

RESEARCH ARTICLE

ATP/P2X7 receptor signaling as a potential anti-inflammatory target of natural polyphenols

Erika Nuka¹, Kohta Ohnishi¹, Junji Terao^{1,2}, Yoshichika Kawai^{1*}

1 Department of Food Science, Graduate School of Biomedical Sciences, Tokushima University, Tokushima, Japan, **2** Faculty of Nursing and Rehabilitation, Konan Women's University, Hyogo, Japan

* kawai.yoshichika@tokushima-u.ac.jp



Abstract

Innate immune cells, such as macrophages, respond to pathogen-associated molecular patterns, such as a lipopolysaccharide (LPS), to secrete various inflammatory mediators. Recent studies have suggested that damage-associated molecular patterns (DAMPs), released extracellularly from damaged or immune cells, also play a role in the activation of inflammatory responses. In this study, to prevent excess inflammation, we focused on DAMPs-mediated signaling that promotes LPS-stimulated inflammatory responses, especially adenosine 5'-triphosphate (ATP)-triggered signaling through the ionotropic purinergic receptor 7 (P2X7R), as a potential new anti-inflammatory target of natural polyphenols. We focused on the phenomenon that ATP accelerates the production of inflammatory mediators, such as nitric oxide, in LPS-stimulated J774.1 mouse macrophages. Using an siRNA-mediated knockdown and specific antagonist, it was found that the ATP-induced enhanced inflammatory responses were mediated through P2X7R. We then screened 42 polyphenols for inhibiting the ATP/P2X7R-induced calcium influx, and found that several polyphenols exhibited significant inhibitory effects. Especially, a flavonoid baicalein significantly inhibited ATP-induced inflammation, including interleukin-1 β secretion, through inhibition of the ATP/P2X7R signaling. These findings suggest that ATP/P2X7R signaling plays an important role in excess inflammatory responses and could be a potential anti-inflammatory target of natural polyphenolic compounds.

OPEN ACCESS

Citation: Nuka E, Ohnishi K, Terao J, Kawai Y (2018) ATP/P2X7 receptor signaling as a potential anti-inflammatory target of natural polyphenols. PLoS ONE 13(9): e0204229. <https://doi.org/10.1371/journal.pone.0204229>

Editor: Martin E. Rottenberg, Karolinska Institutet, SWEDEN

Received: December 17, 2017

Accepted: September 5, 2018

Published: September 24, 2018

Copyright: © 2018 Nuka et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by JSPS KAKENHI Grant Number 26292069, 15K14730.

Competing interests: The authors have declared that no competing interests exist.

Introduction

Inflammation is an essential immune response that enables survival during infection or injury and maintains tissue homeostasis. Under noxious or abnormal conditions, several inflammatory mediators, including cytokines, chemokines, reactive gases, and eicosanoids, produced by immune cells attract other leukocytes to the injured and/or infected tissues to exclude foreign substances through phagocytosis, killing effects, and cell death induction [1–3]. Therefore, inflammation is important for host defense or tissue remodeling. However, abnormal inflammation with over or persistent expression of mediators could lead to the initiation and

progression of various diseases, such as type 2 diabetes [4], cardiovascular diseases [5–7], neurodegenerative diseases [8,9], and cancers [10]. Inflammation is triggered not only by pathogen-associated molecular pattern molecules (PAMPs) derived from microorganisms, such as a lipopolysaccharide (LPS), but also by damage-associated molecular patterns (DAMPs), either actively or passively released from damaged or immune cells, such as uric acid crystals, nucleotides, and the chromatin-associated protein HMGB1. Recent studies have suggested that DAMPs could be implicated in excessive inflammation which plays a key role in the pathogenesis of several diseases, and therefore targeting DAMPs or their receptors might be beneficial [11,12]. In addition, Yang *et al.* reported the importance of therapeutic approaches to selectively target DAMPs-mediated inflammation while preserving physiological protective immune responses, such as PAMPs-elicited inflammatory responses [13].

Adenosine-5'-triphosphate (ATP) is present at high levels (5–8 mM) in the cytoplasm for cellular homeostasis [14,15], and is released through multiple pathways, including the membrane degradation of necrotic cells, connexin hemichannels from immune cells, pannexin hemichannels from apoptotic cells [16], lysosomal exocytosis from dying cells depending on autophagy [17], and vesicular exocytosis from epithelial cells and osteoblasts and nerves [14]. It is suggested that released extracellular ATP has important and diverse effects on many biological processes including inflammasome formation, promoting cytokine release, pain, defective phagocytic clearance, smooth muscle contraction, and neurotransmission [18–20]. These effects are mediated by plasma membrane purinergic receptors comprised of ATP-gated ionotropic P2Xs and guanine nucleotide-binding protein-coupled P2Ys. On the other hand, many researchers have pointed out that extracellular ATP, released from injured or damaged cells under abnormal conditions, causes undesirable responses to the surrounding cells. For example, the ATP levels of bronchoalveolar lavage fluid are increased by smoking, and the extracellular ATP contributes to the pathogenesis of chronic obstructive pulmonary disease [21], lung fibrosis [22], and allergic asthma [23]. Studies using knockout or antagonists of the ionotropic purinergic receptor 7 (P2X7R), preferentially expressed in immune cells, suggested that P2X7R activation by extracellular ATP is linked to colitis [24], systemic lupus erythematosus and rheumatoid arthritis [25], coronary artery diseases [26], short survival during infection [27], and the progression of brain tumors [28]. Recent findings also suggest that P2X7R-mediated ATP signaling may represent a potential therapeutic target to improve health and that natural products or synthetic compounds inhibiting P2X7R could be useful in clinical trials [29,30].

Many reports have suggested that natural polyphenols, present in vegetables or fruits, have potential health-beneficial effects including anti-inflammatory effects through a variety of biological activities, such as antioxidant and radical scavenging activities, regulation of intracellular signaling cascades or transcriptional networks, and improvements of the endothelial structure or function [31,32]. It has been reported that natural polyphenols down-regulate PAMPs-induced inflammatory mediators, such as LPS-induced pro-inflammatory cytokines, both *in vitro* and *in vivo* [32,33]. Toll-like receptor 4 (TLR4) and other pathogens recognition receptors-induced mitogen-activated protein kinases (MAPKs) and nuclear factor κ B (NF- κ B) pathways were shown to be important targets for polyphenols, such as quercetin [34], epigallocatechin gallate (EGCG) [35], luteolin [36], apigenin [37], and others [38,39]. However, it still remains unknown whether DAMPs-mediated excess inflammatory responses could be the target of natural compounds including polyphenols.

In this study, to develop a new strategy for the prevention of inflammatory diseases, we investigated the molecular actions of extracellular ATP on the LPS-induced macrophage inflammation and carried out screening of natural polyphenols for the inhibition of ATP

signaling. We propose that P2X7R-mediated ATP signaling could be a potential anti-inflammatory target of natural polyphenols.

Materials and methods

Reagents

Dulbecco's modified eagle medium (DMEM, 4.5 g/l glucose, liquid), penicillin-streptomycin mixed solution (Stabilized), ATP disodium salt hydrate from yeast, adenosine-5'-diphosphate (ADP) sodium salt from a bacterial source, adenosine-5'-monophosphate (AMP) sodium salt from yeast, apyrase from potato, protease inhibitor cocktail, phosphatase inhibitor cocktail, bovine serum albumin (BSA, F-V), and 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyltetrazolium bromide (MTT) were obtained from Nacalai Tesque, Inc. (Kyoto, Japan). Fetal bovine serum (FBS) and LPS from *Escherichia coli* O127:B8 were obtained from the Sigma-Aldrich (St. Louis, USA). DMEM without phenol red (4.5 g/l glucose, liquid), sulfanilamide, *N*-(1-naphthyl)ethylenediamine dihydrochloride, sodium nitrite, and rabbit IgG were obtained from Wako Pure Chemical Industries (Osaka, Japan). Rabbit polyclonal antibody to inducible nitric oxide synthase (iNOS) (sc-650) and mouse monoclonal antibody to β -actin (sc-47778) were obtained from Santa Cruz Biotechnology, Inc. (Texas, USA). Rabbit polyclonal antibody to P2X7R was obtained from GeneTex, Inc. (LA, USA). Rabbit monoclonal antibodies to phospho-signal transducer and activator of transcription (STAT) 1 (Tyr701) (58D6), phospho-interferon regulatory factor (IRF)-3 (Ser396) (4D4G), phospho-p38 MAPK (Thr180/Tyr182) (D3F9) XP, phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (D13.14.4E) XP, rabbit polyclonal antibody to phospho-stress-activated protein kinases (SAPKs)/c-jun N-terminal kinases (JNKs) (Thr183/Tyr185), and mouse monoclonal antibody to inhibitor κ B (I κ B)- α were obtained from Cell Signaling Technology, Inc. (MA, USA). Rabbit polyclonal antibody to mouse interferon (IFN)- β for neutralization was obtained from Pestka Biomedical Laboratories, Inc. (NJ, USA). BAPTA-AM was obtained from Invitrogen (CA, USA). Fluo 4-AM was obtained from Dojindo Laboratories (Kumamoto, Japan). 4-(*N,N*-Dipropylsulfamoyl) benzoic acid (probenecid) was obtained from Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan). Pluronic F-127 was obtained from Life Technologies Japan, Ltd. (Tokyo, Japan). All other chemicals were commercially available analytical grade reagents. TAK-242 was obtained from Merck.

Cell culture

Murine macrophage-like J774.1 cells were obtained from American Tissue Culture Collection, and cultured in DMEM, which contains glucose (4.5 mg/ml) and L-glutamine (584 μ g/ml), supplemented with 10% heat-inactivated FBS, penicillin (100 units/ml), and streptomycin (100 μ g/ml) at 37°C under 5% CO₂.

Transfection of siRNA

Stealth RNAi siRNA against P2X7R (P2rx7 MSS276199) and a negative control (low GC) were obtained from Invitrogen (CA, USA). The transfection of siRNA was performed using the Neon Transfection System (Life Technologies Japan, Ltd., Tokyo, Japan). J774.1 cells (2×10^6 cells) were resuspended in 100 μ l resuspension buffer R containing 50 nM siRNA against P2X7R or control siRNA and transfected in 100 μ l Neon tip using two pulses (1720 V input pulse voltage, 10 ms input pulse width). The transfected cells were seeded into 24-well plates (5×10^5 cells/well) and cultured in antibiotic-free DMEM for 24–36 h, then used for each experiment.

Measurement of nitric oxide production

Nitric oxide (NO) production was evaluated by measuring the metabolite nitrite (NO_2^-) using the Griess assay. J774.1 cells were seeded into 48-well plates and cultured overnight. After washing with phosphate-buffered saline (PBS), the medium was replaced with phenol red-free DMEM containing LPS (1 $\mu\text{g}/\text{ml}$) and adenosine analogs (1 mM). After incubation for 24 h, each cultured medium (100 μl) was mixed with 100 μl of Griess reagent [1% (w/v) sulfanilamide in 5% (w/w) phosphoric acid and 0.1% (w/v) *N*-(1-naphthyl)ethylenediamine dihydrochloride in water]. The optical density was then measured at 550 nm. The nitrite levels were calculated using a standard curve derived from authentic sodium nitrite.

Western blotting

J774.1 cells were seeded into 24-well plates and cultured overnight. After treatment with each agent for the indicated times, the cells were washed with PBS and lysed with a radio-immunoprecipitation assay buffer [50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 0.25% deoxycholic acid, 1% NP-40, 0.1% sodium dodecyl sulfate (SDS), 1 mM ethylenediaminetetraacetic acid] supplemented with both protease and phosphatase inhibitor cocktails. The protein concentrations were determined by a Protein Assay Bicinchoninate Kit (Nacalai Tesque, Kyoto, Japan). The protein samples were denatured by heating at 95°C for 5 min in the presence of SDS (2%) and 2-mercaptoethanol (5%), then separated using 10% SDS-polyacrylamide gels followed by a semi-dry blotting onto a polyvinylidene difluoride membrane (Hybond P, GE Healthcare). After blocking with EzBlock Chemi (Atto Corp., Tokyo, Japan) in Tris-buffered saline containing 0.05% Tween 20 (TTBS) at room temperature for 30 min, the membrane was washed with TTBS, then incubated overnight with a primary antibody (1:1000) in TTBS containing 5% BSA at 4°C. After washing, the membrane was incubated with horseradish peroxidase-conjugated secondary antibody (1:2000) in TTBS containing 5% skim milk at room temperature for 1 h. After washing, chemiluminescent detection was performed using Chemi-Lumi One L or Chemi-Lumi One Super (Nacalai Tesque, Kyoto, Japan) and visualized by Ez-Capture MG (Atto Corp., Tokyo, Japan).

Real-time reverse transcription-polymerase chain reaction

The total RNA was extracted from the cells using Sepasol-RNA I Super G (Nacalai Tesque, Kyoto, Japan). cDNA was synthesized using ReverTra Ace qPCR RT Master Mix with gDNA Remover (Code No. FSQ-301, Toyobo Co., Ltd., Osaka, Japan). PCR amplification was performed with THUNDERBIRD SYBR qPCR Mix (Code No. QPS-201, Toyobo Co., Ltd., Osaka, Japan) using the LightCycler Nano System (Roche Diagnostics K.K., Tokyo, Japan). The relative expression of each mRNA was calculated according to the $2^{-\Delta\Delta\text{Ct}}$ method and normalized to the glyceraldehyde-3-phosphate dehydrogenase (GAPDH) expression. The primer sequences are listed in the [S1 Table](#).

Measurement of calcium influx

Cells were seeded into 48-well plates and cultured overnight. After washing with PBS, the medium was replaced with phenol red-free DMEM containing Fluo 4-AM (5 $\mu\text{g}/\text{ml}$), Pluronic F-127 (0.02%), and probenecid (1.25 mM). After incubation for 1 h, the medium was replaced with phenol red-free DMEM containing each polyphenol (50 μM) and probenecid (1.25 mM). After incubation for 10 min, an equal volume of phenol red-free DMEM containing ATP (4 mM) and the corresponding polyphenol (50 μM) was added to each well. After incubation for 10 min, the cell plates were scanned using Typhoon FLA 9500 at the following settings: 473

nm blue LD laser, BPB1 emission filter, 500 V photomultiplier, 100- μ m pixel size. The fluorescence intensity of each well was measured using Image Quant TL ver. 8.1.

Cytotoxicity assay

Cells were seeded into 48-well plates and cultured overnight. After each stimulation, the medium was replaced with DMEM containing MTT (0.2 mg/ml) and incubated for 30 min. After removing the medium, the formazan was dissolved in dimethyl sulfoxide (200 μ l/well) and the optical density was measured at 550 nm. The relative cell viability was calculated using the following formula: relative cell viability (%) = optical density of experimental groups / optical density of the control group \times 100.

ELISA for murine interleukin-1 β

The amount of interleukin-1 β (IL-1 β) was determined according to the recommended protocol for the ELISA Development Kit for murine IL-1 β (PeproTech, USA). Colorimetric detection was performed using the TMB Substrate Reagent Set (BD Biosciences, USA). The reaction was terminated by adding 2N sulfuric acid and the optical density was measured at 450 nm.

Statistics

The data represent means \pm S.D. of triplicate determinations. All the statistical analyses were performed by Tukey-Kramer method. The asterisks indicate statistical significance ($P < 0.05$).

Results

ATP accelerates NO production in LPS-stimulated J774.1 mouse macrophages

To examine the effects of ATP on the macrophage inflammation, J774.1 cells were treated with LPS in the presence or absence of ATP for 12–36 h. We observed that the LPS-induced NO production was enhanced after 24 h in the presence of ATP, although NO production was not induced by ATP alone (Fig 1A and Figure A in S1 Fig). The cell viability was not affected by the ATP (Figure B in S1 Fig). Both the protein and mRNA expressions of iNOS, a key enzyme generating NO, were also up-regulated by the addition of ATP (Fig 1B and 1C), but not by ATP alone (Figure C in S1 Fig). Given that the extracellular ATP is rapidly degraded by ectonucleotidases, such as CD39 [40,41], we investigated whether the NO production could certainly be enhanced by ATP, or the degradation products. As shown in Fig 1D, the enhanced NO production by ATP was abolished by the treatment with apyrase, an enzyme that hydrolyzes ATP into AMP. We also confirmed that ADP and AMP did not enhance the LPS-induced NO production (Figure D in S1 Fig).

ATP facilitates IFN- β expression to enhance the NO production

To investigate the molecular mechanisms of the ATP-enhanced NO production, we analyzed the upstream signaling pathway regulating the iNOS transcription. As it is reported that secreted interferons are necessary for full expression of the iNOS mRNA [42,43], we evaluated the involvement of IFN- β , expressed strongly in J774.1 cells, in the LPS/ATP-induced expression of iNOS. As shown in Fig 2A, the addition of anti-IFN- β neutralizing antibody almost completely suppressed the LPS-induced iNOS protein expression in both presence and absence of ATP, suggesting the involvement of autocrine/paracrine signaling of IFN- β in LPS-induced iNOS expression. Indeed, consistent with the induction of iNOS, ATP enhanced the LPS-induced IFN- β mRNA expression and phosphorylation of STAT1, downstream of the

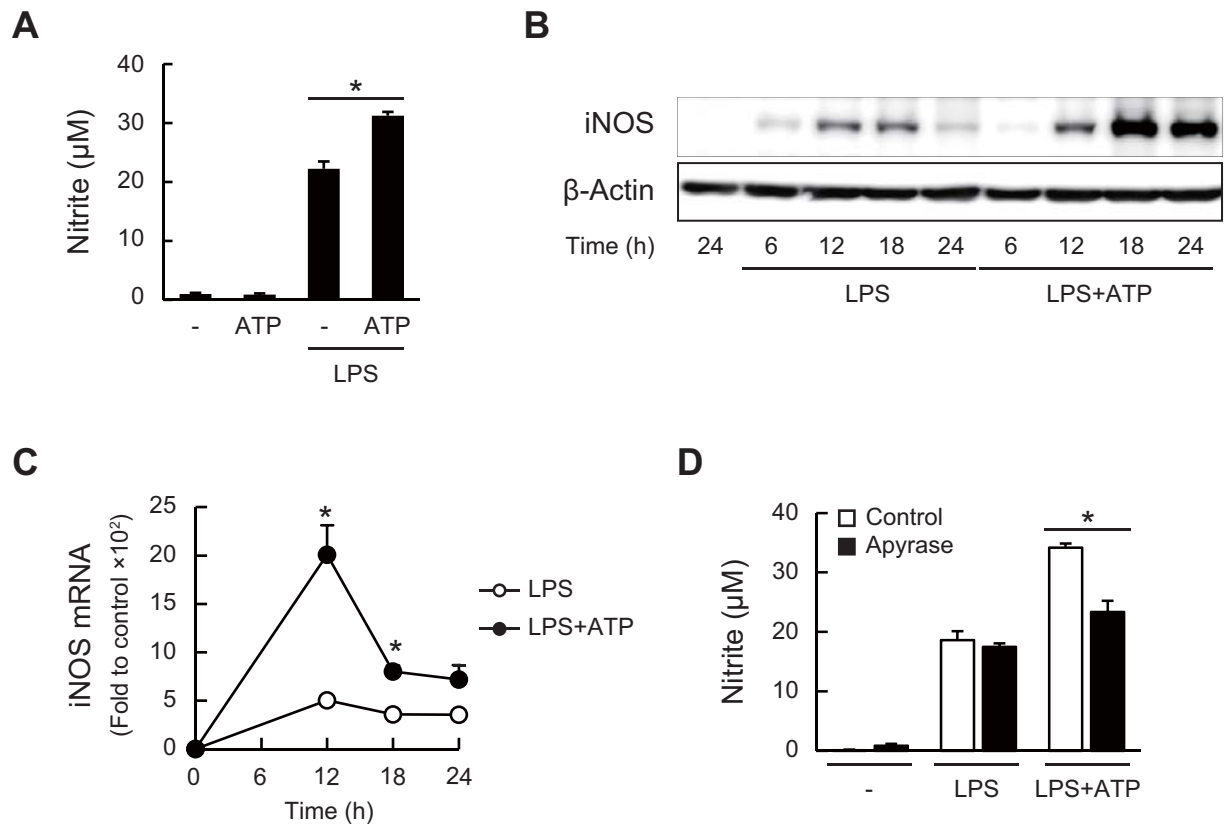


Fig 1. ATP accelerates LPS-induced NO production and iNOS expression. (A) NO production in J774.1 cells was evaluated by measuring the metabolite nitrite (NO₂⁻) in culture supernatants using the Griess assay. The cells were treated with LPS (1 µg/ml) in the presence or absence of ATP (1 mM) for 24 h (**P* < 0.05). (B) iNOS protein expression in LPS/ATP-treated cells was analyzed by western blotting. β-Actin was detected as the internal control. (C) Relative expression of iNOS mRNA measured by quantitative RT-PCR. GAPDH was evaluated as the normalization control (**P* < 0.05 vs LPS-treated group). (D) Effect of apyrase (0.5 units/ml) on LPS/ATP-stimulated NO production (24 h after stimulation, **P* < 0.05). (A, C, and D) Data represent means ± S.D. of triplicate determinations.

<https://doi.org/10.1371/journal.pone.0204229.g001>

IFN-β signaling (Figs 2B and 2C). In addition, the phosphorylation of IRF-3, the essential transcription factor of IFN-β [44], was also up-regulated by the addition of ATP (Fig 2D).

P2X7R activation up-regulates IFN-β expression and NO production

It has been reported that P2X7R, expressed on most immune cells, such as macrophages, is activated through millimolar concentrations of extracellular ATP (0.1–4 mM) and participates in inflammation and pain mechanisms [30,45–48]. Therefore, we next examined the involvement of P2X7R in the enhancement of the LPS-induced inflammation by ATP. As shown in Fig 3A, LPS-induced NO production, especially in the presence of ATP, was significantly reduced in the P2X7R knockdown cells, showing the effects of ATP through P2X7R activation. Since the activated P2X7R has been shown to facilitate the influx of extracellular cations, including calcium ions (Ca²⁺) [48,49], we next investigated the roles of the Ca²⁺ influx in the LPS/ATP-induced IFN-β expression. As shown in Fig 3B, BAPTA-AM, an intracellular Ca²⁺ chelator, significantly inhibited the LPS/ATP-induced IFN-β expression. In addition, A23187, a Ca²⁺ ionophore, also reproduced the enhancing effect of ATP on the IFN-β expression (Fig 3C). We confirmed that the Ca²⁺ influx, monitored using Fluo 4-AM, was induced by the treatment of ATP for 10 min and was abrogated in the P2X7R knockdown cells (Fig 3D).

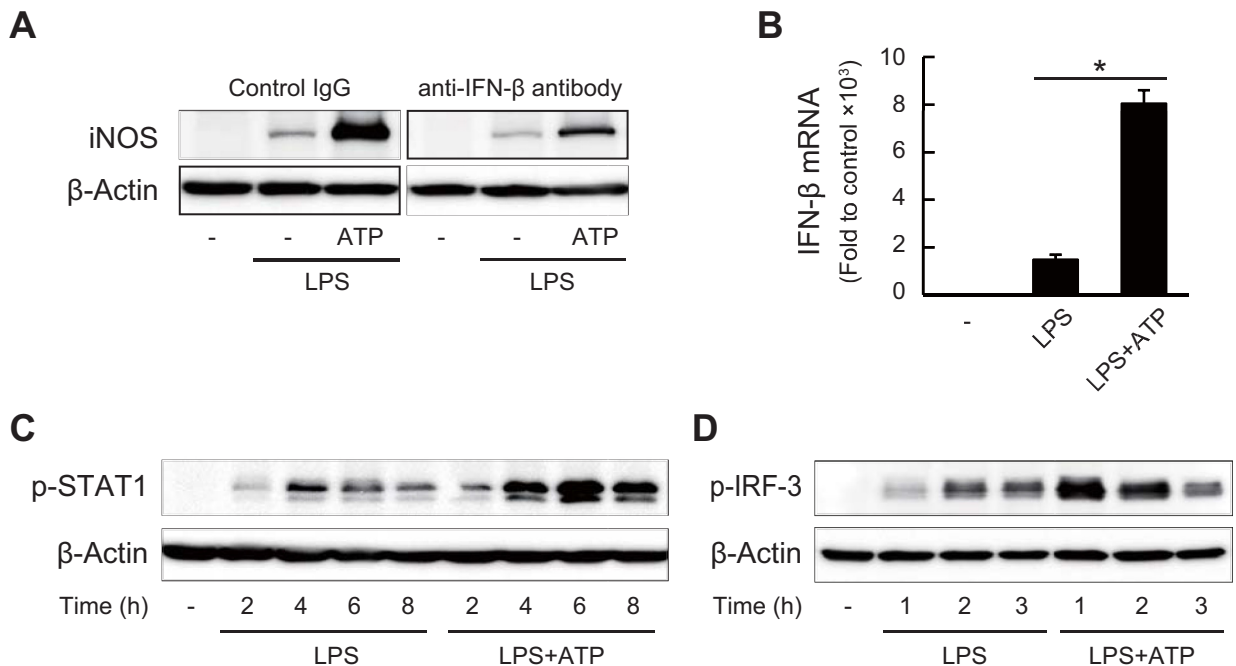


Fig 2. ATP facilitates LPS-induced IFN-β expression leading to NO production. (A) Effects of neutralizing antibody to IFN-β on iNOS protein expression in J774.1 cells treated with LPS (1 μg/ml) and ATP (1 mM) for 12 h. Lanes were noncontiguous but from same blot membrane at same time. (B) Relative expression of IFN-β mRNA in LPS/ATP-treated cells (3 h) measured by quantitative RT-PCR. Data represent means ± S.D. of triplicate determinations (**P* < 0.05). (C) Phosphorylation of STAT1 in LPS/ATP-treated cells after 2–8 h. (D) Phosphorylation of IRF-3 in LPS/ATP-treated cells after 1–3 h.

<https://doi.org/10.1371/journal.pone.0204229.g002>

These results strongly suggest that the ATP/P2X7R-mediated Ca²⁺ influx contributes to the LPS/ATP-induced IFN-β expression.

P2X7R-mediated ATP signaling affects TLR4 signaling to enhance IFN-β expression and NO production

Next, to understand the actions of ATP on enhanced inflammation, we examined the timing of the ATP stimulation for enhancing LPS-induced NO production. As shown in Fig 4A, we found that the LPS-induced NO production was enhanced only when cells were co-treated with LPS and ATP, but hardly affected when ATP was added a few hours after the LPS stimulation. This result suggested that the P2X7R-mediated ATP signaling stimulated upstream of the LPS-induced TLR4 signaling. As shown in Fig 4B, ATP-enhanced NO production in LPS-treated cells was not observed in the presence of a specific TLR4 inhibitor TAK-242, showing the effects of ATP on the TLR4 signaling. We then investigated whether ATP could activate the LPS/TLR4 signaling pathways, such as NF-κB and MAPKs pathways. As shown in Fig 4C, the LPS-induced degradation of IκB-α and phosphorylation of ERK and JNK at 30 min were up-regulated by ATP. Phosphorylation of p38 was not affected by ATP. The phosphorylation of ERK was also induced by ATP alone. It was reported that the activation of these pathways results in the activation of the downstream transcription factors, such as NF-κB and activator protein-1, to induce the IFN-β and iNOS mRNA expression [42,43,50]. In addition, it has been reported that the P2X7R activation induces ERK phosphorylation via Ca²⁺-dependent or -independent activation of the upstream kinases [51–53]. We then investigated the involvement of the ERK phosphorylation on the effects of ATP. As shown in Fig 4D, PD98059, an inhibitor of the MEK1/2-ERK pathway, attenuated the enhancing effects of ATP on the IFN-β

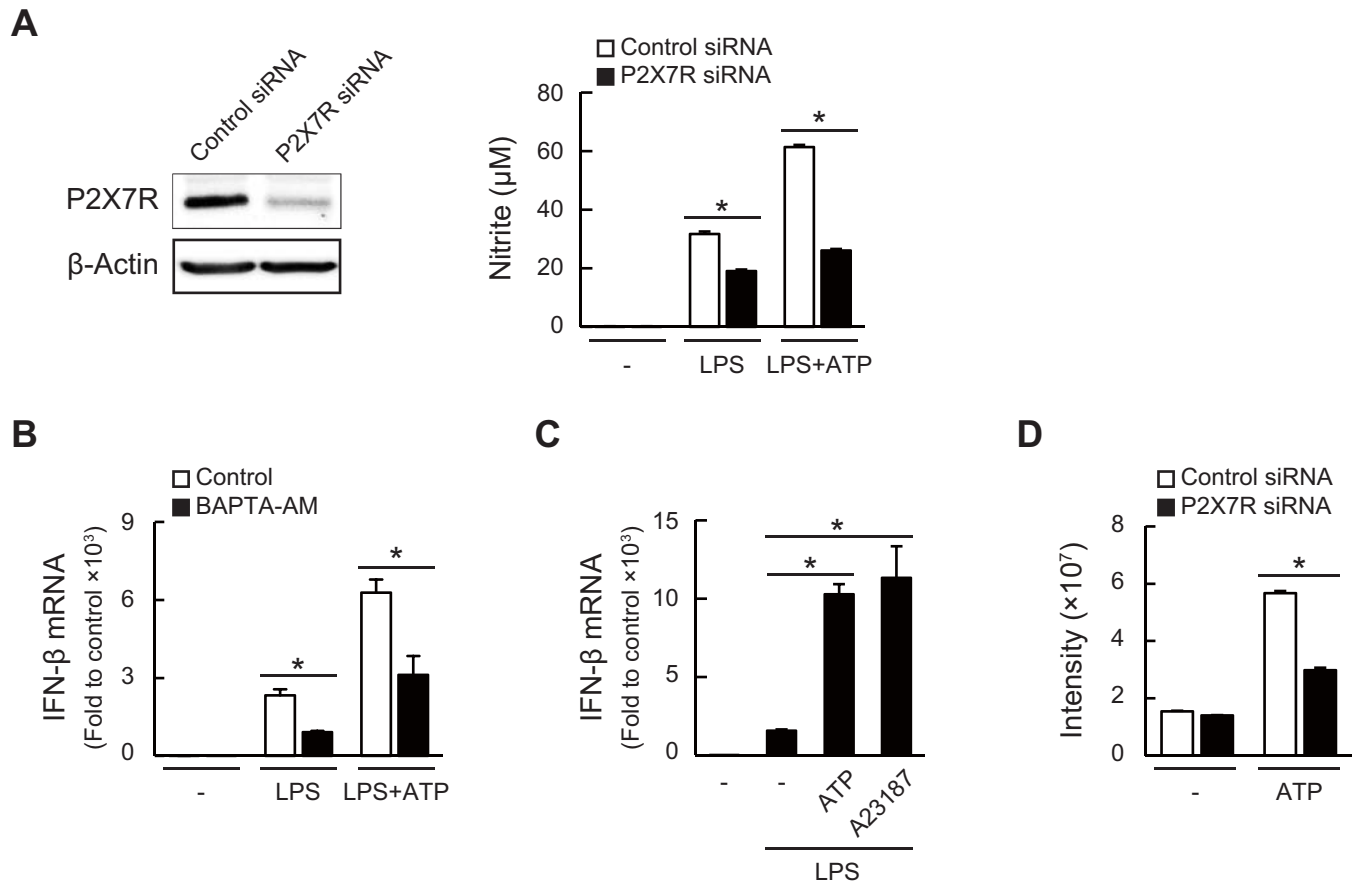


Fig 3. P2X7R activation and following Ca²⁺ influx up-regulate IFN-β expression and NO production. (A) Left, P2X7R protein expression was analyzed 24 h after transfection of control or P2X7R-specific siRNA. Right, NO production in transfected cells was evaluated using Griess assay after stimulation with LPS (1 μg/ml) in the presence or absence of ATP (1 mM). (B) Effects of BAPTA-AM (20 μM) pre-treatment (1 h) on LPS/ATP-induced IFN-β mRNA expression (3 h after stimulation). (C) Effects of A23187 (10 μM) on LPS/ATP-induced IFN-β mRNA expression (3 h after stimulation). (D) Ca²⁺ influx in control or P2X7R knockdown cells was measured using Fluo 4 fluorescence at 10 min after ATP (2 mM) stimulation. All the data represent means ± S.D. of triplicate determinations (**P* < 0.05).

<https://doi.org/10.1371/journal.pone.0204229.g003>

expression. It was also confirmed that the ATP-induced ERK phosphorylation was indeed inhibited in the presence of PD98059 (Fig 4E). Furthermore, the ATP-induced ERK phosphorylation was reduced in the P2X7R knockdown cells and also inhibited by BAPTA-AM (Fig 4F and 4G). These results suggest that ATP sensing by P2X7R and the following Ca²⁺ influx and ERK phosphorylation may be potential pathways for enhancing the LPS-induced IFN-β expression, leading to the increased iNOS expression and NO production.

Inhibitory effects of polyphenols on ATP/P2X7R signaling

We next investigated the inhibitory effects of natural polyphenols on the ATP/P2X7R signaling by measuring the Ca²⁺ influx as an indicator of the P2X7R activation. Among the 42 polyphenols (listed in S2 Table), we found that, as well as brilliant blue G (BBG), a P2X7R antagonist [54], baicalein and resveratrol exhibited the most significant inhibitory effects on the ATP-induced Ca²⁺ influx (Fig 5A). It was also confirmed that these two polyphenols inhibited the P2X7R activation in dose-dependent manners (Fig 5B). On the other hand, inhibitory effects of these polyphenols on the ATP-induced ERK phosphorylation could not be observed (Figure A in S2 Fig). This may be due to the fact that these polyphenols induced ERK phosphorylation

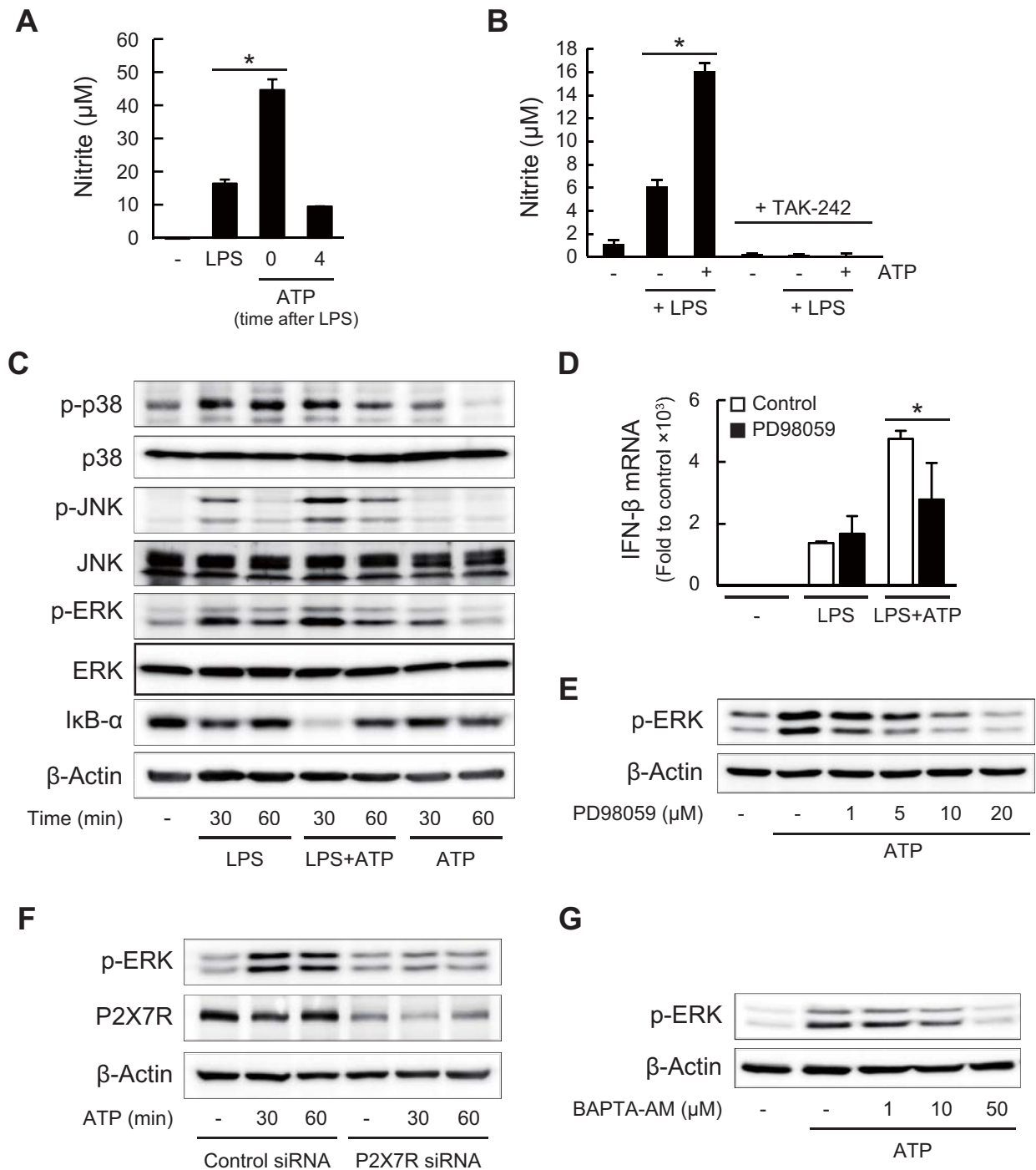


Fig 4. P2X7R-mediated ATP signaling affects LPS/TLR4 signaling. (A) Effects of co- and post-treatment with ATP on LPS-induced NO production in J774.1 cells. ATP (1 mM) was added at 0 or 4 h after LPS (1 µg/ml) treatment. NO production was evaluated by Griess assay 24 h after LPS treatment (**P* < 0.05). (B) Effects of a TLR4 inhibitor TAK-242 (5 µM) on LPS/ATP-induced NO production in J774.1 cells (**P* < 0.05). (C) Phosphorylation of p38, JNK, and ERK, as well as the total proteins, and IκB-α degradation in LPS/ATP-treated cells were analyzed by western blotting. (D) Effects of PD98059 (20 µM) pre-treatment (1 h) on LPS/ATP-induced IFN-β mRNA expression 3 h after stimulation (**P* < 0.05). (E) Effects of PD98059 (pre-treatment for 1 h) on ATP-induced ERK phosphorylation (1 h after ATP stimulation). (F) Effects of P2X7R knockdown on ATP-induced ERK phosphorylation. Cells transfected with control or P2X7R-specific siRNA were stimulated with ATP (1 mM). (G) Effects of BAPTA-AM (pre-treatment for 30 min) on ATP-induced ERK phosphorylation. (A, B and D) Data represent means ± S.D. of triplicate determinations.

<https://doi.org/10.1371/journal.pone.0204229.g004>

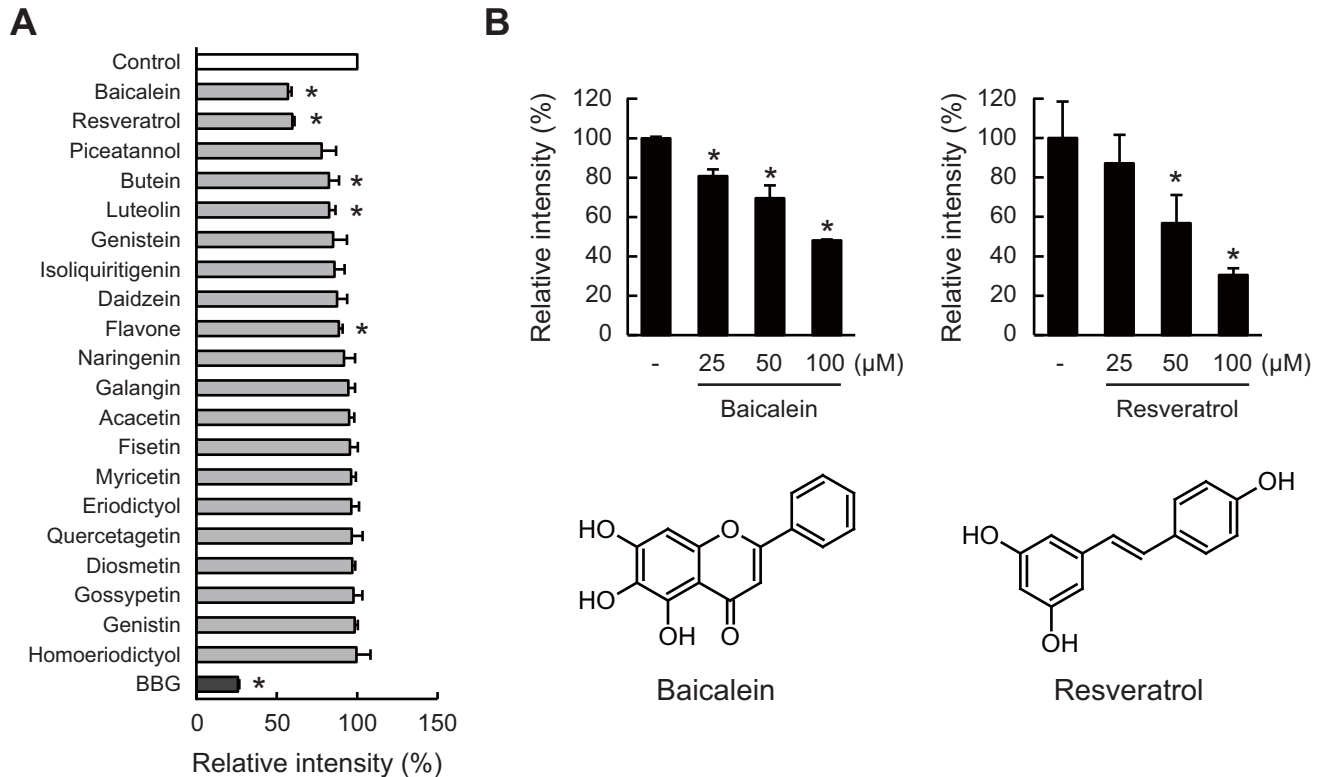


Fig 5. Inhibitory effects of polyphenols on ATP/P2X7R-mediated Ca²⁺ influx. (A) J774.1 cells were pre-treated with each polyphenol (50 μM) or BBG (2 μM) for 10 min followed by ATP stimulation (2 mM) for 10 min. Relative Fluo 4 fluorescence intensities (%) to control (stimulated with ATP alone) were calculated. Data for polyphenols that exhibited inhibitory effects (below 100%) are shown in the graph. (B) Dose-dependent inhibition of ATP-induced Ca²⁺ influx by baicalein (left) and resveratrol (right). Chemical structures of baicalein and resveratrol are also illustrated. All the data represent means ± S.D. of triplicate determinations (*P < 0.05 vs control).

<https://doi.org/10.1371/journal.pone.0204229.g005>

regardless of the presence or absence of ATP. These results suggest that specific polyphenols could target the ATP/P2X7R signaling, probably by antagonizing ATP and suppressing the Ca²⁺ influx.

Anti-inflammatory effects of baicalein through interfering ATP/P2X7R signaling

The results shown above indicated that ATP facilitates inflammatory responses through P2X7R, and that several polyphenols, such as baicalein and resveratrol, inhibit ATP/P2X7R signaling. We next investigated whether these polyphenols could indeed attenuate inflammation through inhibiting ATP/P2X7R signaling. Pre-treatment of baicalein or resveratrol inhibited the LPS/ATP-induced NO production in dose-dependent manners (Fig 6A). We confirmed that these polyphenols did not affect the cell viability under this experimental condition (Figure B in S2 Fig). As reported in many papers, polyphenols could inhibit the LPS/TLR4 signaling [32,38]. Therefore, the inhibitory effects of these polyphenols on NO production were conceivably due to the inhibition of not only the ATP/P2X7R signaling but also the LPS/TLR4 signaling. Thus, to investigate whether the inhibition of the ATP/P2X7R signaling by these polyphenols definitely confers the anti-inflammatory effects, we examined the effects of baicalein and resveratrol on the IFN-β expression, more upstream than the NO production. As shown in Fig 6B, baicalein inhibited the ATP-enhanced IFN-β expression at a lower concentration that did not affect the expression induced by LPS alone, whereas resveratrol rather induced IFN-β expression under this condition.

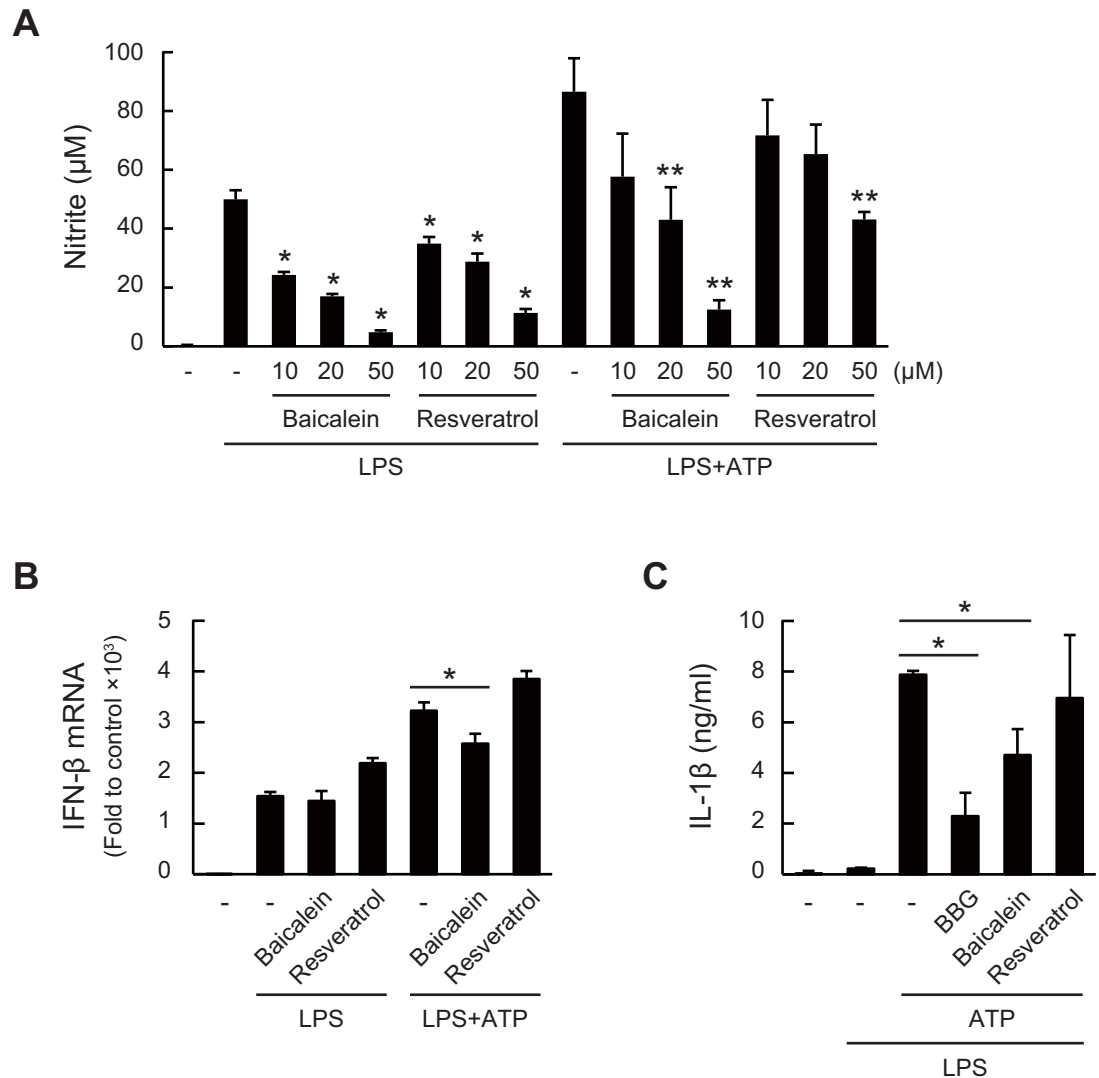


Fig 6. Anti-inflammatory effects of baicalein through interfering ATP/P2X7R signaling. (A) Dose-dependent effects of baicalein and resveratrol on LPS/ATP-induced NO production. Cells were pre-treated with each polyphenol for 1 h followed by LPS/ATP stimulation for 24 h (* $P < 0.05$ vs. LPS group and ** $P < 0.05$ vs. LPS + ATP group). (B) Effects of baicalein and resveratrol (10 µM pre-treatment for 1 h) on LPS/ATP-induced IFN-β mRNA expression 3 h after stimulation (* $P < 0.05$). (C) Effects of baicalein and resveratrol on ATP-triggered IL-1β secretion in LPS-stimulated J774.1 cells. Cells were treated with LPS (1 µg/ml) for 4 h followed by the addition of ATP (5 mM). Each polyphenol (50 µM) or BBG (10 µM) was added 30 min before the ATP treatment (* $P < 0.05$). All the data represent means ± S.D. of triplicate determinations.

<https://doi.org/10.1371/journal.pone.0204229.g006>

To further propose that the polyphenols could attenuate the ATP/P2X7R signaling, we then examined the inhibitory effects of baicalein and resveratrol on the secretion of IL-1β, an LPS-induced pro-inflammatory cytokine, the production of which is tightly regulated and specifically triggered by ATP as secondary stimuli [55]. As shown in Fig 6C, IL-1β was hardly detectable in cultured supernatants of the cells treated with the vehicle and LPS alone. On the other hand, the secretion of IL-1β was significantly induced upon ATP treatment 4 h after the LPS treatment. The secretion was significantly inhibited in the presence of BBG, showing that ATP sensing by P2X7R specifically contributes to IL-1β secretion. Under this experimental condition, the IL-1β secretion was significantly inhibited by pre-treatment with baicalein for 10 min before the ATP stimulation. On the other hand, similar to the results on IFN-β expression,

resveratrol did not inhibit the IL-1 β secretion. These results demonstrated that baicalein could be a potential candidate as a natural inhibitor of the ATP/P2X7R signaling.

Discussion

Inflammation is an important defense response in noxious conditions, such as infection, whereas abnormal inflammatory responses are shown to cause various diseases. LPS-induced inflammation is a well-studied model of infection and diseases, therefore, the molecular signaling through the TLR4 receptor could be a target for therapy. Recent studies have shown that released endogenous molecules from damaged or immune cells, called DAMPs, are also implicated in the activation of inflammatory responses. In this study, we investigated the molecular actions of extracellular ATP, considered as one of the prototypical DAMP molecules [56,57], on the LPS-induced macrophage inflammation. Based on the molecular actions of ATP, to develop a new strategy for the prevention of inflammatory diseases, we evaluated the inhibitory effects of natural polyphenols on ATP signaling. We first observed that NO production and iNOS expression in the LPS-stimulated J774.1 macrophages were enhanced by ATP (Fig 1A–1C). It has been reported that the extracellular ATP is rapidly degraded to ADP and AMP, whereas the LPS-induced NO production was not enhanced by these degradation products (Figure D in S1 Fig). Although we did not examine the actual enzyme activity of ectonucleotidases in J774.1 cells, we scarcely detected the degradation products of ATP by a liquid chromatography-tandem mass spectrometry in culture supernatants of J774.1 treated with 1 mM ATP, strongly suggesting that the effects of ATP in this study reflect the action of ATP itself. NO has been recognized not only as a vasodilator and a neurotransmitter, but also as anti-microbial, anti-tumor, tissue-damaging, and immune-modulating products in the immune system. NO plays a dual role during inflammation by exerting protective and toxic effects in parallel, and this dual effect has been explained as being dependent on the NO concentration [58–60]. Although we did not investigate the phenotypic consequences of the up-regulated NO production, we speculated that LPS/ATP-enhanced NO production might lead to disorders and diseases associated with excess inflammation.

We then investigated the activation of the transcription factors and signal transduction pathways involved in the LPS/ATP-induced iNOS expression. We observed that the LPS-induced IFN- β expression and the downstream STAT1 phosphorylation were up-regulated by ATP. STAT1 has been reported to be an essential transcription factor for the iNOS expression [43]. In addition, we found that ATP facilitated the LPS-induced TLR4 signaling, such as the NF- κ B and MAPK pathways, and the phosphorylation of IRF-3, the transcription factor for IFN- β . TLR4 signaling is generally separated into the myeloid differentiation primary response gene 88 (MyD88)-dependent pathway for expression of the pro-inflammatory cytokines and the TIR domain-containing adaptor protein-inducing IFN- β (TRIF)-dependent pathway for expression of the Type I IFNs. It has been suggested that NF- κ B and MAPK could be activated through both pathways and that IRF-3 could be through the TRIF-dependent pathway [61,62]. Our results indicated that ATP accelerated both pathways, leading to the activation of the iNOS transcription factors. We also found that the mRNA expression of other LPS-induced inflammatory mediators, such as tumor necrosis factor- α (TNF- α), cyclooxygenase-2 (COX-2), IL-6, IL-10, and IL-1 β , were also affected in the presence of ATP (S3 Fig). These results suggested that ATP accelerated upstream pathways in the LPS/TLR4 signaling resulting in perturbation of the downstream mRNA expression of iNOS and other inflammatory mediators.

We focused on the involvement of P2X7R regarding the effects of ATP because P2X7R is highly expressed in immune cells and responds to higher ATP concentrations. Using the siRNA-induced knockdown and antagonist of P2X7R, we demonstrated that P2X7R is indeed

involved in the ATP-enhanced inflammatory responses. Our results that the LPS-induced NO production without ATP addition was also reduced in the P2X7R knockdown cells (Fig 3A) suggest the activation of the P2X7R signaling during stimulation with only LPS. Although similar observations that ATP or the P2X7R agonist enhanced LPS-induced inflammatory responses including IFN- β expression have been reported [63,64], our results clearly revealed the molecular actions of ATP through P2X7R, leading to enhanced NO production. For example, we demonstrated that ATP stimulated the Ca²⁺ influx and activated intracellular signaling, such as ERK phosphorylation, which subsequently enhanced the expression of the LPS-induced IFN- β expression and NO production. In addition, the LPS/ATP-enhanced IFN- β expression was reduced by BAPTA-AM and PD98059 (Figs 3 and 4), suggesting that the intracellular Ca²⁺ and the activation of MEK/ERK are both important for the LPS/ATP-induced inflammatory signaling. It has been reported that ATP binding to the extracellular loop of P2X7R results in not only channel opening but also structural and/or functional changes of the intracellular proteins or membrane lipids, such as calmodulin, heat shock proteins, cytoskeletal proteins, and phosphatidylinositol 4,5-phosphate, all of which bind to the C-terminal of P2X7R [65–67]. Therefore, these observations suggest that both the Ca²⁺-dependent and -independent signals of P2X7R are potential pathways in the ATP-induced inflammatory regulation.

Many studies, including the pre-clinical studies, have suggested that P2X7R could be therapeutically important targets because of the involvement in inflammatory diseases and pain. Although various negative or positive modulators of the P2X7R activation including natural products, synthetic molecules, and monoclonal antibodies, have been demonstrated, there is no approved medicine in clinical use [30,68–70]. In addition, little is known about the beneficial effects of dietary compounds targeting P2X7R [29]. Therefore, to prevent ATP/P2X7R-mediated abnormal inflammation, we explored the inhibitory effects of polyphenols, the largest group of phytochemicals, on the ATP/P2X7R signaling. We then found that a flavonoid baicalein (5,6,7-trihydroxyflavone) and a stilbenoid resveratrol significantly inhibited the ATP/P2X7R-mediated Ca²⁺ influx (Fig 5). We further investigated the effects of baicalein and resveratrol on the LPS/ATP-induced inflammation, and found that, as expected, these polyphenols indeed inhibited the LPS/ATP-induced NO production (Fig 6A). However, at least under this experimental condition, we could not distinguish whether these polyphenols modulated the LPS/TLR4 signaling or ATP/P2X7R signaling because many reports have demonstrated the inhibitory effects of polyphenols on the TLR4 signaling. For instance, resveratrol inhibits the kinase activity of TANK binding kinase 1 associating with TRIF, which is the TLR4 downstream component, resulting in decreasing the NF- κ B and IRF-3 activation [71,72]. Another study also reported that quercetin inhibits the Src- and Syk-mediated PI3K phosphorylation and subsequent TLR4/MyD88/PI3K complex formation, resulting in the reduced activation of downstream signaling [73]. Therefore, to elucidate that the inhibition of the ATP/P2X7R signaling by baicalein or resveratrol definitely contributes to the anti-inflammatory or therapeutic effects, we evaluated the effects of these polyphenols on IL-1 β secretion. In culture experiments, IL-1 β is neither matured nor secreted by the treatment with only LPS. It is generally recognized that ATP addition after a few hours of LPS stimulation triggered inflammasome activation leading to the formation and secretion of the matured IL-1 β [18,55]. We then examined the effects of two polyphenols on the ATP-triggered IL-1 β secretion and found that the IL-1 β secretion was significantly inhibited by baicalein, but unfortunately, not by resveratrol (Fig 6C). Although we did not investigate the activation of inflammasome in this study, the result strongly suggested that baicalein reduced the maturation and secretion of IL-1 β through inhibition of the ATP/P2X7R signaling. Effects of antagonizing ATP/P2X7R signaling by baicalein or other natural products on inflammatory responses *in vivo* have not

yet been determined within this study. Utilization of P2X7R-deficient mice in the near future will clarify the involvements of ATP in inflammatory responses and the possible interactions between natural polyphenols and P2X7R *in vivo*.

Baicalein and its glucuronide (baicalin), the major components of *Scutellaria baicalensis* Georgi, a popular herb used in traditional Chinese herbal medicine, are shown to prevent cancer, inflammation, cardiovascular diseases, and hypertension [74]. He *et al.* reported that baicalein inhibited TLR4 expression, NF- κ B, and MAPK signal pathways and subsequently reduced the levels of pro-inflammatory cytokines [75]. Our current results also proposed a new health-beneficial activity of baicalein through inhibition of ATP/P2X7R signaling. Although the inhibitory effect of baicalin was not examined in this study, we previously reported that the glucuronides of flavonoids could be deconjugated into the aglycones by macrophages, resulting in increased anti-inflammatory activity as compared to the parent glucuronides [76]. Therefore, interaction between macrophages and baicalin could result in the inhibitory effect on the ATP/P2X7R signaling, similar to the action of the aglycone baicalein. On the other hand, in spite of the inhibitory effect on ATP-induced Ca^{2+} influx, resveratrol did not inhibit ATP-enhanced IFN- β expression and IL-1 β secretion (Fig 6B and 6C). Resveratrol rather induced LPS/ATP-induced IFN- β expression. Although we predicted that both baicalein and resveratrol could also inhibit the ATP/P2X7R-mediated ERK phosphorylation, as well as Ca^{2+} influx, these polyphenols, especially resveratrol, conversely enhanced the ATP-induced ERK phosphorylation (Figure A in S2 Fig). Similarly, several papers also demonstrated the ERK activation by polyphenols [77–80]. Within this study, it is still unclear how tight the link between Ca^{2+} influx and ERK phosphorylation is in the ATP signaling. Resveratrol, found in grape products such as red wine, has been shown to improve cardiovascular performance and extend the lifespans of non-mammals and mice [81]. However, we could not conclude that, at least from our observations, resveratrol is a potent inhibitor of ATP/P2X7R signaling.

Overall, our results indicated the potential preventive effects of baicalein on exacerbating the ATP/P2X7R signaling, not only LPS signaling, under noxious or abnormal conditions. We demonstrated the molecular actions of the extracellular ATP through P2X7R on the LPS signaling, resulting in excess inflammation, and also proposed a new strategy for the prevention of excess inflammation by inhibiting the ATP/P2X7R signaling using natural polyphenols, such as baicalein. As well as ATP/P2X7R, involvements of other pairs of DAMPs/receptors such as advanced glycation end-product/RAGEs and β -amyloid, in the inflammatory responses have been suggested [82–83]. Therefore, our conclusion is that the ATP/P2X7R signaling and other DAMPs signaling could be a new potential target of drugs and natural compounds including polyphenols for the prevention of disorders and diseases associated with excess inflammation.

Supporting information

S1 Fig. Effects of ATP on NO production and cell viability. (A) NO production in J774.1 cells was evaluated using the Griess assay. Cells were treated with LPS (1 $\mu\text{g}/\text{ml}$) in the presence or absence of ATP (1 mM) for the indicated time periods ($*P < 0.05$ vs LPS-treated group). (B) Cell viability (%) was analyzed by cytotoxicity assay after LPS/ATP stimulation at the indicated time points. (C) Protein and mRNA expression of iNOS in LPS/ATP-stimulated J774.1 cells. Cells were treated with LPS (1 $\mu\text{g}/\text{ml}$) in the presence or absence of ATP (1 mM) or ATP alone for the indicated time periods. iNOS protein expression (β -actin as a loading control) was evaluated by immunoblotting. iNOS mRNA expression was determined by RT-qPCR (GAPDH as endogenous control, duplicate determinations). (D) Effects of ATP, ADP, and

AMP on the LPS-induced NO production ($*P < 0.05$). All the data represent means \pm S.D. of triplicate determinations.

(PDF)

S2 Fig. Effects of baicalein and resveratrol on ATP-induced ERK phosphorylation and cell viability. (A) Dose-dependent effects of baicalein and resveratrol on ATP-induced ERK phosphorylation in J774.1 cells. Cells were pre-treated with each polyphenol for 1 h followed by ATP (1 mM) stimulation for 1 h. (B) Cell viability (%) was analyzed by cytotoxicity assay after treatment with each polyphenol for 24 h. Data represent means \pm S.D. of triplicate determinations.

(PDF)

S3 Fig. Effects of ATP on the expression of LPS-induced inflammatory mediators. Relative mRNA expression of TNF- α (A), COX-2 (B), IL-6 (C), IL-10 (D), and IL-1 β (E) in J774.1 cells were measured by quantitative RT-PCR after LPS/ATP treatment. All the data represent means \pm S.D. of triplicate determinations ($*P < 0.05$, A, vs LPS + ATP-treated group; B-E, vs LPS-treated group).

(PDF)

S1 Table. Primer sequences used for quantitative RT-PCR.

(PDF)

S2 Table. Chemical structures and names of selected polyphenols used in this study.

(PDF)

Author Contributions

Conceptualization: Yoshichika Kawai.

Funding acquisition: Yoshichika Kawai.

Investigation: Erika Nuka.

Methodology: Kohta Ohnishi, Yoshichika Kawai.

Resources: Junji Terao.

Supervision: Junji Terao, Yoshichika Kawai.

Writing – original draft: Erika Nuka, Yoshichika Kawai.

References

1. Medzhitov R. Inflammation 2010: new adventures of an old flame. *Cell*. 2010 Mar 19; 140(6):771–6. <https://doi.org/10.1016/j.cell.2010.03.006> PMID: 20303867
2. Medzhitov R. Origin and physiological roles of inflammation. *Nature*. 2008 Jul 24; 454(7203):428–35. <https://doi.org/10.1038/nature07201> PMID: 18650913
3. Vazquez-Torres A, Jones-Carson J, Mastroeni P, Ischiropoulos H, Fang FC. Antimicrobial actions of the NADPH phagocyte oxidase and inducible nitric oxide synthase in experimental salmonellosis. I. Effects on microbial killing by activated peritoneal macrophages in vitro. *J Exp Med*. 2000 Jul 17; 192(2):227–36. PMID: 10899909
4. Donath MY, Shoelson SE. Type 2 diabetes as an inflammatory disease. *Nat Rev Immunol*. 2011 Feb; 11(2):98–107. <https://doi.org/10.1038/nri2925> PMID: 21233852
5. Golia E, Limongelli G, Natale F, Fimiani F, Maddaloni V, Parigiano I, et al. Inflammation and cardiovascular disease: from pathogenesis to therapeutic target. *Curr Atheroscler Rep*. 2014 Sep; 16(9):435. <https://doi.org/10.1007/s11883-014-0435-z> PMID: 25037581
6. Libby P. Inflammation in atherosclerosis. *Nature*. 2002 Dec 19–26; 420(6917):868–74. <https://doi.org/10.1038/nature01323> PMID: 12490960

7. Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. *Circulation*. 2002 Mar 5; 105(9):1135–43. PMID: [11877368](https://pubmed.ncbi.nlm.nih.gov/11877368/)
8. Lee YJ, Han SB, Nam SY, Oh KW, Hong JT. Inflammation and Alzheimer's disease. *Arch Pharm Res*. 2010 Oct; 33(10):1539–56. <https://doi.org/10.1007/s12272-010-1006-7> PMID: [21052932](https://pubmed.ncbi.nlm.nih.gov/21052932/)
9. Bagyinszky E, Giau VV, Shim K, Suk K, An SSA, Kim S. Role of inflammatory molecules in the Alzheimer's disease progression and diagnosis. *J Neurol Sci*. 2017 May 15; 376:242–254. <https://doi.org/10.1016/j.jns.2017.03.031> PMID: [28431620](https://pubmed.ncbi.nlm.nih.gov/28431620/)
10. Grivennikov SI, Greten FR, Karin M. Immunity, inflammation, and cancer. *Cell*. 2010 Mar 19; 140(6):883–99. <https://doi.org/10.1016/j.cell.2010.01.025> PMID: [20303878](https://pubmed.ncbi.nlm.nih.gov/20303878/)
11. Vénéreau E, Ceriotti C, Bianchi ME. DAMPs from Cell Death to New Life. *Front Immunol*. 2015 Aug 18; 6:422. <https://doi.org/10.3389/fimmu.2015.00422> PMID: [26347745](https://pubmed.ncbi.nlm.nih.gov/26347745/)
12. Piccinini AM, Midwood KS. DAMPening inflammation by modulating TLR signalling. *Mediators Inflamm*. 2010; 2010:672395. <https://doi.org/10.1155/2010/672395> PMID: [20706656](https://pubmed.ncbi.nlm.nih.gov/20706656/)
13. Yang H, Wang H, Ju Z, Ragab AA, Lundbäck P, Long W, et al. MD-2 is required for disulfide HMGB1-dependent TLR4 signaling. *J Exp Med*. 2015 Jan 12; 212(1):5–14. <https://doi.org/10.1084/jem.20141318> PMID: [25559892](https://pubmed.ncbi.nlm.nih.gov/25559892/)
14. Eltzschig HK, Sitkovsky MV, Robson SC. Purinergic signaling during inflammation. *N Engl J Med*. 2012 Dec 13; 367(24):2322–33. <https://doi.org/10.1056/NEJMra1205750> PMID: [23234515](https://pubmed.ncbi.nlm.nih.gov/23234515/)
15. Leist M, Single B, Castoldi AF, Kühnle S, Nicotera P. Intracellular adenosine triphosphate (ATP) concentration: a switch in the decision between apoptosis and necrosis. *J Exp Med*. 1997 Apr 21; 185(8):1481–6. PMID: [9126928](https://pubmed.ncbi.nlm.nih.gov/9126928/)
16. Lohman AW, Isakson BE. Differentiating connexin hemichannels and pannexin channels in cellular ATP release. *FEBS Lett*. 2014 Apr 17; 588(8):1379–88. <https://doi.org/10.1016/j.febslet.2014.02.004> PMID: [24548565](https://pubmed.ncbi.nlm.nih.gov/24548565/)
17. Wang Y, Martins I, Ma Y, Kepp O, Galluzzi L, Kroemer G. Autophagy-dependent ATP release from dying cells via lysosomal exocytosis. *Autophagy*. 2013 Oct; 9(10):1624–5. <https://doi.org/10.4161/auto.25873> PMID: [23989612](https://pubmed.ncbi.nlm.nih.gov/23989612/)
18. Giuliani AL, Sarti AC, Falzoni S, Di Virgilio F. The P2X7 Receptor-Interleukin-1 Liaison. *Front Pharmacol*. 2017 Mar 16; 8:123. <https://doi.org/10.3389/fphar.2017.00123> PMID: [28360855](https://pubmed.ncbi.nlm.nih.gov/28360855/)
19. Ralevic V, Burnstock G. Receptors for purines and pyrimidines. *Pharmacol Rev*. 1998 Sep; 50(3):413–92. PMID: [9755289](https://pubmed.ncbi.nlm.nih.gov/9755289/)
20. Elliott MR, Chekeni FB, Trampont PC, Lazarowski ER, Kadl A, Walk SF, et al. Nucleotides released by apoptotic cells act as a find-me signal to promote phagocytic clearance. *Nature*. 2009 Sep 10; 461(7261):282–6. <https://doi.org/10.1038/nature08296> PMID: [19741708](https://pubmed.ncbi.nlm.nih.gov/19741708/)
21. Lommatzsch M, Cicko S, Müller T, Lucattelli M, Bratke K, Stoll P, et al. Extracellular adenosine triphosphate and chronic obstructive pulmonary disease. *Am J Respir Crit Care Med*. 2010 May 1; 181(9):928–34. <https://doi.org/10.1164/rccm.200910-1506OC> PMID: [20093639](https://pubmed.ncbi.nlm.nih.gov/20093639/)
22. Riteau N, Gasse P, Fauconnier L, Gombault A, Couegnat M, Fick L, et al. Extracellular ATP is a danger signal activating P2X7 receptor in lung inflammation and fibrosis. *Am J Respir Crit Care Med*. 2010 Sep 15; 182(6):774–83. <https://doi.org/10.1164/rccm.201003-0359OC> PMID: [20522787](https://pubmed.ncbi.nlm.nih.gov/20522787/)
23. Idzko M, Hammad H, van Nimwegen M, Kool M, Willart MA, Muskens F, et al. Extracellular ATP triggers and maintains asthmatic airway inflammation by activating dendritic cells. *Nat Med*. 2007 Aug; 13(8):913–9. <https://doi.org/10.1038/nm1617> PMID: [17632526](https://pubmed.ncbi.nlm.nih.gov/17632526/)
24. Wan P, Liu X, Xiong Y, Ren Y, Chen J, Lu N, et al. Extracellular ATP mediates inflammatory responses in colitis via P2X7 receptor signaling. *Sci Rep*. 2016 Jan 7; 6:19108. <https://doi.org/10.1038/srep19108> PMID: [26739809](https://pubmed.ncbi.nlm.nih.gov/26739809/)
25. Portales-Cervantes L, Niño-Moreno P, Doníz-Padilla L, Baranda-Candido L, García-Hernández M, Salgado-Bustamante M, et al. Expression and function of the P2X₇ purinergic receptor in patients with systemic lupus erythematosus and rheumatoid arthritis. *Hum Immunol*. 2010 Aug; 71(8):818–25. <https://doi.org/10.1016/j.humimm.2010.05.008> PMID: [20493226](https://pubmed.ncbi.nlm.nih.gov/20493226/)
26. Zanin RF, Bergamin LS, Morrone FB, Coutinho-Silva R, de Souza Wyse AT, Battastini AM. Pathological concentrations of homocysteine increases IL-1 β production in macrophages in a P2X₇, NF- κ B, and erk-dependent manner. *Purinergic Signal*. 2015 Dec; 11(4):463–70. <https://doi.org/10.1007/s11302-015-9464-5> PMID: [26265456](https://pubmed.ncbi.nlm.nih.gov/26265456/)
27. Leyva-Grado VH, Ermler ME, Schotsaert M, Gonzalez MG, Gillespie V, Lim JK, et al. Contribution of the Purinergic Receptor P2X₇ to Development of Lung Immunopathology during Influenza Virus Infection. *MBio*. 2017 Mar 28; 8(2):e00229–17. <https://doi.org/10.1128/mBio.00229-17> PMID: [28351919](https://pubmed.ncbi.nlm.nih.gov/28351919/)

28. Ryu JK, Jantaratnotai N, Serrano-Perez MC, McGeer PL, McLarnon JG. Block of purinergic P2X7R inhibits tumor growth in a C6 glioma brain tumor animal model. *J Neuropathol Exp Neurol*. 2011 Jan; 70(1):13–22. <https://doi.org/10.1097/NEN.0b013e318201d4d4> PMID: 21157381
29. Soares-Bezerra RJ, Calheiros AS, da Silva Ferreira NC, da Silva Frutuoso V, Alves LA. Natural Products as a Source for New Anti-Inflammatory and Analgesic Compounds through the Inhibition of Purinergic P2X Receptors. *Pharmaceuticals (Basel)*. 2013 Apr 29; 6(5):650–8. <https://doi.org/10.3390/ph6050650> PMID: 24276172
30. Alves LA, Bezerra RJ, Faria RX, Ferreira LG, da Silva Frutuoso V. Physiological roles and potential therapeutic applications of the P2X7 receptor in inflammation and pain. *Molecules*. 2013 Sep 5; 18(9):10953–72. <https://doi.org/10.3390/molecules180910953> PMID: 24013409
31. Tangney CC, Rasmussen HE. Polyphenols, inflammation, and cardiovascular disease. *Curr Atheroscler Rep*. 2013 May; 15(5):324. <https://doi.org/10.1007/s11883-013-0324-x> PMID: 23512608
32. Santangelo C, Vari R, Scazzocchio B, Di Benedetto R, Filesi C, Masella R. Polyphenols, intracellular signalling and inflammation. *Ann Ist Super Sanita*. 2007; 43(4):394–405. PMID: 18209273
33. Gupta SC, Tyagi AK, Deshmukh-Taskar P, Hinojosa M, Prasad S, Aggarwal BB. Downregulation of tumor necrosis factor and other proinflammatory biomarkers by polyphenols. *Arch Biochem Biophys*. 2014 Oct 1; 559:91–9. <https://doi.org/10.1016/j.abb.2014.06.006> PMID: 24946050
34. Lee KW, Kang NJ, Heo YS, Rogozin EA, Pugliese A, Hwang MK, et al. Raf and MEK protein kinases are direct molecular targets for the chemopreventive effect of quercetin, a major flavonol in red wine. *Cancer Res*. 2008 Feb 1; 68(3):946–55. <https://doi.org/10.1158/0008-5472.CAN-07-3140> PMID: 18245498
35. Youn HS, Lee JY, Saitoh SI, Miyake K, Kang KW, Choi YJ, et al. Suppression of MyD88- and TRIF-dependent signaling pathways of Toll-like receptor by (-)-epigallocatechin-3-gallate, a polyphenol component of green tea. *Biochem Pharmacol*. 2006 Sep 28; 72(7):850–9. <https://doi.org/10.1016/j.bcp.2006.06.021> PMID: 16890209
36. Xagorari A, Papapetropoulos A, Mauromatis A, Economou M, Fotsis T, Roussos C. Luteolin inhibits an endotoxin-stimulated phosphorylation cascade and proinflammatory cytokine production in macrophages. *J Pharmacol Exp Ther*. 2001 Jan; 296(1):181–7. PMID: 11123379
37. Nicholas C, Batra S, Vargo MA, Voss OH, Gavrilin MA, Wewers MD, et al. Apigenin blocks lipopolysaccharide-induced lethality in vivo and proinflammatory cytokines expression by inactivating NF-kappaB through the suppression of p65 phosphorylation. *J Immunol*. 2007 Nov 15; 179(10):7121–7. PMID: 17982104
38. Prasad S, Phromnoi K, Yadav VR, Chaturvedi MM, Aggarwal BB. Targeting inflammatory pathways by flavonoids for prevention and treatment of cancer. *Planta Med*. 2010 Aug; 76(11):1044–63. <https://doi.org/10.1055/s-0030-1250111> PMID: 20635307
39. Rahimifard M, Maqbool F, Moeini-Nodeh S, Niaz K, Abdollahi M, Braidy N, et al. Targeting the TLR4 signaling pathway by polyphenols: A novel therapeutic strategy for neuroinflammation. *Ageing Res Rev*. 2017 Jul; 36:11–19. <https://doi.org/10.1016/j.arr.2017.02.004> PMID: 28235660
40. Burnstock G, Boeynaems JM. Purinergic signalling and immune cells. *Purinergic Signal*. 2014 Dec; 10(4):529–64. <https://doi.org/10.1007/s11302-014-9427-2> PMID: 25352330
41. Antonioli L, Pacher P, Vizi ES, Haskó G. CD39 and CD73 in immunity and inflammation. *Trends Mol Med*. 2013 Jun; 19(6):355–67. <https://doi.org/10.1016/j.molmed.2013.03.005> PMID: 23601906
42. Lowenstein CJ, Alley EW, Raval P, Snowman AM, Snyder SH, Russell SW, et al. Macrophage nitric oxide synthase gene: two upstream regions mediate induction by interferon gamma and lipopolysaccharide. *Proc Natl Acad Sci USA*. 1993 Oct 15; 90(20):9730–4. PMID: 7692452
43. Farlik M, Reutterer B, Schindler C, Greten F, Vogl C, Müller M, et al. Nonconventional initiation complex assembly by STAT and NF-kappaB transcription factors regulates nitric oxide synthase expression. *Immunity*. 2010 Jul 23; 33(1):25–34. <https://doi.org/10.1016/j.immuni.2010.07.001> PMID: 20637660
44. Sakaguchi S, Negishi H, Asagiri M, Nakajima C, Mizutani T, Takaoka A, et al. Essential role of IRF-3 in lipopolysaccharide-induced interferon-beta gene expression and endotoxin shock. *Biochem Biophys Res Commun*. 2003 Jul 11; 306(4):860–6. PMID: 12821121
45. Coddou C, Yan Z, Obsil T, Huidobro-Toro JP, Stojilkovic SS. Activation and regulation of purinergic P2X receptor channels. *Pharmacol Rev*. 2011 Sep; 63(3):641–83. <https://doi.org/10.1124/pr.110.003129> PMID: 21737531
46. Jacobson KA, Müller CE. Medicinal chemistry of adenosine, P2Y and P2X receptors. *Neuropharmacology*. 2016 May; 104:31–49. <https://doi.org/10.1016/j.neuropharm.2015.12.001> PMID: 26686393
47. Cekic C, Linden J. Purinergic regulation of the immune system. *Nat Rev Immunol*. 2016 Mar; 16(3):177–92. <https://doi.org/10.1038/nri.2016.4> PMID: 26922909

48. Junger WG. Immune cell regulation by autocrine purinergic signalling. *Nat Rev Immunol.* 2011 Mar; 11(3):201–12. <https://doi.org/10.1038/nri2938> PMID: 21331080
49. Feske S, Skolnik EY, Prakriya M. Ion channels and transporters in lymphocyte function and immunity. *Nat Rev Immunol.* 2012 Jun 15; 12(7):532–47. <https://doi.org/10.1038/nri3233> PMID: 22699833
50. Gough DJ, Messina NL, Clarke CJ, Johnstone RW, Levy DE. Constitutive type I interferon modulates homeostatic balance through tonic signaling. *Immunity.* 2012 Feb 24; 36(2):166–74. <https://doi.org/10.1016/j.immuni.2012.01.011> PMID: 22365663
51. Gendron FP, Neary JT, Theiss PM, Sun GY, Gonzalez FA, Weisman GA. Mechanisms of P2X7 receptor-mediated ERK1/2 phosphorylation in human astrocytoma cells. *Am J Physiol Cell Physiol.* 2003 Feb; 284(2):C571–81. <https://doi.org/10.1152/ajpcell.00286.2002> PMID: 12529254
52. Tsao HK, Chiu PH, Sun SH. PKC-dependent ERK phosphorylation is essential for P2X7 receptor-mediated neuronal differentiation of neural progenitor cells. *Cell Death Dis.* 2013 Aug 1; 4:e751. <https://doi.org/10.1038/cddis.2013.274> PMID: 23907465
53. da Cruz CM, Ventura AL, Schachter J, Costa-Junior HM, da Silva Souza HA, Gomes FR, et al. Activation of ERK1/2 by extracellular nucleotides in macrophages is mediated by multiple P2 receptors independently of P2X7-associated pore or channel formation. *Br J Pharmacol.* 2006 Feb; 147(3):324–34. <https://doi.org/10.1038/sj.bjp.0706559> PMID: 16341234
54. Jiang LH, Mackenzie AB, North RA, Surprenant A. Brilliant blue G selectively blocks ATP-gated rat P2X(7) receptors. *Mol Pharmacol.* 2000 Jul; 58(1):82–8. PMID: 10860929
55. Pelegrin P, Barroso-Gutierrez C, Surprenant A. P2X7 receptor differentially couples to distinct release pathways for IL-1beta in mouse macrophage. *J Immunol.* 2008 Jun 1; 180(11):7147–57. PMID: 18490713
56. Bours MJ, Swennen EL, Di Virgilio F, Cronstein BN, Dagnelie PC. Adenosine 5'-triphosphate and adenosine as endogenous signaling molecules in immunity and inflammation. *Pharmacol Ther.* 2006 Nov; 112(2):358–404. <https://doi.org/10.1016/j.pharmthera.2005.04.013> PMID: 16784779
57. Chen GY, Nuñez G. Sterile inflammation: sensing and reacting to damage. *Nat Rev Immunol.* 2010 Dec; 10(12):826–37. <https://doi.org/10.1038/nri2873> PMID: 21088683
58. Colasanti M, Suzuki H. The dual personality of NO. *Trends Pharmacol Sci.* 2000 Jul; 21(7):249–52. PMID: 10979862
59. Moncada S, Higgs A, Furchgott R. International Union of Pharmacology Nomenclature in Nitric Oxide Research. *Pharmacol Rev.* 1997 Jun; 49(2):137–42. PMID: 9228663
60. Bogdan C. Nitric oxide and the immune response. *Nat Immunol.* 2001 Oct; 2(10):907–16. <https://doi.org/10.1038/ni1001-907> PMID: 11577346
61. Lu YC, Yeh WC, Ohashi PS. LPS/TLR4 signal transduction pathway. *Cytokine.* 2008 May; 42(2):145–51. <https://doi.org/10.1016/j.cyto.2008.01.006> PMID: 18304834
62. Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. *Nat Immunol.* 2010 May; 11(5):373–84. <https://doi.org/10.1038/ni.1863> PMID: 20404851
63. Gavala ML, Liu YP, Lenertz LY, Zeng L, Blanchette JB, Guadarrama AG, et al. Nucleotide receptor P2RX7 stimulation enhances LPS-induced interferon-β production in murine macrophages. *J Leukoc Biol.* 2013 Oct; 94(4):759–68. <https://doi.org/10.1189/jlb.0712351> PMID: 23911869
64. Fujimoto Y, Nakatani N, Kubo T, Semi Y, Yoshida N, Nakajima H, et al. Adenosine and ATP affect LPS-induced cytokine production in canine macrophage cell line DH82 cells. *J Vet Med Sci.* 2012 Jan; 74(1):27–34. PMID: 21873810
65. Kim M, Jiang LH, Wilson HL, North RA, Surprenant A. Proteomic and functional evidence for a P2X7 receptor signalling complex. *EMBO J.* 2001 Nov 15; 20(22):6347–58. <https://doi.org/10.1093/emboj/20.22.6347> PMID: 11707406
66. Costa-Junior HM, Marques-da-Silva C, Vieira FS, Monção-Ribeiro LC, Coutinho-Silva R. Lipid metabolism modulation by the P2X7 receptor in the immune system and during the course of infection: new insights into the old view. *Purinergic Signal.* 2011 Dec; 7(4):381–92. <https://doi.org/10.1007/s11302-011-9255-6> PMID: 21845440
67. Costa-Junior HM, Sarmiento Vieira F, Coutinho-Silva R. C terminus of the P2X7 receptor: treasure hunting. *Purinergic Signal.* 2011 Mar; 7(1):7–19. <https://doi.org/10.1007/s11302-011-9215-1> PMID: 21484094
68. North RA, Jarvis MF. P2X receptors as drug targets. *Mol Pharmacol.* 2013 Apr; 83(4):759–69. <https://doi.org/10.1124/mol.112.083758> PMID: 23253448
69. Bartlett R, Stokes L, Sluyter R. The P2X7 receptor channel: recent developments and the use of P2X7 antagonists in models of disease. *Pharmacol Rev.* 2014 Jul; 66(3):638–75. <https://doi.org/10.1124/pr.113.008003> PMID: 24928329

70. Mehta N, Kaur M, Singh M, Chand S, Vyas B, Silakari P, et al. Purinergic receptor P2X₇: a novel target for anti-inflammatory therapy. *Bioorg Med Chem*. 2014 Jan 1; 22(1):54–88. <https://doi.org/10.1016/j.bmc.2013.10.054> PMID: 24314880
71. Zhao L, Lee JY, Hwang DH. Inhibition of pattern recognition receptor-mediated inflammation by bioactive phytochemicals. *Nutr Rev*. 2011 Jun; 69(6):310–20. <https://doi.org/10.1111/j.1753-4887.2011.00394.x> PMID: 21631512
72. Sato S, Sugiyama M, Yamamoto M, Watanabe Y, Kawai T, Takeda K, et al. Toll/IL-1 receptor domain-containing adaptor inducing IFN-beta (TRIF) associates with TNF receptor-associated factor 6 and TANK-binding kinase 1, and activates two distinct transcription factors, NF-kappa B and IFN-regulatory factor-3, in the Toll-like receptor signaling. *J Immunol*. 2003 Oct 15; 171(8):4304–10. PMID: 14530355
73. Endale M, Park SC, Kim S, Kim SH, Yang Y, Cho JY, et al. Quercetin disrupts tyrosine-phosphorylated phosphatidylinositol 3-kinase and myeloid differentiation factor-88 association, and inhibits MAPK/AP-1 and IKK/NF-kB-induced inflammatory mediators production in RAW 264.7 cells. *Immunobiology*. 2013 Dec; 218(12):1452–67. <https://doi.org/10.1016/j.imbio.2013.04.019> PMID: 23735482
74. Li-Weber M. New therapeutic aspects of flavones: the anticancer properties of Scutellaria and its main active constituents Wogonin, Baicalein and Baicalin. *Cancer Treat Rev*. 2009 Feb; 35(1):57–68. <https://doi.org/10.1016/j.ctrv.2008.09.005> PMID: 19004559
75. He X, Wei Z, Zhou E, Chen L, Kou J, Wang J, et al. Baicalein attenuates inflammatory responses by suppressing TLR4 mediated NF-kB and MAPK signaling pathways in LPS-induced mastitis in mice. *Int Immunopharmacol*. 2015 Sep; 28(1):470–6. <https://doi.org/10.1016/j.intimp.2015.07.012> PMID: 26202808
76. Ishisaka A, Kawabata K, Miki S, Shiba Y, Minekawa S, Nishikawa T, et al. Mitochondrial dysfunction leads to deconjugation of quercetin glucuronides in inflammatory macrophages. *PLoS One*. 2013 Nov 19; 8(11):e80843. <https://doi.org/10.1371/journal.pone.0080843> PMID: 24260490
77. Chen CY, Jang JH, Li MH, Surh YJ. Resveratrol upregulates heme oxygenase-1 expression via activation of NF-E2-related factor 2 in PC12 cells. *Biochem Biophys Res Commun*. 2005 Jun 17; 331(4):993–1000. <https://doi.org/10.1016/j.bbrc.2005.03.237> PMID: 15882976
78. Na HK, Kim EH, Jung JH, Lee HH, Hyun JW, Surh YJ. (-)-Epigallocatechin gallate induces Nrf2-mediated antioxidant enzyme expression via activation of PI3K and ERK in human mammary epithelial cells. *Arch Biochem Biophys*. 2008 Aug 15; 476(2):171–7. <https://doi.org/10.1016/j.abb.2008.04.003> PMID: 18424257
79. Zhao M, Chen J, Zhu P, Fujino M, Takahara T, Toyama S, et al. Dihydroquercetin (DHQ) ameliorated concanavalin A-induced mouse experimental fulminant hepatitis and enhanced HO-1 expression through MAPK/Nrf2 antioxidant pathway in RAW cells. *Int Immunopharmacol*. 2015 Oct; 28(2):938–44. <https://doi.org/10.1016/j.intimp.2015.04.032> PMID: 25916679
80. Lin CW, Wu MJ, Liu IY, Su JD, Yen JH. Neurotrophic and cytoprotective action of luteolin in PC12 cells through ERK-dependent induction of Nrf2-driven HO-1 expression. *J Agric Food Chem*. 2010 Apr 14; 58(7):4477–86. <https://doi.org/10.1021/jf904061x> PMID: 20302373
81. Bhullar KS, Hubbard BP. Lifespan and healthspan extension by resveratrol. *Biochim Biophys Acta*. 2015 Jun; 1852(6):1209–18. <https://doi.org/10.1016/j.bbadis.2015.01.012> PMID: 25640851
82. Chuah YK, Basir R, Talib H, Tie TH, Nordin N. Receptor for advanced glycation end products and its involvement in inflammatory diseases. *Int J Inflam*. 2013 Sep 11; 2013:403460. <https://doi.org/10.1155/2013/403460> PMID: 24102034
83. Gasic-Milenkovic J, Dukic-Stefanovic S, Deuther-Conrad W, Gärther U, Münch G. β -Amyloid peptide potentiates inflammatory responses induced by lipopolysaccharide, interferon- γ and 'advanced glycation endproducts' in a murine microglia cell line. *Eur J Neurosci*. 2003 Feb; 17(4):813–21. PMID: 12603271