

Production of molecularly imprinted polymer particles with amide-decorated cavities for CO₂ capture using membrane emulsification/suspension polymerisation

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1. INTRODUCTION

The increased level of CO₂ in the atmosphere due to excessive combustion of fossil fuels is one of the main causes of global warming and climate change [1,2]. Capture of CO₂ from large point sources and sequestration in geological formations is considered as the most viable short-term solution for reducing the level of CO₂ in the atmosphere [3]. Currently, post-combustion carbon capture by amine scrubbing is the most established technology for the removal of CO₂ from flue gases [4]. However, amine solutions are corrosive and degrade into toxic products after repetitive regeneration cycles at elevated temperatures [5,6]. In addition, amine scrubbing systems exhibit a low specific interfacial area and require high regeneration energy, which can reduce energy output of the plant by 25-40% [7,8].

Solid adsorbents can be attractive alternatives, because they are nontoxic, nonvolatile and noncorrosive, require lower regeneration energy, and display a higher surface area-to-volume ratio [7,9]. Zeolites can have a high CO₂ capture capacity and high CO₂-over-N₂ selectivity under atmospheric conditions [10,11]. However, the presence of moisture in flue gases greatly reduces their capture capacity and requires higher regeneration temperatures, resulting in higher energy penalties [12].

Metal organic frameworks (MOFs) show high CO₂ capture capacity at elevated pressures [13], but under typical conditions of post-combustion carbon capture, their capture capacity is reduced, especially in the presence of moisture, NO_x and SO_x [10,14]. Carbonaceous materials are cheap and stable in the presence of moisture and impurities [15]. However, they display a low selectivity for CO₂ over N₂ due to the physisorption mechanism of CO₂ capture [2]. One of the main drawbacks of highly porous materials such as activated carbon and MOFs is their low density, which can limit their application in fluidised bed systems [15,16].

25 Physical impregnation or covalent tethering of amines inside mesopores is an effective way
26 of increasing both CO₂ capture capacity and selectivity of porous CO₂ adsorbents [17,18].
27 CO₂ has a higher affinity towards polar amine or amide groups than other flue gases, due to
28 its larger quadrupole moment and polarisability [19].

29 Polymer-based materials, such as hyper cross-linked polymers (HCPs), porous aromatic
30 frameworks (PAFs), and covalent organic polymers (COPs) are new classes of CO₂
31 adsorbents characterised by a high CO₂ selectivity and capture capacity, high hydrothermal
32 stability and ease of surface modification [20–22]. Acrylamide-based molecularly imprinted
33 polymer (MIP) particles for CO₂ capture with a separation factor of up to 340 at a CO₂ partial
34 pressure of 15 kPa have been fabricated using bulk polymerisation [23]. MIPs contain
35 inherently functionalised nanocavities, which are complementary in shape to the target
36 molecule, and can act as active sites for capturing the target molecules (Fig. 1). Unlike amine
37 impregnation or tethering, which often leads to reduction in the total pore volume and
38 specific surface area of the particles [24], molecular imprinting increases porosity of the
39 particles, leading to a higher rate of diffusion of CO₂ to active sites [23,25].

40 However, bulk polymerisation is not suitable for large-scale production, because the resulting
41 bulk polymer must be crushed, ground, and sieved to obtain particles of optimum size, which
42 is time-consuming, laborious, and expensive, as only 30-40% of the particles can be
43 recovered. In addition, the produced particles have irregular shape and sharp edges and are
44 prone to attrition [25]. Proper shape, size, and surface morphology of the particles are the key
45 factors for the applicability of adsorbents. Adsorbents with very fine particles cannot be
46 directly used in industrial CO₂ capture systems. In a fixed-bed system, fine particles cause
47 excessive pressure drop through the bed, whereas in fluidised and moving bed systems, they
48 can readily be entrained in fluidising gas and are prone to plugging, channelling, and
49 agglomerating [26,27]. Because of these limitations, fine particles are usually used in

50 palletised form, which can block their active sites and reduce CO₂ capture capacity and
51 capture rate.

52 In suspension polymerisation, each individual monomer droplet represents a miniature batch
53 reactor leading to higher rates of heat transfer and shorter polymerisation times compared
54 with bulk polymerisation [28]. Since crushing and grinding steps are not involved, higher
55 particle yields can be achieved and synthesised particles are regular spheres due to the
56 spherical shape of the monomer droplets. However, traditional “top-down” emulsification
57 methods, e.g. mixing in a stirred tank, typically lead to highly polydispersed droplets whose
58 size cannot easily be controlled. Membrane emulsification is a “bottom-up” approach based
59 on injection of one liquid through a microporous membrane into another immiscible liquid
60 phase, leading to generation of uniform droplets [29]. Continuous membrane emulsification
61 systems enable large-scale production and can involve oscillatory (pulsed) flow of the
62 continuous phase [29,30] or nonstationary membrane, such as rotating [31] or vibrating
63 [32,33] membrane. Membrane emulsification followed by suspension polymerisation has
64 already been used for preparation of porous and nonporous, coherent and structured, and
65 homogeneous and heterogeneous polymeric particles from various monomers such as
66 styrene, methacrylic acid, dimethylaminoethyl methacrylate, 2-hydroxyethyl methacrylate,
67 methyl methacrylate, and methyl acrylate [34,35].

68 In this study, membrane emulsification/suspension polymerisation method will be explored
69 for fabrication of spherical molecularly imprinted poly[acrylamide-co-(ethyleneglycol
70 dimethacrylate)] particles with amide-decorated cavities for CO₂ capture. The particles will
71 be characterised by their thermal stability, surface morphology, porosity, and CO₂ capture
72 capacity. The material is highly selective to CO₂ due to shape specificity of the cavities and
73 the presence of amide groups covalently incorporated within the 3D network of the polymer.

2. EXPERIMENTAL SECTION

2.1. Materials

Oxalic acid (OA), acrylamide (AAM), acetonitrile (AN), toluene (TL), methanol, and 0.1 M hydrochloric acid were purchased from Fisher Scientific (UK). Ethylene glycol dimethacrylate (EGDMA), azobisisobutyronitrile (AIBN), and polyvinyl alcohol (PVA, $M_w = 13,000\text{--}23,000$, 87–89% hydrolysed) were purchased from Sigma Aldrich (UK). All reagents were of analytical reagent grade. Reverse osmosis (DI) water was supplied using Millipore 185 Milli-Q Plus apparatus. All gases were supplied by BOC (UK) with purity higher than 99.999%.

2.2. Particle Synthesis

The MIP particles were synthesised through the following five steps:

Step 1. Monomer-template self-assembly. A crucial first step in the molecular imprinting process is the self-assembly of functional monomer and template in a pre-polymerised organic phase (Fig. 1a), which enables creation of template-shaped cavities within the polymer matrix. Since CO_2 has a very low solubility in organic solvents under ambient conditions, oxalic acid, a structural analogue of two CO_2 molecules, was used as a dummy template. AAM was selected as a functional monomer, due to high affinity of amide groups towards CO_2 . Monomer-template self-assembly occurred in a 50/50 mixture (by volume) of TL and AN using a mass ratio of AAM to OA of 2.27/1. The mass-to-volume ratio of AAM to porogenic solvents (AN and TL) was 1/11.76. The mixture was stirred for 2 h before EGDMA and AIBN were added as a crosslinker and initiator, respectively. The mass ratio of AAM to EGDMA and AAM to AIBN in the organic phase was 1/4.66 and 17/1, respectively. The composition of the organic phase was optimised in our preliminary investigation.

99 **Step 2. Membrane emulsification.** The oil-in-water (O/W) emulsion was produced using a
100 commercial Micropore Dispersion Cell (MDC) equipped with a flat disc membrane and a
101 paddle-blade stirrer driven by a 24 V DC motor (Fig. 2a). The organic phase was injected at a
102 constant flow rate through the membrane into 50 mL of a 0.5 wt% aqueous solution of PVA
103 using a syringe pump (World Precision Instruments, Sarasota, US). The paddle stirrer fitted
104 above the membrane provided an adjustable shear on the membrane surface to control the
105 droplet size. A nickel membrane containing hexagonal arrays of cylindrical pores with a
106 diameter of 20 μm and a pore spacing of 200 μm was fabricated by the LIGA (Lithographie,
107 Galvanik und Abformung) process, which is based on lithography, nickel electroplating and
108 moulding. The membrane and MDC were supplied by Micropore Technologies Ltd (Redcar,
109 UK).

110 Once the desired amount of oil phase passed through the membrane, the emulsion droplets
111 were transferred to the reactor and polymerised. The membrane was cleaned by immersing in
112 a 15 vol% NaOH for 10 min, followed by 10 min soaking in 7 vol% citric acid and 20 min
113 treatment with DI water in an ultrasonic bath at 30°C.

114 **Step 3. Suspension polymerization.** The O/W emulsion produced in Step 2 was poured into a
115 preheated 500-mL jacketed reactor equipped with a four-neck lid and a four-blade impeller
116 with a diameter of 50 mm (Fig. 2b). The polymerisation took 3 h at an agitation rate of 75
117 rpm and 60°C. The temperature was controlled using a water recirculating heater/chiller
118 system. The emulsion was purged with N₂ for 10 min prior to the reaction, after which
119 nitrogen blanketing was used to prevent the presence of oxygen within the reaction mixture.

120 **Step 4. Surfactant removal.** After polymerisation, the suspension was filtered using a
121 Buchner funnel apparatus with a filter paper (Gard 3, Whatman 6 μm), and the particles were
122 washed with DI water at 85-90°C for 5 min in a water bath while stirring. The washing was

123 repeated 8-10 times until PVA was completely removed from the particles. If the surfactant
124 was not completely removed, the particles were highly agglomerated. The particle
125 agglomeration was investigated by taking SEM images of the washed particles. After
126 complete removal of the surfactant, the particle agglomeration was negligible. Once the
127 washing procedure was optimised, SEM imaging was not necessary.

128 ***Step 5. Template removal.*** OA was extracted from the particles by washing the sample with a
129 10/90 mixture (by volume) of hydrochloric acid and methanol, until no OA was detected in
130 the rinses by a Lambda 35 UV/VIS spectrometer (PerkinElmer, US). Finally, the particles
131 were washed with DI water and dried overnight in a vacuum oven at 80°C. The removal of
132 OA from the polymer matrix created cavities decorated with CONH₂ groups capable of
133 binding CO₂ via Lewis acid-base type interactions (Fig. 1d). It should be noted that the
134 cavities created by molecular imprinting differ from the pores formed by phase separation
135 between polymer and solvent during polymerisation. The size of the cavities is similar to that
136 of the template molecules (< 1 nm), while the pores are larger and lie in a mesoscale range
137 (2-50 nm).

138

139 **2.3. Material Characterization**

140

141 ***Scanning Electron Microscopy (SEM).*** The particle morphology was evaluated using a
142 TM3030 bench-top Scanning Electron Microscope (Hitachi, Tabletop Microscope Europe)
143 operating at an accelerating voltage of 15 keV. The accumulation of electrostatic charges on
144 the particles was prevented by coating the samples with gold/palladium (80/20) prior to SEM
145 measurements. The sputter coating speed was 0.85 nm per second at an applied voltage of 2
146 kV and a plasma current of 25 mA.

147 **Pore size analysis.** The pore size analysis of the samples was performed using an ASAP 2020
148 Micromeritics Accelerated Surface Area and Porosimetry system at 77 K within a liquid
149 nitrogen bath. Prior to each measurement, the samples were degassed at 80°C overnight. The
150 Brunauer–Emmett–Teller (BET) model was used to estimate the specific surface area over a
151 range of relative pressures, P/P_0 , of 0.06-0.3. The Barrett-Joyner-Halenda (BJH) model was
152 used to determine the pore size distribution, and the total pore volume was obtained from the
153 amount of N_2 adsorbed at P/P_0 of 0.99.

154 **Thermal analysis.** The thermal stability of the samples was investigated using a
155 thermogravimetric analyser (TGA) (Q5000 IR, TA Instruments, US). In each test, 10-20 mg
156 of the sample was heated from 50 to 600°C at a ramp rate of 10°C/min under a nitrogen flow
157 rate of 20 mL/min.

158 **Density measurement.** The particle density was measured using a helium pycnometer
159 (Micromeritics, US), over five runs and the average value was reported. The samples were
160 dried overnight in a vacuum oven at 80°C prior to the test.

161 **Particle size analysis.** The droplet and particle size distribution were measured using a laser
162 diffraction particle size analyser (Malvern Mastersizer S, Malvern Instruments, UK). The
163 average size was expressed as the volume median diameter, $D_{v,0.5}$, corresponding to 50 vol%
164 on the cumulative particle size distribution curve. The particle (droplet) size uniformity was
165 estimated using a relative span factor given by

$$\text{span} = \frac{D_{v,0.9} - D_{v,0.1}}{D_{v,0.5}} \quad (1)$$

166 where $D_{v,0.9}$ and $D_{v,0.1}$ are the diameters corresponding to 90 and 10 vol% of the particles,
167 respectively.

168 **CO₂ capture.** The CO₂ capture capacity of the particles was measured using a fixed-bed
 169 adsorption column (15.8 mm O.D. and 9.25 mm I.D., stainless steel). A schematic of the rig
 170 is shown in Fig. 3. In each test the column was packed with 3 g of the particles. A stainless
 171 steel mesh with an average pore size of 0.075 mm and quartz wool plugs were placed at both
 172 sides of the column to prevent particles from escaping from the bed and contaminating the
 173 system. A uniform temperature distribution over the bed was achieved using an in-house
 174 temperature controller system. The gas flow rate was controlled by a mass flow controller
 175 (Alicat Scientific Inc, UK). Prior to each test, the column was purged with 130 mL/min of N₂
 176 for 2 h at 120°C and then allowed to cool down to the test temperature. The CO₂ capture was
 177 done by blowing through the bed a simulated gas mixture of CO₂ and N₂ containing 15 %
 178 (v/v) N₂ at 130 mL/min. The CO₂ concentration in the effluent stream was continuously
 179 monitored using a CO₂ infrared analyser (Quantek Instruments, USA).

180 **3. RESULTS AND DISCUSSION**

181 **3.1. Membrane emulsification**

182 Fig. 4 shows the effect of stirring rate in the MDC on the volume median droplet diameter,
 183 $D_{v,0.5}^d$, and span at the transmembrane flux of 20 and 200 Lm⁻²h⁻¹. The dispersed phase
 184 content at the end of the process was 9 vol%. An increase in stirring speed from 100 to 1200
 185 rpm caused a reduction in $D_{v,0.5}^d$ from 157 to 34 μm at 20 Lm⁻²h⁻¹, and from 137 to 47 μm at
 186 200 Lm⁻²h⁻¹. For membrane with a hexagonal pore array, $D_{v,0.5}^d$ can be correlated with the
 187 droplet formation time, t_d and the transmembrane flux, J_d , using the following equation [36]:
 188
 189

$$D_{v,0.5}^d = \left(\frac{3\sqrt{3}}{\pi k} J_d L^2 t_d \right)^{1/3} \quad (2)$$

190 where L is the membrane pore spacing and k is the fraction of active pores. At constant J_d
 191 and k values, an increase in stirring rate leads to an increase in the drag force acting on the

192 droplets during formation on the membrane surface and the droplet formation time is shorter,
193 causing a reduction in $D_{v,0.5}^d$ based on Eq. (2). The effect of flux on $D_{v,0.5}^d$ differs depending
194 on the rotational speed of the stirrer. At higher stirring rates (800-1200 rpm), the droplets are
195 smaller and do not touch each other while they are growing on the membrane surface. In this
196 case, at higher dispersed phase flux, a higher amount of the dispersed phase will flow into the
197 growing droplet during pinch off and larger droplets will be formed (Fig. 4). At lower stirring
198 rates (100-600 rpm), the size of adjacent growing droplets may become comparable with the
199 membrane pore spacing ($D_{v,0.5}^d \approx L$), causing the forming drops to touch each other on the
200 membrane surface and exert an additional force, the push-off force [37]. The push-off force
201 arises from droplet deformation and acts normal to and away from the membrane surface, i.e.,
202 in the opposite direction to the interfacial tension force, leading to shorter droplet formation
203 time and lower $D_{v,0.5}^d$ values at higher fluxes. As shown in Fig. 4, the most uniform droplets
204 with a span below 0.8 were obtained at the intermediate rotation speeds (600 and 800 rpm).

205 The effect of dispersed phase content ϕ_o in the prepared emulsion on $D_{v,0.5}^d$ and span at a
206 stirring rate of 800 rpm and flux of $200 \text{ Lm}^{-2}\text{h}^{-1}$ is shown in Fig. 5a. No significant variations
207 in $D_{v,0.5}^d$ were observed when ϕ_o increased from 9% to 29%. The same trend was observed in
208 preparation of sunflower oil-in-water emulsions stabilised by 2 wt% Tween 20 in the MDC
209 over a ϕ_o range from 5 to 60% [36]. This behaviour can be attributed to the short injection
210 time that ranged from less than 2 min at $\phi_o = 9 \text{ vol}\%$ to 7 min at $\phi_o = 29 \text{ vol}\%$ during which
211 no significant changes in membrane fouling occurred. The most uniform emulsion droplets
212 with a span of 0.76 were obtained at ϕ_o of 9%. The stability of emulsion droplets against
213 coalescence over 4 weeks of storage at ambient temperature is shown in Fig. 5b. Since AIBN
214 slowly decomposes at room temperature, in this stability study no AIBN was added in the
215 organic phase to prevent polymerisation during storage. No noticeable change in $D_{v,0.5}^d$ was

216 observed over the entire 4-week period implying stable emulsion formulation, although the
217 droplet size uniformity deteriorated slightly over time.

218 219 220 **3.2. Characterisation of synthesised MIP particles** 221

222 Fig. 6 shows SEM images of produced particles after downstream processing, i.e. surfactant
223 removal and template extraction. All the particles are non-agglomerated, spherical and have a
224 smooth surface without cracks. Fig. 7a gives a comparison between the size of the droplets
225 before polymerisation and the corresponding size of the particles after polymerisation and
226 washing. All emulsions were prepared with a dispersed phase content of 23 vol% at J_d of 200
227 $\text{Lm}^{-2}\text{h}^{-1}$ and 400-1000 rpm. The solid diagonal line indicates the data points for which $D_{v,0.5}^d$
228 is equal to $D_{v,0.5}^p$, and the two dashed lines represent $\pm 10\%$ deviation of $D_{v,0.5}^p$ from $D_{v,0.5}^d$. It
229 can be seen that $D_{v,0.5}^p$ deviates from $D_{v,0.5}^d$ only by 4% for the largest droplets produced at
230 400 rpm. The deviation of $D_{v,0.5}^p$ from $D_{v,0.5}^d$ increases with decreasing droplet size and
231 reaches 10% for the smallest droplets produced at 1000 rpm. This observation was not
232 surprising, since very fine particles can be removed during washing, shifting the particle size
233 distribution curve towards the larger particle sizes. The $D_{v,0.1}^d$ value for the droplets produced
234 at 1000 and 400 rpm was 49.9 and 71.6 μm , respectively. After polymerisation and washing,
235 the corresponding $D_{v,0.1}^p$ values were 66.5 and 83.1 μm (33% and 16% higher than the $D_{v,0.1}^d$
236 values), meaning that in the former case the fines were more substantially washed away. Fig.
237 7b shows that the size distribution for the particles is similar to that for the droplets and span
238 values for the particles are around 0.9 in all cases.

Fig. 8 shows a nitrogen adsorption-desorption isotherm and pore size distribution of the particles.
The isotherm is characterised by a hysteresis loop between relative pressure, P/P_0 , of 0.4 and 0.95

associated with capillary condensation occurring in mesopores, implying a type IV isotherm, according to IUPAC classification [38]. The adsorption volume rapidly increases at low P/P_0 values of less than 0.05 due to strong interaction of the N_2 molecules with a bare surface. When the monolayer formation is completed, multilayer formation starts to take place corresponding to the sharp knee of the isotherm. The total surface area, S_{BET} , of the sample was $239 \text{ m}^2/\text{g}$, whereas the external surface area, S_p , was $11 \text{ m}^2/\text{g}$. Since $S_p \ll S_{BET}$, the surface area of the particles was mainly due to internal surface and the particle size had a negligible effect on the total surface area. The pore size distribution was bimodal, with two maxima occurring at diameters of 4 and 9 nm. Compared to MIP particles synthesised by bulk polymerisation that follow a type II adsorption isotherm with an average pore size of 10-24 nm [23], the pore size distribution of the sample in Fig. 8 was considerably narrower with a smaller average pore size.

Fig. 9 shows the TGA curve of the sample heated from 100 to 600°C at a ramp rate of $10^\circ\text{C}/\text{min}$ in an inert atmosphere of N_2 . There was no obvious weight loss up to 210°C and 5% mass loss occurred at 245°C , which is well above anticipated desorption temperature. The average density of the particles measured with a helium pycnometer was $1.3 \text{ g}/\text{cm}^3$. Based on their size and density, the particles belong to Geldart Group A [39], referred to as aeratable particles. These particles can easily be fluidised, with homogeneous fluidisation at low superficial gas velocities and relatively small bubbles at higher velocities [40].

3.3. CO_2 capture capacity of MIP particles

The CO_2 capture capacity of the MIP particles was determined in a fixed-bed reactor at constant adsorption temperatures of 273, 298, and 353 K. The equilibrium CO_2 capture capacity, q_{eq} (mmol/g) was calculated using Eq. (3):

$$q_{eq} = \frac{QC_i t_{ad}}{m_s} \quad (3)$$

244 where Q (mL/min) is the feed gas flow rate, m_s (g) is the mass of the particles in the column,
245 and C_i (mmol/mL) is the molar concentration of CO_2 in the feed stream:

$$C_i = \frac{y_i P}{RT} \quad (4)$$

246 where y_i is the CO_2 molar fraction in the feed stream which was 0.15 in this work, P (kPa) is
247 the total gas pressure in the reactor (~ 102 kPa), T (K) is the operating temperature, R is the
248 universal gas constant, 8.314 J/(K·mol), and t_{ad} is the stoichiometric time, which is the
249 breakthrough time in an ideal adsorption column with a vertical breakthrough curve. The
250 ideal adsorption column operates at 100% CO_2 removal efficiency until the adsorbent
251 becomes fully saturated with CO_2 , after which the removal efficiency instantaneously drops
252 to zero. The breakthrough time is equivalent to the area between the breakthrough curve and
253 the C_o/C_i axis, as shown by the hatched area in Fig. 10, and can be calculated from [41]

$$t_{\text{ad}} = \int_0^{\infty} \left(1 - \frac{C_o}{C_i}\right) dt \quad (5)$$

254 where C_o (mmol/mL) is the molar concentration of CO_2 in the effluent stream.

255 The q_{eq} values determined from the breakthrough curves in Fig. 10 were 0.59, 0.30, and 0.16
256 mmol/g at 273, 298, and 353 K, respectively. These CO_2 capture capacities are comparable to
257 those of N_2 -phobic nanoporous azo-covalent organic polymers synthesised by Patel et al.
258 [42]. The lower capture capacities at higher temperatures can be attributed to the exothermic
259 nature of the adsorption process and weaker hydrogen bonding interactions between CO_2
260 molecules and amide groups within the cavities. The mass transfer resistance in the column
261 can be estimated from the slope of the breakthrough curves in the mass transfer zone [43]. In
262 Fig. 10, the steeper slope of the breakthrough curve at higher temperatures can be attributed
263 to the lower mass transfer resistance, due to faster diffusion of CO_2 .

4. CONCLUSIONS

Spherical molecularly imprinted polymeric particles containing CONH₂ decorated cavities with CO₂ recognition properties were fabricated using membrane emulsification and subsequent suspension polymerisation. The amide groups of the functional monomer (AAM) stayed intact during polymerisation, meaning that the particles were inherently functionalised with polar amide groups and an additional surface functionalisation step was not necessary. The droplet size was controlled by shear stress at the membrane surface during the membrane emulsification process, associated with varying stirring rate. The size of the particles after polymerisation was consistent with the initial droplet size, but the median particle size increased by 4-10% after washing due to removal of fine particles. The droplets maintained their physical stability during storage for 4 weeks and their size was independent of the dispersed phase content reached at the end of the emulsification process. The nitrogen adsorption-desorption isotherm of the material was of type IV, with pore diameters smaller than 20 nm and a specific surface area of 239 m²/g. Based on their density of 1.3 g/cm³ and mean diameters of 80-140 μm, the particles were classified into Geldart group A ('aeratable' and easily fluidisable). The particles were thermally stable up to 210°C and exhibited a CO₂ capture capacity of 0.59 mmol/g at 273 K and 0.15 bar CO₂ partial pressure. Future work will be focused on scale-up of the production process using continuous membrane emulsification systems.

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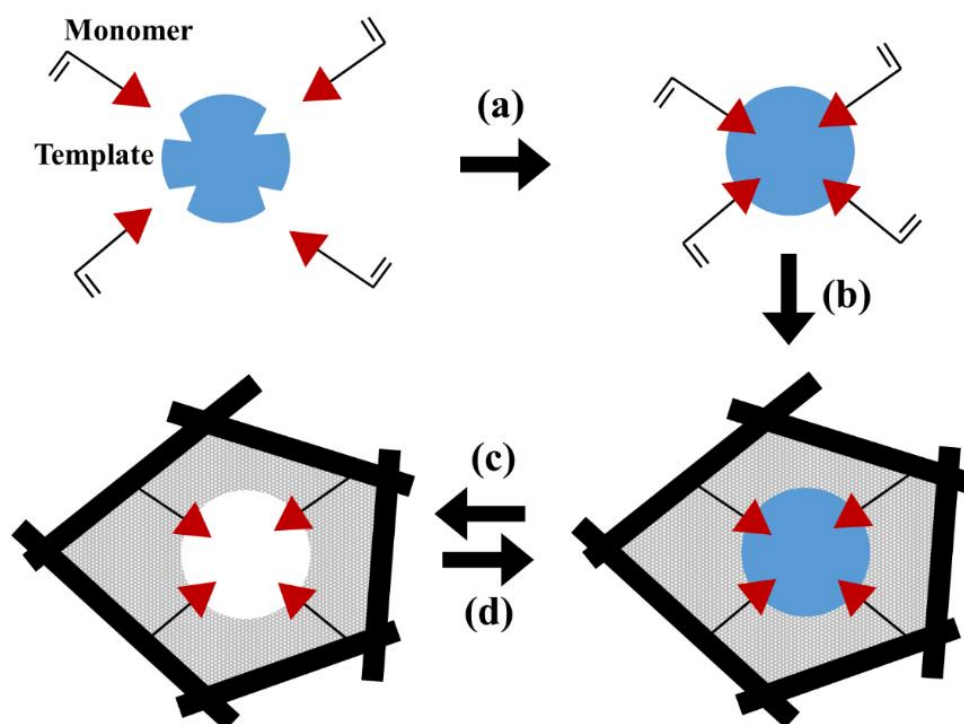


Fig. 1. Formation of molecularly imprinted polymer matrix in this study: (a) Self-assembly of oxalic acid (dummy template) and AAM (monomer); (b) Cross-linking of monomer-template complex using EGDMA; (c) Template removal by methanolic HCl and formation of cavities decorated with amide groups; (d) Capture of CO₂ molecules within the cavity. Template and CO₂ molecules are shown as blue spheres, while amide (CONH₂) groups are shown as red triangles.

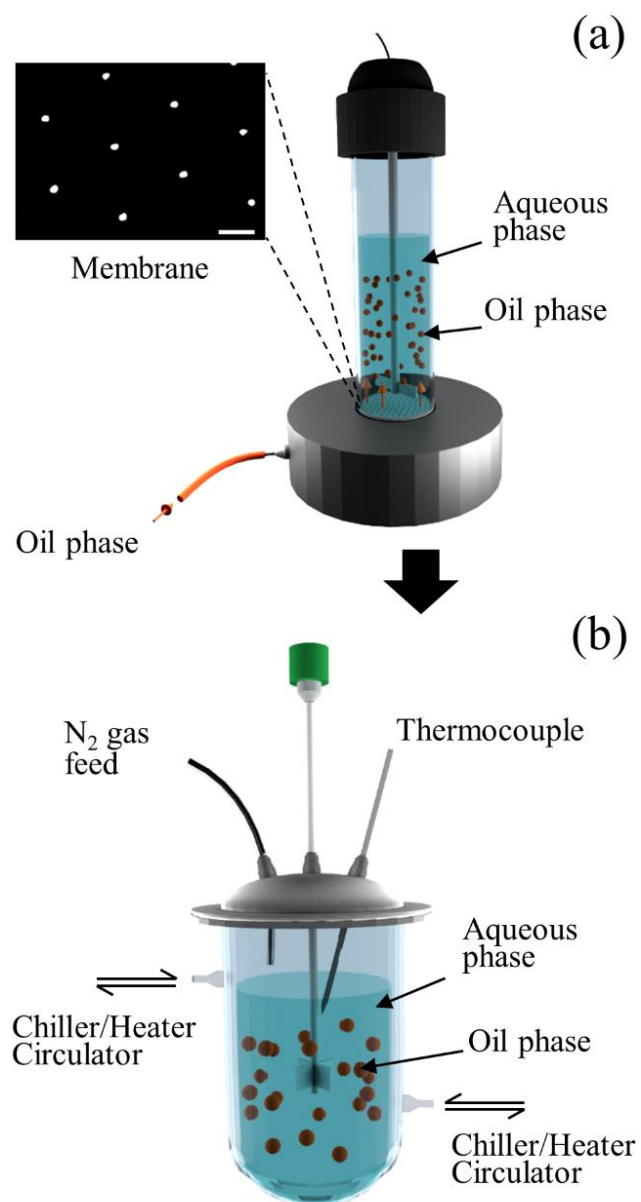


Fig. 2. A schematic of the experimental set-up for the production of MIP particles: (a) Micropore Dispersion Cell for membrane emulsification of the organic phase (AAM, OA, EGDMA, and AIBN dissolved in a mixture of AN and TL) into a 0.5 wt% PVA solution. The inset image is a micrograph of the membrane surface, and the scale bar is 100 μm ; (b) Jacketed batch reactor for suspension polymerisation at controlled temperature.

Figure 3

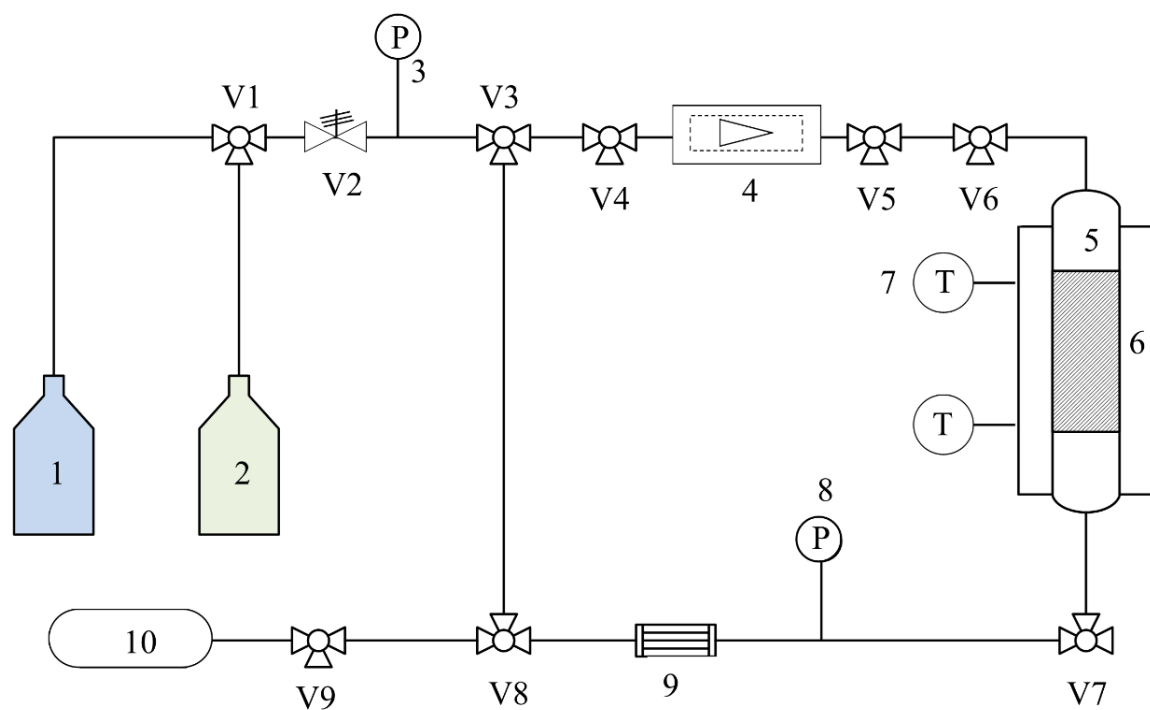


Fig. 3. A schematic of the rig used to test CO₂ capture capacity of the synthesised MIP particles: (1) CO₂ cylinder; (2) N₂ cylinder; (3 & 8) pressure gauge; (4) mass flow controller; (5) fixed bed column; (6) heating system; (7) thermocouples; (9) Peltier cooling module; (10) CO₂ gas analyser; (V1 & V3-V9) 3-way valves; (V2) relief valve.

Figure 4

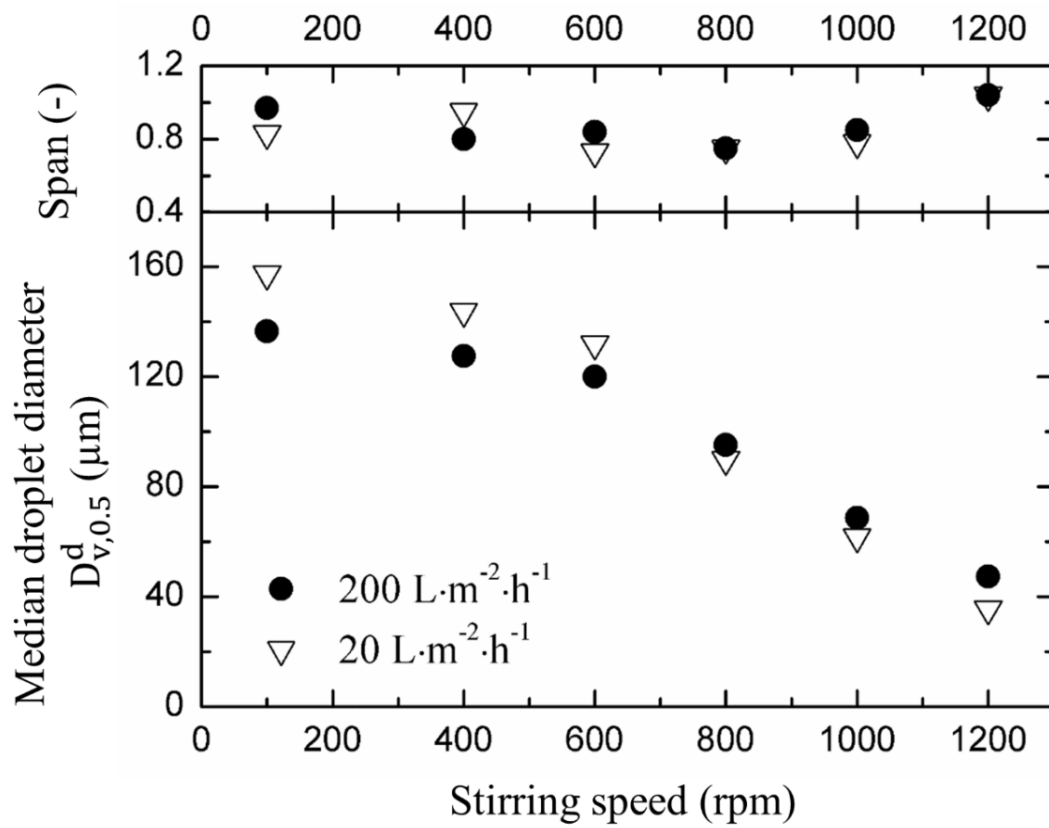


Fig. 4. The effect of stirring rate during membrane emulsification on the volume median diameter, $D_{v,0.5}^d$, and the span at $J_d = 20$ and $200 \text{ L}\cdot\text{m}^{-2}\cdot\text{h}^{-1}$. The dispersed (oil) phase content ϕ_o at the end of the emulsification process was 9 vol%.

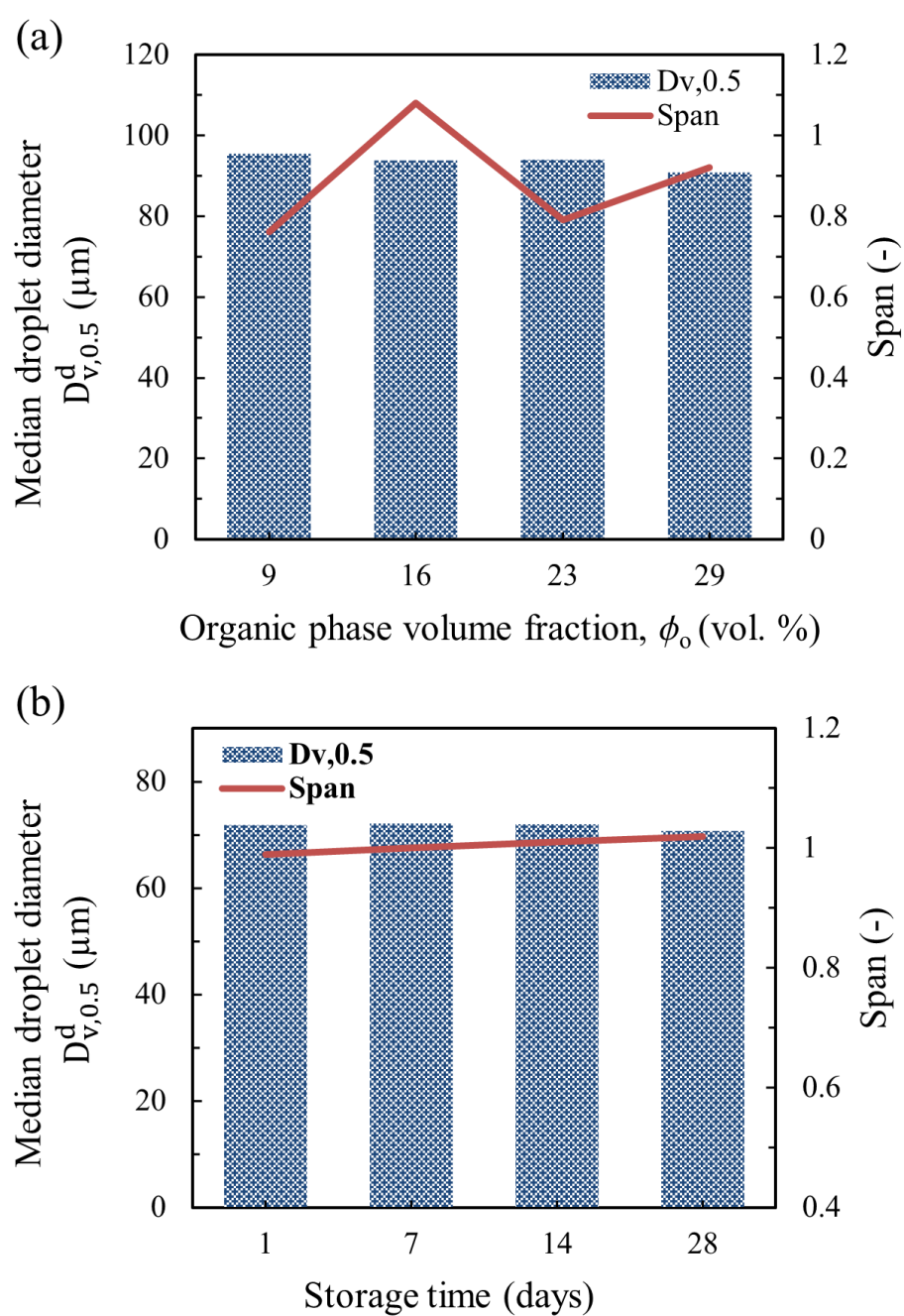


Fig. 5. (a) The effect of dispersed phase content on the volume median diameter, $D_{v,0.5}^d$, and the span at a stirring rate of 800 rpm and $J_d = 200 \text{ Lm}^{-2}\text{h}^{-1}$. (b) Long-term emulsion stability prior to suspension polymerisation.

Figure 6

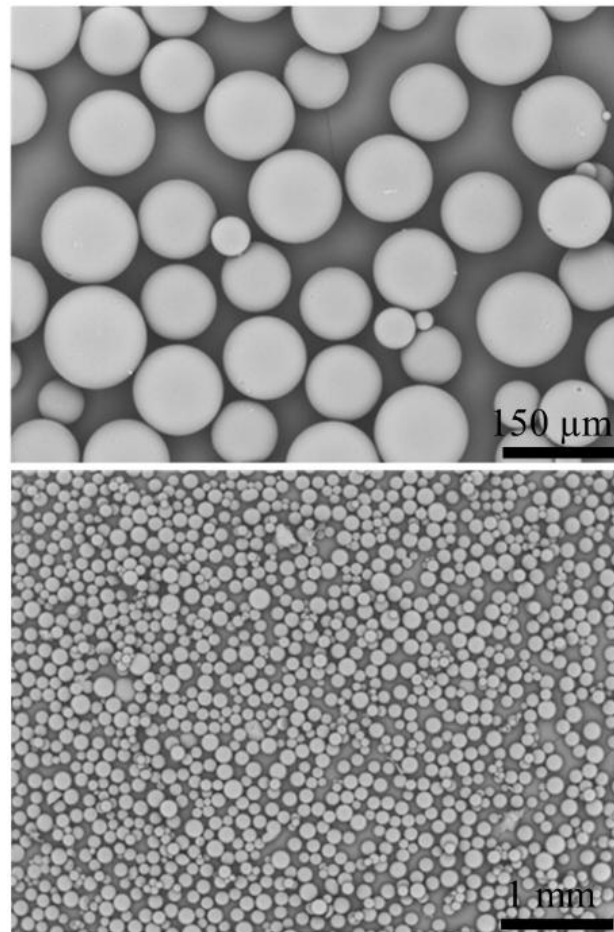


Fig. 6. SEM images of the MIP particles ($D_{v,0.5}^p = 141 \mu\text{m}$, span = 0.96) taken at different magnifications. The emulsion was prepared at 400 rpm, $J_d = 200 \text{ Lm}^{-2}\text{h}^{-1}$, and $\phi_o = 23 \text{ vol}\%$.

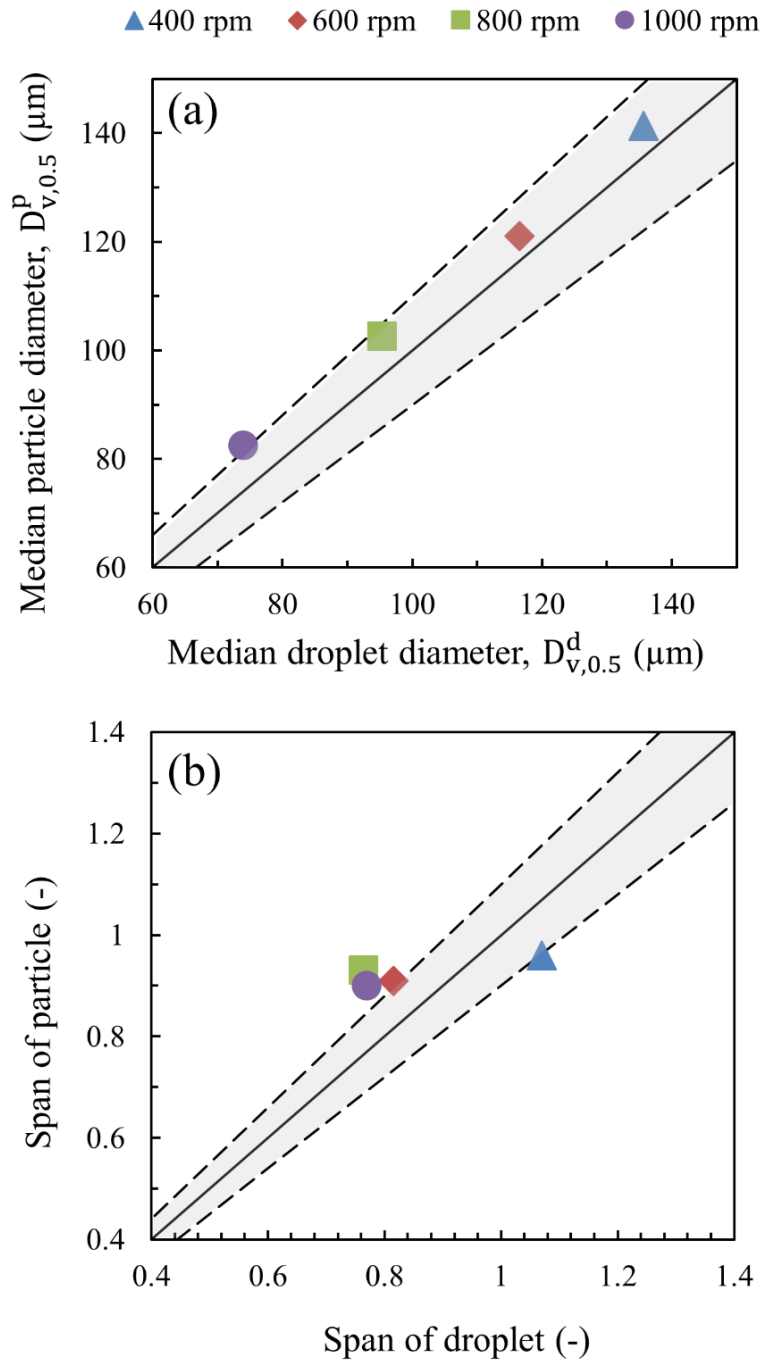


Fig. 7. A comparison of the median particle diameter (a) and the relative span factor (b) before polymerisation and after polymerisation and downstream processing. The solid lines follow the equations (a): $D_{v,0.5}^p = D_{v,0.5}^d$ and (b): particle span = droplet span. The dashed lines represent $\pm 10\%$ deviation from the diagonal lines. The emulsions were prepared at $J_d = 200 \text{ Lm}^{-2}\text{h}^{-1}$ and $\phi_o = 23 \text{ vol}\%$.

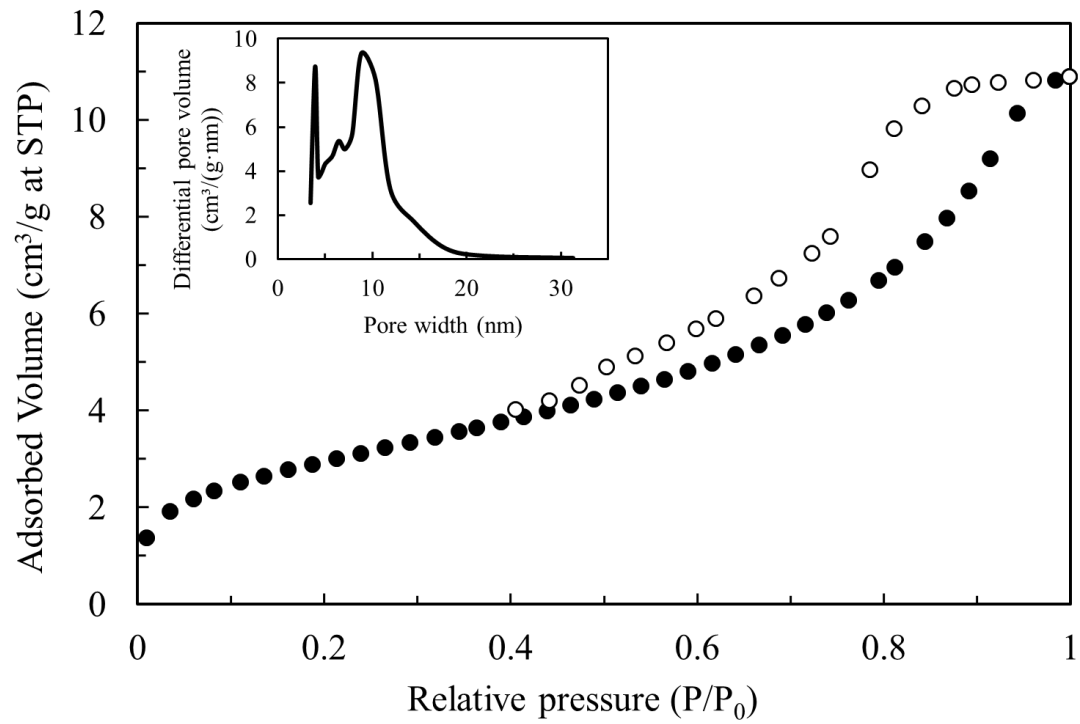


Fig. 8. Nitrogen adsorption-desorption isotherm of the particles produced from droplets at 800 rpm, 200 Lm⁻²h⁻¹, and $\phi_0 = 23$ vol%. The solid and open symbols denote the adsorption and desorption curve, respectively. The inset graph shows the pore size distribution curve.

Figure 9

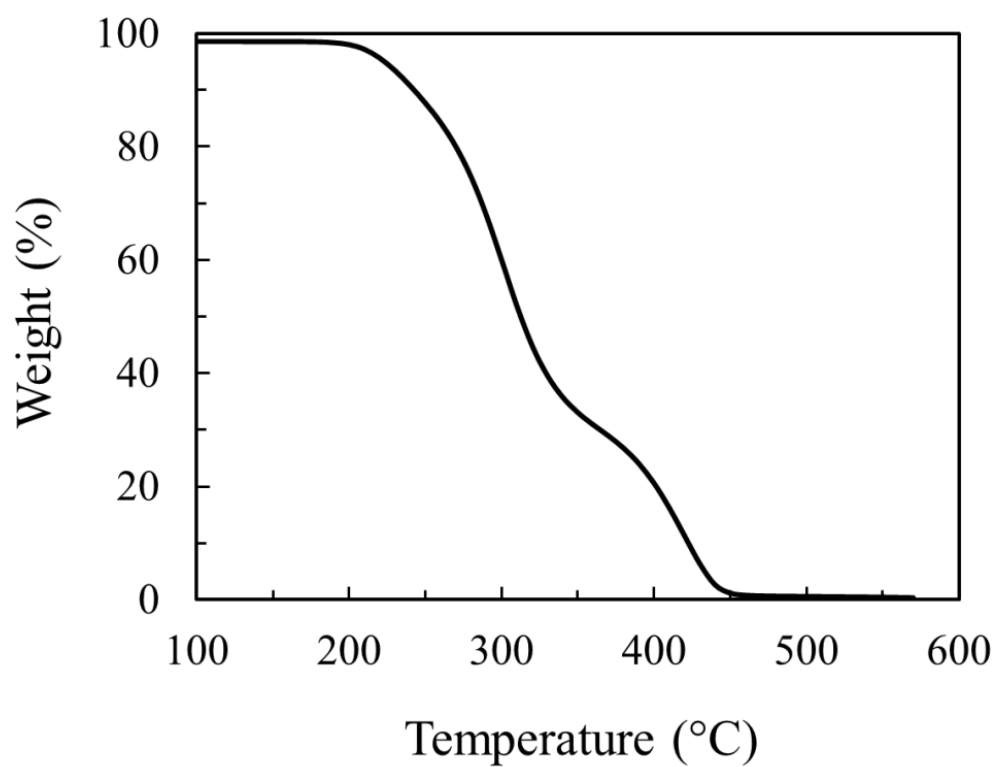


Fig. 9. Thermal degradation of the particles over the temperature range of 100- 600°C at a ramp rate of 10 °C/min and under the nitrogen flow. The monomer droplets were produced at 800 rpm, $J_d = 200 \text{ Lm}^{-2}\text{h}^{-1}$, and $\phi_o = 23 \text{ vol\%}$.

Figure 10

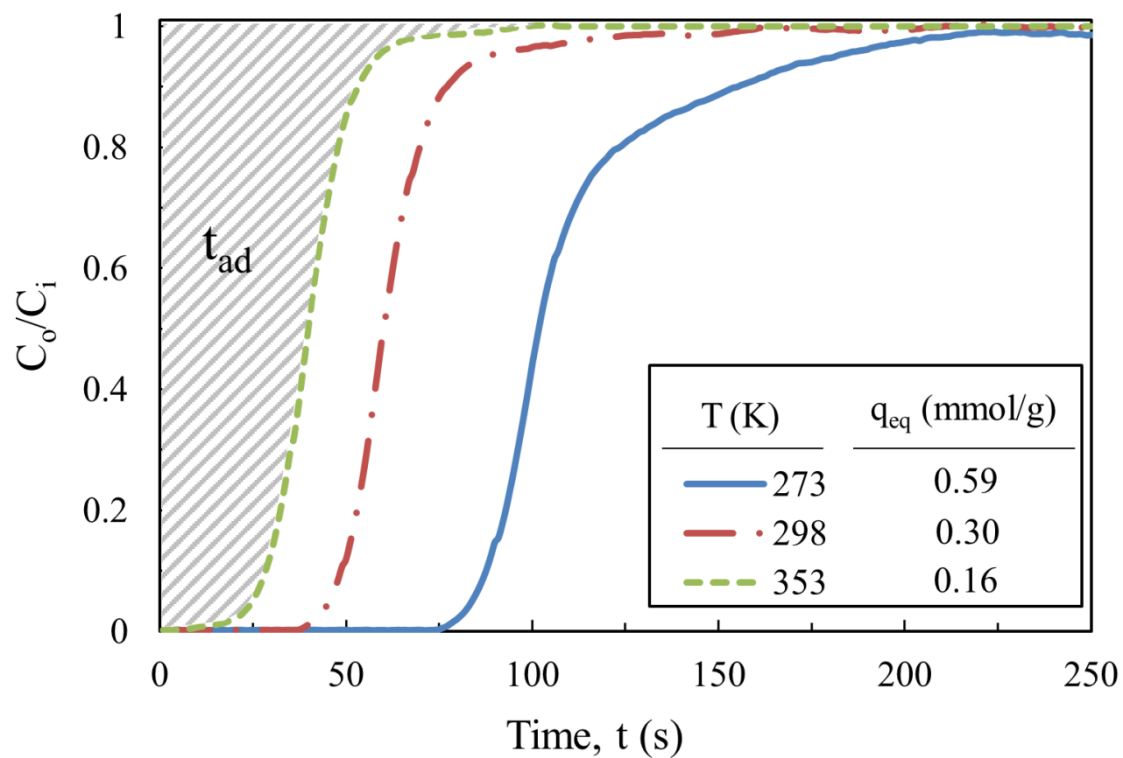


Fig. 10. The breakthrough curves of the MIP particles at different temperatures. The CO_2 partial pressure in the reactor was 0.15 bar. The inset table shows the equilibrium CO_2 capture capacity of the particles at different temperatures calculated using Eq. (3). The hatched area corresponds to the stoichiometric time at 353 K. The emulsion was produced at 800 rpm, $J_d = 200 \text{ Lm}^{-2}\text{h}^{-1}$, and $\phi_o = 23 \text{ vol}\%$.

ABSTRACT

Highly uniform amide-based molecularly imprinted polymer (MIP) particles containing CO₂-philic cavities decorated with amide groups were produced using membrane emulsification and subsequent suspension polymerisation. The organic phase containing acrylamide (functional monomer), oxalic acid (dummy template), ethylene glycol dimethacrylate (crosslinker) and azobisisobutyronitrile (initiator) dissolved in a 50/50 mixture (by volume) of acetonitrile and toluene (porogenic solvents) was injected through a microengineered nickel membrane with a pore diameter of 20 µm and a pore spacing of 200 µm into agitated 0.5 wt% aqueous solution of poly(vinyl alcohol) to form droplets that have been polymerised at 60°C for 3 h. The volume median diameter of the droplets was controlled between 35 and 158 µm by shear stress at the membrane surface. The droplets maintained their physical stability during storage for 4 weeks and their size was independent of the dispersed phase content. The particle size after polymerisation was consistent with the initial droplet size. The particles were stable up to 210°C and had a specific surface area of 239 m²/g and a CO₂ capture capacity of 0.59 mmol/g at 273 K and 0.15 bar CO₂ partial pressure.