column, DO, TOC, Geology	55.09	72.16	0.805	<i>E. coli</i> concentrations correlate negatively with water columns, redox potential (Eh, DO) and organic carbon
			C2: NO ₃ , GWL	17.07			The largest villages with NO ₃ pollution are located near the coast where the groundwater level is shallow
Sanitary risk + <i>E. coli</i>	9	PCA _d	C1: Q1 , Q12, Q13, <i>E. coli</i>	26.97	69.2	0.573	<i>E. coli</i> positively correlated with unsanitary practices, and inversely with the presence of cement floor
			C2: Q6, Q8	14.4			No animal physical access to latrines located in the main villages.
			C3: Q11	13.97			Isolated variable representing a statistical component
			C4: Q2, Q3	13.86			Sanitary conditions caused by direct water infiltration
	9	PCA _e	C1: Q1 , Q12, Q13	20.64	68.91	0.517	Unsanitary practices correlated inversely to presence of cement floor

Physicochemical + <i>E. coli</i> +Sanitary risk data			C2:Log Eh, <i>E. coli</i> , Water column, GWL	19.45			<i>E. coli</i> inverse correlated with depth to groundwater level, water column and Eh
			C3:Q8, Q13, Water column	15.93			Latrines and scattered waste inside open well
			C4: Geology	12.89			Isolated variable representing a statistical component
PCA _e without <i>E. coli</i>	8	PCA _{e.1}	C1: Q1 , Q12, Q13	24.41	60.71	0.497	Unsanitary practices correlated inversely to presence of cement floor
			C2:Log Eh, Water column, GWL	18.71			<i>E. coli</i> inverse correlated with depth to groundwater level, water column and Eh
			C3:Q8, Water column	17.59			Latrines and scattered waste inside open well

304

305 As in the previous section, the first PCA_a performed (Table 5) was a preliminary screening of variables to
 306 select the ones being significant in terms of information, thus allowing eliminating those that were
 307 redundant or irrelevant. A second analysis was conducted (PCA_b) (Table 5), considering only the sanitary
 308 risk factors from the questionnaire for this particular subset of waterpoints (thus, without the need to
 309 include here the variable “type of well”). Following the scheme reported previously, these two PCAs were
 310 followed by two more where the variable *E. coli* was added. PCA_c included selected hydrochemical
 311 variable and *E. coli*. PCA_d included sanitary risk factors (selected from the results of PCA_b) and the
 312 variable *E. coli*, for a total of nine variables.

313 PCA_e involved nine variables including hydrochemical, selected risk factor variables and presence of *E.*
 314 *coli*. In component two, *E. coli* showed inverse correlation with depth to groundwater level, water column
 315 and Eh. In general, despite the uniformity in the physical and chemical properties in the water column, a
 316 prominent stratification of microbial groups was observed (consistent with Karlov et al., 2008). The
 317 inverse correlation between *E. coli* and the water column suggested preferential presence of faecal
 318 bacteria when the water column was small. Results of PCA_e are represented in Figure 5 as a projection
 319 on the plane corresponding to variofactors 1 and 2.



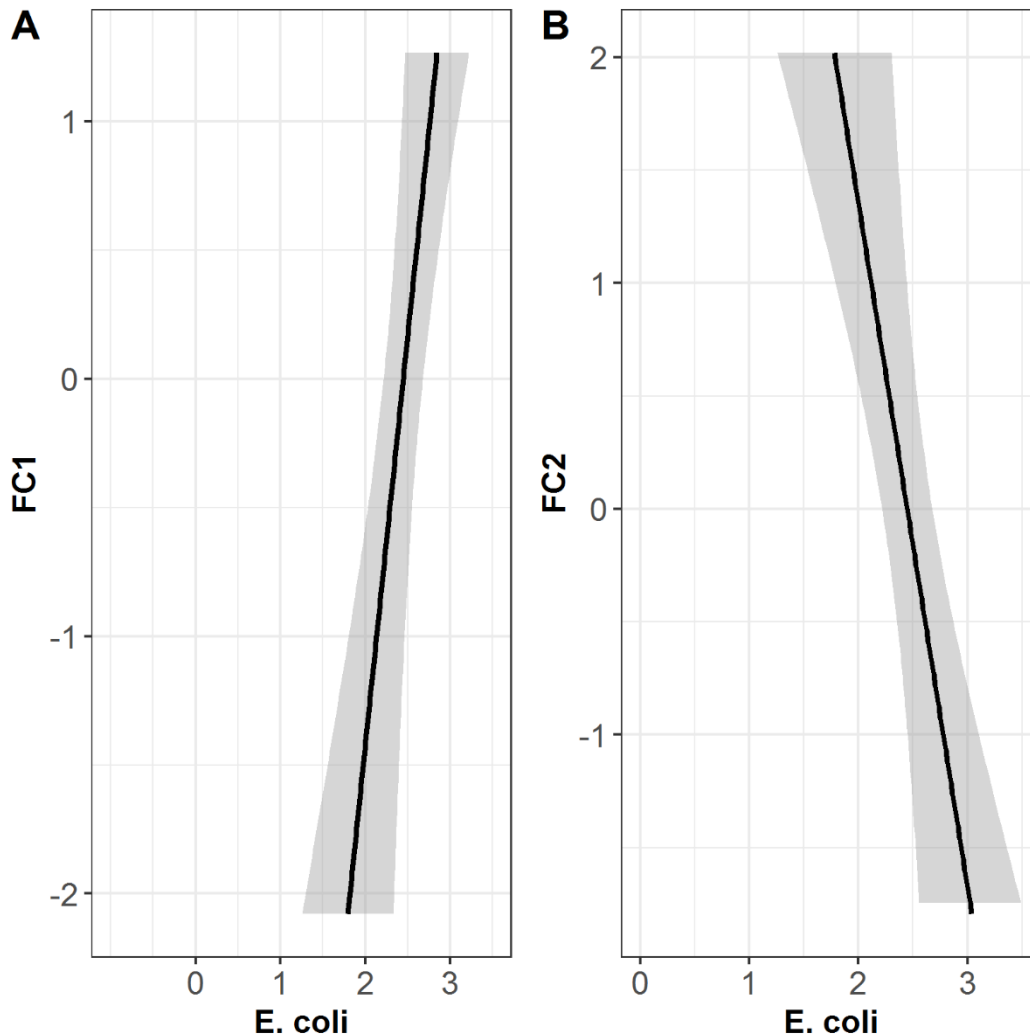
320

321 *Figure 5. Main results of PCA_e. Samples are projected to variofactor space (VF1 and VF2 axes), and position of samples*
 322 *are scaled for visualization purposes. Size of the points increase with *E. coli* measurements. Grey arrows represent the*
 323 *contribution of each variable projected into the variofactor plane, so that some components can be easily identified.*

324

325 Finally, PCA_{e.1} was performed including the same variables as PCA_e, after excluding *E. coli*. In short, the
 326 components obtained were very similar to those from PCA_e, thus indicating robustness in the analysis.

327 From the results of PCA_{e.1}, a generalised mixed model with Poisson error distribution was performed
 328 including principal components variofactors as covariates. The analysis indicates that the covariates
 329 affecting significantly the presence of *E. coli* only in hand-dug wells were C1 (Figure65a; $\chi^2_1= 7.399$; $p =$
 330 0.006 ; $\beta =0.15$) -sanitary issues- and C2 (Figure 6b; $\chi^2_1= 4.496$; $p = 0.033$; $\beta=-0.15$) -redox state related
 331 to GW levels-, while C3 ($\chi^2_1= 1.388$; $p = 0.238$) and subsequent components, were not found significant.



332

333 *Figure 6. Significant relation between E. coli and variofactors V1 (A) and V2 (B) from PCA_{e.1} considering only hand-dug*
 334 *wells and hand-dug wells with handpumps. E. coli ranges from Safe (0) to Unsafe (3). FC1 is positively correlated with*
 335 *the extension of the cement floor (Q1), the cover of the hand-dug well (Q12) and the presence of waste inside the well*
 336 *(Q13). FC2 is positively correlated with log Eh, GWL depth and water column.*

337

338 4. Discussion

339 In the study area, a coastal rural area in South-East Kenia, microbiological pollution levels exceeded the
 340 WHO drinking water quality recommendations in almost all the waterpoints analysed in two campaigns

341 in 2016. We could not find any direct relation of geology to *E. coli* pollution, although in other cases we
342 believe that geology could be a significant factor, as it might drive fast/slow recharge.

343 Most bacteriological problems in groundwater supply points can be associated to improper well design,
344 bad construction, and/or insufficient maintenance practices. Lutterodt et al. (2018) already points out
345 that shallow hand-dug wells have more pollution and sanitary issues as compared to boreholes. Actually,
346 in this study, well type is the primary variable controlling the presence of *E. coli*. Inadequate maintenance
347 of hand pumps, improper sanitation and unhygienic conditions around the waterpoints, are factors that
348 may contribute to faecal contamination, in line with the existing literature (Sukumaran et al., 2015;
349 Ercumen et al., 2017; Godfrey et al., 2006; Lin et al., 2018), since unsanitary covers and litter scattered
350 inside (or around) the well strongly result in the presence of *E. coli* in hand-dug wells, regardless of the
351 presence of handpumps. Furthermore, the extension of the cement floor around the waterpoints and its
352 maintenance state, significantly affects *E. coli* presence, since a small protection by cementation could
353 imply short transit times and direct injection of bacteria through the non-saturated zones.

354 The highest counts of faecal bacteria were observed near human settlements. Unlike other studies that
355 suggest that groundwater faecal pollution is highly variable in a monthly basis (Knappett et al., 2012b;
356 van Geen et al., 2011), influenced by seasonal changes, and being significantly largest during the wet
357 season (Howard et al., 2003; Kayembe et al., 2018), the present study does not show any difference in *E.*
358 *coli* quantification between seasons. This could be explained due to the low precipitation during the wet
359 season in 2016, when the study area was affected by La Niña event, with an estimated 69% reduction in
360 recharge compared to average values (Ferrer et al., 2019). For this reason, more sampling campaigns
361 would be needed in order to study the effect of seasonality on the faecal bacteria in the study area.
362 Actually, it was observed that *E. Coli* concentration values increased with low groundwater levels mostly
363 in the dry season, most probably related to direct input of bacteria (either for well construction or
364 maintenance conditions) into small volumes of water. Future research is needed to understand the actual
365 causality of this correlation, since longer and more recurrent droughts will be expected under future
366 climate change conditions that in sub-Saharan Africa might imply the lowering of groundwater levels,
367 causing a potential cascading effect on water availability and quality.

368 Some geochemical variables displayed a strong correlation with the registered concentrations of *E. coli*.
369 Yet, in some cases it is only due to some external factor that explains both variables together. An example,
370 is Na⁺ concentrations. In the study area, Na⁺ and *E. coli* concentrations display a significant negative
371 correlation, mainly related to low Na⁺ and high *E. coli* concentration values found in the wells located in
372 the Margarini and Kilindini sands. These geological formations show low transit time through the
373 unsaturated zone (Ferrer et al. 2019) and thus less attenuation capacity of the soil, with high *E. coli* counts
374 reaching the shallow aquifer. These observations are in line with Howard et al., (2003), who suggested
375 that fast recharge is the major cause of microbiological contamination, and underpinned that

376 hydrochemical and isotopical data, routinely used to evaluate transit times in aquifer systems, might also
377 be used as indicators of the presence or absence of faecal pollution in other study areas in similar
378 realities. Furthermore, these results are in line with the studies of Leber et al., (2011) and van Geen et al.,
379 (2011), where low permeability layers and large residence times of groundwater were suggested as the
380 cause of the little to none *E. coli* presence.

381 Regarding the risk factors affecting all type of waterpoints, this study confirms that the presence of
382 leaching pit latrines in the vicinity of supply wells is a clear driver of faecal pollution, causing serious
383 concerns for the public, as already shown in Howard et al., 2003; Graham and Polizzotto, 2013; Martínez-
384 Santos et al., 2017; Prüss-Ustün et al., 2016; Schmoll et al., 2006. This effect increases whenever there is
385 a general lack of physical barriers (e.g., concrete) in the latrines between stored excreta and soil and/or
386 groundwater (Van Ryneveld and Fourie, 1997). It is relevant to note that despite the presence of *E. coli*
387 in the study area is correlated to the presence of pit latrines within 30 m from the well, it is not correlated
388 to the actual number of latrines in the vicinity. Thus, it seems that the presence of just one single latrine
389 is enough to cause pollution at the well, while the actual number of latrine just becomes irrelevant.

390 Redox condition shows a positive correlation with dissolved oxygen, number of latrines and *E. coli*
391 concentration. Latrines are an obvious source of oxygenated water with organic matter and bacteria
392 loads and so, despite the presence of organic matter usually leads to a fast depletion of oxygen, they can
393 coexist for short times in particular in heterogeneous soils (see e.g., Freixa et al., 2015). Low values of Eh
394 result in enhanced transport of bacteria in groundwater. *E. coli* is also correlated again to water levels;
395 thick non-saturated zones increase water transit times from the surface to the aquifer, reducing aquifer
396 vulnerability to pollution. As Weldeyohannes et al. (2018) show, the levels of *E. coli* decrease dramatically
397 (below detection limits) when the vadose zone is more than 0.9 m thick. This could be due to the
398 additional mechanisms in the unsaturated zone favouring colloid/bacterial retention at the solid-water
399 interfaces (Sepehrnia et al., 2018a). This effect might counteract that of bacteria increasing with reducing
400 water levels mentioned before; a reduction of the saturated thickness also results in a large retention of
401 faecal bacteria in the unsaturated zone.

402 A management strategy to reduce sanitary risks related with groundwater supply should focus on the
403 correct construction of the wells to improve the isolation of the waterpoints to external sources of
404 pollution. One possible solution would imply drilling of shallow boreholes equipped with handpumps
405 and totally protected on the top, and also using vertical seals consisting of cement or expanding bentonite
406 clay along the annulus between the casing and the borehole. Notice that vertical seals were not explored
407 in this study because of the Kenia reality. Furthermore, despite they are less affordable, drilling deep
408 boreholes seem to be the safest solution, but could result in groundwater in anaerobic conditions, with
409 the need for additional treatment. Well maintenance, protection of waterpoints (preventing water
410 ponding around, and with sanitation coverage implementation), and sanitary practices are a must, and

411 should be emphasized; as a consequence, awareness and sensitization campaigns to eradicate
412 malpractices should be carried out.

413

414 5. Conclusions

415 While the presence of faecal bacteria in domestic supply wells has been acknowledged for decades, no
416 study until the present discriminate and quantify how the combination of hydrogeological and non-
417 hydrogeological parameters correlate with the presence of *E. coli* as a proxy of faecal pollution. Therefore,
418 a number of qualitative and quantitative variables combining geological, hydrological, geochemical,
419 sanitary risk factors, well types, and maintenance variables have been statistical analysed for
420 correlations with *E. coli* concentrations in a coastal area of Sub-Saharan Africa, with high presence of
421 faecal bacteria in the groundwater used to supply the population.

422 This study demonstrates that including in a PCA such an interdisciplinary set of variables can be a useful
423 methodology to obtain precise information of the relations between different types of variables, most
424 times separated in analysis (e.g., in modelling efforts). Furthermore, this study goes a step forward when
425 trying to assess which variables are related to faecal bacteria pollution; that is, including PCA variofactors
426 as a covariate in mixed models might become a useful tool to assess the factors influencing significantly
427 the presence of pathogenic organisms. Despite the geological formation itself did not show a direct
428 relation with *E. coli* pollution, some hydrogeologically related parameters and variables (flow velocity,
429 redox condition, water column, etc) were found significant in the analyses. Therefore, geology and
430 hydrogeology can be combined when explaining risk pollution in shallow aquifer wells.

431 This methodology has confirmed in a quantitative way that the well constructive characteristics are most
432 important to avoid the presence of pathogenic bacteria in groundwater. Extended cement floor would
433 reduce the presence of faecal bacteria pollution, being more important in those areas where the water
434 infiltrates fast through the unsaturated zone. Furthermore, knowing the geochemical elements,
435 indicators of transit time, and groundwater depth, could just be simple and good indicators of the
436 presence of faecal bacteria. Hence, easy to identify and measure physical and geochemical
437 measurements, such as water column and Eh, may be used to assess a priori faecal pollution. Actually, Eh
438 can be related to the presence of input water with high organic matter load (indicative of the presence of
439 latrines nearby), while variations in the water column are driven by climate and well operation
440 conditions.

441

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447

448 **Bibliography**

449 Adelana, S.M.A., MacDonald, A.M., 2008. Groundwater research issues in Africa. IAH Sel. Pap. Hydrogeol.
450 13, 1–7.

451 Aquagenx, 2015. Compartment Bag Test (CBT) Instructions for Use: Drinking Water. [WWW Document].
452 URL www.aquagenx.com

453 Bain, R., Cronk, R., Hossain, R., Bonjour, S., Onda, K., Wright, J., Yang, H., Slaymaker, T., Hunter, P., Prüss-
454 Ustün, A., Bartram, J., 2014. Global assessment of exposure to faecal contamination through drinking
455 water based on a systematic review. *Trop. Med. Int. Heal.* 19, 917–927.

456 Barba, C., Folch, A., Gaju, N., Sanchez-Vila, X., Carrasquilla, M., Grau-Martínez, A., Martínez-Alonso, M.,
457 2019a. Microbial community changes induced by Managed Aquifer Recharge activities: linking
458 hydrogeological and biological processes. *Hydrol. Earth Syst. Sci* 23, 139–154.

459 Barba, C., Folch, A., Sanchez-Vila, X., Martínez-Alonso, M., Gaju, N., 2019b. Are dominant microbial sub-
460 surface communities affected by water quality and soil characteristics? *J. Environ. Manage.* 237, 332–
461 343.

462 Bates, D., Mächler, M., Bolker, B., Walker, S., 2015. Fitting Linear Mixed-Effects Models Using lme4. *J. Stat.*
463 *Softw.* 67, 1–48.

464 Bhattacharjee, S., Ryan, J.N., Elimelech, M., 2002. Virus transport in physically and geochemically
465 heterogeneous subsurface porous media. *J. Contam. Hydrol.* 57, 161–187.

466 Blaschke, A.P., Derx, J., Zessner, M., Kirnbauer, R., Kavka, G., Strelec, H., Farnleitner, A.H., Pang, L., 2016.
467 Setback distances between small biological wastewater treatment systems and drinking water wells
468 against virus contamination in alluvial aquifers. *Sci. Total Environ.* 573, 278–289.

469 Broström, G., Holmberg, H., 2011. Generalized linear models with clustered data: Fixed and random
470 effects models. *Comput. Stat. Data Anal.* 55, 3123–3134.

471 Carles-Brangari, A., Sanchez-Vila, X., Freixa, A., Romani, A., Rubol, S., Fernandez, D., 2017. A mechanistic
472 model (BCC-PSSICO) to predict changes in the hydraulic properties for bio-amended variably saturated
473 soils. *Water Resources Research*, 53(1), 93-109.

474 Carles-Brangari, A., Fernandez, D., Sanchez-Vila, X., Manzoni, S., 2018. Ecological and soil hydraulic
475 implications of microbial responses to stress: a modeling analysis. *Advances in Water Resources*, 116,
476 178-194.

477 Charles, K.J., Souter, F.C., Baker, D.L., Davies, C.M., Schijven, J.F., Roser, D.J., Deere, D.A., Priscott, P.K.,
478 Ashbolt, N.J., 2008. Fate and transport of viruses during sewage treatment in a mound system. *Water Res.*
479 42, 3047–3056.

480 CWSB, 2013. Coastal Water Services Board-Water Point Mapping Report: Kwale County [WWW
481 Document]. URL www.cwsb.go.ke

482 Dayanti, M.P., Fachrul, M.F., Wijayanti, A., 2018. *Escherichia coli* as bioindicator of the groundwater
483 quality in Palmerah District, West Jakarta, Indonesia, in: IOP Conference Series: Earth and Environmental
484 Science. 106, 1-7.

485 Devane, M.L., Weaver, L., Singh, S.K., Gilpin, B.J., 2018. Fecal source tracking methods to elucidate critical
486 sources of pathogens and contaminant microbial transport through New Zealand agricultural
487 watersheds – A review. *J. Environ. Manage.* 222, 293–303.

488 Elangovan, N.S., Lavanya, V., Arunthathi, S., 2018. Assessment of groundwater contamination in a
489 suburban area of Chennai, Tamil Nadu, India. *Environ. Dev. Sustain.* 20, 2609–2621.

490 Ferguson, A.S., Layton, A.C., Mailloux, B.J., Culligan, P.J., Williams, D.E., Smartt, A.E., Sayler, G.S., Feighery,
491 J., McKay, L.D., Knappett, P.S.K., Alexandrova, E., Arbit, T., Emch, M., Escamilla, V., Ahmed, K.M., Alam, M.J.,
492 Streatfield, P.K., Yunus, M., van Geen, A., 2012a. Comparison of fecal indicators with pathogenic bacteria
493 and rotavirus in groundwater. *Sci. Total Environ.* 431, 314–322.

494 Ferguson, A.S., Layton, A.C., Mailloux, B.J., Culligan, P.J., Williams, D.E., Smartt, A.E., Sayler, G.S., Feighery,
495 J., McKay, L.D., Knappett, P.S.K., Alexandrova, E., Arbit, T., Emch, M., Escamilla, V., Ahmed, K.M., Alam, M.J.,
496 Streatfield, P.K., Yunus, M., van Geen, A., 2012b. Comparison of fecal indicators with pathogenic bacteria
497 and rotavirus in groundwater. *Sci. Total Environ.* 431, 314–322.

498 Ferrer, N., Folch, A., Lane, M., Olago, D., Odida, J., Custodio, E., 2019. Groundwater hydrodynamics of an
499 Eastern Africa coastal aquifer, including La Niña 2016–17 drought. *Sci. Total Environ.* 661, 575–597.

500 Foster, T., Willetts, J., 2018. Multiple water source use in rural Vanuatu: are households choosing the
501 safest option for drinking? *Int. J. Environ. Health Res.* 28, 579–589.

502 Freixa, A., Rubol, S., Carles, A., Fernandez, D., Butturini, A., Sanchez-Vila, X., Romani, A., 2015. The effects
503 of sediment depth and oxygen concentration on the use of organic matter: An experimental study using
504 an infiltration sediment tank. *Science of the total environment*, 540, 20-31.

505 Goyal, S.M., Keswick, B.H., Gerba, C.P., 1984. Viruses in groundwater beneath sewage irrigated cropland.
506 Water Res. 18, 299–302.

507 Graham, J.P., Polizzotto, M.L., 2013. Pit Latrines and Their Impacts on Groundwater Quality: A Systematic
508 Review. Environ. Health Perspect. 121, 521–530.

509 Gronewold, A.D., Sobsey, M.D., McMahan, L., 2017. The compartment bag test (CBT) for enumerating fecal
510 indicator bacteria: Basis for design and interpretation of results. Sci. Total Environ. 587–588, 102–107.

511 Hair, J., Anderson, R., Tatham, R., Black, W., 1995. Multivariate Data Analysis, 4th ed, Technometrics.
512 Prentice-Hall Inc, New Jersey.

513 Howard, G., Pedley, S., Barrett, M., Nalubega, M., Johal, K., 2003. Risk factors contributing to
514 microbiological contamination of shallow groundwater in Kampala, Uganda. Water Res. 37, 3421–3429.

515 Howard, G., Pedley, S., Barrett, M., Nalubega, M., Johal, K., 2003. Risk factors contributing to
516 microbiological contamination of shallow groundwater in Kampala, Uganda. Water Res. 37, 3421–3429.

517 Karlov, D.S., Marie, D., Danil, •, Sumbatyan, A., Chuvochina, M.S., Kulichevskaya, I.S., Alekhina, I.A., Sergey,
518 •, Bulat, A., 2008. Microbial communities within the water column of freshwater Lake Radok, East
519 Antarctica: predominant 16S rDNA phlotypes and bacterial cultures. Polar Biol. 40.

520 Kayembe, J.M., Thevenon, F., Laffite, A., Sivalingam, P., Ngelinkoto, P., Mulaji, C.K., Otamonga, J.P., Mubedi,
521 J.I., Poté, J., 2018. High levels of faecal contamination in drinking groundwater and recreational water due
522 to poor sanitation, in the sub-rural neighbourhoods of Kinshasa, Democratic Republic of the Congo. Int.
523 J. Hyg. Environ. Health 221, 400–408.

524 Kilungo, A., Powers, L., Arnold, N., Whelan, K., Paterson, K., Young, D., 2018. Evaluation of well designs to
525 improve access to safe and clean water in rural Tanzania. Int. J. Environ. Res. Public Health 15, 1–11.

526 Knappett, P.S.K., Emelko, M.B., Zhuang, J., McKay, L.D., 2008. Transport and retention of a bacteriophage
527 and microspheres in saturated, angular porous media: Effects of ionic strength and grain size. Water Res.
528 42, 4368–4378.

529 Knappett, P.S.K., McKay, L.D., Layton, A., Williams, D.E., Alam, M.J., Huq, M.R., Mey, J., Feighery, J.E.,
530 Culligan, P.J., Mailloux, B.J., Zhuang, J., Escamilla, V., Emch, M., Perfect, E., Sayler, G.S., Ahmed, K.M., Van
531 Geen, A., 2012a. Implications of fecal bacteria input from latrine-polluted ponds for wells in sandy
532 aquifers. Environ. Sci. Technol. 46, 1361–1370.

533 Knappett, P.S.K., Mckay, L.D., Layton, A., Williams, D.E., Alam, M.J., Mailloux, B.J., Ferguson, A.S., Culligan,
534 P.J., Serre, M.L., Emch, M., Ahmed, K.M., Sayler, G.S., Geen, A. Van, 2012b. Unsealed tubewells lead to
535 increased fecal contamination of drinking water. J. Water Health 10, 565–578.

536 Leber, J., Rahman, M.M., Ahmed, K.M., Mailloux, B., van Geen, A., 2011. Contrasting Influence of Geology
537 on E. coli and Arsenic in Aquifers of Bangladesh. *Ground Water* 49, 111–123.

538 MacDonald, A.M., Bonsor, H.C., Dochartaigh, B.É.Ó., Taylor, R.G., 2012. Quantitative maps of groundwater
539 resources in Africa. *Environ. Res. Lett.* 7, 1–7.

540 Macler, B.A., Merkle, J.C., 2000. Current knowledge on groundwater microbial pathogens and their
541 control, *Hydrogeology Journal*. 8, 29-40.

542 Martínez-Santos, P., Martín-Loeches, M., García-Castro, N., Solera, D., Díaz-Alcaide, S., Montero, E., García-
543 Rincón, J., 2017. A survey of domestic wells and pit latrines in rural settlements of Mali: Implications of
544 on-site sanitation on the quality of water supplies. *Int. J. Hyg. Environ. Health* 220, 1179–1189.

545 Matthess, G., Pekdeger, A., Schroeter, J., 1988. Persistence and transport of bacteria and viruses in
546 groundwater - a conceptual evaluation. *J. Contam. Hydrol.* 2, 171–188.

547 Mzuga, J.M., Tole, M.P., Ucakuwun, E.K., 1998. The impact of geology and pit latrines on groundwater
548 quality in Kwale District, Dunes, groundwater, mangroves and birdlife in coastal Kenya. Chapter 6, 85-96

549 Nowicki, S., Lapworth, D.J., Ward, J.S.T., Thomson, P., Charles, K., 2019. Tryptophan-like fluorescence as a
550 measure of microbial contamination risk in groundwater. *Sci. Total Environ.* 646, 782–791.

551 Olajuyigbe, A.E., Olamiju, I.O., Ola-Omole, C.M., 2017. Vulnerability of hand-dug wells in the core area of
552 Akure, Nigeria. *Urban Water J.* 14, 797–803.

553 Oteng-Peprah, M., de Vries, N.K., Acheampong, M.A., 2018. Greywater characterization and generation
554 rates in a peri urban municipality of a developing country. *J. Environ. Manage.* 206, 498–506.

555 Paliy, O., Shankar, V., 2016. Application of multivariate statistical techniques in microbial ecology. *Mol.*
556 *Ecol.* 25, 1032–1057.

557 Perujo, N., Sanchez-Vila, X., Proia, L., Romani, A., 2017. Interaction between physical heterogeneity and
558 microbial processes in subsurface sediments: a laboratory-scale column experiment. *Environmental*
559 *Science and Technology*", 51 (11), 6110-6119.

560 Rao, V.C., Metcalf, T.G., Melnick, J.L., 1986. Articles in the Update series Human viruses in sediments,
561 sludges, and soils*, *Bulletin of the World Health Organization*.

562 Prüss-Ustün, A., Wolf, J., Corvalán, C., Bos, R., Neira, M., 2016. Global Burden of Diseases From
563 Environmental Risks.

564 Rohmah, Y., Rinanti, A., Hendrawan, D.I., 2018. The determination of ground water quality based on the
565 presence of Escherichia coli on populated area (a case study: Pasar Minggu, South Jakarta). *IOP Conf. Ser.*
566 *Earth Environ. Sci.* 106.

567 Saiers, J.E., Ryan, J.N., 2005. Colloid deposition on non-ideal porous media: The influences of collector
568 shape and roughness on the single-collector efficiency. *Geophys. Res. Lett.* 32, 1–5.

569 Schmoll, O., Howard, G., Chilton, J., Chorus, I., 2006. Protecting Groundwater for Health: Managing the
570 Quality of Drinking-water Sources, *Protecting Groundwater for Health: Managing the Quality of Drinking-*
571 *water Sources.*

572 Sepehrnia, N., Bachmann, J., Hajabbasi, M., Afyuni, M., Horn, M., 2018a. Modeling *Escherichia coli* and
573 *Rhodococcus erythropolis* transport through wetttable and water repellent porous media. *Colloids*
574 *Surfaces B Biointerfaces* 172, 280–287.

575 Sepehrnia, N., Memarianfard, L., Moosavi, A.A., Bachmann, J., Rezanezhad, F., Sepehri, M., 2018b.
576 Retention modes of manure-fecal coliforms in soil under saturated hydraulic condition. *J. Environ.*
577 *Manage.* 227, 209–215.

578 Sharma, P.K., Srivastava, R., 2011. Numerical analysis of virus transport through heterogeneous porous
579 media. *J. Hydro-Environment Res.* 5, 93–99.

580 Stauber, C., Miller, C., Cantrell, B., Kroell, K., 2014. Evaluation of the compartment bag test for the
581 detection of *Escherichia coli* in water. *J. Microbiol. Methods* 99, 66–70.

582 Tabachnik, B., Fidell, L., 2007. *Using multivariate statistics.* Pearson Education Inc, Boston, MC.

583 Team, 2018. *R: A Language and Environment for Statistical Computing.*

584 Thompson, B., 2004. *Exploratory and Confirmatory Factor Analysis: Understanding Concepts and*
585 *Applications.*

586 Tole, M.P., 1997. Pollution of groundwater in the coastal Kwale Distric, Kenya. *Sustain. Water Resour.*
587 *under Increasing Uncertain.* 287–297.

588 Ugochukwu, U.C., Ojike, C., 2019. Assessment of the groundwater quality of a highly populated district in
589 Enugu State of Nigeria. *Environ. Dev. Sustain. Online ISSN* 1573-2975.

590 Van Geen, A., Ahmed, K.M., Akita, Y., Alam, M.J., Culligan, P.J., Emch, M., Escamilla, V., Feighery, J., Ferguson,
591 A.S., Knappett, P., Layton, A.C., Mailloux, B.J., McKay, L.D., Mey, J.L., Serre, M.L., Streatfield, P.K., Wu, J.,
592 Yunus, M., 2011. Fecal contamination of shallow tubewells in Bangladesh inversely related to arsenic.
593 *Environ. Sci. Technol.* 45, 1199–1205.

594 Van Ryneveld, M., Fourie, A., 1997. A strategy for evaluating the environmental impact of on-site
595 sanitation systems. *Water SA* 23, 279–291.

596 Weldeyohannes, A.O., Kachanoski, G., Dyck, M., 2018. Wastewater Flow and Pathogen Transport from At-
597 Grade Line Sources to Shallow Groundwater. *J. Environ. Qual.* 47, 1051.

598 Wright, J.A., Cronin, A., Okotto-Okotto, J., Yang, H., Pedley, S., Gundry, S.W., 2013. A spatial analysis of pit
 599 latrine density and groundwater source contamination. Environ. Monit. Assess. 185, 4261–4272.

600 Yates, M. V, Gerba, C.P., Kelley, L.M., 1985. Virus persistence in groundwater. Appl. Environ. Microbiol. 49,
 601 778–781.

602

603 **Supplementary material**

604 *Table 1S. E. coli quantification results from CBT in March 2016. Green colour means safe, yellow means*
 605 *intermediate risk, orange means high risk and red means unsafe.*

Code	Aquagenx (bags) MPN/100 ml	Code	Aquagenx (bags) MPN/100 ml	Code	Aquagenx (bags) MPN/100 ml
Footprints School	0,0	A/14/10	0,0	Z2-112	48,3
Z4-11	48,3	Z3-87	1,5	Z1-140	0,0
Z4-09	>100	Z3-98	>100	Z2-104	2,6
Z4-01	>100	Z3-90	>100	Z1-110	>100
A/04/12	0,0	A/05/11	>100	DB/FI/HP	0,0
Z4-18	>100	HOTSPRING	0,0	Z3-96	>100
A/06/12	1,2	C108HWL	>100	E/29/01	13,6
Z4-78B	>100	3KD01	48,3	A/09/11	0,0
Z4-08	48,3	S1-3KD06	>100	MIVUMONI	0,0
Z4-06	>100	GD31	0,0	C/15/10	0,0
D/100/16	13,6	MUK DAM	>100	C/109/21	0,0
Z4-04	>100	MUK DWS	>100	C/12/12	0,0
Z4-MS	>100	Z1-122	13,6	C/06/12	0,0
D/82/14	0,0	Z1-125	>100	C/19/10	0,0
Z4-85	>100	Z1-124	>100	D/129/19	0,0
Z4-24	>100	D/16/10	1,2	DB/MH/CO	0,0
Z3-25	>100	Z1-121B	>100	Z1-141	>100
D/63/13	0,0	Z1-116	13,6	UK-WL	0,0
D/68/13	0,0	C/07/09	0,0	A/06/13	0,0
Z3-30	48,3	A/01/11	0,0	D/103/16	0,0
Z3-29	13,6	Z2-103	>100	LUKORE-SH	48,3
DB/BM/HP	0,0	D/203/27	1,1	Z1-118	>100
BH310	13,6	DB/MS/LST	0,0	VIN-WL	0,0
BH402	0,0	Z1-135	>100	Base_BH_1	0,0
NK-03	0,0	DB/KI/ST	0,0	Base_BH_3	0,0

Z1-70	>100	Z1-33	>100	Base_BH_7	0,0
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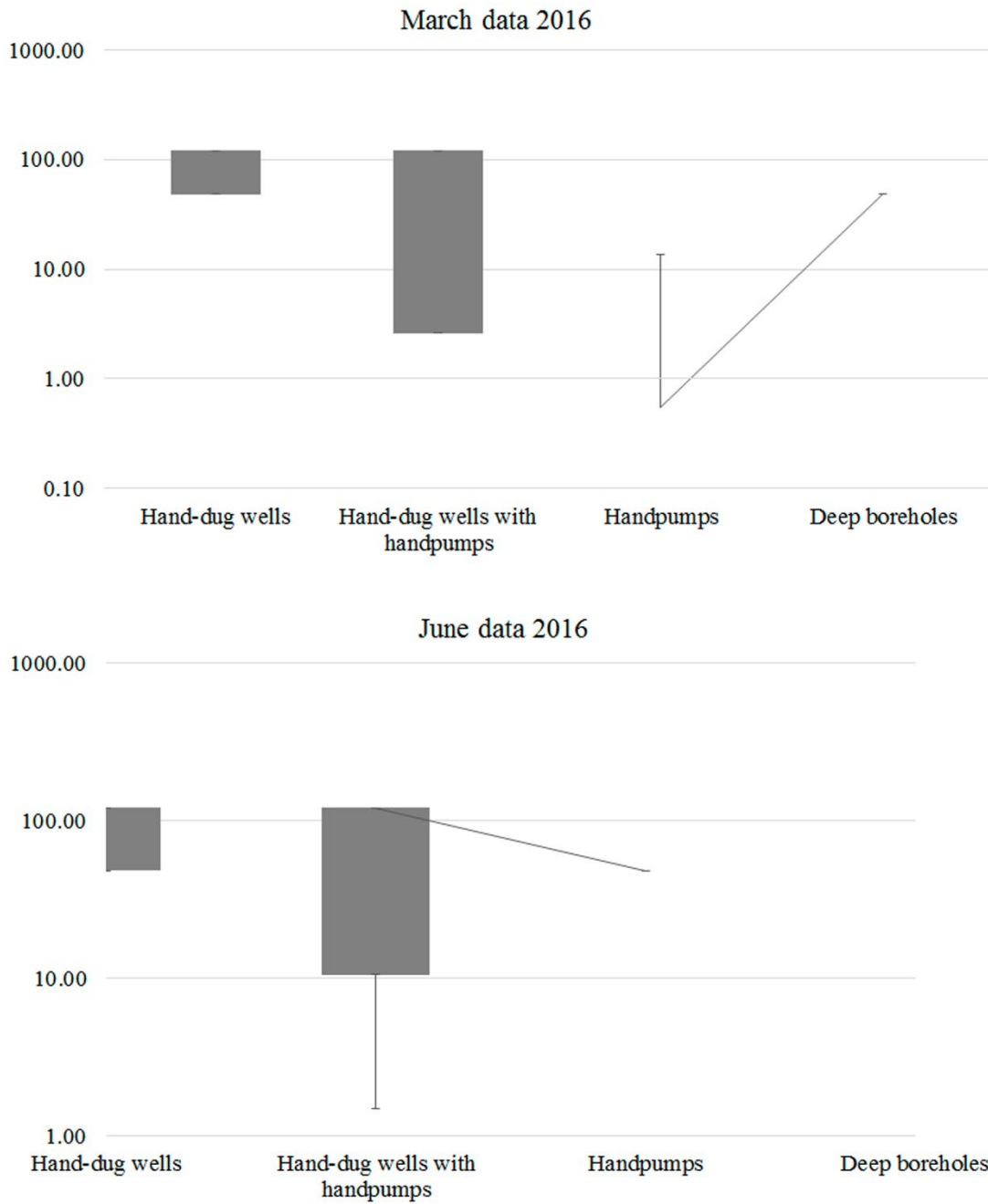
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609 *Table 2S. E. coli quantification results from CBT in June 2016. Green colour means safe, yellow means*
610 *intermediate risk, orange means high risk and red means unsafe.*

Code	Aquagenx (bags) <i>MPN/100 ml</i>	Code	Aquagenx (bags) <i>MPN/100 ml</i>	Code	Aquagenx (bags) <i>MPN/100 ml</i>
Footprints School	0,0	A/05/11	>100	Z1-110	>100
Z4-11	48,3	HOTSPRING	0,0	DB/FI/HP	0,0
Z4-01	>100	C108HWL	>100	Z3-96	48,3
A/04/12	0,0	3KD01	48,3	E/29/01	9,6
Z4-18	48,3	MUACHEMA	>100	A/09/11	0,0
A/06/12	0,0	S1-3KD06	>100	MIVUMONI	0,0
Z4-78B	>100	GD31	0,0	C/15/10	0,0
Z4-08	>100	MUK DAM	>100	C/109/21	0,0
Z4-06	>100	MUK DWS	>100	C/12/12	0,0
D/100/16	48,3	Z1-122	13,6	C/06/12	0,0
Z4-04	>100	Z1-125	>100	C/19/10	0,0
Z4-MS	48,3	Z1-124	>100	D/129/19	0,0
D/82/14	0,0	D/16/10	0,0	DB/MH/CO	0,0
Z4-85	48,3	Z1-121B	>100	Z1-141	>100
Z4-24	>100	Z1-116	13,6	UK-WL	0,0
D/63/13	0,0	C/07/09	0,0	D/103/16	0,0
D/68/13	0,0	A/01/11	0,0	LUKORE- SH	0,0
Z3-30	>100	Z2-103	>100	Z1-118	>100
Z3-29	>100	D/203/27	13,6	VIN-WL	0,0
DB/BM/HP	0,0	DB/MS/LST	0,0	Base_BH_3	0,0
BH310	0,0	Z1-135	>100	Base_BH_7	0,0
Z1-70	>100	Z2-112	48,3	DB/KI/ST	0,0
Z1-33	48,3	Z1-140	1,5	Z3-102B	13,6
A/14/10	0,0	Z2-104	4,7	BH302	0,0
Z3-87	>100	Z3-98	>100	C/05/09	48,3
Z3-90	13,6	C/03/09	0,0		



611

612 *Figure SM. E. coli presence for each type of each water point in each field survey.*