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Platelet lamellipodia formation is not required for thrombus formation and stability

Short running title: Cyfip1 in platelet function

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Key points

- Cyfip1 plays a crucial role for branching of actin filaments and for lamellipodia formation.
- Lamellipodia formation is not required for the formation of a hemostatic plug or thrombus.

Abstract

During platelet spreading, the actin cytoskeleton undergoes rapid rearrangement, forming filopodia and lamellipodia. Controversial data have been published on the role of lamellipodia in thrombus formation and stability. The WAVE regulatory complex, which has been shown in other cells to drive lamellipodia formation by enhancing actin nucleation via the Arp2/3 complex, is activated by Rac1 interaction with the WAVE complex subunit Cyfip1. We analyzed *Cyfip1^{flox/flox} Pf4-Cre* mice to investigate the role of Cyfip1 in platelet function. These mice displayed normal platelet counts and a slight reduction in platelet volume. Activation of mutant platelets was only moderately reduced to all tested agonists as measured by α IIb β 3 integrin activation and P-Selectin surface exposure. However, lamellipodia formation of mutant platelets was completely abolished on different matrices. Nevertheless, Cyfip1-deficient platelets formed stable thrombi on collagen fibers *ex vivo* and in two models of occlusive arterial thrombosis *in vivo*. Similarly, the hemostatic function and maintenance of vascular integrity during inflammation of the skin and lung were unaltered in the mutant mice. Investigation of platelet morphology in an induced thrombus under flow revealed that platelets rather form filopodia in the thrombus shell, and are flattened with filopodia-like structures when in direct contact to collagen fibers at the bottom of the thrombus. We provide for the first time direct evidence that platelet lamellipodia formation is not required for stable thrombus formation, and that morphological changes of platelets differ between a static spreading assay and thrombus formation under flow.

Introduction

Circulating platelets are essential players in hemostasis and thrombosis. Upon vascular injury, platelets rapidly adhere to exposed extracellular matrix proteins and form a hemostatic plug which limits excessive blood loss, but also represents a major pathomechanism of ischemic cardio- and cerebrovascular diseases.¹ Thrombus formation is a multi-step process. At first, platelet tethering is mediated by the interaction of glycoprotein (GP) Ib with immobilized von Willebrand factor on exposed subendothelial collagen. This interaction enables binding of GPVI to collagen, which in turn initiates cellular activation. Subsequent inside-out upregulation of integrin affinity, synthesis of soluble mediators, granule release, and coagulant activity contribute to firm platelet adhesion and thrombus growth.²

The cytoskeleton ensures normal platelet size and shape, and plays a fundamental role for platelet function. Upon agonist-induced platelet activation, a rapid rearrangement of the cytoskeleton takes place, which results in platelet shape change and contributes to thrombus formation and stability.³ *In vitro* studies have shown that platelets getting in contact with immobilized adhesive proteins such as fibrinogen, start to spread and form first finger-like protrusions, so-called filopodia, which are finally remodeled to lamellipodia, which are flat, undulating cellular protrusions with Arp2/3-mediated orthogonally arrayed short actin filaments. A fully spread platelet displays a circumferential zone of orthogonally arrayed short filaments within lamellipodia.⁴ Controversial data have been published on the role of lamellipodia in thrombus formation and stability, which were mainly based on inhibitor studies or on analyses of platelets with additional functional defects.⁵⁻⁹ A role for the small GTPase Rac1 in platelet lamellipodia formation was demonstrated using mice with deletion of Rac1 in the hematopoietic system (Mx1-Cre).^{5,6} However, while it was suggested that Rac1-regulated lamellipodia formation might be essential for maintaining integrity of platelet aggregates under shear,⁵ another study showed that Rac1 deficiency leads to a specific GPVI-dependent PLC γ 2 activation defect, resulting in impaired platelet adhesion and defective thrombus formation on collagen under flow. Supplementation with ADP and a thromboxane A2 analogue could rescue this defect.⁶ Further, analysis of mice with a markedly decreased expression of the Arp2/3 complex⁹ revealed no significant defects in the hemostatic function or thrombus formation, although mutant platelets were unable to form lamellipodia. However, deletion of Arpc2 was not completely successful in megakaryocytes and platelets (>95% reduction),⁹ which could potentially rescue platelet function. Thus, the role of lamellipodia in this process is still unclear and direct proof has been lacking.

The WASp family verprolin-homologous protein (WAVE) regulatory complex is a five-subunit protein complex, which includes the proteins WAVE-1/-2/-3, ABI1/2, NAP1 (also known as NCKAP1) or NCKAP1L, Cytoplasmic FMR1-interacting protein (Cyfip) 1/2 and HSPC300.¹⁰

The WAVE complex is *per se* inactive, but is activated upon interaction of the subunit Cyfip1/2 and Rac1-GTP.¹¹ The active WAVE complex interacts with Arp2/3, which in turn is crucial for the formation of branched actin filaments. WAVE-1 and -2 are the two major isoforms of WAVE in platelets.¹² Homozygous disruption of the WAVE1 gene resulted in postnatal lethality with an average lifespan of 23.6 days.¹³ Analysis of platelets of young mutant mice (age of 14-20 days) revealed that WAVE-1 plays an important role downstream of GPVI.¹⁴ WAVE-2-null mice died at embryonic day 12.5 and their *in vitro* differentiated megakaryocytes exhibited spreading defects.¹⁵ The Cyfip1 protein is mainly in a complex with the WAVE-2 isoform,¹⁶⁻¹⁸ however its role for platelet function has not been studied.

We investigated the role of Cyfip1 in platelet physiology by generating *Cyfip1^{fl/fl} Pf4-Cre* mice specifically lacking Cyfip1 in megakaryocytes and platelets. We demonstrate that *Cyfip1^{fl/fl} Pf4-Cre* mice display normal platelet counts, and that activation of their platelets is only moderately decreased. However, mutant platelets are unable to form lamellipodia on different matrices, making this mouse line a unique model to study the function of platelet lamellipodia. We finally provide definite evidence that lamellipodia formation is dispensable for thrombus growth and stability, and that morphological changes of platelets differ between a static spreading assay and thrombus formation under flow.

Material and Methods

Animals. Animal studies were approved by the district government of Lower Franconia (Bezirksregierung Unterfranken). Male and female mice were analyzed between 4 and 20 weeks of age. The *Cyfip1* flox mouse strain was created from ES cell clone (DEPD00521_7_E11) obtained from the NCCR-NIH supported KOMP Repository (www.komp.org) and generated by the CSD consortium for the NIH funded Knockout Mouse Project (KOMP). Following importation, the ES cells were cultured on a MEF feeder layer. Correct targeting of *Cyfip1* was confirmed on DNA extracted from the ES cell clone by PCR on the 5' side (CATTCTAACGTTCAAGATCGCAGACCAG and CACAACGGGT TCTTCTGTTAGTCC; 7.5kb) and on the 3' side (CATGTCTGGATCCGGGGGTACC GCGTCGAG and GATCTCCTGGTGCGGCATGCAAGC; 7.1kb). The presence of the isolated 3' loxP site was also confirmed by PCR (AAGCAGTCCGTGCTGAGATG and GAACCAAGCTCCCAGATTCC; 262bp). Following confirmation of the genotyping of the targeted clone, mouse lines were derived by injection of ES cells into C57BL/6J blastocysts. After breeding of chimeras, germline offspring were identified by coat color and the presence of the modified allele was confirmed with the 3' loxP primers described above. Mice were subsequently crossed with a mouse line expressing Flpe (Tg(ACTFLPe)9205Dym) to delete the selectable marker by recombination at the FRT sites.¹⁹ Correct removal of the lacZ gene

and Neo cassette was confirmed by PCR across the remaining FRT site (CTCTTCTTGTGGACCGCATC and CACGGTAGCCTGACAAGAGG; 562bp). To study the function of Cyfip1 in platelets, *Cyfip1^{fl/fl}* mice (supplemental Figure 1A) were intercrossed with mice carrying the Cre-recombinase under the platelet factor 4 (*Pf4*) promoter²⁰ to generate platelet- and megakaryocyte-specific Cyfip1-knockout mice. *Cyfip1^{fl/fl Pf4-Cre}* is further referred to as *Cyfip1^{-/-}*, *Cyfip1^{fl/fl}* is further referred to as *Cyfip1^{+/+}* which served as controls.

Platelet preparation, immunoblotting, aggregometry, flow cytometry, platelet spreading, electron and super-resolution microscopy, bleeding time, clot retraction, ex vivo and in vivo thrombus formation, reverse passive Arthus reaction and LPS-induced inflammation are described in supplemental Material and methods.

Data analysis. The presented results are mean \pm standard deviation from at least two to three independent experiments per group, if not stated otherwise. Differences between control and knockout mice were statistically analyzed using the Mann-Whitney-U test. *P*-values <0.05 were considered as statistically significant: **P* < 0.05 ; ***P* <0.01 ; ****P* <0.001 . Results with a *P*-value > 0.05 were considered as not significant.

Data sharing statement. Data supplement is available with the online version of this article.

Results

***Cyfip1^{fl/fl Pf4-Cre}* mice display normal platelet counts and only moderately impaired platelet activation**

We confirmed lack of the Cyfip1 protein in platelet lysates of *Cyfip1^{-/-}* mice by Western blot analysis (Figure 1A, supplemental Figure 1B). WAVE2 expression was reduced in mutant platelets, indicating instability of the complex in the absence of Cyfip1, however, expression of Rac1, Arpc2 and WAVE1 remained unaltered (Figure 1A, supplemental Figure 1C). *Cyfip1^{-/-}* mice displayed normal blood cell count parameters, including platelet count, and only a slight but significant reduction in platelet volume (Figure 1B-C, MPV: *Cyfip1^{+/+}* 5.11 ± 0.16 fL, *Cyfip1^{-/-}* 4.83 ± 0.07 fL, ****P* $<.001$, supplemental Figure 1D). Expression of prominent glycoproteins and platelet ultrastructure were comparable between *Cyfip1^{+/+}* and *Cyfip1^{-/-}* mice (supplemental Figures 1E and 2).

Activation of mutant platelets was only moderately reduced in response to all tested agonists as measured by α IIb β 3 integrin activation and P-Selectin surface exposure (Figure 1D-E). To test the functional consequences of this moderate activation defect, we performed *in vitro* aggregation studies. *Cyfip1^{-/-}* platelets aggregated normally to the G protein-coupled agonist

thrombin (Figure 1F) and the GPVI-specific agonist collagen-related peptide (CRP) even at very low concentrations (Figure 1F). We only observed consistently a delay in onset of aggregation in response to collagen. However, the amplitude as a measure for maximum platelet aggregation was comparable between control and mutant platelets (Figure 1F). Next, we preincubated washed platelets with apyrase (2 U/ml) and indomethacin (10 μ M) to prevent the effect of second wave mediators, and then activated with high concentrations of collagen. We found that *Cyfp1*^{-/-} platelets aggregated normally at high collagen concentrations under these conditions (10 and 20 μ g/mL collagen, Figure 1F), excluding a selective GPVI signaling defect in *Cyfp1*^{-/-} platelets as observed for *Rac1*-deficient platelets.⁶ Next, we assessed changes in protein tyrosine phosphorylation upon stimulation with CRP and detected a comparable total tyrosine phosphorylation pattern in *Cyfp1*^{-/-} platelets (Figure 1G).

The Arp2/3 complex is activated by the WAVE regulatory complex downstream of *Rac1*. Therefore, we performed activation studies with control platelets in the presence of the Arp2/3 inhibitor, CK-666, and the fungal toxin, cytochalasin D, a potent actin polymerization inhibitor. Control platelets incubated with CK-666 (50 μ M) displayed reduced activation similar to the reduction seen with *Cyfp1*^{-/-} platelets (α IIb β 3 activation: supplemental Figure 3A; P-selectin exposure: supplemental Figure 3B). Preincubation of platelets with cytochalasin D had a stronger effect on agonist-induced activation as compared to *Cyfp1*-deficient platelets (supplemental Figure 3). These data suggest that the moderately reduced activation of *Cyfp1*^{-/-} platelets is rather caused by an Arp2/3-related defect than an upstream signaling defect. Taken together, lack of *Cyfp1* only moderately and non-selectively impairs platelet activation, which has no major effect on platelet-platelet interaction.

Absent lamellipodia formation of *Cyfp1*^{-/-} platelets on different matrices in static spreading assays

It has been shown that the WAVE regulatory complex drives lamellipodia formation in other cell types.²¹⁻²³ This prompted us to study the role of the WAVE complex subunit *Cyfp1* in platelet lamellipodia formation. Four consecutive stages of platelet spreading can be observed: (I) adhesion of resting platelets, (II) formation of filopodia, (III) a combination of filopodia and lamellipodia, and (IV) only lamellipodia. *Cyfp1*^{-/-} platelets adhered to fibrinogen under static conditions, formed filopodia, but strikingly, lamellipodia formation (phases III and IV) was completely abolished (Figure 2A-B, supplemental Videos 1 and 2). However, the transition from resting platelet to filopodia-forming platelet was unaltered between both groups as revealed by live-imaging of platelet spreading (Figure 2C). At the 30 min time point, spread mutant platelets were smaller but had an increased perimeter, and formed more filopodia with increased length (Figure 2D-G). To compare the defect of lamellipodia

formation of *Cyfp1*^{-/-} platelets to reagents interfering with actin branching or polymerization, we incubated control platelets with different concentrations of CK-666 and cytochalasin D prior spreading, and observed that a concentration of 50 μ M CK-666 or higher resulted in a significant reduction of fully spread platelets (phase IV), however, even at 200 μ M CK-666 approx. 30% of platelets could be observed in phase III (supplemental Figure 4A). Nevertheless, this data shows that the spreading defect of CK-666 treated platelets resembles to a large extent the phenotype of the *Cyfp1*-deficient platelets. In contrast, platelets were unable to change their shape when actin polymerization was inhibited by cytochalasin D (supplemental Figure 4B). These results suggest that *Cyfp1*-deficient platelets cannot form Arp2/3-dependent branched actin filaments, which are necessary for lamellipodia formation.

Therefore, we investigated the localization of Arp2/3 in spread platelets and found that Arp2/3 was strongly expressed at the circumferential zone of control platelets (supplemental Figure 5). Interestingly, Arp2/3 also localized to filopodia in control and mutant platelets, indicating that not altered expression or localization, but impaired *Cyfp1*-mediated Arp2/3 activation is causative for abolished lamellipodia formation. To assess this in more detail, we analyzed the cytoskeletal organization of spread platelets by platinum replica electron²⁴ (Figure 3A) and super-resolution microscopy by *direct* stochastic optical reconstruction microscopy (*d*STORM)²⁵ (supplemental Figure 6). Orthogonally arrayed short actin filaments were formed in the circumferential zone of spread control platelets. In contrast, *Cyfp1*^{-/-} platelets showed only filopodia consisting of parallel bundles of actin filaments (Figure 3A). In agreement, *Cyfp1*^{-/-} platelets were principally able to assemble filamentous actin as we determined the F-actin content in resting platelets and in platelets after activation with CRP or thrombin by flow cytometry. Only a slightly decreased content was observed in CRP-activated platelets but not resting or thrombin-activated platelets (Figure 3B). However, the F-actin ratio of activated platelets over resting platelets revealed no significant differences, demonstrating that mutant platelets can assemble F-actin after activation similar to controls (Figure 3B). Next, we tested whether *Cyfp1*-deficient platelets form lamellipodia on other surfaces. Likewise, mutant platelets could not form lamellipodia on collagen IV, laminin and CRP and were unable to reorganize actin filaments into short, branched filaments (Figure 4A-B).

Lamellipodia are not required for *ex vivo* thrombus formation

Next, we tested the role of lamellipodia in thrombus formation in an *in vitro* system. Therefore, we perfused whole blood over a collagen-coated surface in a flow chamber at different shear rates (150 s⁻¹, 1000 s⁻¹, 1700 s⁻¹). Interestingly, surface coverage (Figure 5A-B) and thrombus volume (Figure 5A,C) were comparable between samples from control and

mutant mice (supplemental Videos 3 and 4). Kinetics of single platelet adhesion and embolization of smaller platelet aggregates appeared normal. Similarly, *Cyfp1^{-/-}* platelets showed effective thrombus compaction under flow (supplemental Video 4) and clot retraction *in vitro* (Figure 5D-E). Video analyses revealed that single platelets at times formed filopodia to grab collagen fibers, before they became activated and initiated thrombus formation (supplemental Video 5). Surprisingly, we have never observed fully spread platelets (equivalent to phase IV platelets in static spreading assay) with circumferential lamellipodia under these conditions. These findings were corroborated when we removed the flow chamber from the glass coverslip after the experiment and prepared the sample for scanning (Figure 6A) and platinum replica (Figure 6B) electron microscopic analyses. Investigation of control and *Cyfp1^{-/-}* platelet morphology showed that in general, platelets form filopodia in the thrombus shell (Figure 6A) and are flattened with filopodia structures and parallel actin bundles when in direct contact to collagen fibers at the bottom of the thrombus (Figure 6A-B, supplemental Figure 7A-B), contrary to the abundant fully spread platelets with circumferential lamellipodia observed in a static spreading assay (Figures 2 and 3). Some unspread platelets at collagen fibers were found in control and mutant samples and only in a few control platelets we could identify small parts of branched actin filaments at the cell boundary (supplemental Figure 7A-B). Taken together, these results demonstrate that lamellipodia formation is not required for stable thrombus formation *ex vivo* and that morphological changes of platelets differ between a static spreading assay and thrombus formation under flow.

***Cyfp1^{-/-}* platelets can expose phosphatidylserine on the surface**

A distinct population of highly activated platelets has been described residing close to collagen fibers and as patches around a thrombus. These platelets are characterized by surface-exposed phosphatidylserine (PS), which supports coagulation factor complex assembly, and hence is a key regulatory event in murine arterial thrombus formation.^{26,27} To test whether *Cyfp1^{-/-}* platelets can regulate coagulant activity, washed platelets were stimulated with ionomycin, a combination of CRP and thrombin or alone, and PS exposure was determined by Annexin V binding. While ionomycin efficiently induced PS exposure, platelets stimulated with thrombin or CRP alone could hardly bind Annexin V. The combination of CRP and thrombin induced PS exposure on more than 40% of control and mutant platelets (supplemental Figure 8A-B). Next, we perfused whole blood over immobilized collagen and observed that the number and surface coverage of PS-positive collagen-adherent platelets were comparable between control and mutant samples (supplemental Figure 8C-D). These data show that platelets can regulate coagulant activity without the necessity to form lamellipodia.

Normal bleeding times, arterial thrombus formation and maintenance of vascular integrity in *Cytip1*^{-/-} mice

Next, we analyzed the relevance of lamellipodia structures for *in vivo* platelet function. Bleeding times after tail tip amputation were comparable between control and *Cytip1*^{-/-} mice (Figure 7A). The application of 20% FeCl₃ on the exteriorized mesenteric arterioles of *Cytip1*^{-/-} mice resulted in comparable platelet adhesion, appearance of first thrombi (Figure 7B-C) and time to formation of occlusive thrombi (Figure 7D, supplemental Videos 6 and 7). Similarly, thrombus growth was studied in a second model of arterial thrombosis, in which the abdominal aorta is mechanically injured and blood flow is monitored by an ultrasonic perivascular Doppler flowmeter. In this model, thrombus formation and occlusion of the vessel in *Cytip1*^{-/-} mice occurred with similar kinetics as in controls (Figure 7E, supplemental Figure 9A). Finally, we isolated the aorta and analyzed the shape of single platelets in the experimentally-induced *in vivo* thrombus (Figure 7F, supplemental Figure 9B). Again, we could not observe platelets with lamellipodia, but rather platelets which formed filopodia. This shows that lamellipodia structures are not required for classical formation of a hemostatic plug or a pathological thrombus *in vivo*.

It has been reported that inflammation represents an unconventional hemostatic situation in which individual platelets are sufficient to prevent bleeding.²⁸ To test whether platelet lamellipodia structures play a role in this process, we challenged mice in two different models, namely the reverse passive Arthus reaction (intravenous injection of antigen, followed by intradermal injection of antibody) and LPS-induced inflammation. As expected, when we depleted platelets in mice by injection of an anti-GPIIb/IIIa antibody, we observed severe inflammatory bleeding in the skin (Figure 7G-H) and into the lung (Figure 7I-J), demonstrating the importance of platelets to maintain vascular integrity as reported previously.²⁹ Finally, control and *Cytip1*^{-/-} mice were subjected to the reverse passive Arthus reaction of the skin and LPS-induced lung inflammation. *Cytip1*^{-/-} mice did not show signs of bleeding in these experimental settings, suggesting that platelet lamellipodia formation is dispensable for maintaining vascular integrity at sites of acute inflammation (Figure 7G-J).

Discussion

Static platelet spreading assays, particularly on fibrinogen-coated surfaces, are frequently performed as a measure of the contribution of outside-in signaling and cytoskeletal rearrangement in platelet function³⁰ as well as readout for functional properties of *in vitro* produced platelets.³¹⁻³⁴ *In vitro*, four stages of platelet spreading can be observed: (I) adhesion of resting platelets, (II) formation of finger-like protrusions, called filopodia, (III) a

combination of filopodia and plate-like extensions, called lamellipodia, and (IV) only lamellipodia. Under dynamic conditions, platelets adhere to the exposed extracellular matrix, become activated and thereby change their shape, which is accompanied by the secretion and synthesis of several prothrombotic factors, which together contribute to the activation of more platelets and finally leads to aggregate formation.

It has been debated over years whether the stages of *in vitro* platelet spreading, particularly lamellipodia formation, also contribute to platelet adhesion and activation under dynamic conditions *in vivo*. Platelet lamellipodia have been suggested to be important for the initiation of thrombus growth when platelets are in direct contact with extracellular matrix components, most notably collagen.^{5,7,8} In contrast, other publications provided indirect evidence that lamellipodia formation is not crucial for thrombus formation and hemostasis.^{6,9,35} However, the mouse models of Rac1 and Arp2/3 deficiency displayed severe defects in GPVI signaling,⁶ and platelet production as well as knockout efficiency,⁹ respectively. In this study, we provide for the first time definitive evidence that platelet lamellipodia formation is not required for stable thrombus formation *ex vivo* and *in vivo*, hemostatic plug formation as well as for maintenance of vascular integrity in inflammatory settings (Figures 5-7) by analyzing Cyfip1-deficient mice with abolished lamellipodia formation but only minimal side-effects in platelet function (Figures 1-4). Moreover, we demonstrate by direct observation that morphological changes of platelets differ between a static spreading assay and thrombus formation under flow, where platelets only formed filopodia to adhere to collagen fibers (supplemental Video 5) and to form a stable thrombus as visualized by scanning electron microscopy in an experimentally-induced arterial thrombus (Figure 7F). We could further show that control and mutant platelets in direct contact with collagen fibers at the bottom of the thrombus were flattened but formed filopodia structures with parallel actin filaments. Fully spread platelets with circumferential lamellipodia, as can be observed in the static spreading assay (phase IV), were absent under these conditions (Figure 6, supplemental Figure 7). However, although platelet lamellipodia are not required for the formation and overall stability of thrombi, we cannot exclude that lamellipodia formation might still be important in other thrombi-related processes, such as vessel repair.

Interestingly, it was previously demonstrated that Arp2/3 complex activation is normal in platelets lacking WASp and hypothesized that other proteins, probably WAVE2 or cortactin or both, activate the Arp2/3 complex.³⁶ We could show that Cyfip1-deficient platelets have a reduced WAVE2 expression (Figure 1A) and cannot form Arp2/3-dependent branched actin filaments (Figure 3A). Thus, our data strongly suggest for the first time that the WAVE2 protein/complex regulates Arp2/3 activity in platelets.

It was recently shown that collagen-adherent platelets can transform into PS-exposing balloon-like structures, which was termed procoagulant spreading and likely contributes to hemostatic and thrombotic responses *in vivo* by amplifying procoagulant responses.³⁷ We tested the ability of mutant platelets to expose PS on the surface after stimulation with CRP and thrombin *in vitro* and under dynamic conditions *ex vivo*, and obtained comparable results for control and mutant mice (supplemental Figure 8). Therefore, we suggest that platelets do not need to form lamellipodia in order to expose PS on the surface. Hence, the bridging mechanism of platelet and coagulation response seems to be intact in *Cyfip1*-deficient platelets, enabling them to initiate thrombus formation.

However, one open question remains: what is the role of lamellipodia in platelet function? There are several possibilities: (I) Lamellipodia structures could be an *in vitro* phenomenon during static platelet spreading. However, this is rather unlikely, since many other cells also produce this dense network of branched actin filaments contributing to important cellular functions. (II) The platelet machinery to form lamellipodia might be a relic of evolution no longer needed for the specialized functions platelets have, and thus dispensable. Until now, we cannot exclude either possibility, however, most likely is that (III) platelet lamellipodia play an important role in other processes besides the initiation of thrombus formation and thrombus stability. These membrane protrusions at the leading edge of cells drive cell migration in many physiological and pathological situations.³⁸ Recently, it was reported that platelets actively migrate at sites of a formed thrombus.³⁹ The authors concluded from their findings that platelet migration might be too slow to contribute to thrombus formation, but might play a role in thrombus reorganization and consolidation.³⁹ Our data of the *Cyfip1*^{-/-} mice show that platelet lamellipodia, which are most likely a prerequisite for platelet migration, are not important for thrombus formation, supporting the conclusion by Gaertner *et al.* of the dispensable role of platelet migration in thrombus growth. We did not directly analyze thrombus reorganization, however, we visualized single platelets in an experimentally-induced arterial thrombus and found that platelets exclusively form filopodia or filopodia-like structures (Figure 7F). Therefore, we suggest that at least lamellipodia are not important in the reorganization of a thrombus. Interestingly, Gaertner and colleagues could show that platelets also migrate to sites of infection in order to help trap bacteria and clear the vascular surface.³⁹ The authors used platelets from *Myh9*^{-/-} mice, which are unable to migrate,³⁹ however can extend lamellipodia but without stress fiber-like formation.⁴⁰ It will be interesting to investigate whether *Cyfip1*-deficient platelets with absent lamellipodia formation can migrate *in vitro* and *in vivo*, and whether *Cyfip1* is a key protein in bacteria scavenging and bundling at sites of infection.

In summary, we show that Cyfip1-deficiency does not alter platelet biogenesis and has no major impact on platelet activation, but Cyfip1 plays a crucial role for branching of actin filaments and consequently for lamellipodia formation *in vitro*. We took advantage of this unique mouse model and could demonstrate that lamellipodia formation is not required for the formation of a hemostatic plug or a thrombus. Moreover, Cyfip1-deficient mice could maintain vascular integrity at site of inflammation. Finally, we suggest that megakaryocyte-/platelet-specific Cyfip1-deficient mice represent a unique mouse system to study the role of platelet lamellipodia in other platelet-dependent processes *in vivo*. Further studies will be required to address this.

References

1. Vogtle T, Cherpokova D, Bender M, Nieswandt B. Targeting platelet receptors in thrombotic and thrombo-inflammatory disorders. *Hamostaseologie*. 2015;35(3):235-243.
2. Nieswandt B, Pleines I, Bender M. Platelet adhesion and activation mechanisms in arterial thrombosis and ischaemic stroke. *J Thromb Haemost*. 2011;9 Suppl 1:92-104.
3. Ono A, Westein E, Hsiao S, et al. Identification of a fibrin-independent platelet contractile mechanism regulating primary hemostasis and thrombus growth. *Blood*. 2008;112(1):90-99.
4. Hartwig JH. Mechanisms of actin rearrangements mediating platelet activation. *J Cell Biol*. 1992;118(6):1421-1442.
5. McCarty OJ, Larson MK, Auger JM, et al. Rac1 is essential for platelet lamellipodia formation and aggregate stability under flow. *J Biol Chem*. 2005;280(47):39474-39484.
6. Pleines I, Elvers M, Strehl A, et al. Rac1 is essential for phospholipase C-gamma2 activation in platelets. *Pflugers Arch*. 2009;457(5):1173-1185.
7. Aslan JE, McCarty OJ. Rho GTPases in platelet function. *J Thromb Haemost*. 2013;11(1):35-46.
8. Yusuf MZ, Raslan Z, Atkinson L, et al. Prostacyclin reverses platelet stress fibre formation causing platelet aggregate instability. *Sci Rep*. 2017;7(1):5582.
9. Paul DS, Casari C, Wu C, et al. Deletion of the Arp2/3 complex in megakaryocytes leads to microthrombocytopenia in mice. *Blood Adv*. 2017;1(18):1398-1408.
10. Alekhina O, Burstein E, Billadeau DD. Cellular functions of WASP family proteins at a glance. *J Cell Sci*. 2017;130(14):2235-2241.
11. Derivery E, Lombard B, Loew D, Gautreau A. The Wave complex is intrinsically inactive. *Cell Motil Cytoskeleton*. 2009;66(10):777-790.
12. Oda A, Miki H, Wada I, et al. WAVE/Scars in platelets. *Blood*. 2005;105(8):3141-3148.
13. Dahl JP, Wang-Dunlop J, Gonzales C, Goad ME, Mark RJ, Kwak SP. Characterization of the WAVE1 knock-out mouse: implications for CNS development. *J Neurosci*. 2003;23(8):3343-3352.
14. Calaminus SD, McCarty OJ, Auger JM, et al. A major role for Scar/WAVE-1 downstream of GPVI in platelets. *J Thromb Haemost*. 2007;5(3):535-541.
15. Eto K, Nishikii H, Ogaeri T, et al. The WAVE2/Abi1 complex differentially regulates megakaryocyte development and spreading: implications for platelet biogenesis and spreading machinery. *Blood*. 2007;110(10):3637-3647.
16. Gautreau A, Ho HY, Li J, Steen H, Gygi SP, Kirschner MW. Purification and architecture of the ubiquitous Wave complex. *Proc Natl Acad Sci U S A*. 2004;101(13):4379-4383.
17. Innocenti M, Zucconi A, Disanza A, et al. Abi1 is essential for the formation and activation of a WAVE2 signalling complex. *Nat Cell Biol*. 2004;6(4):319-327.
18. Steffen A, Rottner K, Ehinger J, et al. Sra-1 and Nap1 link Rac to actin assembly driving lamellipodia formation. *EMBO J*. 2004;23(4):749-759.
19. Rodriguez CI, Buchholz F, Galloway J, et al. High-efficiency deleter mice show that FLPe is an alternative to Cre-loxP. *Nat Genet*. 2000;25(2):139-140.
20. Tiedt R, Schomber T, Hao-Shen H, Skoda RC. Pf4-Cre transgenic mice allow the generation of lineage-restricted gene knockouts for studying megakaryocyte and platelet function in vivo. *Blood*. 2007;109(4):1503-1506.
21. Chen Z, Borek D, Padrick SB, et al. Structure and control of the actin regulatory WAVE complex. *Nature*. 2010;468(7323):533-538.
22. Davidson AJ, Insall RH. Actin-based motility: WAVE regulatory complex structure reopens old SCARs. *Curr Biol*. 2011;21(2):R66-68.
23. Pollitt AY, Insall RH. WASP and SCAR/WAVE proteins: the drivers of actin assembly. *J Cell Sci*. 2009;122(Pt 15):2575-2578.

24. Spindler M, van Eeuwijk JMM, Schurr Y, et al. ADAP deficiency impairs megakaryocyte polarization with ectopic proplatelet release and causes microthrombocytopenia. *Blood*. 2018.
25. Heilemann M, van de Linde S, Schuttpelz M, et al. Subdiffraction-resolution fluorescence imaging with conventional fluorescent probes. *Angew Chem Int Ed Engl*. 2008;47(33):6172-6176.
26. Kuijpers MJ, Munnix IC, Cosemans JM, et al. Key role of platelet procoagulant activity in tissue factor- and collagen-dependent thrombus formation in arterioles and venules in vivo differential sensitivity to thrombin inhibition. *Microcirculation*. 2008;15(4):269-282.
27. Munnix IC, Strehl A, Kuijpers MJ, et al. The glycoprotein VI-phospholipase Cgamma2 signaling pathway controls thrombus formation induced by collagen and tissue factor in vitro and in vivo. *Arterioscler Thromb Vasc Biol*. 2005;25(12):2673-2678.
28. Gros A, Syvannarath V, Lamrani L, et al. Single platelets seal neutrophil-induced vascular breaches via GPVI during immune-complex-mediated inflammation in mice. *Blood*. 2015;126(8):1017-1026.
29. Goerge T, Ho-Tin-Noe B, Carbo C, et al. Inflammation induces hemorrhage in thrombocytopenia. *Blood*. 2008;111(10):4958-4964.
30. Durrant TN, van den Bosch MT, Hers I. Integrin alphaIIb beta3 outside-in signaling. *Blood*. 2017;130(14):1607-1619.
31. Di Buduo CA, Wray LS, Tozzi L, et al. Programmable 3D silk bone marrow niche for platelet generation ex vivo and modeling of megakaryopoiesis pathologies. *Blood*. 2015;125(14):2254-2264.
32. Ito Y, Nakamura S, Sugimoto N, et al. Turbulence Activates Platelet Biogenesis to Enable Clinical Scale Ex Vivo Production. *Cell*. 2018;174(3):636-648 e618.
33. Moreau T, Evans AL, Vasquez L, et al. Large-scale production of megakaryocytes from human pluripotent stem cells by chemically defined forward programming. *Nat Commun*. 2016;7:11208.
34. Thon JN, Mazutis L, Wu S, et al. Platelet bioreactor-on-a-chip. *Blood*. 2014;124(12):1857-1867.
35. Kahr WH, Pluthero FG, Elkadri A, et al. Loss of the Arp2/3 complex component ARPC1B causes platelet abnormalities and predisposes to inflammatory disease. *Nat Commun*. 2017;8:14816.
36. Falet H, Hoffmeister KM, Neujahr R, Hartwig JH. Normal Arp2/3 complex activation in platelets lacking WASp. *Blood*. 2002;100(6):2113-2122.
37. Agbani EO, van den Bosch MT, Brown E, et al. Coordinated Membrane Ballooning and Procoagulant Spreading in Human Platelets. *Circulation*. 2015;132(15):1414-1424.
38. Krause M, Gautreau A. Steering cell migration: lamellipodium dynamics and the regulation of directional persistence. *Nat Rev Mol Cell Biol*. 2014;15(9):577-590.
39. Gaertner F, Ahmad Z, Rosenberger G, et al. Migrating Platelets Are Mechano-scavengers that Collect and Bundle Bacteria. *Cell*. 2017;171(6):1368-1382 e1323.
40. Leon C, Eckly A, Hechler B, et al. Megakaryocyte-restricted MYH9 inactivation dramatically affects hemostasis while preserving platelet aggregation and secretion. *Blood*. 2007;110(9):3183-3191.

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Author contributions

Y.S. performed experiments, analyzed data and wrote the manuscript. A.S., S.B., J.V., L.R., C.K., and P.E. performed experiments and analyzed data. Sh.B. generated the mice and analyzed data. M.S. and B.N. analyzed data. L.M. analyzed data and contributed to the writing of the manuscript. M.B. supervised research, performed experiments, analyzed data and wrote the manuscript.

Disclosure of conflict of interest

The authors declare no competing interests.

Figure legends

Figure 1. Moderately reduced activation of *Cyfp1*^{-/-} platelets. (A) Expression of indicated proteins in control and mutant platelets was assessed by Western blot analysis. GAPDH served as loading control (n=3). (B) Platelet count per μL and (C) platelet size given as mean forward scatter (FSC) was determined via flow cytometry (n=4, representative for four independent experiments). Values are mean \pm s.d. ***P<.001. (D) Flow cytometric determination of inside-out activation of the $\alpha\text{IIb}\beta\text{3}$ integrin (JON/A-PE antibody) and (E) degranulation-dependent P-selectin exposure (FITC-labelled anti-P-selectin antibody) in response to the agonists ADP, U46619 (thromboxane analogue), Thrombin (Thr), collagen related peptide (CRP), convulxin (CVX) and Rhodocytin (RC). (n=5, representative for four independent experiments); Values are mean \pm s.d. **P<.01, ***P<.001. (F) Washed platelets from *Cyfp1*^{+/+} and *Cyfp1*^{-/-} mice were activated with the indicated agonists. 2nd wave inhibitors: Indomethacin 10 μM , EDTA, Apyrase 2 U/ml (representative curve of at least six independent samples). (G) Determination of tyrosine phosphorylation after stimulation with 1 $\mu\text{g/ml}$ CRP under stirring conditions (1000 rpm) at 37°C. Fifty-microliter aliquots were taken at the indicated time points. Samples were blotted on a PVDF membrane and stained using the 4G10 antibody. GAPDH served as loading control (n=3).

Figure 2. *Cyfp1*-deficient platelets are unable to form lamellipodia on a fibrinogen-coated surface. (A) Washed platelets were allowed to spread on fibrinogen, following fixation at the indicated time points. (n=3, representative for three independent experiments). Scale bar: 5 μm . (B) Quantification of the different spreading phases of fixed platelets after 5, 15 and 30 minutes from differential interference contrast images (Zeiss Axiovert 200 inverted microscope (100x/1.4 oil objective, Zeiss)). (C) Phase abundance of live cells over time (WT n=58; KO n=32). Quantification of (D) platelet size and (E) platelet perimeter (WT n=100, KO n=58) of all *Cyfp1*^{+/+} and *Cyfp1*^{-/-} platelets present in a visual field after 30 min spreading. (F) Length and (G) number of filopodia (WT n=58; KO n=32) of live platelets in phase II spreading. Values are mean \pm s.d. ***P<.001.

Figure 3. *Cyfp1*-deficient platelets cannot form branched actin filaments. (A) Platinum replica electron microscopic images of the cytoskeleton ultrastructure of platelets spread on fibrinogen after 15 min; Scale bars: 1 or 4 μm . (B) F-actin content under resting conditions and upon stimulation with CRP or thrombin was assessed by measuring the mean fluorescence intensity of bound FITC labeled phalloidin in flow cytometry (left). Ratio of mean fluorescent intensity of activated vs. resting samples reflecting F-actin polymerization rates (right). (n=4, representative for three independent experiments); Values are mean \pm s.d. *P<.05.

Figure 4. Abolished lamellipodia formation of mutant platelets on different matrices.

(A) Washed platelets were allowed to spread on collagen IV, laminin or CRP for 30 min. (n=3) Scale bars: 5 μ m. (B) Images of the platelet cytoskeleton ultrastructure on CRP after 15 min. (n=5) Scale bars: 1 μ m.

Figure 5. Normal *ex vivo* thrombus formation and clot retraction of Cyfip1-deficient platelets.

(A) Thrombus formation was assessed on collagen at different shear rates. Shown are representative fluorescence pictures of platelets stained with a Dylight-488 anti-GPIX antibody. (n=5, representative for three independent experiments) Scale bar: 30 μ m. (B) Surface coverage and (C) relative thrombus volume were quantified. (D) Clot formation was observed over time and (E) residual serum volume was measured (n=3, representative for two independent experiments).

Figure 6. Flattened platelets with filopodia-like structures at the bottom of the thrombus.

(A) Scanning electron microscopic image of platelets after experimentally-induced *ex vivo* thrombus formation on collagen. White arrow indicates a filopodium, black arrowhead indicates collagen fiber; Scale bar: 4 μ m. (B) Platinum replica of platelets after experimentally-induced *ex vivo* thrombus formation on collagen. White arrow indicates a filopodium with unbranched actin filaments; Scale bars: 1 and 2 μ m.

Figure 7. Lamellipodia formation is not required for hemostatic function and arterial thrombus formation.

(A) A 2-mm segment of the tail tip was cut, and bleeding was determined to have ceased when no blood drop was observed on the filter paper. Each symbol represents 1 individual. (B) Mesenteric arterioles were treated with 20% FeCl₃, and adhesion and thrombus formation of fluorescently labeled platelets were monitored by *in vivo* fluorescence microscopy. Representative images are shown. (C) Statistical evaluation of the time to appearance of a first thrombus and (D) time to occlusion. (E) The abdominal aorta was mechanically injured using forceps (compression for 15 s), and blood flow was monitored. Each symbol represents one individual. (F) Scanning electron microscopy of an *in vivo* formed thrombus. (n=4); Scale bar: 2 μ m. (G-H) Mice were subjected to the rpA reaction to induce local skin inflammation. (G) Representative images are shown and (H) Hb content in tissue punch biopsies from inflammatory spots was quantified (n=4, representative for three independent experiments). (I) Representative images of bronchoalveolar lavage liquid at 4 hours after LPS application. (J) Quantification of the Hb content in bronchoalveolar lavage liquid. Platelet depleted animals served as positive control of inflammatory bleeding in both models (n=5, representative for three independent experiments).