

**HHS PUBLIC ACCESS**

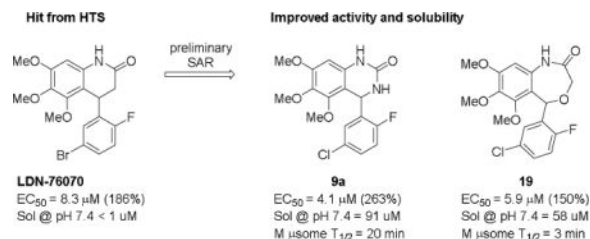
Author manuscript

Bioorg Med Chem Lett. Author manuscript; available in PMC 2018 December 01.

Published in final edited form as:

Bioorg Med Chem Lett. 2017 December 01; 27(23): 5144–5148. doi:10.1016/j.bmcl.2017.10.066.**Optimization of a series of heterocycles as survival motor neuron gene transcription enhancers****Sungwoon Choi^a, Alyssa N. Calder^a, Eliza H. Miller^a, Kierstyn P. Anderson^a, Dawid K. Fiejtek^a, Anne Rietz^b, Hongxia Li^b, Jonathan J. Cherry^b, Kevin M. Quist^b, Xuechao Xing^a, Marcie A. Glicksman^a, Gregory D. Cuny^a, Christian L. Lorson^c, Elliot A. Androphy^b, and Kevin J. Hodgetts^a**^aLaboratory for Drug Discovery in Neurodegeneration, Brigham and Women's Hospital and Harvard Medical School, 65 Landsdowne Street, Cambridge, MA, USA^bDepartment of Dermatology, Indiana University School of Medicine, Indianapolis, IN, USA^cDepartment of Veterinary Pathobiology, Bond Life Sciences Center, University of Missouri, Columbia, Missouri, USA**Abstract**

Spinal muscular atrophy (SMA) is a neurodegenerative disorder that results from mutations in the *SMN1* gene, leading to survival motor neuron (SMN) protein deficiency. One therapeutic strategy for SMA is to identify compounds that enhance the expression of the *SMN2* gene, which normally only is a minor contributor to functional SMN protein production, but which is unaffected in SMA. A recent high-throughput screening campaign identified a 3,4-dihydro-4-phenyl-2(1H)-quinolinone derivative (**2**) that increases the expression of *SMN2* by 2-fold with an EC₅₀ = 8.3 μM. A structure-activity relationship (SAR) study revealed that the array of tolerated substituents, on either the benzo portion of the quinolinone or the 4-phenyl, was very narrow. However, the lactam ring of the quinolinone was more amenable to modifications. For example, the quinazolinone (**9a**) and the benzoxazepin-2(3H)-one (**19**) demonstrated improved potency and efficacy for increase in *SMN2* expression as compared to **2**. 2017 Elsevier Ltd. All rights reserved.

Graphical abstract

Publisher's Disclaimer: This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final citable form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Keywords

Spinal muscular atrophy; Survival motor neuron

Spinal muscular atrophy (SMA) is a neurodegenerative disease characterized by progressive muscle wasting, loss of motor function and premature death in the most severe cases.¹⁻³ Two genes, *SMN1* and *SMN2*, produce survival motor neuron (SMN) protein,⁴⁻⁶ which is ubiquitously expressed with the highest levels in the spinal cord⁷ and functions in the assembly of spliceosomal small nuclear ribonucleoproteins.⁸ The majority of SMN protein normally is produced from the *SMN1* gene, while the almost identical *SMN2* gene only produces approximately 10% of functional protein.^{9,10}

SMA results from mutations within exon 7 of *SMN1*, leading to SMN protein deficiency.¹¹ The clinical severity of this disease is, therefore, indirectly proportional to the copy number of *SMN2* genes, the sole source of protein in these patients.^{12,13} Although the threshold level of SMN protein necessary to maintain healthy motor neurons is not known, it has been estimated that increasing the amount of functional protein by only 2- or 3-fold may be clinically significant.¹⁴ Thus, the *SMN2* gene has become a therapeutic target, in which multiple strategies (*i.e.*, small molecules, antisense oligonucleotide) work to increase the transcription of *SMN2*, increase the inclusion of exon 7 *SMN2*, stabilize the full-length exon-7 included *SMN2* mRNA, or stabilize the SMN protein.¹⁵⁻¹⁷ As SMA is the leading heritable cause of infant mortality, it is critical to build upon the one recently FDA-approved treatment, Nusinersen, an antisense oligonucleotide¹⁸ and to provide a variety of treatment options with different modes of action. Several repurposed drugs, such as riluzole, phenylbutyrate, valproic acid, albuterol and hydroxyurea, have advanced into clinical trials, but none has elicited convincing improvement in muscle function or survival in SMA. Currently, there are several small molecules in Phase I to Phase III clinical trials for SMA, including compounds that increased exon 7 inclusion of SMN2.¹⁹

Previously, we reported an *SMN2*-luciferase reporter assay for identifying compounds that increase SMN expression from the *SMN2* gene and its use in high-throughput screening.²⁰ Using the reporter assay, we discovered two hit compounds, **LDN-75654 (1)** and **LDN-76070 (2)** (Figure 1), that increase expression of *SMN2* by 2-fold, but that have different mechanisms of action than the small molecules already in clinical trials to treat SMA. Compound **1** increases the stability of SMN protein, whereas compound **2** acts in a transcriptional manner.²¹

We recently reported the structure-activity relationship (SAR) of analogs of the 5-isopropylisoxazole-3-carboxamide **1**.²² In this paper, we report preliminary SAR of the 3,4-dihydro-4-phenyl-2(1H)-quinolinone **2** for increasing SMN expression.

The 3,4-dihydro-4-phenyl-2(1H)-quinolinone derivatives were prepared according to the procedure outlined in Scheme 1. A three-component coupling of an aniline (**3**), aromatic aldehyde (**4**) and Meldrum's acid in refluxing ethanol generated the quinolinones **5a-m** in good yield.²³ These compounds were transformed further into the methyl substituted derivatives **6a-b**, which were isolated as mixture of *cis*- and *trans*-isomers.

A series of 3,4-dihydro-quinazolinone derivatives **9a–9l** was prepared according to the procedures outlined in Scheme 2.²⁴ Anilines **3** were treated with potassium isocyanate to give the ureas **8** (*e.g.*, R = H). Alternatively, phenyl isocyanates **7** were treated with primary amines to yield substituted ureas **8** (*e.g.*, R = alkyl). The ureas then were treated with catalytic methane sulfonic acid in refluxing toluene, with azeotropic removal of water, to produce the 3,4-dihydro-quinazolinones **9a–9l**.

The synthesis of the 2-quinazolinone **13** and the seven-membered 1,4-benzodiazepin-2-one **14** is outlined in Scheme 3. The pivalate-protected 3,4,5-trimethoxyaniline **10** was lithiated with butyllithium in THF at -78°C , and the resulting anion was quenched with methyl 5-chloro-2-fluorobenzoate to give the ketone **11** in modest yield. The pivalate group was cleaved with refluxing sulfuric acid and gave the versatile intermediate **12**. For example, the ketone **12** was treated with trichloroacetyl chloride, and the product, upon treatment with ammonium acetate, cyclized to **13**. On the other hand, the ketone **12** also was treated with glycine ethyl ester, with azeotropic removal of water, and gave the 1,3-dihydro-5-phenyl-2H-1,4-benzodiazepin-2-one **14**.

Several oxygen-containing heterocycles (*e.g.*, **16**, **19–21**) also were prepared from the ketone **12** (Scheme 4 and Scheme 5). Ketone **12** was reduced to the racemic alcohol **15** with sodium borohydride in ethanol. Treatment of **15** with carbonyl diimidazole (CDI) directly gave the oxazinone **16** in good yield. Acylation of **15** with 2-bromoacetyl chloride or 2-bromopropionyl chloride gave the amides **17** and **18**, respectively. Treatment of **17** with two equivalents of sodium hydride gave the 7-membered benzoxazepin-2(3H)-one **19** in moderate yield.²⁵ Similarly, cyclization of **18** gave the 3-methyl-benzoxazepin-2(3H)-one **20** as a mixture of diastereoisomers. Finally, treatment of **19** with methyl iodide gave the *N*-methyl amide **21**.

The synthesis of the 5-methyl substituted benzoxazepinone **23** is outlined in Scheme 5. Treatment of the ketone **12** with methyl magnesium bromide gave the racemic tertiary alcohol **22** in good yield. Reaction of **22** with 2-bromoacetyl chloride and cyclization gave **23**.

Compounds were assessed for activity in the luciferase reporter assay, as previously described, at various concentrations (0.1 – 50 μM), and were compared to DMSO (0.1%) controls.¹⁹ Dose-response curves were generated to determine EC_{50} values and percent maximum increase in *SMN2* expression. Unless otherwise stated, all values are the mean \pm S.E.M. of 3 separate experiments. First, the importance of the tri-methoxy group on the aryl ring and methyl substitution at each of the 1- and 3-positions of the dihydro-quinolinone was investigated (Figure 2). Removal of either the 7- or 6-methoxy group (*e.g.*, compounds **5a** and **5b**) resulted in the complete loss of activity. With this information, the methoxy groups were determined to be necessary for activity, and the tri-methoxy group was retained in the remaining analogs described in this optimization study. Similarly, the 1- and 3-methyl analogs (*e.g.*, **6a** and **6b**) were inactive, and these two positions were left unsubstituted in subsequent analogs.

We next investigated the effects on *SMN2* expression of substituents on the 4-aryl ring (Table 1). Compounds were prepared following the cyclization procedure outlined in Scheme 1. Removal of either the 5-bromo (*e.g.*, **5c**) or the 2-fluoro (*e.g.*, **5d**) led to a complete loss in activity, indicating the importance of a substituent at both positions. Therefore, we retained the 2-fluoro substituent and varied the 5-substituent. Replacing the 5-bromo by 5-chloro gave an analog (**5e**) of similar activity to **2**, but other electron withdrawing groups were not active (*e.g.*, **5f**, **5g**, **5h** and **5i**). Several other halogenated analogs also were found to be inactive (*e.g.*, **5j**, **5k**, **5l** and **5m**).

After the disappointing finding that limited substitution was tolerated on the 4-aryl ring, we chose to explore other cyclic cores. The goal was to identify a more potent core and then to revisit the SAR on the 4-aryl ring. Although the 5-bromo-2-fluoro aryl analog **2** was the most active compound, for chemical stability reasons, we chose to retain the 5-chloro-2-fluoro aryl ring at the benzylic position of the new cores (Table 2). The 3,4-dihydroquinazolinone derivative **9a** had very promising activity, although the *N*-methyl (**9b**) was less active, and both the *N*-propyl (**9c**) and the 2-quinazolinone (**13**) were inactive. The seven-membered benzodiazepin-2-one **14** retained potency similar to the lead **2**, although its efficacy was reduced. Several of the oxygen containing heterocycles had interesting activity.

The benzoxazin-2-one **16** had encouraging activity, and the 7-membered benzoxazepin-2(3H)-ones, **19** and **20**, had similar activities as **9a**. Interestingly, the 3-methyl analog **20** was tested as a mixture of diastereoisomers, and it is likely that one isomer will retain all activity. *N*-Methylation of the amide (**21**) was not tolerated, and introduction of a methyl group at the benzylic position (*e.g.*, **23**) also was not tolerated.

Given the promising activity of the 3,4-dihydroquinazolinone core (*e.g.*, **9a**), we evaluated the SAR for aryl substitution with this heterocyclic core (Table 3). Removal of either the 5-chloro (*e.g.*, **9b**) or the 2-fluoro (*e.g.*, **9c**) led to a partial, but not a complete, loss in activity. Other mono-halogenated analogs retained some activity (*e.g.*, **9d**, **9e** and **9g**), although the methoxy analogs (**9h–9j**) were inactive. In the next set of analogs, the 2-fluoro substituent was retained, and the 5-substituent was varied. The 5-methyl analog **9k** had similar potency, and the 5-fluoro **9n** was less active. The other 5-substituted analogs (**9l**, **9m** and **9o**) were inactive but moving the 5-chloro to the 4-position (**9p**) was tolerated. Although the 2-methoxy analog (**9h**) activity was restored on incorporation of a 4-chloro (**9q**) or 5-chloro (**9r**).

In the preliminary SAR of the quinazolinone series, we did not establish the importance of the 5-methoxy group on SMN enhancing activity (*e.g.*, deletion of the 5-OMe). We did, however, establish the importance of a methoxy group at the analogous position in the benzoxazepin-2(3H)-one series. The synthesis of **26** is outlined in Scheme 6. Palladium-catalyzed addition of the commercially available 5-chloro-2-fluorophenylboronic acid and 2-amino-4,5-dimethoxybenzotrile **24** gave the corresponding ketone, which then was reduced with sodium borohydride in ethanol to give the racemic alcohol **25**. Reaction of **25** with 2-bromoacetyl chloride, and cyclization as described previously, gave the 7,8-dimethoxy analog **26**. Compound **26** was determined to be inactive in the luciferase reporter

assay. Combined, the lack of activity for **5a**, **5b**, and **26** establishes the importance of all three methoxy groups for activity.

Two promising analogs, the quinazolinone (**9a**) and the benzoxazepin-2(3H)-one (**19**), were evaluated for aqueous solubility and stability in mouse microsomes (Figure 3). Both **9a** and **19** had improved aqueous solubility, **9a** had promising stability in mouse microsomes although **19** was poor.

Compound **9a** was further characterized in secondary assays. We observed a ~2-fold increase in total SMN protein as well as a 2-fold increase in the number of gems in SMA derived fibroblasts following treatment with **9a**. Consistent with the earlier discovery that compound **2** acts in a transcriptional manner,²¹ compound **9a** increased SMN-luciferase protein expression from both the SMN1 and SMN2-luciferase constructs. qRT-PCR analysis revealed that compound **9a** increased the amount of total SMN-luciferase reporter transcripts with a corresponding increase in the amount of exon 7 included transcripts. This effect required 24 hours for full activation and is consistent with the hypothesis that this series is regulating SMN2 expression at the transcriptional level.

Compound **9a** was next tested in a preliminary study in SMN^{-/-} mice. Animals were injected with 20 mg/kg of compound **9a** in DMSO by intraperitoneal injection once daily starting on PND 2. Compound **9a** gave a significant increase in lifespan (*e.g.*, 15-day median survival) compared to vehicle treated animals (*e.g.*, 6-day median survival). Compound **9a** treated animals also displayed an increase in the ability to right themselves when compared to vehicle treated animals.

Racemic **9a** was next separated by supercritical fluid chromatography, using a ChiralPak IC column, into its two individual enantiomers in > 99% enantiomeric excess. Testing of the individual enantiomers revealed that the fastest eluting enantiomer was inactive, whereas the second enantiomer retained all activity in the SMN2 luciferase reporter assay (EC₅₀ = 1.3 μM, 185%).

In summary, we established preliminary SAR of the quinazolinone series and discovered that most changes to the aryl ring substituents led to a loss in activity. We discovered several alternative cores (*e.g.*, quinazolinone the benzoxazepin-2(3H)-one) that had similar or better activity than **2** and the SAR of the aryl ring was expanded. Compound **9a** was identified as a more active analog with improved solubility and stability in mouse microsomes. In an efficacy study in SMN^{-/-} mice, a 2-fold extension in survival was observed following daily treatment with **9a**. The synthesis, determination of absolute stereochemistry and biological evaluation of single enantiomers of **9a** and related analogs is in progress and will be reported in due course.

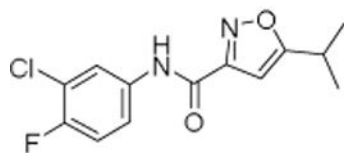
Acknowledgments

The authors thank the NIH (R01 HD064850, R21 NS064349, R21 HD057402), FightSMA, CureSMA, and Gwendolyn Strong Foundation for grant support.

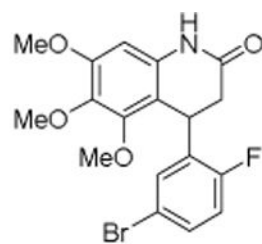
References and Notes

1. Oskoui, M., Darras, BT., De Vivo, DC. Spinal muscular atrophy: 125 years later and on the verge of a cure. Academic Press; San Diego: p. 2017
2. Pearn J. *Lancet*. 1980; 1(8174):919. [PubMed: 6103267]
3. Ahmad S, Bhatia K, Kannan A, Gangwani L. *J Exp Neuroscience*. 2016; 10:39.
4. Lefebvre S, Bürglen L, Reboullet S, Clermont O, Burlet P, Viollet L, Benichou B, Cruaud C, Millasseau P, Zeviani M, Le Paslier D, Frézal J, Cohen D, Weissenbach, Munnich A, Melki J. *Cell*. 1995; 80:155. [PubMed: 7813012]
5. Echaniz-Laguna A, Miniou P, Bartholdi D, Melki J. *Am J Hum Genet*. 1999; 64:1365. [PubMed: 10205267]
6. Coovert DD, Le TT, McAndrew PE, Strasswimmer J, Crawford TO, Mendell JR, Couslon SE, Androphy EJ, Prior TW, Burghes AHM. *Hum Mol Genet*. 1997; 6:1205. [PubMed: 9259265]
7. Boda B, Mas C, Giudicelli C, Nepote V, Guimiot F, Levacher B, Zvara A, Santha M, LeGall I, Simonneau M. *Eur J Hum Genet*. 2004; 12:729. [PubMed: 15162126]
8. Gubitza AK, Feng W, Dreyfuss G. *Exp Cell Res*. 2004; 296:51. [PubMed: 15120993]
9. Lorson CL, Hahnen E, Androphy EJ, Wirth B. *Proc Natl Acad Sci USA*. 1999; 96:6307. [PubMed: 10339583]
10. Monani UR, Lorson CL, Parsons DW, Prior TW, Androphy EJ, Burghes AHM, McPherson JD. *Hum Mol Genet*. 1999; 8:1177. [PubMed: 10369862]
11. Wirth B. *Hum Mut*. 2000; 15:228. [PubMed: 10679938]
12. Gavrilov DK, Shi XY, Das K, Gilliam TC, Wang CH. *Nat Genet*. 1998; 20:230. [PubMed: 9806538]
13. Harada Y, Sutomo R, Sadewa AH, Akutsu T, Takeshima Y, Wada H, Matsuo M, Nishio H. *J Neurol*. 2002; 249:1211. [PubMed: 12242541]
14. Meyer K, Marquis J, Trub J, Nlend Nlend R, Verp S, Reupp MD, Imboden H, Barde I, Trono D, Schumperli D. *Hum Mol Genet*. 2009; 18:546. [PubMed: 19010792]
15. Sunshine, SS., Jarecki, J., MacKenzie, A., Chen, KS. Spinal muscular atrophy therapeutics development. Academic Press; San Diego: 2017.
16. Calder AN, Androphy EJ, Hodgetts KJ. *Small Molecules in Development for the Treatment of Spinal Muscular Atrophy*. *J Med Chem*. 2016; 59:10067. [PubMed: 27490705]
17. Cherry, JJ., Calder, AN., Hodgetts, KJ., Androphy, EJ. *Small molecule approaches to upregulate SMN expression from the SMN 2 locus*. Academic Press; San Diego: p. 2017
18. Ottesen EW. *Translational Neuroscience*. 2017; 8:1. [PubMed: 28400976]
19. (a) Ratni H, Karp GM, Weetall M, Naryshkin NA, Paushkin SV, Chen KS, McCarthy KD, Qi H, Turpoff A, Woll MG, Zhang X, Zhang N, Yang T, Dakka A, Vazirani P, Zhao X, Pinard E, Green L, David-Pierson P, Tuerck D, Poirier A, Muster W, Kirchner S, Mueller L, Gerlach I, Metzger F. Specific correction of alternative survival motor neuron 2 splicing by small molecules: Discovery of a potential novel medicine to treat spinal muscular atrophy. *J Med Chem*. 2016; 59:6086–6100. [PubMed: 27299419] (b) Palacino J, Swalley SE, Song C, Cheung AK, Shu L, Zhang X, Van Hoosear M, Shin Y, Chin DN, Keller CG, Beibel M, Renaud NA, Smith TM, Salcius M, Shi X, Hild M, Servais R, Jain M, Deng L, Bullock C, McLellan M, Schuierer S, Murphy L, Blommers MJ, Blaustein C, Berenshteyn F, Lacoste A, Thomas JR, Roma G, Michaud GA, Tseng BS, Porter JA, Myer VE, Tallarico JA, Hamann LG, Curtis D, Fishman MC, Dietrich WF, Dales NA, Sivasankaran R. SMN2 splice modulators enhance U1-pre-mRNA association and rescue SMA mice. *Nat Chem Biol*. 2015; 11:511–517. [PubMed: 26030728]
20. (a) Cherry JJ, Evans MC, Ni J, Cuny GD, Glicksman MA, Androphy EJ. *J Biomol Screen*. 2012; 17:481. [PubMed: 22233647] (b) Cherry JJ, Androphy EJ. *Future Med Chem*. 2012; 4:1733. [PubMed: 22924510] (c) Xiao J, Marugan JJ, Zheng W, Titus SA, Southall N, Cherry JJ, Evans M, Androphy EJ, Austin CP. *J Med Chem*. 2011; 54:6215. [PubMed: 21819082]
21. Cherry JJ, Osman EY, Evans MC, Choi S, Xing X, Cuny GD, Glicksman MA, Lorson CL, Androphy E. *J EMBO Mol Med*. 2013; 5:1035–50.

22. Rietz A, Li H, Quist KM, Cherry JJ, Lorson CL, Burnett BG, Kern NL, Calder AN, Fritsche M, Lusic H, Boaler PJ, Choi S, Xing X, Glicksman MA, Cuny GD, Androphy EJ, Hodgetts KJ. Discovery of a Small Molecule Probe That Post-Translationally Stabilizes the Survival Motor Neuron Protein for the Treatment of Spinal Muscular Atrophy. *J Med Chem.* 2017; 60:4594–4610. [PubMed: 28481536]
23. Wang X-S, Zhang M-M, Zeng Z-S, Shi D-Q, Tu S-J, Wei X-Y, Zong Z-M. *Tetrahedron Lett.* 2005; 46:7169.
24. Lin AJ, Pardini RS, Lillis BJ, Sartorelli AC. *J Med Chem.* 1974; 17:668.
25. Houlihan WJ, Cooke G, Denzer M, Nicoletti J. *J Heterocycl Chem.* 1982; 19:1453.



LDN-75654 (1)
EC₅₀ = 1.8 μM (242%)



LDN-76070 (2)
EC₅₀ = 8.3 μM (186%)

Figure 1.
Two SMN mRNA expression enhancers identified following high-throughput screening.

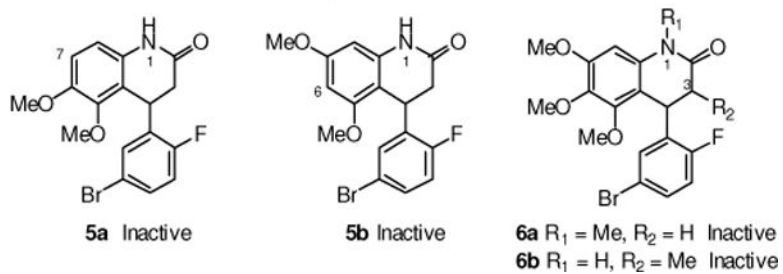


Figure 2. Effects of deletion of individual methoxy substituents and methyl substitution to the dihydro-quinolinone core.

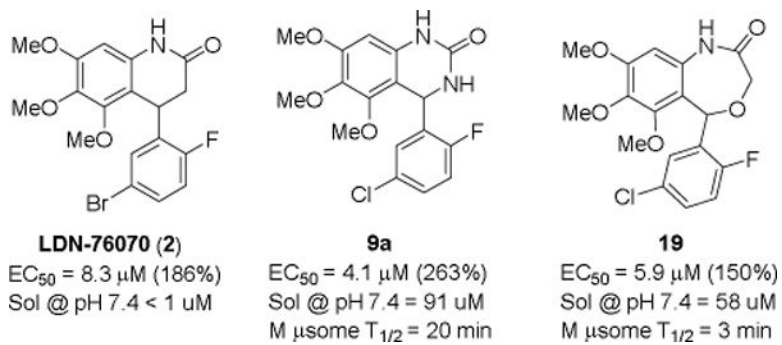
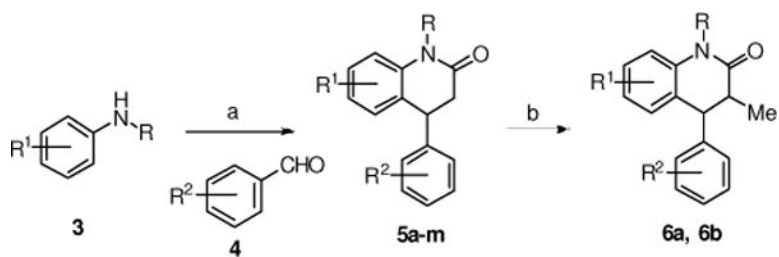
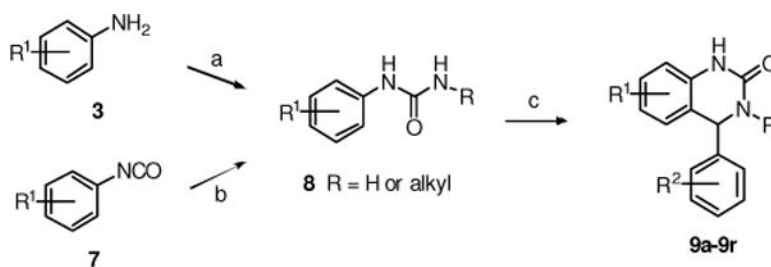


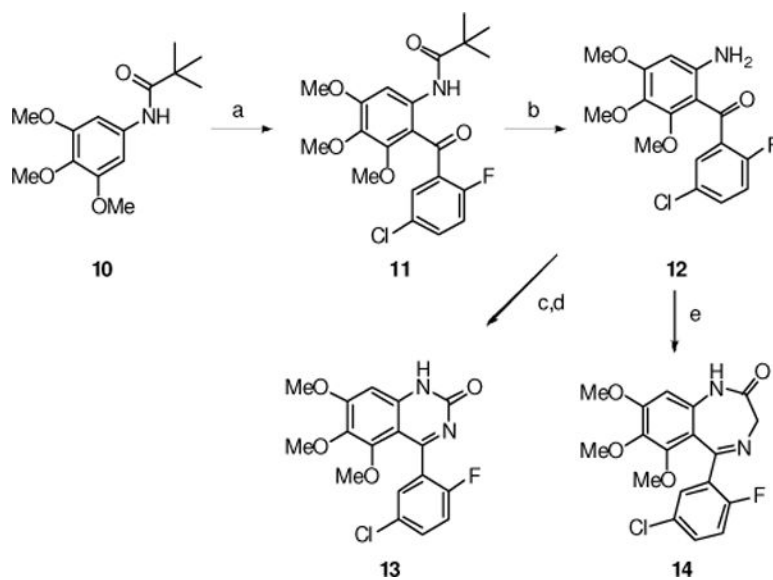
Figure 3.
Solubility and stability of **9a** and **19**.

**Scheme 1.**

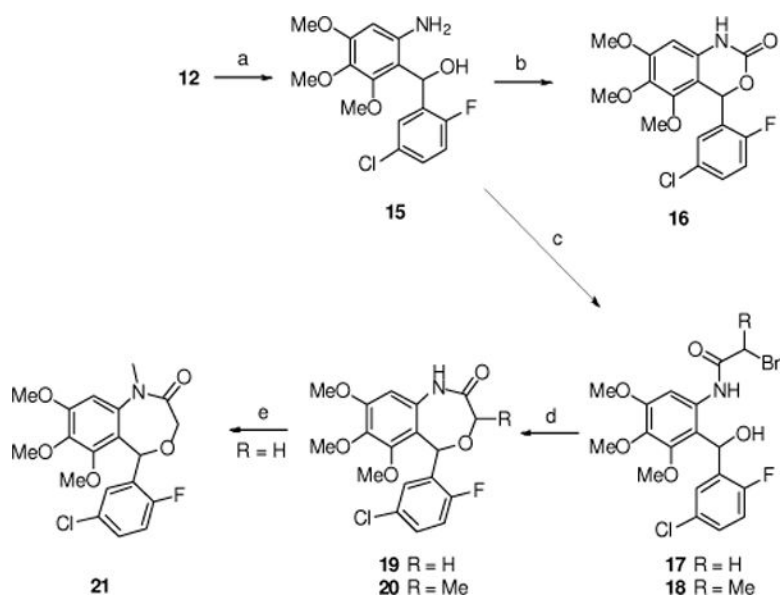
(a) Meldrum's acid, EtOH, 85°C, 16 h (55–78%); (b) BuLi, THF, 0°C then MeI (48%).

**Scheme 2.**

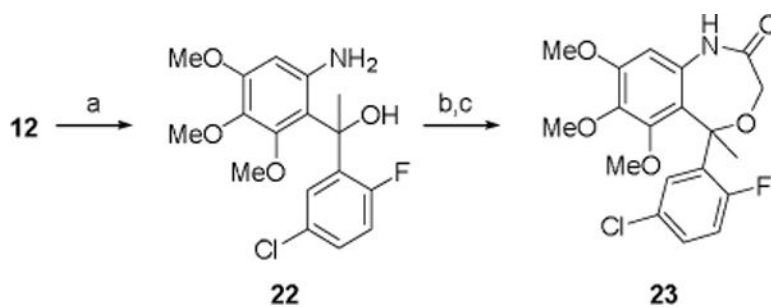
(a) KNCO, AcOH, RT; (84–97%); (b) RNH₂, THF, 0 °C to RT (63–89%); (c) R²PhCHO, cat. MeSO₃H, toluene, reflux (31–81%).

**Scheme 3.**

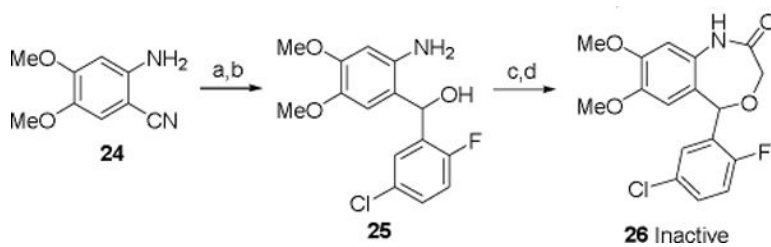
(a) BuLi, THF -78°C then methyl 5-chloro-2-fluorobenzoate (15%); (b) H_2SO_4 , EtOH, reflux (57%); (c) Cl_3CCOCl , DCM, Et_3N , RT; (d) NH_4OAc , DMSO, 80°C (41% over 2-steps); (e) $\text{HCl}\cdot\text{NH}_2\text{CH}_2\text{CO}_2\text{Et}$, Py, Dean-Stark trap, reflux, 24 h (38%).

**Scheme 4.**

(a) NaBH_4 , EtOH, RT (82%); (b) CDI, THF, RT (76%); (c) BrCHRC(O)Cl , Et_2O , Et_3N , 0°C to RT (62–73%); (d) NaH, THF, RT then reflux (43–72%); (e) MeI, Cs_2CO_3 , THF, RT (81%).

**Scheme 5.**

(a) MeMgBr, THF, 0°C (67%); (b) BrCH₂C(O)Cl, Et₂O, Et₃N, 0°C to RT; (c) NaH, THF, RT then reflux (54% over 2-steps).

**Scheme 6.**

(a) 2-F,5-Cl-PhB(OH)₂, Pd(OAc)₂, DMSO, 24 h, 90°C (79%); (b) NaBH₄, EtOH, RT (77%);
(c) BrCH₂C(O)Cl, Et₂O, Et₃N, 0°C to RT (67%); (d) NaH, THF, RT to reflux (71%).

Table 1

Effects of substitution on the aryl ring of the 3,4-dihydro-4-aryl-2(1H)-quinolinone core.

#	Ar	EC50 (μM)	#	Ar	EC50 (μM)
2	2-F,5-Br	8.3 (186%)	5c	2-F	IA
5d	3-Br	IA	5e	2-F,5-Cl	12 (150%)
5f	2-F,5-F	IA	5g	2-F,5-CN	IA
5h	2-F,5-NO ₂	IA	5i	2-F,5-CONH ₂	IA
5j	2-Cl,3-Cl	IA	5k	2-Cl,4-Cl	IA
5l	3-F,4-F	IA	5m	3-Cl,4-Cl	IA

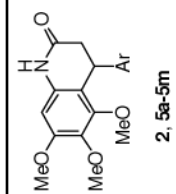


Table 2

Effects of the heterocyclic core (Ar = 2-F,5-Cl-phenyl)

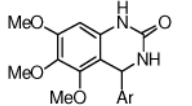
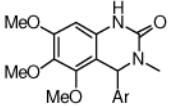
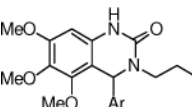
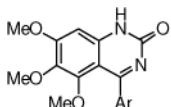
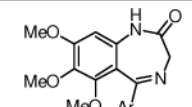
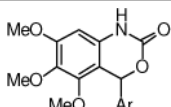
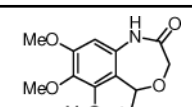
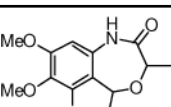
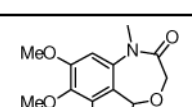
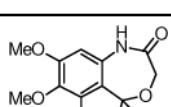
#	Compound	#	Compound
9a	 $EC_{50} = 4.1 \mu\text{M}$ (263%)	9b	 $EC_{50} = 6 \mu\text{M}$ (180%)
9c	 Inactive	13	 Inactive
14	 $EC_{50} = 8.4$ (125%)	16	 $EC_{50} = ?$ (125%)
19	 $EC_{50} = 5.9$ (150%)	20	 $EC_{50} = 2.6$ (125%)
21	 Inactive	23	 Inactive

Table 3
Effects of substitution on the aryl ring of the 3,4-dihydro-4-aryl-quinazolin-2(1*H*)-one core.

#	Ar	EC ₅₀ (μM)	#	Ar	EC ₅₀ (μM)
9a	2-F,5-Cl	4.1 (263%)	9b	2-F	10 (100%)
9c	3-Cl	5.2 (100%)	9d	3-F	25 (150%)
9e	4-F	14.0 (150%)	9f	2-Cl	IA
9g	4-Cl	6.6 (150%)	9h	2-OMe	IA
9i	3-OMe	18 (100%)	9j	4-OMe	IA
9k	2-F,5-Me	4.3 (125%)	9l	2-F,5-Ph	IA
9m	2-F,5-CN	IA	9n	2-F,5-F	17 (100%)
9o	2-F,5-OMe	IA	9p	2-F,4-Cl	6.3 (150%)
9q	2-OMe,4-Cl	7.8 (200%)	9r	2-OMe,5-Cl	3.6 (225%)

