

**UCC Library and UCC researchers have made this item openly available.  
Please [let us know](#) how this has helped you. Thanks!**

<b>Title</b>	The importance of the calcium-to-magnesium ratio for phytoremediation of dairy industry wastewater using the aquatic plant <i>Lemna minor</i> L.
<b>Author(s)</b>	Walsh, Éamonn; Paolacci, Simona; Burnell, Gavin M.; Jansen, Marcel A. K.
<b>Publication date</b>	2020-01-07
<b>Original citation</b>	Walsh, É., Paolacci, S., Burnell, G. and Jansen, M. A. K. (2020) 'The importance of the calcium-to-magnesium ratio for phytoremediation of dairy industry wastewater using the aquatic plant <i>Lemna minor</i> L.', <i>International Journal of Phytoremediation</i> . doi: 10.1080/15226514.2019.1707478
<b>Type of publication</b>	Article (peer-reviewed)
<b>Link to publisher's version</b>	<a href="http://dx.doi.org/10.1080/15226514.2019.1707478">http://dx.doi.org/10.1080/15226514.2019.1707478</a> Access to the full text of the published version may require a subscription.
<b>Rights</b>	© 2020, Taylor & Francis Group. This is an Accepted Manuscript of an article published by Taylor & Francis in <i>International Journal of Phytoremediation</i> on 7 January 2020, available online: <a href="http://www.tandfonline.com/10.1080/15226514.2019.1707478">http://www.tandfonline.com/10.1080/15226514.2019.1707478</a>
<b>Embargo information</b>	Access to this article is restricted until 12 months after publication by request of the publisher.
<b>Embargo lift date</b>	2021-01-07
<b>Item downloaded from</b>	<a href="http://hdl.handle.net/10468/9644">http://hdl.handle.net/10468/9644</a>

Downloaded on 2021-11-27T09:47:18Z

1

2 **The importance of the calcium-to-magnesium ratio for phytoremediation of**  
3 **dairy industry wastewater using the aquatic plant *Lemna minor* L..**

4

5 Éamonn Walsh<sup>a,b,\*</sup>, Simona Paolacci<sup>a,b</sup>, Gavin Burnell<sup>a,b</sup>, Marcel A.K. Jansen<sup>a,b</sup>

6 <sup>a</sup> School of Biological, Earth and Environmental Science, University College Cork, Distillery

7 Fields, North Mall, Cork, Ireland

8 <sup>b</sup> Environmental Research Institute, University College Cork, Lee Road, Cork, Ireland

9 \* Corresponding author

## 10 **Abstract**

11 Lemnaceae are being exploited to remediate a variety of different wastewaters. Dairy  
12 processing waste is produced in large amounts, and contains a range of valuable plant nutrients,  
13 for example, nitrate, ammonium, phosphate, iron and calcium. Our aim was to remediate dairy  
14 processing waste with the duckweed plant *Lemna minor*. However, initial trials failed to  
15 establish growth of *L. minor* on this medium. A lack of growth can be due to a lack of essential  
16 plant micro- and macro-nutrients, or the presence of phytotoxic ingredients. In this study we  
17 show that not just nutrient concentrations, but also the ratios between them can be important in  
18 facilitating growth. Using lab-scale experiments in which *L. minor* were grown on 100 mL of  
19 synthetic wastewater, we demonstrated that the skewed Ca:Mg ratio in synthetic dairy industry  
20 wastewater is a key obstacle to good growth. Experiments showed that a ratio which favours  
21 magnesium over calcium negatively affects *L. minor* growth and photosynthetic yield, leading  
22 to RGRs as low as  $0.05 \text{ day}^{-1}$ . A change in this ratio to favour calcium, through the addition of  
23 calcium sulphate, leads to RGRs of  $0.2 - 0.3 \text{ day}^{-1}$ . Experiments lead us to conclude that a  
24 Ca:Mg ratio of 1:1.6 (by molar concentration) or greater is necessary for *Lemna minor* growth,  
25 and therefore phytoremediation of dairy industry processing wastewater.

26

27 *Keywords: phytoremediation, dairy processing, Lemna, duckweed, wastewater, toxicity*

## 28 **Introduction**

29 Phytoremediation refers to the process whereby plants, and associated microorganisms, are  
30 used to remove and/or degrade contaminants from soils and waters. Lemnaceae species,  
31 commonly referred to as duckweed (Landolt 1986), have been extensively studied for their  
32 phytoremediation potential. This potential relates to fast growth rates, relative tolerance to a  
33 range of pollutants, and high pollutant removal rates (Zayed et al. 1998; Cheng & Stomp 2009;  
34 Ziegler et al. 2015). Furthermore, the high protein content and good protein quality, i.e.  
35 desirable amino acid composition, make Lemnaceae biomass attractive as a potential  
36 component in animal feeds (Cheng & Stomp 2009; Anderson et al. 2011; Appenroth et al.  
37 2017). Thus, where Lemnaceae are used to remediate uncontaminated agricultural waste  
38 streams, a circular economy approach can be considered. In this scenario, nutrients (most  
39 importantly N- and P-containing compounds) present in wastewater are recycled into animal  
40 feed. Re-using plant nutrients, such as phosphate, nitrate and ammonia, present in wastewater  
41 can generate income from waste, reduce the costs associated with storage and tertiary  
42 wastewater treatments, and prevent environmental damage (i.e. eutrophication) associated with  
43 release of nutrient-rich waste on to surface waters (Diaz & Rosenberg 2008; Conley et al.  
44 2009).

45 Dairy industry processing wastewater is generated during the production of dairy products such  
46 as milk powder, cheese and yogurt from raw milk. Typically, large volumes are produced. In  
47 Europe dairy processing is seen as the largest industrial food wastewater source, with 0.5–37  
48 m<sup>3</sup> of effluent per m<sup>3</sup> of processed milk (Kolev Slavov 2017). This processing wastewater is  
49 rich in nitrogen and phosphorous as well as other essential plant nutrients such as calcium,  
50 potassium and magnesium (Ince 1998; Demirel & Yenigun 2004; Goyal & Gandhi 2009;  
51 Carvalho et al. 2013; Ryan & Walsh 2016). As expected for a waste product from the food  
52 industry, dairy industry processing wastewater contains only low concentrations of

53 contaminants such as heavy metals (Ince 1998; Demirel & Yenigun 2004). Therefore, dairy  
54 industry processing wastewater is well suited to remediation using a circular economy  
55 approach.

56 Duckweed species have been shown to be tolerant of a wide range of conditions and nutrient  
57 concentrations (Landolt & Kandeler 1987). Nevertheless, any particular wastewater needs to  
58 fulfil minimal criteria to facilitate growth and phytoremediation. An important criterion is the  
59 presence of adequate levels of essential plant growth nutrients. Taking nitrogen as an example,  
60 *Lemna minor* will grow on either ammonium or nitrate as a nitrogen source and can tolerate  
61 concentrations ranging between 0.2 to 150 mM with optimal concentrations varying depending  
62 on the nitrogen source (Landolt & Kandeler 1987; Paolacci et al. 2016). Duckweed can tolerate  
63 a pH range from 4 – 8 (Landolt & Kandeler 1987), but also the pH is also important in  
64 determining the tolerance of duckweed to ammonia (Körner et al. 2003). In the case of  
65 phosphate, duckweed tolerates concentrations ranging between 0.001 to 10 mM (Landolt &  
66 Kandeler 1987; Paolacci et al. 2016). For calcium and magnesium acceptable concentrations  
67 range between 0.2-20 mM and 0.2-10 mM, respectively (Landolt & Kandeler 1987; Van Dam  
68 et al. 2010; Paolacci et al. 2016), while the Ca:Mg ratio is also an important determinant of  
69 plant growth. Other criteria for plant growth and phytoremediation relate to the presence of,  
70 potentially phytotoxic, pollutants. In the specific case of dairy industry processing waste there  
71 is a heavy load of organic matter in the wastewater which is measured as biochemical oxygen  
72 demand (BOD<sub>5</sub> mg/L), chemical oxygen demand (COD mg/L) and fats (mg/L) (Janczukowicz  
73 et al. 2008; Carvalho et al. 2013). Duckweed do not require organic compounds in the medium  
74 for survival and growth (Körner et al. 1998), however, they can contribute to the reduction in  
75 the amount of organic matter as part of a phytoremediation approach (Körner et al. 1998; Li et  
76 al. 2017). All compounds mentioned are present in dairy processing wastewaters but there is a  
77 high degree of variability in their concentrations, and this relates to different factories and

78 processes (Demirel & Yenigun 2004; Goyal & Gandhi 2009; Carvalho et al. 2013; Tikariha &  
79 Sahu 2014), as well as strong seasonal influences on milk production.

80 In order to facilitate reproducible laboratory phytoremediation studies, a synthetic dairy  
81 industry wastewater has been developed (Tarpey 2016; Gil-Pulido et al. 2018). The  
82 composition of this synthetic wastewater (Table 1) is based on measurements of the  
83 composition of real wastewater. Unfortunately, preliminary experiments showed that  
84 duckweed did not grow well in this synthetic dairy wastewater. The aim of the present study  
85 was to identify the reasons responsible for the poor performance of duckweed on the synthetic  
86 dairy wastewater. Synthetic dairy industry wastewater has a Ca:Mg ratio of 1:14.6. It is known  
87 that an imbalance in favour of magnesium can have a negative effect on the growth and health  
88 of duckweed (Landolt & Kandeler 1987; Paolacci et al. 2016). This antagonistic Ca:Mg  
89 relationship was first studied in terrestrial plants (Loew & May 1901). It has been found that  
90 in soils magnesium decreases the calcium uptake in the plant, while calcium reduces  
91 magnesium uptake to a lesser extent (Halstead et al. 1958). Thus, imbalances in the soil Ca:Mg  
92 ratio can potentially aggravate calcium or magnesium deficiencies as well as magnesium  
93 toxicity (Brady et al. 2005). Therefore, based on existing literature, we hypothesised that a ratio  
94 between calcium and magnesium in favour of magnesium causes acute toxicity in *L. minor*,  
95 and that changing the ratio in favour of calcium removes this acute toxicity for *L. minor*. In this  
96 study, different levels of this imbalance between calcium and magnesium were tested in short-  
97 and long-term experiments in order to identify the concentration of calcium and magnesium  
98 most suitable for duckweed in the particular chemical environment of the synthetic dairy  
99 wastewater. This study underpins an important aspect of phytoremediation, assessing and  
100 amending wastewater composition to facilitate plant growth.

## 101 **Materials and Methods**

### 102 *Cultivation of stock and experimental plants*

103 The duckweed strain used in this study was *Lemna minor* - Blarney (international strain ID  
104 number 5500). A stock of sterile *L. minor* was kept in optimised growing conditions on half-  
105 strength Hutner's medium (Hutner 1953) in a growth room at a constant temperature of 22°C,  
106 light intensity of 52.66  $\mu\text{mol m}^{-2} \text{s}^{-1}$  and a photoperiod of 14 hours light to 10 hours dark.

107 A synthetic wastewater was used as a growing media (Table 1) for experimental purposes. The  
108 composition of this synthetic wastewater was based on analysis of real dairy industry  
109 processing wastewater (Tarpey 2016). Control plants were grown in 100 mL of half-strength  
110 Hutner's medium and experimental plants were grown in 100 mL of synthetic dairy processing  
111 wastewater. For all experiments plants were grown in their respective media in magenta vessels  
112 (Magenta GA-7 Plant Culture Box) for 7 days in a growth room at a constant temperature of  
113 21°C, light intensity of 80.82  $\mu\text{mol m}^{-2} \text{s}^{-1}$  and a photoperiod of 16 hours light to 8 hours dark.  
114 This light intensity is lower than that used in standard toxicological protocols, thus growth rates  
115 were moderately lower than standard, not exceeding an RGR of 0.3  $\text{day}^{-1}$ .

### 116 *Experimental design*

117 This study contains two types of experiments; short-term (7 days) and long-term (42 days). In  
118 both types of studies *L. minor* plants were grown on a number of differently modified versions  
119 of synthetic dairy wastewater (Table 1) with half-strength Hutner's medium as a control. At  
120 the start of each experiment, three three-frond colonies were added to each magenta. On day  
121 zero of each experiment, the starting mass (fresh weight), colony number and frond number  
122 were determined. The synthetic wastewater was not changed during the 7-day experiments. In  
123 the case of long-term experiments (42 days), the 100mL of synthetic wastewater was replaced

124 weekly. Furthermore, the density of plants in the long-term experiment was returned to a  
125 constant amount, three colonies, at the start of each week.

126 [Table 1 near here]

### 127 *Synthetic wastewater modifications*

128 A reduction in the concentration of chloride was achieved by replacing ammonium chloride in  
129 the synthetic wastewater with ammonium sulphate, thus removing the majority of chloride. A  
130 reduction in the concentration of sodium was achieved by the removal of sodium bicarbonate.  
131 Iron and manganese concentrations were reduced through the addition of less iron sulphate  
132 heptahydrate and manganese chloride tetrahydrate, respectively. Potassium was increased  
133 through the addition of potassium bicarbonate. Sulphate was increased through the substitution  
134 of ammonium chloride for ammonium sulphate and the addition of potassium sulphate. The  
135 calcium concentration was increased by adding calcium sulphate ( $\text{CaSO}_4$ ) to the synthetic  
136 wastewater. The magnesium concentration was increased or decreased by altering the  
137 concentration of magnesium sulphate heptahydrate ( $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$ ). A secondary impact of  
138 altering the calcium and magnesium concentrations is that sulphate concentration is also  
139 affected. The highest concentration to which sulphate is increased, 8.3 mM, is still significantly  
140 below the maximum concentration of sulphate tolerated by *L. minor*, 60 mM (see Table 2 for  
141 maximum tolerated and ‘optimal’ concentrations). The pH of synthetic dairy wastewater is  
142 naturally around 8. However, for experiments the pH was reduced to between 4.5 – 5, a pH  
143 similar to that of half-strength Hutner’s medium, to ensure differences observed were not due  
144 to differences in pH.

### 145 *Measured parameters*

146 On day seven of the short-term experiments mass (fresh biomass), colony number, frond  
147 number and chlorophyll *a* fluorescence were measured. Frond and colony numbers were



148 counted by eye. Before weighing, plants were wrapped in tissue paper to remove excess water.  
149 The Relative Growth Rate (RGR) was calculated based on fresh weight measurements using  
150 the formula below (Connolly & Wayne 1996):

$$151 \quad RGR = \frac{\ln \left( \frac{W_2}{W_1} \right)}{\Delta T}$$

152 Where  $W_1$  is initial fresh biomass (Day 0),  $W_2$  is final fresh biomass (Day 7),  $\Delta T$  is length of  
153 the experiment in days and  $\ln$  is the natural logarithm.

154 Chlorophyll *a* fluorescence was measured using pulse amplitude modulated chlorophyll *a*  
155 fluorometry (WALZ Imaging fluorometer, Effeltrich, Germany). For chlorophyll *a*  
156 fluorescence analysis, plants were dark adapted for 15 minutes immediately before  
157 measurements. Then, three random colonies from each Magenta were taken for analysis; the  
158 measured values of these three colonies were averaged together and treated as one replicate.  
159 The chlorophyll fluorescence analysis procedure is as follows; first, a low intensity modulated  
160 measuring light was turned on to measure  $F_0$  on the dark-adapted plant, and secondly a  
161 saturating pulse of light ( $2700 \mu\text{mol}/\text{m}^2/\text{s}$ ) was applied to obtain the maximum fluorescence  
162  $F_m$ . Subsequently, actinic light (photosynthetically active light of  $186 \mu\text{mol}/\text{m}^2/\text{s}$ ) was applied  
163 to the plants and at 20 second intervals saturating pulses were applied to measure  $F_m'$ , the  
164 maximum fluorescence under light-adapted conditions.  $F_t$  is the value of fluorescence  
165 immediately before the saturating pulse is applied, i.e. the steady-state value of fluorescence.  
166  $F_v/F_m$ , the maximum quantum efficiency of photosystem II (PSII), and  $Y(II)$ , the quantum  
167 efficiency of PSII under steady state light conditions were calculated according to Maxwell  
168 and Johnson (2000) using the following equations:

$$169 \quad F_v/F_m = (F_m - F_0)/F_m$$

$$170 \quad Y(II) = (F_m' - F_t)/F_m'$$

171 For the long-term 42-day experiment, mass (fresh biomass), colony number and frond number  
172 were measured every seven days before media were replaced with fresh synthetic wastewater.  
173 RGR was calculated based on growth over seven days.

#### 174 ***Data analysis***

175 Statistical analyses were conducted using R software (R 3.4.3). Numbers of independent  
176 replicates were 3 to 4, as stated in legends. One-way ANOVA and Welch's ANOVA were used  
177 to examine whether there were significant differences in RGR, Y(II) and Fv/Fm between  
178 treatment groups (excluding the Hutner's treatment group). Post-hoc tests Tukey and Games-  
179 Howell were used in pairwise comparisons of treatment groups (also excluding the Hutner's  
180 treatment group). Welch's ANOVA and Games-Howell tests were used when a dataset was  
181 not homoscedastic thus failing one of the assumptions of an ANOVA and Tukey test. In the  
182 42-day experiment average RGR between treatment groups was compared each week, Hutner  
183 being included in this analysis.

## 184 **Results and Discussion**

### 185 ***Investigation of components of synthetic dairy wastewater***

186 As part of a phytoremediation approach for dairy industry wastewater, *L. minor* was grown  
187 under laboratory conditions on synthetic dairy wastewater. However, it was found that growth  
188 rates were poor, and that colonies displayed extensive chlorosis. Chlorosis occurred relatively  
189 fast (within days) indicating toxicity rather than deficiency symptoms. Thus, a systematic desk-  
190 top study of all individual chemical components present in the synthetic medium was  
191 conducted. The concentration of each element in the synthetic wastewater was first calculated  
192 and then compared with the minimum required, the maximum tolerated and the optimal range  
193 of values (Table 2). Based on the data in table 2, a number of elements were selected that were  
194 present in non-optimal concentrations and that could potentially have a negative impact on *L.*

195 *minor* growth. The elements iron, manganese, sodium, chloride, potassium and sulphate were  
196 identified as falling into this category. Iron, chloride and manganese were present at  
197 concentrations at upper end of their optimal ranges. Potassium and sulphate were both present  
198 in concentrations at the lower end of their respective optimal ranges. Reductions of 100% in  
199 chloride, or sodium concentrations did not improve growth. Reductions of some 70% in iron  
200 or manganese concentrations did not improve growth, and neither did 10-fold increases in  
201 potassium or sulphate (data not shown). These experiments did not reveal any candidate to  
202 explain observed growth impairment on unmodified synthetic wastewater.

203 [Table 2 near here]

#### 204 ***Increasing concentrations of calcium relative to magnesium***

205 Synthetic dairy processing water has a low concentration of calcium (0.014 mM), notably lower  
206 than the optimal range of concentrations (0.2 – 20 mM). Furthermore, it was noted that the  
207 Ca:Mg ratio in the synthetic wastewater was 1:14.6, and this relative lack of calcium compared  
208 to magnesium might potentially also cause growth problems (Landolt & Kandeler 1987; Van  
209 Dam et al. 2010). To explore the roles of calcium, magnesium and the Ca:Mg ratio in  
210 controlling growth, an initial experiment was conducted in which *L. minor* plants were grown  
211 in medium containing increasing amounts of calcium, whilst the magnesium concentration was  
212 kept constant at 0.2 mM. The calcium concentrations ranged from 0.014 to 1.21 mM,  
213 translating to Ca:Mg ratios of 1:14.6 to 6.1:1, respectively. As a control, plants were grown on  
214 half-strength Hutner's medium which has a Ca:Mg ratio of 1:1.

215 Under control growth conditions, plants on half-strength Hutner's medium displayed vigorous  
216 growth (average RGR of 0.226 day<sup>-1</sup>) and had a healthy appearance. In contrast, plants growing  
217 on unmodified synthetic wastewater (Ca:Mg ratio of 1:14.6) had an average RGR of just 0.061  
218 day<sup>-1</sup> (Figure 1). These plants appeared chlorotic. When the synthetic dairy processing medium

219 was modified through the addition of calcium, RGR values increased significantly. In modified  
220 synthetic wastewater with a concentration of calcium greater than or nearly equal to that of  
221 magnesium, RGR values were 0.164 (Ca:Mg – 1:1.6), 0.163 (Ca:Mg – 1.2:1), 0.163 (Ca:Mg –  
222 3:1) and 0.148 (Ca:Mg – 6.1:1) day<sup>-1</sup>. From a low growth rate on medium with a Ca:Mg ratio  
223 of 1:14.6, RGR increased up until a Ca:Mg of 1:1.6 ratio, where RGR-values plateaued (Figure  
224 1). Analysis of variance (ANOVA) between treatments (excluding Hutner) showed significant  
225 variance among them,  $F(6, 14) = 7.606$ ,  $p < 0.001$ , indicating that there is a positive association  
226 between the Ca:Mg ratio and the RGR (day<sup>-1</sup>) of *L. minor*. A post-hoc Tukey test revealed  
227 which differences in RGR were significant. The RGR values of plants grown on synthetic  
228 wastewater of a Ca:Mg ratio of 1:3.3, 1:1.6, 1.2:1, 3:1 and 6.1:1 did not differ significantly  
229 from each other. The RGR of plants on synthetic waste of Ca:Mg ratio 1:14.6 differed  
230 significantly from that of plants on a Ca:Mg ratio of 1:1.6, 1.2:1 and 3:1,  $p < 0.05$ . While the  
231 RGR of plants on synthetic wastewater of Ca:Mg ratio of 1:8.2 differed significantly from that  
232 of plants on a Ca:Mg ratio of 1:3.3, 1:1.6, 1.2:1, 3:1 and 6.1:1,  $p < 0.05$ .

233 [Figure 1 near here]

234 To complement growth rate measurements, key photosynthetic parameters were measured in  
235 parallel. In particular Fv/Fm and Y(II) were analysed. Both of these parameters test if, and to  
236 what degree, a stressor is affecting photosystem II in the plant. The former refers to the plant  
237 in a dark-adapted state, while the latter refers to plants during photosynthesis at steady-state  
238 conditions. The control plants on half-strength Hutner's medium displayed good  
239 photosynthetic activity, as shown by average values of 0.65 for Fv/Fm and 0.39 for Y(II)  
240 (Figure 2). On synthetic wastewater with Ca:Mg ratios of 1:14.6 and 1:8.2, Fv/Fm and Y(II)  
241 values were considerably lower than those of the control plants. In synthetic media with Ca:Mg  
242 ratios of 1:3.3, 1:1.6, 1.2:1, 3:1 and 6.1:1, Fv/Fm and Y(II) values were similar, or higher, than  
243 those of plants on the half-strength Hutner's control. As the calcium concentration was

244 increased, Fv/Fm and Y(II) values increased up until a Ca:Mg ratio of 1.2:1 where values  
245 started to plateau. For Fv/Fm, ANOVA showed significant differences between synthetic  
246 wastewater treatments,  $F(6, 14) = 7.843$ ,  $p < 0.01$ , indicating that there was a positive association  
247 between changing the Ca:Mg ratio and Fv/Fm, an indicator of the health of *L. minor*. A post-  
248 hoc Tukey test showed that the significant variance (shown as letters above the bars in Figure  
249 2) between the groups ( $p < 0.01$ ) was between plants growing on a Ca:Mg ratio of 1:8.2 and  
250 those on a ratio of 1:3.3, 1:1.6, 1.2:1, 3:1 and 6.1:1. For Y(II) ANOVA also showed significant  
251 differences between treatment groups,  $F(6, 14) = 22.62$ ,  $p < 0.001$ . This indicates, similarly to  
252 Fv/Fm, there was a positive association between the Ca:Mg ratio and Y(II), an indicator of the  
253 photosynthetic efficiency of the plant. A post-hoc Tukey test shows this significant variation  
254 was between plants growing on synthetic wastewater of Ca:Mg ratios 1:14.6 and 1:8.2, which  
255 had low Y(II) values, and plants growing on Ca:Mg ratios of 1:3.3, 1:1.6, 1.2:1, 3:1 and 6.1:1,  
256 which had higher Y(II) values.

257 [Figure 2 near here]

258 These observations are important from a management perspective and show that the addition  
259 of calcium turns synthetic dairy industry wastewater in a suitable medium for growth of *L.*  
260 *minor*. However, the observations do not inform whether the positive effects of calcium on *L.*  
261 *minor* growth and photosynthesis are due to a rise in calcium concentration and/or an increase  
262 in the Ca:Mg ratio.

### 263 ***Increasing concentrations of magnesium relative to calcium***

264 To explore in more detail the importance of the Ca:Mg ratio, the ratio was changed in favour  
265 of magnesium at adequate calcium concentrations. Calcium concentrations were kept constant  
266 at 0.12 mM, which was previously shown to accommodate good growth (Figure 1). *L. minor*  
267 was grown with magnesium concentrations ranging from 0.2 to 4.99 mM, yielding Ca:Mg

268 ratios of 1:1.6 through to 1:41.2, respectively. It can be seen (Figure 3) that RGR progressively  
269 decreased as the concentration of magnesium was increased in the synthetic wastewater. The  
270 average RGR at a Ca:Mg ratio of 1:1.6 was 0.21 day<sup>-1</sup>. RGR decreased to a value of 0.035 day<sup>-1</sup>  
271 at a Ca:Mg ratio of 1:41.2. The *L. minor* plants at the high magnesium concentration (4.99  
272 mM) were characterised by both poor growth and chlorosis. An analysis of variance (ANOVA)  
273 showed significant differences in RGR between groups,  $F(3, 12) = 7.996$ ,  $p < 0.05$ , indicating  
274 the significant influence of magnesium on plant growth. A post-hoc Tukey test showed  
275 significant differences between the RGR of plants growing on a Ca:Mg ratio of 1:1.6 and that  
276 of plants in Ca:Mg ratios 1:16.5 ( $p < 0.05$ ) and 1:41.2 ( $p < 0.01$ ).

277 [Figure 3 near here]

278 Fv/Fm and Y(II) values reflect the same trend seen in the RGR data in Figure 3. As the  
279 magnesium concentration increased in the synthetic wastewater the Fv/Fm and Y(II) values  
280 were reduced (Figure 4). At a Ca:Mg ratio of 1:1.6, the average Fv/Fm was 0.752 and Y(II)  
281 was 0.426. The lowest photosynthetic efficiencies were found at a ratio of Ca:Mg of 1:41.2,  
282 where Fv/Fm was 0.099 and Y(II) was 0.007. A Welch's ANOVA test (used because of  
283 heteroscedasticity in the dataset) showed that there was a significant difference between the  
284 treatment groups,  $p < 0.01$ . A Games-Howell post-hoc test (also used because of  
285 heteroscedasticity in the dataset) showed that there were significant differences in Fv/Fm and  
286 Y(II), shown as letters above bars in Figure 4. The Fv/Fm of plants growing in Ca:Mg ratio of  
287 1:41.2 differed significantly from that of plants growing in Ca:Mg ratio of 1:1.6 ( $p < 0.05$ ) and  
288 of 1:8.2 ( $p < 0.05$ ). Similarly, the Y(II) of plants growing in Ca:Mg ratio of 1:41.2 differed  
289 significantly from that of plants growing in a ratio of 1:1.6 ( $p < 0.001$ ) and 1:8.2 ( $p < 0.05$ ).

290 [Figure 4 near here]

291 The observed negative effects of magnesium additions on both RGR and photosynthetic  
292 parameters can be attributed to the rise in magnesium concentration and/or the decrease in the  
293 Ca:Mg ratio. High concentrations of magnesium have been reported to be toxic to Lemnaceae,  
294 although this effect can be countered by increasing the Ca:Mg ratio (Van Dam et al. 2010). To  
295 further explore this point, an experiment was conducted in which the concentrations of both  
296 calcium and magnesium were increased, so as to maintain an optimised 1:1.6 Ca:Mg ratio,  
297 while using magnesium concentrations that were earlier found to be toxic (Figures 3 and 4).

### 298 ***Simultaneous increase of calcium and magnesium concentrations***

299 *L. minor* was grown in synthetic dairy wastewater with a Ca:Mg ratio of 1:1.6 and a range of  
300 calcium (0.12 – 3.12 mM) and magnesium (0.2 – 5.14 mM) concentrations. The RGR for plants  
301 growing on synthetic wastewater was slightly lower than that of the control on half-strength  
302 Hutner's medium, across all concentrations. The highest RGR for plants growing on synthetic  
303 waste came from those growing on a Ca:Mg ratio of 1:1.6 at the lowest concentration. As the  
304 absolute concentrations of calcium and magnesium increased plant RGR decreased moderately  
305 (Figure 5). An ANOVA showed a significant difference between the RGR of *L. minor* in  
306 different synthetic wastewater treatments,  $F(4,15) = 3.89$ ,  $p < 0.05$  indicating that increased  
307 concentrations of both calcium and magnesium had a significant impact on *L. minor* RGR. A  
308 post-hoc Tukey test revealed significant differences in RGR between plants growing on  
309 wastewater of calcium and magnesium concentrations of 0.21 and 0.2 mM, respectively, and  
310 that of plants growing on the two highest concentrations ( $p < 0.05$  for both). Once the  
311 concentrations of calcium and magnesium were increased to 2.5 and 4.1 mM, respectively, or  
312 above, RGR significantly decreased.

313 [Figure 5 near here]

314 Fv/Fm and Y(II) measurements revealed non-significant differences between plants that were  
315 grown in synthetic wastewater containing different concentrations of calcium and magnesium  
316 (Figure 6). When treatments were compared using an ANOVA test no significant differences  
317 were found; Fv/Fm,  $F(4, 15)=0.524$ ,  $p=0.72$ ; Y(II),  $F(4, 15)=0.809$ ,  $p=0.538$ . Furthermore,  
318 Fv/Fm and Y(II) values for plants grown on synthetic wastewater were similar to those on half-  
319 strength Hutner's medium, the control.

320 [Figure 6 near here]

321 A comparison of figures 3 and 4 with figures 5 and 6 shows that a magnesium concentration  
322 of 1 mM or more can have a significantly negative impact on *L. minor* growth and  
323 photosynthesis (Figures 3 and 4) when calcium concentrations are low. A magnesium  
324 concentration of 4.99 mM (Figures 3 and 4) can even cause death of plants. However, the same  
325 high magnesium concentrations have only a minor effect on RGR, and no effect on  
326 photosynthesis, when the ratio of Ca:Mg is kept at 1:1.6 (Figures 5 and 6). Thus, the data  
327 emphasise the antagonism between calcium and magnesium, and the relative importance of the  
328 Ca:Mg ratio.

329 In Lemnaceae, antagonistic interactions between calcium and magnesium have been observed,  
330 and these impacted upon the aquatic toxicity of magnesium sulphate (Van Dam et al. 2010),  
331 the degradation process of starch (Appenroth & Gabrys 2003) and the germination of turions  
332 (Appenroth et al. 1999). Van Dam *et al.* (2010) observed the toxic effects of magnesium on  
333 *Lemna aequinoctialis* (as well as on algae and other freshwater species), and the alleviation of  
334 these toxic effects through the addition of calcium. Similarly, Appenroth *et al.* (1999) observed  
335 that a high magnesium concentration, in a near calcium-free environment, inhibited turion  
336 germination in *Spirodela polyrhiza*. Furthermore, these authors also observed that the  
337 inhibiting effect of magnesium could be abolished by either adding calcium or by reducing the



338 concentration of magnesium. The explanation for this observed antagonism is that magnesium  
339 is capable of competing with and inhibiting calcium uptake while also affecting calcium-  
340 dependent processes (for example turion germination).

341 Thus, for phytoremediation approaches the calcium concentration, the magnesium  
342 concentration and the Ca:Mg ratio in wastewater all need to be considered. This conclusion is  
343 important in the context of remediation of dairy industry processing waste which can have a  
344 highly variable Ca:Mg ratio. For example, standardised synthetic dairy wastewater has a  
345 Ca:Mg ratio of 1:14.6 but Demirel and Yenigun (2004) found a Ca:Mg ratio of 1:1.5 (1.37:2.06  
346 mM) in milk processing waste while Goyal and Gandhi (2009) found a Ca:Mg ratio of 5:1  
347 (7.26:1.48 mM) from cheese whey. Thus, an unfavourable Ca:Mg ratio is a fact that needs to  
348 be considered when developing a protocol for the phytoremediation of dairy processing waste.

#### 349 ***Long-term growth on modified synthetic dairy wastewater***

350 Notwithstanding the importance of the Ca:Mg ratio, we note a small decrease in RGR at higher  
351 magnesium concentrations, even where the Ca:Mg ratio is constant (Figure 5). It is possible  
352 that this is “pure” magnesium toxicity, which is slowly building up over time. To explore this  
353 in more detail, short term (7 day) experiments were complemented by 42-day experiments, in  
354 which the Ca:Mg ratio was kept at 1:1.6, but at two different concentrations of calcium and  
355 magnesium. Under these conditions, healthy growth of *L. minor* was observed for the full  
356 length of the experiment, and growth rates were very similar to those obtained on half-strength  
357 Hutner’s medium. ANOVA tests, confirmed this observation, showing no significant  
358 difference between the RGR of *L. minor* grown on synthetic wastewater with a Ca:Mg ratio of  
359 1:1.6, and those grown on Hutner’s throughout the 42-day experiment (Figure 7). Neither was  
360 there a difference between plants on low concentrations (0.12 mM Ca and 0.2 mM Mg ) of  
361 calcium and magnesium, and those on high ones (3.12 mM Ca and 5.14 mM Mg). The general

362 trend is that the RGR of *L. minor* increased each week up to the end of the experiment at day  
363 42, and this is presumably due to the frequent replacement of medium.

364 [Figure 7 near here]

## 365 **Conclusions**

366 Lemnaceae can be used to clean-up a variety of different wastewaters. However, some waste  
367 streams are not suitable for growth of Lemnaceae and need to be modified to facilitate growth  
368 and phytoremediation. Here we confirm the hypothesis that the Ca:Mg ratio can be a major  
369 determinant of growth and photosynthesis, both in short and long-term trials. Yet, the data also  
370 show that growth can be restored by the addition of calcium-sulphate, a procedure that is  
371 feasible in a commercial setting. An addition of calcium to the synthetic wastewater to reach a  
372 concentration of 0.12 mM, compared to 0.2 mM of magnesium, resulted in RGRs of 0.164 -  
373 0.330 day<sup>-1</sup>. These rates compare well with those of plants growing in half-strength Hutner's  
374 medium, which achieve RGRs between 0.226 – 0.330 day<sup>-1</sup>. It is acknowledged that calcium  
375 can act as an antagonist for other elements, such as iron, manganese and potassium. However,  
376 initial experiments showed that a reduction in the concentrations of iron and manganese did  
377 not reverse the acute toxicity seen in plants. Thus, higher concentrations of calcium in later  
378 experiments acting as an antagonist towards these elements would not have reversed acute  
379 toxicity either. The data presented in this paper show convincingly that it is the ratio between  
380 calcium and magnesium, which is causing acute toxicity to *L. minor* in this case. A Ca:Mg ratio  
381 of 1:1.6 or greater is necessary for *Lemna minor* growth, and therefore phytoremediation of the  
382 dairy industry processing wastewater.

## 383 **Acknowledgements**

384 We wish to thank the Irish Environmental Protection Agency (grant 2016-W-LS-11) for  
385 funding this study. MJ acknowledges support by WoB.

## 386 **Declaration of interest statement**

387 The authors wish to disclose no conflict of interest.

## 388 References

- 389 Anderson KE, Lowman Z, Stomp A-M, Chang J. 2011. Duckweed as a Feed Ingredient in Laying  
390 Hen Diets and its Effect on Egg Production and Composition. *Int J Poult Sci.* 10:4–7. doi:  
391 10.3923/ijps.2011.4.7
- 392 Appenroth K-J, Gabrys H. 2003. Ion antagonism between calcium and magnesium in phytochrome-  
393 mediated degradation of storage starch in *Spirodela polyrhiza*. *Plant Sci.* 165:1261–1265. doi:  
394 10.1016/S0168-9452(03)00334-0
- 395 Appenroth KJ, Gabrys H, Scheuerlein RW. 1999. Ion antagonism in phytochrome-mediated calcium-  
396 dependent germination of turions of *Spirodela polyrhiza* (L.) Schleiden. *Planta.* 208:583–587. doi:  
397 10.1007/s004250050596
- 398 Appenroth KJ, Sree KS, Böhm V, Hammann S, Vetter W, Leiterer M, Jahreis G. 2017. Nutritional  
399 value of duckweeds (Lemnaceae) as human food. *Food Chem.* 217:266–273. doi:  
400 10.1016/j.foodchem.2016.08.116
- 401 Brady KU, Kruckeberg AR, Bradshaw Jr. HD. 2005. Evolutionary Ecology of Plant Adaptation to  
402 Serpentine Soils. *Annu Rev Ecol Evol Syst.* 36:243–266. doi:  
403 10.1146/annurev.ecolsys.35.021103.105730
- 404 Caicedo JR, Van Der Steen NP, Arce O, Gijzen HJ. 2000. Effect of total ammonia nitrogen  
405 concentration and pH on growth rates of duckweed (*Spirodela polyrrhiza*). *Water Res.* 34:3829–3835.  
406 doi: 10.1016/S0043-1354(00)00128-7
- 407 Carvalho F, Prazeres AR, Rivas J. 2013. Cheese whey wastewater: Characterization and treatment.  
408 *Sci Total Environ.* 445–446:385–396. doi: 10.1016/j.scitotenv.2012.12.038
- 409 Cheng JJ, Stomp AM. 2009. Growing Duckweed to recover nutrients from wastewaters and for  
410 production of fuel ethanol and animal feed. *Clean - Soil, Air, Water.* 37:17–26. doi:  
411 10.1002/clen.200800210
- 412 Conley DJ, Paerl HW, Howarth RW, Boesch DF, Seitzinger SP, Havens KE, Lancelot C, Likens GE.  
413 2009. Controlling eutrophication: nitrogen and phosphorus. *Science* (80- ). 323:1014–1015. doi:  
414 10.1126/science.1167755
- 415 Connolly J, Wayne P. 1996. Asymmetric competition between plant species. *Oecologia.* 108:311–  
416 320. doi: 10.1007/BF00334656
- 417 Van Dam RA, Hogan AC, McCullough CD, Houston MA, Humphrey CL, Harford AJ. 2010. Aquatic  
418 toxicity of magnesium sulfate, and the influence of calcium, in very low ionic concentration water.  
419 *Environ Toxicol Chem.* 29:410–421. doi: 10.1002/etc.56
- 420 Demirel B, Yenigun O. 2004. Anaerobic acidogenesis of dairy wastewater: The effects of variations  
421 in hydraulic retention time with no pH control. *J Chem Technol Biotechnol.* 79:755–760. doi:  
422 10.1002/jctb.1052
- 423 Diaz RJ, Rosenberg R. 2008. Spreading Dead Zones and Consequences for Marine Ecosystems.  
424 *Science* (80- ). 321:926–929. doi: 10.1126/science.1156401
- 425 Gil-Pulido B, Tarpey E, Almeida EL, Finnegan W, Zhan X, Dobson ADW, O’Leary N. 2018.  
426 Evaluation of dairy processing wastewater biotreatment in an IASBR system: Aeration rate impacts  
427 on performance and microbial ecology. *Biotechnol Reports.* 19:e00263. doi:  
428 10.1016/j.btre.2018.e00263
- 429 Goyal N, Gandhi DN. 2009. Comparative Analysis of Indian Paneer and Cheese Whey for Electrolyte  
430 Whey Drink. *World J Dairy Food Sci.* 4:70–72.

- 431 Halstead RL, MacLean AJ, Nielsen KF. 1958. Ca: Mg ratios in soil and the yield and composition of  
432 alfalfa. *Can J Soil Sci.* 38:85–93. doi: 10.4141/cjss58-014
- 433 Hutner SH. 1953. Comparative physiology of heterotrophic growth in higher plants. In: *Growth Differ*  
434 *plants.* Ames (IA): Iowa State College Press; p. 417–447.
- 435 Ince O. 1998. Performance of a two-phase anaerobic digestion system when treating dairy  
436 wastewater. *Water Res.* 32:2707–2713. doi: 10.1016/S0043-1354(98)00036-0
- 437 Janczukowicz W, Zieliński M, Debowski M. 2008. Biodegradability evaluation of dairy effluents  
438 originated in selected sections of dairy production. *Bioresour Technol.* 99:4199–4205. doi:  
439 10.1016/j.biortech.2007.08.077
- 440 Kolev Slavov A. 2017. General characteristics and treatment possibilities of dairy wastewater—A  
441 review. *Food Technol Biotechnol.* 55:14–28. doi: 10.17113/ftb.55.01.17.4520
- 442 Körner S, Das SK, Veenstra S, Vermaat JE. 2001. The effect of pH variation at the  
443 ammonium/ammonia equilibrium in wastewater and its toxicity to *Lemna gibba*. *Aquat Bot.* 71:71–  
444 78. doi: 10.1016/S0304-3770(01)00158-9
- 445 Körner S, Lyatuu GB, Vermaat JE. 1998. The influence of *Lemna gibba* L. on the degradation of  
446 organic material in duckweed-covered domestic wastewater. *Water Res.* 32:3092–3098. doi:  
447 10.1016/S0043-1354(98)00054-2
- 448 Körner S, Vermaat JE, Veenstra S. 2003. The Capacity of Duckweed to Treat Wastewater. *J Environ*  
449 *Qual.* 32:1583–1590. doi: 10.2134/jeq2003.1583
- 450 Lahive E, O’Halloran J, Jansen MAK. 2012. Frond development gradients are a determinant of the  
451 impact of zinc on photosynthesis in three species of Lemnaceae. *Aquat Bot.* 101:55–63. doi:  
452 10.1016/j.aquabot.2012.04.003
- 453 Landolt E. 1986. Biosystematic investigations in the family of duckweeds (Lemnaceae). The family  
454 of Lemnaceae—a monographic study, volume 1. Stiftung Rübel, Zürich: Veröffentlichungen des  
455 Geobotanischen Institutes der ETH.
- 456 Landolt E, Kandeler R. 1987. Biosystematic investigations in the family of duckweeds (Lemnaceae).  
457 The family of Lemnaceae - a monographic study, volume 2. Stiftung Rübel, Zürich:  
458 Veröffentlichungen des Geobotanischen Institutes der ETH.
- 459 Li L, Liu M, Wu M, Jiang C, Chen X, Ma X, Liu J, Li W, Tang X, Li Z. 2017. Effects of duckweed  
460 (*Spirodela polyrrhiza*) remediation on the composition of dissolved organic matter in effluent of scale  
461 pig farms. *J Environ Sci.* 55:247–256. doi: 10.1016/j.jes.2016.06.033
- 462 Liu C, Dai Z, Sun H. 2017. Potential of duckweed (*Lemna minor*) for removal of nitrogen and  
463 phosphorus from water under salt stress. *J Environ Manage.* 187:497–503. doi:  
464 10.1016/j.jenvman.2016.11.006
- 465 Loew O, May DW. 1901. The Relation of Lime and Magnesia to Plant Growth: I. Liming of Soils  
466 from a Physiological Standpoint. Washington: US Government Printing Office.
- 467 Maxwell K, Johnson GN. 2000. Chlorophyll fluorescence - A practical guide. *J Exp Bot.* 51:659–668.  
468 doi: 10.1093/jxb/51.345.659
- 469 Paolacci S, Harrison S, Jansen MAK. 2016. A comparative study of the nutrient responses of the  
470 invasive duckweed *Lemna minuta*, and the native, co-generic species *Lemna minor*. *Aquat Bot.*  
471 134:47–53. doi: 10.1016/j.aquabot.2016.07.004
- 472 Ryan M, Walsh G. 2016. The Characterisation of Dairy Waste and the Potential of Whey for  
473 Industrial Fermentation. Limerick: University of Limerick. Report No: 173.
- 474 Sree KS, Adelman K, Garcia C, Lam E, Appenroth KJ. 2015. Natural variance in salt tolerance and

- 475 induction of starch accumulation in duckweeds. *Planta*. 241:1395–1404. doi: 10.1007/s00425-015-  
476 2264-x
- 477 Tarpey E. 2016. An Investigation into the Use of IASBRs for Treatment of Dairy Processing  
478 Wastewater, MEng Thesis. Galway: National University of Ireland.
- 479 Tikariha A, Sahu O. 2014. Study of Characteristics and Treatments of Dairy Industry Waste Water. *J*  
480 *Appl Environ Microbiol*. 2:16–22. doi: 10.12691/jaem-2-1-4
- 481 Wang W. 1986. Toxicity tests of aquatic pollutants by using common duckweed. *Environ Pollution*  
482 *Ser B, Chem Phys*. 11:1–14. doi: 10.1016/0143-148X(86)90028-5
- 483 Zayed A, Gowthaman S, Terry N. 1998. Phytoaccumulation of Trace Elements by Wetland Plants: I.  
484 Duckweed. *J Environ Qual*. 27:715–721. doi: 10.2134/jeq1998.00472425002700030032x
- 485 Ziegler P, Adelman K, Zimmer S, Schmidt C, Appenroth KJ. 2015. Relative in vitro growth rates of  
486 duckweeds (Lemnaceae) - the most rapidly growing higher plants. *Plant Biol*. 17:33–41. doi:  
487 10.1111/plb.12184
- 488

489 **Tables**

490 Table 1. Composition synthetic dairy wastewater (Tarpey 2016).

<b>Chemical</b>	<b>Concentration mg/L<sup>1</sup></b>	<b>Molar concentration mM<sup>1</sup></b>
Ammonium chloride	167.3	3.13
Urea	129.9	2.16
Disodium phosphate	50	0.35
Potassium bicarbonate	50	0.50
Sodium bicarbonate	130	1.55
Calcium chloride dihydrate	2	0.018
Magnesium sulphate heptahydrate	50	0.20
Manganese sulphate monohydrate	2	0.012
Iron sulphate heptahydrate	35	0.126
Zinc sulphate heptahydrate	2.15	0.007
Cobalt chloride hexahydrate	1.2	0.005
Manganese chloride tetrahydrate	4.95	0.025
Copper sulphate pentahydrate	1.25	0.005
Nickel chloride hexahydrate	0.95	0.004
Sodium molybdate dihydrate	1.1	0.005
Boric acid	0.07	0.001
Sodium selenite	0.49	0.003
EDTA	100	0.342

491 <sup>1</sup>Indicates the final concentration of each compound in the synthetic wastewater media

492 Table 2. Compound concentrations in half-strength Hutner's medium and synthetic wastewater  
 493 with duckweed requirements and tolerance concentrations

Compound	Hutner (mM)	Synthetic wastewater (mM)	Minimum Required (mM)	Maximum Tolerated (mM)	'Optimal' Range (mM)
Ammonia/ammonium <sub>a,b,c</sub>	NP	3.33	0.18	> 5.87	1.2 - 2.9
Nitrate <sup>a</sup>	9.06	NP	0.05	> 16.13	0.05 - 4.8
Urea	NP <sup>i</sup>	2.16	ND	ND	ND
Total Nitrogen <sup>d</sup>	9.07	8.17	0.005	149.93	0.2 - 25
Phosphate <sup>a,d</sup>	3.00	0.35	0.0003	9.48	0.001 - 0.95
Total Phosphorous <sup>a,d</sup>	3.00	0.35	0.0003	9.69	0.001 - 0.97
Potassium <sup>d</sup>	5.96	0.50	0.01	51.15	0.50 - 9.97
Sodium <sup>d,e,f</sup>	0.02	1.55	0	217.49	0 - 10.87
Magnesium <sup>a,d</sup>	2.97	0.20	0.004	32.92	0.21 - 9.87
Calcium <sup>a,d</sup>	3.04	0.014	0.01	39.92	0.20 - 19.96
Sulphate <sup>d,g</sup>	3.00	0.35	0.01	59.33	0.52 - 20.82
Chloride <sup>e,g</sup>	NP	3.21	0.001	28.21	0.001 - 5.64
EDTA <sup>d</sup>	0.01	0.34	0	> 2.05	0 - 1.71
Iron <sup>d,g</sup>	0.004	0.13	0.004	0.39	0.004 - 0.18
Zinc <sup>g,h</sup>	0.003	0.007	0.001	0.31	0.002 - 0.09
Cobalt <sup>d</sup>	NP	0.005	0	0.05	0
Manganese <sup>g</sup>	0.001	0.037	0.0005	1.09	0.001 - 0.05
Copper <sup>d,h</sup>	0.0001	0.005	0.0001	0.03	0.00001 - 0.01
Nickel <sup>d,h</sup>	NP	0.004	0	0.02	0.00 - 0.002
Molybdenum <sup>d,h</sup>	0.0004	0.005	0.0002	1.04	0.0002 - 0.10
Boron <sup>d,h</sup>	0.02	0.001	0.0001	5.55	0.0001 - 0.46
Selenium <sup>d,h</sup>	NP	0.003	0	0.06	0

494 <sup>a</sup>(Paolacci et al. 2016)

495 <sup>b</sup>(Caicedo et al. 2000)

496 <sup>c</sup>(Körner et al. 2001)

497 <sup>d</sup>(Landolt & Kandeler 1987)

498 <sup>e</sup>(Sree et al. 2015)

499 <sup>f</sup>(Liu et al. 2017)

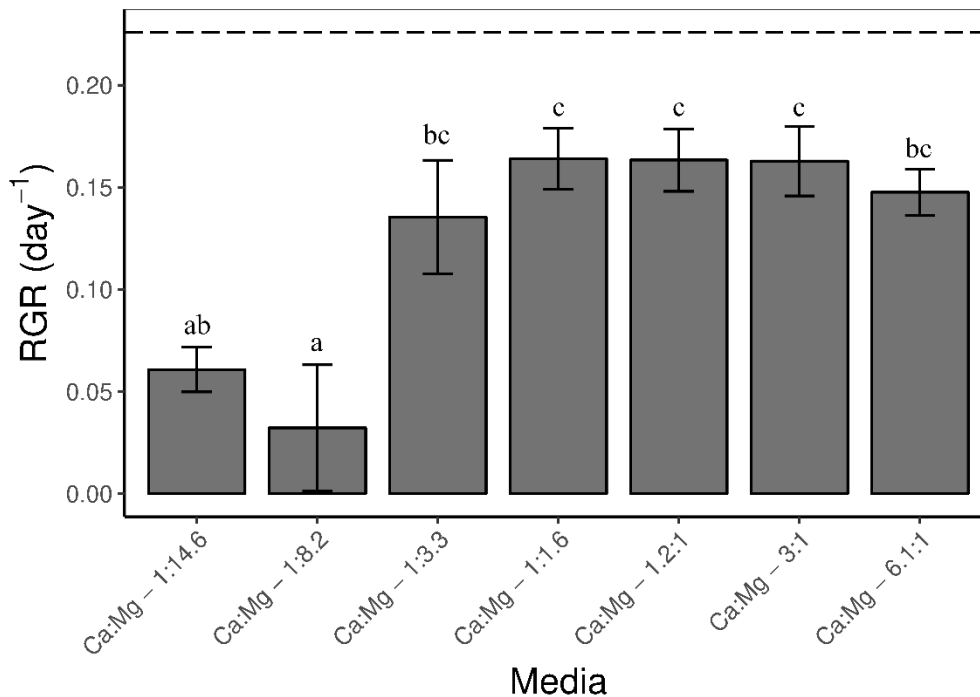
500 <sup>g</sup>(Wang 1986)

501 <sup>h</sup>(Lahive et al. 2012)

502 <sup>i</sup>NP – Not present



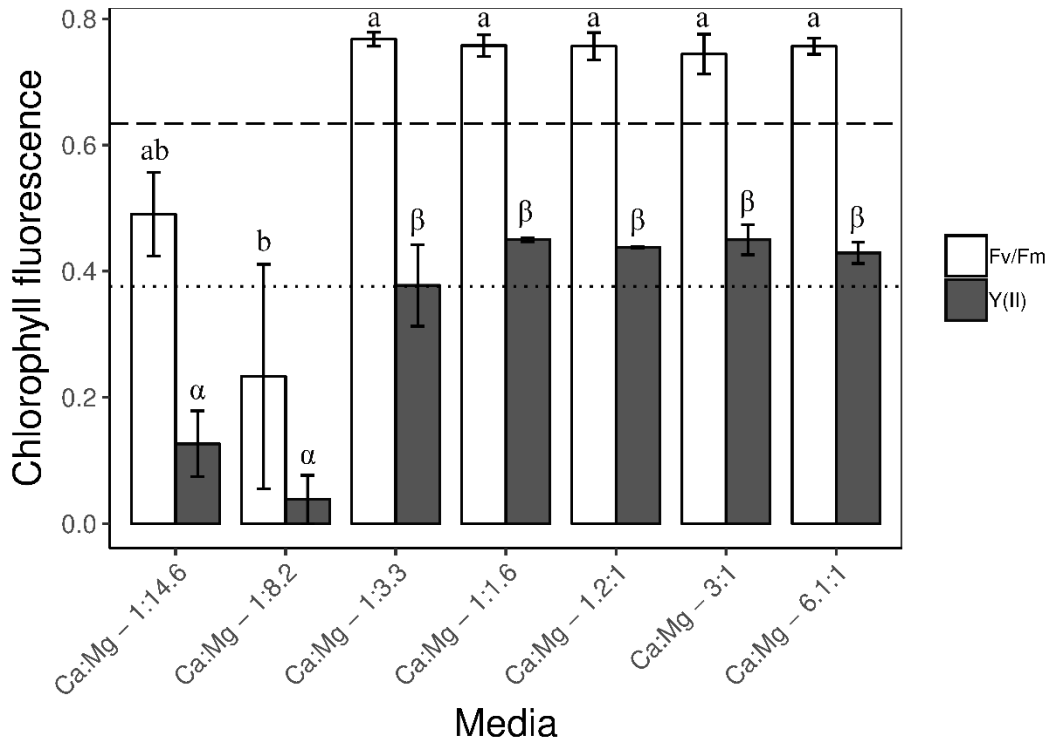
503 **Figure captions**



504

505 Figure 1. Biomass RGR of *L. minor* growing on synthetic wastewater medium with various  
506 Ca:Mg ratios (bars) and on half-strength Hutner's medium (dashed line). The unmodified  
507 synthetic waste medium has a Ca:Mg ratio of 1:14.6. Error bars represent standard error (n=3).  
508 ANOVA shows a positive association between Ca:Mg ratio and RGR ( $p < 0.05$ ). A post-hoc  
509 Tukey test which is represented on the graph as letters above the bars, shows the significant  
510 variance between groups. Bars that do not share a similar letter differ significantly ( $p < 0.05$ )  
511 from one another.

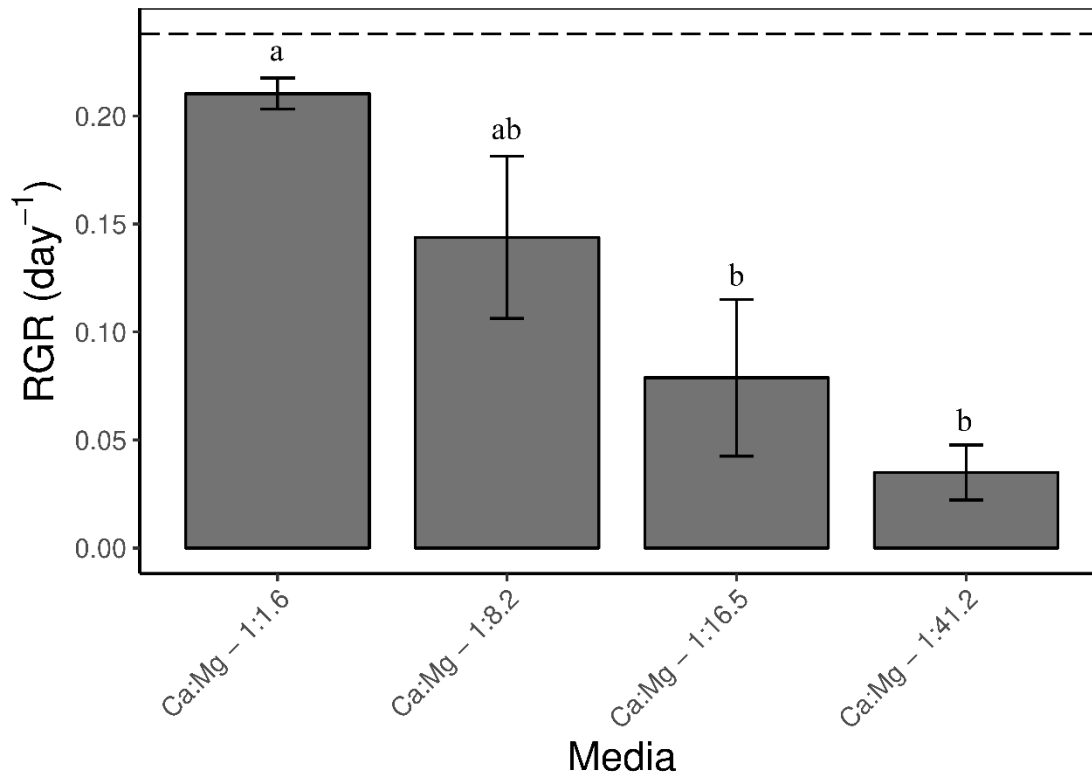
512



513

514 Figure 2. Fv/Fm and Y(II) values of *L. minor* growing on synthetic wastewater with different  
515 Ca:Mg ratios (bars) and on half-strength Hutner's medium (dashed and dotted lines, Fv/Fm  
516 and Y(II), respectively). Error bars represent standard error (n=3). ANOVA shows a positive  
517 association between Ca:Mg ratio and Fv/Fm, as well as Y(II) (p<0.01). A post-hoc Tukey test,  
518 indicated by letters above the bars, shows significant difference between Fv/Fm of plants at  
519 Ca:Mg ratio of 1:8.2 and all other ratios (excluding 1:14.6), p<0.01. For Y(II), a post-hoc Tukey  
520 test shows significant differences between plants growing at a Ca:Mg ratio of 1:14.6 and 1:8.2  
521 and plants growing in all other ratios (p<0.01). Bars that do not share a similar letter differ  
522 significantly from one another.

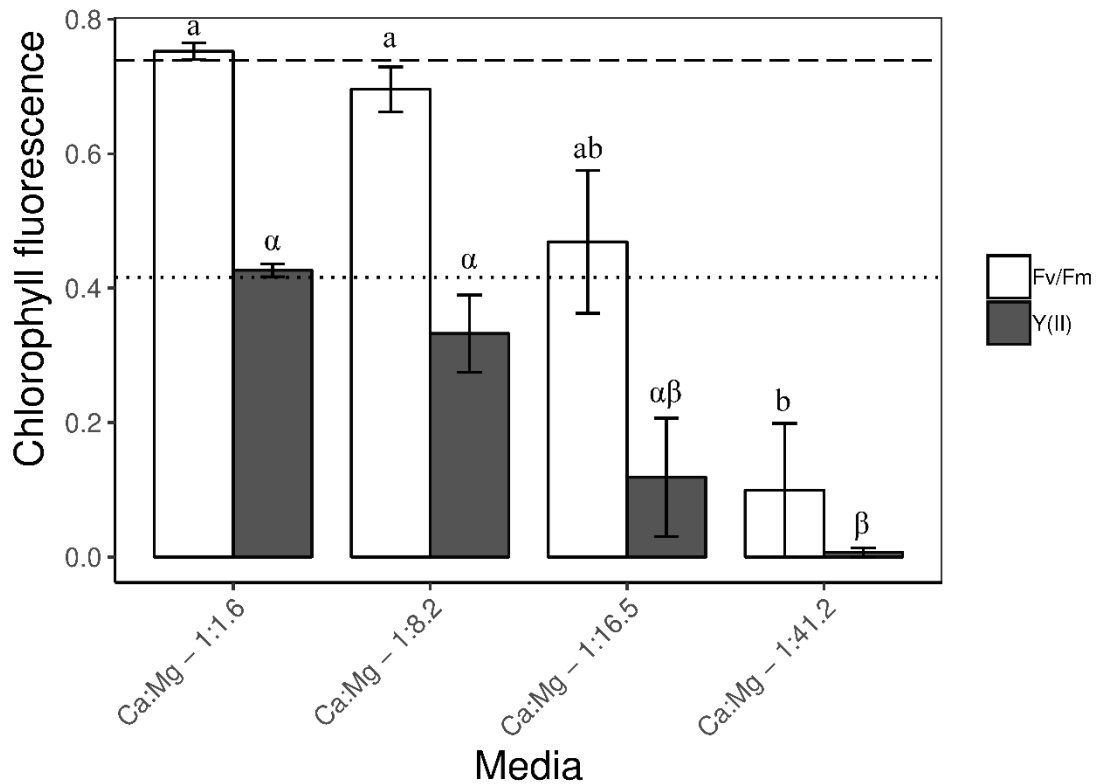
523



524

525 Figure 3. RGR of *L. minor* growing on synthetic wastewater with various Ca:Mg ratios (bars)  
 526 and on half-strength Hutner's medium (dashed line). Error bars represent standard error (n=4).  
 527 ANOVA showed significant differences in RGR between groups,  $F(3, 12) = 7.996$ ,  $p < 0.05$ .  
 528 Letters above the bars show the results of a Tukey post-hoc test. Bars that do not share a similar  
 529 letter differ significantly from one another.

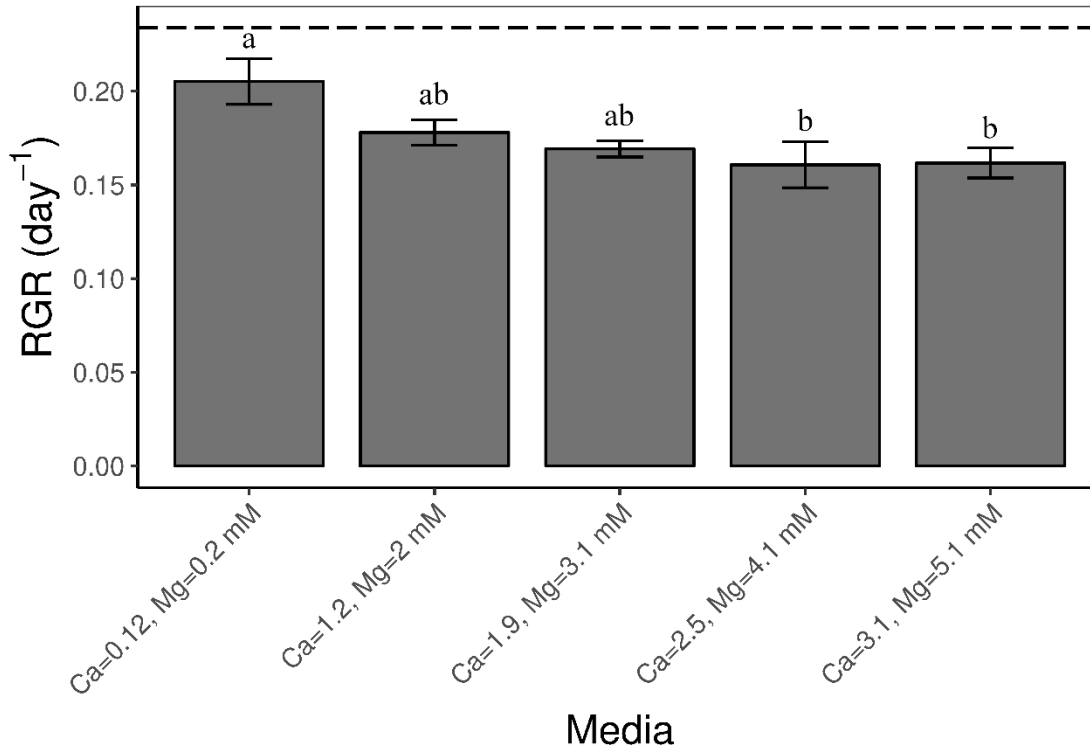
530



531

532 Figure 4. Fv/Fm and Y(II) of *L. minor* growing on synthetic wastewater with various Ca:Mg  
 533 ratios (bars) and on half-strength Hutner's medium (dashed and dotted lines, Fv/Fm and Y(II),  
 534 respectively). Error bars represent standard error (n=4). A Welch's ANOVA test showed that  
 535 there is significant difference between the treatment groups,  $p < 0.01$ . Letters above bars show  
 536 significant differences between groups as indicted in Games-Howell post-hoc test. Bars that do  
 537 not share a similar letter differ significantly from one another.

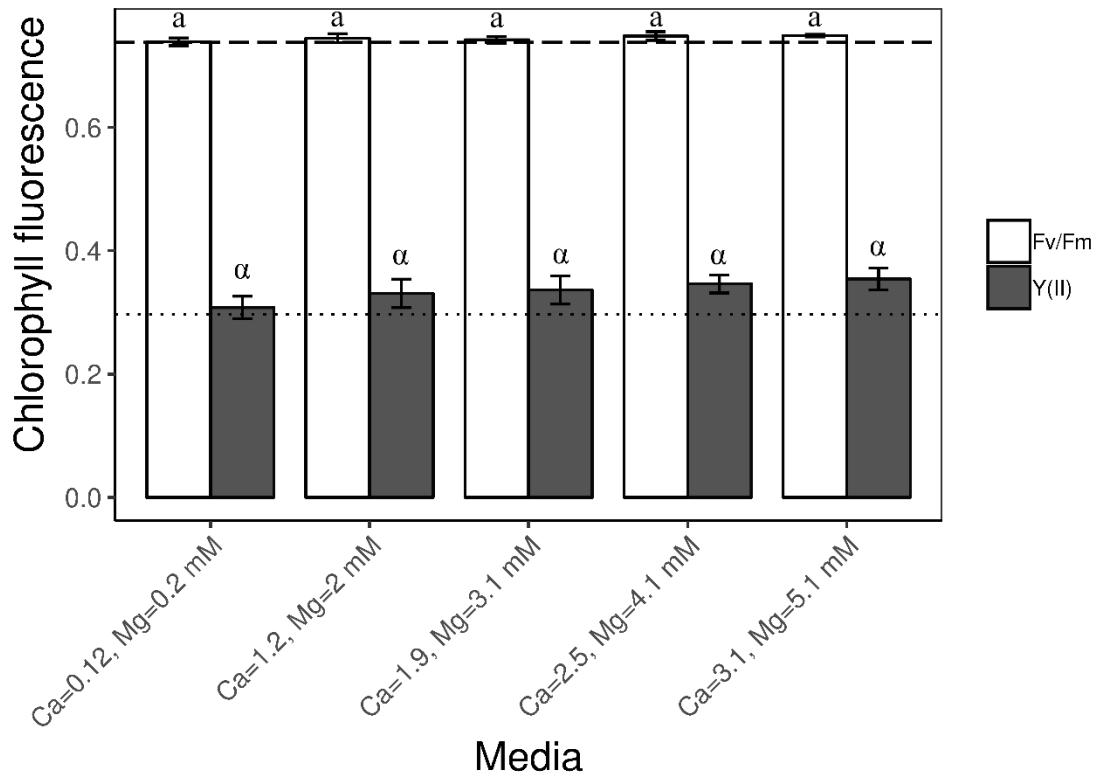
538



539

540 Figure 5. RGR of *L. minor* growing on synthetic wastewater of various concentrations of  
 541 calcium and magnesium (bars) and on half-strength Hutner's medium (dashed line). Error bars  
 542 represent standard error (n=4). ANOVA showed a significant difference between the RGR of  
 543 *L. minor* in different synthetic wastewater treatments,  $F(4,15) = 3.89$ ,  $p < 0.05$ . A post-hoc  
 544 Tukey test, indicated by letters above the bars, shows significant differences between the RGR  
 545 of *L. minor* in a calcium and magnesium concentration of 0.12 and 0.2 mM, respectively, and  
 546 those in the two highest concentrations. Bars that do not share a similar letter differ significantly  
 547 from one another.

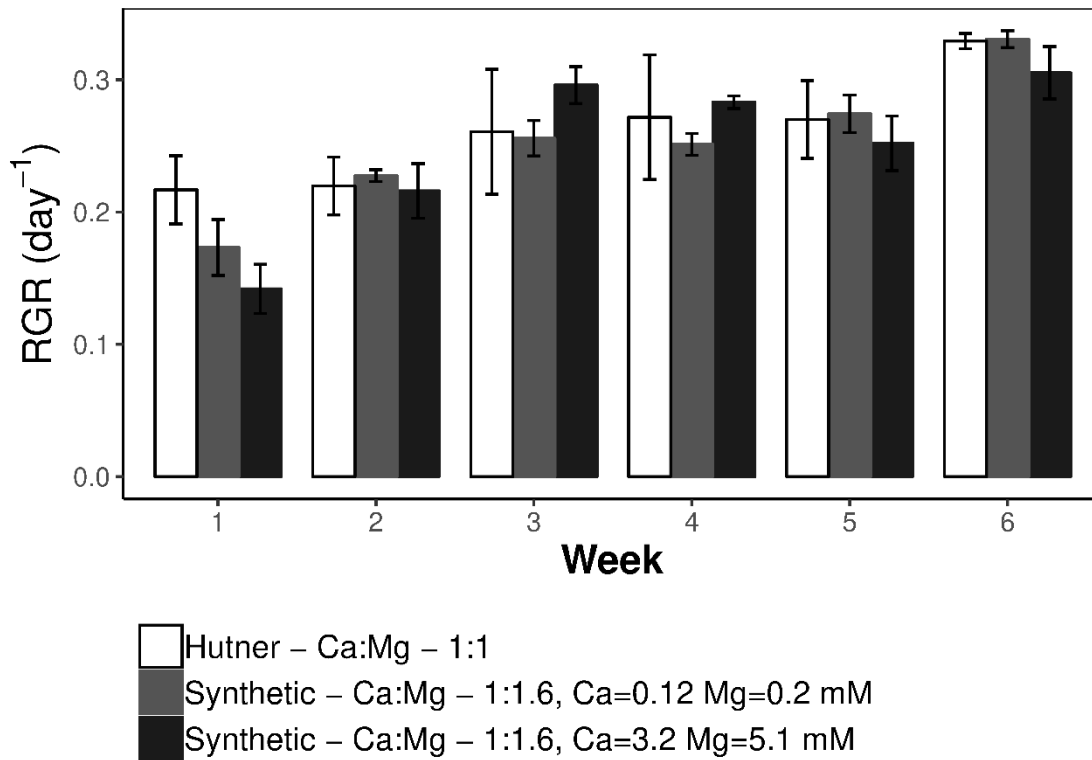
548



549

550 Figure 6. Fv/Fm and Y(II) for *L. minor* growing in synthetic wastewater of various  
 551 concentrations of calcium and magnesium (bars) and on half-strength Hutner's medium  
 552 (dashed and dotted lines, Fv/Fm and Y(II), respectively). Error bars represent standard error  
 553 (n=4). ANOVA did not reveal significant effects of concentration of either Fv/Fm or Y(II).  
 554 Measurement values of Fv/Fm and Y(II) do not differ significantly between treatments.

555



556

557 Figure 7. Weekly RGR of *L. minor* growing on synthetic wastewater (Ca:Mg ratio of 1:1.6 at  
 558 two concentrations) and half-strength Hutner's medium throughout a 42-day experiment.  
 559 ANOVA tests showed the RGR between treatments was not significantly different in any week  
 560 of the experiment (n=4). Results for week 1-6 respectively:  $F(2,9)=2.918$ ,  $p=0.105$ ;  
 561  $F(2,9)=0.112$ ,  $p=0.896$ ;  $F(2,9)=0.554$ ,  $p=0.593$ ;  $F(2,9)=0.337$ ,  $p=0.723$ ;  $F(2,9)=0.28$ ,  $p=0.762$ ;  
 562  $F(2,9)=1.319$ ,  $p=0.314$ .

563