Added dietary cobalt or vitamin B<sub>12</sub>, or injecting vitamin B<sub>12</sub> does not improve performance or indicators of ketosis in pre- and post-partum Holstein-Friesian dairy cows W.A.D.V. Weerathilake<sup>1\*</sup>, A.H. Brassington<sup>2</sup>, S.J. Williams<sup>1</sup>, W.Y. Kwong<sup>2</sup>, L.A. Sinclair<sup>1†</sup> and K.D. Sinclair<sup>2</sup> <sup>1</sup>Department of Animal Production, Welfare and Veterinary Sciences, Harper Adams University, Edgmond, Newport, Shropshire, UK, TF10 8NB <sup>2</sup>School of Biosciences, University of Nottingham, Sutton Bonington Campus, Loughborough, Leicestershire, UK, LE12 5RD \*Current address: Department of Livestock and Avian Science, Faculty of Livestock, Fisheries and Nutrition, Wayamba University of Sri Lanka, Makandura, Gonawila (NWP), 60170, Sri Lanka. †E-mail: lsinclair@harper-adams.ac.uk Short title: Cobalt and vitamin B<sub>12</sub> metabolism in dairy cows 

## **Abstract**

28

29

30

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

Vitamin  $B_{12}$  is synthesised in the rumen from cobalt and has a major role in metabolism in the peripaturient period, although few studies have evaluated the effect of the dietary inclusion of cobalt (Co), vitamin  $B_{12}$  or injecting vitamin  $B_{12}$  on the metabolism, health and performance of high yielding dairy cows. Fifty-six Holstein-Friesian dairy cows received one of four treatments from 8 weeks prior to calving to 8 weeks post calving: C, no added Co; DC, additional 0.2 mg Co/kg DM; DB, additional 0.68 mg vitamin B<sub>12</sub>/kg DM; IB, intra-muscular injection of vitamin B<sub>12</sub> to supply 0.71 mg/cow/day pre-partum and 1.42 mg/cow/day post-partum. The basal and lactation rations both contained 0.21 mg Co/kg DM. Cows were weighed and condition scored at drying off, 4 weeks prior to calving, within 24 h of calving and at 2, 4 and 8 weeks post-calving, with blood samples collected at drying off, 2 weeks pre-calving, calving and 2, 4 and 8 weeks post-calving. Liver biopsy samples were collected from all animals at drying off and 4 weeks post-calving. Live weight changed with time, but there was no effect of treatment (P>0.05), whereas cows receiving IB had the lowest mean body condition score and DB the highest (P<0.05). There was no effect of treatment on post-partum DM intake, milk yield or milk fat concentration (P>0.05) with mean values of 21.6 kg/day, 39.6 kg/day and 40.4 g/kg respectively. Cows receiving IB had a higher plasma vitamin B<sub>12</sub> concentration than those receiving any of the other treatments (P<0.001), but there was no effect (P>0.05) of treatment on homocysteine or succinate concentrations, although mean plasma methylmalonic acid concentrations were lower (P=0.019) for cows receiving IB than for Control cows. Plasma β-hydroxybutyrate concentrations increased sharply at calving followed by a decline, but there was no effect of treatment. Similarly, there was no effect (*P*>0.05) of treatment on plasma non-esterified fatty acids or glucose. Whole tract digestibility of DM and fibre measured at week 7 of lactation were similar between treatments, and there was little effect of treatment on the milk fatty acid profile except for C15:0, which was lower in cows receiving DC than IB (P<0.05). It is concluded that a basal dietary concentration of 0.21 mg Co/kg DM is sufficient to meet the requirements of high yielding dairy cows during the transition period, and there is little benefit from additional Co or vitamin B<sub>12</sub>.

**Key words:** dairy cow, digestibility, liver metabolism, milk, minerals

# **Implications**

The microbes in the rumen of dairy cows require cobalt to synthesise vitamin  $B_{12}$  which the cows then requires for efficient energy and protein metabolism, particularly during the transition from pregnancy to early lactation. This study investigated the effect of feeding different levels of dietary cobalt and supplementing the diet or injecting vitamin  $B_{12}$ , and found little benefit from providing additional amounts on cow performance, health or milk quality. This information can be used to more accurately formulate diets for dairy cows to improve health and performance and reduce diet costs.

### Introduction

During the transition period from late gestation to early lactation it is usual for dairy cows to experience an imbalance between dietary energy intake and nutrient demand for milk production (Lean et al., 2013; Raboisson *et al.*, 2014). This results in varying degrees of negative energy balance and consequently mobilisation of body adipose tissue (Raboisson *et al.*, 2014). The inability of the liver to metabolise mobilised fat can result in sub-clinical or clinical ketosis, normally defined when plasma  $\beta$ -hydroxybutyrate (3-OHB) concentrations are  $\geq$  1.2 mmol/L (McArt *et al.*, 2013). The prevalence of sub-clinical ketosis (SCK) has been reported to be approximately 22% in European dairy herds (Suthar *et al.*, 2013), whilst in North America a rate of 43%

has been reported (McArt *et al.*, 2012). Cows with SCK have been reported to have an increased odds of developing clinical ketosis, displaced abomasum and metritis of 9.5, 5.0 and 1.5 respectively (Suthar *et al.*, 2013), and a decreased probability of pregnancy (Raboisson *et al.*, 2014).

Vitamin B<sub>12</sub> has a major role in the metabolism of lipid by the liver and subsequently plays a central role in controlling SCK. One of the two main functions of vitamin B<sub>12</sub> in the dairy cow is its action as a co-factor in the transformation of methylmalonyl CoA to succinyl CoA which is then used in the Krebs cycle within the liver for the synthesis of glucose from propionate (McDowell 2000). Insufficient tissue supply of vitamin B<sub>12</sub> therefore results in an accumulation of methylmalonic acid (MMA) and ketones in the blood. The second role of vitamin B<sub>12</sub> is its requirement as a co-factor for the synthesis of methionine via the transfer of a methyl group from 5-methyl-tetrahydrofolate to homocysteine (McDowell 2000). Methionine is generally regarded as one of the first limiting amino acids in milk protein synthesis (NRC, 2001) and plays a key role in the synthesis of S-adenosylmethionine as a methyl donor (McDowell 2000), and

Within the rumen, elemental cobalt (Co) is used by bacteria to synthesise vitamin B<sub>12</sub>, and it has traditionally been considered that the rumen bacteria are able to synthesise sufficient amounts to meet the cows requirements over the periparturient period and throughout lactation, provided there is a sufficient dietary supply of Co (NRC, 2001). The current recommended level of dietary Co for dairy cows, based on NRC (2001), is 0.11 mg/kg DM, but vitamin B<sub>12</sub> synthesis in the rumen has been shown to increase linearly between 0.1 and 1.0 mg Co per kg DM (Tiffany *et al.*, 2003; 2006), and growing beef cattle are known to respond to dietary Co levels up to 0.25 mg/kg DM (Stangl *et al.*, 2000). In the well-fed dairy cow, serum vitamin B<sub>12</sub> concentrations decline during the dry period and throughout early lactation (Kincaid

consequently has a major impact on milk production.

and Socha, 2007), but crucially, production and liver-health related responses to dietary Co spanning current recommended levels have not been determined in the transition cow, highlighting an area where investigation is required. Additionally, whilst elemental Co is nontoxic, dietary Co sources have recently been re-classified under EU legislation (EU 601/2013 amended 2014) as carcinogenic, so that mineral premixes have been set to contain a maximum of 0.34 mg Co/kg DM. Rumen protected forms of vitamin B<sub>12</sub> are available, but given that vitamin B<sub>12</sub> is important in rumen function, it is questionable if rumen protected sources of vitamin B<sub>12</sub> alone are desirable substitutes for dietary Co. Consequently, there is a need to re-evaluate dairy cow Co requirements and the benefits of vitamin B<sub>12</sub> supplementation over the periparturient period in higher yielding dairy cows. The objectives of the study were to determine the effects of the dietary addition of Co, vitamin B<sub>12</sub> or the injection of vitamin B<sub>12</sub> in late gestation/early lactation on dairy cow metabolism, intake and performance.

#### Materials and methods

121 Animals and treatments

Fifty-six Holstein-Friesian dairy cows (12 primiparous and 44 multiparous) commenced the study 57 (SE ± 0.9) days pre-partum, and remained on study until 56 days post-partum. Cows were blocked according to parity (1st, 2nd and 3rd+), and previous 305 day milk yield, and randomly allocated to one of four dietary treatments. Pregnant, non-lactating cows were housed in a free stall-building from the end of lactation (approximately 8 weeks prior to calving), with *ad libitum* access to the same basal dry cow diet which was fed as a total mixed ration (TMR; Table 1) via a barrier with individual head locks. Approximately 3 days prior to calving the cows were transferred to a straw bedded calving yard where they remained until approximately 24 h post-calving. For approximately the first 7 to 10 days post-calving cows were

housed in a loose yard where they had access to a lactation TMR that contained no added Co via a feed barrier with individual head locks. Cows were then transferred to free-stall housing containing individual feeders. All cows had continual access to water.

132

133

134

135

136

137

138

139

140

141

142

143

144

145

146

147

148

149

150

151

152

153

154

155

156

157

Prior to calving the cows received one of four dietary treatments: C = no added Co; DC = an additional 0.2 mg Co/kg DM (supplied as cobalt carbonate; Anima, Krakow, Poland); DB = 0.68 mg added vitamin B<sub>12</sub>/kg DM (Impextraco, Heist-op-den Burg, Belgium) or IB = injected vitamin  $B_{12}$  (5 ml of 1000 µg vitamin  $B_{12}$ /ml resulting in 0.71 mg/day; AnimalCare, York, UK; intra muscular every 7 days). The additional level of 0.2 mg Co/kg DM in DC was chosen to provide a daily Co supply approximately twice that of C, but to be within the permitted EU legal limit of 0.34 mg/kg DM. The 0.68 mg vitamin B<sub>12</sub>/kg DM provided in DB was calculated to provide a similar predicted tissue supply of vitamin B<sub>12</sub> as DC assuming a 4 % conversion of Co to cobalamin, a loss of 80 % of supplementary vitamin  $B_{12}$  in the rumen, and 10 % absorption of vitamin B<sub>12</sub> at the duodenum (Stemme et al., 2008; Girard et al., 2009; Akins et al., 2013). The level of injected vitamin B<sub>12</sub> was chosen to provide a supranutritional tissue supply based on Girard and Matte (2005a), and therefore act as a positive control. All animals were individually supplemented with 200 g/day of ground wheat which acted as a carrier for treatments DC and DB by restraining each cow using the individual head-lock gates until the daily allowance had been consumed, and assuming a DM intake of 12 kg/cow/day (NRC, 2001). From calving until approximately 8-10 days post-calving all cows were fed a TMR (Table 1) containing no additional Co or vitamin B<sub>12</sub>. During this period all animals were supplemented with 200 g/day of ground wheat as a carrier to provide treatments DC and DB, assuming a DM intake of approximately 20 kg/day (NRC 2001), with IB receiving 10 ml of 1000 µg vitamin B<sub>12</sub>/ml injected every 7 days, resulting in 1.42 mg/day. For the remainder of the study the supplements were included into the TMR, and intake monitored using roughage intake feeders (Hokofarm, Marknesse, The Netherlands).

Experimental routine

158

159

160

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

The cows were weighed and condition scored (Ferguson et al., 1994) at drying off, 4 weeks post-drying off, within 24 h of calving, and at weeks 2, 4 and 8 post-calving. Samples of the TMR and individual forages were collected weekly: the forage sample was oven dried and the quantity of lucerne and maize silage adjusted to achieve the desired ratio, and the TMR sample frozen at -20°C prior to analysis. Blood samples were collected by jugular venepuncture at 1000 h on the day of drying off, 6 weeks post-drying (approximately 2 weeks prior to calving), within 24 h of calving, and 2, 4 and 8 weeks post calving. Liver samples were collected by biopsy through the 11th intercostal space from all animals at drying off and at 4 weeks post-calving; samples were snap frozen in liquid N and stored at -80°C until subsequent analysis. Cows were milked twice daily at approximately 0600 and 1700 h, with yield recorded at each milking, and samples taken weekly on consecutive am and pm milkings for subsequent analysis. Additional milk samples were collected during week 7 of lactation for the determination of the fatty acid (FA) profile. Whole tract digestibility was estimated during week 7 of lactation by collecting faecal samples from each cow over 5 days at approximately 0800 and 1400 h (Van Keulen and Young, 1977), and stored at -20°C for subsequent analysis.

Chemical analysis

Weekly TMR samples were bulked within month and analysed according to AOAC (2012) for DM (934.01) and CP (988.05), whilst NDF and ADF were determined according to Van Soest *et al.* (1991). The determination of NDF was conducted without sodium sulphite, with alpha-amylase and was corrected for ash. Samples were also

analysed for major and trace minerals as described by Cope et al. (2009). Plasma samples were analysed for urea, glucose, non-esterified fatty acids and 3-OHB (Randox Laboratories, County Antrim, UK; kit catalogue no. TP245, UR221, AB362, GL1611, FA115 and RB 1008 with an intra-assay CV of 3.5, 1.8, 3.0 and 4.7% respectively) using a Cobas Miras Plus auto-analyser (ABX Diagnostics, Bedfordshire, UK). Plasma samples were analysed for minerals as described by Cope et al. (2009). Analysis of plasma vitamin B<sub>12</sub>, homocysteine, MMA, succinic acid (SA) and hepatic triacylglycerol (TAG) concentration (along with their intra-assay CV) are provided in Supplementary Material S1. For MMA and SA only the two most extreme treatments (C and IB) were analysed. Milk samples were analysed for fat, protein, lactose and somatic cell count by Eurofins UK (Wolverhampton, UK). Fatty acid methyl esters (FAME) in hexane were prepared from milk fat and individual FAME were determined by GLC (Hewlett Packard 6890, Wokingham, UK) fitted with a CP-Sil 88 column (100 m x 0.25 mm i.d. x 0.2 µm film) as described previously by Sinclair et al., (2015). Faecal samples were bulked between days and sampling times and analysed for acid insoluble ash (Van Keulen and Young 1977), ash (AOAC, 2012; 942.05), CP, NDF and ADF.

Statistical analysis

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

206

207

208

209

Performance and blood parameters were analysed as repeated measures analysis of variance, using Genstat 17.1 (VSN Int. Ltd., Oxford, UK). The performance in the week prior to allocation was used as a co-variate where appropriate:

205 
$$Y_{ijk} = \mu + B_i + C_j + T_k + C_j \cdot T_l + \epsilon_{ijk}$$

Where  $Y_{ijk}$  = dependent variable;  $\mu$  = overall mean;  $B_i$  = fixed effect of blocks;  $C_j$  = effect of treatment;  $T_k$  = effect of time;  $C_j.T_k$  = interaction between treatment and time, and  $\epsilon_{ijk}$  = residual error. Milk SCC and hepatic TAG concentrations were log transformed prior to analysis to homogenise variance. A subsequent analysis of TAG

levels substituted parity for treatment. Milk FA concentration and diet digestibility were analysed as a randomised block design by analysis of variance as:

 $Y_{ij} = \mu + B_i + C_j + \varepsilon_{ij}$ 

Where  $Y_{ij}$  = dependent variable;  $\mu$  = overall mean;  $B_i$  = fixed effect of blocks;  $C_j$  = effect of treatment, and  $\epsilon_{ij}$  = residual error. Results are presented as treatment means with a SED, with post-hoc analysis using Tukey's test at a 5% level of significance.

# Results

Diet analysis, intake and animal performance.

The chemical composition of the basal dry-cow diet was similar to that predicted, whereas the background concentration of Co was higher in both the dry cow and lactation diets at 0.21 mg/kg DM (Table 2). The four lactation diets had a similar mean DM, OM, CP, NDF and ADF concentration of 469 g DM/kg, 940, 168, 368 and 231 g/kg DM respectively. The four diets also had a similar concentration of macro and trace elements. In contrast, the treatments with no added Co (C, and IB) had a dietary concentration of approximately 0.21 mg Co/kg DM, whilst DC had almost double the dietary concentration of Co, with DB being 0.02 mg Co/kg DM higher than the Control.

There was an increase in DM intake (*P*<0.001) from 18.6 kg/d during week 2 of lactation to 23.7 kg/d during week 8, with a mean intake of 21.6 kg/d over the study period, but there was no effect (*P*>0.05) of dietary treatment or treatment x time interaction (Table 3). Similarly, milk yield increased (*P*<0.001) from 30.3 kg/d in week 1 to 43.6 kg/d in week 8 of lactation, but there was no effect (*P*>0.05) of treatment or time x treatment interaction on milk performance or composition, with mean values of 39.6 kg/day, 40.4 g/kg and 33.1 g/kg for milk yield, fat and protein concentration respectively. There was an effect of time (*P*<0.001) on live weight, which increased

between drying off and 4 weeks pre-calving, before decreasing immediately postcalving but there was no effect (P>0.05) of treatment, or time x treatment interaction. There was also an effect of time on BCS (P<0.001), which declined post-calving, and treatment, with cows receiving IB having the lowest and DB the highest BCS (P<0.05). Plasma metabolite and mineral concentrations, and hepatic triacylglycerol content Plasma glucose concentration decreased from 8 weeks pre-calving to calving, then increased to week 8 post calving (P<0.001), but there was no effect (P>0.05) of dietary treatment on weekly or mean concentration (Table 4 and Supplementary Figure 1a). In contrast, plasma 3-OHB concentrations increased pre-calving, with a sharp increase at calving followed by a decrease at week 2 of lactation (Supplementary Figure 1b), but there was no effect (P>0.05) of dietary treatment. There was an effect of time on plasma NEFA concentration (Supplementary Figure 1c; P<0.001), which decreased pre-calving, before increasing post-calving, but there was no effect (P>0.05) of treatment. Plasma urea concentrations declined rapidly at calving (Supplementary Figure 1d; P<0.001) but were not affected (P>0.05) by treatment. There was no effect (P>0.05) of dietary treatment on mean plasma mineral concentration, except Co, which was higher (P<0.05) in cows receiving DC or IB than C or DB, and Zn, which was higher in cows receiving DC than IB (P<0.05).

236

237

238

239

240

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

There was a treatment x time interaction (P<0.001) for plasma vitamin B<sub>12</sub> concentration, which decreased post calving in all treatments (Figure 1a), and overall was higher (P<0.001) in cows receiving IB than any of the other treatment groups (246 vs 192, 195 and 190 pmol/L for IB vs C, DC and DB respectively). There was no treatment effect on plasma concentrations of homocysteine, which decreased (P<0.001) following calving (Figure 1b). In contrast, plasma concentrations of both MMA (Figure 1c) and SA (Figure 1d) increased (P<0.001) following parturition, with an interaction between treatment and time for plasma MMA concentrations (P=0.033);

MMA being lower in IB than Control cows post-calving. There was no treatment effect on hepatic TAG concentrations pre- or post-partum (P>0.05), however, TAG concentrations were higher (P<0.001) in samples collected during the early post-partum period than during late gestation.

Diet digestibility and milk fatty acid profile

There was no effect (P>0.05) of dietary treatment on the digestibility of any of the parameters measured (P>0.05; Table 5). There were few differences in the FA profile of milk from cows fed any of the treatments, except C15:0 which was lower (P<0.05) in cows receiving supplementary Co (DC) compared to injected vitamin B<sub>12</sub> (IB; Table 6). The desaturase index calculated using *cis*-9, *trans*-11 CLA was lower (P<0.01) in cows receiving IB compared to C or DC, but there was no other effect of dietary treatment.

## Discussion

To attempt to provide a low dietary Co concentration in the current study, feed ingredients were selected to be low in Co, with the mineral/vitamin supplement formulated to contain no Co. Despite this, both the dry cow and lactation control diets contained 0.21 mg Co/kg DM, above the recommended dietary requirement of 0.11 mg/kg DM stated by NRC (2001), but considerably lower than a number of similar studies that have attempted to provide low dietary Co diets (Akins *et al.*, 2013; Girard *et al.*, 2009). The addition of 0.2 mg Co/kg DM in DC resulted in a dietary concentration approximately twice that of C, but was well within the permitted EU legal limit of 0.34 mg added Co/kg DM, or the 1.14 mg Co/kg DM limit in the total diet.

Performance and dry matter intake

The peri-parturient period in dairy cows is characterised by a phase of negative energy balance as cows transition from late gestation into early lactation, with associated

homeorhetic and homeostatic regulation of metabolic functions (McArt et al., 2013). In particular, there is a substantial increase in demand for glucose for foetal growth in late pregnancy, and for the synthesis of lactose for milk production by the mammary gland in early lactation (Lean et al., 2013). Vitamin B<sub>12</sub>, through its role as a co-enzyme for methylmalonyl-CoA mutase (EC:5.4.99.2), is crucial for the synthesis of glucose from propionate (McDowell, 2000), and a deficiency in this vitamin can be translated into a reduction in milk synthesis (NRC, 2001). Milk secretion also requires a considerable supply of amino acids, with methionine generally being regarded as one of the first rate limiting AA in most dairy cow rations (NRC, 2001), and vitamin B<sub>12</sub> has a key role in the activity of *methionine synthase* (EC:2.1.1.13) which regenerates homocysteine to methionine (McDowell 2000). In the current study, additional dietary Co or vitamin B<sub>12</sub> or parenteral supply of vitamin B<sub>12</sub> did not influence milk production or composition in early lactation. Similarly, Kincaid and Socha (2007) reported no effect of dietary Co concentration (0.15, 0.89 and 1.71 mg/kg DM) on early lactation milk yield or composition. In contrast, Juchem et al., (2012) reported a decrease in milk fat concentration, but not fat yield, when rumen protected B-vitamins were included in dairy cow rations, although the combination of vitamins used precludes any conclusion to be drawn on the relative efficacy of vitamin  $B_{12}$ . Duplessis et al., (2017) also reported no effect of weekly injections of 10 mg of vitamin B<sub>12</sub> on milk performance or composition over the transition period. Weekly intramuscular injections of vitamin B<sub>12</sub>, however, resulted in a significant increase in energy corrected milk yield of early lactation primiparous dairy cows supplemented with folic acid and methionine in the study of Girard and Matte (2005a), due to a combination of an increase in milk yield and milk fat concentration, despite dietary Co levels being stated as adequate. In contrast, NRC (2001) concluded that there was no evidence of a performance response to vitamin B<sub>12</sub> injections in dairy cows when fed adequate (i.e.

288

289

290

291

292

293

294

295

296

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311

312

above 0.11 mg/kg DM) amounts of Co. Results from the current study support that the recent EU restriction on the amount of supplementary Co that can be added to the diet should not impair dairy cow performance or health. There may, however, be some value to higher dietary levels of Co, or rumen protected vitamin  $B_{12}$  to increase the vitamin  $B_{12}$  content in milk for human consumption as suggested by Girard and Matte (2005b), particularly in situations where foods are fortified with folic acid.

Regulation of lipogenesis and lipolysis in the transition period involves a complex interaction between neural, hormonal and paracrine stimuli, along with immune cell migration that changes the structure and cellular distribution of adipose tissue (Contreras *et al.*, 2017). In the current study, both live weight and body condition score were characterised by an increase pre-partum, followed by a rapid decline post-partum, reaching a nadir at approximately 4 weeks post-calving. This rapid decline in body condition and live weight is greater than that reported in other studies that have investigated the effect of diet on body energy reserves where the nadir in energy balance is typically not reached until approximately week 8 to 10 post-partum (Duplessis *et al.*, 2017). Parenteral administration of vitamin B<sub>12</sub> resulted in the lowest mean BCS post-partum, but this was not reflected in an increase in milk or milk fat yield. Duplessis *et al.* (2017) reported no benefit to injections of vitamin B<sub>12</sub> on body condition over the transition period, although dietary levels of Co (1.45 and 1.05 mg/kg DM for the dry period and lactation respectively) were substantially higher than that used here.

Cows fed any of the dietary treatments in the current study continued to increase their DM intake throughout the first 8 weeks of lactation, but there was no effect of dietary Co level or vitamin  $B_{12}$  on intake. Several other studies have also failed to demonstrate a benefit to Co supplementation or parenteral administration of vitamin  $B_{12}$  on DM intake either pre-partum or during lactation (Akins *et al.*, 2013; Girard and

Matte, 2005a). Indeed, supra-nutritional levels of Co have been demonstrated to result in a reduction in DM intake in some studies, particularly in early lactation (Kincaid and Socha, 2007).

Blood metabolites, minerals and hepatic triacylglycerol content

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

355

356

357

358

359

360

361

362

363

364

365

Poor adaptation to negative energy balance through the transition period is associated with an elevation in plasma NEFA and 3-OHB concentrations (McArt et al., 2013). Whilst there is much debate regarding definitive cut-point concentrations of these metabolites for milk production and disease outcomes, NEFA values of 0.3 to 0.5 mmol/l pre-partum and 0.7 to 1.0 mmol/l post partum, and 3-OHB concentrations of 1.2 mmol/l, have been suggested as a good combination between sensitivity and specificity (McArt et al., 2013). Based on these assumptions, mean plasma NEFA in the current study were high at drying off and again at calving, whereas mean plasma 3-OHB concentrations were always well within accepted limits. Plasma glucose concentrations were also affected by day of sampling in the current study, decreasing sharply at 2 weeks post-calving before returning to pre-partum levels by week 4. There was, however, no effect of dietary treatment on any of these metabolites pre or postpartum, and it is apparent that vitamin B<sub>12</sub> was not limiting for cows. A number of other studies have also reported no effect of dietary Co level (Akins et al., 2013; Kincaid and Socha, 2007), dietary supplementation with vitamin B<sub>12</sub> (Graulet et al., 2007) or following parenteral administration of vitamin B<sub>12</sub> (Akins et al., 2013; Duplessis et al., 2017) on serum NEFA concentration.

A threshold for plasma vitamin  $B_{12}$  of 150 µmol/l has been suggested, above which there is little further benefit to milk performance in dairy cows (Duplessis *et al.*, 2017). Plasma vitamin  $B_{12}$  in the current study was above this threshold in cows fed any of the dietary treatments, and were approximately 28% higher in cows receiving weekly injections of vitamin  $B_{12}$ . The high demands for metabolism and secretion into

milk (Kincaid and Socha 2007) may predispose higher yielding dairy cows to vitamin B<sub>12</sub> deficiency and can be exhibited in an increase in plasma concentrations of key intermediaries in energy and amino acid metabolism. The dependency of methionine synthase and methylmalonyl-CoA mutase on vitamin B<sub>12</sub> results in plasma concentrations of homocysteine and MMA acid being good indicators of vitamin B<sub>12</sub> adequacy (Stangl et al., 2000). These authors reported a significant decline in both plasma homocysteine and MMA when dietary Co concentrations were increased above 0.2 mg/kg DM in growing beef cattle. However, in the current study there was little effect of supplementary Co, vitamin B<sub>12</sub> or parental administration of vitamin B<sub>12</sub>, and supports the performance and blood metabolite results that the basal dietary Co concentration of 0.21 mg/kg DM was sufficient to meet requirements in late pregnancy and early lactation. Despite the lack of an effect of treatment on hepatic TAG concentrations in the current study there was a 3-fold increase in concentration postcompared to pre-partum (mean values of 19 and 6 mg/g liver respectively), although mean values were still within the 1-5% liver TAG concentration that is associated with mild fatty liver (Bobe et al., 2004).

382 Whole tract digestibility

366

367

368

369

370

371

372

373

374

375

376

377

378

379

380

381

383

384

385

386

387

388

389

390

391

In addition to the requirement for the metabolism of glucose and methionine in the dairy cow, vitamin B<sub>12</sub> has been demonstrated to be an essential growth factor for efficient rumen microbial metabolism, with ruminal synthesis of vitamin B<sub>12</sub> and subsequent flow to the duodenum being significantly related to the rate of microbial protein synthesis in the rumen (Castagnino *et al.*, 2016). This has been suggested to improve fibre digestion in the rumen (Lopez Guisa and Satter, 1992), and increase the ruminal production of propionate in some studies (Tiffany *et al.*, 2003), but not others (Stemme *et al.*, 2008). Increasing dietary Co concentration from sub-optimal levels has been shown to increase whole tract digestibility quadratically in sheep, being

highest at a dietary level of 0.6 mg/kg DM (Wang *et al.*, 2007). Similarly, Kadim *et al.* (2003) reported that the parental provision of vitamin B<sub>12</sub> increased diet digestibility in goats, which was speculated to be due to an increase in intestinal absorption of nutrients or microbial production in the rumen. In the current study digestibility coefficients for DM, N, OM and fibre fractions were similar to that reported by others when feeding similar diets (Sinclair *et al.*, 2015), but there was no effect of dietary treatment, and it can be concluded that there was sufficient vitamin B<sub>12</sub> available for ruminal metabolism.

Milk fatty acids

392

393

394

395

396

397

398

399

400

401

402

403

404

405

406

407

408

409

410

411

412

413

414

415

416

The concentration of saturated FA in milk may be reduced via the action of steroyl-CoA desaturase (SCD) on both de novo and absorbed FA such as C14:0, C16:0 and C18:0, and is responsible for the majority of the *cis*-9, *trans*-11 CLA from its action on C18:1 trans-11 (Shingfield et al., 2013). Several studies have investigated the impact of the dietary inclusion, ruminal infusion or intravenous injection of Co on SCD activity, and reported a dose dependent decrease in activity by up to 72% (Leskinen et al., 2016). Infusing Co does not impact on gene expression of SCD, and its effects would therefore appear to be at a post-transcriptional level (Karlengen *et al.*, 2012). However, in previous studies that have reported a reduction in SCD activity supra-nutritional levels of Co have been fed. For example, Karlengen et al. (2012) reported no effect on the milk desaturase index in cows when fed approximately 0.2 or 19 mg Co/kg DM, but it was reduced when 270 mg/kg DM was infused into the rumen. In the current study, there was little evidence of an effect of adding approximately 0.2 mg Co/kg DM, or additional vitamin B<sub>12</sub> administered per os, on SCD activity. In contrast, SCD activity was reduced when supra-nutritional levels of vitamin B<sub>12</sub> were injected, although few other studies have examined the effect of vitamin B<sub>12</sub> on the activity of this enzyme.

## Conclusions

A dietary Co concentration of 0.21 mg/kg DM provides sufficient Co for the synthesis of vitamin  $B_{12}$  required for the metabolism, performance and diet digestibility of high-yielding dairy cows. Recent restrictions on the inclusion of Co in the diet in the European Union are therefore unlikely to have an impact on intake, performance or health over the periparturient period, and there is little justification for the use of supplementary sources of vitamin  $B_{12}$ .

# Acknowledgements

The authors would like to acknowledge the input of Jess Marshall, Paul Daley, Rosie Barraclough and Marcus Doig for assistance with collecting and analysing the samples, and to AHDB Dairy for funding the study. WADV Weerathilake was supported by a Commonwealth Studentship, and AH Brassington by a scholarship from The Perry Foundation.

# **Declaration of interest**

434 None

### **Ethics Statement**

The procedures involving animals were conducted in accordance with the UK Animals

(Scientific Procedures) Act 1986 (amended 2012), and were approved by the Harper

Adams University Local Ethical Review.

## Software and data repository resources

442 None.

# References

- Akins MS, Bertics SJ, Socha MT and Shaver RD 2013. Effects of cobalt supplementation
- and vitamin B<sub>12</sub> injections on lactation performance and metabolism of Holstein dairy cows.
- 447 Journal of Dairy Science 96, 1755-1768.
- 448 Association of Analytical Chemists (AOAC) 2012. Official methods of analysis, volume 1,
- 449 19th edition, AOAC, Arlington, VA, USA.
- Bobe G, Young JW and Beitz DC 2004. Invited review: pathology, etiology, prevention, and
- treatment of fatty liver in dairy cows. Journal of Dairy Science 87, 3105-3124.
- Castagnino DS, Seck M, Beaudet V, Kammes KL, Voelker Linton JA, Allen MS, Gervais R,
- 453 Chouinard PY and Girard CL 2016. Effects of forage family on apparent ruminal synthesis of
- B vitamins in lactating dairy cows. Journal of Dairy Science 99, 1884-1894.
- 455 Contreras GA, Strieder-Barboza C and Raphael W 2017. Adipose tissue lipolysis and
- 456 remodelling during the transition period of dairy cows. Journal of Animal Science and
- 457 Biotechnology 8, 41.
- Cope CM, Mackenzie AM, Wilde D and Sinclair LA 2009. Effects of level and form of dietary
- 459 zinc on dairy cow performance and health. Journal of Dairy Science 92, 2128-2135.
- 460 Duplessis M, Lapierre H, Pellerin D, Laforest JP and Girard CL 2017. Effects of intramuscular
- injections of folic acid, vitamin B<sub>12</sub>, or both, on lactational performance and energy status of
- 462 multiparous dairy cows. Journal of Dairy Science100, 4051-4064.
- 463 Ferguson JD, Galligan DT, and Thomsen N 1994. Principal descriptors of body condition score
- in Holstein cows. Journal of Dairy Science 77, 2695-2703.
- 465 Girard CL, Santschi DE, Stabler SP and Allen RH 2009. Apparent ruminal synthesis and
- 466 intestinal disappearance of vitamin B<sub>12</sub> and its analogs in dairy cows. Journal of Dairy Science
- 467 92, 4524-4529.

- 468 Girard CL and Matte JJ 2005a. Effects of intramuscular injections of vitamin B<sub>12</sub> on lactation
- 469 performance of dairy cows fed dietary supplements of folic acid and rumen-protected
- 470 methionine. Journal of Dairy Science 88, 671-676.
- 471 Girard Cl and Matte JJ 2005b. Folic acid and vitamin B<sub>12</sub> requirements of dairy cows: A concept
- 472 to be revised. Livestock Production Science 98, 123-133.
- 473 Graulet B, Matte JJ, Desrochers A, Doepel L, Palin MF and Girard CL 2007. Effects of dietary
- supplements of folic acid and vitamin B<sub>12</sub> on metabolism of dairy cows in early lactation.
- 475 Journal of Dairy Science 90, 3442-3455.
- Juchem SO, Robinson PH and Evans E 2012. A fat based rumen protection technology post-
- ruminally delivers a B vitamin complex to impact performance of multiparous Holstein cows.
- 478 Animal Feed Science and Technology 174, 68-78.
- Kadim IT, Johnson EH, Mahgoub O, Srikandakumar A, Al-Ajmi D, Ritchie A, Annamalai K and
- 480 Al-Halhali AS 2003. Effect of low levels of dietary cobalt on apparent nutrient digestibility in
- Omani goats. Animal Feed Science and Technology 109, 209-216.
- 482 Kanakkaparambil R, Singh R, Li D, Webb R and Sinclair KD 2009. B-vitamin and
- 483 homocysteine status determines ovarian response to gonadotrophin treatment in sheep.
- 484 Biology of Reproduction 80, 743-752.
- Karlengen IJ, Harstad OM, Taugbøl O, Berget I, Aastveit AH and Våge DI 2012. The effect
- of excess cobalt on milk fatty acid profiles and transcriptional regulation of SCD, FASN,
- DGAT1 and DGAT2 in the mammary gland of lactating dairy cows. Journal of Animal
- 488 Physiology and Animal Nutrition. 96 1065-1073.
- 489 Kincaid RL and Socha MT 2007. Effect of cobalt supplementation during late gestation and
- 490 early lactation on milk and serum measures. Journal of Dairy Science 90, 1880-1886.

- Lean IJ, Van Saun R and DeGaris PJ 2013. Energy and protein nutrition management of
- transition dairy cows. Veterinary Clinics Food Animal Practice 29, 337–366.
- Leskinen H, Viitala S, Mutikainen M, Kairenius Piia, Tapio I, Taponen J, Bernard L, Vilkki J
- 494 and Shingfield KJ 2016. Ruminal infusions of cobalt EDTA modify milk fatty acid composition
- 495 via decreases in fatty acid desaturation and altered gene expression in the mammary gland
- 496 of lactating cows. Journal of Nutrition 146, 976-985.
- 497 Lopez-Guisa JM and Satter LD 1992. Effect of copper and cobalt addition on digestion and
- 498 growth in heifers fed diets containing alfalfa silage or corn crop residues. Journal of Dairy
- 499 Science 75, 247-256.
- 500 McArt JAA, Nydam DV and Oetzel GR 2012. Epidemiology of subclinical ketosis in early
- lactation dairy cattle. Journal of Dairy Science 95, 5056-5066.
- 502 McArt JAA, Nydam DV, Oetzel GR, Overton TR and Ospina PA 2013. Elevated non-esterified
- fatty acids and ß-hydroxybutyrate and their association with transition dairy cow performance.
- 504 The Veterinary Journal 198, 560-570.
- 505 McDowell LR 2000. Vitamins in animal and human nutrition, 2nd edition. Iowa State University
- 506 Press, Ames, IA, USA.
- 507 National Research Council (NRC) 2001. Nutrient requirements of dairy cattle. 7<sup>th</sup> revised
- 508 edition. National Academy Press.
- Preynat A, Lapierre H, Thivierge MC, Palin MF, Matte JJ Desrochers A and Girard CL 2009.
- 510 Effects of supplements of folic acid, vitamin B<sub>12</sub>, and rumen-protected methionine on whole
- 511 body metabolism of methionine and glucose in lactating cows. Journal of Dairy Science 92,
- 512 677-689.

- 513 Raboisson D, Mounié M and Maigné E 2014. Diseases, reproductive performance, and 514 changes in milk production associated with subclinical ketosis in dairy cows: A meta-analysis 515 and review. Journal of Dairy Science 97, 7547-7563. 516 Shingfield KJ, Arölä A, Ahenjärvi S, Vanhatalo A, Toivonen V, Griinari JM and Huhtanen P 517 2013. Ruminal infusions of cobalt-EDTA reduce mammary  $\Delta 9$ -desturase index and later milk 518 fatty acid composition in lactating dairy cows. The Journal of Nutrition 138, 710-717. 519 Sinclair LA, Edwards R, Errington KA, Holdcroft AM and Wright M 2015. Replacement of 520 grass and maize silages with lucerne silage: effects on performance, milk fatty acid profile 521 and digestibility in Holstein-Friesian dairy cows. Animal 9: 1970-1978. 522 Stangl GJ, Schwarz FJ, Müller H and Kirchgessner M 2000. Evaluation of the cobalt 523 requirement of beef cattle based on vitamin B<sub>12</sub>, folate, homocysteine and methylmalonic 524 acid. British Journal of Nutrition 84, 645-653. 525 Stemme K, Lebzien P, Flachowsky G and Scholz H 2008. The influence of an 526 increased cobalt supply on ruminal parameters and microbial vitamin B<sub>12</sub> synthesis in 527 the rumen of dairy cows. Archives of Animal Nutrition 62, 207-218. 528 Suthar VS, Canelas-Raposo J, Deniz A and Heuwieser W 2013. Prevelance of subclinical 529 ketosis and relationships with postpartum diseases in European dairy cows. Journal of Dairy 530 Science 96, 2925-2938. 531 Tiffany ME, Spears JW, Xi L and Horton J 2003. Influence of supplemental cobalt source 532 and concentration on performance, vitamin B<sub>12</sub> status, and ruminal and plasma metabolites 533 in growing and finishing steers. Journal of Animal Science 81, 3151-3159.
- 535 production and fermentation of mixed ruminal microorganisms grown in continuous culture 536 flow-through fermentors. Journal of Animal Science 84, 635-640.

Tiffany ME, Fellner V and Spears JW 2006. Influence of cobalt concentration on vitamin B<sub>12</sub>

537 Van Keulen J and Young BA 1977. Evaluation of acid-insoluble ash as a natural marker in 538 ruminant digestibility studies. Journal of Animal Science 44, 282-287. 539 Van Soest PJ, Robertson JB and Lewis BA 1991. Methods for dietary fiber, neutral detergent 540 fiber, and nonstarch polysaccharides in relation to animal nutrition. Journal of Dairy Science 541 74, 3583-3597. 542 Wang RL, Hong XH, Zhang YZ, Zhu XP, Narenbatu and Jia ZH 2007. Influence of dietary 543 cobalt on performance, nutrient digestibility and plasma metabolites in lambs. Animal Feed 544 Science and Technology 135, 346-352.

Table 1 Basal dry cow and lactation total mixed ration composition

Tubic I Basar ary sow and last	Dry cow <sup>1</sup>	Lactation
Ingredient, g/kg DM		
Chopped wheat straw	471	
Maize silage	218	389
Lucerne silage	87	111
Wheat	109	79
Molassed sugar beet feed		79
Soy hulls		66
Molasses	3	7
Protected fat		16
Rapeseed meal	31	91
Corn gluten feed	26	76
Hipro soybean meal	19	55
Palm kernel meal	9	25
Feed grade urea	10	1
Dry cow minerals <sup>2</sup>	13	
Lactation minerals <sup>3</sup>		5
Magnesium chloride	4	

<sup>&</sup>lt;sup>1</sup>Water was added at 0.43 kg/kg DM

<sup>&</sup>lt;sup>2</sup>Dry cow minerals g/kg: Ca 20; P 60; Mg 200; Na 5; mg/kg I 400; Se 30; Cu 750; Mn 4 000; Zn 6 000; iu/kg Vit A 800 000; Vit D 200 000; Vit E 10 000; Biotin 75 mg/kg.

<sup>&</sup>lt;sup>3</sup>Lactation minerals g/kg: Ca 180; P 25; Mg 80; Na 120; mg/kg I 400; Se 40; Cu 1 400; Mn 4 000; Zn 6 000; iu/kg Vit A 1 000 000; Vit D 300 000; Vit E 4 500; Biotin 135 mg/kg.

Table 2 Chemical and mineral composition of dry cow and lactation diets that contained no added cobalt (C), added dietary Co (DC), added dietary vitamin

B<sub>12</sub> (DB) or injected vitamin B<sub>12</sub> (IB)

Nutrient	Dry cow <sup>1</sup>	C and IB	DC	DB
DM, g/kg	474	471	466	468
Organic matter, g/kg DM	932	941	939	937
Ash, g/kg DM	67.8	59.2	60.9	63.1
CP, g/kg DM	139	168	169	165
NDF, g/kg DM	561	364	359	386
ADF, g/kg DM	359	230	226	238
Hemicellulose, g/kg DM	202	134	133	148
Fat, g/kg DM	16.7	33.5	34.4	31.7
Fatty acids, g/100 g FA				
C16:0	25.1	25.9	24.1	25.9
C18:0	nd²	2.10	1.99	2.10
C18:2 cis-9, cis-12	28.5	27.5	27.1	27.5
C18:3 cis-9, cis-12, cis-15	3.15	2.69	2.44	2.69
Macro minerals, g/kg DM				
Ca	5.82	7.84	7.71	7.87
Р	2.92	4.03	4.15	4.05
Mg	4.25	2.69	2.71	2.74
Na	1.32	1.66	1.66	1.66
K	12.8	15.7	15.8	15.9
S	1.77	2.50	2.50	2.32
Micro minerals, mg/kg DM				
Со	0.21	0.21	0.36	0.23
Mn	76.0	58.9	57.2	59.4
Fe	279	281	274	285
Zn	102	67.1	64.0	65.6
Мо	1.34	1.16	1.20	1.48

<sup>&</sup>lt;sup>1</sup>Dry cow treatments achieved by individually providing either no added Co (C), 0.2 mg Co/kg DM (DC), 0.68 mg vitamin B<sub>12</sub>/kg DM (DB) or injected vitamin B<sub>12</sub> (IB). <sup>2</sup>nd = not detected

**Table 3** Intake (post calving), milk performance and body condition (mean value pre- and post-calving) of dairy cows offered diets that contained no added cobalt (C), added dietary Co (DC), added dietary vitamin  $B_{12}$  (DB) or injected vitamin  $B_{12}$  (IB)

	Treatment					<i>P</i> -value <sup>1</sup>		
	С	DC	DB	IB	SED	Treatment	Time	
DM intake (kg/day)	21.5	21.3	21.7	21.8	0.78	0.918	<0.001	
Milk yield (kg/day)	38.3	39.5	40.8	39.6	1.58	0.480	<0.001	
Milk fat (g/kg)	40.6	40.8	40.6	39.6	1.92	0.922	<0.001	
Milk protein (g/kg)	32.9	33.2	33.3	33.1	0.67	0.936	<0.001	
Milk lactose (g/kg)	45.0	45.0	45.4	45.0	0.37	0.668	<0.001	
Milk fat (kg/d)	1.54	1.59	1.63	1.55	0.921	0.715	0.035	
Milk protein (kg/d)	1.24	1.28	1.34	1.30	0.046	0.235	<0.001	
Milk lactose (kg/d)	1.72	1.75	1.86	1.79	0.073	0.256	<0.001	
SCC (log <sub>10</sub> /ml) <sup>2</sup>	1.74	1.59	1.67	1.83	0.152	0.452	<0.001	
Body condition	3.00 <sup>ab</sup>	3.08 <sup>ab</sup>	3.14 <sup>b</sup>	2.94ª	0.069	0.042	<0.001	
Live weight (kg)	665	657	683	651	22.0	0.504	<0.001	

<sup>&</sup>lt;sup>1</sup>There were no (*P*>0.05) Treatment x Time interactions

<sup>&</sup>lt;sup>2</sup>Scc = somatic cell count

<sup>&</sup>lt;sup>a,b</sup>Means within a row with a different superscript differ (*P* <0.05).

**Table 4** Mean plasma metabolite and mineral concentration, and hepatic triacylglycerol concentrations in dairy cows offered diets that contained no added cobalt (C), added dietary Co (DC), added dietary vitamin B<sub>12</sub> (DB) or injected vitamin B<sub>12</sub> (IB)

	-	Treatment				<i>P</i> -value <sup>1</sup>	
	С	DC	DB	IB	SED	Treatment	Time
Metabolites <sup>2</sup>							
Glucose (mmol/l)	3.52	3.39	3.48	3.54	0.107	0.480	<0.001
3-OHB (mmol/l)	0.48	0.52	0.51	0.51	0.030	0.598	<0.001
NEFA (mmol/l)	0.40	0.40	0.41	0.44	0.054	0.848	<0.001
Urea (mmol/l)	4.36	4.24	4.29	4.27	0.214	0.953	<0.001
Minerals							
P (mmol/l)	3.01	3.18	3.19	3.19	0.141	0.519	<0.001
Mg (mmol/l)	0.86	0.79	0.90	0.73	0.133	0.582	0.611
Fe (µmol/l)	37.5	35.9	39.7	36.8	2.05	0.294	<0.001
Zn (µmol/l)	12.4 <sup>ab</sup>	13.5 <sup>b</sup>	12.8 <sup>ab</sup>	11.6ª	0.59	0.023	<0.001
Cu (µmol/l)	14.5	15.1	14.9	15.0	0.70	0.863	<0.001
Co (µmol/l²)	0.014	0.018	0.015	0.019		<0.001	0.045
	±0.0022	±0.0022	±0.0022	±0.0022			
Se (µmol/l)	1.04	1.03	1.04	1.04	0.031	0.965	<0.001
Mo (µmol/l)	0.25	0.23	0.24	0.25	0.025	0.817	<0.001
Hepatic triacylglycerol, mg/g fresh <sup>3</sup>							
Pre-partum	5.81	5.85	5.56	7.23		0.616	
	±2.349	±2.349	±2.349	±2.349			
Post-partum	16.1	18.4	22.0	19.3		0.862	
	±2.651	±2.651	±2.651	±2.651			

<sup>&</sup>lt;sup>1</sup>There were no (*P*>0.05) Treatment x Time interactions

<sup>&</sup>lt;sup>2</sup>3-OHB = \(\beta\)-hydroxybutyrate, NEFA = non-esterified fatty acids

<sup>&</sup>lt;sup>3</sup>Data were not normally distributed and were analysed by firstly converting to natural log.

The antiln least square means along with the interval of confidence at 95% are presented.

a,b,c Means within a row with a different superscript differ (P < 0.05).

**Table 5** Diet digestibility (kg/kg) of DM, organic matter (OM), nitrogen (N) and fibre in dairy cows offered diets that contained no added cobalt (C), added dietary Co (DC), added dietary vitamin  $B_{12}$  (DB) or injected vitamin  $B_{12}$  (IB)

Treatment						
	С	DC	DB	IB	SED	<i>P</i> -value
DM	0.71	0.72	0.73	0.73	0.016	0.630
OM	0.73	0.74	0.74	0.75	0.015	0.631
N	0.66	0.67	0.66	0.69	0.019	0.240
NDF	0.54	0.53	0.58	0.56	0.029	0.257
ADF	0.45	0.45	0.50	0.48	0.031	0.234
Hemicellulose <sup>1</sup>	0.68	0.65	0.73	0.70	0.031	0.136

<sup>&</sup>lt;sup>1</sup>Calculated as NDF-ADF

**Table 6** Milk fatty acid profile during week 7 of lactation in dairy cows offered diets that contained no added cobalt (C), added dietary Co (DC), added dietary vitamin  $B_{12}$  (DB)

or injected vitamin B<sub>12</sub> (IB)

Of Injected Vitamin B <sub>12</sub> (IB)	С	DC	DB	IB	SED	<i>P</i> -value
Fatty acid (g/100g)						
C4:0	4.87	4.78	4.94	4.42	0.322	0.401
C6:0	2.77	2.66	2.62	2.67	0.099	0.356
C8:0	1.56	1.33	1.34	1.60	0.124	0.066
C10:0	3.27	3.01	2.64	3.22	0.170	0.164
C12:0	3.68	3.44	3.32	3.62	0.190	0.242
C14:0	10.7	10.4	10.1	10.6	0.353	0.366
C14:1 <i>cis</i> -9	0.87	0.94	0.89	0.90	0.079	0.817
C15:0	0.99 <sup>ab</sup>	0.93ª	0.94 <sup>ab</sup>	1.09 <sup>b</sup>	0.056	0.025
C16:0	27.5	27.6	27.1	27.9	0.671	0.651
C16:1 <i>cis</i> -9	1.42	1.61	1.58	1.50	0.121	0.383
C17:0	0.53	0.55	0.51	0.58	0.028	0.150
C18:0	7.83	7.96	7.57	7.40	0.352	0.393
C18:1 <i>trans</i> 6 to 8	0.40	0.41	0.39	0.38	0.047	0.936
C18:1 trans-9	0.30	0.36	0.35	0.31	0.057	0.692
C18:1 trans-10	0.61	0.54	0.67	0.73	0.104	0.323
C18:1 trans-11	0.61	0.66	0.66	0.91	0.116	0.061
C18:1 trans-12	0.53	0.60	0.66	0.48	0.091	0.222
C18:1 <i>cis</i> -9	18.3	18.8	18.5	17.3	0.926	0.447
C18:2 <i>cis</i> -9, cis-12	2.53	2.41	2.40	2.48	0.124	0.686
C18:2 cis-9, trans-11	0.56	0.62	0.56	0.56	0.069	0.769
C18:2 trans-10, cis-12	0.03	0.05	0.05	0.05	0.008	0.221
C18:3 cis-9, cis-12, cis-15	0.06	0.10	0.10	0.12	0.036	0.383
C20:0	0.07	0.06	0.06	0.07	0.006	0.801
C20:1 <i>cis</i> -9	0.35	0.31	0.30	0.35	0.038	0.428
C20:5 n-3	0.05	0.05	0.06	0.05	0.008	0.647
C22:6 n-3	0.06	0.08	0.08	0.08	0.014	0.259
Other	9.64	9.69	11.37	10.65	1.528	0.625
Summation						
Saturated	64.0	62.9	61.7	63.4	1.37	0.386
Monounsaturated	23.4	24.3	24.0	22.9	1.07	0.579
Polyunsaturated	3.30	3.32	3.30	3.37	0.181	0.984
<16	28.8	27.6	27.2	28.3	0.88	0.284
16:0 and 16:1	29.0	29.4	28.8	29.5	0.70	0.693
>16	32.9	33.6	33.0	31.9	1.33	0.651
Desaturase index						
14:1/(14:0+14:1)	0.07	0.08	0.08	0.08	0.005	0.475
16:1/(16:0+16:1)	0.05	0.06	0.06	0.05	0.004	0.264
18:1/(18:0+18:1)	0.70	0.70	0.71	0.70	0.010	0.861
c-9, t-11 CLA/( 18:1 t-11+	0.50 <sup>b</sup>	0.51 <sup>b</sup>	0.47 <sup>ab</sup>	0.40 <sup>a</sup>	0.033	0.006
c-9, t-11 CLA <sup>1</sup> 1CLA = conjugated lingleic acid						

<sup>&</sup>lt;sup>1</sup>CLA = conjugated linoleic acid. <sup>a,b</sup>Means within a row with a different superscript differ (*P* < 0.05).

**Figure 1** Pre- and post-partum blood concentrations of vitamin B<sub>12</sub> (a) and secondary metabolites, homocysteine (b), methylmalonic acid (c) and succinate (d). Cows offered diets that contained no added cobalt (Co; C: ■), added dietary Co (DC: •), added dietary vitamin B<sub>12</sub> (DB: ○) or injected vitamin B<sub>12</sub> (IB: △). For plasma B<sub>12</sub>, interaction between Treatment and Time: P=0.007; SED = 22.7. For homocysteine, main effect of Time: P<0.001; SED = 0.265. For methylmalonic acid, interaction between Treatment x Time: P=0.033; SED = 0.031. For succinate, main effect of Time: P<0.001; SED = 0.231.

## **Supplementary Material S1**

Added dietary cobalt or vitamin  $B_{12}$ , or injecting vitamin  $B_{12}$  does not improve performance or indicators of ketosis in pre- and post-partum Holstein-Friesian dairy cows

W.A.D.V. Weerathilake<sup>1\*</sup>, A.H. Brassington<sup>2</sup>, S.J. Williams<sup>1</sup>, W.Y. Kwong<sup>2</sup>, L.A. Sinclair<sup>1†</sup> and K.D. Sinclair<sup>2</sup>

### **Animal** Journal

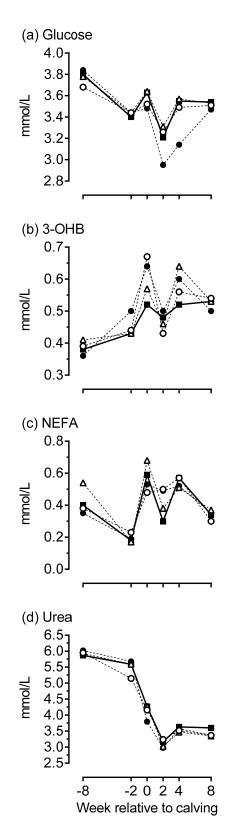
Plasma vitamin  $B_{12}$  analysis was conducted using the ADIVA Centaur CP VB12 assay at the AHVLA laboratory, Shrewsbury, Shropshire, UK. The limit of detection was 33 pmol/l, and coefficients of variation (CVs) for low, medium and high quality controls were 12.2, 12.4 and 12.4% respectively. Plasma homocysteine (Hcy) was determined using an Imola Auto analyser (RX imola; Randox Laboratories Ltd., Antrim, U.K.) using a kit supplied by Randox Laboratories (catalogue no. HY4036). The limit of detection was 1.74  $\mu$ M and CV's for low, medium and high quality controls were 1.03, 0.83 and 0.95% respectively.

Plasma methylmalonic acid (MMA) and succinic acid (SA) concentrations were determined by GC with mass spectroscopic detection (GC-MS) following derivatisation and extraction, modified from the method of Kanakkarambil *et al.* (2009). Briefly, 50 μl of plasma and 5 μl of internal standard (4-chlorobutyric acid (CBA) 250 μM) were added to 250 μl 12% BF<sub>3</sub>-methanol in a 2.5 ml screw capped glass vial, vortexed for 30 s and heated at 95°C for 15 minutes on a heating block. After cooling, 250 μl distilled water and 250 μl dichloromethane (CH<sub>2</sub>Cl<sub>2</sub>) was added to the vial and vortexed

for 30 s. The mixture was then centrifuged for 8 minutes at 2500 g at 4°C to separate the layers. The lower dichloromethane layer was transferred to a screw capped autosampler vial with insert for GC-MS analysis. The method used a DB-WAX (crosslinked polyethylene glycol; J&W Scientific Agilent technology) 30 m long column of 0.25mm i.d and 0.15 µm film thickness. The carrier gas was He and flow rate was 1.0 ml/min. Injection mode was splitless and volume was 1 µl for both SCAN mode, for qualification, and SIM mode, for quantification. Injection port temperature was 260°C. The MS selective detector interference temperature was 280°C. The chromatograph was programmed for an initial temperature of 50°C for 2 minutes, increased to 150°C at 8°C per minute, then increased to 220°C at 100°C per minute and held for 5 minutes at the final temperature. The MS was operated in electron impact (EI) ionization mode with the ionization energy of 70eV. SCAN mode measured at m/z: 20-500 and SIM (selected ion-monitoring) ions were set at 105 (for CBA), 115 (for MMA and SA). The same method was used for standards of MMA and SA at concentrations 0.156, 0.313, 0.625, 1.25, 2.5, 5, 10 and 20 µmol/l. Calibration was carried out by comparison of peak areas of CBA with MMA and SA in the standards and the final results expressed in µmol/l plasma. The limit of detection and limit of quantification was 0.156 µmol/l. The CV's for low, medium and high quality controls were 0.44, 1.03, 0.83% for MMA and 1.2, 1.1 and 2.3% for SA respectively.

For hepatic triacylglycerol (TAG) analyses, samples were reduced to powder under liquid N, and known amounts (around 90-120 mg) weighed and homogenised in 1.6 ml of 0.47 M sodium sulphate, followed by the addition of 2 ml of 0.47 M sodium sulphate and 5.4 ml hexane:isopropanol (3:2, v/v). The mixtures were vortexed for 30 s and centrifuged at 2000 g for 5 minutes at room temperature, then 2.5 ml of the top layer transferred to a glass tube and dried under nitrogen. The dried lipid was

reconstituted in 1 ml of hexane and 10 or 60 µl transferred to a second tube and dried under nitrogen. Dried samples were then re-suspended in 120 µl of isopropanol and 50 µl was mixed with 250 µl of colour reagent from Wako LabAssay triglyceride kit (Catalogue No. 290-63701; Alpha Laboratories, Hampshire, UK) for triacylglycerol measurement. Different concentrations of triolein standard (Catalogue No. T7140, Sigma-Aldrich, Gillingham, UK) were processed in parallel with liver samples to generate a standard curve. Different concentrations of glycerol and non-extracted triolein were included in each plate to monitor enzyme activity and completeness of lipolysis by lipoprotein lipase. In addition, a single liver analysed in each plate served as a quality control. The inter-assay CV was 8.4%.



Supplementary Figure 1 Plasma glucose (a), β-hydroxybutyrate (3-OHB) (b), non esterified fatty acids (NEFA) (c) and urea (d) in pre- and post-partum of dairy cows offered diets that contained no added cobalt (Co) (C: ■), added dietary Co (DC: •), added dietary B12 (DB: ○) or injected B12 (IB: Δ).

