



Cork Institute of Technology
SWORD - South West Open Research
Deposit

Articles

Biological Sciences

2014-07-14

Preparation and Biological Properties of Ring-Substituted Naphthalene-1-Carboxanilides

Eoghan Nevin

Department of Biological Sciences, Cork Institute of Technology

Rodney Govender

Department of Biological Sciences, Cork Institute of Technology

Jim O'Mahony


Department of Biological Sciences, Cork Institute of Technology

Aidan Coffey

Department of Biological Sciences, Cork Institute of Technology

Et. al.

Follow this and additional works at: <https://sword.cit.ie/dptbiosciart>

 Part of the [Biology Commons](#)

Recommended Citation

Gonec, T. et al., 2014. Preparation and Biological Properties of Ring-Substituted Naphthalene-1-Carboxanilides. *Molecules*, 19(7), pp.10386–10409. Available at: <http://dx.doi.org/10.3390/molecules190710386>.

This Article is brought to you for free and open access by the Biological Sciences at SWORD - South West Open Research Deposit. It has been accepted for inclusion in Articles by an authorized administrator of SWORD - South West Open Research Deposit. For more information, please contact sword@cit.ie.

Article

Preparation and Biological Properties of Ring-Substituted Naphthalene-1-Carboxanilides

Tomas Gonec¹, Jiri Kos¹, Eoghan Nevin², Rodney Govender², Matus Pesko³, Jan Tengler¹, Ivan Kushkevych⁴, Vendula Stastna⁴, Michal Oravec⁵, Peter Kollar⁴, Jim O'Mahony², Katarina Kralova⁶, Aidan Coffey² and Josef Jampilek^{1,*}

¹ Department of Chemical Drugs, Faculty of Pharmacy, University of Veterinary and Pharmaceutical Sciences, Palackeho 1/3, 61242 Brno, Czech Republic

² Department of Biological Sciences, Cork Institute of Technology, Bishopstown, Cork, Ireland

³ Department of Environmental Ecology, Faculty of Natural Sciences, Comenius University, Mlynska dolina Ch-2, 84215 Bratislava, Slovakia

⁴ Department of Human Pharmacology and Toxicology, Faculty of Pharmacy, University of Veterinary and Pharmaceutical Sciences, Palackeho 1/3, 61242 Brno, Czech Republic

⁵ Global Change Research Centre AS CR, Belidla 986/4a, 60300 Brno, Czech Republic

⁶ Institute of Chemistry, Faculty of Natural Sciences, Comenius University, Mlynska dolina Ch-2, 84215 Bratislava, Slovakia

* Author to whom correspondence should be addressed; E-Mail: josef.jampilek@gmail.com; Tel.: +420-541-562-926.

Received: 10 May 2014; in revised form: 13 July 2014 / Accepted: 14 July 2014 /

Published: 17 July 2014

Abstract: In this study, a series of twenty-two ring-substituted naphthalene-1-carboxanilides were prepared and characterized. Primary *in vitro* screening of the synthesized carboxanilides was performed against *Mycobacterium avium* subsp. *paratuberculosis*. *N*-(2-Methoxyphenyl)naphthalene-1-carboxamide, *N*-(3-methoxyphenyl)naphthalene-1-carboxamide, *N*-(3-methylphenyl)naphthalene-1-carboxamide, *N*-(4-methylphenyl)naphthalene-1-carboxamide and *N*-(3-fluorophenyl)naphthalene-1-carboxamide showed against *M. avium* subsp. *paratuberculosis* two-fold higher activity than rifampicin and three-fold higher activity than ciprofloxacin. The most effective antimycobacterial compounds demonstrated insignificant toxicity against the human monocytic leukemia THP-1 cell line. The testing of biological activity of the compounds was completed with the study of photosynthetic electron transport (PET) inhibition in isolated spinach (*Spinacia oleracea* L.) chloroplasts. The PET-inhibiting activity expressed

by IC₅₀ value of the most active compound *N*-[4-(trifluoromethyl)phenyl]naphthalene-1-carboxamide was 59 µmol/L. The structure-activity relationships are discussed.

Keywords: naphthalene; lipophilicity; *in vitro* antimycobacterial activity; *in vitro* cytotoxicity; photosynthetic electron transport inhibition; spinach chloroplasts

1. Introduction

Mycobacterium tuberculosis is the pathogen responsible for tuberculosis (TB). This disorder is more prevalent in the world today than at any other time. The number of notified cases of multi-drug-resistant strains of *M. tuberculosis* (MDR-TB) is increasing in 27 high MDR-TB burden countries. In addition, other non-tuberculous mycobacteria (NTM) are now recognized as significant human pathogens and cause difficult-to-treat or incurable diseases, such as pulmonary disease, lymphadenitis, skin and soft tissue disease, gastrointestinal and skeletal infections, and potentially resulting in death, especially in cases involving immunocompromised patients. The emergence of NTM underlines the urgency of searching for new structure types of drugs; it also seems to be important to re-engineer and re-test old drug families to achieve effective control of both TB and NTM [1–3].

As biologically active compounds, including drugs and pesticides, target particular biological systems, some herbicides can also have molecular sites of action in mammals/non-plant organisms. However, targeting compounds to biological systems with similar physicochemical properties can lead to completely different biological responses in plants and animals. Therefore, many innovative/original pharmaceutical companies perform screening of biologically active compounds, not only as potential drugs, but also as potential pesticides. For example, fluconazole was firstly discovered as a potent pesticide, and subsequently it was confirmed as an antifungal drug [4–6]. Moreover, good correlation between microbiological activities and herbicidal or antialgal effects was found [7–18].

The presence of an amide (-NHCO-) group [19,20] is characteristic of a number of biologically active compounds, such as antimicrobials, antiprotozoals, antineoplastics, antivirals, anti-inflammatory agents or herbicides [7,9,12,13,21–28]. The wide spectrum of biological effects of substituted (mono/diaza)naphthalene scaffolds includes especially anti-infective and antineoplastics activity [9–12,21,29–42]. In addition, some (aza)naphthalene derivatives also showed noteworthy herbicidal activity [9–13,24,30,32,34] explained by reversible binding to photosystem II (PS II), a membrane-protein complex in the thylakoid membranes, which catalyses the oxidation of water and the reduction of plastoquinone [43], and, thereby, inhibit photosynthesis [44–46].

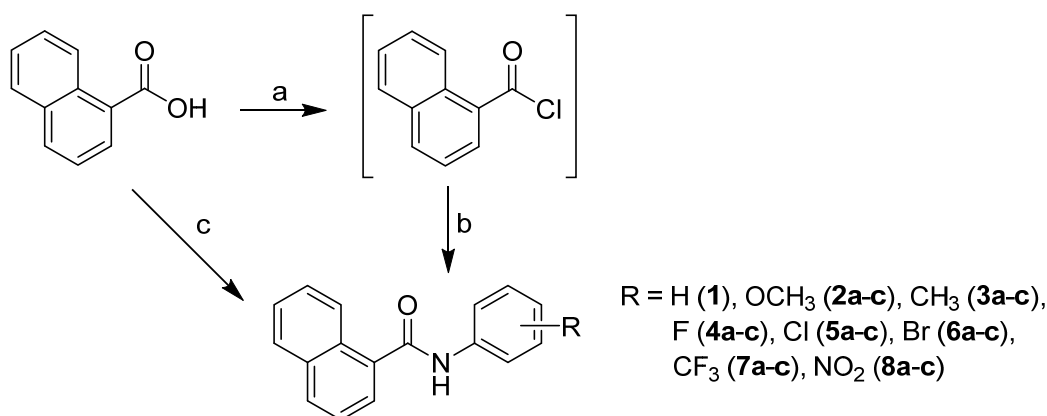
In the context of the previously-described azanaphthalenes [24,30–37] or various amides [8–12,25–28], ring-substituted naphthalene-1-carboxanilides as biologically active compounds were investigated. All the compounds were tested for activity against NTM species—*M. avium* subsp. *paratuberculosis* and additionally they were assessed for their potency to inhibit photosynthetic electron transport in isolated spinach chloroplasts (*Spinacia oleracea* L.). Relationships between the structure and their *in vitro* antimycobacterial activity or/and activity related to inhibition of photosynthetic electron transport (PET) in isolated spinach chloroplasts are discussed.

2. Results and Discussion

2.1. Chemistry

The studied compounds were prepared using two methods according to Scheme 1. The discussed compounds were synthesized by using two-step synthesis (Method A: a,b) via 1-naphthoyl chloride as intermediate [9] or by one-pot microwave-assisted synthesis (Method B: c) [10]. Both methods yielded a series of twenty-two *N*-substituted naphthalene-1-carboxanilides **1–8c**.

Scheme 1. Synthesis of ring-substituted naphthalene-1-carboxanilides **1–8c**.



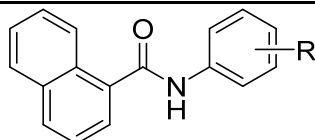
Reagents and conditions: (a) SOCl₂, toluene; (b) triethylamine, dichloromethane, (substitute) aniline; (c) PCl₃, chlorobenzene, (substitute) aniline, MW.

2.2. Lipophilicity Characteristics and Electronic Properties

Within structure-activity relationship investigations of various molecular descriptors, topological indexes or parameters describing physicochemical properties are used. Thus, they represent numerical values that characterize properties of molecules that influence biological activity (receptor binding). In our previous studies biological activity was strongly influenced especially by lipophilicity and electronic parameters [7–13,24–28,30–37], expressed either as Hammett's σ parameters or Taft polar substituent constants σ^* .

Lipophilicity is a property that has a major effect on bioavailability, biotransformation as well as pharmacological activity, because drugs cross biological membranes through passive transport, which strongly depends on their lipophilicity. Lipophilicity of the studied compounds was determined by RP-HPLC as capacity factor logarithm ($\log k$). The procedure was performed under isocratic conditions with methanol as an organic modifier in the mobile phase using an end-capped non-polar C₁₈ stationary RP column. The results of ring-substituted naphthalene-1-carboxanilides **1–8c** are shown in Table 1 and illustrated in Figure 1. The highest experimental lipophilicity was found for *N*-[4-(trifluoromethyl)-phenyl]naphthalene-1-carboxamide (**7c**), while *N*-(4-methoxyphenyl)naphthalene-1-carboxamide (**2c**) showed the lowest $\log k$ value. Lipophilicity was also calculated as $\log P$ using ACD/Percepta ver. 2012 (Advanced Chemistry Development, Inc., Toronto, ON, Canada).

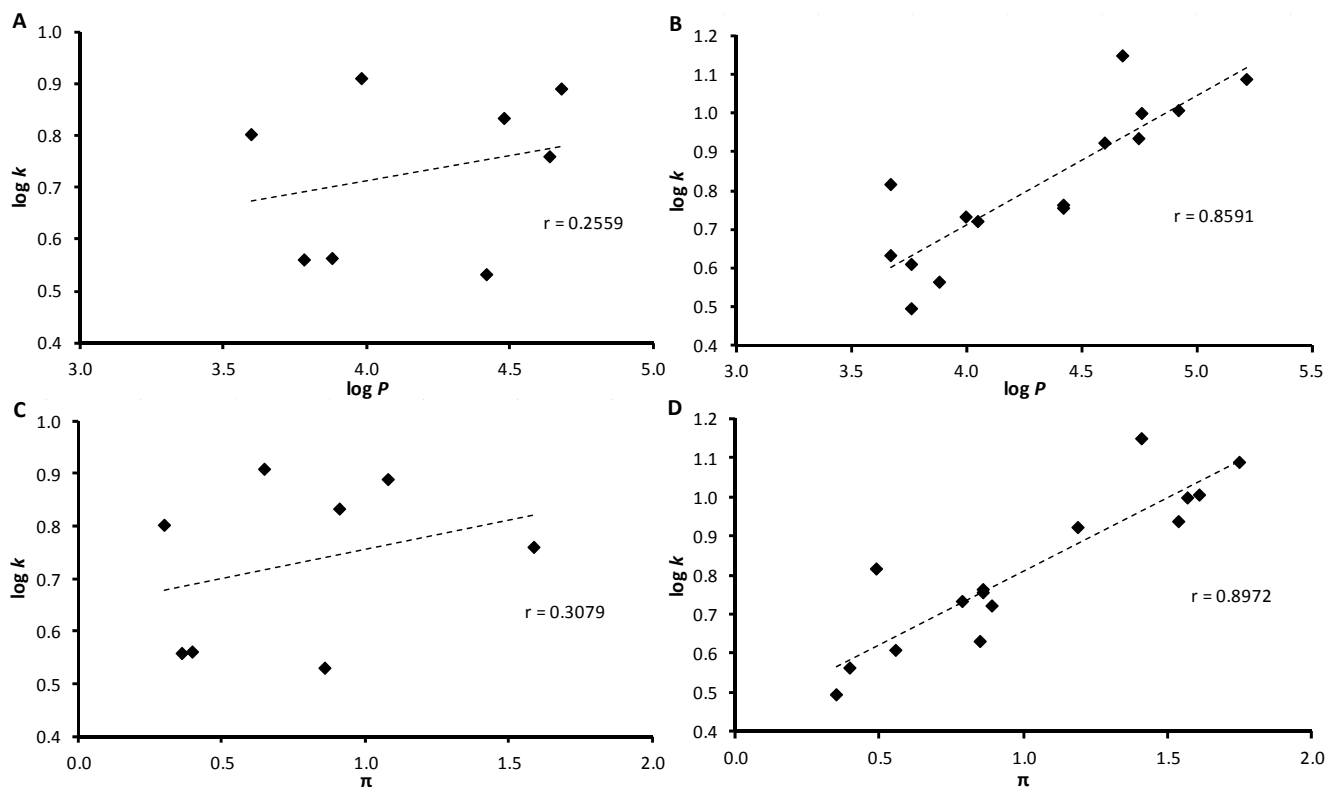
Table 1. Structure of ring-substituted naphthalene-1-carboxanilides **1–8c**, experimentally determined values of lipophilicity $\log k$, calculated values of lipophilicity ($\log P$), predicted hydrophobic distributive parameter π , predicted polar substituent constants σ^* , *in vitro* antimycobacterial activity (MIC/IC₉₀) of compounds against clinical isolate of *Mycobacterium avium* subsp. *paratuberculosis* CIT03 (MAP) in comparison with isoniazid (INH), ciprofloxacin (CPX), and rifampicin (RIF) standards, *in vitro* cytotoxicity assay (LD₅₀) of choice compounds and IC₅₀ values related to PET inhibition in isolated spinach chloroplasts in comparison with 3-(3,4-dichlorophenyl)-1,1-dimethylurea (DCMU) standard.



Comp.	R ¹	$\log k$	$\log P^a$	π^a	σ^{*a}	MAP MIC/IC ₉₀ [$\mu\text{mol/L}$]	LD ₅₀ [$\mu\text{mol/L}$]	PET IC ₅₀ [$\mu\text{mol/L}$]
1	H	0.5620	3.88	0.40	0.26	<i>b</i>	—	<i>c</i>
2a	2-OCH ₃	0.8013	3.60	0.30	0.29	54	>30	<i>c</i>
2b	3-OCH ₃	0.6087	3.76	0.56	0.30	54	>30	<i>c</i>
2c	4-OCH ₃	0.4953	3.76	0.35	0.38	<i>b</i>	—	<i>c</i>
3a	2-CH ₃	0.5309	4.42	0.86	0.29	<i>b</i>	—	<i>c</i>
3b	3-CH ₃	0.7624	4.42	0.86	0.30	57	>30	136
3c	4-CH ₃	0.7551	4.42	0.86	0.38	57	>30	381
4a	2-F	0.5592	3.78	0.36	0.31	<i>b</i>	—	<i>c</i>
4b	3-F	0.7204	4.05	0.89	0.32	57	>30	131
4c	4-F	0.6315	3.67	0.85	0.39	<i>b</i>	—	<i>c</i>
5a	2-Cl	0.8322	4.48	0.91	0.33	<i>b</i>	—	<i>c</i>
5b	3-Cl	0.9357	4.75	1.54	0.32	<i>b</i>	—	107
5c	4-Cl	0.9223	4.60	1.19	0.40	<i>b</i>	—	102
6a	2-Br	0.8892	4.68	1.08	0.31	<i>b</i>	—	<i>c</i>
6b	3-Br	1.0047	4.92	1.61	0.31	<i>b</i>	—	278
6c	4-Br	0.9985	4.76	1.57	0.40	184	—	365
7a	2-CF ₃	0.7595	4.64	1.59	0.27	<i>b</i>	—	<i>c</i>
7b	3-CF ₃	1.0862	5.22	1.75	0.33	<i>b</i>	—	114
7c	4-CF ₃	1.1480	4.68	1.41	0.41	793	—	59
8a	2-NO ₂	0.9090	3.98	0.65	0.27	<i>b</i>	—	476
8b	3-NO ₂	0.8155	4.00	0.79	0.34	<i>b</i>	—	492
8c	4-NO ₂	0.7328	3.67	0.49	0.43	<i>b</i>	—	129
DCMU	—	—	—	—	—	—	—	1.9
INH	—	—	—	—	—	1823	—	—
CPX	—	—	—	—	—	181	—	—
RIF	—	—	—	—	—	109	—	—

^a calculated using ACD/Percepta (Advanced Chemistry Development, Inc., Toronto, ON, Canada, 2012); π : hydrophobicity contribution of the substituent in position 1 of naphthalene core, σ^* : electron properties contribution of the substituent in position 1 of naphthalene core; ^b no activity; ^c precipitation during experiment.

Figure 1. Match of experimentally found $\log k$ values with calculated $\log P$ data of *ortho*-substituted (A) and *meta*- and *para*-substituted derivatives (B) and with calculated distributive parameter π (hydrophobicity contribution of the substituent in position 1 of naphthalene) of *ortho*-substituted (C) and *meta*- and *para*-substituted derivatives (D).



The results obtained with all the ring-substituted naphthalene-1-carboxanilides show that the experimentally-determined lipophilicity ($\log k$) of the discussed compounds poorly correlates with the calculated values $\log P$ of compounds **1–8c** as shown in Figure 1A,B. For the *ortho*-substituted derivatives no correlation was found. Similar poor correlation between experimentally and predicted lipophilicity values was described for ring-substituted 2-hydroxynaphthalene-1-carboxanilides and 1-hydroxynaphthalene-2-carboxanilides [11,12] and for hydroxyquinolinecarboxamides [24]; nevertheless, this poor correlation was attributed to the phenolic moiety in the position vicinal to the carboxamide group. As there is no any phenolic moiety in these structures, lipophilicity was additionally expressed as a hydrophobic distributive parameter π of the carboxamide moiety and ring-substituted phenyl; it was also calculated using ACD/Percepta, see Table 1. Again, it can be stated that no match of $\log k$ to π of the *ortho*-substituted compounds was found (Figure 1C), while the *meta*- and *para*-substituted derivatives showed better match, see Figure 1D. The most significant deviations within the dependence illustrated in Figure 1D can be observed for 4-CF₃ (**7c**) and 4-NO₂ (**8c**).

Based on these observations, some intramolecular interactions between the carboxamide moiety and the phenyl substituents could be supposed that influence the resulting lipophilicity of compounds. These interactions are characterized rather by using distributive parameter π of substituents in position 1 of naphthalene than by using $\log P$ parameter of the whole molecule. Differences between expected and found values were also observed during TLC, when the purity of the final compounds was

checked. *Ab initio*/DFT calculations of charges in individual structures could provide more detailed information, nevertheless, such investigation was not the aim of this paper.

The influence of R substituents on lipophilicity ($\log k$) is as follows within *ortho*-substituted derivatives: $\text{CH}_3 < \text{F} = \text{H} < \text{CF}_3 < \text{OCH}_3 < \text{Cl} < \text{Br} < \text{NO}_2$ and as follows within *meta*- and *para*-substituted derivatives: $\text{OCH}_3 \approx \text{H} < \text{F} < \text{CH}_3 < \text{NO}_2 < \text{Cl} < \text{Br} < \text{CF}_3$. Within the individual series the lipophilicity determined by $\log k$ values increases for halogens and methyl substituents as follows: *ortho* < *para* < *meta*; for OCH_3 and NO_2 as follows: *para* < *meta* < *ortho*. It can be assumed that $\log k$ values specify lipophilicity within individual series of the studied compounds.

Based on the above-mentioned observations, electronic properties of prepared compounds **1–8c** were expressed as polar substituent constants σ^* of the whole substituent in position 1 of naphthalene, *i.e.*, influence of carboxamide moiety and ring-substituted phenyl was included. The individual σ^* parameters were predicted using the ACD/Percepta software; they ranged from 0.26 (**1**, R = H) to 0.43 (**8c**, R = 4- NO_2).

2.3. In Vitro Antimycobacterial Evaluation

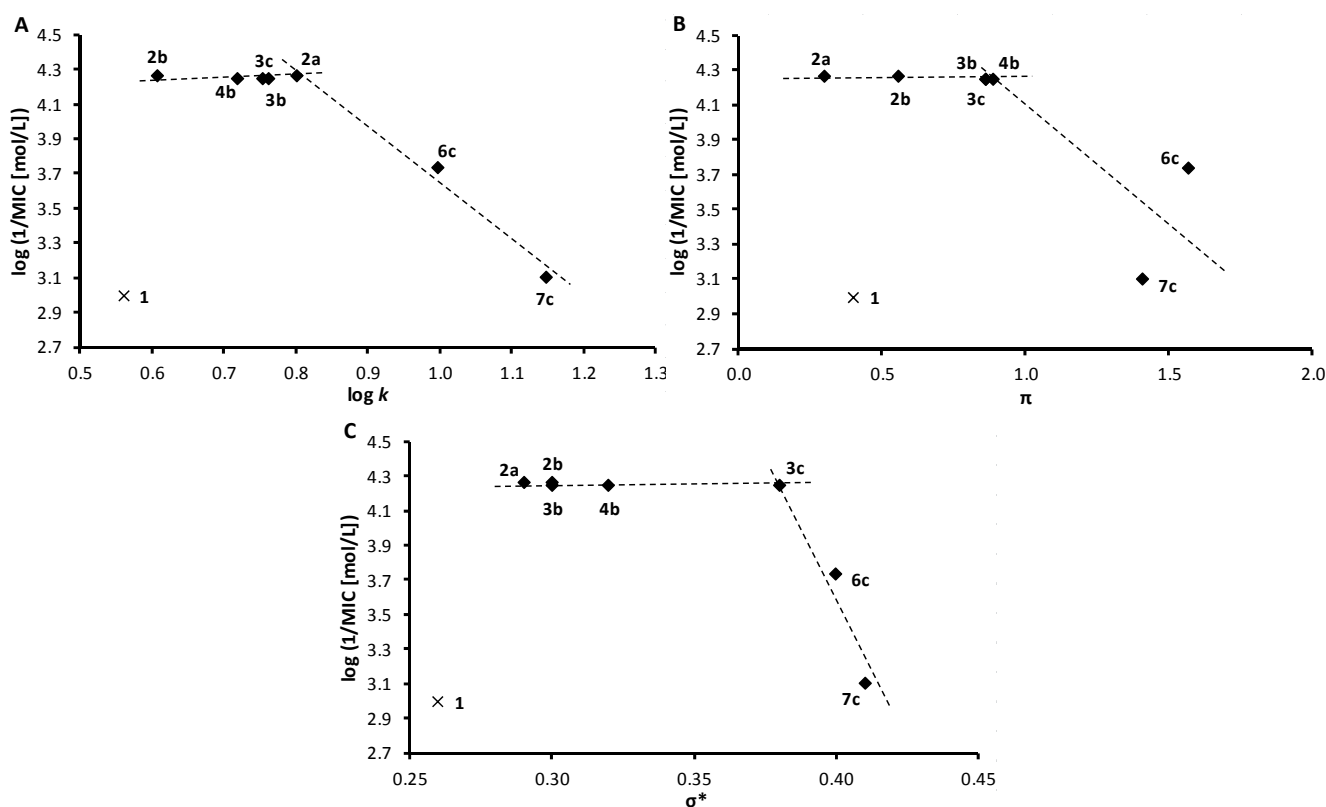
In vitro antimycobacterial screening of all the compounds against *Mycobacterium avium* subsp. *paratuberculosis* was performed, however most compounds did not show any activity (higher than 793 $\mu\text{mol/L}$). Only the eight compounds presented in Table 1 showed antimycobacterial effectivity. The antimycobacterial activity was expressed as minimum inhibitory concentration (MIC) that is defined for mycobacteria as a 90% or greater (IC_{90}) reduction of growth in comparison with the control. The MIC/ IC_{90} value is routinely and widely used in bacterial assays and is a standard detection limit according to the Clinical and Laboratory Standards Institute [47]. *N*-(2-Methoxyphenyl)naphthalene-1-carboxamide (**2a**), *N*-(3-methoxyphenyl)naphthalene-1-carboxamide (**2b**), *N*-(3-methylphenyl)naphthalene-1-carboxamide (**3b**), *N*-(4-methylphenyl)-naphthalene-1-carboxamide (**3c**) and *N*-(3-fluorophenyl)naphthalene-1-carboxamide (**4b**) demonstrated two-fold higher activity than rifampicin and three-fold higher activity than ciprofloxacin.

Figure 2 shows dependence of the antimycobacterial activities expressed as $\log(1/\text{MIC} [\text{mol/L}])$ on lipophilicity expressed as $\log k$ (Figure 2A), distributive parameter π (Figure 2B) and electronic properties expressed as polar substituent constants σ^* (Figure 2C). When inactive compound **1** (R = H) was eliminated (marked by cross), the dependences of $\log(1/\text{MIC})$ on $\log k$ and of $\log(1/\text{MIC})$ on σ^* were biphasic: in the range of π from 0.6087 (**2b**) to 0.8013 (**2a**), as well as in the range of σ^* from 0.29 (**2a**) to 0.38 (**3c**) the antimycobacterial activity remained practically constant while a further increase of $\log k$ or σ^* resulted in a strong activity decrease. Consequently, it can be concluded that the lipophilicity of compounds higher than $\log k = 0.8013$ is adverse in terms of antimycobacterial activity; increased electron-withdrawing properties of the whole substituent in position 1 of naphthalene (expressed as polar substituent constants σ^*) exceeding value $\sigma^* = 0.38$ are also unfavourable.

When antimycobacterial activities of both series of naphthalene-2-carboxanilides [9] and naphthalene-1-carboxanilides are compared, it can be stated that the discussed 1-carboxanilides demonstrated significantly higher activity. Comparison of 2-hydroxynaphthalene-1-carboxanilides [11] with naphthalene-1-carboxanilides is more complicated. It can be stated that 2-hydroxy derivatives showed wider spectrum of antimycobacterial effect; nevertheless when MICs of compounds and the

standards are compared, it seems that compounds presented herein demonstrated more potent activity. In general it can be stated that in both series derivatives with lipophilicity $\log k > 0.6$ can show antimycobacterial effect, and within both series rather *meta*-substituted derivatives demonstrated higher activity, which can be connected with the planarity of the structure [9,11,12], nevertheless in the series of 2-hydroxynaphthalene-1-carboxanilides substituents with significant electron-withdrawing effect are favoured [11], while among naphthalene-1-carboxanilides rather electron-neutral or slightly electron-withdrawing substituents are preferable, *i.e.*, phenyl substituents with minimal effect on electronic density at the carboxamide moiety [10–12].

Figure 2. Dependence of *in vitro* antimycobacterial activity against *Mycobacterium avium* subsp. *paratuberculosis* $\log(1/\text{MIC} [\text{mol/L}])$ on lipophilicity expressed as $\log k$ (A) or distributive parameter π (B), as well as polar substituent constants σ^* (C) of selected studied compounds. (π : hydrophobicity contribution of the substituent in position 1 of naphthalene, σ^* : electron properties contribution of the substituent in position 1 of naphthalene; eliminated compound **1** marked by cross)

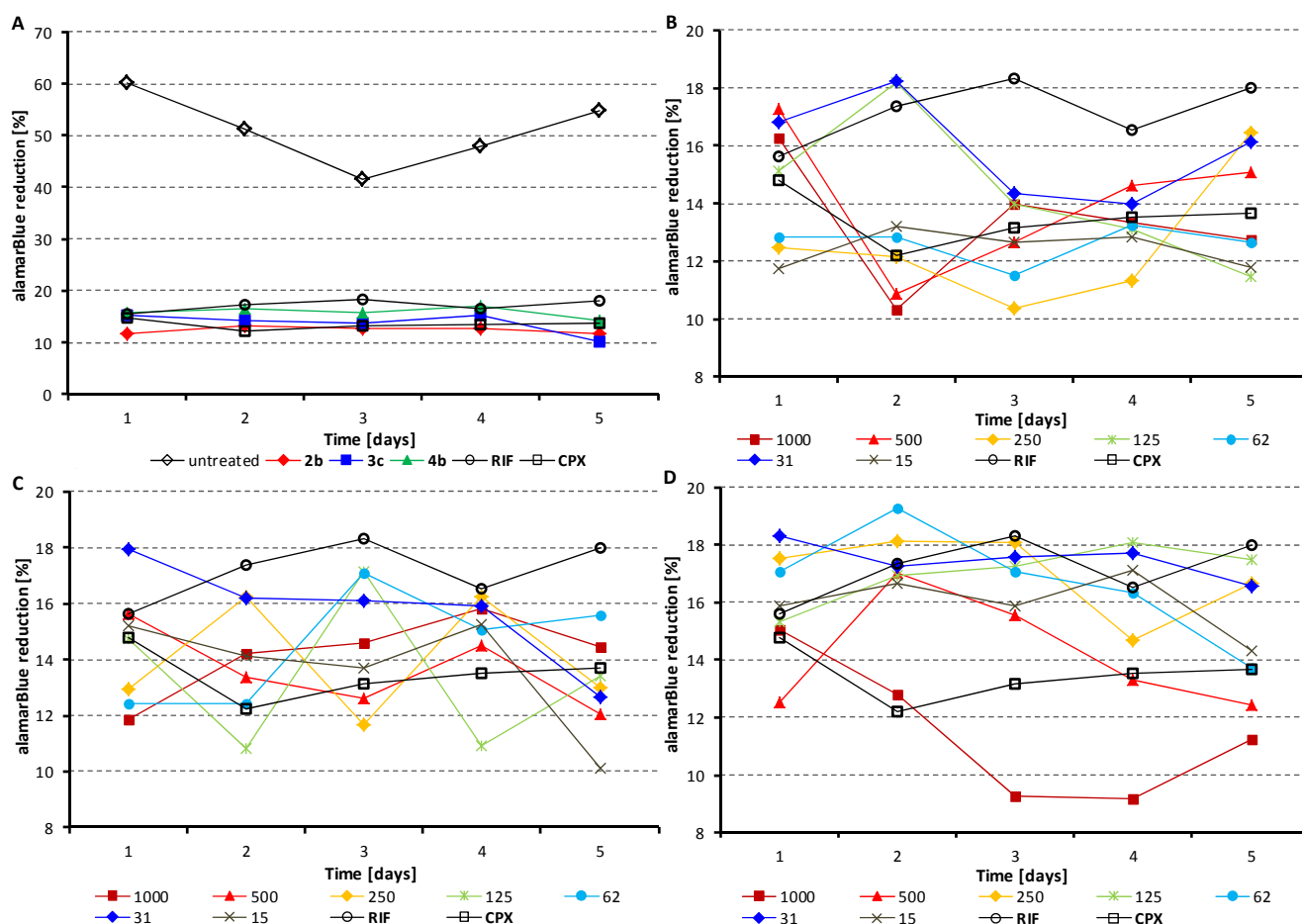


Of the eight compounds the three with the best efficacy were used to monitor their effect on the viability of *M. avium* subsp. *paratuberculosis* CIT03 over a period of five days, see Figure 3. A concentration range starting at 1,000 $\mu\text{g/mL}$ to 15 $\mu\text{g/mL}$ was used for compounds **2b**, **3c**, and **4b**. The percent reduction of alamarBlue of these three compounds was compared to an untreated control of *M. avium* subsp. *paratuberculosis* and the standards rifampicin and ciprofloxacin. Figure 3A illustrates the dependence of the percent reduction of alamarBlue of compounds **2b**, **3c**, and **4b** at their MICs (15 $\mu\text{g/mL}$, *i.e.*, 54, 57, and 57 $\mu\text{mol/L}$) on time in comparison with ciprofloxacin (CPX) and rifampicin (RIF) standards at their MICs (60 and 90 $\mu\text{g/mL}$, *i.e.*, 181 and 109 $\mu\text{mol/L}$) and the

untreated control of *M. avium* subsp. *paratuberculosis*. Figure 3B–D show in detail individual dependences of the percent reduction of alamarBlue on time (in days) at various concentrations of the individual compounds (range from 1000 to 15 $\mu\text{g/mL}$) in comparison with the standards at their MICs. The percent reduction values for all treated *M. avium* subsp. *paratuberculosis* samples were less than 20% over the five-day period of testing. As previously outlined by Carroll *et al.* [48], percent reduction values less than 20% indicate insufficient cell metabolism and hence lack of cell viability.

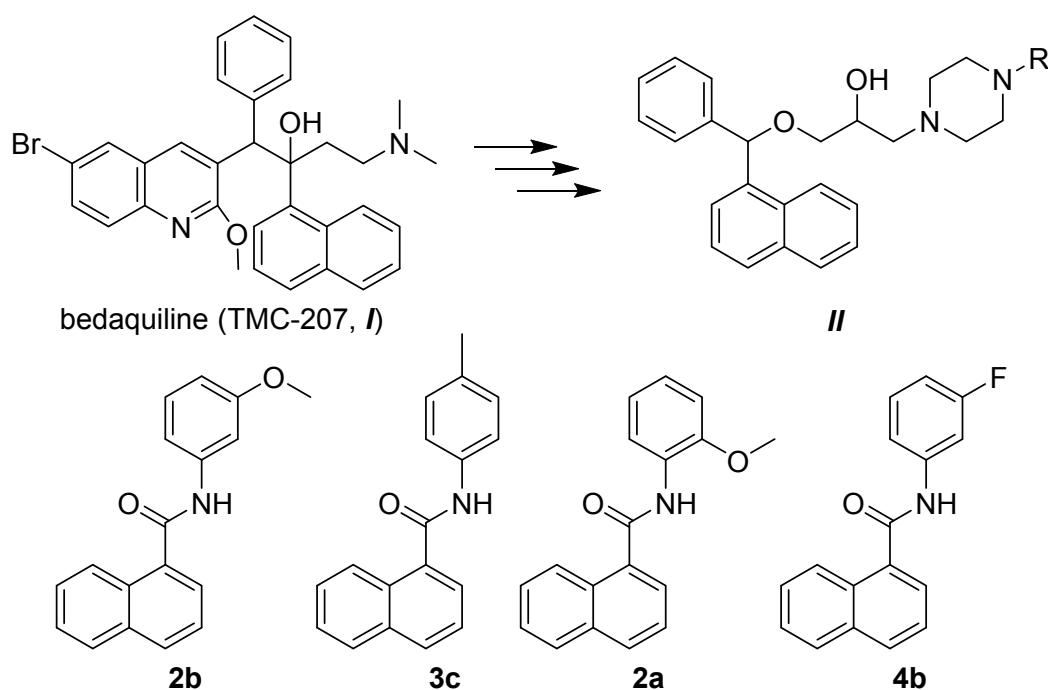
Additionally, a standard MTT assay was performed on these same compounds. The MTT assay is a well-characterized method of assessing cell growth through measurement of respiration. As such, a low level of cell viability may suggest inhibition of cell growth through respiratory inhibition [49]. 3-Methoxy (**2b**) and 4-methyl (**3c**) derivatives showed more than 80% reduction in activity at the lowest tested concentration (8 $\mu\text{g/mL}$, *i.e.*, *ca.* 30 $\mu\text{mol/L}$) after four hours of incubation, similar to the reduction observed in the rifampicin and ciprofloxacin standards. Compounds **2a** and **4b** achieved similar levels of inhibition at 16 $\mu\text{g/mL}$ (*ca.* 60 $\mu\text{mol/L}$) concentration.

Figure 3. Viability of *M. avium* subsp. *paratuberculosis* expressed as percent reduction of alamarBlue over a period of five days after treatment with selected studied compounds **2b**, **3c**, and **4b** in comparison with untreated control of *M. avium* subsp. *paratuberculosis* and standards ciprofloxacin (CPX) and rifampicin (RIF) at their MICs (A). Viability of *M. avium* subsp. *paratuberculosis* expressed as percent reduction of alamarBlue over a period of five days after treatment with selected studied compounds **2b** (B), **3c** (C), and **4b** (D) at different concentrations (1,000–15 $\mu\text{g/mL}$) in comparison with CPX and RIF at their MICs.



Janin mentioned that a similar type of carboxamides can interfere with the mycobacterial proton pump $F_0F_1H^+ATPase$ or inhibit biosynthesis of amino acids [50]. This hypothesis was confirmed in recently published papers [3,51,52], where the mechanism of action of bedaquiline (TMC-207, **I**) and its naphthalene analogues (**II**) as potent compounds inhibiting the respiratory chain of mycobacteria is discussed. Based on structure analogy, the discussed ring-substituted naphthalene-1-carboxanilides can be considered as simplified derivatives of bedaquiline and its naphthalene analogues, see Figure 4. Since the change in colour of alamarBlue is caused by a decrease of mycobacterial cell metabolism, it is possible that these compounds bind to the mycobacterial respiratory chain components. However, another possible site of action of the studied compounds in the mycobacteria cannot be excluded.

Figure 4. Structural analogy of the most effective compounds **2a**, **2b**, **3c**, **4b** to bedaquiline (TMC-207, **I**) and its naphthalene analogues (**II**) [3,50].



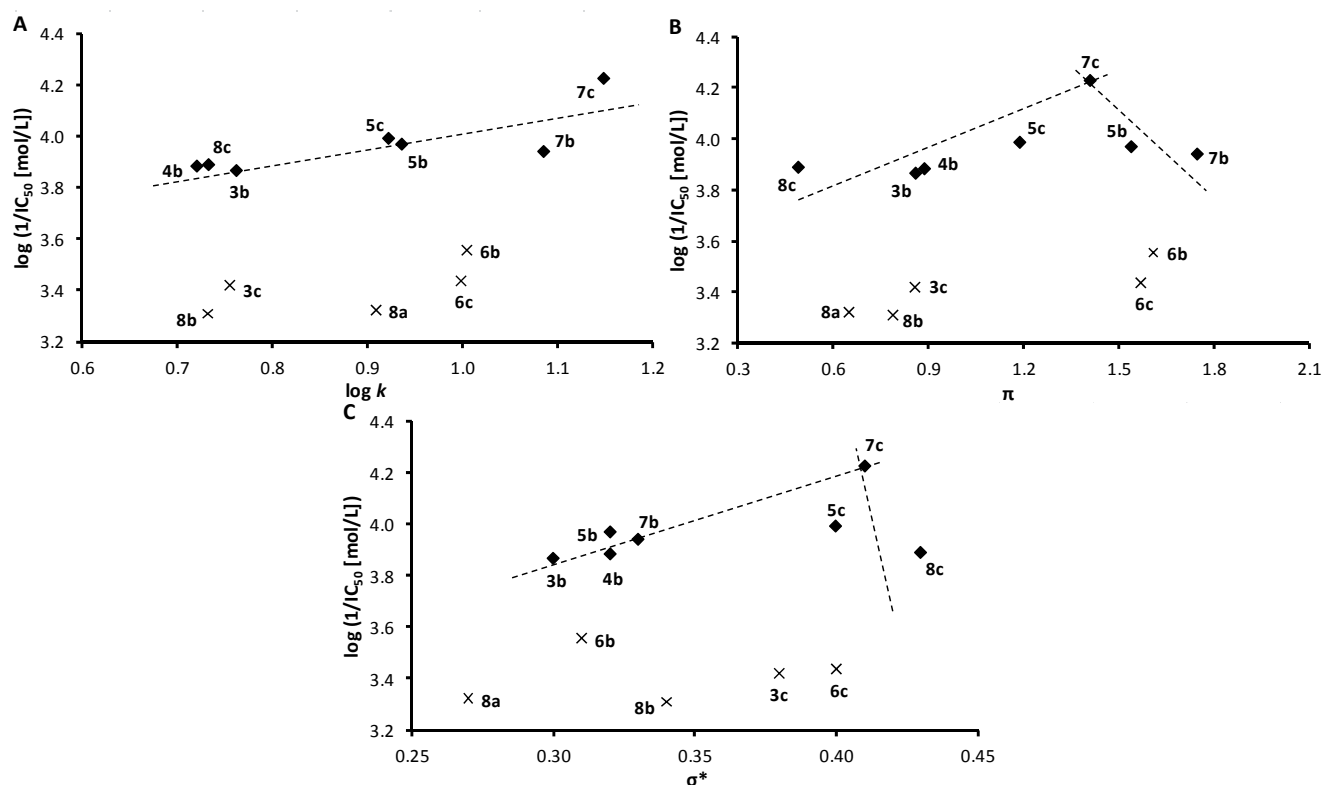
2.4. In Vitro Cytotoxicity Assay

The preliminary *in vitro* screening of cytotoxicity of the most effective antimycobacterial compounds **2a**, **2b**, **3b**, **3c**, and **4b** was performed using the human monocytic leukemia THP-1 cell line. The cytotoxicity was evaluated as the LD_{50} value (LD_{50} -lethal dose to 50% of the cell population), see Table 1. The discussed anilides **2a**, **2b**, **3b**, **3c** and **4b** were not soluble at concentrations above 30 $\mu\text{mol/L}$ in testing medium containing 0.1% DMSO, therefore the highest available concentration 30.0 $\mu\text{mol/L}$ was used for the toxicity assay. Treatment with 30 $\mu\text{mol/L}$ of **2a**, **2b**, **3b**, **3c**, and **4b** did not lead to significant lethal effect on THP-1 cells (e.g., LD_{50} of oxaliplatin and of camptothecin assessed in this line formerly showed similar values: $1.7 \pm 0.64 \mu\text{mol/L}$ and $0.16 \pm 0.07 \mu\text{mol/L}$, respectively). Based on these observations it can be concluded that the discussed anilides with antimycobacterial efficacy are nontoxic [53] and therefore they can be considered as promising agents for subsequent design of novel antimycobacterial agents.

2.5. Inhibition of Photosynthetic Electron Transport (PET) in Spinach Chloroplasts

The evaluated naphthanilide derivatives showed moderate or low inhibition of photosynthetic electron transport (PET) in isolated spinach (*Spinacia oleracea* L.) chloroplasts compared with the standard, see Table 1. The PET-inhibiting activity was expressed by IC_{50} value, *i.e.*, compound concentration in mol/L causing 50% inhibition of PET. Generally compounds showed poor aqueous solubility and IC_{50} values of **1–3a**, **4a**, **4c**, **5a**, **6a**, and **7a** could not be determined due to precipitation of the compounds during the experiments. From rest twelve compounds compound **7c** ($R = 4-CF_3$) expressed the highest PET-inhibiting activity ($IC_{50} = 59 \mu\text{mol/L}$), while compound **8b** ($R = 3-NO_2$) expressed the lowest PET-inhibiting activity ($IC_{50} = 492 \mu\text{mol/L}$). However, it could be noted that limited solubility in tested medium at higher compound concentrations was observed also for **3c**, **6b**, **6c**, **8a**, and **8b**.

Figure 5. Relationships between PET inhibition $\log(1/IC_{50})$ [mol/L] in isolated spinach chloroplasts and lipophilicity expressed as $\log k$ (A) or distributive parameter π (B), as well as polar substituent constants σ^* (C) of selected studied compounds. (π : hydrophobicity contribution of the substituent in position 1 of naphthalene, σ^* : electron properties contribution of the substituent in position 1 of naphthalene; eliminated compounds **3c**, **6b**, **6c**, **8a**, **8b** marked by crosses)



Dependences of $\log(1/IC_{50})$ on the lipophilicity of compounds expressed as $\log k$ (Figure 5A) as well as distributive parameter π (Figure 5B) and electronic properties expressed as polar substituent constants σ^* (Figure 2C) are presented in Figure 2. When compounds with reduced solubility in tested medium, namely **3c** ($R = 4-CH_3$), **6b,c** ($R = 3-Br, 4-Br$), and **8a,b** ($R = 2-NO_2, 3-NO_2$) were eliminated (marked by crosses) the dependence of $\log(1/IC_{50}$ [mol/L]) on $\log k$ illustrated for seven compounds

(**3b**, **4b**, **5b**, **5c**, **7b**, **7c**, **8c**) seems to be approximately linear. On the other hand, the dependence of PET-inhibiting activity on distributive parameter π of the whole substituents in position 1 of naphthalene showed bilinear dependence, the activity increased up to $\pi = 1.41$ (R = 4-CF₃, compound **7c**) and further lipophilicity increase resulted in activity decrease. The dependence of PET inhibition on the electronic properties σ^* of the whole substituents in position 1 of naphthalene, showed also biphasic course with the optimum $\sigma^* = 0.41$ (compound **7c**).

When the PET-inhibiting activity is compared with that of 2-hydroxynaphthalene-1-carboxanilides [11], it can be stated that 2-hydroxy derivatives showed higher solubility and higher activity, nevertheless similar relationships between PET inhibition and lipophilicity and electronic parameters of substituents were observed. Completely different dependences of PET inhibition on lipophilicity and electronic parameters of substituents were found within the series of naphthalene-2-carboxanilides [9].

3. Experimental Section

3.1. General

All reagents were purchased from Aldrich. TLC experiments were performed on alumina-backed silica gel 40 F254 plates (Merck, Darmstadt, Germany) using chloroform/acetone (2:1) as a mobile phase. The plates were illuminated under UV (254 nm) and evaluated in iodine vapor. The melting points were determined on Kofler hot-plate apparatus HMK (Franz Kustner Nacht KG, Dresden, Germany) and are uncorrected. Infrared (IR) spectra were recorded on a Smart MIRacle™ ATR ZnSe for Nicolet™ Impact 410 FT-IR spectrometer (Thermo Scientific, West Palm Beach, FL, USA). The spectra were obtained by accumulation of 256 scans with 2 cm⁻¹ resolution in the region of 4,000–600 cm⁻¹. All ¹H and ¹³C-NMR spectra were recorded on a Bruker Avance III 400 MHz FT-NMR spectrometer (400 MHz for ¹H and 100 MHz for ¹³C, Bruker Comp., Karlsruhe, Germany). Chemical shifts are reported in ppm (δ) using internal Si(CH₃)₄ as the reference with diffuse, easily exchangeable signals being omitted. Mass spectra were measured using a LTQ Orbitrap Hybrid Mass Spectrometer (Thermo Electron Corporation, USA) with direct injection into an APCI source (400 °C) in the positive mode.

3.2. Synthesis

General Procedure for Synthesis of Carboxamide Derivatives **1–8c**

Method A: Naphthalene-1-carboxylic acid (5.8 mmol) was dissolved in dry hot toluene (40 mL) and thionyl chloride (8.0 mmol) was added. After 2 h of refluxing, solvent and excessive thionyl chloride were evaporated under reduced pressure giving naphthalene-1-carbonyl chloride as colourless liquid in quantitative yield. The product forms colourless needles after cooling. Mp. 18–19 °C (26 °C [54]). Naphthalene-1-carbonyl chloride (5.8 mmol), triethylamine (8.7 mmol) and corresponding substituted aniline (5.8 mmol) were dissolved in dry dichloromethane (30 mL) and the mixture was stirred for 12 h at ambient temperature. The solvent was evaporated under reduced pressure, the solid residue washed with 10% HCl, and the crude product was recrystallized from propan-2-ol with addition of active carbon.

Method B: Naphthalene-1-carboxylic acid (5.8 mmol) and the corresponding substituted aniline (5.8 mmol) were suspended in dry chlorobenzene (25 mL). Phosphorous trichloride (2.9 mmol) was added dropwise, and the reacting mixture was heated in the microwave reactor at maximal allowed power 500 W and 130 °C, using infrared flask-surface control of temperature, for 15 min. The solvent was evaporated under reduced pressure, the solid residue washed with 2 M HCl, and the crude product was recrystallized from propan-2-ol with addition of active carbon.

N-Phenyl*naphthalene-1-carboxamide* (**1**). Method B; Yield 46%; Mp. 164 °C (164–165°C [55], 163 °C [56], 160 °C [57]); $R_f = 0.87$; IR (Zn/Se ATR, cm^{-1}): 3281 m , 3045 w , 1650 s , 1594 s , 1520 s , 1499 s , 1433 s , 1312 m , 1253 m , 1078 s , 781 s , 770 s , 744 s , 686 m ; $^1\text{H-NMR}$ (DMSO- d_6) [55,58,59], δ : 10.58 (s, 1H), 8.22–8.17 (m, 1H), 8.08 (d, $J = 8.1$ Hz, 1H), 8.06–8.00 (m, 1H), 7.83 (d, $J = 8.1$ Hz, 2H), 7.76 (d, $J = 5.9$ Hz, 1H), 7.65–7.58 (m, 3H), 7.38 (t, $J = 8.1$ Hz, 2H), 7.13 (td, $J = 7.3$ Hz, $J = 0.7$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.14, 139.17, 134.73, 133.04, 129.86, 129.57, 128.51, 128.13, 126.76, 126.14, 125.21, 124.96, 124.83, 123.51, 119.80; HR-MS: for $\text{C}_{17}\text{H}_{14}\text{NO}$ $[\text{M}+\text{H}]^+$ calculated 248.1070 m/z , found 248.1079 m/z .

N-(2-Methoxyphenyl)*naphthalene-1-carboxamide* (**2a**). Method A; Yield 89%; Mp. 117 °C; $R_f = 0.72$; IR (Zn/Se ATR, cm^{-1}): 3321 m , 3042 w , 1651 s , 1591 m , 1505 m , 1486 s , 1456 s , 1429 m , 1259 m , 1250 m , 1220 m , 1112 m , 1043 m , 1026 s , 901 m , 813 m , 802 m , 785 s , 760 s , 738 m , 723 m ; $^1\text{H-NMR}$ (DMSO- d_6), δ : 9.60 (s, 1H), 8.32–8.27 (m, 1H), 8.07 (d, $J = 8.4$ Hz, 1H), 8.04–7.99 (m, 1H), 7.92 (d, $J = 7.3$ Hz, 1H), 7.77 (d, $J = 6.6$ Hz, 1H), 7.63–7.56 (m, 3H), 7.26–7.09 (m, 2H), 7.01 (td, $J = 7.5$ Hz, $J = 1.5$ Hz, 1H), 3.83 (s, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.12, 151.25, 134.51, 133.10, 129.99, 129.70, 128.14, 126.96, 126.73, 126.15, 125.59, 125.27, 125.15, 124.89, 123.83, 120.16, 111.50, 55.69; HR-MS: for $\text{C}_{18}\text{H}_{16}\text{NO}_2$ $[\text{M}+\text{H}]^+$ calculated 278.1176 m/z , found 278.1181 m/z .

N-(3-Methoxyphenyl)*naphthalene-1-carboxamide* (**2b**). Method A; Yield 93%; Mp. 161 °C; $R_f = 0.69$; IR (Zn/Se ATR, cm^{-1}): 3280 m , 3046 w , 2966 w , 1648 s , 1592 s , 1538 s , 1493 m , 1327 m , 1307 s , 1251 s , 1027 m , 845 s , 815 m , 776 s , 736 m , 726 m , 688 s ; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.57 (s, 1H), 8.20–8.15 (m, 1H), 8.08 (d, $J = 8.0$ Hz, 1H), 8.05–8.00 (m, 1H), 7.74 (dd, $J = 7.0$ Hz, $J = 1.5$ Hz, 1H); 7.65–7.57 (m, 3H), 7.53 (t, $J = 1.5$ Hz, 1H), 7.40–7.31 (m, 1H), 7.27 (t, $J = 8.1$ Hz, 1H), 6.71 (ddd, $J = 8.1$ Hz, $J = 2.6$ Hz, $J = 1.1$ Hz, 1H); 3.76 (s, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.23, 159.47, 140.39, 134.71, 133.07, 129.96, 129.57, 129.37, 128.20, 126.85, 126.23, 125.27, 124.99, 124.89, 112.09, 109.11, 105.67, 54.96; HR-MS: for $\text{C}_{18}\text{H}_{16}\text{NO}_2$ $[\text{M}+\text{H}]^+$ calculated 278.1176 m/z , found 278.1181 m/z .

N-(4-Methoxyphenyl)*naphthalene-1-carboxamide* (**2c**). Method A; Yield 91%; Mp. 177 °C; $R_f = 0.66$; IR (Zn/Se ATR, cm^{-1}): 3227 m , 3125 w , 3050 w , 1647 s , 1603 s , 1506 s , 1469 m , 1414 m , 1327 m , 1227 m , 1180 m , 1030 m , 1026 m , 834 m , 821 m , 799 s , 777 s , 747 s , 730 m ; $^1\text{H-NMR}$ (DMSO- d_6) [58], δ : 10.45 (s, 1H), 8.22–8.17 (m, 1H), 8.07 (d, $J = 8.7$ Hz, 1H), 8.05–8.00 (0, 1H), 7.75 (d, $J = 7.7$ Hz, 1H), 7.73 (d, $J = 8.8$ Hz, 2H), 7.64–7.57 (m, 3H), 6.96 (d, $J = 9.1$ Hz, 2H), 3.76 (s, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 166.74, 155.51, 134.88, 133.07, 132.37, 129.78, 129.63, 128.16, 126.75, 126.17, 125.17, 125.07, 124.89, 121.33, 113.77, 55.14; HR-MS: for $\text{C}_{18}\text{H}_{16}\text{NO}_2$ $[\text{M}+\text{H}]^+$ calculated 278.1176 m/z , found 278.1177 m/z .

N-(2-Methylphenyl)naphthalene-1-carboxamide (**3a**). Method B; Yield 76%; Mp. 187 °C; $R_f = 0.70$; IR (Zn/Se ATR, cm^{-1}): 3267m, 3046w, 1642s, 1572s, 1524s, 1456s, 1303m, 1258m, 718s, 771s, 747s, 736s; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.06 (s, 1H), 8.32–8.28 (m, 1H), 8.08 (d, $J = 8.4$ Hz, 1H), 8.06–8.01 (m, 1H), 7.83 (d, $J = 6.6$ Hz, 1H), 7.66–7.50 (m, 4H), 7.32–7.15 (m, 3H), 2.34 (s, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.35, 136.23, 134.64, 133.15, 133.4, 130.31, 129.93, 129.81, 128.22, 126.81, 126.24, 126.23, 125.96, 125.87, 125.38, 125.18, 124.96, 17.96; HR-MS: for $\text{C}_{18}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+$ calculated 262.1226 m/z , found 262.1231 m/z .

N-(3-Methylphenyl)naphthalene-1-carboxamide (**3b**). Method A; Yield 88%; Mp. 162 °C; $R_f = 0.69$; IR (Zn/Se ATR, cm^{-1}): 3227m, 3133w, 3053w, 1651s, 1593s, 1538s, 1428m, 1305m, 1266m, 1252m, 1206w, 805m, 776s, 745s, 734s, 691s; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.52 (s, 1H), 8.21–8.17 (m, 1H), 8.08 (d, $J = 8.1$ Hz, 1H), 8.06–8.01 (m, 1H), 7.75 (d, $J = 6.6$ Hz, 1H), 7.70 (s, 1H), 7.76–7.58 (m, 4H), 7.26 (t, $J = 7.6$ Hz, 1H), 6.95 (d, $J = 7.2$ Hz, 1H), 2.33 (s, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.15, 139.13, 137.76, 134.82, 133.07, 129.87, 129.60, 128.40, 128.17, 126.81, 126.18, 125.21, 125.02, 124.89, 124.27, 120.34, 117.03, 21.10; HR-MS: for $\text{C}_{18}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+$ calculated 262.1226 m/z , found 262.1230 m/z .

N-(4-Methylphenyl)naphthalene-1-carboxamide (**3c**). Method A; Yield 95%; Mp. 196 °C (183 °C [56], 190–193 °C [60], 187–189 °C [61]); $R_f = 0.69$; IR (Zn/Se ATR, cm^{-1}): 3227m, 3180w, 3111w, 3031w, 2909w, 1651s, 1598s, 1532s, 1514s, 1404m, 1322s, 1301m, 1257m, 817m, 805s, 778s, 747s, 730m; $^1\text{H-NMR}$ (DMSO- d_6) [58,59], δ : 10.51 (s, 1H), 8.21–8.16 (m, 1H), 8.07 (d, $J = 8.4$ Hz, 1H), 8.05–8.00 (m, 1H), 7.74 (dd, $J = 7.0$ Hz, $J = 1.1$ Hz, 1H), 7.71 (d, $J = 8.1$ Hz, 2H), 7.64–7.55 (m, 3H), 7.18 (d, $J = 8.4$ Hz, 2H), 2.30 (s, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 166.97, 136.70, 134.83, 133.06, 132.53, 129.83, 129.61, 128.93, 128.16, 126.78, 126.17, 125.20, 125.03, 124.88, 119.81, 20.35; HR-MS: for $\text{C}_{18}\text{H}_{16}\text{NO}$ $[\text{M}+\text{H}]^+$ calculated 262.1226 m/z , found 262.1232 m/z .

N-(2-Fluorophenyl)naphthalene-1-carboxamide (**4a**). Method A; Yield 94%; Mp. 141 °C (Mp. 137–138 °C [62]); $R_f = 0.69$; IR (Zn/Se ATR, cm^{-1}): 3172m, 3118w, 3013w, 1656s, 1530s, 1498s, 1450s, 1313s, 1291s, 1261s, 1106m, 804m, 781s, 754s; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.38 (s, 1H), 8.29–8.24 (m, 1H), 8.09 (d, $J = 8.4$ Hz, 1H), 8.05–8.00 (m, 1H), 7.80 (d, $J = 6.6$ Hz, 1H), 7.79–7.74 (m, 1H), 7.65–7.55 (m, 3H), 7.39–7.23 (m, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.39, 155.33 (d, $J = 245.8$ Hz), 133.94, 133.07, 130.16, 129.70, 128.19, 126.84, 126.64 (d, $J = 7.6$ Hz), 126.45 (d, $J = 2.3$ Hz), 126.20, 125.64 (d, $J = 12.2$ Hz), 125.52, 125.03, 124.85, 124.21 (d, $J = 3.1$ Hz), 115.68 (d, $J = 19.7$ Hz); HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOF}$ $[\text{M}+\text{H}]^+$ calculated 266.0976 m/z , found 266.0982 m/z .

N-(3-Fluorophenyl)naphthalene-1-carboxamide (**4b**). Method A; Yield 96%; Mp. 150 °C; $R_f = 0.68$; IR (Zn/Se ATR, cm^{-1}): 3277m, 3039w, 1651s, 1611s, 1592s, 1522s, 1484m, 1414m, 1391m, 1249m, 1148m, 1129m, 855m, 770s, 676m; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.80 (s, 1H), 8.21–8.16 (m, 1H), 8.10 (d, $J = 8.1$ Hz, 1H), 8.06–8.00 (m, 1H), 7.85–8.76 (m, 2H), 7.66–7.54 (m, 4H), 7.47–7.36 (m, 1H), 6.96 (tdd, $J = 8.4$ Hz, $J = 2.6$ Hz, $J = 1.1$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.42, 162.05 (d, $J = 239.7$ Hz), 140.90 (d, $J = 10.7$ Hz), 134.29, 133.06, 130.22 (d, $J = 9.1$ Hz), 130.19, 129.51, 128.22, 126.96, 126.29, 125.44, 124.91, 124.88, 115.49 (d, $J = 2.4$ Hz), 110.03 (d, $J = 21.3$ Hz), 106.50 (d, $J = 25.8$ Hz); HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOF}$ $[\text{M}+\text{H}]^+$ calculated 266.0976 m/z , found 266.0981 m/z .

N-(4-Fluorophenyl)naphthalene-1-carboxamide (**4c**). Method A; Yield 78%; Mp. 185 °C; $R_f = 0.69$; IR (Zn/Se ATR, cm^{-1}): 3284 m , 3046 w , 1648 s , 1521 s , 1505 s , 1404 s , 1390 s , 1308 m , 1206 m , 1097 m , 826 s , 807 s , 778 s , 760 m , 701 m ; $^1\text{H-NMR}$ (DMSO- d_6) [58,59], δ : 10.66 (s, 1H), 8.22–8.17 (m, 1H), 8.09 (d, $J = 8.1$ Hz, 1H), 8.06–8.01 (m, 1H), 7.86–7.81 (m, 2H), 7.77 (dd, $J = 7.0$ Hz, $J = 1.5$ Hz, 1H), 7.65–7.56 (m, 3H), 7.23 (t, $J = 9.0$ Hz, 2H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.08, 158.21 (d, $J = 238.2$ Hz), 135.58 (d, $J = 2.3$ Hz), 134.54, 133.07, 130.01, 129.57, 128.20, 126.87, 126.23, 125.32, 125.00, 124.89, 121.57 (d, $J = 7.6$ Hz), 115.15 (d, $J = 22.0$ Hz); HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOF}$ $[\text{M}+\text{H}]^+$ calculated 266.0976 m/z , found 266.0981 m/z .

N-(2-Chlorophenyl)naphthalene-1-carboxamide (**5a**). Method A; Yield 83%; Mp. 157 °C (Mp. 154–156 °C [63]); $R_f = 0.72$; IR (Zn/Se ATR, cm^{-1}): 3263 m , 3046 w , 1648 s , 1581 s , 1520 s , 1505 s , 1475 m , 1439 m , 1301 s , 1253 m , 1059 m , 815 m , 786 s , 777 s , 743 s ; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.30 (s, 1H), 8.37–8.32 (m, 1H), 8.10 (d, $J = 8.4$ Hz, 1H), 8.06–8.01 (m, 1H), 7.85 (d, $J = 8.4$ Hz, 1H), 7.76–7.57 (m, 5H), 7.43 (td, $J = 7.7$ Hz, $J = 1.5$ Hz, 1H), 7.33 (td, $J = 7.7$ Hz, $J = 1.8$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.38, 134.85, 133.91, 133.10, 130.24, 129.73, 129.49, 129.16, 128.17, 128.14, 127.39, 127.38, 126.84, 126.23, 125.59, 125.15, 124.86; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOCl}$ $[\text{M}+\text{H}]^+$ calculated 282.0680 m/z , found 282.0685 m/z .

N-(3-Chlorophenyl)naphthalene-1-carboxamide (**5b**). Method A; Yield 87%; Mp. 144 °C; $R_f = 0.71$; IR (Zn/Se ATR, cm^{-1}): 3270 m , 3050 w , 1647 s , 1585 s , 1524 s , 1478 s , 1304 s , 1249 s , 863 m , 775 s , 744 m , 682 s ; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.78 (s, 1H), 8.21–8.16 (m, 1H), 8.11 (d, $J = 8.1$ Hz, 1H), 8.06–8.01 (m, 2H), 7.78 (dd, $J = 7.2$ Hz, $J = 1.1$ Hz, 1H), 7.70 (d, $J = 9.2$ Hz, 1H), 7.63–7.56 (m, 3H), 7.41 (t, $J = 8.0$ Hz, 1H), 7.20 (ddd, $J = 7.7$ Hz, $J = 1.8$ Hz, $J = 0.7$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.44, 140.63, 134.23, 133.07, 132.98, 130.30, 130.22, 129.51, 128.23, 126.97, 126.29, 125.46, 124.92, 124.88, 123.29, 119.25, 118.16; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOCl}$ $[\text{M}+\text{H}]^+$ calculated 282.0680 m/z , found 282.0685 m/z .

N-(4-Chlorophenyl)naphthalene-1-carboxamide (**5c**). Method A; Yield 90%; Mp. 182 °C; $R_f = 0.72$; IR (Zn/Se ATR, cm^{-1}): 3270 m , 3042 w , 1647 s , 1590 s , 1514 s , 1489 s , 1389 m , 1310 m , 1092 m , 1014 m , 820 s , 809 m , 783 s , 774 s , 709 m ; $^1\text{H-NMR}$ (DMSO- d_6) [58,59], δ : 10.73 (s, 1H), 8.20–8.15 (m, 1H), 8.09 (d, $J = 8.1$ Hz, 1H), 8.06–8.01 (m, 1H), 7.86 (d, $J = 8.8$ Hz, 2H), 7.77 (dd, $J = 8.1$ Hz, $J = 1.1$ Hz, 1H), 7.65–7.56 (m, 3H), 7.44 (d, $J = 8.8$ Hz, 2H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.24, 138.14, 134.38, 133.06, 130.11, 129.52, 128.49, 128.22, 127.22, 126.91, 126.26, 125.40, 124.94, 124.88, 121.31; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOCl}$ $[\text{M}+\text{H}]^+$ calculated 282.0680 m/z , found 282.0685 m/z .

N-(2-Bromophenyl)naphthalene-1-carboxamide (**6a**) Method A; Yield 79%; Mp. 155 °C (Mp. 147–150 °C [62], 149 °C [64]); $R_f = 0.73$; IR (Zn/Se ATR, cm^{-1}): 3256 m , 3042 w , 1647 s , 1575 m , 1520 s , 1471 m , 1432 m , 1299 m , 1253 m , 1026 m , 784 s , 774 s , 742 s , 734 s ; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.27 (s, 1H), 8.41–8.36 (m, 1H), 8.10 (d, $J = 8.4$ Hz, 1H), 8.06–8.01 (m, 1H), 7.88 (d, $J = 7.0$ Hz, 1H), 7.83–7.60 (m, 5H), 7.48 (td, $J = 7.7$ Hz, $J = 1.5$ Hz, 1H), 7.26 (td, $J = 7.7$ Hz, $J = 1.8$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.29, 136.27, 133.88, 133.10, 132.62, 130.24, 129.73, 128.64, 128.14, 128.01, 127.85, 126.79, 126.21, 125.55, 125.23, 124.82, 120.21; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOBr}$ $[\text{M}+\text{H}]^+$ calculated 326.0175 m/z , found 326.0183 m/z .

N-(3-Bromophenyl)naphthalene-1-carboxamide (**6b**). Method A; Yield 90%; Mp. 152 °C; $R_f = 0.71$; IR (Zn/Se ATR, cm^{-1}): 3266m, 3042w, 1647s, 1575m, 1520s, 1471s, 1432s, 1299s, 1253m, 1026m, 784s, 774s, 742s, 734s; $^1\text{H-NMR}$ (DMSO- d_6) [59], δ : 10.77 (s, 1H), 8.21–8.16 (m, 2H), 8.11 (d, $J = 8.1$ Hz, 1H), 8.05–8.01 (m, 1H), 7.78 (dd, $J = 7.0$ Hz, $J = 1.1$ Hz, 1H), 7.74–7.71 (m, 1H), 7.66–7.56 (m, 3H), 7.40–7.33 (m, 2H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.41, 140.77, 134.20, 133.06, 130.58, 130.22, 129.51, 128.22, 126.97, 126.29, 126.18, 125.46, 124.91, 124.86, 122.10, 121.42, 118.54; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOBr}$ $[\text{M}+\text{H}]^+$ calculated 326.0175 m/z , found 326.0183 m/z .

N-(4-Bromophenyl)naphthalene-1-carboxamide (**6c**). Method A; Yield 84%; Mp. 195 °C; $R_f = 0.72$; IR (Zn/Se ATR, cm^{-1}): 3216m, 3162w, 3100w, 3028w, 1648s, 1532m, 1486s, 1390m, 1318m, 1255m, 1070m, 1010m, 814s, 788s, 774s, 742m, 726m; $^1\text{H-NMR}$ (DMSO- d_6) [59], δ : 10.73 (s, 1H), 8.22–8.14 (m, 1H), 8.10 (d, $J = 8.1$ Hz, 1H), 8.06–7.99 (m, 1H), 7.81 (d, $J = 8.8$ Hz, 2H), 7.77 (d, $J = 7.0$ Hz, 1H), 7.66–7.56 (m, 3H) 7.57 (d, $J = 8.8$ Hz, 2H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 168.26, 138.55, 134.35, 133.06, 131.40, 130.13, 129.51, 128.20, 126.91, 126.24, 125.40, 124.92, 124.86, 121.69, 115.23; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{NOBr}$ $[\text{M}+\text{H}]^+$ calculated 326.0175 m/z , found 326.0184 m/z .

N-[2-(Trifluoromethyl)phenyl]naphthalene-1-carboxamide (**7a**). Method B; Yield 74%; Mp. 151 °C; $R_f = 0.87$; IR (Zn/Se ATR, cm^{-1}): 3249s, 3050w, 2999w, 1651s, 1585m, 1520s, 1452s, 1418s, 1297s, 1256s, 1170s, 1110s, 1058s, 1036s, 910m, 797m, 778s, 764s, 726m; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.38 (s, 1H), 8.33–8.28 (m, 1H), 8.10 (d, $J = 8.1$ Hz, 1H), 8.05–8.01 (m, 1H), 7.82–7.76 (m, 3H), 7.72–7.58 (m, 5H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 168.29, 135.46 (q, $J = 3.9$ Hz), 133.80, 133.15, 133.14, 131.16, 130.27, 129.73, 128.22, 127.43, 126.85, 126.45 (q, $J = 4.6$ Hz), 126.29, 126.16 (q, $J = 29.6$ Hz), 125.44, 125.14, 124.89, 123.70 (q, $J = 276.2$ Hz); HR-MS: for $\text{C}_{18}\text{H}_{13}\text{NOF}_3$ $[\text{M}+\text{H}]^+$ calculated 316.0944 m/z , found 316.0953 m/z .

N-[3-(Trifluoromethyl)phenyl]naphthalene-1-carboxamide (**7b**). Method B; Yield 78%; Mp. 148 °C; $R_f = 0.87$; IR (Zn/Se ATR, cm^{-1}): 3267m, 3046w, 1647s, 1529s, 1489s, 1330s, 1306s, 1254m, 1230m, 1162s, 1131s, 1115s, 1094s, 1065s, 1012w, 929m, 880s, 793s, 780s, 744s, 698m; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.29 (s, 1H), 8.35 (s, 1H), 8.24–8.19 (m, 1H), 8.11 (d, $J = 8.4$ Hz, 1H), 8.10–8.01 (m, 2H), 7.81 (d, $J = 6.6$ Hz, 1H), 7.67–7.59 (m, 4H), 7.49 (d, $J = 7.7$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.65, 139.99, 134.09, 133.10, 130.39, 129.90, 129.54, 129.45 (q, $J = 31.9$ Hz), 128.29, 127.06, 126.37, 125.62, 124.97, 124.91, 124.10 (q, $J = 270.9$ Hz), 123.30, 119.94 (q, $J = 3.9$ Hz), 115.83 (q, $J = 3.3$ Hz); HR-MS: for $\text{C}_{18}\text{H}_{13}\text{NOF}_3$ $[\text{M}+\text{H}]^+$ calculated 316.0944 m/z , found 316.0953 m/z .

N-[4-(Trifluoromethyl)phenyl]naphthalene-1-carboxamide (**7c**). Method B; Yield 68%; Mp. 193 °C; $R_f = 0.88$; IR (Zn/Se ATR, cm^{-1}): 3299m, 1657s, 1613m, 1597m, 1522s, 1511s, 1407m, 1328s, 1315s, 1253m, 1155s, 1109s, 1067s, 1018s, 904m, 834s, 789m, 777s, 756m; $^1\text{H-NMR}$ (DMSO- d_6), δ : 10.95 (s, 1H), 8.21–8.16 (m, 1H), 8.15 (d, $J = 8.4$ Hz, 1H), 8.09–8.02 (m, 3H), 7.83–7.74 (m, 3H), 7.67–7.59 (m, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.70, 142.80, 134.12, 133.10, 130.39, 129.52, 128.31, 127.08, 126.37, 125.96 (q, $J = 3.8$ Hz), 125.64, 124.93, 124.92, 124.36 (q, $J = 269.6$ Hz), 123.67 (q, $J = 31.9$ Hz), 119.68; HR-MS: for $\text{C}_{18}\text{H}_{13}\text{NOF}_3$ $[\text{M}+\text{H}]^+$ calculated 316.0944 m/z , found 316.0952 m/z .

N-(2-Nitrophenyl)naphthalene-1-carboxamide (**8a**). Method B; Yield 43%; Mp. 98 °C (Mp. 138–139 °C [59]); $R_f = 0.87$; IR (Zn/Se ATR, cm^{-1}): 3333 m , 1677 s , 1608 m , 1584 s , 1495 s , 1418 s , 1336 s , 1272 s , 897 m , 779 s , 727 s ; $^1\text{H-NMR}$ (DMSO- d_6), δ : 11.00 (s, 1H), 8.33–8.28 (m, 1H), 8.14 (d, $J = 8.1$ Hz, 1H), 8.07–8.02 (m, 2H), 7.85 (dd, $J = 7.0$ Hz, $J = 1.1$ Hz, 1H), 7.79–7.60 (m, 5H), 7.46 (td, $J = 7.5$ Hz, $J = 2.2$ Hz, 1H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.12, 143.21, 133.88, 133.16, 133.12, 131.01, 130.89, 129.70, 128.31, 127.09, 126.43, 125.97, 125.93, 125.73, 124.99, 124.92, 124.85; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$ calculated 293.0921 m/z , found 293.0925 m/z .

N-(3-Nitrophenyl)naphthalene-1-carboxamide (**8b**). Method A; Yield 79%; Mp. 172–175 °C; $R_f = 0.87$; IR (Zn/Se ATR, cm^{-1}): 3278 m , 3093 w , 3045 w , 1647 s , 1526 s , 1423 m , 1340 s , 1325 s , 1290 s , 1256 s , 902 m , 881 s , 834 m , 806 s , 777 s , 735 s ; $^1\text{H-NMR}$ (DMSO- d_6) [59], δ : 11.08 (s, 1H), 8.89 (t, $J = 2.2$ Hz, 1H), 8.25–8.20 (m, 1H), 8.16–8.11 (m, 2H), 8.07–7.97 (m, 2H), 7.84 (dd, $J = 7.0$ Hz, $J = 1.1$ Hz, 1H), 7.68 (t, $J = 8.1$ Hz, 1H), 7.66–7.59 (m, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.74, 147.97, 140.34, 133.85, 133.12, 130.54, 130.10, 129.52, 128.32, 127.13, 126.41, 125.75, 125.74, 124.93, 124.92, 118.14, 113.88; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$ calculated 293.0921 m/z , found 293.0931 m/z .

N-(4-Nitrophenyl)naphthalene-1-carboxamide (**8c**). Method B; Yield 60%; Mp. 213–215 °C; $R_f = 0.87$; IR (Zn/Se ATR, cm^{-1}): 3195 m , 3046 w , 1655 s , 1613 m , 1593 s , 1552 s , 1504 s , 1407 s , 1327 s , 1305 s , 1260 s , 1107 s , 856 s , 845 s , 821 s , 766 s , 749 s , 686 s ; $^1\text{H-NMR}$ (DMSO- d_6) [59], δ : 11.20 (s, 1H), 8.31 (d, $J = 9.2$ Hz, 2H), 8.21–8.15 (m, 2H), 8.08 (d, $J = 9.5$ Hz, 2H), 8.05–8.02 (m, 1H), 7.84 (dd, $J = 7.0$ Hz, $J = 1.1$ Hz, 1H), 7.68–7.59 (m, 3H); $^{13}\text{C-NMR}$ (DMSO- d_6), δ : 167.85, 145.36, 142.51, 133.71, 133.09, 130.65, 129.46, 128.34, 127.19, 126.41, 125.85, 124.89, 124.82, 124.81, 119.48; HR-MS: for $\text{C}_{17}\text{H}_{13}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$ calculated 293.0921 m/z , found 293.0926 m/z .

3.3. Lipophilicity Determination Using HPLC (Capacity Factor k / Calculated $\log k$)

A HPLC system Agilent 1200 equipped with DAD detector (Agilent, USA) was used. A chromatographic column Symmetry[®] C₁₈ 5 μm , 4.6 \times 250 mm, Part No. WAT054275 (Waters Corp., Milford, MA, USA) was used. The HPLC separation process was monitored and evaluated by EZChrom Elite software ver. 3.3.2 (Agilent, USA). Isocratic elution by a mixture of MeOH p.a. (60%) and H₂O-HPLC Mili-Q grade (40%) as a mobile phase was used. The total flow of the column was 1.0 mL/min, injection 20 μL , column temperature 40 °C, and sample temperature 10 °C. The detection wavelength 210 nm was chosen. The KI methanolic solution was used for the dead time (t_D) determination. Retention times (t_R) were measured in minutes. The capacity factors k were calculated according to formula $k = (t_R - t_D)/t_D$, where t_R is the retention time of the solute, whereas t_D denotes the dead time obtained using an unretained analyte. $\log k$, calculated from the capacity factor k , is used as the lipophilicity index converted to $\log P$ scale. The $\log k$ values of individual compounds are shown in Table 1.

3.4. In Vitro Antimycobacterial Evaluation

A well-characterized clinical isolate of *Mycobacterium avium* subsp. *paratuberculosis* (CIT03) was grown in Middlebrook broth (MB), supplemented with Oleic-Albumin-Dextrose-Catalase supplement

(OADC, Becton, Dickinson and Comp., Franklin Lakes, NJ, USA) and mycobactin J (2 µg/mL). Identification of this isolate was performed using biochemical and molecular protocols. At log phase growth, a culture sample (10 mL) was centrifuged at 15,000 rpm/20 min using a bench top centrifuge (Model CR 4-12, Jouan Inc., Winchester, VA, USA). Following removal of the supernatant, the pellet was washed in fresh Middlebrook 7H9GC broth and re-suspended in fresh supplemented MB (10 mL). The turbidity was adjusted to match McFarland standard No. 1 (3×10^8 cfu) with MB broth. A further 1:20 dilution of the culture was then performed in MB broth. The antimicrobial susceptibility of the mycobacterial species was investigated in a 96-well plate format. In these experiments, sterile deionized water (300 µL) was added to all outer-perimeter wells of the plates to minimize evaporation of the medium in the test wells during incubation. Each evaluated compound (100 µL) was incubated with the mycobacterial species (100 µL). Dilutions of each compound were prepared in duplicate. For all synthesized compounds, final concentrations ranged from 1,000 µg/mL to 8 µg/mL. All compounds were prepared in DMSO and subsequent dilutions were made in supplemented MB. The plates were sealed with parafilm and incubated at 37 °C for 11 days.

Following incubation, a 10% addition of alamarBlue (AbD Serotec, Kidlington, UK) was mixed into each well and readings at 570 nm and 600 nm were taken, initially for background subtraction and subsequently after 24 h re-incubation. The background subtraction is necessary for strongly coloured compounds, where the colour may interfere with the interpretation of any colour change. For non-interfering compounds, a blue colour in the well was interpreted as an absence of growth and a pink colour was scored as growth. Furthermore percent reduction of alamarBlue was calculated to generate quantitative data on cell viability during exposure to antimycobacterial compounds over a period of 5 days. The formula used to determine percent reduction of alamarBlue has been standardized for colorimetric analysis by Abd Serotec.

For the MTT assay, the same 96-well plate set up and incubation conditions listed above were used. After the incubation period, a 10% addition of MTT reagent was mixed into each well and incubated at 37 °C for 4 h. The reagent and media were then aspirated from the wells to which 50 µL DMSO was then added and plates were read at 550 nm.

The antimycobacterial activity was expressed as the minimum inhibitory concentration (MIC) that is defined for mycobacteria as a 90% or greater (IC_{90}) reduction of growth in comparison with the control. The MIC value is routinely and widely used in bacterial assays and is a standard detection limit according to the Clinical and Laboratory Standards Institute [47]. Isoniazid, rifampicin and ciprofloxacin (Sigma-Aldrich) were used as reference antimycobacterial drugs. The results are summarized in Table 1.

3.5. *In Vitro* Cytotoxicity Assay

Human monocytic leukemia THP-1 cells were obtained from the European Collection of Cell Cultures (ECACC, Salisbury, UK; Methods of characterization: DNA Fingerprinting (Multilocus probes) and isoenzyme analysis). These cells were routinely cultured in RPMI 1640 (Lonza, Verviers, Belgium) medium supplemented with 10% fetal bovine serum (FBS, Sigma-Aldrich, St. Louis, MO, USA), 2% L-glutamine, 1% penicillin and streptomycin (Lonza) at 37 °C with 5% CO₂. Cells were passaged at approximately one-week intervals. Cells were routinely tested for the absence of

mycoplasma (Hoechst 33258 staining method). The tested compounds were dissolved in DMSO (Sigma-Aldrich) and added in five increasing concentrations to the cell suspension in the culture medium. The maximum concentration of DMSO in the assays never exceeded 0.1%. Subsequently, the cells were incubated for 24 h at 37 °C with 5% CO₂ to various compound concentrations ranging from 0.37 to 20 µmol/L in RPMI 1640 medium. Cell toxicity was determined using a Cytotoxicity Detection Kit^{PLUS} Lactate dehydrogenase (LDH) assay kit (Roche Diagnostics, Mannheim, Germany) according to the manufacturer's instructions, as described previously [10–13]. For LDH assays, cells were seeded into 96-well plates (5 × 10⁴ cells/well in 100 µL culture medium) in triplicate in serum-free RPMI 1640 medium, and measurements at 492 nm wavelength (Synergy 2 Multi-Mode Microplate Reader, BioTek, Winooski, VT, USA) were taken 24 h after the treatment with tested compounds. The median lethal dose values, LD₅₀, were deduced through the production of a dose-response curve. All data were evaluated using GraphPad Prism 5.00 software (GraphPad Software, San Diego, CA, USA). The results are summarized in Table 1.

LDH Test Principle: The LDH activity is determined by a coupled enzymatic reaction, whereby the tetrazolium salt INT is reduced to formazan. An increase in the amount of dead or plasma membrane damaged cells results in an increase of LDH enzyme activity in the culture supernatant. This increase in the amount of enzyme activity in the supernatant directly correlates to the amount of formazan formed during a limited time period. The formazan dye formed is water soluble and shows a broad absorption maximum at approximately 500 nm.

3.6. Study of Inhibition Photosynthetic Electron Transport (PET) in Spinach Chloroplasts

Chloroplasts were prepared from spinach (*Spinacia oleracea* L.) according to Masarovicova and Kralova [65]. The inhibition of photosynthetic electron transport (PET) in isolated spinach chloroplasts was determined spectrophotometrically (Genesys 6, Thermo Scientific, USA), using an artificial electron acceptor 2,6-dichlorophenol-indophenol (DCIPP) according to Kralova *et al.* [66], and the rate of photosynthetic electron transport was monitored as a photoreduction of DCPIP. The measurements were carried out in phosphate buffer (0.02 mol/L, pH 7.2) containing sucrose (0.4 mol/L), MgCl₂ (0.005 mol/L) and NaCl (0.015 mol/L). The chlorophyll content was 30 mg/L in these experiments and the samples were irradiated (~100 W/m² with 10 cm distance) with a halogen lamp (250 W) using a 4 cm water filter to prevent warming of the samples (suspension temperature 22 °C). The studied compounds were dissolved in DMSO due to their limited water solubility. The applied DMSO concentration (up to 4%) practically did not affect the photochemical activity in isolated spinach chloroplasts (observed differences in DCPIP photoreduction due DMSO addition were within experimental error). The inhibitory efficiency of the studied compounds was expressed by IC₅₀ values, *i.e.*, by molar concentration of the compounds causing 50% decrease in the oxygen evolution rate relative to the untreated control. The comparable IC₅₀ value for a selective herbicide 3-(3,4-dichlorophenyl)-1,1-dimethylurea, DCMU (Diuron[®]) was about 1.9 µmol/L. The results are summarized in Table 1.

4. Conclusions

A series of twenty-two substituted naphthalene-1-carboxanilides were prepared and characterized. The prepared compounds were tested for their antimycobacterial activity against clinical isolate of *Mycobacterium avium* subsp. *paratuberculosis* and for their ability to inhibit photosynthetic electron transport (PET) in isolated spinach chloroplasts (*Spinacia oleracea* L.). Generally compounds showed poor aqueous solubility, which significantly reduced their biological effect. Five compounds, *N*-(2-methoxyphenyl)naphthalene-1-carboxamide (**2a**), *N*-(3-methoxyphenyl)naphthalene-1-carboxamide (**2b**), *N*-(3-methylphenyl)naphthalene-1-carboxamide (**3b**), *N*-(4-methylphenyl)-naphthalene-1-carboxamide (**3c**) and *N*-(3-fluorophenyl)naphthalene-1-carboxamide (**4b**) showed antimycobacterial activity two-fold higher than rifampicin and three-fold higher activity than ciprofloxacin. It was found that dependence of the antimycobacterial activities on lipophilicity and electronic properties were bilinear. Based on the results of the MTT assay of the most efficient compounds **2a**, **2b**, **3c**, and **4b** it seems that they could influence the mycobacterial respiratory chain. The most effective antimycobacterial compounds **2a**, **2b**, **3b**, **3c** and **4b** were tested for their *in vitro* cytotoxicity against THP-1 cells and within this preliminary screening they demonstrated insignificant toxicity. Based on these facts, it can be concluded that the discussed anilides can be considered as promising agents for subsequent design of novel antimycobacterial agents. The PET-inhibiting activity was rather moderate or low, the most active compound *N*-[4-(trifluoromethyl)-phenyl]naphthalene-1-carboxamide (**7c**) expressed $IC_{50} = 59 \mu\text{mol/L}$.

Acknowledgements

This study was supported by the IGA VFU Brno 37/2014/FaF, by the Slovak Grant Agency VEGA 1/0612/11, by the Projects APVV-0061-11, CZ.1.07/2.3.00/30.0053, CZ.1.05/1.1.00/02.0073 and by the Irish Department of Agriculture Fisheries and Food (FIRM) 08RDCIT601 & 08RDCIT617.

Author Contributions

Tomas Gonec, Jiri Kos, Josef Jampilek—design, synthesis of the compounds, SAR, writing of the paper. Jan Tengler, Michal Oravec—analysis/characterization of the compounds. Eoghan Nevin, Rodney Govender, Jim O'Mahony, Aidan Coffey—antimycobacterial evaluation of the compounds, writing of the paper. Matus Pesko, Katarina Kralova—study of PET inhibition in spinach chloroplasts of the compounds, writing of the paper. Ivan Kushkevych, Vendula Stastna, Peter Kollar—evaluation of cytotoxicity of the compounds.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. World Health Organization. *Global Tuberculosis Report 2013*; WHO Press: Geneva, Switzerland, 2013.

2. Wagner, D.; Young, L.S. Nontuberculous mycobacterial infections: A clinical review. *Infection* **2004**, *32*, 257–270.
3. Koul, A.; Arnoult, E.; Lounis, N.; Guillemont, J.; Andries, K. The challenge of new drug discovery for tuberculosis. *Nature* **2011**, *469*, 483–490.
4. Shaner, D.L. Herbicide safety relative to common targets in plants and mammals. *Pest. Manag. Sci.* **2004**, *60*, 17–24.
5. Delaney, J.; Clarke, E.; Hughes, D.; Rice, M. Modern agrochemical research: A missed opportunity for drug discovery? *Drug Discov. Today* **2006**, *11*, 839–845.
6. Duke, S.O. Herbicide and pharmaceutical relationships. *Weed Sci.* **2010**, *58*, 334–339.
7. Dolezal, M.; Cmedlova, P.; Palek, L.; Vinsova, J.; Kunes, J.; Buchta, V.; Jampilek, J.; Kralova, K. Synthesis and antimycobacterial evaluation of substituted pyrazinecarboxamides. *Eur. J. Med. Chem.* **2008**, *43*, 1105–1113.
8. Imramovsky, A.; Pesko, M.; Kralova, K.; Vejsova, M.; Stolarikova, J.; Vinsova, J.; Jampilek, J. Investigating spectrum of biological activity of 4- and 5-chloro-2-hydroxy-*N*-[2-(arylamino)-1-alkyl-2-oxoethyl]benzamides. *Molecules* **2011**, *16*, 2414–2430.
9. Gonec, T.; Bobal, P.; Suján, J.; Pesko, M.; Guo, J.; Kralova, K.; Pavlacka, L.; Vesely, L.; Kreckova, E.; Kos, J.; *et al.* Investigating the spectrum of biological activity of substituted quinoline-2-carboxamides and their isosteres. *Molecules* **2012**, *17*, 613–644.
10. Kos, J.; Zadrazilova, I.; Pesko, M.; Keltosova, S.; Tengler, J.; Gonec, T.; Bobal, P.; Kauerova, T.; Oravec, M.; Kollar, P.; *et al.* Antibacterial and herbicidal activity of ring-substituted 3-hydroxynaphthalene-2-carboxanilides. *Molecules* **2013**, *18*, 7977–7997.
11. Gonec, T.; Kos, J.; Zadrazilova, I.; Pesko, M.; Govender, R.; Keltosova, S.; Chambel, B.; Pereira, D.; Kollar, P.; Imramovsky, A.; *et al.* Antibacterial and herbicidal activity of ring-substituted 2-hydroxynaphthalene-1-carboxanilides. *Molecules* **2013**, *18*, 9397–9419.
12. Gonec, T.; Kos, J.; Zadrazilova, I.; Pesko, M.; Keltosova, S.; Tengler, J.; Bobal, P.; Kollar, P.; Cizek, A.; Kralova, K.; *et al.* Antimycobacterial and herbicidal activity of ring-substituted 1-hydroxynaphthalene-2-carboxanilides. *Bioorg. Med. Chem.* **2013**, *21*, 6531–6541.
13. Fajkusova, D.; Pesko, M.; Keltosova, S.; Guo, J.; Oktabec, Z.; Vejsova, M.; Kollar, P.; Coffey, A.; Csollei, J.; Kralova, K.; *et al.* Anti-infective and herbicidal activity of *N*-substituted 2-aminobenzothiazoles. *Bioorg. Med. Chem.* **2012**, *20*, 7059–7068.
14. Kralova, K.; Sersen, F.; Cizmarik, J. Inhibitory effect of piperidinoethylesters of alkoxyphenylcarbamic acids on photosynthesis. *Gen. Physiol. Biophys.* **1992**, *11*, 261–267.
15. Kralova, K.; Bujdakova, H.; Kuchta, T.; Loos, D. Correlation between biological activity and the structure of 6-amino-2-*R*-thiobenzothiazoles. Anti-yeast activity and inhibition of photochemical activity of chloroplasts. *Pharmazie* **1994**, *49*, 460–461.
16. Kralova, K.; Kallova, J.; Loos, D.; Devinsky, F. Correlation between biological activity and the structure of *N,N'*-bis(alkyldimethyl)-1,6-hexanediammonium dibromides. Antibacterial activity and inhibition of photochemical activity of chloroplasts. *Pharmazie* **1994**, *49*, 857–858.
17. Bujdakova, H.; Kralova, K.; Sidoova, E. Antifungal and antialgal activity of 3-(2-alkylthio-6-benzothiazolylaminomethyl)-2-benzoxazolinethiones. *Pharmazie* **1995**, *50*, 156–156.
18. Kralova, K.; Bujdakova, H.; Cizmarik, J. Antifungal and antialgal activity of piperidinopropyl esters of alkoxy substituted phenylcarbamic acids. *Pharmazie* **1995**, *50*, 440–441.

19. Laursen, J.S.; Engel-Andreasen, J.; Fristrup, P.; Harris, P.; Olsen, C.A. Cis-trans amide bond rotamers in β -peptoids and peptoids: Evaluation of stereoelectronic effects in backbone and side chains. *J. Am. Chem. Soc.* **2013**, *135*, 2835–2844.
20. Pattabiraman, V.R.; Bode, J.W. Rethinking amide bond synthesis. *Nature* **2011**, *480*, 471–479.
21. Roth, H.J.; Fenner, H. *Arzneistoffe*, 3rd ed.; Deutscher Apotheker Verlag: Stuttgart, Germany, 2000; pp. 51–114.
22. Sinning, C.; Watzler, B.; de Petrocellis, L.; di Marzo, V.; Imming, P. Dopamides, vanillylamides, ethanolamides, and arachidonic acid amides of anti-inflammatory and analgesic drug substances as TRPV1 ligands. *Chem. Med. Chem.* **2008**, *3*, 1956–1964.
23. Good, N.E. Inhibitors of the Hill reaction. *Plant Physiol.* **1961**, *36*, 788–803.
24. Musiol, R.; Tabak, D.; Niedbala, H.; Podeszwa, B.; Jampilek, J.; Kralova, K.; Dohnal, J.; Finster, J.; Mencil, A.; Polanski, J. Investigating biological activity spectrum for novel quinoline analogues 2: Hydroxyquinolinecarboxamides with photosynthesis inhibiting activity. *Bioorg. Med. Chem.* **2008**, *16*, 4490–4499.
25. Imramovsky, A.; Vinsova, J.; Monreal-Ferriz, J.; Dolezal, R.; Jampilek, J.; Kaustova, J.; Kunc, F. New antituberculotics originated from salicylanilides with promising *in vitro* activity against atypical mycobacterial strains. *Bioorg. Med. Chem.* **2009**, *17*, 3572–3579.
26. Otevrel, J.; Mandelova, Z.; Pesko, M.; Guo, J.; Kralova, K.; Sersen, F.; Vejsova, M.; Kalinowski, D.; Kovacevic, Z.; Coffey, A.; *et al.* Investigating the spectrum of biological activity of ring-substituted salicylanilides and carbamoylphenylcarbamates. *Molecules* **2010**, *15*, 8122–8142.
27. Imramovsky, A.; Pesko, M.; Monreal-Ferriz, J.; Kralova, K.; Vinsova, J.; Jampilek, J. Photosynthesis-inhibiting efficiency of 4-chloro-2-(chlorophenylcarbamoyl)phenyl alkyl-carbamates. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 4564–4567.
28. Pauk, K.; Zadrazilova, I.; Imramovsky, A.; Vinsova, J.; Pokorna, M.; Masarikova, M.; Cizek, A.; Jampilek, J. New derivatives of salicylamides: Preparation and antimicrobial activity against various bacterial species. *Bioorg. Med. Chem.* **2013**, *21*, 6574–6581.
29. Andries, K.; Verhasselt, P.; Guillemont, J.; Gohlmann, H.W.; Neefs, J.M.; Winkler, H.; van Gestel, J.; Timmerman, P.; Zhu, M.; Lee, E.; *et al.* A diarylquinoline drug active on the ATP synthase of *Mycobacterium tuberculosis*. *Science* **2005**, *307*, 223–227.
30. Jampilek, J.; Dolezal, M.; Kunes, J.; Buchta, V.; Kralova, K. Quinaldine derivatives: Preparation and biological activity. *Med. Chem.* **2005**, *1*, 591–599.
31. Musiol, R.; Jampilek, J.; Buchta, V.; Niedbala, H.; Podeszwa, B.; Palka, A.; Majerz-Maniecka, K.; Oleksyn, B.; Polanski, J. Antifungal properties of new series of quinoline derivatives. *Bioorg. Med. Chem.* **2006**, *14*, 3592–3598.
32. Musiol, R.; Jampilek, J.; Kralova, K.; Richardson, D.R.; Kalinowski, D.; Podeszwa, B.; Finster, J.; Niedbala, H.; Palka, A.; Polanski, J. Investigating biological activity spectrum for novel quinoline analogues. *Bioorg. Med. Chem.* **2007**, *15*, 1280–1288.
33. Podeszwa, B.; Niedbala, H.; Polanski, J.; Musiol, R.; Tabak, D.; Finster, J.; Serafin, K.; Wietrzyk, J.; Boryczka, S.; Mol, W.; *et al.* Investigating the antiproliferative activity of quinoline-5,8-dione analogues on tumour cell lines. *Bioorg. Med. Chem. Lett.* **2007**, *17*, 6138–6141.

34. Jampilek, J.; Musiol, R.; Finster, J.; Pesko, M.; Carroll, J.; Kralova, K.; Vejsova, M.; Coffey, A.; Polanski, J. Investigating biological activity spectrum for novel styrylquinazoline analogues. *Molecules* **2009**, *14*, 4246–4265.
35. Mrozek-Wilczkiewicz, A.; Kalinowski, D.; Musiol, R.; Finster, J.; Kovacevic, Z.; Jampilek, J.; Rzeszowska-Wolny, J.; Richardson, D.R.; Polanski, J. Investigating anti-proliferative activity of styrylazanaphthalenes and azanaphthalenediones. *Bioorg. Med. Chem.* **2010**, *18*, 2664–2671.
36. Serda, M.; Mrozek-Wilczkiewicz, A.; Jampilek, J.; Pesko, M.; Kralova, K.; Vejsova, M.; Musiol, R.; Polanski, J. Investigation of biological properties for (hetero)aromatic thiosemicarbazones. *Molecules* **2012**, *17*, 13483–13502.
37. Cieslik, W.; Musiol, R.; Nycz, J.; Jampilek, J.; Vejsova, M.; Wolff, M.; Machura, B.; Polanski, J. Contribution to investigation of antimicrobial activity of styrylquinolines. *Bioorg. Med. Chem.* **2012**, *20*, 6960–6968.
38. Rokade, Y.B.; Sayyed, R.Z. Naphthalene derivatives: A new range of antimicrobials with high therapeutic value. *Rasayan J. Chem.* **2009**, *2*, 972–980.
39. Durrant, J.D.; Hall, L.; Swift, R.V.; Landon, M.; Schnauffer, A.; Schnauffer, A.; Amaro, R.E. Novel naphthalene-based inhibitors of Trypanosoma brucei RNA editing ligase 1. *PLoS Negl. Trop. Dis.* **2010**, doi:10.1371/journal.pntd.0000803.
40. Parineeta, B.N. Derivatives of 1-chloromethyl naphthalene: Synthesis and microbiological evaluation as potential antifungal agents. *Der Pharma Chem.* **2011**, *3*, 105–111.
41. Kanno, T.; Tanaka, A.; Shimizu, T.; Nakano, T.; Nishizaki, T. 1-[2-(2-Methoxyphenylamino)ethylamino]-3-(naphthalene-1-yloxy)propan-2-ol as a potential anticancer drug. *Pharmacology* **2013**, *91*, 339–345.
42. Damu, G.L.V.; Wang, Q.P.; Zhang, H.Z.; Zhang, Y.Y.; Lv, J.S.; Zhou, C.H. A series of naphthalimide azoles: Design, synthesis and bioactive evaluation as potential antimicrobial agents. *Sci. Chin. Chem.* **2013**, *56*, 952–969.
43. Draber, W.; Tietjen, K.; Kluth, J.F.; Trebst, A. Herbicides in photosynthesis research. *Angew. Chem.* **1991**, *3*, 1621–1633.
44. Tischer, W.; Strotmann, H. Relationship between inhibitor binding by chloroplasts and inhibition of photosynthetic electron-transport. *Biochim. Biophys. Acta* **1977**, *460*, 113–125.
45. Trebst, A.; Draber, W. Structure activity correlations of recent herbicides in photosynthetic reactions. In *Advances in Pesticide Science*; Greissbuehler, H., Ed.; Pergamon Press: Oxford, UK, 1979; pp. 223–234.
46. Bowyer, J.R.; Camilleri, P.; Vermaas, W.F.J. Photosystem II and its interaction with herbicides. In *Herbicides, Topics in Photosynthesis*; Baker, N.R., Percival, M.P., Eds.; Elsevier: Amsterdam, The Netherlands, 1991; Volume 10, pp. 27–85.
47. National Committee for Clinical Laboratory Standards. *Susceptibility Testing of Mycobacteria, Nocardiae, and Other Aerobic Actinomycetes; Approved Standard, M24-A2*, 2nd ed; National Committee for Clinical Laboratory Standards: Wayne, PA, USA, 2011.
48. Carroll, J.; Douarre, P.; Coffey, A.; Buckley, J.; Cashman, B.; O'Farrell, K.; O'Mahony, J. Optimization of a rapid viability assay for Mycobacterium avium paratuberculosis by using alamarBlue. *Appl. Environ. Microbiol.* **2009**, *75*, 7870–7872.

49. Bueno, J. Antitubercular *in vitro* drug discovery: Tools for begin the search. In *Understanding Tuberculosis-New Approaches to Fighting against Drug Resistance*; Cardona, P.J., Ed.; In Tech: Rijeka, Croatia, 2012; pp. 147–168.
50. Janin, Y.L. Antituberculosis drugs: Ten years of research. *Bioorg. Med. Chem.* **2007**, *15*, 2479–2513.
51. Bueno, R.V.; Braga, R.C.; Segretti, N.D.; Ferreir, E.I.; Trossini, G.H.; Andrade, C.H. New tuberculostatic agents targeting nucleic acid biosynthesis: Drug design using QSAR approaches. *Curr. Pharm. Des.* **2014**, *20*, 4474–4485.
52. Koul, A.; Vranckx, L.; Dhar, N.; Gohlmann, H.W.H.; Ozdemir, E.; Neefs, J.M.; Schulz, M.; Lu, P.; Mortz, E.; McKinney, J.D.; *et al.* Delayed bactericidal response of Mycobacterium tuberculosis to bedaquiline involves remodelling of bacterial metabolism. *Nat. Commun.* **2014**, doi:10.1038/ncomms4369.
53. Suffness, M.; Douros, J. Current status of the NCI plant and animal product program. *J. Nat. Prod.* **1982**, *45*, 1–14.
54. Bell, F. CCLVII.—The migration of acyl groups in o-aminophenols. *J. Chem. Soc.* **1930**, 1981–1987. doi:10.1039/JR9300001981.
55. Kumar, K.N.; Sreeramamurthy, K.; Palle, S.; Mukkanti, K.; Das, P. Dithiocarbamate and DBU-promoted amide bond formation under microwave condition. *Tetrahedron Lett.* **2010**, *51*, 899–902.
56. Kobs, U.; Neumann, W.P. Facile and effective synthesis of unusually substituted aromatic *N*-phenylamides. *Chem. Ber.* **1990**, *123*, 2191–2194.
57. Shah, R.; Deshpande, R.K. A simplified method for the preparation of anilides. *J. Uni. Bombay* **1933**, *2*, 125–127.
58. Strukil, V.; Bartolec, B.; Portada, T.; Dilovic, I.; Halasz, I.; Margetic, D. One-pot mechanosynthesis of aromatic amides and dipeptides from carboxylic acids and amines. *Chem. Commun.* **2012**, *48*, 12100–12102.
59. Liu, L.H.; Guo, L.; Liu, C.H.; Zhang, X.; Jiang, Y.B. Intramolecular charge transfer with 1-naphthanilides and 2-naphthanilides. *Chin. J. Chem.* **2005**, *23*, 857–864.
60. Zhang, Z.; Yu, Y.; Liebeskind, L.S. N-Amidation by copper-mediated cross-coupling of organostannanes or boronic acids with *O*-acetyl hydroxamic acids. *Org. Lett.* **2008**, *10*, 3005–3008.
61. Bahrami, K.; Khodaei, M.M.; Targhan, H.; Arabi, M.S. Preparation of esters and amides from carboxylic acids and *N*-formylation of amines promoted by 1,3,5-triazo-2,4,6-triphosphorine-2,2,4,4,6,6-hexachloride. *Tetrahedron Lett.* **2013**, *54*, 5064–5068.
62. Tambade, P.J.; Patil, Y.P.; Bhanage, B.M. Palladium bis(2,2,6,6-tetramethyl-3,5-heptanedionate) catalyzed alkoxycarbonylation and aminocarbonylation reactions. *Appl. Organometal. Chem.* **2009**, *23*, 235–240.
63. El-Sheikh, M.I.; Marks, A.; Biehl, E.R. Investigation of the synthesis of benzoxazole via aryne reaction. *J. Org. Chem.* **1981**, *46*, 3256–3259.
64. Aluri, B.R.; Niaz, B.; Kindermann, M.K.; Jones, P.G.; Heinicke, J. *P-C-N*-Heterocycles: Synthesis of biaryl-type 1,3-benzazaphospholes with *ortho*-substituted phenyl or 2-heteroaryl groups. *Dalton Trans.* **2011**, *40*, 211–224.

65. Masarovicova, E.; Kralova, K. Approaches to measuring plant photosynthesis activity. In *Handbook of Photosynthesis*, 2nd ed.; Pessaraki, M., Ed.; Taylor & Francis Group: Boca Raton, FL, USA, 2005; pp. 617–656.
66. Kralova, K.; Sersen, F.; Sidoova, E. Photosynthesis inhibition produced by 2-alkylthio-6-R-benzothiazoles. *Chem. Pap.* **1992**, *46*, 348–350.

Sample Availability: Samples of the compounds are available from the authors.

© 2014 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).