

A candidate single nucleotide polymorphism in the 3' untranslated region (rs17878624) of survivin gene for NSCLC

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Abstract: Survivin is a gene that locates on human chromosome 17q25 and contains 142 amino acid. Survivin (BIRC5) is the first one of the found inhibitors of apoptosis proteins (IAPs) that is an important protein family and regulates apoptosis. It is expressed particularly in cancer cells. 3'UTR region of gene has components that is necessary for gene function and this region plays a critical role in the regulation of posttranscriptional regulation of the gene expression. Therefore, polymorphisms in this region may affect the function of the gene. The purpose of the study is to investigate possible relationship, that is associated with development and prognosis of the disease, between the 3'UTR region (rs17878624) polymorphism and NSCLC in a Turkish society.

Key words: NSCLC; Survivin; Gene polymorphism; 3'UTR.

Introduction

Lung cancer is the most widely-spread cancer in the world since 1985. It constitutes about 12-13% of all new cancer cases and 29% of all cancer death worldwide (1). Lung cancer has been classified into Small Cell Lung Cancer (SCLC) and Non-Small Cell Lung Cancer (NSCLC) by the World Health Organization. Although, conventional cancer treatment options, surgery and chemotherapy, are applied to this cancer, overall outcome is poor due to recurrence of disease in most of cases and resistance of tumor to great number of anti-cancer agents (2). New, marker is needed to identify high-risk individuals and predict disease outcomes. Survivin seems to be candidate gene for these purposes in lung cancer (especially for NSCLC). It is the gene that encodes for survivin protein which belongs to the human Inhibitors of Apoptosis Protein (IAP) family. IAPs family regulates the cell division preventing the programmed cell death – apoptosis - through inhibiting certain apoptotic proteins called caspases (3,4). Hence, survivin plays an essential role in the development and progression of neoplastic processes suppressing the cancer cell death, thus facilitating the growth of these cells (2). To date, there are eight members of IAPs family. These proteins usually consist of one to three baculovirus IAP repeat (BIR) domains, which are responsible for anti-apoptotic activity, and either a C-terminal RING or caspase activation recruitment (CARD) domain. However, survivin has unique structure containing only a single BIR domain and lacking RING and (CARD) domains (5). Survivin protein is significantly expressed in embryonic and fetal tissues and homozygous survivin deletion leads to early embryonic death, showing its essential

role in cell development, differentiation and homeostasis. Whereas its amount is undetectable or very low in terminally differentiated adult tissues. Studies indicate that in contrast to normal cases in variety human cancers including lung cancer survivin gene is dramatically up-regulated (6,7). These findings prove the importance of above mentioned gene in carcinogenesis and thus makes it potential marker for early diagnosis and prognostic point of view. Genetic variation is one of the main factors that can affect the gene expression and thus can mediate the individual's predisposition to cancer. Single-nucleotide polymorphism (SNP) is most common DNA sequence variation type. Identifying the distribution frequency of certain SNP in a population is broadly used strategy in order to predict the risk and prognosis of a cancer (8). There are several SNPs identified to be related with the different region of survivin gene (9,10). The aim of this study to identify whether there is any correlation between a certain SNP (rs17878624) that is located on 3'UTR of survivin gene and NSCLC in Turkish population.

Materials and Methods

Study population and collection of specimens

In the study 163 diagnosed NSCLC patients and 151 healthy controls were examined. NSCLC patients were recruited from the Yedikule Chest Diseases and Thoracic Surgery Training Research Hospital, Istanbul. Samples were collected from Istanbul Yedikule Chest Diseases and Thoracic Surgery Training Hospital Department of Thoracic Surgery clinic. The mean ages of the patients and controls were 60.7 ± 9.57 years and 57.9 ± 9.39 years respectively. The percentage of females was 5.5%

Table 1. Characteristics and laboratory parameters of non-small cell lung cancer patients and controls.

Characteristics	NSCLC patients (N=163)	Control group (N=151)	P value
Male N (%)	154 (94.5)	114 (75.5)	
Female N (%)	9 (5.5)	37 (24.5)	
Age (years, means \pm SD)	60,7 \pm 9,57	57,9 \pm 9,39	0,814
Smoking history (pack/years)	48,78 \pm 30,850	38,98 \pm 27,874	0,817
WBC ($10^3/mm^3$)	9,11 \pm 3,171	6,62 \pm 1,679	0,001
FEV1	65,64 \pm 18,213	96,59 \pm 19,509	0,808
FVC	68,8934 \pm 17,18806	98,0595 \pm 16,54910	0,537
FEV1/FVC	78,8142 \pm 16,07904	96,2329 \pm 13,86570	0,129

*P obtained by the Student *t*-test. Data are reported as number (percentage in parentheses) or as means \pm standard deviation.

for patients and 24.5% for controls, and percentage of males was 94.5% for patients and 75.5% for controls (Table 1.). In NSCLC group, all subjects were diagnosed and confirmed with histopathological examination. One hundred fifty-one healthy persons without any malignancy were selected for the control group that comprised only individuals with a negative family history of cancer. All participants signed an informed consent before enrollment and Institutional Ethical committee approval was obtained for the study. Pulmonary function test and routine biochemical examinations were done after test for voluntaries. The study protocol was approved by both the Ethical Committee of the Istanbul Faculty of Medicine (July 08, 2010 No:376).

DNA extraction

DNA was isolated from the blood leukocytes in 10 ml EDTA by the method of Miller et al. based on sodium dodecyl sulphate lysis, ammonium acetate extraction, and ethanol precipitation (11,12). The concentration and purity of DNA were determined using an ND-1000 spectrophotometer (NanoDrop Technologies Inc., Wilmington, DE, USA) at A260 and A280. Template DNA (0.5–1.0 μ g) was used in a PCR under sterile conditions.

Genotyping

For the genotyping studies, the polymerase chain reaction and restriction fragment length polymorphism (PCR-RFLP) methods were used. We investigated the polymorphism in the 3' untranslated region of the Survivin gene. 0.25 μ mol/L of each primer was used for the reaction. The primers shown in Table 2. In a volume of 25 μ l containing 1,5 mM MgCl₂, 50 mM KCl, 10 mM Tris- HCl (pH:8.4), 0.16 mM each of dNTP (MBI Fermentas), and 1 unit of Taq polymerase (MBI Fermentas). Amplification was performed by initial denaturation at 94°C for 5 minutes, followed by 35 cycles with denaturation steps at 94°C for 30 seconds, annealing at 56°C for 30 seconds, and extension at 72°C for 30 seconds. The PCR programme was completed by a final extension cycle at 72°C for 5 minutes (Miller et al., 1988). The appropriate primers (Table 2.) were used to amplify the corresponding gene of the subjects by

PCR and the reaction products were digested by using the HphI restriction enzyme at 37°C. The PCR product exhibited a 741 base pair fragment for 3' untranslated region (rs17878624) polymorphism. The PCR product was digested with HphI (MBI Fermentas). Then visualized by electrophoresis on 3% agarose containing 0.5 mg/ml ethidium bromide and examined under transillumination. If there is any conflict, samples were repeated (Figure 1).

Statistical analysis

All statistical analyses were carried out using the SPSS version 17.0 for Windows. Numeric values were analyzed by student's *t*-test. Chi-square test used that Survivin 3' untranslated region of the prevalence of the genotype with alleles differences between groups with for assessing together. The relative associations between NSCLC patients and controls were assessed by calculating crude Gart's odds ratios (ODs) and 95% confidence intervals (95%CI). Threshold for significance was $p < 0.05$.

Results

Genotype and allele frequencies for Survivin 3'UTR region (rs17878624) in NSCLC patients and controls are listed in Table 3. The distribution of the survivin 3'UTR

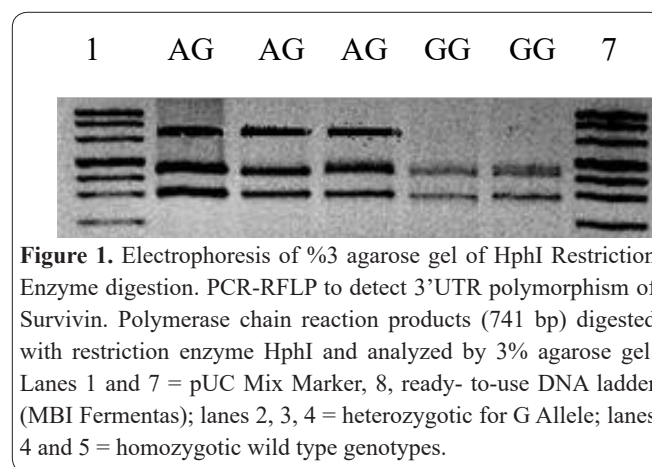


Figure 1. Electrophoresis of 3% agarose gel of HphI Restriction Enzyme digestion. PCR-RFLP to detect 3'UTR polymorphism of Survivin. Polymerase chain reaction products (741 bp) digested with restriction enzyme HphI and analyzed by 3% agarose gel. Lanes 1 and 7 = pUC Mix Marker, 8, ready- to-use DNA ladder (MBI Fermentas); lanes 2, 3, 4 = heterozygotic for G Allele; lanes 4 and 5 = homozygotic wild type genotypes.

Table 2. PCR-RFLP products of Survivin 3' UTR region (rs17878624).

Primers	PCR product	Restriction enzyme	Restriction products
3'UTR region (rs17878624) R:5-TCATCTTACGCCAGACTTCAG-3' F:5-GAGAAGTGAGGGAGGAGG-3'	741 bp	HphI	AG: 741/401/340 bp GG: 401/340 bp AA: 741 bp

bp = base pair; F = Forward; R = Reverse.

Table 3. Survivin 3'UTR polymorphism allele and genotype frequencies in patients with NSCLC and controls.

	NSCLC patients (N=163)	Control group (N=151)	P value
Survivin 3'UTR Genotype			
GG	92 (56.4%)	105 (69.5%)	0.016
AG	71 (43.6%)	46 (30.5%)	
AA	0 (0%)	0 (0%)	
Survivin 3'UTR allele			
G	255 (78.23%)	256 (84.76%)	0.035
A	71 (21.77%)	46 (15.24%)	

genotypes in control and patients was found to be significantly different ($p=0,016$). The prevalence of the survivin 3'UTR GG homozygosity was 56.4% (92/163) in patients and 69.5% (105/151) in the control group. Individuals carrying survivin 3'UTR GG genotype had a 2-fold decreased risk for NSCLC cancer ($*p=0.016$, $\chi^2:5.75$; OR: 0.568 %95CI:0.357-0.903). There was no association between 3'UTR region (rs17878624) frequencies and tumor stage, lymph node and metastasis status in NSCLC patients. Survivin 3'UTR region (rs17878624) genotype and smoking (pack/years) did not reach significance.

Discussion

During cancer formation in healthy differentiated tissues, unlike Bcl-2 and other IAPs, abnormalities in apoptosis mechanisms play a critical role. In cancer the apoptosis mechanism is highly suppressed by anti-apoptotic proteins. One of the most important of these proteins is survivin which is responsible for cell cycle regulation. Apart from survivin there have been various proteins identified which are found to be upregulated in cancer cells. One other mechanism how the apoptotic signal is bypassed is the regions in the protein structure called BIR (Baculovirus IAP repeat) which binds to the caspase proteins who are the main effectors of apoptosis in a cell. One research on survivin (13) suggests that polymorphisms in the promotor sequence of the survivin gene can cause genetic modification in many other cancer cells other than small cell lung cancer cell lines. Another research (14) demonstrates the correlation between the formation of hepatocellular carcinomas and T9809C polymorphisms located at survivin gene's 3' UTR. Studies show that nasopharyngeal cancer development is dependant on the genetic make-up mir-218 and survivin genes, 3' UTR (15). Another case demonstrated Mityaev *et al*, suggested for the treatment of lung cancer by gene therapy up regulation of survivin promoter sequences can be paramount (16). A 2012 study by Chang *et al*. proved the relationship between development of esophageal cancer and the 3' UTR sequences of survivin and CUG-BP1 (17). From a clinical perspective, prediction of individual patterns in treatment is mainly reliant on the patients genetic make-up of survivin gene regulation and genetic variations. It is safe to say these factors influence and affect the responses against treatment and acts as a precursor. For example, the high survivin upregulation in pleural effusions are proved to be a precursor of a bad prognosis by Lan *et al*. in their publication in 2010 (18-20). These studies show that survivin inhibits apoptosis and it regulates cell cycle which develops angiogenesis. These studies also show that survivin has significant expression in many kinds of human tumors.

So it can be therapeutic target.

To conclude, our case study of Turkish population genetics on the survivin gene 3' UTR polymorphisms (rs17878624) show that GG genotype provides about twice as much protection from forementioned diseases. In addition to this; we also proved that survivin is association with development and prognosis of diseases, between 3'UTR region (rs17878624) polymorphism and NSCLC in Turkish population.

Interest conflict

There is no conflict of interest among the author.

References

- Park SK, Cho LY, Yang JJ, Park B, Chang SH, Lee KS, *et al*. Scientific Committee, Korean Academy of Tuberculosis and Respiratory Diseases. Lung cancer risk and cigarette smoking, lung tuberculosis according to histologic type and gender in a population based case-control study. *Lung Cancer*. 2010;68(1):20-6.
- Rosato A, Menin C, Boldrin D, Dalla Santa S, Bonaldi L, Scaini MC, *et al*. Survivin expression impacts prognostically on NSCLC but not SCLC. *Lung Cancer*. 2013;79(2):180-6.
- Fangusaro JR, Caldas H, Jiang Y, Altura RA. Survivin: an inhibitor of apoptosis in pediatric cancer. *Pediatr Blood Cancer*. 2006 Jul;47(1):4-13.
- Hassan M, Watari H, AbuAlmaaty A, Ohba Y, Sakuragi N. Apoptosis and molecular targeting therapy in cancer. *Biomed Res Int*. 2014;150845. doi:10.1155/2014/150845.
- Gazouli M, Tzanakis N, Rallis G, Theodoropoulos G, Papaconstantinou I, Kostakis A, *et al*. Survivin -31G/C promoter polymorphism and sporadic colorectal cancer. *Int J Colorectal Dis*. 2009;24(2):145-50.
- Wagner M, Schmelz K, Dörken B, Tamm I. Epigenetic and genetic analysis of the survivin promoter in acute myeloid leukemia. *Leuk Res*. 2008;32(7):1054-60.
- Feng C, Yang F, Wang J. FBXO4 inhibits lung cancer cell survival by targeting Mcl-1 for degradation. *Cancer Gene Ther*. 2017;24(8): 342-347.
- Hsieh YS, Tsai CM, Yeh CB, Yang SF, Hsieh YH, Weng CJ. Survivin T9809C, an SNP located in 3'-UTR, displays a correlation with the risk and clinicopathological development of hepatocellular carcinoma. *Ann Surg Oncol*. 2012; Suppl 3:S625-33.
- Jang JS, Kim KM, Kang KH, Choi JE, Lee WK, Kim CH, *et al*. Polymorphisms in the survivin gene and the risk of lung cancer. *Lung Cancer*. 2008;60(1):31-9.
- Mallolas J, Rodríguez R, Gubern C, Camós S, Serena J, Castellanos M. A polymorphism in the promoter region of the survivin gene is related to hemorrhagic transformation in patients with acute ischemic stroke. *Neuromolecular Med*. 2014;16(4):856-61.
- Miller SA, Dykes DD, Polesky HF. A simple salting out procedure for extracting DNA from human nucleated cells. *Nucleic Acids Res*. 1988;16(3):1215.
- Drábek J, Petrek M. A sugar, laundry detergent, and salt method

for extraction of deoxyribonucleic acid from blood. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub.* 2002;146(2):37-9.

13. Dai J, Jin G, Dong J, Chen Y, Xu L, Hu Z, et al. Prognostic significance of survivin polymorphisms on non-small cell lung cancer survival. *J Thorac Oncol.* 2010;5(11):1748-54.

14. Huang CY, Lin CS, Tai WT, Hsieh CY, Shiau CW, Cheng AL, et al. Sorafenib enhances radiation-induced apoptosis in hepatocellular carcinoma by inhibiting STAT3. *Int J Radiat Oncol Biol Phys.* 2013;86(3):456-62.

15. Alajez NM, Lenarduzzi M, Ito E, Hui AB, Shi W, Bruce J, et al. MiR-218 suppresses nasopharyngeal cancer progression through downregulation of survivin and the SLIT2- ROBO1 pathway. *Cancer Res.* 2011;71(6):2381-91.

16. Mityaev MV, Kopantzev EP, Buzdin AA, Vinogradova TV, Sverdlov ED. Enhancer element potentially involved in human survivin gene promoter regulation in lung cancer cell lines. *Biochemistry*

(Mosc). 2010;75(2):182-91.

17. Chang ET, Donahue JM, Xiao L, Cui Y, Rao JN, Turner DJ, et al. The RNA-binding protein CUG-BP1 increases survivin expression in oesophageal cancer cells through enhanced mRNA stability. *Biochem J.* 2012; 446(1):113-23.

18. Lan CC, Wu YK, Lee CH, Huang YC, Huang CY, Tsai YH, et al. Increased survivin mRNA in malignant pleural effusion is significantly correlated with survival. *Jpn J Clin Oncol.* 2010;40(3):234-40.

19. Andre F, Schartz NE, Movassagh M, Flament C, Pautier P, Morrice P, et al. Malignant effusions and immunogenic tumour-derived exosomes. *Lancet.* 2002 Jul 27;360(9329):295-305.

20. Janviere Kabagwira and Nathan R. Wall. An Argument to Examine Exosomal Survivin Splice Variant Expression and Patient Survival in Pancreatic Cancer. *Clin Oncol.* 2017; 2: 1310.