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Research Article

Disruption of Lipid Profile and Alteration of Hepatic Lipoprotein Metabolism Gene Expression in Anaemia-induced Rat

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Abstract

Background and Objective: Iron metabolism in animals is altered by haemolytic anaemia induced by phenylhydrazine (PHZ), however, its effects on lipid metabolism remains elusive. The aim of this study was to examine the impact of anaemia on lipid profiles and lipoprotein metabolism gene expression in rats. **Materials and Methods:** Fourteen adult male Wistar rats were randomly classified into normal control and anaemia-induced group ($n = 7$), respectively. Anaemia was induced in rats by daily administration of PHZ at 10 mg kg^{-1} for 8 consecutive days, after which blood was collected and liver excised. Lipid profiles of plasma and liver were determined spectrophotometrically while the expression of genes associated with lipid and lipoprotein metabolism was assayed by reverse transcriptase polymerase chain reaction. **Results:** The induced-anaemia resulted in hypotriglyceridemia and hypophospholipidosis, with concurrent hypercholesteromia compared to control, respectively. Liver triglycerides, phospholipids, cholesterol were observed to be up-regulated. Anaemic rats showed a significant ($p < 0.05$) up-regulation of the relative expression of hepatic lecithin-cholesterol acyltransferase (Lcat), paraoxonase-1 (Pon-1), aryl hydrocarbon receptor (Ahr), 3-hydroxy-3-methylglutaryl-CoA reductase (Hmgcr) and down-regulation of Scavenger Receptor Class B Type I (Scarb1). **Conclusion:** The induced-anaemia alter the expression of lipoprotein metabolizing genes which might be the underlying mechanism of anaemia to disrupt lipid metabolism.

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INTRODUCTION

Anaemia, known as the reduction of oxygen carrying ability of the blood, is a common blood disorder affecting people of all ages and posing a great global healthcare^{1,2}. It can be due to failure of red cell proliferation, defective maturation of red blood cells, blood loss and haemolysis^{1,3}. Haemolytic anaemia is the accelerated destruction of mature red cells outside the bone marrow or a consequence of the destruction of imperfectly formed red cells^{1,4}. Hemolytic conditions with substantial intravascular hemolysis include paroxysmal nocturnal hemoglobinuria (PNH), sickle-cell disease (SCD), thalassemias, hereditary spherocytosis and drug or chemical-induced anaemia^{5,6}. The chemicals or drugs can caused haemolysis by interacting with sulfhydryl groups, inhibit various enzymes, disrupt immune mechanisms and fragment erythrocytes as they pass through the platelet-fibrin mesh⁷. Phenylhydrazine (PHZ) is an antipyretic drug that is well known for its ability to produce hemolysis in rats and humans leading to the formation of meta-haemoglobin which contributed to the oxidation of oxyhemoglobin⁶⁻⁹. Report shows that PHZ induced haemolytic anaemia in rats based on hepatic changes in the expression of genes that are mechanistically linked to haematotoxicity⁷. It has been suggested that PHZ induces haemolytic anaemia as a consequence of peroxidation of membrane lipids^{10,11}.

Cholesterol, triglycerides, phospholipids and fatty acids are lipids molecules that play key roles in metabolic pathways which are transported in the blood as lipoproteins¹²⁻¹⁴. Disturbances in the homeostasis of these lipids and lipoprotein resulted in dyslipidemia¹⁵⁻¹⁹ which has effects on health, thus great attention is paid to abnormal levels of lipids and its associated factors²⁰⁻²². The present study was undertaken to investigate the effect of PHZ-induced anaemia on the lipid profile and expression of lipoprotein metabolism genes in a rat model.

MATERIALS AND METHODS

Acclimatization of experimental animals: Ten weeks old inbred male albino rats (n = 14) weighing between 100 and 150 g were housed in clean cages. The rats were acclimatize to their environment for one week under standard 12 h light and dark cycles with free access to feed and clean tap water ad libitum before the experiment. The study was approved by the ad hoc Animal Ethical Committee of the Department of Biochemistry, Lagos State University, Ojo-Lagos, Nigeria and conducted in accordance with the ethical norms guiding principles of laboratory animal care²³.

Treatment protocol and tissue collection: The rats were randomly distributed into two groups of seven rats each. About 10 mg kg⁻¹ b.wt., PHZ (Sigma-Aldrich Chemical Company, St Louis, MO, USA) was administered daily by oral gavage for 8 consecutive days²⁴ while the control rats were allowed free access to water. The rats were sacrificed on the 9th day, after an overnight fast, under ketamine anaesthesia and blood collected by cardiac puncture. Blood and liver were processed as previously described by Ogunrinola²⁵ and Rotimi et al. ²⁶ while a portion of the left lobe was preserved in RNAlater[®].

Biochemical analysis: Total plasma cholesterol and triglyceride concentrations were determined using commercially available kits according to the manufacturer's instructions. Plasma phospholipids was determined as described by Rifai et al. ²⁷. Lipids were extracted from the liver according to the method of Folch et al.²⁸ and the cholesterol, triglyceride and phospholipid determination was performed by the methods earlier described by Ogunrinola et al.²⁹ and Rotimi et al.³⁰, respectively.

Ribonucleic acid (RNA) extraction: The RNA was extracted from RNAlater[®]-stabilized liver using the Aidlab spin column RNA extraction kit according to the instructions of the manufacturer. Concentration and purity of extracted RNA was determined at 260 and 280 nm using a NanoDrop[®] 2000 spectrophotometer (Thermo Scientific). The RNA samples were kept at -80°C until gene expression analysis.

Expression of hepatic lipid metabolizing genes: The levels of expression of five lipid metabolizing genes (lecithin-cholesterol acyltransferase (Lcat), paraoxonase-1 (Pon-1), aryl hydrocarbon receptor (Ahr), 3-hydroxy-3-methylglutaryl-CoA reductase (Hmgcr) and scavenger receptor class B Type I (Scarb1)) were assessed in the liver using semi-quantitative reverse transcriptase polymerase chain reaction (RT-PCR) as previously described by Rotimi et al.³¹ using gene specific primers (GSP), which are designed based on known sequences of the target RNA (Table 1). The intensity of the migrated RNAs bands was analysed using Image J software³². Results were presented as relative expression of each gene in comparison with a housekeeping (β -actin, Actb) gene (ratio of intensity of each gene to that of β -actin, Actb).

Statistical analysis: All statistical analyses were performed using IBM SPSS[®] version 20.0 statistical software (IBM Corp., Armonk, NY, USA). Data were expressed as Mean \pm SEM of five replicates.

Table 1: Sequences of gene-specific primers

Analysis of variance (ANOVA) was carried out to test for level of homogeneity at $p < 0.05$ among the groups.

RESULTS AND DISCUSSION

Lipid profiles results indicates that induced-anaemia with PHZ significantly increased plasma cholesterol (Fig. 1a) and down-regulate triglyceride (Fig. 1b) and phospholipid (Fig. 1c). However, the levels of hepatic cholesterol (Fig. 2a), triglyceride (Fig. 2b) and phospholipid (Fig. 2c) were significantly ($p < 0.05$) increased in the anaemia-induced group compared to control group.

The expression of five genes was assessed in liver samples from all treatment groups. The relative expression of hepatic aryl hydrocarbon receptor (Ahr) (Fig. 3a), lecithin-cholesterol acyltransferase (Lcat) (Fig. 3b), paraoxonase-1 (Pon-1) (Fig. 3c) and 3-hydroxy-3-methylglutaryl-CoA reductase (Hmgcr) (Fig. 3d) were significantly increased in anaemic-induced rats corresponding to 38.29, 5.26, 6.48 and 29.80%, respectively while hepatic scavenger receptor class B type I (Scarb1) (Fig. 3e) was significantly ($p = 0.05$) down-regulated.

Lipid and lipoprotein abnormalities have been shown to be the predictors for the metabolic disturbances, including dyslipidemia, hypertension, diabetes, cardiovascular disease and liver dysfunction^{29,33}. Abnormal lipid homeostasis has been reported in haematological disorders such as sickle cell anaemia which alter membrane fluidity and function of red blood cell^{34,35}. These finding of elevated plasma cholesterol concentrations was in agreement with that of Mazandarani and Hoseini³⁶.

Fig. 1(a-c):

Effects of PHZ induced-anaemia on plasma lipid profiles (a) Cholesterol, (b) Triglyceride and (c) Phospholipid

Bars represent the Means \pm SEM (n = 7). Bars with different alphabets are significantly ($p < 0.05$) different from each other

Fig. 2(a-c): Effect of PHZ induced-anaemia on hepatic lipid profiles (a) Cholesterol, (b) Triglycerides and (c) Phospholipid

Bars represent the Means \pm SEM (n = 7). Bars with different alphabets are significantly ($p < 0.05$) different from each other

This may be due to the concentration of red blood cell that is altered by cholesterol synthesis or its displacement from tissue to plasma¹⁶ and can lead to abnormal cholesterol loading of the erythrocyte membrane in hemolytic anaemia patients with liver disease³⁷. This study revealed hepatic accumulation of lipids which can either be due to excessive production and release of the lipids into circulation or by defective removal from the blood or the combination of both and could cause liver dysfunctioning^{33,38}. The mechanisms underlying the observed accumulation of liver triglycerides may include increased liver fatty acid mobilization and delivery to the liver, increased hepatic lipogenesis and decreased secretion of very low density lipoprotein³⁹. Also, the observed increase hepatic phospholipid may be due to heighten free fatty acid availability and/or increased cholesterologenesis^{40,41}.

Another finding of this study was that induced-anaemia elevated cholesterol concentration. This is in agreement with reports of Peng et al.⁴² and some studies that there is relationship between hypercholesterolemia and anaemia having effect on atherosclerosis^{16,43-45}. The up-regulation expression of hepatic 3-hydroxy-3-methylglutaryl-CoA Reductase (Hmgcr) gene contributed to the overall cholesterol status as observed in the present study. This is consistent with the reports of Ness and Chambers⁴⁶, Eisa-Beygi et al.⁴⁷ and Tian et al.⁴⁸. Likewise, the present study showed that induced-anaemia resulted in activation of Lecithin cholesterol acyltransferase (Lcat) gene expression (important enzyme in the reverse cholesterol transport process)⁴⁹⁻⁵¹ which in turns elevated plasma cholesterol. The result also showed the synergistic effect of mRNA gene expression for Hmgcr and Lcat activation and increase in the level of plasma cholesterol in the induced anaemic rats.

The observed reduced gene expression of scavenger receptor class B Type I (Scarb 1) due to induced-anaemia in the rat may correlate with elevation of cholesterol levels. This result is consistent with report of Assanasen et al.⁵² that the cholesterol flux mediated by Scarb1 plays a role in the regulation of signal-transduction initiation. In this present study the significant up-regulation of paraoxonase-1 (Pon-1) gene expression is in consistent with the report of Gong et al.⁵³. The increase may be partially explained by the ability to reduce oxidative stress by scavenging oxidative free radicals and found to be associated with aryl hydrocarbon receptor (Ahr) dependent mechanism as supported by the report of Guedard et al.⁵⁴. The current study also revealed the elevation of Ahr gene expression in the liver which is similar to the report of Wada et al.⁵⁵.

Fig. 3(a-e):

Effects of induced-anaemia on relative expression of hepatic lipid metabolizing genes (a) Ahr, (b) Lcat, (c) Pon-1, (d) Hmgcr and (e) Scarb1

Bars represent the Means \pm SEM (n = 3). Relative expression is ratio of intensity of each gene to that of housekeeping gene (β -actin, Actb). Bars with different alphabets are significantly ($p < 0.05$) different from each other

This may be involve in protecting the liver against lipotoxicity, transcription of cholesterol-biosynthetic genes-Hmgcr and elevated cholesterol, triglyceride and phospholipid through interaction with the transcription factor-sterol element binding protein 2, although this was not investigated.

CONCLUSION

Phenylhydrazine anaemic-induction in rat model for 8 days revealed hepatic dysfunction and dysregulation of lipid and lipoprotein metabolism through altered expression of Hmgcr, Pon1, Lcat, Scarb1 and Ahr genes. Notably, down-regulation of Scarb1 may be more sensitive indicator of anaemic-induction. Overall, the changes observed in this study may be associated to be the underlying mechanism whereby induced-anaemia signal liver damage.

SIGNIFICANCE STATEMENT

This study provides significant insight into the molecular changes sustained in expression of lipoprotein genes and lipid metabolism by PHZ induced-anaemia in a rat model. The mechanism of action of these changes may be due to the observed dyslipidemia, up regulation of liver lipids and dysregulation of lipoprotein metabolizing gene expression, which can ultimately be associated with the risk of cardiovascular diseases.

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References

Abramoff, M.D., P.J. Magelhaes and S.J. Ram, 2004. Image processing with image. *Biophotonics Int.*, 11: 36-42.

[Direct Link](#) |

Akinlade, K.S., C.O. Adewale, S.K. Rahamon, F.A. Fasola, J.A. Olaniyi and A.D. Atere, 2014. Defective lipid metabolism in sickle cell anaemia subjects in vaso-occlusive crisis. *Niger. Med. J.*, 55: 428-431.

[CrossRef](#) | [Direct Link](#) |

Alferez, M.J., I. Lopez-Aliaga, T. Nestares, J. Diaz-Castro, M. Barrionuevo, P.B. Ros and M.S. Campos, 2006. Dietary goat milk improves iron bioavailability in rats with induced ferropenic anaemia in comparison with cow milk. *Int. Dairy J.*, 16: 813-821.

[CrossRef](#) | [Direct Link](#) |

Anandakumar, P. and M.K. Vanitha, 2014. Drug induced hepatotoxicity-a review. *Adv. J. Pharm. Life Sci. Res.*, 2: 39-45.

[Direct Link](#) |

Ashar, S., S. Sultan and A. Sheeraz, 2015. Serum fasting lipid profile in children and adolescents with β -thalassaemia major in Southern Pakistan. *Malays J. Pathol.*, 37: 233-238.

[PubMed](#) | [Direct Link](#) |

Ashour, T.H., 2014. Hematinic and anti-anemic effect of thymoquinone against phenylhydrazine-induced hemolytic anemia in rats. *Res. J. Med. Sci.*, 8: 67-72.

[Direct Link](#) |

Assanasen, C., C. Mineo, D. Seetharam, I.S. Yuhanna and Y.L. Marcel et al., 2005. Cholesterol binding, efflux and a PDZ-interacting domain of scavenger receptor-BI mediate HDL-initiated signaling. *J. Clin. Invest.*, 115: 969-977.

[CrossRef](#) | [Direct Link](#) |

Berger, J., 2007. Phenylhydrazine haematotoxicity. *J. Applied Biomed.*, 5: 125-130.

[Direct Link](#) |

Beutler, E., 2001. Hemolytic Anemia Due to Chemical and Physical Agents. In: Williams Hematology, Beutler, E., B.S. Coller, M.A. Lichtman, T.J. Kipps and U. Seligsohn (Eds.). 6th Edn., McGraw-Hill, New York, pp: 629-632.

Chowta, N.K., S.B. Reddy, M.N. Chowta, A. Shet, B. Achappa and D.R. Madi, 2017. Lipid profile in anemia: Is there any correlation? *Ann. Trop. Med. Public Health*, 10: 837-840.

[CrossRef](#) | [Direct Link](#) |

Dabbagh, A.J., G.T. Shwaery, J.F. Keaney, Jr. and B. Frei, 1997. Effect of iron overload and iron deficiency on atherosclerosis in the hypercholesterolemic rabbit. *Arteriosclerosis Thrombosis Vascul. Biol.*, 17: 2638-2645.

[Direct Link](#) |

Dabbagh, A.J., T. Mannion, S.M. Lynch and B. Frei, 1994. The effect of iron overload on rat plasma and liver oxidant status in vivo. *Biochem. J.*, 300: 799-803.

[CrossRef](#) | [PubMed](#) |

Diamanti-Kandarakis, E., A.G. Papavassiliou, S.A. Kandarakis and G.P. Chrousos, 2007. Pathophysiology and types of dyslipidemia in PCOS. *Trends Endocrinol. Metab.*, 18: 280-285.

Dobiasova, M. and J.J. Frohlich, 1999. Advances in understanding of the role of Lecithin Cholesterol Acyltransferase (LCAT) in cholesterol transport. *Clin. Chim. Acta*, 286: 257-271.

Eisa-Beygi, S., M. Ekker, T.W. Moon, R.L. Macdonald and X.Y. Wen, 2014. Developmental processes regulated by the 3-hydroxy-3-methylglutaryl-CoA reductase (HMGCR) pathway: Highlights from animal studies. *Reprod. Toxicol.*, 46: 115-120.

El Hilaly, J., Z.H. Israili and B. Lyoussi, 2004. Acute and chronic toxicological studies of *Ajuga iva* in experimental animals. *J. Ethnopharmacol.*, 91: 43-50.

Folch, J., M. Lees and G.H.S. Stanley, 1957. A simple method for the isolation and purification of total lipides from animal tissues. *J. Biol. Chem.*, 226: 497-509.

Gkamprela, E., M. Deutsch and D. Pectasides, 2017. Iron deficiency anemia in chronic liver disease: Etiopathogenesis, diagnosis and treatment. *Ann. Gastroenterol.*, 30: 405-413.

Gong, M., M. Garige, R. Varatharajalu, P. Marmillot, C. Gottipatti, L.C. Leckey and R.M. Lakshman, 2009. Quercetin up-regulates paraoxonase 1 gene expression with concomitant protection against LDL oxidation. *Biochem. Biophys. Res. Commun.*, 379: 1001-1004.

Gouedard, C., R. Barouki and Y. Morel, 2004. Dietary polyphenols increase paraoxonase 1 gene expression by an aryl hydrocarbon receptor-dependent mechanism. *Mol. Cell. Biol.*, 24: 5209-5222.

Herz, J., 2009. ApoE receptors in the nervous system. *Curr. Opin. Lipidol.*, 20: 190-196.

Jain, S.K. and D. Subrahmanyam, 1978. On the mechanism of phenylhydrazine-induced hemolytic anemia. *Biochem. Biophys. Res. Commun.*, 82: 1320-1324.

Jollow, D.J. and D.C. McMillan, 2001. Oxidative Stress, Glucose-6-Phosphate Dehydrogenase and the Red Cell. In: *Biological Reactive Intermediates VI*, Dansette, P.M., R. Snyder, M. Delaforge, G.G. Gibson and H. Greim et al. (Eds.). Springer, USA., pp: 595-605.

Katsiki, N., D.P. Mikhailidis and C.S. Mantzoros, 2016. Non-alcoholic fatty liver disease and dyslipidemia: An update. *Metabolism*, 65: 1109-1123.

Kaur, S.C., 2015. *Biochemistry of Atherosclerosis*. Springer Science and Business Media, New York, pp: 23-38.

Kelesidis, T. and J.S. Currier, 2014. Dyslipidemia and cardiovascular risk in human immunodeficiency virus infection. *Endocrinol. Metab. Clin.*, 43: 665-684.

Lee, C.H., P. Olson and R.M. Evans, 2003. Minireview: Lipid metabolism, metabolic diseases, and peroxisome proliferator-activated receptors. *Endocrinology*, 144: 2201-2207.

PubMed |

Mazandarani, M. and S.M. Hoseini, 2017. Anaemia and plasma lipid profile in common carp (*Cyprinus carpio*) exposed to ambient copper sulphate and nano-scale copper oxide. *Aquacult. Res.*, 48: 844-852.

McMillan, D.C., C.B. Jensen and D.J. Jollow, 1998. Role of lipid peroxidation in dapsone-induced hemolytic anemia. *J. Pharmacol. Exp. Ther.*, 287: 868-876.

Muller, A., H. Jacobsen, E. Healy, S. McMickan and F. Istace et al., 2006. Hazard classification of chemicals inducing haemolytic anaemia: An EU regulatory perspective. *Regul. Toxicol. Pharmacol.*, 45: 229-241.

NRC., 2011. *Guide for the Care and Use of Laboratory Animals*. 8th Edn., National Academies Press, Washington, DC., USA., ISBN-13: 9780309154000, Pages: 220.

Ness, G.C. and C.M. Chambers, 2000. Feedback and hormonal regulation of hepatic 3-Hydroxy-3-methylglutaryl coenzyme a reductase: The concept of cholesterol buffering capacity. *Exp. Biol. Med.*, 224: 8-19.

Newberry, E.P., S.M. Kennedy, Y. Xie, J. Luo and S.E. Stanley et al., 2008. Altered hepatic triglyceride content after partial hepatectomy without impaired liver regeneration in multiple murine genetic models. *Hepatology*, 48: 1097-1105.

Ogunrinola, O.O., 2015. Lipid profile and malondialdehyde concentrations in cadmium-induced rats: A study with relation to doses. *MOJ Toxicol.*, 1: 149-154.

Ogunrinola, O.O., O.O. Fajana, B.O. Williams, E. Ogedengbe, A.A. Onifade, F.C. Ekeocha and K.O. Shasore, 2016. The therapeutic potential of *Cocos nucifera* water on cadmium-induced lipid toxicity in male rat. *Int. J. Sci. Res. Environ. Sci. Toxicol.*, Vol. 1.

Peng, M., W. Xu, K. Mai, H. Zhou and Y. Zhang et al., 2014. Growth performance, lipid deposition and hepatic lipid metabolism related gene expression in juvenile turbot (*Scophthalmus maximus* L.) fed diets with various fish oil substitution levels by soybean oil. *Aquaculture*, 433: 442-449.

Rifai, N., G.R. Warnick and M.H. Dominiczak, 2000. *Handbook of Lipoprotein Testing*. 2nd Edn., American Association Clinical Chemistry, Washington, DC., USA.

Rolim, A.E.H., R. Henrique-Araujo, E.G. Ferraz, F.K.D.A.A. Dultra and L.G. Fernandez, 2015. Lipidomics in the study of lipid metabolism: Current perspectives in the omic sciences. *Gene*, 554: 131-139.

Rother, R.P., L. Bell, P. Hillmen and M.T. Gladwin, 2005. The clinical sequelae of intravascular hemolysis and extracellular plasma hemoglobin: A novel mechanism of human disease. *JAMA.*, 293: 1653-1662.

Rotimi, O.A., I.O. Olayiwola, O. Ademuyiwa and E.A. Balogun, 2012. Effects of fibre-enriched diets on tissue lipid profiles of MSG obese rats. *Food Chem. Toxicol.*, 50: 4062-4067.

Rotimi, O.A., S.O. Rotimi, C.U. Duru, O.J. Ebebeinwe, A.O. Abiodun, B.O. Oyeniya and F.A. Faduyile, 2017. Acute aflatoxin B1-Induced hepatotoxicity alters gene expression and disrupts lipid and lipoprotein metabolism in rats. *Toxicol. Rep.*, 4: 408-414.

Rotimi, O.A., S.O. Rotimi, F. Oluwafemi, O. Ademuyiwa and E.A. Balogun, 2016. Coexistence of aflatoxicosis with protein malnutrition worsens hepatic oxidative damage in rats. *J. Biochem. Mol. Toxicol.*, 30: 269-276.

Rotimi, S.O., D.A. Ojo, O.A. Talabi, R.N. Ugbaja, E.A. Balogun and O. Ademuyiwa, 2015. Amoxicillin-and pefloxacin-induced cholesterogenesis and phospholipidosis in rat tissues. *Lipids Health Dis.*, Vol. 14. 10.1186/s12944-015-0011-8

Rotimi, S.O., G.E. Bankole, I.B. Adelani and O.A. Rotimi, 2016. Hesperidin prevents lipopolysaccharide-induced endotoxicity in rats. *Immunopharmacol. Immunotoxicol.*, 38: 364-371.

Sawada, H., K. Takami and S. Asahi, 2004. A toxicogenomic approach to drug-induced phospholipidosis: Analysis of its induction mechanism and establishment of a novel in vitro screening system. *Toxicol. Sci.*, 83: 282-292.

Shirvani, M., M.V. Sadeghi, S.R. Hosseini, A. Bijani and R. Ghadimi, 2017. Does serum lipid profile differ in anemia and non-anemic older subjects? *Caspian J. Int. Med.*, 8: 305-310.

Shukla, P., N.K. Yadav, P. Singh, F.W. Bansode and R.K. Singh, 2012. Phenylhydrazine induced toxicity: A review on its haematotoxicity. *Int. J. Basic Applied Med. Sci.*, 2: 86-91.

Sotuneh, N., S.R. Hosseini, J. Shokri-Shirvani, A. Bijani and R. Ghadimi, 2014. Helicobacter pylori infection and metabolic parameters: Is there an association in elderly population? *Int. J. Prevent. Med.*, 5: 1537-1542.

Thacker, S.G., X. Rousset, S. Esmail, A. Zarzour and X. Jin et al., 2015. Increased plasma cholesterol esterification by LCAT reduces diet-induced atherosclerosis in SR-BI knockout mice. *J. Lipid Res.*, 56: 1282-1295.

Tian, L., Y. Song, M. Xing, W. Zhang and G. Ning et al., 2010. A novel role for thyroid-stimulating hormone: Up-regulation of hepatic 3-hydroxy-3-methyl-glutaryl-coenzyme a reductase expression through the cyclic adenosine monophosphate/protein kinase A/cyclic adenosine monophosphate-responsive element binding protein pathway. *Hepatology*, 52: 1401-1409.

Unami, A., N. Nishina, T. Terai, S. Sato, T. Tamura, K. Noda and Y. Mine, 1996. Effects of cisplatin on erythropoietin production in rats. *J. Toxicol. Sci.*, 21: 157-165.

Wada, T., H. Sunaga, K. Miyata, H. Shirasaki, Y. Uchiyama and S. Shimba, 2016. Aryl hydrocarbon receptor plays protective roles against HFD-induced hepatic steatosis and the subsequent lipotoxicity via direct transcriptional regulation of Socs3 expression. *J. Biol. Chem.*, Vol. 291. 10.1074/jbc.M115.693655

Winther, S.A., N. Finan, A. Sharma, C. Trop-Pedersen and C. Andersson, 2014. Association of anemia with the risk of cardiovascular adverse events in overweight/obese patients. *Int. J. Obes.*, 38: 432-437.

Yang, S., X.Y. Chen and X.P. Xu, 2015. The relationship between lipoprotein-associated phospholipase A (2), cholesteryl ester transfer protein and lipid profile and risk of atherosclerosis in women with iron deficiency anaemia. *Clin. Lab.*, 61: 1463-1469.

Zorca, S., L. Freeman, M. Hildesheim, D. Allen, A.T. Remaley, J.G. Taylor and G.J. Kato, 2010. Lipid levels in sickle-cell disease associated with haemolytic severity, vascular dysfunction and pulmonary hypertension. *Br. J. Haematol.*, 149: 436-445.