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Introduction

Limits of Life and the Habitability of Mars: The ESA Space Experiment BIOMEX on the ISS

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Abstract

BIOMEX (BIOlogy and Mars EXperiment) is an ESA/Roscosmos space exposure experiment housed within the exposure facility EXPOSE-R2 outside the Zvezda module on the International Space Station (ISS). The design of the multiuser facility supports-among others-the BIOMEX investigations into the stability and level of degradation of space-exposed biosignatures such as pigments, secondary metabolites, and cell surfaces in contact with a terrestrial and Mars analog mineral environment. In parallel, analysis on the viability of the investigated organisms has provided relevant data for evaluation of the habitability of Mars, for the limits of life, and for the likelihood of an interplanetary transfer of life (theory of lithopanspermia). In this project, lichens, archaea, bacteria, cyanobacteria, snow/permafrost algae, meristematic black fungi, and bryophytes from alpine and polar habitats were embedded, grown, and cultured on a mixture of martian and lunar regolith analogs or other terrestrial minerals. The organisms and regolith analogs and terrestrial mineral mixtures were then exposed to space and to simulated Mars-like conditions by way of the EXPOSE-R2 facility. In this special issue, we present the first set of data obtained in reference to our investigation into the habitability of Mars and limits of life. This project was initiated and implemented by the BIOMEX group, an international and interdisciplinary consortium of 30 institutes in 12 countries on 3 continents. Preflight tests for sample selection, results from ground-based simulation experiments, and the space experiments themselves are presented and include a complete overview of the scientific processes required for this space experiment and postflight analysis. The presented BIOMEX concept could be scaled up to future exposure experiments on the Moon and will serve as a pretest in low Earth orbit. Key Words: EXPOSE-R2—BIOMEX—Habitability—Limits of life—Extremophiles—Mars. Astrobiology 19, 145–157.

1. Results from Previous Spaceflight and Ground-Based Experiments

PREVIOUS EXPERIMENTS in spaceflight and ground-based studies, which were performed before the BIOMEX (BIOlogy and Mars EXperiment) proposal submission to ESA, showed that, in particular, microcolonies of bacteria, meristematic black fungi, and symbiotic associations of microorganisms such as lichens are able to survive and be reactivated after simulated and direct space experiments (Tarasenko et al., 1990; Horneck et al., 1994; de Vera et al., 2003, 2004a, 2004b, 2007, 2008, 2010; de la Torre Noetzel et al., 2007; Sancho et al., 2007; Onofri et al., 2008, 2010; Olsson-Francis et al., 2009; de la Torre et al., 2010; de Vera and Ott, 2010). Bacteria strains such as Bacillus subtilis and Deinococcus radiodurans have shown a certain radiation and vacuum tolerance (Horneck, 1993; Horneck et al., 1994, 2001; Rettberg et al., 2002, 2004; Möller et al., 2007a, 2007b, 2007c; Pogoda de la Vega et al., 2007; Wassmann et al., 2012; Panitz et al., 2014). Gram-negative endophytic bacteria and cyanobacteria survived a 14-day shuttle flight (within the shuttle interior) and exhibited enhanced plant colonizing activity in microgravity (Tarasenko et al., 1990). During the BIOPAN 5 and 6 experiments, the lichens Rhizocarpon geographicum and Xanthoria elegans were analyzed after exposure to space conditions of about 11-14 days coupled with parallel tests in ground-based facilities. These results have led to the conclusion that the tested symbiotic eukaryotic associations of alga and fungi in the lichen were not seriously damaged, and nearly 70-100% of the tested lichens survived. The lichens were physiologically active and able to germinate and grow. Furthermore, investigations on the mutation rate of photoproducts on the DNA have shown that the mycobiont (the fungal symbiont) is practically unaffected by UV radiation and that the algal symbiont is more sensitive (de Vera et al., 2003, 2004a, 2004b, 2007, 2008, 2010; de Vera, 2005; de la Torre Noetzel et al., 2007; Sancho et al., 2007; de la Torre et al., 2010; de Vera and Ott, 2010). Cyanobacteria, as has been shown by analysis on akinetes (resting-state cells of cyanobacteria), were also able to survive the low Earth orbit and simulated extraterrestrial conditions (Olsson-Francis et al., 2009), while vegetative cells of Chroococcidiopsis sp. CCMEE 029 survived prolonged desiccation periods (Billi, 2009; Fagliarone et al., 2017) and a few minutes of exposure to an attenuated Mars-like UV flux and 4 h of exposure to a Mars-like UV flux (Cockell et al., 2005). Numerous species mentioned in this study were even able to survive simulated catastrophes as induced by asteroid impact simulations (Horneck et al., 2001, 2008; Stöffler et al., 2007; Meyer et al., 2011). Mars simulation tests with methanogenic archaea have also shown a remarkable level of survival and demonstrated physiological activity during exposure to Mars-like environmental conditions (Morozova and Wagner, 2007; Morozova et al., 2007, 2015; Schirmack et al., 2014). The same has been observed for meristematic black fungi during a ground-based experiment in the facilities at the German Aerospace Center (DLR) Cologne named EVT (Experiment Verification Test), which was performed for the Lichens and Fungi Experiment (LIFE) on EXPOSE-E (Onofri et al., 2008) and after the final space experiment (Onofri et al., 2012, 2015). In other ground-based experiments, we were able to show that Paenibacillus sp. caused biocorrosion of anorthosite rock (Lytvynenko et al., 2006). In total, we can presume that a wide variety of different microorganisms, even from higher evolutionary advanced levels than those of archaea or bacteria, are able to resist and survive space and Mars-like conditions for a period of time (at least for 1.5 years). However, because of the limited capacity of the space exposure facilities, further work with replicates and other samples is still needed to finally answer questions on the degree of Mars' habitability or the kind of space and Mars-like environmental conditions that are limiting factors in reference to the most important vital functions of life (de Vera et al., 2014; Schulze-Makuch et al., 2015).

The BIOMEX results presented here further advance our knowledge and address pressing questions as mentioned above

BIOMEX selected sar	nples for spaceflight						
Archaea	Methanosarcina sp. strain SMA-21 (terrestrial permafrost) (GFZ/AWI Potsdam)						
Bacteria	Deinococcus radiodurans wild type and crtI or crtB (nonpigmented) (DLR Cologne)						
	Biofilm containing Leptothrix, Pedomicrobium, Pseudomonas, Hyphomonas, Tetrasphaera (TU Berlin)						
	Cyanobacterium Nostoc sp. strain CCCryo 231-06 (Fraunhofer IZI-BB)						
	Cyanobacterium Gloeocapsa OU-20 (Astrobiology Center Edinburgh)						
	Cyanobacterium Chroococcidiopsis sp. CCMEE 029 (Uni Roma)						
Alga	Green alga Sphaerocystis sp. CCCryo 101-99 (Fraunhofer IZI-BB)						
Lichens	Circinaria gyrosa (INTA)						
	Buellia frigida (Antarctic lichen) (H-H-Uni Düsseldorf)						
Fungi	Cryptoendolithic Antarctic black fungus Cryomyces antarcticus CCFEE 515 (Uni Viterbo)						
Bryophytes	Grimmia sessitana (alpine samples) (Uni Potsdam)						
	Marchantia polymorpha L. (Uni Potsdam)						
Biomolecules	Pigment Chlorophyll (H-H-Uni Düsseldorf)						
	Pigment beta-Carotene (H-H-Uni Düsseldorf)						
	Pigment Naringenin (H-H-Uni Düsseldorf)						
	Pigment Quercitin (H-H-Uni Düsseldorf)						
	Pigment Parietin (H-H-Uni Düsseldorf)						
	Pigment Melanin (H-H-Uni Düsseldorf)						
	Cellulose (H-H-Uni Düsseldorf)						
	Chitin (H-H-Uni Düsseldorf)						
Biofilm	Kombucha biofilm containing: Yeasts: Saccharomyces ludwigii, Schizosaccharomyces pombe, Zygosaccharomyces rouxii, Zygosaccharomyces bailii, Brettanomyces bruxellensis; Bacteria: Paenibacillus sp. IMBG221, Acetobacter nitrogenifigens, Gluconacetobacter kombuchae sp. nov., Gluconacetobacter xylinum (NAS Ukraine)						
Substrates/Minerals	Agar (as a substitute for Murein) (H-H-Uni Düsseldorf)						
	Minerals lunar analog mixture (MfN Berlin)						
	Minerals P-MRS: Early acidic Mars analog (Mixture of Fe ₂ O ₃ , montmorillonite, chamosite, kaolinite, siderite, hydromagnesite, quartz, gabbro, and dunite) (MfN Berlin)						
	Minerals S-MRS: Late basic Mars analog (Mixture of hematite, goethite, gypsum, quartz, gabbro, dunite) (MfN Berlin)						
	Silica discs (glass) (Astrobiology Center Edinburgh)						

Gray shaded cells indicate the samples for which results are available and included in this special collection.

TABLE 1. SELECTED SAMPLES FOR BIOMEX

Component	P-MRS (wt %)	S-MRS (wt %)	LRA (wt %)
Gabbro (Groß-Bieberau, Germany)	3	32	-
Dunite—Olivine Fo ₉₆ (Åheim, Norway)	2	15	5.7
CPx—Diopside (Kragerö, Norway)	-	-	8.9
OPx—Hyperstehn (Egersund, Norway)	-	-	5.7
Anorthosite—Plagioclase (Larvik, Norway)	-	-	66.8
Quarzite (Bayerischen Wald, Germany)	10	3	-
Apatite (Minas Gerais, Brasil)	-	-	1.1
Hematite (Cerro Bolivar, Venezuela)	5	13	-
Illmenite (Flekkefiord, Norway)	-	-	1.1
Iron (Fe)	-	-	1.3
Montmorillonite (Hallertau, Germany)	45	-	-
Chamosite (Nucic, Czech Republic)	20	-	-
Kaolinite (Hirschau, Germany)	5	-	-
Siderite (Hüttenberg, Austria)	5	-	-
Hydromagnesite (Albaner Berge, Italy)	5	-	-
Goethite (Salchendorf, Germany)	-	7	-
Gypsum (Nüttermoor, Germany)	-	30	-
Volcanic slag (Aeolian islands, Italy)	-	-	9.4

TABLE 2. MARS AND LUNAR ANALOG MINERAL MIXTURES

P-MRS: phyllosilicatic martian regolith = early acidic. S-MRS: sulfatic martian regolith = late basic. LRA: lunar regolith analog.

to an extended degree in comparison to previously executed space experiments, which, for the most part, were more restricted and focused on investigating the likelihood of an interplanetary transfer of life as is formulated in the lithopan-spermia hypothesis (Richter, 1865; Thomson, 1894; Arrhenius, 1903; see also Lee *et al.*, 2017). Accordingly, these new BIOMEX experiments in space were intended to address new questions in planetary research and improve future space exploration goals. Nevertheless, it is clear that the results obtained by BIOMEX could also be used to evaluate previously performed space experiments in reference to lithopanspermia.

2. Sample Selection

As a consequence of results obtained in previous space experiments that engendered a significant number of stillopen questions, a proposal named BIOMEX (ILSRA-2009-0834) was submitted in 2009. This was in response to the ESA international research announcement for research in space life sciences at the International Space Station (ISS)—ILSRA-2009—and BIOMEX was successfully selected. The proposal included replicate exposure of known species used in previous space experiments such as the reassessed Antarctic



FIG. 1. Mars analog pellets integrated in the EXPOSE-R2 hardware.

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fungus *Cryomyces antarcticus*, the cyanobacterium *Chroococcidiopsis* sp., the lichen *Circinaria gyrosa* (formerly known as *Aspicilia fruticulosa* before reclassification; Sohrabi *et al.*, 2013), and a set of new, preselected organisms for further preflight experiments (see Table 1). After survival of these organisms was shown, the samples were incorporated into the BIOMEX experiment, integrated into the final EXPOSE-R2 hardware, and sent to space on the ISS.

This new sample set was chosen systematically and comprised a selection from archaea, bacteria, and eukaryotes, which represent the three main domains of the tree of life. Most of these organisms were collected from Mars analog habitats distributed on different continents, which include the Alps (climatic and geomorphologic Mars analogy: gullies, polygons, temperatures below 0°C, dryness, and elevated UV irradiation), the steppe highlands of Central Spain (characterized by extreme insolation, high temperature contrasts, and arid summers Crespo and Barreno, 1978]), and regions in the Arctic and Antarctica. The aim of selecting a wide variety of species was to identify which are able to demonstrate the limits of life with regard to the applied space and Mars-like conditions in low Earth orbit, as well as further our understanding of the kind of species for which Mars could be habitable.

2.1. Biological samples

A set of organisms was tested, which were embedded in, or grown on, Mars (optionally lunar) analogs and other terrestrial minerals. Bacteria, biofilms of bacteria and yeast species, cyanobacteria, archaea, lichens, snow/permafrost algae, meristematic black fungi, and bryophytes of mostly alpine and polar habitats of desiccation- and radiationresistant strains were chosen because some of these organisms are thought to be among the oldest on Earth (Wang et al., 1999; Schidlowski, 2001; Campbell et al., 2003; Yuan et al., 2005) and, over time, have evolutionarily adapted to different environmental conditions. Some of these organisms could even be Mars-relevant because they use Mars-atmosphere resources such as CO₂ to form methane, a trace gas found remotely in the martian atmosphere (Formisano et al., 2004; Mumma et al., 2009) and in situ at Gale Crater by way of the rover Curiosity (Webster et al., 2015). The processes that lead to extreme variations in the methane concentration of the martian atmosphere, in particular the potential for abiogenic origin (Lefèvre and Forget, 2009), have recently been reviewed (Yung et al., 2018). In previous studies, some of the organisms studied in the BIOMEX experiments have also exhibited a high resistance under simulated and real space conditions or simulated martian conditions. Others, like the newly selected bryophytes, provide insights on the resistance capabilities of an evolutionarily younger life-form. Details on the selected species are listed in Table 1.

2.2. Mars and lunar analog mineral mixtures

With the BIOMEX experiment, our goal was to analyze the effects of a space environment that approaches as closely as possible Mars-like environmental conditions and includes the use of Mars analog mixtures that could serve as a substrate or an embedding matrix for biological samples (see Table 2 and Figs. 1 and 2). These Mars analog mineral



FIG. 2. Grain size distribution of martian regolith analog P-MRS (early acidic MRS) and S-MRS (late basic MRS).

mixtures mimic the regolith cover from early and late evolutionary stages of Mars (Böttger et al., 2012). The components of the mixtures were developed in the Museum für Naturkunde (MfN) Berlin (Germany) in the context of the Helmholtz-Alliance "Planetary Evolution and Life" proposal and based on several observational studies (Bibring et al., 2005, 2006; Poulet et al., 2005; Chevrier and Mathé, 2007). It is important to test the effects of space and the martian environment on minerals in parallel biological investigations. A welcome consequence of this space experiment would be that the investigated samples would also be tested for viability and space-resistance capacity and provide valuable data if used in a replicate space experiment with regard to the probability of lithopanspermia in the Earth-Mars system. The lithopanspermia hypothesis has also been investigated in previous space experiments on FOTON/BIOPAN some years ago and on the EXPOSE-E mission on the ISS. However, replicates are still needed. Besides the Mars analog mineral mixtures, the MfN also provided a lunar regolith analog (see Table 2 and Figs. 1 and 3) for investigation into the influence of the lunar surface material on organisms, which could be relevant for lifesupport systems such as the selected and tested cyanobacteria (not part of this special collection of articles).



FIG. 3. Grain size distribution of lunar regolith analog material LRA.

 TABLE 3. EXPERIMENT VERIFICATION TESTS (EVTs)

EXPOSE-R2 EVT part 1	BIOMEX Experiment
Test parameter	performed
Vacuum 10 ⁻⁵ Pa	7 d, pressure: 3.5×10^{-2} ±0.12 Pa
Mars atmosphere (CO ₂ gas composition) 103 Pa	7 d, pressure: 6.5×10^2 ±0.12 Pa
Temperature -10°C to +45°C	48 cycles 8 h each
Temperature max and min –25°C and +60°C	-25°C±0.5°C, 1 h +60°C±0.5°C, 1 h
Irradiation 254 nm Hg low-pressure lamp @ 56 μW/cm ²	$\begin{array}{c} 0s \rightarrow 0J/m^2 \\ 18s \rightarrow 10.1J/m^2 \\ 2min59s \rightarrow 100.2J/m^2 \\ 29min46s \rightarrow 1000.2J/m^2 \\ 4h57min37s \rightarrow 9999.9J/m^2 \end{array}$
EXPOSE-R2 EVT part 2 (run 1+2)	BIOMEX Experiment
Run 1 Irradiation 200–400 nm SOL2000 @ 1,271.2 W/m ² _{200–400nm}	$\begin{array}{l} 0\mathrm{s} \rightarrow \mathrm{dark} \\ 18\mathrm{min} \rightarrow 1.4 \times 10^3 \mathrm{kJ/m^2} \\ 3\mathrm{h} \rightarrow 1.4 \times 10^4 \mathrm{kJ/m^2} \\ 30\mathrm{h} \rightarrow 1.4 \times 10^5 \mathrm{kJ/m^2} \\ 99\mathrm{h} \rightarrow 4.5 \times 10^5 \mathrm{kJ/m^2} \\ 148\mathrm{h} \rightarrow 6.8 \times 10^5 \mathrm{kJ/m^2} \end{array}$
Run 2 Irradiation 200–400 nm	$0s \rightarrow dark$ $432s \rightarrow 5.5 \times 10^{2} \text{ kJ/m}^{2}$ $(0.1\% \text{ ND filter})$ $1 \text{ h } 12 \text{ min } \rightarrow 5.5 \times 10^{3} \text{ kJ/m}^{2}$
SOL2000 @ 1,271.2 $W/m_{200-400nm}^2$ (as for a 12-month mission duration)	$(1.0\% \text{ ND filter})$ $30 \text{ h} \rightarrow 1.4 \times 10^5 \text{ kJ/m}^2$ $60 \text{ h} \rightarrow 2.7 \times 10^5 \text{ kJ/m}^2$ $120 \text{ h} \rightarrow 5.5 \times 10^5 \text{ kJ/m}^2$
Gluing test	>24 h vulcanization, glue: Wacker-silicone

ND: neutral density.

2.3. Ground-based environmental and space simulations

Many preflight tests were performed to discern whether the chosen samples (Fig. 4) are able to resist extreme conditions with reference to space and martian environments. After an array of experiments on Mars-like regolith and desiccation tests, the organisms were exposed to Experiment Verification Tests (EVTs) and Scientific Verification Tests (SVTs) (Rabbow *et al.*, 2017). In the EVTs, a number of parameters were tested individually on the selected samples. Among these tests, vacuum, a low-pressure Mars-like CO₂ atmosphere, extreme temperature cycles from far below zero to more than 40°C, and UVC irradiation were applied (see Table 3). The SVT experiments were conducted inside hardware with conditions that approached those of the space environment at the ISS (see Table 4).

2.4. Radiation conditions in space

For several of the BIOMEX experiments, we used different neutral density filters (Fig. 5) as covers below the sample window or as cutoff filters, which allowed for a radiation spectral range such as that present at the surface of Mars. For the more protected samples, doses varied between 4.13×10^1 kJ/m² and 6.5×10^3 kJ/m² (see Fig. 5 for details). Samples such as the cyanobacteria of the genera Nostoc, Gloeocapsa, and Chroococcidiopsis; the green snow/permafrost alga Sphaerocystis; and the fungus Cryomyces antarcticus were partly shielded by special MgF₂ neutral density filters with transmission of 0.1% for space conditions and quartz filters with transmission of 0.1% for Marslike conditions (for detailed description of the EXPOSE-R2 facility and the space mission, see Rabbow et al. [2017]). These allowed for an approximation of martian subsurface radiation conditions, representing a thin soil cover with a significantly reduced amount of transmission. The reason for these filters, which covered select samples, was also to mimic those conditions that occur in natural habitats where organisms primarily colonize endolithic niches or fissures and cracks in rocks or soils or are embedded by shielding snow or ice. Some of these organisms occur naturally as endoliths, such as Chroococcidiopsis sp. and Cryomyces antarcticus. Furthermore, the neutral density filters with 0.1% transmission

SVT	Duration	Pressure	Atmosphere	Temperature (T)	T extremes	Irradiation
Tray 1	December 2013– January 2014, 38 d	vacuum pressure at 4.1×10 ⁻⁵ Pa		<i>T</i> cycles between -25° C (16 h in the dark) and $+10^{\circ}$ C (8 h during irradiation)		The upper layers of each tray: UVR _{200-400nm} with 1271 Wm ⁻² $(5.7 \times 10^5 \text{ kJ m}^{-2})$ for 5924 min The lower layers of the trays were kept in the dark
Tray 2			Mars atmosphere (95.55% CO ₂ , 2.7% N ₂ , 1.6% Ar, 0.15% O ₂ , and \sim 370 ppm H ₂ O at 1 kPa)	-23°C		

TABLE 4. SCIENTIFIC VERIFICATION TESTS (SVTs)

Tray # 1 comp 1	-			A.C.	EX	POS	SE-			
01-1-t-01 01-1-t-02 11-1-03 1-1-t-04					R2			BIOMEX		
orden alga Sphaereocystis Sphaereocystis					Samp distri	Sample distribution			lission in open space: 014– 016	
1-1-t-05 1-1-t-06 1-1-t-07 1-1-t-08								99/139		
Cyanobacteria Nostoc Tere Tere Nostoc	1	82	8	2	h					
1-1-t-10 Lichens 1-1-t-10 1-1-t-12 Meristematic black fuori										
Cryomyces Chroococcidiopsis Buella frigida antarcticus sp.		ANK P	To and		L Bot		and the second s	ijk.		
1.1.t-13 12.t-14 1.1.t-15 1.1.t-16 Merintematic black Componenteria	Tray	#20	comp	2:						
Cryomyces Chroococcidiopsis gyrosa Buella frigida antarcticus Sp.	2-2-t-01 S-Mars BC1	2-2-t-02 S-Mars Ca1	2-2-t-03 S-Mars Na1	2-2-t-04 S-Mars Qu1	2-2-t-05 P-Mars BC1	2-2-t-06 P-Mars Ca1	2-2-t-07 P-Mars Na1	2-2-t-08 P-Mars Qu1	BC1 = B-Carotene Ca1 = Chlorophyll a	
Tray # 2 comp 1:	2-2-t-09 S-Mars	2-2-t-10 S-Mars	2-2-t-11 S-Mars	2-2-t-12 S-Mars	2-2-t-13 P-Mars	2-2-t-14 P-Mars	2-2-t-15 P-Mars	2-2-t-16 P-Mars	Na1 = Naringenin	
2-1-t-01 2-1-t-02 2-1-t-03 2-1-t-04	Me1	Pa1	Ce1	Ch1	Me1	Pa1	Ce1	Ch1	Qu1 = Quercitin	
Received and Steel algo	2-2-t-17 S-Mars BC1	2-2-t-18 S-Mars Ca1	2-2-t-19 S-Mars Na1	2-2-t-20 S-Mars Qu1	2-2-t-21 P-Mars BC1	2-2-t-22 P-Mars Ca1	2-2-t-23 P-Mars Na1	2-2-t-24 P-Mars Qu1	Me1 = Melanin Pa1 = Parietin	
Sphaereotystis Sphaereocystis Gloeocapsa polymotipia L	2-2-t-25 S-Mars	2-2-t-26 S-Mars	2-2-t-27 S-Mars	2-2-t-28 S-Mars	2-2-t-29 P-Mars	2-2-t-30 P-Mars	2-2-t-31 P-Mars	2-2-t-32 P-Mars	Ce1 = Cellulose	
Cvanobacteria Cvanobacteria	Me1	Pa1	Ce1	Ch1	Me1	Pa1	Ce1	Ch1	Ch1 = Chitin	
Nostoc Nostoc Cyanobacteria Girminia desaltana Marchantia Gleeocapsa	guartz BC1	2-2-t-34 quartz Ca1	2-2-t-35 quartz Na1	quartz Qu1	2-2-t-37 S-Mars Ag1	2-2-t-38 P-Mars Ag1	2-2-t-39 S-Mars	2-2-t-40 P-Mars	Ag1 = Agar S-Mars → Control	
2.1-1-09 2-1-1-11 2-1-1-12	2-2-t-41	2-2-t-42	2-2-t-43	2-2-t-44	2-2-t-45	2-2-t-46	2-2-4-47	2-2-t-48	P-Mars → Control	
Lichens Meristematic black fungi	Me1	Pa1	Ce1	Ch1	Ag1	Ag1	Co1	Ag1	Quarz → Control	
Cryomyces Chrococcellopsis Byroša ¹³ Buella frigida antarcticus sp.	2-2-t-49	2-2-t-50	2-2-t-51	2-2-t-52	2-2-t-53	2-2-t-54	2-2-t-55	2-2-t-56	P-MRS S-MRS	
2.1.1.13 21.1.14 2.1.05 2.1.16	Iron b	acteria	ROMBUC	A-Biofilm	Arc	naea arcina spec.	crt Deinc	wt	hotoniania hatanlaataa	
Lichens Lichens Circinarias Differs Commy Circinarias Differs Commy Circinarias Differs Commy Circinarias Commy Circinarias Commy Circinarias Commy Circinarias Commy Circinarias Commy Circinarias Commy Circinarias Commy Circinarias Circinaria Circinarias Circinarias Circinarias Cir	2-2-t-57 S-Mars	2-2-t-58 P-Mars	2-2-t-59 lunar	2-2-t-60 lunar	2-2-t-61 S-Mars	2-2-t-62 P-Mars	2-2-t-63 S-Mars	2-2-t-64 S-Mars	LRS	
evrosa Duena menda antarcectos	* /1	* /1					crt	wt	O cm 1	

FIG. 4. Visual table of the sample distribution within the EXPOSE-R2 hardware.



FIG. 5. The distribution of neutral density filters and the values of transmission depending on the used material.

1-1-t-01 2 480 ± 2% Green alga Sphaereocystis	1-1-t-02 3 030 ± 2% Green alga Sphaereocystis	1-1-t-03 4 260 ± 2% Cyanobacteria Gloeocapsa	1-1-t-04 5 040 000 ± 6ryophyte Griennia secularias Marchantia polymorpha L	\$	Tray Tray	#1 co #2 co	omp 1 omp 1	UV-, P	POSE AR-Raditio	-R2	Bl	OME)	(
1-1-t-05 2 960 ± 6% Cyanobacteria Nostoc	1-1-t-06 3 550 ± 6% Cyanobacteria Nostoc	1-1-t-07 5 310 ± 8% Cyanobacteria Gloeocapsa	1-1-t-08 6 440 000 ± 10% Bryophytes Grimmia sessitana + Marchantia polymorpha L.		Tray	#2 c	omp 2	kJ/m ²			22.1	0.2014-03.	.02.20	16
1-1-t-09 6 080 000 ± 10% Lichens Circinaria gyrosa	1-1-t-10 6 810 000 ± 8% Lichens Buelia frigida	1-1-t-11 5 400 ± 7% Meristematic black fungi Cryomyces antarcticus	1-1-t-12 3 000 ± 4% Cyanobacteria Chroococcidiopsis sp.		2-2-t-01 3 890 000 ± 3% S-Mars BC1 2-2-t-08 5 000 000 ±	2-2-1-02 5 160 000 ± 3% S-Mars Cal 2-2-1-10 5 470 000 ±	2-2-t-03 5 530 000 ± 3% S-Mars Na1 2-2-t-11 5 850 000 ±	2-2-1-04 5-210-000 ± 3% 5-Mars Qu1 2-2-4-12 6-530-000 ±	2-2-t-05 5 140 000 ± 3% P-Mars BC1 2-2-t-13 5 530 000 ±	2-2-t-06 5 700 000 ± 3% P-Mars Ca1 2-2-t-14 7 000 000 +	2-2-4-07 5-128-060 ± 2% P-Mars Na1 2-2-4-15 6-350-000 ±	2-2-5-08 4 130 000 ± 3% P-Mars Qu1 2-2-5-18 5 150 000 ±		or code: kJ/m ² 0001 - 1500 1501 - 2500 2501 - 3500
1-1-t-13 5 400 000 ± 12% Lichers	1-1-t-14 5 990 000 ± 11% Lichens	1-1-t-15 4 710±9% Meristematic black fungi Cryomyces	1-1-t-16 2 770± 7% Cyanobacteria Chroococcidiopsi		5% 5.Mars Mo1 2-2-t-17 5 800 000 ± 11%	8% S-Mars Pa1 2-2-t-18 7 410 000 ± 9%	8% S-Mars Ce1 2-2-t-19 7 940 000 ± 9%	8% S-Mars Ch1 2-2-t-20 7 470 000 ± 9%	8% P-Mars Me1 2-2-5-21 7 510 000 ± 9%	P-Mars P-Mars Pa1 2-2-t-22 8 040 000 ± 9%	8% P-Mars Ce1 2-2-t-23 7 330 000 ± 9%	8% P-Mars Ch1 2-2-t-24 5 830 000 ± 9%		3501 - 4500 4501 - 5500
2-1-t-01 2 240 ± 3%	Buelia frigida 2-1-t-02 2 740 ± 3%	antarcticus 2-1-t-03 3 840 ± 3%	sp. 2-1-t-04 4 510 000 ± 4%	-	S-Mars BC1 2-2-t-25 5 510 000 ± 13% S-Mars	S-Mars Ca1 2-2-t-26 7 120 000 ± 11% S-Mars	S-Mars Na1 2-2-t-27 7 500 000 ± 11% S-Mars	S-Mars Gut 2-2-t-28 6 770 000 ± 11% S-Mars	P-Mars 8C1 2-2-t-29 6 880 000 ± 11% P-Mars	P-Mars Ca1 2-2-t-30 7 560 000 ± 11% P-Mars	P-Mars Na1 2-2-t-31 6 930 000 ± 11% P-Mars	P-Mars Qu1 2 2 5 32 5 340 600 2 9% P-Mars		5501 - 6500 4500000 - 5500000 5500001 - 6500000
Green alga Sphaereocystis 2-1-t-05	Green alga Sphaereocystis 2-1-t-06	Cyanobacteria Gloeocapsa 2-1-t-07	Grimmia setsitana + Marchantia polymorpha L 2-1-t-08	2	Me1 2-2-1-33 4 290 000 ± 6% quartz	Pa1 2-2-1-34 5 340 000 ± 5% quartz	2-2-t-35 5 830 000 ± 5% quartz	Ch1 2-2-1-36 5-110-000 ± 4% quartz	Me1 2-2-t-37 6 460 000 ± 8% S-Mars	2-2-t-38 7 270 000 ± 7% P-Mars	Ce1 2-2-t-39 6 610 000 ± 8% S-Mars	Ch1 2.2 t 40 5 210 600 t 7% P-Mars		6500001 - 7400000 7400001 - 8400000
2 650 ± 7% Cyanobacteria Nostoc	3 180 ± 6% Cyanobacteria Nostoc	4 800 ± 9% Cyanobacteria Gloeocapsa	5 810 000 ± 10% Bryophytes Grimmia sessitana + Marchantia polymorpha L.		801 2-2-1-41 5 120 000 5 13% quartz Me1	Ca1 2-2-t-42 6 190 000 ± 11% quartz Pa1	Na1 2-2-t-43 6 690 000 ± 11% quartz Ce1	Qu1 2-2-t-44 6 270 000 ± 11% quartz Ch1	Ag1 2-2-t-45 7 840 000 ± 11% S-Mars Ag1	Ag1 2-2-t-46 8 340 000 ± 11% P-Mars Ag1	2-2-t-47 6 160 000 ± 11% quartz Co1	2.2 t 48 5 020 000 ± 9% guart± Ag1		
2-1-t-09 5 540 000 ± 10% Lichens Gircinaria gyrosa	2-1-t-10 6 220 000 ± 9% Lichens Buelia frigida	2-1-t-11 4 740 ± 8% Meristematic black fungi Cryomyces antarcticus	2-1-t-12 2 700 ± 5% Cyanobacteria Chroococcidiopsis sp.		2 2 5 49 4 820 900 ± 11% S-Mars R Bio	2-2-t-50 5 670 000 ± 9% P-Mars film	2-2-t-51 6 120 000 ± 9% Junar	2-2-t-52 5 770 000 ± 9% Iunar A-Biofilm	2-2-t-53 5 870 ± 9% S-Mars K Arc	2-2-t-54 6 130 ± 8% P-Mars Maea	2-2-t-55 52.7 ± 9% S-Mars ort Deinco	2-2-4-56 44.6 ± 8% S-Mars wt coccus		
2-1-t-13 4 920 000 ± 13% Lichens Cridinaria gyresa	2-1-t-14 5 480 000 ± 11% Lichens Buelia frigida	2-1-t-15 4 260 ± 10% Meristematic black fungi Cryomyces antarcticus	2-1-t-16 2 480 ± 7% Cyanobacteria Chroococcidiopsis sp.		2-2-1-57 4 550 000 ± 17% 5-Mars	2-2-t-58 5 600 000 ± 14% P-Mars	2-2-t-59 5 980 000 ± 14% Iunar	2-2-t-60 5 520 000 ± 14% Iunar	2-2-t-61 5 670 ± 14% S-Mars	2-2-t-62 5 960 ± 14% P-Mars	2-2-t-63 52.3 ± 14% S-Mars crt	2-2-t-64 41.3 ± 11% S-Mars wt		

FIG. 6. Final UV/PAR radiation dose distribution on the BIOMEX compartments within EXPOSE-R2 after exposure on the ISS (data provided by RedShift).

Sample category /		Resul	ts on limits of	life	Results on Habitability of Mars				
domain	species	selection tests	EVT/SVT	space	selection tests	EVT/SVT	space		
Archaea	Methanosarcina sp.strain SMA-21 (terrestrial permafrost) (GFZ/AWI Potsdam)	±			+				
Bacteria	Cyanobacterium <i>Chroococcidiopsis sp.</i> CCMEE 029 (Uni Roma)			+			+		
Fungi	Cryptoendolithic Antarctic black fungus Cryomyces antarcticus CCFEE 515 (Uni Viterbo)		+	±		+	±		
Lichens	<i>Circinaria gyrosa</i> (INTA) <i>Buellia frigida</i> (Antarctic lichen) (H-H-Uni Düsseldorf)			± ±			± ±		
Bryophytes	<i>Grimmia sessitana</i> (alpine samples) (Uni Potsdam)		±			±			
Biofilm	KOMBUCHA Biofilm containing: Yeasts: Saccharomyces ludwigii, Schizosaccharomyces pombe, Zygosaccharomyces rouxii, Zygosaccharomyces bailii, Brettanomyces bruxellensis; Bacteria: Paenibacillus sp. IMBG221, Acetobacter nitrogenifigens, Gluconacetobacter kombuchae sp. nov., Gluconacetobacter xylinum.(NAS Ukraine)			±			Ŧ		

TABLE 5. RESULTS LISTED ACCORDING TO THE TOPICS "LIMITS OF LIFE" AND HABITABILITY OF LIFE

(+) Survival / metabolically active / growth capacity, (±) partly survival, more damaged

were additionally applied to the samples of the methanogen archaeon *Methanosarcina soligelidi* SMA 21. In this case, these filters were used because of the specific nature of this organism's original habitat, which is situated within permafrost-affected soils and protected by soil particles with different grain sizes. *Deinococcus radiodurans* was covered by neutral density filters with a transmission of 0.01%. The measured and calculated data with regard to the final doses the samples experienced, which included UVA, UVB, UVC, PAR (photosynthetically active radiation), and Lyman alpha, are represented in Fig. 6 and were kindly provided by ESA via computations completed by the company RedShift Design and Engineering BVBA.

Significant variation was observed in the dose of UV among samples that were placed within sample sites not protected by filters. The observed variations could be explained by the dependence of the sample position in the hardware, which would have been exposed to a variety of shadowing effects during the orbit of the ISS. When filters were not used in the BIOMEX experiments, the final doses varied between 4.5×10^6 and 8.4×10^6 kJ/m². Specific organisms exposed without any neutral density filters include the epilithic lichens *Circinaria gyrosa* and *Buellia frigida*, the epilithically living bryophytes *Grimmia* sp. and *Marchantia polymorpha*, the iron bacteria biofilm, and the kombucha biofilm. Biomolecules exposed on the surface and embedded in the Mars analog mineral pellets also endured the same direct space conditions without any neutral density filters.

3. Overview of Results within This Special Collection

The results presented in this special collection are arranged to provide an overview of each step of the processes involved in the BIOMEX experiment, that is, from selecting

TABLE 6. DETAILED RESULT LIST EXPLAINING THE CLASSIFICATION SHOWN IN TABLE 5

Sample category /	spagios	F	Results on limits of	life	Results on Habitability of Mars				
domain	species	selection tests	EVT/SVT	space	selection tests	EVT/SVT	space		
Archaea	Methanosarcina sp.strain SMA-21 (terrestrial permafrost) (GFZ/AWI Potsdam)	Decrease of CH ₄ production rate on the used Mars-analog high concentration of perchlorate (not applied in BIOMEX space exposure)			CH ₄ production rate on the used Mars-analog minerals stable (but decrease on high concentations of perchlorates / not applied in BIOMEX space exposure)				
Bacteria	Cyanobacterium <i>Chroococcidiopsis</i> sp. CCMEE 029 (Uni Roma)			Survival and recovery on Mars analog minerals, low DNA-damage particularly in the more protected endolithically dark control areas			Survival and recovery on Mars analog minerals, low DNA-damage particularly in the more protected endolithically dark control areas		
Fungi	Cryptoendolithic Antarctic black fungus <i>Cryomyces</i> <i>antarcticus CCFEE 535</i> (Uni Viterbo)		On Mars-analog S-MRS with and without direct irradiation no significant decrease in growth and no DNA damage. On (non)irradiated samples with P-MRS slight decrease of growth and slight increase of DNA damage. Original sandstone material and LRA under simulated space and Mars conditions slightly affect the growth	On irradiated S-MRS 40 % still growing, on irradiated P-MRS 79 % growing. In Martian atmosphere without irradiation on S- MRS about 55 % growth and on P-MRS about less than 20 % growth. nall cases within both MRS significant membrane damage. On LRA no significant changings in growth and less damaged membranes		On Mars-analog S-MRS with and without direct irradiation no significant decrease in growth or increase of DNA damage and on (non)irradiated samples with P-MRS slight decrease of growth and slight increase of DNA damage.	On irradiated S-MRS 40 % still growing, on irradiated P- MRS 79 % growing. In Martian atmosphere without irradiation on S-MRS about 55 % growth and on P-MRS about less than 20 % growth.In all cases within both MRS significant membrane damage.		
Lichens	Circinaria gyrosa (INTA)			Quick moderate-high recovery of the PSII activity in the space dark control (vacuum and Mars atmosphere), where also morphology and DNA stability was observed. But irradiated samples under the same conditions were significantly affected.			Quick moderate-high recovery of the PSII activity in the space dark control (vacuum and Mars atmosphere), where also morphology and DNA stability was observed. But irradiated samples under the same conditions were significantly affected.		
	Buellia frigida (antarctic lichen) (H-H-Uni Düsseldorf)			In the cultivation assay only the space exposed algal symbionts are still able to grow and form colonies. These results indicate that the lichen Buellia frigida is partly able to survive low earth orbit conditions.			The post-exposed lichen symbionts show after LIVE/DEAD staining viability rates of up to 23.6% for the algal and up to 10.5% for the fungal symbiont. This means Mars is less habitable for Buelia frigida.		
Bryophytes	Grimmia sessitana (alpine samples) (Uni Potsdam)		The mosses were still vital after doses of radiation expected during the EXPOSE-R2 mission on the ISS. This earliest extant lineage of land plants is highly resistant to extreme abiotic simulated space and Mars- like conditions.			The mosses were still vital after doses of radiation expected during the EXPOSE-R2 mission on the ISS. This earliest extant lineage of land plants is highly resistant to extreme abiotic simulated space and Mars-like conditions.			
Biofilm	KOMBUCHA Biofilm containing: Years: Saccharomyces ludwigii, Schizosaccaromyces pombe, Zygosaccharomyces rouxii, Brettanomyces bruxellensis; Bacteria: Paenibacillus sp. IMBG221, Acetobacter nitrogenifigens, Gluconacetobacter kombuchae sp. nov., Gluconacetobacter sylinum.(NAS Ukraine)			After returning to Earth, the space-flown hacterial-yeast community recovered in two months. With the UV-irradiation of DNA, changes in the cellular membranes, and an inhibition of cellulose synthesis were observed. After a series of culture experiments, the revived communities restored partially their composition and the associated activities.			After returning to Earth, the space-flown bacterial-yeast community recovered in two momths. Within the UV- irradiated samples, a degradation of DNA, changes in the cellular membranes, and an inhibition of cellulose synthesis were observed. After a series of culture experiments, the communities restored partially their composition and the associated activities.		

S-MRS: sulfatic martian regolith. P-MRS: phyllosilicatic martian regolith. LRA: lunar regolith analog.

samples for simulation experiments to the final exposure experiments in space. Results from pretests on the chosen methanogenic archaeon of the genus Methanosarcina at the preselection level are reported in Serrano et al. (2019). This archaeon was chosen because of its relevance with regard to its potential for being metabolically active on Mars. Therefore, the first tests were designed to use different substrates that contained magnesium perchlorate to establish a Mars-relevant perchlorate environment and mineral mixture before attempting the next selection step of applying atmospheric and radiation-related environmental conditions within the preflight experiments EVTs and SVTs. With regard to the EVTs and SVTs, we present the results obtained by analysis of the fungus Cryomyces antarcticus (Pacelli et al., 2019) and the moss Grimmia sp. (Huwe et al., 2019). Results obtained after space exposure are shown from a series of analyses on the biofilm kombucha (Podolich *et al.*, 2019), the cyanobacterium Chroococcidiopsis (Billi et al., 2019), the cryptoendolithic Antarctic fungus Cryomyces antarcticus (Onofri et al., 2019), and the lichens Buellia frigida (Backhaus et al., this issue) and Circinaria gyrosa (de la Torre et al., not part of this issue). A rough summary of the different studies is given in Table 5, and more details are shown in Table 6. Survival, physiological activity, and growth capacity were detected in all organisms tested. However, life's vital functions decreased from slight to significant, and the reader is directed to specific articles in this collection for detailed discussion of these findings. Several of the selected archaea, bacteria, and heterogenic multilayered biofilms formed by a multitude of species were found to be the most resistant to simulated or direct space and Mars-like conditions. Less resistance and a significant decrease in cell numbers and vitality with regard to the Mars-like environment were shown for multicellular life-forms such as the tested fungus Cryomyces antarcticus (Onofri et al., 2019) and the lichens Buellia frigida (Backhaus et al., 2019) and Circinaria gyrosa (de la Torre Noetzel et al., 2019). The bryophyte Grimmia sp. was an exception (Huwe et al., 2019), but further analysis after space exposure might show whether this specific moss is also resistant to the conditions in space. Actual results in reference to the bryophytes were shown only for the preflight selection mode of EVT and SVT. Our results so far indicate that present Mars seems to be habitable for archaea and bacteria over longer timescales. However, a clearer understanding of the limits of life would be achievable with the implementation of extended space exposure experiments on the Moon, for example, with similar space exposure facilities as those used in the present study (see de Vera et al., 2012).

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Author Disclosure Statement

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Abbreviations Used

- BIOMEX = BIOlogy and Mars EXperiment
 - DLR = German Aerospace Center
 - EVTs = Experiment Verification Tests
 - ISS = International Space Station
 - MfN = Museum für Naturkunde
 - PAR = photosynthetically active radiation
 - SVTs = Scientific Verification Tests