Red trap colour of the carnivorous plant Drosera rotundifolia does not serve a prey
attraction or camouflage function.
Foot, G^1 , Rice SP ² , Millett, J^{2*}
¹ School of Life Sciences, The University of Warwick, Coventry, CV4 7AL, UK
² Centre for Hydrological and Ecosystem Science, Department of Geography, Loughborough
University, Loughborough, LE11 3TU, UK
*Corresponding author: email: j.millett@lboro.ac.uk
Short title: Drosera trap colour and prey attraction

17 Summary

18 The traps of many carnivorous plants are red in colour. This has been widely hypothesised to serve a prey attraction function; colour has also been hypothesised to function as camouflage, 19 20 preventing prey avoidance. We tested these two hypotheses *in-situ* for the carnivorous plant Drosera rotundifolia. We conducted three separate studies: (i) prey attraction to artificial 21 traps to isolate the influence of colour; (ii) prey attraction to artificial traps on artificial 22 backgrounds to control the degree of contrast; and (iii) observation of prey capture by D. 23 rotundifolia to determine the effects of colour on prey capture. Prey were not attracted to 24 green traps and were deterred from red traps. There was no evidence that camouflaged traps 25 26 caught more prey. For *D. rotundifolia* there was a relationship between trap colour and prey capture. However, trap colour may be confounded with other leaf traits. Thus we conclude 27 that for D. rotundifolia red trap colour does not serve a prey attraction or camouflage 28 29 function.

Key words: plant-insect interactions; leaf colour; carnivorous plants; *Drosera rotundifolia*;
prey attraction.

32 Introduction

33 Carnivorous plants attract, trap and digest animal prey, utilising the nutrients gained to enhance fitness [1,2]. To attract prev carnivorous plants use a variety of mechanisms such as 34 olfactory [3], nectar [4,5] and visual cues [6]. Red colouration has been widely hypothesised 35 as a visual cue used to lure potential prey, by increasing contrast with the background [7-10]. 36 However, experimental evidence is limited and hampered by methodological issues such as a 37 lack of ecological relevance (see [5,10]) or confounding of attraction and capture mechanisms 38 (for example in [11,12]). As a result, prey attraction to red carnivorous plant traps has not yet 39 been conclusively demonstrated for any species. 40

In this study we investigated *in-situ* the role of red colour in attracting prey onto the adhesive 41 42 traps of the carnivorous plant Drosera rotundifolia. Drosera rotundifolia grows on Sphagnum hummocks on ombrotrophic bogs. Their leaves form small rosettes (c. 5 cm in diameter) and 43 catch prey (predominantly Diptera and Collembola [13]) using sticky mucilage secreted on 44 the end of stalked glands on the leaf. Attraction of prey to these traps has not yet been 45 demonstrated, but their leaves are a distinctive red colour, which has been hypothesised to 46 serve a prey capture function [7,8]. This might be through direct attraction of prey or possibly 47 by camouflaging the trap, as suggested by Jürgens *et al.* [3]; this might be of benefit if prey 48 capture is a passive process where a conspicuous trap might deter prey. We tested the 49 50 hypotheses that: 1) Potential prey are attracted to red coloured traps and 2) that traps that are more cryptic will be more successful at trapping prey. We also evaluated prey capture of 51 differently coloured D. rotundifolia leaves to establish whether any differences are observed 52 naturally. 53

54 Material and methods

Over a three week period during August and September 2012 we conducted three separate 55 studies on an ombrotrophic (rain-fed) raised bog at Thorne Moors, England (53°37'55"N, 56 $0^{\circ}54'21''W$). We used an area of approx. 100 m² with abundant *D. rotundifolia* growing in a 57 Sphagnum substrate. The three experiments were interspersed across the study area, but 58 separated by at least 5 m to avoid interference. We constructed artificial traps by printing red 59 and green *D. rotundifolia* shaped images (four 1.7×1.2 cm leaves per trap) onto photopaper 60 (Fig. S1). Traps were laminated and covered with non-setting adhesive (OecoTak A5, Oecos, 61 Hertfordshire, UK). Neutral coloured traps were prepared by excluding the coloured images 62 (i.e. only laminate and glue), these traps were translucent rather than transparent. The colour 63 of artificial traps mostly fell within the natural range of *D. rotundifolia* at the same site (Table 64 1). After seven days the traps were removed and replaced with new traps in the same 65

66 locations. This was repeated three times for all artificial and natural traps. Captured prey

67 items were identified to order level. The length of each prey item was measured under a

68 binocular microscope to determine treatment impacts on prey size as well as number.

69 *Experiment 1: are prey attracted to green or red artificial traps?*

70 Red, green and neutral artificial traps were placed together (one of each) in random locations

71 (n=12 in total) on hummocks throughout the study area (see Fig. S1).

72 *Experiment 2: is prey attraction a result of trap contrast against its background?*

73 Artificial backgrounds were constructed using the same red and green colouration as the

real artificial traps, printed onto A4 sized standard photo paper and then laminated. Three red and

three green artificial traps were stapled onto each red or green background. Six red and six

76 green backgrounds were randomly distributed in pairs (one red and one green) throughout the

study area resulting in a 2×2 split-plot design.

78 *Natural observations: does* D. rotundifolia *leaf colour influence prey capture?*

60 plants were randomly selected and labelled, before removing all captured arthropods.
Plants were photographed to determine the colour of each leaf and the immediate background
(a 5 × 5 cm square centred on the plant). All the leaves on each plant were measured and prey
capture recorded; data were pooled to give mean values for each plant. Leaf area was
estimated based on measurements of length and width, assuming an ellipsoidal shape. After
seven days the plants were revisited and all captured arthropods removed with tweezers, this
was repeated twice more

86 *Colour analysis*

Leaf colour in CIE 1976 (*L*a*b**) colour space (CIELAB) was determined to enable
assessment of colour difference and how red or green a leaf is (based on *a**). Colour
differences between leaves and background (conspicuousness) were determined by

90 calculating the delta-E 1976 (ΔE_{76}) (details of colour analysis are in Electronic Supplementary 91 Material).

92 Statistical analysis

93 Prey count and length data were pooled within plants and analysed in IBM SPSS Statistics for

94 Windows v.20.0 [14]. Initial exploration using repeated measures ANOVA showed no

statistically significant interaction between sampling date and any of the experimental

96 treatments. Therefore, data from the three sampling dates were pooled for all analyses.

97 Experiment 1 was analysed using a one-way ANOVA, experiment 2 was analysed as a split-

98 plot design. Differences between treatments were assessed with Fisher's LSD.

99 Homoscedasticity was tested using plots of residuals and normality was tested using normal 100 probability plots. The observational study was analysed using multiple logistic regression to 101 determine the impact of the measured leaf variables on the probability of prey capture, and 102 correlation to determine the relationships between other measured variables. Differences in 103 the taxa of captured prey were analysed using Pearson's chi-squared test.

104 **Results**

105 For all trap types Diptera were the most common arthropod order caught (57-84% of total),

106 followed by Hymenoptera (7.5-17.4%) then Collembola (5.8-16.5%) (Fig. 1, χ^2 =1273.84,

107 n=705, d.f.=4, P<0.001). Capture rates differed between trap types ($\chi^2 = 173.58$, n=70, d.f.=7,

108 P < 0.001): traps in experiment 1 (103-139 arthropods/seven days) and natural traps (121

arthropods/seven days) caught more than traps in experiment 2 (32-53 arthropods/seven

110 days). There was a significant interaction between trap type and the distribution of prey taxa

111 (Fig. 1, χ^2 =50.10, *n*=705, d.f.=28, *P*=0.01), due to small differences in capture by natural

112 compared with artificial traps (i.e. Diptera being a smaller component and Hymenoptera and

113 Collembola being a larger component of natural traps).

114 Experiment 1: Artificial traps

- 115 Green and clear artificial traps caught significantly more prey than red traps ($F_{2,33}$ =4.807,
- 116 P < 0.001; Fig. 2a) with no significant difference in the length of the captured invertebrates

117 apparent ($F_{2,33}=0.676$, P=0.516).

- 118 Experiment 2: artificial background
- 119 Trap and background colour significantly affected prey capture rates and the interaction
- between the two was statistically significant (trap colour $F_{1,5}=12.707$, P=0.005; background
- 121 colour $F_{1,5}=7.709$, P=0.020; interaction $F_{1,10}=11.949$, P=0.006). Red traps on red
- backgrounds attracted fewest prey; other combinations did not differ (Fig. 2b). There was no
- 123 significant impact of trap or background colour on the length of prey captured (trap colour –
- 124 $F_{1,34}=0.599$, P=0.457; background colour $-F_{1,10}=0.019$, P=0.892; interaction $-F_{1,10}=3.792$,
- 125 P=0.080).
- 126 Experiment 3: D. rotundifolia
- 127 Plants with redder leaves had a higher probability of prey capture success (Logistic
- regression: Wald(1)=7.052, P=0.008, B±SE=0.059±0.022), as did plants with more leaves
- 129 (Logistic regression: Wald(1)=3.880, P=0.49, $B\pm SE=0.249\pm 0.127$). There was no impact of
- 130 leaf size (Logistic regression: Wald(1)=1.803, P=0.179) or conspicuousness (ΔE_{76})
- 131 (Wald(1)==1.429, P =0.232). (χ^2 (1)=13.793, P=0.008 R^2_{CS} =0.085, R^2_{N} =0.113.)
- 132 There were significant positive correlations between leaf colour (a^*), ΔE_{76} and leaf area, and a
- significant negative correlation between leaf area and the number of leaves on a plant (Table
- 134 2). There was no correlation between the number of leaves and trap colour or ΔE_{76} . For those
- 135 leaves that captured prey, plants with fewer and larger leaves caught more and larger prey, but
- leaf area and number were both negatively correlated with prey capture efficiency (Table 2).
- 137 For these plants there was no correlation between trap colour or conspicuousness and any of

the measures of prey capture, with the exception that more conspicuous traps were lessefficient at catching prey.

140 **Discussion**

141 In our study a red colouration did not fulfil a prey attraction function or serve as advantageous camouflage in D. rotundifolia, as has been previously assumed or suggested [3,7,8]. Instead 142 red colouration might deter potential prey. This is not entirely surprising. Red detection is 143 more difficult for species without red receptors than for those with red receptors [15]. It is 144 unlikely that the red colouration of carnivorous plant traps would evolve as a visual cue to 145 attract prey, unless they capture ecologically significant numbers of these 'red-sensitive' prey. 146 147 Diptera do not possess red receptors [16] and were the most abundant prey species for the D. 148 rotundifolia studied here. There would therefore be no likely ecological advantage of 149 attempting to attract these prey using red traps.

150 The covariation among the various trap characteristics and prey capture suggests trade-offs in terms of investment in traps. Leaf size/number trade-offs have been demonstrated both 151 between and within species [17,18]. We found a leaf size/area trade-off for D. rotundifolia. 152 In addition, having fewer, smaller leaves appears to be the most efficient (i.e. prev capture per 153 unit trap area) way to capture prey, though having more leaves increases the probability of 154 any prey capture at all. This benefit will be balanced against the other benefits of small or 155 156 large leaves, so in D. rotundifolia and maybe other carnivorous plants we might expect this trade-off to alter in relation to resource availability. 157

Artificial and natural trap colour was coincident and they caught similar prey, supporting the
use of artificial traps as a surrogate for natural traps. Interestingly, redder natural *D*. *rotundifolia* traps had a greater likelihood of prey capture than greener traps. The artificial
traps measured true colour attraction but prey presence on the traps of *D. rotundifolia* is a

162	measure of both prey attraction (not limited to colour attraction) and capture, with potential
163	for confounding of factors. Reduced prey attraction for red traps on red backgrounds is likely
164	to be a consequence of potential prey being deterred from red (due to the large area of red
165	present in these experimental units) rather than an explicit effect of the degree of crypsis.
166	Additionally, we did not explicitly test the role of UV reflection which might play a role in
167	prey attraction [6]. The prey captured by <i>D. rotundifolia</i> in our study are able to perceive UV
168	[16] and there may be variation in UV reflectance of red and green traps not accounted for
169	with the artificial traps. Thus UV pigmentation might have a prey attraction function that we
170	could not detect.
171	Acknowledgements
172	We thank Julian Small of Natural England for site access and Lucy Kitcher for fieldwork
173	assistance. Many thanks to Aaron Ellison and two anonymous reviewers for their useful and
174	constructive advice.
175	Data accessibility
176	Data deposited in Dryad Digital Repository: http://dx.doi.org/10.5061/dryad.h7s03 [19]
177	Electronic Supplementary Material
178	Figure S1: Photograph of artificial trap deployment.
179	ESM1: Details of colour analysis.
180	References
181 182	1 Juniper, B. B. E., Robins, R. J. & Joel, D. M. 1989 <i>The Carnivorous Plants</i> . Academic Press. [cited 2013 Oct. 28].
183 184	2 Ellison, A. M. & Gotelli, N. J. 2001 Evolutionary ecology of carnivorous plants. <i>Trends Ecol. Evol.</i> 16 , 623–629.
185 186	Jürgens, A., El-Sayed, A. M. & Suckling, D. M. 2009 Do carnivorous plants use volatiles for attracting prey insects? <i>Funct. Ecol.</i> 23 , 875–887. (doi:10.1111/j.1365-

187 2435.2009.01626.x)

Bauer, U., Willmes, C. & Federle, W. 2009 Effect of pitcher age on trapping efficiency 188 and natural prey capture in carnivorous Nepenthes rafflesiana plants. Ann. Bot. 103, 189 1219–26. (doi:10.1093/aob/mcp065) 190 5 Bennett, K. F. & Ellison, A. M. 2009 Nectar, not colour, may lure insects to their 191 192 death. Biol. Lett. 5, 469–72. (doi:10.1098/rsbl.2009.0161) Joel, D., Juniper, B. & Dafni, A. 1985 Ultraviolet patterns in the traps of carnivorous 193 6 plants. New Phytol. 101, 585–593. 194 Lloyd, F. E. 1942 The Carnivorous Plants. The Chronica Botanica Company. 195 7 Ichiishi, S., Nagamitsu, T., Kondo, Y., Iwashina, T., Kondo, K., Tagashira, N. & 196 8 Orikazu 1999 Effects of macro-components and sucrose in the medium on in vitro red-197 colour pigmentation in Dionea muscipula Ellis and Drosera spathulata Labill. Plant 198 Biotechnol. 16, 235–238. 199 9 Jaffe, K., Michelangeli, F., Gonzalez, J. M., Miras, B. & Ruiz, M. C. 1992 Carnivory in 200 pitcher plants of the genus Heliamphora (Sarraceniaceae). New Phytol. 122, 733-744. 201 10 Schaefer, H. M. & Ruxton, G. D. 2008 Fatal attraction: carnivorous plants roll out the 202 red carpet to lure insects. *Biol. Lett.* **4**, 153–5. (doi:10.1098/rsbl.2007.0607) 203 11 Pavlovic, A., Krausko, M., Libiaková, M. & Adamec, L. 2014 Feeding on prey 204 increases photosynthetic efficiency in the carnivorous sundew Drosera capensis. Ann. 205 206 Bot. 113, 69–78. (doi:10.1093/aob/mct254) 12 Cresswell, J. E. 1993 The morphological correlates of prey capture and resource 207 parasitism in pitchers of the carnivorous plant Sarracenia purpurea. Am. Midl. Nat. 129, 208 35-41. 209 210 13 Ellison, A. M. & Gotelli, N. J. 2009 Energetics and the evolution of carnivorous plants--Darwin's "most wonderful plants in the world". J. Exp. Bot. 60, 19-42. 211 (doi:10.1093/jxb/ern179) 212 213 14 IBM Corp. 2011 IBM SPSS Statistics for Windows. Chittka, L., Spaethe, J., Schmidt, A. & Hickelsberger, A. 2001 Adaptation, constraint, 15 214 and chance in the evolution of flower color and pollinator color vision. In Cognitive 215 Ecology of Pollination (eds L. Chittka & J. Thompson), pp. 106–126. Cambridge 216 University Press. [cited 2013 Nov. 12]. 217 16 Briscoe, A. D. & Chittka, L. 2001 The evolution of color vision in insects. Annu. Rev. 218 *Entomol.* **46**, 471–510. (doi:10.1146/annurev.ento.46.1.471) 219

- 17 Whitman, T. & Aarssen, L. W. 2009 The leaf size/number trade-off in herbaceous 220 221 angiosperms. J. Plant Ecol. 3, 49–58. (doi:10.1093/jpe/rtp018)
- 18 Kleiman, D. & Aarsen, L. W. 2007 The leaf size/number trade-off in trees. J. Ecol. 95, 222 376–382. (doi:10.1111/j.1365-2745.2006.01205.x) 223

Foot, G., Rice, S.. & Millett, J. 2014 Data from: Trap colour of the carnivorous plant
Drosera rotundifolia does not serve a prey attraction or camouflage function. Dryad
Digital Repository. http://dx.doi.org/10.5061/dryad.h7s03

227	Table 1.
228	Colour characteristics of artificial traps, artificial traps on artificial backgrounds, and D.
229	rotundifolia leaves.
230	Table 2.
231	Results of Pearson's Correlation between leaf characteristics for all plants, and between leaf
232	characteristics and prey capture for those plants that captured prey.
233	Figure 1.
234	Proportion of total arthropod capture in each taxa for (A) D. rotundifolia leaves, (B) red,
235	green and clear artificial traps, and (C) artificial red and green traps on red and green artificial
236	backgrounds. Numbers above the bars indicate the number of prey in each group.
237	Figure 2.
238	Numbers of arthropods captured by (A) artificial traps on artificial backgrounds and (B) red,
239	green and clear artificial traps. Bars show mean±se. Different letters above bars indicate
240	significant differences (Fisher's LSD).
241	
242	
243	
244	
245	
246	
247	
248	
249	
250	
251	

Table 1.

		Colour (CIELAB a*) [†]			Colour difference $(\Delta E_{76})^{\ddagger}$		
		Min	Max	Mean	Min	Max	Mean
D. rotundifolia traps		-12.1	47.5	22.7	10.2	56.2	32.9
D. rotundifolia background.		-12.2	24.6	-1.0	-	-	-
Artificial trans on natural	Red	14.7	45.1	27.2	34.6	67.4	46.6
Artificial traps on natural	Green	-15.1	-6.4	-10.4	18.0	73.8	56.2
background	Clear	-3.2	5.8	0.6	18.6	60.5	46.7
	Red on green	-11.5 (b	ackgrou	nd colour)	43.8	53.8	48.8
	Red on red	43.1 (background colour)			10.1	12.9	11.7
Artificial traps on artificial background	Green on green	-11.5 (background colour)			7.6	11.6	9.2
	Green on red	43.1 (b	ackgrour	nd colour)	65.5	71.3	69.0

[†]negative values indicate a green colouration, and positive a red colouration. [‡]larger values indicate greater colour contrasts.

Table 2

		Area	Colour	Leaves	ΔE_{76}
All plants					
Area	Pearson Correlation		0.175	-0.177	0.21
	Р		0.029	0.027	0.00
Calaura	Pearson Correlation			0.027	0.652
Colour	Р			0.742	0.00
Learnes	Pearson Correlation				0.09
Leaves	Р				0.240
Only plants that c	aptured prey				
Prey number	Pearson Correlation	0.267	0.180	-0.501	-0.01
	Р	0.016	0.108	<0.001	0.874
Duran 1	Pearson Correlation	0.212	0.145	-0.283	0.004
Prey length	Р	0.058	0.195	0.010	0.97
Capture efficiency	Pearson Correlation	-0.375	0.020	-0.397	-0.27
	Р	0.001	0.862	<0.001	0.01

prey captured per plant, Prey length=mean length of captured prey, Capture efficiency=umber
of prey captured per unit leaf/trap area.



