



# Genome-Wide Association Study of Cryptosporidiosis in Infants Implicates *PRKCA*

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**ABSTRACT** Diarrhea is a major cause of both morbidity and mortality worldwide, especially among young children. Cryptosporidiosis is a leading cause of diarrhea in children, particularly in South Asia and sub-Saharan Africa, where it is responsible for over 200,000 deaths per year. Beyond the initial clinical presentation of diarrhea, it is associated with long-term sequelae such as malnutrition and neurocognitive developmental deficits. Risk factors include poverty and overcrowding, and yet not all children with these risk factors and exposure are infected, nor do all infected children develop symptomatic disease. One potential risk factor to explain these differences is their human genome. To identify genetic variants associated with symptomatic cryptosporidiosis, we conducted a genome-wide association study (GWAS) examining 6.5 million single nucleotide polymorphisms (SNPs) in 873 children from three independent cohorts in Dhaka, Bangladesh, namely, the Dhaka Birth Cohort (DBC), the Performance of Rotavirus and Oral Polio Vaccines in Developing Countries (PROVIDE) study, and the Cryptosporidiosis Birth Cohort (CBC). Associations were estimated separately for each cohort under an additive model, adjusting for length-for-age Z-score at 12 months of age, the first two principal components to account for population substructure, and genotyping batch. The strongest meta-analytic association was with rs58296998 ( $P = 3.73 \times 10^{-8}$ ), an intronic SNP and expression quantitative trait locus (eQTL) of protein kinase C alpha (*PRKCA*). Each additional risk allele conferred 2.4 times the odds of *Cryptosporidium*-associated diarrhea in the first year of life. This genetic association suggests a role for protein kinase C alpha in pediatric cryptosporidiosis and warrants further investigation.

**IMPORTANCE** Globally, diarrhea remains one of the major causes of pediatric morbidity and mortality. The initial symptoms of diarrhea can often lead to long-term consequences for the health of young children, such as malnutrition and neurocognitive developmental deficits. Despite many children having similar exposures to infectious causes of diarrhea, not all develop symptomatic disease, indicating a possible role for human genetic variation. Here, we conducted a genetic study of susceptibility to symptomatic disease associated with *Cryptosporidium* infection (a leading cause of diarrhea) in three independent cohorts of infants from Dhaka, Bangladesh. We identified a genetic variant within protein kinase C alpha (*PRKCA*) asso-

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ciated with higher risk of cryptosporidiosis in the first year of life. These results indicate a role for human genetics in susceptibility to cryptosporidiosis and warrant further research to elucidate the mechanism.

**KEYWORDS** *Cryptosporidium*, genetics, genome analysis

**C**ryptosporidiosis is a leading cause of diarrhea and is estimated to be responsible for greater than 200,000 deaths in young children in South Asia and sub-Saharan Africa each year (1). Beyond the immediate infection, cryptosporidiosis is also associated with long-term sequelae, including malnutrition and neurocognitive developmental deficits (2–5). The majority of human infections are caused by the *Cryptosporidium hominis*, *C. meleagridis*, and *C. parvum* species (4, 6, 7). As cryptosporidiosis is transmitted fecal-orally, contact with any reservoir with possible fecal contamination could serve as a point of transmission. In the developed world, cryptosporidia represent an important cause of diarrhea in individuals living with HIV and are the most common pathogens causing waterborne outbreaks (7).

In regions of endemicity, cryptosporidiosis mostly impacts young children, and risk factors for infection include poverty and overcrowding (4, 8–10). Livestock serve as an environmental reservoir for *C. parvum*, and transmission after contact with infected animals or with drinking water contaminated by human or animal waste has been reported previously (11). In regions where *Cryptosporidium* infection is endemic, there is heterogeneity in clinical courses and outcomes. In an eight-site multicenter international study of enteric infection and malnutrition (MAL-ED), the rate of *Cryptosporidium* infection, age of onset, number of repeat infections, and clinical manifestation differed significantly by site (9). In a recent study in Dhaka, Bangladesh, we found that two-thirds of children living in an urban slum were infected with *Cryptosporidium* by 2 years of age and that one-fourth had had more than one episode of cryptosporidiosis. Fully three-fourths of the infections were subclinical, but, regardless of the symptoms, children with cryptosporidiosis were more likely to become malnourished by 2 years of age (4). Potential explanations for the *Cryptosporidium* infection heterogeneity include differences in the pathogenicity of various *Cryptosporidium* species or genotypes (12) and in host genetic susceptibility.

Candidate gene studies identified an increased risk of *Cryptosporidium* infection associated with specific alleles in HLA class I and II genes and with single nucleotide polymorphisms (SNPs) in the mannose binding lectin (*MBL*) gene (13–15). Bangladeshi preschool children with multiple *Cryptosporidium* infections ( $\geq 2$  infections) were more likely to carry the -221 *MBL2* promoter variant (rs7906206; odds ratio [OR] = 4.02,  $P = 0.025$ ) and to have the YO/XA haplotype (OR = 4.91), as well as to be deficient in their *MBL* serum levels (OR = 10.45) (14). Since the findings with respect to the *MBL* and HLA alleles explained *Cryptosporidium* susceptibility only partially, we conducted a genome-wide association study (GWAS) of cryptosporidiosis occurring in the first year of life using three existing birth cohorts of children in Dhaka, Bangladesh: the Performance of Rotavirus and Oral Polio Vaccines in Developing Countries (PROVIDE) study, the Dhaka Birth Cohort (DBC), and the Cryptosporidiosis Birth Cohort (CBC).

(This article was submitted to an online preprint archive [16].)

## RESULTS

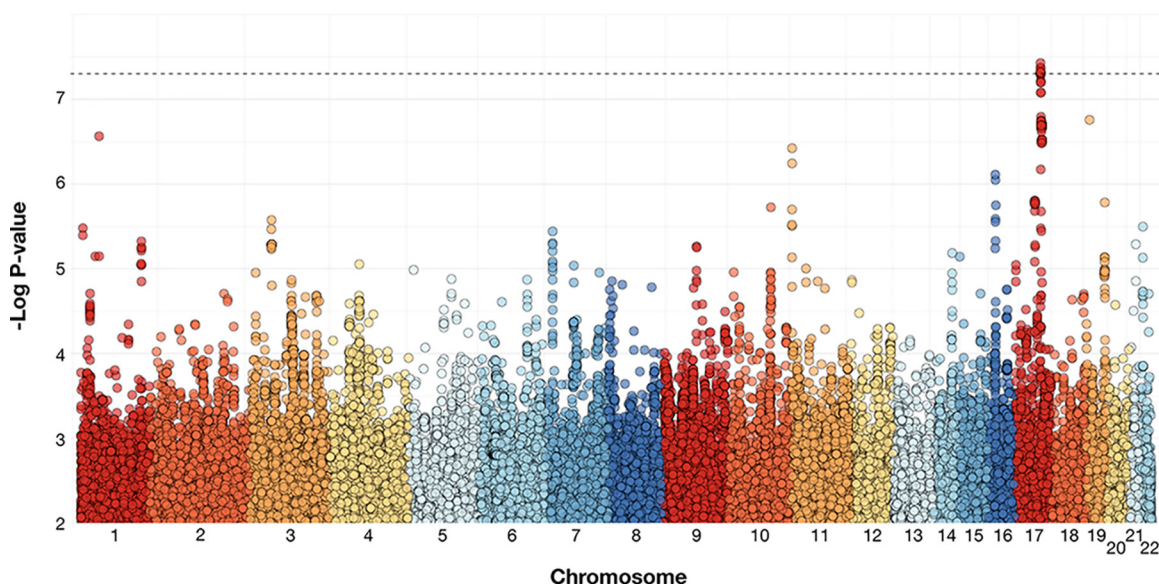
Across these three cohorts, there were a total of 183 children with at least one symptomatic (diarrheal) sample that tested positive for *Cryptosporidium* within the first year of life (“cases”) (Table 1). A total of 873 children did not test positive for *Cryptosporidium* in either symptomatic (diarrheal) or surveillance samples within the first year of life (“controls”). There were no significant differences in length-for-age Z-score (LAZ) at birth ( $LAZ_{birth}$ ), the number of days exclusively breastfed, or sex between cases and controls ( $P > 0.05$ ). To control for a possible role of malnutrition affecting susceptibility to infection, we compared the LAZ at 12 months of age ( $LAZ_{12}$ ) between cases and controls. We observed increased levels of stunting in cases (lower

**TABLE 1** Demographics of study populations

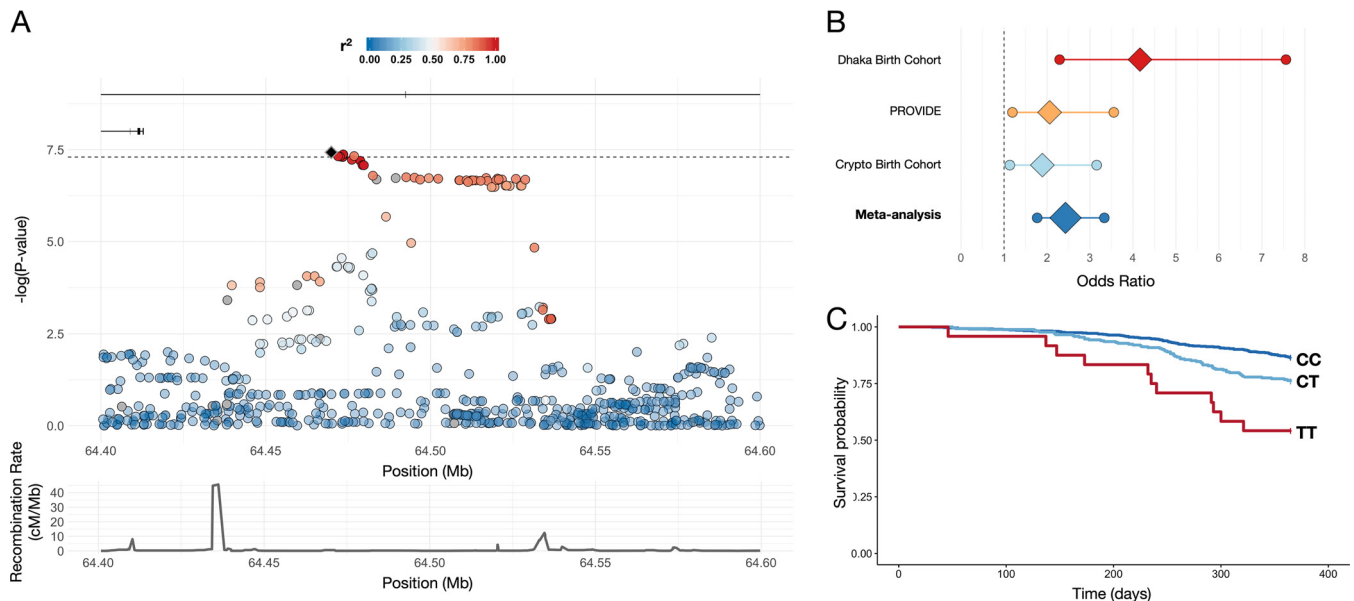
Parameter	Value for:									
	Dhaka Birth Cohort (DBC)			PROVIDE			Cryptosporidiosis Birth Cohort (CBC)			
	Mean for controls (n = 267)	Mean for cases (n = 46)	P	Mean for controls (n = 354)	Mean for cases (n = 60)	P	Mean for controls (n = 252)	Mean for cases (n = 77)	P	$P_{\text{het}}$
LAZ at 12 mos	-1.75	-1.74	0.97	-1.40	-1.79	$7.28 \times 10^{-3}$	-1.34	-1.63	0.02	0.12
Exclusive breast feeding (no. of days)	130.2	114.6	0.16	127.2	112.1	0.06	110.9	103.7	0.42	0.74
Sex (% female subjects)	46.3	34.8	0.15	45.9	46.7	0.91	52.8	57.7	0.45	0.28

LAZ<sub>12</sub>) versus controls within PROVIDE ( $P = 0.007$ ) and CBC ( $P = 0.02$ ), while no differences were observed in stunting between cases and controls in DBC ( $P = 0.97$ ). Additionally, there was no statistically significant evidence of heterogeneity in LAZ<sub>12</sub>, number of days exclusively breastfed, or sex between the three studies (heterogeneity  $P [P_{\text{het}}], >0.05$ ).

**GWAS of cryptosporidiosis within the first year of life.** We tested the association between 6.5 million SNPs across the human genome and symptomatic *Cryptosporidium* infection in the first year of life. Effects were estimated separately for the three birth cohorts and subsequently combined using a fixed-effects meta-analysis, filtered for heterogeneity ( $P_{\text{het}}$ ), minor allele frequency (MAF) ( $>5\%$ ), and imputation quality (INFO; score,  $>0.6$ ) (Fig. 1; see also Fig. S1 in the supplemental material). A total of 6 SNPs in an intron of *PRKCA* (protein kinase c, alpha) were significantly associated with *Cryptosporidium* infection ( $P < 5 \times 10^{-8}$ ) (Fig. 2A). For the SNP most highly associated with *Cryptosporidium* infection (rs58296998), each copy of the risk allele (T) conferred 2.4 times the odds of cryptosporidiosis within the first year of life ( $P = 3.73 \times 10^{-8}$ ). The effect size and risk allele were consistent across all three studies ( $P_{\text{het}}$  value of 0.11) (Fig. 2B). After conditioning performed on the basis of rs58296998 (by including this SNP in the logistic regression model as a covariate), the evidence for association with the remaining SNPs in the region was no longer statistically significant, suggesting that the observed association in *PRKCA* is explained by a single SNP (rs58296998) or by one



**FIG 1** Manhattan plot for cryptosporidiosis within the first year of life. Each dot indicates the association of a single SNP with cryptosporidiosis in the first year of life. SNPs are sorted by chromosome (each color) and position along the x axis. The y axis is the  $-\log_{10}$   $P$  value for the SNP association in the meta-analysis of study-specific logistic regressions adjusting for length-for-age Z-score at 12 months, the first two study-specific principal components, and the genotyping batch for the Dhaka Birth Cohort (DBC). Genome-wide significance ( $5 \times 10^{-8}$ ) is denoted by the dashed line. This plot is limited to associations with a  $P$  value below 0.01.



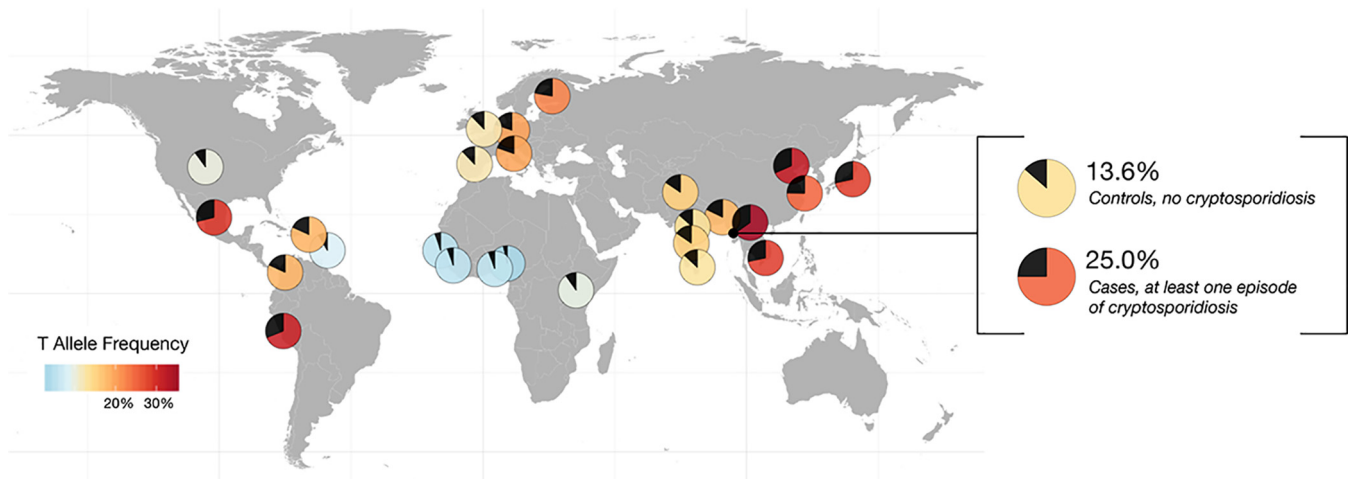
**FIG 2** Association between variants in *PRKCA* and cryptosporidiosis. (A) Regional association on chromosome 17 between variants in *PRKCA* and cryptosporidiosis. Fill denotes linkage disequilibrium ( $r^2$ ) between the top SNP (rs58296998) and surrounding SNPs. cM/Mb, centimorgan/megabase. (B) Forest plot of odds ratios and 95% confidence intervals for top signal rs58296998 by individual cohort and meta-analysis. Crypto Birth Cohort, Cryptosporidiosis Birth Cohort. (C) Survival analysis of first episode of cryptosporidium-associated diarrhea among all participants by rs58296998 genotype within the first year of life.

highly correlated with this SNP (Fig. S2A). Among the 26 children homozygous for the risk allele (TT) at rs58296998, 46% developed symptomatic cryptosporidiosis during the first year of life. This proportion decreased to 24% for children heterozygous (CT) for this risk allele ( $n = 272$ ), compared to 13% of children homozygous (CC) for the risk allele ( $n = 745$ ).

The rs58296998 T allele frequencies (15.0% to 16.7%) for all three cohorts in this region are consistent with the Bangladeshi reference population (1000 Genomes phase 3) frequency of 18% and the overall South Asian frequency of 15% (17). Globally, the highest frequencies of rs58296998 T allele are found in East Asian populations, with the highest T allele frequency of 34% of the Chinese Dai in Xishuangbanna, China. The rs58296998 T allele is at lower frequencies within Africa, at 9% within the Luhya in Kenya, and is even less frequent in West Africa (3.5% to 5.5%) (Fig. 3).

Cases had their first diarrheal episode positive for *Cryptosporidia* at a mean of 242 days of age. We confirmed the GWAS results with respect to the dosage of rs58296998 risk alleles significantly associated with time to first diarrheal sample positive for *Cryptosporidia* among cases versus right-censored controls (up to the child's first birthday) ( $P = 6.37 \times 10^{-8}$ ). All children homozygous for the risk allele (TT) had their first episode in the first year of life (Fig. 2C). Among cases, however, there was no statistically significant association between rs58296998 genotype and time to infection ( $P = 0.095$ ) (Fig. S2B). In PROVIDE, the rs58296998 genotype was associated with severity of diarrhea as determined by the Ruuska score ( $P = 0.028$ ) (Fig. S2C).

Suggestive SNP associations with *Cryptosporidium* ( $P < 10^{-6}$ ) were also identified on chromosome 11 (chr11) and chr16. The strongest association on chromosome 11 (rs4758351) was found within an intergenic region of a cluster of olfactory receptor genes. Each copy of the rs4758351 A allele (MAF of 14%) conferred 2.39 times the odds of *Cryptosporidium* within the first year of life ( $P = 3.78 \times 10^{-7}$ ) (Fig. S3A). Multiple SNPs in this region of chr11 (position 6015194 to position 6024551) had similar magnitudes and strengths of association with *Cryptosporidium* (OR, 2.13 to 2.39). The strongest association on chromosome 16 was with the rs9937140 SNP, located upstream of apolipoprotein O pseudogene 5 (*APOOP5*). Each copy of the rs9937140 G allele (MAF, 23%) conferred 1.99 times the odds of cryptosporidiosis ( $P = 7.75 \times 10^{-7}$ ) (Fig. S3B).

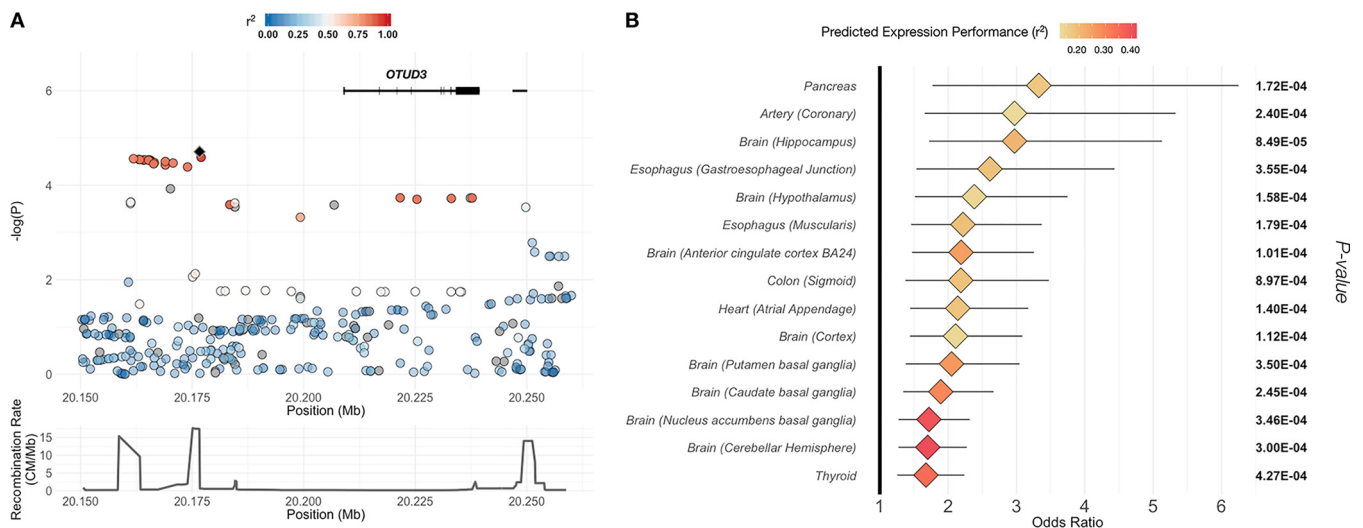


**FIG 3** Allele frequencies for allele T at top signal rs58296998 as determined by analysis of 1000 Genomes phase 3 data, as well as by analysis of case/control status in the three cohorts combined. Each pie chart on the map shows the frequency of the T allele with the black wedge. The remainder of each pie chart is colored in accordance with that T allele frequency. The inset provides the T allele frequency for children without any symptomatic cryptosporidiosis in the first year of life (controls; MAF = 13.6%) and for those with at least one diarrheal episode (cases; MAF = 25.0%).

**Expression and PrediXcan.** We used a publicly available resource, the Genotype-Tissue Expression (GTEx) Project, to estimate the influence of human genetic variation on human gene expression in multiple tissues (18, 19). The associated rs58296998 SNP, located in the *PRKCA* gene, is also associated with *PRKCA* expression. This expression quantitative trait locus (eQTL), or a genetic variant previously shown to influence the expression of a gene, showed decreasing expression of *PRKCA* with each T allele in the esophageal muscularis ( $P = 3.12 \times 10^{-5}$ ), the sigmoid colon ( $P = 4.61 \times 10^{-4}$ ), and the esophageal mucosa ( $P = 7.50 \times 10^{-4}$ ) (19). These expression data, coupled with the GWAS result, suggested that decreased expression of *PRKCA* is correlated with increased risk of symptomatic *Cryptosporidium* infection within the first year of life.

**Additional genome-wide expression and gene set analyses.** In the absence of direct gene expression measurement, we relied on previously estimated tissue-specific associations between genome-wide SNPs and gene expression, which quantify the genetic component of gene expression. We estimated predicted patterns of genome-wide differential gene expression between cases and controls by weighting the summary statistics from our GWAS of cryptosporidiosis in the first year of life by the use of tissue-specific PredictDB weights. These SNP-level estimates were then combined for each gene to infer association between imputed gene expression and cryptosporidiosis (20, 21). No association of predicted gene expression with cryptosporidiosis reached statistical significance. A total of 13 genes showed a nominally significant ( $P < 0.001$ ) association in more than one tissue-specific model (see Table S1 in the supplemental material; see also Fig. S4). Variants in the gene *OTUD3* (OTU deubiquitinase 3) (chr1; position 20208356 to position 20239438) were associated with cryptosporidiosis in 18 different tissue-specific models ( $P < 0.001$ ). In all tissue-specific models, individuals with predicted increased expression of *OTUD3* had an increased risk of cryptosporidiosis within the first year of life (OR, 1.68 to 6.63;  $P = 8.46 \times 10^{-5}$  to  $8.97 \times 10^{-4}$ ) (Fig. 4).

We also performed gene set enrichment analysis using MSigDB hallmark gene sets ( $n = 50$ ), KEGG ( $n = 186$ ) and BioCarta ( $n = 217$ ) by combining gene-level summary statistics to examine aggregate signals within biological pathways. No pathways reached statistical significance after adjusting for multiple comparisons; however, data from several gene sets were suggestive (Table S2). The two top-ranked gene sets are among the hedgehog signaling pathways, namely, the hallmark hedgehog signaling pathway (empirical  $P$  value [ $P_{\text{emp}}$ ] =  $5.04 \times 10^4$ ) (Bayes factor [BF] = 515.65) and KEGG hedgehog signaling pathway ( $P_{\text{emp}}$  =  $1.47 \times 10^{-3}$ ) (BF = 235.59).



**FIG 4** OTUD3 region showing association with cryptosporidiosis in the first year of life. (A) Association of SNPs on chromosome 1 region, colored by linkage disequilibrium ( $r^2$ ) with index SNP (black diamond). (B) Association of case status with imputed gene expression in all tissues with  $P$  value of  $<0.001$  and predicted expression performance of  $r^2 = >0.1$ .

## DISCUSSION

Here, we present the results of the first genome-wide association study of symptomatic *Cryptosporidium* infection. Specifically, we tested the role of host genetics in susceptibility to *Cryptosporidium* infection associated with diarrhea within the first year of life. A region on chromosome 17 was identified, with each additional T allele of rs58296998, an intronic SNP in *PRKCA*, conferring 2.4 times the odds of cryptosporidiosis within the first year of life. Additionally, this SNP was previously identified as an eQTL of *PRKCA*, with decreased expression of *PRKCA* associated with the T allele. This suggests that this SNP may influence *Cryptosporidium* infection through decreased expression of *PRKCA*.

The protein kinase C alpha gene (*PRKCA*) is an isotype of the protein kinase C (PKC) family, whose members are serine and threonine specific and are known to be involved in diverse cellular signaling pathways. Specifically, PKCs have numerous roles in the development and function of the gastrointestinal tract (22) and in the immune response (23). This relationship was confirmed with knockout experiments, where  $PKC\alpha$  was shown to be a positive regulator of Th17 cell effector functions.  $PKC\alpha$ -deficient [ $Prkca(-/-)$ ] cells failed to produce the appropriate levels of interleukin-17A (IL-17A) *in vitro* (23). An analysis of *Cryptosporidium parvum*-infected mice demonstrated the importance of the Th17 response to infection, showing increased levels of IL-17 mRNA and Th17 cell-related cytokines in gut tissue after infection (24). Additionally, both pharmacological inhibition and genetic  $PKC\alpha$  inhibition have been shown to prevent NHE3 internalization,  $Na^+$  malabsorption, and tumor necrosis factor (TNF)-mediated diarrhea, despite continued barrier dysfunction (25), supporting the idea of a role for *PRKCA* in symptomatic cryptosporidiosis. This link between *PRKCA* and Th17 may be critical to gut infections and, specifically, to infection of *Cryptosporidium* in the developing infant gut. We identified a SNP that was associated with decreased expression of *PRKCA* and thus was less able to mediate the IL-17 immune response during *Cryptosporidium* infection. *PRKCA* has also been shown to be associated with numerous other infections, including infections by *Staphylococcus aureus* (26); with progression of sepsis (27) and toxoplasmosis (28); with *Burkholderia cenocepacia* infections in cystic fibrosis patients (29); and with hepatitis E virus replication (30).

As an obligate intracellular parasite, *Cryptosporidium* relies on host cells to complete its life cycle in the human host; thus, it is also plausible that *PRKCA* directly mediates susceptibility via impacts on parasite invasion. Sporozoites invade brush border intestinal epithelial cells by inducing volume increases (31) and cytoskeletal remodeling at

the site of host cell attachment (32), leading to engulfment via host membrane protrusions. Studies have shown that inhibition of host factors, including actin remodeling proteins and PKC enzymes, is sufficient to inhibit sporozoite invasion *in vitro* (32). Interestingly, PKC $\alpha$  has been shown to play an important role in *Escherichia coli* pathogenesis (33). Like *Cryptosporidium*, *E. coli* induces host actin condensation at the site of host cell invasion, and immunocytochemical studies indicate that activated PKC $\alpha$  colocalized with actin condensation at the bacterial entry site (34).

While our top SNP within *PRKCA* has previously been shown to influence the expression of *PRKCA* in GTEx, our imputed gene expression analysis using PrediXcan did not reveal a significant difference in predicted levels of *PRKCA* expression between cases and controls. This was likely due to the difference between a single SNP being examined in GTEx and the combined effects of multiple eQTLs estimated from a European descent reference population in PrediXcan. A major limitation of predicted gene expression analyses is the lack of population specificity for non-European groups (35). The PrediXcan models were derived from individuals of European descent, as were the covariance structures used to infer correlations between eQTLs. We saw a direct relationship between population differences in allele frequencies for the weighted SNPs and impaired performance. Specifically, we observed the lowest predictive performance in tissues for which the informative SNPs had large differences in allele frequencies between European and South Asian populations in the 1000 Genomes Project phase 3 data (17) (see Fig. S5 in the supplemental material). These included two tissues, namely, esophageal mucosa and the colon sigmoid tissue, in which rs58296998 was identified as an eQTL for *PRKCA*. These trends highlight the importance of reference populations representative of global populations to ensure that tools are useful in non-European populations, such as ours. We also identified an association of increased expression of *OTUD3* with increased odds of cryptosporidiosis within the first year of life. This gene is associated with ulcerative colitis (36–42) and inflammatory bowel disease (43, 44). This finding is consistent with the hypothesis of a pathway shared between enteric infection and autoimmune intestinal disease, as indicated in a previous genetic analysis of *Entamoeba histolytica* infection in the same study population (45).

Collapsing the predicted patterns of differentially expressed genes into gene sets, we found enrichment in the hedgehog signaling pathway. A previous study examined the gene expression profiles of long noncoding RNA (lncRNA) and mRNA in HCT-8 cells infected with *C. parvum* subtype 11d (46). Of note, *PRKCA* was the most significantly differentially expressed gene in infected HCT8 cells 24 h postinfection (2.24-fold decreased expression in infected cells;  $P = 3.82 \times 10^{-5}$ ). Pathway analysis of the differentially expressed mRNAs found that genes in the hedgehog signaling pathway were significantly enriched during *Cryptosporidium* infection. This finding, in combination with our identification of hedgehog signaling in imputed gene expression profiles, is suggestive of a potential link between decreased *PRKCA* expression and hedgehog signaling; however, further research to confirm these findings and to elucidate the role of *PRKCA* genetic variation in gene expression and hedgehog pathway perturbation is needed.

A potential limitation of our study was that, due to the use of sensitive molecular diagnostics, multiple enteropathogens were frequently detected in each diarrheal sample. However, we did not detect the same genetic signatures as that seen in our previous study of *Entamoeba histolytica* in this same study population for *Cryptosporidium* (45). Further, coinfection with multiple pathogens would dilute the statistical signal for any one pathogen, and yet we found a statistically significant result for *Cryptosporidium*. Therefore, we are confident that our results are specific to cryptosporidiosis, despite cooccurrence with other enteric pathogens.

Through a GWAS meta-analysis of three separate birth cohorts, we identified a region in *PRKCA* on chromosome 17 as being associated with increased risk of symptomatic cryptosporidiosis in the first year of life among Bangladeshi infants. This gene has previously been implicated in other infectious outcomes, indicating pleiotropy with the immune system's reaction to numerous pathogens. Publicly available data support

a link between our top SNP and expression of *PRKCA*, suggesting a mechanism operating via Th17 inflammatory control. Clinical trials are currently proposed for PKC isotypes, including PKC- $\alpha$ , for treatment of autoimmune disease (47). These treatments may also be important for cryptosporidiosis, which lacks treatment for young children, due to an underlying shared pathway identified in this study. Identifying host genetic variations associated with cryptosporidiosis, such as those in *PRKCA*, can help us identify viable drug targets to improve treatment and prevention of this major cause of morbidity and mortality. Further research is needed to elucidate the mechanism underlying this relationship and to better understand the complex interplay of genetic susceptibility and environmental influences in the development of intestinal disease.

## MATERIALS AND METHODS

**Study protocol.** The study protocol was approved by the Research and Ethical Review Committee of the International Center for Diarrheal Disease Research, Bangladesh, and by the Institutional Review Board of the University of Virginia and the Institutional Review Board of the Johns Hopkins Bloomberg School of Public Health. The parents or guardians of all individuals provided informed consent.

**Dhaka Birth Cohort study design.** Designed to study the influence of malnutrition in child development, the Dhaka Birth Cohort (DBC) is a subset of a larger birth cohort recruited from the urban slum in the Mirpur Thana in Dhaka, Bangladesh. Children were enrolled within the first week after birth and followed up biweekly with household visits by trained field research assistants (FRAs) for the first year of life. Anthropometric measurements were collected at the time of enrollment and every 3 months thereafter. Length-for-age adjusted Z-scores (LAZ) were calculated by comparing the lengths and weights of study subjects with those of the World Health Organization (WHO) reference population, adjusting for age and sex, using WHO Anthro software, version 3.0.1. Field research assistants (FRAs) collected diarrheal stool samples from the home or study field clinic every time that the mother of the child reported diarrhea. To maintain a cold chain, the samples were transported to the Centre for Diarrheal Disease Research, Bangladesh (ICDDR,B) parasitology laboratory. The presence of *Cryptosporidium* was determined using enzyme-linked immunosorbent assay (ELISA). More details can be found in previously published reports by Steiner et al. (4) and Korpe et al. (9). We used a nested case-control design, where children with at least one diarrheal sample positive for *Cryptosporidium* within the first year were defined as “cases.” Children with diarrheal samples that were not positive for *Cryptosporidium* were defined as “controls.”

**PROVIDE study design.** The “Performance of Rotavirus and Oral Polio Vaccines in Developing Countries” (PROVIDE) Study consists of a randomized controlled clinical trial and birth cohort from the same urban slum in the Mirpur Thana in Dhaka, Bangladesh, as the DBC and Cryptosporidiosis Birth Cohort (CBC) (see below). PROVIDE was specifically designed to assess the influence of various factors on oral vaccine efficacy among children in areas with high poverty, urban overcrowding, and poor sanitation. The 2-by-2 factorial design looked specifically at the efficacy of the 2-dose Rotarix oral rotavirus vaccine and oral polio vaccine (OPV) with an inactivated polio vaccine (IPV) boost over the first 2 years of life. All participants were from the Mirpur area of Dhaka, Bangladesh, with pregnant mothers recruited from the community by female Bangladeshi FRAs. Each participant had 15 scheduled follow-up clinic visits, as well as biweekly diarrhea surveillance through home visits by FRAs. The presence of *Cryptosporidium* in diarrheal samples was determined by ELISA. Consistently with the DBC phenotype definition, cases had at least one diarrheal sample positive for *Cryptosporidium* within the first year of life. Controls had at least one diarrheal sample available for testing, but none were positive for *Cryptosporidium*. Severity of diarrhea was determined with the Ruuska score, which assesses severity as a function of diarrhea length, clinical symptoms, and other clinical features (48).

**Cryptosporidiosis Birth Cohort study design.** The Cryptosporidiosis Birth Cohort (“Cryptosporidiosis and Enteropathogens in Bangladesh”; ClinicalTrials.gov registration no. NCT02764918) is a prospective longitudinal birth cohort study in two sites in Bangladesh. The first site is in an urban, economically depressed neighborhood of Mirpur, and the second is in Mirzapur, a rural subdistrict 60 km northwest of Dhaka. The two birth cohorts were established in parallel, with the objective of understanding the incidence of cryptosporidiosis, the acquired immune response, and host genetic susceptibility to cryptosporidiosis in Bangladeshi children. Pregnant women were recruited and screened, and infants were enrolled at birth. Participants were followed twice-weekly with in-home visits to monitor for child morbidity and diarrhea for 24 months. Infant length and weight were measured every 3 months, and weight-for-age and length-for-age adjusted Z-scores were determined using World Health Organization Anthro software (version 3.2.2). Stool samples were collected during diarrheal illness and once per month for surveillance. Stool was tested for *Cryptosporidium* by quantitative PCR (qPCR) assay modified from a method reported previously by Liu et al. (49). A cycle threshold value of 40 was used. The pan-*Cryptosporidium* primers and probes target the 18S gene in multiple species known to infect humans (4).

**Genotype data.** DNA for all three cohorts was extracted from blood samples collected in the first few months of follow-up. The Dhaka Birth Cohort (DBC) and PROVIDE Study data were generated and cleaned as described previously (45). A summary of quality control (QC) procedures is provided in Fig. S1 in the supplemental material. Briefly, a total of 396 children in the DBC were genotyped on three different Illumina arrays. Imputation to 1000Genomes phase 3 data was performed for all individuals. After postimputation QC, which included additional filtering for relatedness and for poorly imputed variants, a total of 396 individuals and 10.2 million SNPs were included in the DBC data freeze. For PROVIDE, a total



of 541 individuals were genotyped on a Multi-Ethnic Genotyping Array (MEGA) (Illumina). After standard quality control measures (including the use of minor allele frequency values of  $>0.5\%$  and missingness values of  $<5\%$ ) were applied and first-degree-related individuals removed, a total of 499 individuals remained. After imputation to 1000Genomes and subsequent postimputation QC, a total of 499 individuals and 10.8 million genetic variants remained. For CBC, a total of 630 individuals were genotyped on a Multi-Ethnic Global Array (MEGA) (Illumina). One individual was removed for first-degree relatedness ( $PI\_HAT > 0.2$ ), 31 individuals were removed as PCA outliers, and 3 individuals were removed for heterozygosity. No individuals or SNPs were removed for missingness ( $>5\%$ ). Additional SNP-level filters included the use of minor allele frequency (MAF) values of  $<0.5\%$  ( $M = 751,869$ ) and Hardy-Weinberg equilibrium  $P$  values of  $<10^{-5}$  ( $M = 85$ ). After all QC steps, CryptoCohort genotype data included 594 individuals and 826,228 SNPs. Phasing in of SHAPEIT2 (50) was followed by imputation to 1000 Genomes phase 3 data (1000Genomes) (17) performed with IMPUTE2 (51, 52). All three studies were separately imputed to 1000Genomes.

**Cross-study genetic data harmonization.** After imputation, all three data sets (DBC, PROVIDE, and CBC) were double-checked for relatedness (both within each study and between studies) to ensure independence. One individual from each pair of related individuals was dropped in a manner consistent with the first or second degree of relatedness ( $PI\_HAT > 0.2$ ). Individual outliers for heterozygosity ( $F = >5$  standard deviations from the mean) were also excluded from further analysis. A total of 85 individuals were dropped from DBC, 9 from PROVIDE, and 34 from CBC. Only the top principal component from the combined data set was found to be significantly associated with outcome (Fig. S6).

**Statistical analysis.** All three studies (DBC, PROVIDE, and CBC) were analyzed separately using logistic regression with an additive model accounting for imputed genotype weights in SNPTTEST (51, 53, 54). All three analyses were adjusted for length-for-age Z-score (LAZ) at 1 year of age, for sex, and for the first two principal components. The Dhaka Birth Cohort was additionally conditioned on the genotyping array to account for batch effects. We combined the three analyses in a fixed-effects meta-analysis within META. Results were filtered for  $P_{\text{net}}$  values of  $>0.05$ , minor allele frequency (MAF) of  $>5\%$ , and INFO score of  $>0.6$  in all three studies, resulting in 6,504,706 SNPs. The conditional analyses were run separately by cohort for the *PRKCA* region, with each analysis being conditioned on rs58296998 in addition to the original covariates with SNPTTEST. Results were again filtered for heterogeneity or  $P_{\text{het}}$  values of  $>0.05$ , MAF of  $>5\%$ , and INFO score of  $>0.6$  in all three studies.

**Allele frequencies.** The allele frequencies were derived from the 1000 Genomes Project phase 3 data, v5a (17). Individuals were stratified by their denoted population with first degree related individuals removed.

**GTEx and eQTL overlap GWAS results.** Expression quantitative trait loci (eQTLs) were identified through the use of the GTEx Portal (<https://www.gtportal.org/home/>) on 6 August 2018 (19). The top SNP was identified as an eQTL for *PRKCA* with  $P$  values of  $<0.001$  for multiple tissues. PrediXcan measured gene expression in 48 tissues and subsequently mapped genetic variation across the human genome to tissue-specific gene expression levels. Therefore, eQTLs are identified in a tissue-specific manner and annotated as such on the GTEx Portal.

**MetaXcan imputation and association analysis.** To impute gene expression and association with outcome from our GWAS summary statistics, we applied MetaXcan (S-PrediXcan and packaged best practices) (21). Weights were previously derived with GTEx v7 data in a population of subjects of European descent, with accompanying European-descent linkage disequilibrium metrics for the SNP covariance matrices (PredictDB Data Respository; <http://predictdb.org/>). MetaXcan was used instead of the original PrediXcan to ensure consistency in models with our GWAS. All 48 tissues were run separately for the meta-analysis results previously described. Following imputation and estimation of gene expression with outcome, we calculated weights for each gene-tissue pair as the ratio between the number of SNPs used in the model and the total number that were prespecified in the model multiplied by predicted expression performance. To determine associations across many tissues, a  $P$  value threshold of 0.001 was utilized. A strict Bonferroni correction performed for the 242,686 comparisons resulted in a  $P$  value threshold of  $0.05/242,686 = 2.06 \times 10^{-7}$ , according to which no comparison yielded a statistically significant result. The relationships of allele frequencies in European and South Asian populations with PrediXcan weights were examined to assess prediction capacity (Fig. S5 and S7).

**Gene set enrichment analysis.** Gene set enrichment analysis was conducted on the described previously imputed gene expression data summary statistics from MetaXcan. For each gene, we selected the tissue corresponding to the smallest  $P$  value. Using the program GIGSEA (Genotype Imputed Gene Set Enrichment Analysis [55]), we tested for associations of 453 curated gene sets defined by MSigDB hallmark gene sets (56), as well as KEGG (Kyoto Encyclopedia of Genes and Genomes; <https://www.kegg.jp>) and BioCarta (57) gene sets (58). To account for redundancy with overlapping gene sets, we utilized the weighted multiple linear regression model, using the matrix operation to increase speed, with a total of 1,000 permutations. A false-discovery rate of 0.05 was calculated on the ranked results.

**Data availability.** Data are publicly available from the NIH, via dbGAP, phs001478.v1.p1 (Exploration of the Biologic Basis for Underperformance of Oral Polio and Rotavirus Vaccines in Bangladesh), or by request from us. All analysis programs used are detailed above, but the actual code in R for each analysis is also available by request from us.

## SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

**FIG S1**, PDF file, 0.1 MB.

**FIG S2**, PDF file, 2.8 MB.

**FIG S3**, PDF file, 0.1 MB.

**FIG S4**, PDF file, 0.2 MB.

**FIG S5**, PDF file, 0.1 MB.

**FIG S6**, PDF file, 0.2 MB.

**FIG S7**, PDF file, 0.1 MB.

**TABLE S1**, XLSX file, 0.03 MB.

**TABLE S2**, XLSX file, 0.03 MB.

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## REFERENCES

- Sow SO, Muhsen K, Nasrin D, Blackwelder WC, Wu Y, Farag TH, Panchalingam S, Sur D, Zaidi AKM, Faruque ASG, Saha D, Adegbola R, Alonso PL, Breiman RF, Bassat Q, Tamboura B, Sanogo D, Onwuchekwa U, Manna B, Ramamurthy T, Kanungo S, Ahmed S, Qureshi S, Quadri F, Hossain A, Das SK, Antonio M, Hossain MJ, Mandomando I, Nhampossa T, Acácio S, Omoro R, Oundo JO, Ochieng JB, Mintz ED, O'Reilly CE, Berkeley LY, Livio S, Tennant SM, Sommerfelt H, Nataro JP, Ziv-Baran T, Robins-Browne RM, Mishcherkin V, Zhang J, Liu J, Houpt ER, Kotloff KL, Levine MM. 2016. The burden of cryptosporidium diarrheal disease among children <24 months of age in moderate/high mortality regions of sub-Saharan Africa and South Asia. *PLoS Negl Trop Dis* 10:e0004729. <https://doi.org/10.1371/journal.pntd.0004729>.
- Guerrant DI, Moore SR, Lima AA, Patrick PD, Schorling JB, Guerrant RL. 1999. Association of early childhood diarrhea and cryptosporidiosis with impaired physical fitness and cognitive function four–seven years later in a poor urban community in northeast Brazil. *Am J Trop Med Hyg* 61:707–713. <https://doi.org/10.4269/ajtmh.1999.61.707>.
- Mondal D, Haque R, Sack RB, Kirkpatrick BD, Petri WA. 2009. Attribution of malnutrition to cause-specific diarrheal illness: evidence from a prospective study of preschool children in Mirpur, Dhaka, Bangladesh. *Am J Trop Med Hyg* 80:824–826. <https://doi.org/10.4269/ajtmh.2009.80.824>.
- Steiner KL, Ahmed S, Gilchrist CA, Burkey C, Cook H, Ma JZ, Korpe PS, Ahmed E, Alam M, Kabir M, Tofail F, Ahmed T, Haque R, Petri WA, Faruque A. 2018. Species of cryptosporidia causing subclinical infection associated with growth faltering in rural and urban Bangladesh: a birth cohort study. *Clin Infect Dis* 67:1347–1355. <https://doi.org/10.1093/cid/ciy310>.
- Korpe PS, Haque R, Gilchrist C, Valencia C, Niu F, Lu M, Ma JZ, Petri SE, Reichman D, Kabir M, Duggal P, Petri WA. 2016. Natural history of cryptosporidiosis in a longitudinal study of slum-dwelling Bangladeshi children: association with severe malnutrition. *PLoS Negl Trop Dis* 10:e0004564. <https://doi.org/10.1371/journal.pntd.0004564>.
- Chalmers RM, Robinson G, Elwin K, Elson R. 2019. Analysis of the *Cryptosporidium* spp. and gp60 subtypes linked to human outbreaks of cryptosporidiosis in England and Wales, 2009 to 2017. *Parasit Vectors* 12:95. <https://doi.org/10.1186/s13071-019-3354-6>.
- Khalil IA, Troeger C, Rao PC, Blacker BF, Brown A, Brewer TG, Colombara DV, De Hostos EL, Engmann C, Guerrant RL, Haque R, Houpt ER, Kang G, Korpe PS, Kotloff KL, Lima AAM, Petri WA, Platts-Mills JA, Shultz DA, Forouzanfar MH, Hay SI, Reiner RC, Mokdad AH. 2018. Morbidity, mortality, and long-term consequences associated with diarrhoea from Cryptosporidium infection in children younger than 5 years: a meta-analysis study. *Lancet Glob Health* 6:e758–e768. [https://doi.org/10.1016/S2214-109X\(18\)30283-3](https://doi.org/10.1016/S2214-109X(18)30283-3).
- Kattula D, Jeyavelu N, Prabhakaran AD, Premkumar PS, Velusamy V, Venugopal S, Geetha JC, Lazarus RP, Das P, Nithyanandhan K, Gunasekaran C, Muliylil J, Sarkar R, Wanke C, Ajjampur SSR, Babji S, Naumova EN, Ward HD, Kang G. 2017. Natural history of cryptosporidiosis in a birth cohort in southern India. *Clin Infect Dis* 64:347–354. <https://doi.org/10.1093/cid/ciw730>.
- Korpe PS, Valencia C, Haque R, Mahfuz M, McGrath M, Houpt E, Kosek M, McCormick BJJ, Penataro Yori P, Babji S, Kang G, Lang D, Gottlieb M, Samie A, Bessong P, Faruque ASG, Mduma E, Nshama R, Havt A, Lima IFN, Lima AAM, Bodhidatta L, Shreshtha A, Petri WA, Ahmed T, Duggal P. 2018. Epidemiology and risk factors for cryptosporidiosis in children from 8 low-income sites: results from the MAL-ED study. *Clin Infect Dis* 67:1660–1669. <https://doi.org/10.1093/cid/ciy355>.
- Sarkar R, Kattula D, Francis MR, Ajjampur SSR, Prabhakaran AD, Jayavelu N, Muliylil J, Balraj V, Naumova EN, Ward HD, Kang G. 2014. Risk factors for cryptosporidiosis among children in a semi urban slum in southern India: a nested case-control study. *Am J Trop Med Hyg* 91:1128–1137. <https://doi.org/10.4269/ajtmh.14-0304>.
- Bennett JE, Dolin R, Blaser MJ. 2015. Mandell, Douglas, and Bennett's principles and practice of infectious diseases, 8th ed. Elsevier/Saunders, Philadelphia, PA.
- Gilchrist CA, Cotton JA, Burkey C, Arju T, Gilmartin A, Lin Y, Ahmed E, Steiner K, Alam M, Ahmed S, Robinson G, Zaman SU, Kabir M, Sanders M, Chalmers RM, Ahmed T, Ma JZ, Haque R, Faruque ASG, Berriman M, Petri WA. 2018. Genetic diversity of *Cryptosporidium hominis* in a Bangladeshi community as revealed by whole-genome sequencing. *J Infect Dis* 218:259–264. <https://doi.org/10.1093/infdis/jiy121>.
- Kirkpatrick BD, Haque R, Duggal P, Mondal D, Larsson C, Peterson K, Akter J, Lockhart L, Khan S, Petri WA. 2008. Association between Cryptosporidium infection and human leukocyte antigen class I and class II alleles. *J Infect Dis* 197:474–478. <https://doi.org/10.1086/525284>.
- Carmolli M, Duggal P, Haque R, Lindow J, Mondal D, Petri WA, Mourningstar P, Larsson CJ, Sreenivasan M, Khan S, Kirkpatrick BD. 2009. Deficient serum mannose-binding lectin levels and MBL2 polymorphisms increase the risk of single and recurrent *Cryptosporidium* infec-

- tions in young children. *J Infect Dis* 200:1540–1547. <https://doi.org/10.1086/606013>.
15. Kelly P, Jack DL, Naeem A, Mandanda B, Pollok RC, Klein NJ, Turner MW, Farthing MJ. 2000. Mannose-binding lectin is a component of innate mucosal defense against *Cryptosporidium parvum* in AIDS. *Gastroenterology* 119:1236–1242. <https://doi.org/10.1053/gast.2000.19573>.
  16. Wojcik GL, Korpe P, Marie C, Mychaleckyj J, Kirkpatrick BD, Rich SS, Concannon P, Faruque ASG, Haque R, Petri WA, Jr, Duggal P. 2019. Genome-wide association study of cryptosporidiosis in infants implicates PRKCA. *bioRxiv* <https://doi.org/10.1101/819052>.
  17. 1000 Genomes Project Consortium, Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, Korbel JO, Marchini JL, McCarthy S, McVean GA, Abecasis GR. 2015. A global reference for human genetic variation. *Nature* 526:68–74. <https://doi.org/10.1038/nature15393>.
  18. eGTEx Project. 2017. Enhancing GTEx by bridging the gaps between genotype, gene expression, and disease. *Nat Genet* 49:1664–1670. <https://doi.org/10.1038/ng.3969>.
  19. GTEx Consortium. 2015. Human genomics. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue gene regulation in humans. *Science* 348:648–660. <https://doi.org/10.1126/science.1262110>.
  20. Gamazon ER, Wheeler HE, Shah KP, Mozaffari SV, Aquino-Michaels K, Carroll RJ, Eyler AE, Denny JC, GTEx Consortium, Nicolae DL, Cox NJ, Im HK. 2015. A gene-based association method for mapping traits using reference transcriptome data. *Nat Genet* 47:1091–1098. <https://doi.org/10.1038/ng.3367>.
  21. Barbeira A, Dickinson SP, Torres JM, Bonazzola R, Zheng J, Torstenson ES, Wheeler HE, Shah KP, Edwards T, Garcia T, GTEx Consortium, Nicolae D, Cox NJ, Im HK. 2016. Integrating tissue specific mechanisms into GWAS summary results. *BioRxiv* <https://doi.org/10.1101/045260>.
  22. Di Mari JF, Mifflin RC, Powell DW. 2005. The role of protein kinase C in gastrointestinal function and disease. *Gastroenterology* 128:2131–2146. <https://doi.org/10.1053/j.gastro.2004.09.078>.
  23. Meisel M, Hermann-Kleiter N, Hinterleitner R, Gruber T, Wachowicz K, Pfeiffer-Obermair C, Fresser F, Leitges M, Soldani C, Viola A, Kaminski S, Baier G. 2013. The kinase PKC $\alpha$  selectively upregulates interleukin-17A during Th17 cell immune responses. *Immunity* 38:41–52. <https://doi.org/10.1016/j.immuni.2012.09.021>.
  24. Zhao GH, Fang YQ, Ryan U, Guo YX, Wu F, Du SZ, Chen DK, Lin Q. 2016. Dynamics of Th17 associating cytokines in *Cryptosporidium parvum*-infected mice. *Parasitol Res* 115:879–887. <https://doi.org/10.1007/s00436-015-4831-2>.
  25. Clayburgh DR, Musch MW, Leitges M, Fu Y-X, Turner JR. 2006. Coordinated epithelial NHE3 inhibition and barrier dysfunction are required for TNF-mediated diarrhea in vivo. *J Clin Invest* 116:2682–2694. <https://doi.org/10.1172/JCI29218>.
  26. Sun A, Zhang H, Pang F, Niu G, Chen J, Chen F, Zhang J. 2018. Essential genes of the macrophage response to *Staphylococcus aureus* exposure. *Cell Mol Biol Lett* 23:25. <https://doi.org/10.1186/s11658-018-0090-4>.
  27. Wu Y, Zhang L, Zhang Y, Zhen Y, Liu S. 2018. Bioinformatics analysis to screen for critical genes between survived and non-survived patients with sepsis. *Mol Med Rep* 18:3737–3743. <https://doi.org/10.3892/mmr.2018.9408>.
  28. Arenas AF, Salcedo GE, Gomez-Marin JE. 2017. R script approach to infer toxoplasma infection mechanisms from microarrays and domain-domain protein interactions. *Bioinform Biol Insights* 11:1177932217747256. <https://doi.org/10.1177/1177932217747256>.
  29. Assani K, Shrestha CL, Robledo-Avila F, Rajaram MV, Partida-Sanchez S, Schlesinger LS, Kopp BT. 2017. Human cystic fibrosis macrophages have defective calcium-dependent protein kinase C activation of the NADPH oxidase, an effect augmented by Burkholderia cenocepacia. *J Immunol* 198:1985–1994. <https://doi.org/10.4049/jimmunol.1502609>.
  30. Wang W, Wang Y, Debing Y, Zhou X, Yin Y, Xu L, Herrera Carrillo E, Brandsma JH, Poot RA, Berkhout B, Neyts J, Peppelenbosch MP, Pan Q. 2017. Biological or pharmacological activation of protein kinase C alpha constrains hepatitis E virus replication. *Antiviral Res* 140:1–12. <https://doi.org/10.1016/j.antiviral.2017.01.005>.
  31. Chen X-M, O'Hara SP, Huang BQ, Splinter PL, Nelson JB, LaRusso NF. 2005. Localized glucose and water influx facilitates *Cryptosporidium parvum* cellular invasion by means of modulation of host-cell membrane protrusion. *Proc Natl Acad Sci U S A* 102:6338–6343. <https://doi.org/10.1073/pnas.0408563102>.
  32. Chen X-M, Splinter PL, Tietz PS, Huang BQ, Billadeau DD, LaRusso NF. 2004. Phosphatidylinositol 3-kinase and frabin mediate *Cryptosporidium parvum* cellular invasion via activation of Cdc42. *J Biol Chem* 279:31671–31678. <https://doi.org/10.1074/jbc.M401592200>.
  33. Crane JK, Oh JS. 1997. Activation of host cell protein kinase C by enteropathogenic *Escherichia coli*. *Infect Immun* 65:3277–3285. <https://doi.org/10.1128/IAI.65.8.3277-3285.1997>.
  34. Sukumaran SK, Prasadarao NV. 2002. Regulation of protein kinase C in *Escherichia coli* K1 invasion of human brain microvascular endothelial cells. *J Biol Chem* 277:12253–12262. <https://doi.org/10.1074/jbc.M110740200>.
  35. Keys KL, Mak ACY, White MJ, Eckalbar WL, Dahl AW, Mefford J, Mikhaylova AV, Contreras MG, Elhawary JR, Eng C, Hu D, Hunstman S, Oh SS, Salazar S, Lenoir MA, Ye JC, Thornton TA, Zaitlen N, Burchard EG, Gignoux CR. 2019. On the cross-population generalizability of gene expression prediction models. *bioRxiv* <https://doi.org/10.1101/552042>.
  36. Yang S-K, Hong M, Zhao W, Jung Y, Tayebi N, Ye BD, Kim K-J, Park SH, Lee I, Shin HD, Cheong HS, Kim LH, Kim H-J, Jung S-A, Kang D, Youn H-S, Liu J, Song K. 2013. Genome-wide association study of ulcerative colitis in Koreans suggests extensive overlapping of genetic susceptibility with Caucasians. *Inflamm Bowel Dis* 19:954–966. <https://doi.org/10.1097/MIB.0b013e3182802ab6>.
  37. Genetics Consortium UI, Barrett JC, Lee JC, Lees CW, Prescott NJ, Anderson CA, Phillips A, Wesley E, Parnell K, Zhang H, Drummond H, Nimmo ER, Massey D, Blaszczyk K, Elliott T, Cotterill L, Dallal H, Lobo AJ, Mowat C, Sanderson JD, Jewell DP, Newman WG, Edwards C, Ahmad T, Mansfield JC, Satsangi J, Parkes M, Mathew CG, Wellcome Trust Case Control Consortium 2, Donnelly P, Peltonen L, Blackwell JM, Bramon E, Brown MA, Casas JP, Corvin A, Craddock N, Deloukas P, Duncanson A, Jankowski J, Markus HS, Mathew CG, McCarthy MI, Palmer CNA, Plomin R, Rautanen A, Sawcer SJ, Samani N, et al. 2009. Genome-wide association study of ulcerative colitis identifies three new susceptibility loci, including the HNF4A region. *Nat Genet* 41:1330–1334. <https://doi.org/10.1038/ng.483>.
  38. Ellinghaus D, Psoriasis Association Genetics Extension (PAGE), Jostins L, Spain SL, Cortes A, Bethune J, Han B, Park YR, Raychaudhuri S, Pouget JG, Hübenenthal M, Folseraas T, Wang Y, Esko T, Metspalu A, Westra H-J, Franke L, Pers TH, Weersma RK, Collif J, D'Amato M, Halfvarson J, Jensen AB, Lieb W, Degenhardt F, Forstner AJ, Hofmann A, Schreiber S, Mrowietz U, Juran BD, Lazaridis KN, Brunak S, Dale AM, Trembath RC, Weidinger S, Weichenthal M, Ellinghaus E, Elder JT, Barker JNWN, Andreassen OA, McGovern DP, Karlsen TH, Barrett JC, Parkes M, Brown MA, Franke A. 2016. Analysis of five chronic inflammatory diseases identifies 27 new associations and highlights disease-specific patterns at shared loci. *Nat Genet* 48:510–518. <https://doi.org/10.1038/ng.3528>.
  39. Silverberg MS, Cho JH, Rioux JD, McGovern DPB, Wu J, Annesse V, Achkar J-P, Goyette P, Scott R, Xu W, Barmada MM, Klei L, Daly MJ, Abraham C, Bayless TM, Bossa F, Griffiths AM, Ippoliti AF, Lahaie RG, Latiano A, Paré P, Proctor DD, Regueiro MD, Steinhardt AH, Targan SR, Schumm LP, Kistner EO, Lee AT, Gregersen PK, Rotter JI, Brant SR, Taylor KD, Roeder K, Duerr RH. 2009. Ulcerative colitis-risk loci on chromosomes 1p36 and 12q15 found by genome-wide association study. *Nat Genet* 41:216–220. <https://doi.org/10.1038/ng.275>.
  40. Anderson CA, Boucher G, Lees CW, Franke A, D'Amato M, Taylor KD, Lee JC, Goyette P, Imielinski M, Latiano A, Lagacé C, Scott R, Amininejad L, Bumpstead S, Baidoo L, Baldassano RN, Barclay M, Bayless TM, Brand S, Büning C, Colombel J-F, Denson LA, De Vos M, Dubinsky M, Edwards C, Ellinghaus D, Fehrmann RSN, Floyd JAB, Florin T, Franchimont D, Franke L, Georges M, Glas J, Glazer NL, Guthery SL, Haritunians T, Hayward NK, Hugot J-P, Jobin G, Laukens D, Lawrence J, Lémann M, Levine A, Libioulle C, Louis E, McGovern DP, Milla M, Montgomery GW, Morley KI, Mowat C, et al. 2011. Meta-analysis identifies 29 additional ulcerative colitis risk loci, increasing the number of confirmed associations to 47. *Nat Genet* 43:246–252. <https://doi.org/10.1038/ng.764>.
  41. Franke A, Balschun T, Sina C, Ellinghaus D, Häslér R, Mayr G, Albrecht M, Wittig M, Buchert E, Nikolaus S, Gieger C, Wichmann HE, Sventoraityte J, Kupcinskas L, Onnie CM, Gazouli M, Anagnou NP, Strachan D, McArdle WL, Mathew CG, Rutgeerts P, Vermeire S, Vatn MH, IBSEN Study Group, Krawczak M, Rosenstiel P, Karlsen TH, Schreiber S. 2010. Genome-wide association study for ulcerative colitis identifies risk loci at 7q22 and 22q13 (IL17REL). *Nat Genet* 42:292–294. <https://doi.org/10.1038/ng.553>.
  42. McGovern DPB, NIDDK IBD Genetics Consortium, Gardet A, Törkvist L, Goyette P, Essers J, Taylor KD, Neale BM, Ong RTH, Lagacé C, Li C, Green T, Stevens CR, Beauchamp C, Fleshner PR, Carlson M, D'Amato M, Halfvarson J, Hibberd ML, Lördal M, Padyukov L, Andriulli A, Colombo E, Latiano A, Palmieri O, Bernard E-J, Deslandres C, Hommes DW, de Jong DJ, Stokkers PC, Weersma RK, Sharma Y, Silverberg MS, Cho JH, Wu J,

- Roeder K, Brant SR, Schumm LP, Duerr RH, Dubinsky MC, Glazer NL, Haritunians T, Ippoliti A, Melmed GY, Siscovick DS, Vasiliauskas EA, Targan SR, Annese V, Wijmenga C, et al. 2010. Genome-wide association identifies multiple ulcerative colitis susceptibility loci. *Nat Genet* 42: 332–337. <https://doi.org/10.1038/ng.549>.
43. Jostins L, International IBD Genetics Consortium (IBDGC), Ripke S, Weersma RK, Duerr RH, McGovern DP, Hui KY, Lee JC, Schumm LP, Sharma Y, Anderson CA, Essers J, Mitrovic M, Ning K, Cleyne I, Theatre E, Spain SL, Raychaudhuri S, Goyette P, Wei Z, Abraham C, Achkar J-P, Ahmad T, Amininejad L, Ananthakrishnan AN, Andersen V, Andrews JM, Baidoo L, Balschun T, Bampton PA, Bitton A, Boucher G, Brand S, Büning C, Cohain A, Cichon S, D'Amato M, De Jong D, Devaney KL, Dubinsky M, Edwards C, Ellinghaus D, Ferguson LR, Franchimont D, Fransen K, Geary R, Georges M, Gieger C, et al. 2012. Host-microbe interactions have shaped the genetic architecture of inflammatory bowel disease. *Nature* 491:119–124. <https://doi.org/10.1038/nature11582>.
44. de Lange KM, Moutsianas L, Lee JC, Lamb CA, Luo Y, Kennedy NA, Jostins L, Rice DL, Gutierrez-Achury J, Ji S-G, Heap G, Nimmo ER, Edwards C, Henderson P, Mowat C, Sanderson J, Satsangi J, Simmons A, Wilson DC, Tremelling M, Hart A, Mathew CG, Newman WG, Parkes M, Lees CW, Uhlig H, Hawkey C, Prescott NJ, Ahmad T, Mansfield JC, Anderson CA, Barrett JC. 2017. Genome-wide association study implicates immune activation of multiple integrin genes in inflammatory bowel disease. *Nat Genet* 49:256–261. <https://doi.org/10.1038/ng.3760>.
45. Wojcik GL, Marie C, Abhyankar MM, Yoshida N, Watanabe K, Mentzer AJ, Carstensen T, Mychaleckyj J, Kirkpatrick BD, Rich SS, Concannon P, Haque R, Tsokos GC, Petri WA, Duggal P, Wojcik GL, Marie C, Abhyankar MM, Yoshida N, Watanabe K, Mentzer AJ, Carstensen T, Mychaleckyj J, Kirkpatrick BD, Rich SS, Concannon P, Haque R, Tsokos GC, Petri WA, Duggal P. 2018. Genome-wide association study reveals genetic link between diarrhea-associated *Entamoeba histolytica* infection and inflammatory bowel disease. *mBio* 9:e01668-18. <https://doi.org/10.1128/mBio.01668-18>.
46. Liu T-L, Fan X-C, Li Y-H, Yuan Y-J, Yin Y-L, Wang X-T, Zhang L-X, Zhao G-H. 2018. Expression profiles of mRNA and lncRNA in HCT-8 cells infected with *Cryptosporidium parvum* Ild subtype. *Front Microbiol* 9:1409. <https://doi.org/10.3389/fmicb.2018.01409>.
47. Baier G, Wagner J. 2009. PKC inhibitors: potential in T cell-dependent immune diseases. *Curr Opin Cell Biol* 21:262–267. <https://doi.org/10.1016/j.ceb.2008.12.008>.
48. Ruuska T, Vesikari T. 1990. Rotavirus disease in Finnish children: use of numerical scores for clinical severity of diarrhoeal episodes. *Scand J Infect Dis* 22:259–267. <https://doi.org/10.3109/00365549009027046>.
49. Liu J, Gratz J, Amour C, Kibiki G, Becker S, Janaki L, Verweij JJ, Taniuchi M, Sobuz SU, Haque R, Haverstick DM, Houpt ER. 2013. A laboratory-developed TaqMan array card for simultaneous detection of 19 enteropathogens. *J Clin Microbiol* 51:472–480. <https://doi.org/10.1128/JCM.02658-12>.
50. Delaneau O, Marchini J, Zagury J-F. 2011. A linear complexity phasing method for thousands of genomes. *Nat Methods* 9:179–181. <https://doi.org/10.1038/nmeth.1785>.
51. Marchini J, Howie B, Myers S, McVean G, Donnelly P. 2007. A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat Genet* 39:906–913. <https://doi.org/10.1038/ng2088>.
52. Howie BN, Donnelly P, Marchini J. 2009. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. *PLoS Genet* 5:e1000529. <https://doi.org/10.1371/journal.pgen.1000529>.
53. Wellcome Trust Case Control Consortium. 2007. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature* 447:661–678. <https://doi.org/10.1038/nature05911>.
54. Marchini J, Howie B. 2010. Genotype imputation for genome-wide association studies. *Nat Rev Genet* 11:499–511. <https://doi.org/10.1038/nrg2796>.
55. Zhu S, Qian T, Hoshida Y, Shen Y, Yu J, Hao K. 2019. GIGSEA: genotype imputed gene set enrichment analysis using GWAS summary level data. *Bioinformatics* 35:160–163. <https://doi.org/10.1093/bioinformatics/bty529>.
56. Liberzon A, Birger C, Thorvaldsdóttir H, Ghandi M, Mesirov JP, Tamayo P. 2015. The Molecular Signatures Database (MSigDB) hallmark gene set collection. *Cell Syst* 1:417–425. <https://doi.org/10.1016/j.cels.2015.12.004>.
57. Nishimura D. 2001. BioCarta. Biotech Software & Internet Report 2:117–120. <https://doi.org/10.1089/152791601750294344>.
58. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, Paulovich A, Pomeroy SL, Golub TR, Lander ES, Mesirov JP. 2005. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A* 102:15545–15550. <https://doi.org/10.1073/pnas.0506580102>.