

1 **Thermophilic and mesophilic temperature phase anaerobic co-digestion (TPAcD)**
2 **compared with single-stage co-digestion of sewage sludge and sugar beet pulp**
3 **lixiviation.**

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10

11 **Abstract**

12 The performance of temperature phase anaerobic co-digestion (TPAcD) for sewage
13 sludge and sugar beet pulp lixiviation (using the exchange process of the digesting
14 substrate between spatially separated thermophilic and mesophilic digesters) was
15 tested and compared to single-stage mesophilic and thermophilic anaerobic co-
16 digestions. The volatile solids removal efficiency from the TPAcD system was
17 dependent on the sludge exchange rate, but was in the range 72.6–64.6%, which was
18 higher than 46.8% with single-stage thermophilic digestion as well as 40.5% with
19 mesophilic digestion. The specific methane yield was 424-468 ml CH₄ per gram volatile
20 solids removed and similar to single-stage mesophilic anaerobic digestion. The increase
21 in microbial activity inside the reactor was directly proportional to the organic loading
22 rate (OLR) (or inversely proportional to the HRT) and inversely proportional to the size
23 of the microbial population in single-stage anaerobic co-digestion systems.

24

25 **Keywords:** Two-phase anaerobic co-digestion, sewage sludge, sugar beet pulp
26 lixiviation, microbial activity

27

28 **1. Introduction**

29 Single-stage mesophilic completely mixed anaerobic digestion has been widely used
30 for the reduction in volume of organic sludge from wastewater treatment processes
31 and for obtaining energy in the form of methane gas. Mesophilic digestion with
32 sewage sludge usually requires over a 20-day retention time, but it is not so efficient in
33 the reduction of volatile solids and the deactivation of pathogenic organisms. To
34 overcome these limitations, interest in thermophilic digestion and co-digestion has
35 increased in recent years.

36 Co-digestion is the simultaneous anaerobic digestion of a mixture of two or more
37 substrates. The technology is similar to anaerobic digestion, but it is an attractive
38 option due to the increase in methane yields, because of the positive synergism
39 established in the digestion medium. This fact that increases the economic viability of
40 biogas plants [1].

41 Co-digestion technology could lead to the following benefits [1, 2]: (1) dilution of
42 inhibitory and/or toxic compounds, (2) increase of the organic content inside the
43 digester, with better utilization of the digester volume, (3) enhancement of digestate
44 stabilization, (4) accomplishment of the required moisture content in the digester
45 feed, with easier handling of blended wastes, (5) a greater reduction in the emission of
46 greenhouse gases to the atmosphere and (6) economic advantages from sharing
47 equipment and costs. However, some drawbacks exist as well: (1) the high cost of
48 waste transfer from the cosubstrate generation point to the anaerobic plant, and (2)
49 the harmonization of different policies regarding waste generators.

50 To overcome the limitations of mesophilic digestion, interest in thermophilic digestion,
51 using the higher metabolic rate of thermophilic microorganisms, has increased [3-5].

52 Although better performance in the reduction of volatile solids and the deactivation of
53 pathogenic organisms can be obtained from thermophilic digestion, the effluent
54 quality and ability to dewater the residual sludge are poor, and require additional
55 energy to heat the digester [4, 6]. Especially, thermophilic digestion is not much more
56 sensitive to operational conditions, such as temperature, and the organic loading rate,
57 as well as to the characteristics of the influent sludge [6, 7]. Generally, anaerobic
58 processes can be characterized by the digestion environment, microorganisms and
59 process configuration, and each process has its unique advantages.

60 According to previous studies [8, 9], two-phase or two-stage anaerobic processes have
61 shown good performance in terms of effluent quality, methane yield, volatile solids
62 reduction and process stability. This implies that the performance of an anaerobic
63 process could be improved with the proper combination of anaerobic process
64 characteristics. Recently, the temperature phased anaerobic digestion (TPAD) process,
65 which consists of thermophilic and mesophilic digesters in series, has been studied in
66 order to incorporate the advantages of both thermophilic and mesophilic digestion
67 [10-12]. The TPAD process can be operated at higher loading rates compared to single-
68 stage processes [11, 13] and is better for the deactivation of pathogenic organisms [13]
69 and in its ability to absorb shock loadings, like other two-stage or two phase anaerobic
70 processes [14].- The first-stage of the TPAD process is sensitive to environmental
71 conditions, and has a notable influence on the overall TPAD process. In addition, the
72 degree of maximum volatile solids reduction and specific methane yield obtainable
73 from the TPAD process are not much different from that of single-stage anaerobic
74 processes with sufficient solid retention time [11]. Recently, phased anaerobic

75 digestion systems have gained attention as a sustainable technology for sludge
76 digestion and methane production [15].

77 Although large-scale TPACD systems have not been applied widely, researchers have
78 demonstrated the potential superiority of TPAD systems over single-stage digesters
79 and other AD processes. Improved total volatile solids (TVS) and pathogen removal,
80 increased methane yield, process stability and organic loading rate (OLR), a shorter
81 hydraulic retention time (HRT), as well as decreased foaming and short-chain fatty
82 acids in the effluent are some of the positive aspects of anaerobic co-digestion.

83 Although the determination of the number of microorganisms is important in many
84 microbial ecology studies [16], these studies have not assessed the activities
85 associated with the methanogen population. Microbial activity will correlate with
86 number only as long as the environmental conditions remain constant. Any change in
87 substrate and operating conditions in the reactors will alter these parameters.

88 Microbial number and activity represent distinct ecological parameters.

89 The stability of the system depends on the viable bacterial groups, and HRT is a
90 significant factor in selecting the predominant microbial species [17, 18].

91 Understanding the functioning of anaerobic reactors requires quantitative information
92 on microbial numbers, biomass and activities of the bacterial groups involved in the
93 process. The measurement of biomass as volatile solids is a significant limitation in
94 studies on the kinetics of the process, development, operation and monitoring of
95 reactors. Direct count procedures by microscopy methods yield the highest estimates
96 of members of micro-organisms and are occasionally used for an indirect calculation of
97 biomass. Epifluorescence microscopy is widely used for direct counting of bacteria,
98 since it does not require culturing [19]. A characteristic peculiarity of methanogens is

99 their UV-induced blue-green autofluorescence which permits counting by
100 autofluorescence microscopy [20]. However, this method is subjective: it only shows
101 methanogens with a high content of F420 such as hydrogen-utilizing methanogens;
102 acetate-utilizing methanogens belonging to the genus Methanosaeta cannot be
103 counted at all and the genus Methanosarcina is found in clumps made up of many
104 individual cells. Nevertheless, it is a frequently used method to count autofluorescent
105 methanogens in anaerobic reactors [18, 21].

106 The aim of this research was to test the configuration of anaerobic co-digestion, using
107 a temperature phased anaerobic co-digestion (TPAcD) process which consists of a
108 thermophilic digester followed by a mesophilic one, to improve the efficiency of single
109 phase anaerobic co-digestion of sewage sludge and sugar beet pulp lixiviation. Thus,
110 the performance of the single-stage completely mixed thermophilic and mesophilic
111 digestions were examined and their characteristics compared with the results obtained
112 in temperature phase anaerobic co-digestion. Mesophilic and thermophilic single-stage
113 anaerobic co-digestion for sewage sludge and sugar beet pulp lixiviation were
114 compared between them.

115 Relationships between OLR, methane generation and both methanogenic anaerobic
116 micro-organisms and the activity of those microorganisms were also considered.

117 The most important novelty of the data presented in this study is the direct
118 experimental evidence regarding the influence of HRT on the population levels of
119 methanogenic anaerobic micro-organisms in the digester.

120 **Notations**

121 **AD** Anaerobic digestion

122 **AcD** Anaerobic co-digestion

- 123 **COD** Soluble carbon oxygen demand
- 124 **CSTR** Continuous stirred-tank reactor
- 125 **HRT** Hydraulic retention time
- 126 **ORL** Organic loading rate
- 127 **SBPL** Sugar beet pulp lixiviation
- 128 **SS** Sewage sludge
- 129 **TPAcD** Two phase anaerobic co-digestion
- 130 **TPAD** Two phase anaerobic digestion
- 131 **TS** Total solids
- 132 **TVS** Total volatile solids
- 133 **VFA** Volatile fatty acids
- 134 **WWTP** Waste water treatment plant

135 **2. Materials and methods**

136 **2.1. Experimental process**

137 The schematic diagrams of the anaerobic co-digestion systems used for the
138 experiments are shown in Figure 1. For the temperature co-phase anaerobic co-
139 digestion system, (a) the lab-scale system consisted of a 6 l thermophilic reactor (5 l
140 working volume) followed by a 6 l mesophilic digester (5 l working volume). Both
141 experimental digesters shared similar characteristics: the cover of each reactor
142 incorporated three separate ports for different functions: feeding, mechanical
143 agitation, and measurement of biogas generation (using a 10 l Tedlar bag). The
144 reactors were kept at the selected temperature by water circulating in the water jacket
145 surrounding the reactors.

146 For the single-stage anaerobic co-digestion systems, semi-continuous laboratory-scale
147 stirred tank reactors were used (Figure 1b). The equipment consisted of a reactor with
148 a stainless steel vessel that was agitated and heated. The total volume was 6 l and the
149 working volume was 5 l. ~~To maintain the operating temperature, the digesters were~~
150 ~~heated by recirculating water through a thermostatic jacket.~~ Biogas was collected in
151 10-l Tedlar bags, and a special syringe was used for sampling the gases.

152 Six tests were developed (Table 1).

153 In TPACD systems, the thermophilic digester was fed with a mix of sewage sludge and
154 sugar beet pulp lixiviation (50-50 w/w) and the mesophilic digester was fed with the
155 effluent generated in the previous thermophilic digester.

156 Two TPACD experiments were carried out. The first-stage thermophilic (55°C) digester
157 was operated at 10 and 6 days of retention time, respectively; its effluent was used to
158 provide feed for the second-stage mesophilic (35°C) digester. The second-stage
159 digester was operated at an HRT of 10 days in both cases. Therefore, two HRT
160 combinations were assayed: 20 (TPACD10/10) and 16 (TPACD6/10). Each condition was
161 maintained for an operational period lasting three times the duration of the HRT to
162 ensure that steady state conditions were reached by checking whether constant
163 effluent characteristic values (carbon oxygen demand soluble (COD), total solids (TS),
164 total volatile solids (TVS), gas production and composition, volatile fatty acids (VFA)
165 and alkalinity levels. Sampling during each steady-state period was performed for five
166 consecutive days.

167 **2.2. Anaerobic inocula and substrates**

168 The digester was initially loaded with a mixture of inoculum and substrate, resulting in
169 a final concentration of 20% w/w of inoculum, which is considered optimum for biogas
170 production [22].

171 Primary sludge from the WWTP of San Fernando-Cádiz was used as the inoculum in the
172 mesophilic reactor. The mixed anaerobic culture used as the thermophilic inoculum of
173 the CSTR reactor was obtained from a lab digester running at 20 days of HRT. The
174 inoculum was obtained through a direct change from mesophilic (35°C) to
175 thermophilic conditions (55°C), as described by Riau et al. (2010). The characteristics of
176 the inoculum used in the start-up process are presented in Table 2. The substrate was
177 composed of sewage sludge and sugar beet pulp lixiviation.

178 **Sewage sludge:** The digesters were fed with sewage sludge collected from the
179 aforementioned WWTP.

180 **Lixiviation of sugar beet pulp:** Pellets were collected from Azucarera Ebro Company in
181 Jerez de la Frontera (Cádiz). Sugar beet pulp used as the co-substrate was subjected to
182 physical pretreatment before the co-digestion process in order to promote hydrolysis
183 and solubilization of the organic matter and, therefore, improve anaerobic digestion in
184 the generation of biogas and enhance the final residue's agronomic valorization [22].

185 Once the inoculum was mixed with the substrate, i.e. a mixture of sewage sludge and
186 lixiviation of sugar beet pulp, the system remained unfed for a period of one week to
187 acclimatize the inoculum to the waste at the selected temperatures (35 and 55°C).

188 The average feeding compositions for each reactor in all experiments carried out are
189 summarized in Table 3.

190 Initially, the organic loading rate (OLR) applied to the single-stage thermophilic and
191 mesophilic reactors were 1.2 and 2.1 g TVS/l/d for T20-M20 and T10-M10,
192 respectively.

193 For TPACD, the initial OLR applied were 2.2 and 2.5 g TVS/l/d for TPACD10/10 and
194 TPACD6/10, respectively.

195 These conditions were maintained until steady state conditions were reached.

196 **2.4. Chemical and microbial analyses**

197 The volume and composition of biogas were determined daily. The biogas produced
198 was quantified using a gas flow meter (Ritter TG1) and a gas suction pump (KNF
199 Laboport). Gas chromatography was used to analyze the different components of the
200 biogas. The gases analyzed were: H₂, CH₄, CO₂, O₂ and N₂ (GC-2010 Shimadzu).

201 The following analytical determinations were performed to monitor and control the
202 process in the substrate and the effluent: TS, TVS, pH, soluble COD, alkalinity and VFA
203 (acetic, propionic, iso-butyric, butyric, iso-valeric, valeric, iso-caproic, caproic and
204 heptanoic). The pH was measured daily using a Crison 20 Basic pH meter. TVS, COD
205 and VFA were analyzed three times a week. These determinations were performed
206 according to APHA (1995) [23]. Organic matter removal was calculated as the
207 percentage difference between the TVS of the influent and the TVS of the effluent
208 within the substrate TVS. Total acidity was calculated by addition of the individual fatty
209 acids.

210 Quantification assays were performed when reactors reached steady-state conditions.

211 The attainment of the steady state was verified after an initial period (three times the
212 HRT) by checking whether the effluent characteristic values continued at the mean of
213 the previous measurements. The autofluorescent methanogens in the reactors were

214 counted by autofluorescence microscopy. The experimental protocol was performed
215 according to Solera et al., 2001 [18].

216 **3. Results and discussion**

217 The operational conditions were applied to reactors in the mesophilic range (M20 and
218 M10), in the thermophilic range (T20 and T10), and temperature phased (TPAcD10/10
219 and TPAcD6/10). The operational conditions are presented in the Table 4
220 Table 5 shows Effluent quality and performance of the single-stage mesophilic and
221 thermophilic co-digestion processes. The data shown the results of the stability period
222 for each HRT studied.

223 **3.1. Single-stage mesophilic and thermophilic digestion**

224 During the operation time of the single-stage anaerobic processes, the alkalinity level
225 of the thermophilic digestion process was higher than that of the mesophilic process,
226 as shown in Figure 2b. It is well-known that alkalinity in an anaerobic digestion can be
227 generated from the degradation of nitrogenous organic compounds, sulfate reduction,
228 the release of orthophosphate and an increase in VFAs [24, 25]. In this study, the
229 ammonia nitrogen from the thermophilic digestion process was 808 mg/l, which was
230 higher than the 707 mg/l in the mesophilic process at 20 days of HRT. The same
231 behavior was observed at 10 days of HRT (Table 5).

232 The pH value of the effluent substrates gradually decreased between 20 days HRT and
233 10 in both temperature regimes, as shown in Figure 2b. Although, the pH values of the
234 mesophilic process at 20 and 10 days HRT were below 7.5, the digestion showed good
235 behavior and was stable at this value. The pH of the thermophilic process was
236 generally higher than that of the mesophilic process. This was a result of the higher
237 alkalinity of the thermophilic anaerobic digestion process. The increased alkalinity and

238 thus pH from the degradation of nitrogenous compounds in our experiments is in
239 agreement with previous studies [26].

240 The COD level of the thermophilic process was much higher than that in the mesophilic
241 process, as shown in Table 5. At the steady state, the mean values of soluble COD were
242 4.9 and 1.6 kg/m³ for the thermophilic and mesophilic processes, respectively (Table 5)
243 for the optimum hydraulic retention time (10 days). The VFA level in the thermophilic
244 process was generally higher than that in the mesophilic process, which was consistent
245 with the COD data (Figure 2a). This clearly shows that mesophilic digestion was
246 superior to thermophilic digestion in terms of the effluent quality, which can be
247 explained by the low substrate affinity of some thermophilic organisms [4, 6, 7].

248 The main component of VFA in the mesophilic and thermophilic processes was
249 acetate, but in the thermophilic process at 20-days of HRT, propionate was present at a
250 very high value (Figure 2e). Based on the literature [6, 7], the higher level of
251 propionate in the thermophilic digester occurred due to the higher hydrogen partial
252 pressure, and the acetate was from the higher organic loading rate conditions. In this
253 study, the accumulation of propionate in the thermophilic digester was probably due
254 to the wide fluctuation in the influent characteristics. This indicates that acetogens and
255 hydrogenotrophs under thermophilic conditions are more sensitive to environmental
256 changes. At 10 days HRT, the thermophilic process was able to compensate for the
257 variations in feeding because it was working with an optimum organic loading rate.

258 The VFA to alkalinity ratio for the four single-stage anaerobic systems were monitored
259 to compare the buffering capacities for the change in pH (Figure 2.a). It has been
260 reported that the buffering capacity is sufficient when the VFA-to-alkalinity ratio is
261 maintained below 0.4 [10]. In this study, this ratio in the mesophilic process was below

262 0.1. For the thermophilic anaerobic digestion process, this ratio was a little higher in
263 both HRTs studied in this work. The slightly higher VFA-to-alkalinity ratio of the
264 thermophilic process was primarily a result of the higher VFA concentration. This
265 indicates that single-stage mesophilic anaerobic co-digestion had better buffering
266 capabilities than thermophilic co-digestion.

267 The performance of the digesters with respect to solids removal for different tests is
268 presented in Table 5 and Figure 2d. For single-stage reactors, thermophilic conditions
269 resulted in higher removal than the corresponding mesophilic operated reactors.

270 There was a noticeable increase in terms of volatile solids removal when the reactor
271 temperature was raised, with removal rates increasing from 40.5 to 76.5% for 10 days
272 of HRT. For a longer retention time (20 days HRT), the difference between the
273 mesophilic and thermophilic regimes was lower since at this HRT the bacteria in the
274 mesophilic range are capable of biodegrading all biodegradable solids, although 20
275 days is not the optimum retention time. Maibaum and Kuehn (1999) [4] reported that
276 the difference in the degradation rates of solids substrates under thermophilic and
277 mesophilic conditions becomes significant in relation to the decrease in the retention
278 time.

279 As shown in Table 5, the average methane content of the biogas from the mesophilic
280 process was higher, at around 70%, than that of the thermophilic process. This was
281 probably a result of the reduced solubility of carbon dioxide under thermophilic
282 conditions [26]. In previous studies, the methane content of the biogas was mainly
283 affected by the type of substrate, rather than the temperature conditions, for
284 anaerobic digestion [5, 26]. However, the specific methane yield of the mesophilic
285 process, based on the removed VS, was a little more sensitive to the influent

286 characteristics of feeding, indicating a higher capacity of mesophilic methanogens for
287 coping with variations in influent characteristics compared to thermophilic
288 methanogens. The average specific methane yield of the thermophilic process was
289 lower, at 210 ml CH₄/gTVS_{removal}, than the 630 ml CH₄/gTVS_{removal} by the mesophilic
290 digester for the optimum retention time (Table 5). This was presumably due to the
291 higher maintenance energy of the anaerobic thermophilic microorganisms [6, 7], as
292 well as the higher hydrogen content of the biogas [26]. In comparison with the
293 thermophilic reactor, the mesophilic reactor produced a greater quantity of methane
294 per gram of TVS destroyed at the optimum HRT. This suggests that the thermophilic
295 reactor was not efficient in converting all the intermediate products to methane.
296 The biomethanation process involves stepwise degradations of complex biomass by
297 diverse microbial populations that interact with each other. Four guilds of microbes,
298 which include hydrolytic acidogens, non-hydrolytic acidogens, syntrophic acetogens,
299 and methanogens, drive the biomethanation process in a sequential and concerted
300 manner.

301 After the analysis of the single-stage anaerobic co-digestion of sewage sludge and
302 sugar beet pulp lixiviation, the condition employing 10 days as the hydraulic retention
303 time in the mesophilic regime were determined to be the best option. Once this HRT
304 was chosen, the next goal was to compare this optimum with two-phase anaerobic
305 digestion technology.

306 **3.2. The thermophilic and mesophilic co-phase anaerobic digestion**

307 An increase in biogas production was observed in the TPACD10/10 process: biogas
308 generation increased from 1.70 l/d under thermophilic conditions for 10 days of HRT to
309 3.42 l/d under mesophilic conditions to 3.59 l/d in the temperature phased system.
310 The alkalinity levels of the temperature co-phase thermophilic and mesophilic
311 digesters were influenced by the variation in the alkalinity of the influent substrate, as
312 shown in Figure 3a. The average level of alkalinity in the co-phase thermophilic
313 digester was around 3400–5300 mg/l as CaCO₃, which was higher than 3200–3800
314 mg/l as CaCO₃ in the co-phase mesophilic digester. The greater alkalinity under
315 thermophilic conditions was similar to that of the single-stage anaerobic processes, as
316 shown in Figure 2b, and reflects the higher degradation activity toward nitrogenous
317 organic compounds, such as proteins, under thermophilic conditions [26]. The pH
318 levels of the co-phase thermophilic and mesophilic digesters in TPACD10/10 were in
319 the range of the methanogenic process; nevertheless, the pH in TPACD6/10 decreased
320 to below 7 in the thermophilic range due to VFA accumulation. In the first TPACD test,
321 the pH levels in the mesophilic and thermophilic digesters were similar to those in the
322 single-stage mesophilic and thermophilic anaerobic processes.

323 The influence of the substrate exchange rate on the pH of the TPACD system was not
324 observed in the first assay. However, when the HRT in the thermophilic phase was
325 decreased, the accumulation of VFA occurred, causing a decrease in pH in the
326 thermophilic digester and, in addition, a fault in the mesophilic reactor. Therefore,
327 with the substrates used in those test the optimum TPACD system is TPACD 10/10
328 where thermophilic reactor is a pretreatment of the mesophilic anaerobic co-
329 digestion.

330 In TPACD10/10, the VFA values in the co-phase thermophilic digester became stable
331 after the operation time, as well as that of the mesophilic digester, and were not
332 influenced by the wide change in the influent characteristics. At the steady state, the
333 VFA value in the co-phase mesophilic digester was 537 mg ACH/l, which was lower
334 than 761 mg ACH/l found in the mesophilic digester. This indicates that the mesophilic
335 digester of the co-phase system was stable and functioned well. The affinity of the
336 thermophilic substrate for VFA was quite a bit lower than that of the feeding from the
337 single-stage mesophilic digester (Table 5). This seems to suggest that the higher
338 substrate affinity methanogenic bacteria were selected and dominated in the co-phase
339 mesophilic digester by the substrate exchange between the thermophilic and
340 mesophilic digesters. In the case of the co-phase thermophilic digester, the VFA value
341 was slightly higher than that of the single-stage thermophilic digester.

342 In the TPACD6/10 test, an accumulation of VFA in the thermophilic digester occurred
343 because of the reduced HRT. Due to this circumstance, anaerobic co-digestion in the
344 mesophilic digester failed.

345 The main VFA component of the co-phase mesophilic digester was acetate, as in the
346 single-stage mesophilic process (Figure 2 and Figure 3). However, in the co-phase
347 thermophilic digester, the propionate content was considerable at both HRTs. This
348 higher propionate content (Table 6) at a higher substrate exchange rate in the co-
349 phase thermophilic digester was probably related to the higher hydrogen partial
350 pressure [6, 26].

351 Individual and total VFA concentrations in the effluent of the first-stage reactor
352 increased when the total HRT decreased in each assay. This indicates that the HRT of
353 the thermophilic phase is a more important factor affecting the VFA content.

354 The reduction of HRT in thermophilic reactor of the TPACD process and the subsequent
355 VFA accumulation conditioned the pH of the digester (Table 6).

356 Table 6 shows the SCOD values of the thermophilic and mesophilic temperature co-
357 phase co-digestion systems. At steady state, in TPACD10/10, the SCOD values in the co-
358 phase thermophilic and thermophilic digesters were 5800 and 3000 mg/l, which were
359 higher than those of single-stage mesophilic and thermophilic processes, respectively.

360 The good effluent quality in terms of COD was mainly attributable to the low VFA
361 levels in the co-phase thermophilic and mesophilic digesters, probably due to the
362 higher methanogenic activity and higher affinity of the anaerobic substrate for VFA in
363 the co-phase system in the first TPACD test.

364 Figure 3a shows the VFA-to-alkalinity ratio required to evaluate the buffering capacity
365 of the temperature co-phase anaerobic co-digestion system; values higher than 0.5
366 clearly indicate that the reactor does not contain a good equilibrium between
367 acidogenic and methanogenic microbiota. In TPACD10/10, the VFA-to-alkalinity ratios
368 were 0.21 for the thermophilic digester and 0.20 for the mesophilic digester, which is
369 an indicator of a high level of stability. These values indicate that the buffering capacity
370 in the temperature co-phase anaerobic system was sufficient for SS and SBPL co-
371 digestion, as with the single-stage mesophilic anaerobic processes. The slightly higher
372 buffering capacity in the co-phase thermophilic digester was attributable to both a
373 higher alkalinity level from the enhanced degradation of nitrogenous compounds and
374 as well as the VFA level. The higher buffering capacity in the co-phase thermophilic
375 digester also contributed to the good buffering capacity in the mesophilic digester
376 through substrate exchange between the thermophilic and mesophilic digesters.

377 Nevertheless, a reduction in the HRT in the thermophilic phase (TPAcD6/10) caused an
378 increase in the acidity/alkalinity ratio in the thermophilic effluent with a value of 0.77.
379 The overall specific methane yields were as good as the single-stage mesophilic
380 anaerobic process (T10), although some portion of the overall yield was from the
381 thermophilic digester of the co-phase digestion system. The HRT of the mesophilic
382 digester was 10 days, in both cases, but the specific methane yield was lower in
383 TPAcD10/10, showing 340 ml CH₄/gTVS_{removal}. The methane generated from the wastes
384 calculated with respect to TVS removal was higher in mesophilic phase of the TPAcD
385 process in comparison with the thermophilic stage. This suggests that the thermophilic
386 reactor was not efficient at converting all the intermediate products into methane. In
387 TPAcD6/10, there was a drop in biogas production in the mesophilic phase, due to the
388 accumulation of VFA in the previous stage.

389 The TVS in the co-phase mesophilic and thermophilic digesters were stable, and were
390 not influenced by the TVS variation in the influent substrate, as shown in Figure 3b.

391 The reduction in volatile solids was around 77.2% in TPAcD10/10, and remained stable
392 in TPAcD6/10 although the global methane yield was lower, as shown in Table 6. In the
393 literature [13], the reduction in volatile solids obtained using the TPAcD process for
394 waste-activated sludge was about 50% at 28 days of SRT, which was around 10%
395 higher than that of the single-stage mesophilic digester. In this study, the reduction in
396 volatile solids that could be obtained in the co-phase digestion system was over 36.5%
397 higher than that of the single-stage mesophilic digester and around 1% higher than the
398 single-stage thermophilic digester. The enhanced performance in terms of TVS
399 reduction obtained from the temperature co-phase anaerobic digestion system was
400 mainly attributable to the higher hydrolytic activity of the thermophilic digester. On

401 the other hand, the additional energy for substrate exchange and for heating the
402 thermophilic digester in the co-phase digestion system should be considered.
403 However, these additional energy requirements could be compensated for by the
404 advantages of the co-phase digestion system, including the reduction of volatile solids,
405 better effluent quality and process stability, and increased methane production,
406 compared to the single-stage mesophilic or the thermophilic processes.

407 **3.3. Microbial population dynamics**

408 Microbial populations in anaerobic digestion have previously been investigated, with
409 the finding that HRT is a significant factor in selecting the predominant microbial
410 species [18, 32]. One of the objectives of the present study was to obtain direct
411 experimental evidence for the influence of HRT on the population levels of
412 methanogenic anaerobic microorganisms in the digester.

413 The results show the evolution of the methanogenic bacteria concentration at
414 different HRT (days). The methanogenic counts were performed at the end of each
415 period [19, 20, 33] when the microbial population had adapted to the new organic
416 loading rate conditions in the mesophilic and thermophilic single-stage anaerobic co-
417 digestion process as well as the TPACD processes (TPACD10/10 and TPACD6/10).

418 Anaerobic effluent from the mesophilic anaerobic digester of sewage sludge from a
419 waste water treatment industrial plant was used as the inoculum. In single phase
420 anaerobic co-digestion, the microbial community is not dependent on the imposed
421 OLR. However, the microbial community was larger in the mesophilic range than in the
422 thermophilic range in both HRTs assayed. In the TPACD process, a slight increase in the
423 microbial population took place, compared with 10 days HRT in mesophilic and

424 thermophilic single anaerobic co-digestion, as the result of the higher content of
425 microorganisms in the substrate.

426 Methanogenic microorganism activity was determined by comparing the amount of
427 methane generated for each HRT tested with the size of the population in the
428 methanogenic reactor analyzed by epifluorescence microscopy. The results are shown
429 in Table 7.

430 Microbial activity increased between 20 and 10 days of HRT in mesophilic and
431 thermophilic single anaerobic co-digestion, and was much higher when the microbial
432 content in the reactor decreased. In systems with no biomass retention, a decreased
433 HRT is reflected by a lower number of microorganisms exiting the system daily in the
434 effluent. Consequently, the population inside the reactor is very active. Due to the
435 increase in biogas and methane generation when the HRT decreased, the activity
436 increased when the HRT decreased. In the single phase anaerobic system,
437 independently of the operated HRT, the positive correlation between activity and
438 methane generation was high. There was a high correlation between OLR and
439 microbial activity in single-stage anaerobic co-digestion of sewage sludge and sugar
440 beet pulp lixiviation.

441 In the TPACD processes, the individual microbial activity of each phase decreased in
442 concordance with the reduction in methane generation at each stage. These results
443 seem to show that the activity of anaerobic microorganisms in the reactor could be
444 more related to the OLR than to microbial concentrations.

445 Under some conditions, microbial number and activity showed proportional
446 correlations, whereas this is not the case in many realistic circumstances. This requires

447 caution and critical thinking when one parameter is calculated or estimated from
448 another.

449 This study shows that the increase in microbial activity inside the reactor is directly
450 proportional to the OLR (or inversely proportional to the HRT) and inversely
451 proportional to the size of the microbial population in the system in single-stage
452 anaerobic co-digestion. These results are in accordance with those previously reported
453 by Solera et al. (2001b) [19], in contrast to results from other studies showing a direct
454 correlation between the methanogenic population and the organic loading rate [17,
455 34].

456 **4. Discussion**

457 Most full-scale biomethanation systems in use are single-stage mesophilic digesters, in
458 which it is difficult to provide optimal conditions for all four of the guilds of microbes.
459 As such, the metabolic activities of the microbial guilds are compromised and the
460 performance of single-stage mesophilic digesters is often suboptimal; the reduction of
461 TVS is rather slow and only a portion of TVS can be converted. Although pretreatments
462 using heat and diluted acid or base can improve the digestibility of the feedstock, they
463 inevitably increase capital and operational costs and potentially produce inhibitory
464 compounds. In addition, up to two thirds of the methane is produced from acetate in
465 anaerobic digesters [27], but syntrophic acetogens and acetoclastic methanogens have
466 extremely slow growth due to their thermodynamically unfavorable pathways [28].
467 Consequently, the entire biomethanation process in single-stage mesophilic AD
468 systems is often suboptimal and prone to being disrupted by the accumulation of
469 propionate and butyrate, especially at high organic loading rates [29].

470 Thermophilic AD is considered one of the most promising approaches to improve
471 biometanation by accelerating the hydrolysis of the polymeric feedstock and other
472 metabolic pathways [30]. For microbial biomass-laden feedstocks, high temperatures
473 help to lyse intact microbial cells, making the cellular components available for
474 bioconversion. However, several studies have shown that thermophilic digesters suffer
475 from poor stability due to the accumulation of VFA, especially propionate, reduced
476 methane production, and an increased carbon dioxide content [29]. The above
477 limitations associated with thermophilic AD are thought to be attributable to several
478 factors. First, elevated temperatures decrease the diversity and robustness of
479 methanogens in digesters, as only three species of methanogens have been identified
480 in thermophilic anaerobic digesters [31]. Second, high temperature decreases the
481 solubility of H₂. Third, some microbes, especially syntrophic acetogens and
482 methanogens, are more susceptible to inhibitory metabolites (e.g., NH₃, H₂S, and
483 propionic and butyric acids) at thermophilic temperatures than at mesophilic
484 temperatures [27].

485 **5. Conclusions**

486 The single-stage mesophilic AcD was superior to the thermophilic AcD in terms of the
487 specific methane yield, effluent quality and process stability. However, TVS reduction
488 in the thermophilic AcD was higher than in the mesophilic AcD.

489 The performance of TPACD was dependent on the HRT of the thermophilic digester,
490 but the advantages of single-stage mesophilic and the thermophilic AD could be
491 obtained in the TPACD system. The effluent quality (in terms of specific methane yield
492 and process stability) was higher for the TPAD process than for the single-stage
493 mesophilic AcD, but not in terms of soluble COD and VFA. The TVS reduction in the

494 TPACD process was much higher than in the single-stage mesophilic AcD and similar to
495 that in the single-stage thermophilic AcD.
496

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503 **References**

- 504 [1] J. Mata-Alvarez, S. Mac, P. Llabrés. Anaerobic digestion of organic solid wastes, an
505 overview of research achievements and perspectives. *Bioresour. Technol.* 74,(2000) 3–
506 16.
- 507 [2] F. Alatríste-Mondragón, P. Samar, H.H. Cox, B.K. Ahring, R. Iranpour. Anaerobic co-
508 digestion. *Water Environ. Res.* 78,(2006) 607–636.
- 509 [3] N. Aoki M. Kawase. Development of high performance thermophilic two-phase
510 digestion process. *Water Sci Technol*; 23(1991):1147–56.
- 511 [4] C. Maibaum, V. Kuehn. Thermophilic and mesophilic operation of an anaerobic
512 treatment of chicken slurry together with organic residual substances. *Water Sci*
513 *Technol*; 40(1)(1999):231–6.
- 514 [5] J. Zabranska, J. Stepova, R. Wachtl, P. Jenicek, M. Dohanyos. The activity of
515 anaerobic biomass in thermophilic and mesophilic digesters at different loading rates.
516 *Water Sci Technol.*, 32(9)(2000):49–56.
- 517 [6] M. Kim, Y.H. Ahn, R.E. Speece. Comparative process stability and efficiency of
518 anaerobic digestion; mesophilic vs. thermophilic; 36 (2002):4369–85.
- 519 [7] J.B. van Lier. Limitation of thermophilic anaerobic wastewater treatment and the
520 consequences for process design. *Antonie van Leeuwenhoek*, 69 (1996):1–14.

521 [8] B.N. Azbar, R. Speece. Two-phase, two-stage and singlestage anaerobic process
522 comparison. *J Environ Eng.*, 127(3)(2001):240–7.

523 [9] R. Roberts, L. Son, C.F. Forster. Thermophilic/mesophilic dual digestion system for
524 treating waste activated sludge. *J Chem Technol Biotechnol*, 74 (1999):445–50.

525 [10] Q. Zhao, G. Kugel. Thermophilic/mesophilic digestion of sewage sludge and
526 organic wastes. *J Environ Sci Health*, A31 (1996) (9):2211–31.

527 [11] Y.C. Song, S.H. Park, J.S. Lee. Enhanced anaerobic stabilization of sewage sludge
528 using TPAD process. *J Korean Soc. Civil Eng.* 21(2001) (6B):70 5–13.

529 [12] B. Ferrán. Two-phase anaerobic digestion of municipal sewage sludge
530 optimization of the pathogen destruction. Proceedings of the 75th WEF Annual
531 Conference and Exposition (WEFTEC 2002), Session 46. Chicago: WEF; 2002.

532 [13] Y. Han, S. Sung, R.R. Dague. Temperature-phased anaerobic digestion of
533 wastewater sludge. *Water Sci Technol.*, 36 (1997) (6–7):367–74.

534 [14] N. Azbar, P. Ursillo, R.E. Speece. Effect of process configuration and substrate
535 complexity on the performance of anaerobic processes. *Water Res.*, 35 (2001) (3):817–
536 29.

537 [15] V. Riau, M.A. De la Rubia, M. Perez. Temperature-phased anaerobic digestion
538 (TPAD) to obtain class A biosolids: A semi-continuous study. *Bioresour. Technol.* 101,
539 (2010) 2706–2712.

540 [16] W. Zabriskie A.E. Humphre. Real-time estimation of aerobic batch fermentation
541 biomass concentration by component balancing. *AIChE Journal*, 24 (1978), 138–146.

542 [17] T.C. Zhang, T. Noike. Influence of retention time on reactor performance and
543 bacterial trophic populations in anaerobic digestion processes. *Water Res.*, 28 (1994)
544 (1):27–36.

545 [18] R. Solera, L.I. Romero, D. Sales. Analysis of the methane production in
546 thermophilic anaerobic reactors: use of autofluorescence microscopy. *Biotechnol. Lett.*
547 23 (2001) (22), 1889–1892.

548 [19] R. Solera, L.I. Romero, D. Sales. Determination of the microbial population in
549 thermophilic anaerobic reactor: comparative analysis by different counting methods.
550 *Anaerobe*, 7 (2001) :79–86.

551 [20] H.J. Doddema, G.D. Vogels. Improved identification of methanogenic bacteria by
552 fluorescence microscopy. *Appl Environ Microbiol.*, 36 (1978) (5):752–4.

553 [21] B.K. Ince, O. Ince. Changes to bacterial community makeup in a twophase
554 anaerobic system. *J Chem Technol Biotechnol.*,75 (2000) (6):500–8.

555 [22] R. Montañés, M. Pérez, R. Solera. Mesophilic anaerobic co-digestion of sewage
556 sludge and a lixiviation of sugar beet pulp: Optimisation of the semicontinuous
557 process. *Bioresour. Technol.* 142, (2013) 655-662.

558 [23] APHA, AWWA, WEF, 1995. *Standard Methods for the Examination of Water and*
559 *Wastewater*, 19th ed. Washington, DC, New York, USA.

560 [24] M.G. Capri, Gv.R. Marais. pH adjustment in anaerobic digestion. *Water Res.*, 9
561 (1975) :307–13.

562 [25] E.V. Munch, P.F. Greenfield. Estimating VFA concentrations in prefermenters by
563 measuringpH. *Water Res.*, 32 (1998) (8):2431–41.

564 [26] C. Gallert, J. Winter. Mesophilic and thermophilic anaerobic digestion of source
565 sorted organic wastes: effect of ammonia on glucose degradation and methane
566 production. *Appl Microbial Biotechnol.*, 48 (1997) 405–10.

567 [27] Z. Yu, M. Morrison, F.L. Schanbacher. Production and utilization of methane
568 biogas as renewable fuel. (2009). In: Vertès, A.A., Blaschek, H.P., Yukawa, H.,

569 [28] F.A. de Bok, C.M. Plugge, A.J. Stams. Interspecies electron transfer in
570 methanogenic propionate degrading consortia. *Water Res.* 38, (2004) 1368–1375.

571 [29] R.E. Speece, S. Boonyakitsombut, M. Kim, N. Azbar, P. Ursillo. Overview of
572 anaerobic treatment: thermophilic and propionate implications. *Water Environ. Res.*
573 78, (2006) 460–473.

574 [30] A. Converti, A. del Borghi, M. Zilli, S. Arni, M. del Borghi. Anaerobic digestion of
575 the vegetable fraction of municipal refuse: mesophilic versus thermophilic conditions.
576 *Bioprocess Biosyst. Eng.* 21,(1999) 371–376.

577 [31] T. Hori, S. Haruta, Y. Ueno, M. Ishii, Y. Igarashi. Dynamic transition of a
578 methanogenic population in response to the concentration of volatile fatty acids in a
579 thermophilic anaerobic digester. *Applied and Environmental Microbiology* 72
580 (2)(2006), 1623–1630.

581 [32] Y.Y. Li, T. Noike. Metabolic characteristics and distribution of fatty acid-utilizing
582 bacteria in anaerobic digester. In: *Proceedings of 23rd annual conference of the Japan*
583 *Society on Water Pollution Research*, p.(1999) 445–6.

584 [33] M.K. Jain, G. Zeikus, L Bhatnagar. Methanogens. In: Levett PN, editor. *Anaerobic*
585 *microbiology A practical approach*, New York: Oxford University Press, p. (1991) 223–
586 45.

587 [34] M.A. De la Rubia, M. Pérez, L.I. Romero, D. Sales, D. Effects of solids retention time
588 (SRT) on pilot scale anaerobic thermophilic sludge digestion. *Process Biochem.* 41 (1),
589 (2006) 79-86.

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