

Stability and adaptability of runner peanut genotypes based on nonlinear regression and AMMI analysis

Roseane Cavalcanti dos Santos⁽¹⁾, Alineaura Florentino Silva⁽²⁾, Tarcísio Marcos Souza Gondim⁽¹⁾, José Oscar Lustosa de Oliveira Júnior⁽³⁾, Raimundo Bezerra de Araújo Neto⁽³⁾, Edvaldo Sagrilo⁽³⁾, Ramon Araujo de Vasconcelos⁽¹⁾, Péricles de Albuquerque Melo Filho⁽⁴⁾ and João Luís da Silva Filho⁽¹⁾

⁽¹⁾Embrapa Algodão, Caixa Postal 174, CEP 58428-095 Campina Grande, PB, Brazil. E-mail: roseane.santos@embrapa.br, tarcisio@embrapa.br, ramon@embrapa.br, joaoluiz@cnpa.embrapa.br ⁽²⁾Embrapa Semiárido, Caixa Postal 23, CEP 56302-970 Petrolina, PE, Brazil. E-mail: alinefs@cpatsa.embrapa.br ⁽³⁾Embrapa Meio-Norte, Avenida Duque de Caxias, nº 5.650, Buenos Aires, CEP 64006-220 Teresina, PI, Brazil. E-mail: oscar@cpamn.embrapa.br, rbezerra@cpamn.embrapa.br, sagrilo@cpamn.embrapa.br ⁽⁴⁾Universidade Federal Rural de Pernambuco, Rua Dom Manoel de Medeiros, s/nº, Dois Irmãos, CEP 52171-900 Recife, PE, Brazil. E-mail: pericles@depa.ufrpe.br

Abstract – The objective of this work was to estimate the stability and adaptability of pod and seed yield in runner peanut genotypes based on the nonlinear regression and AMMI analysis. Yield data from 11 trials, distributed in six environments and three harvests, carried out in the Northeast region of Brazil during the rainy season were used. Significant effects of genotypes (G), environments (E), and GE interactions were detected in the analysis, indicating different behaviors among genotypes in favorable and unfavorable environmental conditions. The genotypes BRS Pérola Branca and LViPE-06 are more stable and adapted to the semiarid environment, whereas LGoPE-06 is a promising material for pod production, despite being highly dependent on favorable environments.

Index terms: *Arachis hypogaea*, genotype x environment interaction, pod yield, seed yield, semiarid adaptation.

Estabilidade e adaptabilidade de genótipos de amendoim rasteiro baseadas em regressão não linear e análise AMMI

Resumo – O objetivo deste trabalho foi estimar a estabilidade e a adaptabilidade da produção de vagens e de sementes de genótipos de amendoim rasteiro com base em metodologias de regressão não linear e análise AMMI. Foram utilizados dados de produtividade de 11 ensaios, distribuídos em seis ambientes e três safras agrícolas, realizados na região Nordeste, no período das águas. Efeitos significativos de genótipos (G), ambientes (A) e da interação GA foram detectados nas análises e indicaram comportamento diferente entre os genótipos em condições ambientais favoráveis e desfavoráveis. Os genótipos BRS Pérola Branca e LViPE-06 são os mais estáveis e adaptados ao ambiente semiárido, enquanto o LGoPE-06 é material promissor para a produção de vagens, embora seja altamente dependente de ambiente favorável.

Termos para indexação: *Arachis hypogaea*, interação genótipo x ambiente, produtividade de vagens, produtividade de sementes, adaptação ao semiárido.

Introduction

High yield is the main goal of most plant breeding programs. Although yield traits are governed by a pool of major genes, the best performance of genotypes often depends on environmental conditions, resulting in a strong genotype and environment (GE) interaction. A strategy to reduce this interaction is based on cultivar selection, considering production stability in different locations in order to discriminate cultivars adapted to favorable and unfavorable environments.

Knowledge on the performance and adaptability of genotypes to particular environments is fundamental to

estimate the agronomical value of cultivars and for their recommendation for specific environments (Murakami et al., 2004). Moreover, performance stability allows for the identification of stable genotypes, with a predictable performance in different environments.

Several methodologies have been used to estimate phenotypic adaptability and stability, and differ as to the concepts and biometric procedures by which the GE interaction is measured. These methodologies are usually based on simple linear regression or multiple linear regression, nonlinear models, multivariate methods – including principal components analysis –, and methods that integrate the variance analysis of the

principal component analysis, such as the additive main effects and multiplicative interaction (AMMI) analysis (Eberhart & Russel, 1966; Toler & Burrows, 1998; Pacheco et al., 2005; Oliveira & Godoy, 2006).

Linear regression is the model most often used in the study of adaptability and stability, and provides information important for cultivar recommendation. This methodology describes the average standard response of each genotype to the environment; however, it does not contribute to identify the main relationships of the true causes of GE interaction (Ferreira et al., 2006; Oliveira & Godoy, 2006).

According to Ferreira et al. (2006), among breeders, the main criticisms of linear regression models are: the dependence of the environmental index on the mean performance of genotypes, and the use of biased estimators of the regression coefficients, since the independent variable is measured with error. Simple and bi-segmented models were proposed by Toler & Burrows (1998), which use indicator variables for favorable and unfavorable environments. In these models, the environmental index is a parameter to be estimated and, therefore, not dependent on the phenotypic means of the genotypic group.

Multivariate methods or univariate and multivariate methods are also used to estimate phenotypic stability. Among them, AMMI analysis integrates variance analysis of the main effects – i.e., genotypes and environments – with principal component analysis for the multiplicative effects of GE interaction (Oliveira & Godoy, 2006). AMMI analysis can contribute in the identification of the most stable and productive genotypes and in the further recommendation of region-specific cultivars. It also provides more precise estimates of genotypic responses and easy interpretation of the results in biplot graphs (Zobel et al., 1988). According to Toler & Burrows (1998), there is a relationship between parameters of the AMMI method and simple and bi-segmented regression models. Ferreira et al. (2006) recommend the simultaneous use of the Toler & Burrows (1998) and AMMI models to estimate phenotypic stability.

The objective of this work was to estimate the stability and adaptability of pod and seed yield in runner peanut (*Arachis hypogaea* L.) genotypes based on the nonlinear regression and AMMI analysis.

Materials and Methods

The field trials were carried out in the rainy season, during three years, in six environments of the Northeast region of Brazil (Table 1).

Late and early runner peanut genotypes were cultivated in plots (8-m row lengths), in which 0.5-m edges were considered border lines. Each row was spaced in 70 cm, and the sowing density was eight plants per meter. The evaluated genotypes are shown in Table 2.

Soil correction and fertilization were previously performed according to soil analysis. A randomized block design, with five replicates, was used in all trials. Crop management was done according to Santos et al. (2006).

Harvest was performed between 115 and 130 days after emergence. After post-harvest treatments, pod and seed yield data were subjected to joint analysis of variance to check for the presence of significant GE interaction effects, using the SAS software (SAS Institute, 1999). The heterogeneity of variance for each dependent variable was estimated by the ratio between the highest and lowest mean square observed in the individual analysis. The adjustment of the degrees of freedom for mean error and GE was done by the Cochran method, as described in Pimentel-Gomes (1990).

Each experiment was considered a distinct environment. The phenotypic adaptability and stability analysis (Toler & Burrows, 1998), and AMMI analysis (Zobel et al., 1988) were used. AMMI analysis joins additive analysis, to investigate main effects, and multiplicative analysis, to detect GE interaction. Therefore, it unites analysis of variance and principal component analysis. The equation proposed by Toler & Burrows (1998) is $\hat{Y}_{ij} = \hat{\alpha}_i + [Z_j \hat{\beta}_{1i} + (1 - Z_j) \hat{\beta}_{2i}] \hat{\mu}_j + \varepsilon_{ij}$, in which: \hat{Y}_{ij} is the mean performance of the i^{th} genotype as to yield in the j^{th} environment ($i = 1, 2, \dots, p$ genotypes; and $j = 1, 2, \dots, q$ environments); $\hat{\alpha}_i$ is the parameter that reflects the value of performance of genotype i on the intercept with $\hat{\mu}_j = 0$; $\hat{\beta}_{1i}$ and $\hat{\beta}_{2i}$ are the parameters that reflect the sensibility of the phenotypic performance of genotype i in unfavorable and favorable environments, respectively; $\hat{\mu}_j$ is the parameter that reflects the quality of the j^{th} environment; ε_{ij} is the mean experimental error (residue); and Z_j is the indicator variable, in which $Z_j = 1$ if $\hat{\mu}_j \leq 0$, and $Z_j = 0$ if $\hat{\mu}_j > 0$.

The genotype classification used in the present study was proposed by Toler & Burrows (1998), based on criteria shown in Table 3. According to Toler

Table 1. Environmental data from peanut trials carried out in the Northeast region of Brazil.

Environment	Coordinates	Altitude (m)	Climate	Total rainfall (mm)	Sowing	Soil
Petrolina, PE	09°23'34"S; 40°30'28"W	376	Semiarid	187	May	Oxisol
Abreu e Lima, PE	07°54'43"S; 34°54'10"W	19	Tropical	876	June	Entisol
Barbalha, CE	07°18'18"S; 39°18'07"W	414	Semiarid	613	July	Vertisol
Monteiro, PB	07°53'20"S; 37°07'12"W	599	Semiarid	239	March	Oxisol
Colinas, MA	06°01'33"S; 44°14'57"W	141	Tropical	1,160	February	Entisol
São João do Piauí, PI	08°21'29"S; 42°19'22"W	222	Semiarid	509	February	Oxisol

Table 2. Genealogy and agronomic traits of the evaluated peanut genotypes.

Genotype	Genealogy	Origin	Seed			Seeds per pod	Pod
			Color	Size	Shape		
BRS Pérola Branca	Cultivar	PB, Brazil	White	Large	Long	34	Large
LB308	Top line	PB, Brazil	White	Medium	Round	23	Large
LB408	Top line	PB, Brazil	White	Medium	Round	23	Large
IAC Caiapó	Cultivar	SP, Brazil	Tan	Large	Long	2	Large
Florunner	Cultivar	USA	Tan	Large	Long	2	Large
LGoPE-06	Top line	PE, Brazil	Tan	Extra large	Round	2	Extra large
LViPE-06	Top line	PE, Brazil	Tan	Extra large	Round	2	Extra large

Table 3. Criteria used for genotype classification according to Toler & Burrows (1998).

Group	Criterion	Classification of genotypes
A	Reject H ($\beta_1 = \beta_2$), accept $\beta_1 < 1 < \beta_2$	Convex and doubly desirable response
B	Accept H ($\beta_1 = \beta_2$), reject H ($\beta = 1$), but $\beta_1 > 1$	Simple linear response, desirable in high quality environments
C	Accept H ($\beta_1 = \beta_2$), accept H ($\beta = 1$)	Simple linear response, not deviating from mean response in the environments
D	Accept H ($\beta_1 = \beta_2$), reject H ($\beta = 1$), but $\beta < 1$	Simple linear response, desirable in poor quality environments
E	Reject H ($\beta_1 = \beta_1$), accept $\beta_1 > 1 > \beta_2$	Concave response and doubly undesirable.

(1990), a promising genotype should have doubly desirable response, with low response to undesirable environments ($\mu_j < 0$), and high performance in favorable conditions ($\mu_j > 0$).

In the AMMI model, the effects of interaction deviations are analyzed by singular value decompositions. The variation contained in the sum of squares of interactions is decomposed into "pattern", when the variation captured by the auto value is significant, and "noisy", when this variation is not significant. The interpretation of adaptability and stability is performed by graphical analysis, with the scores of the first two principal components plotted in a biplot. The closer the point is to the biplot origin, the more stable is

the genotype or the environment; the more distant, the greater the contribution to the interaction. Genotypes or environments close to each other have the same pattern as to the GE interaction (Dias, 2005).

Results and Discussion

Joint analysis of variance showed significant effect for all sources of variation for both pod and seed yield, indicating differential behavior of genotypes, which was not consistent with different environments (Table 4). Auto values, obtained by principal components analysis, over 70% of the variation found in GE interactions were captured by the first two principal components for both pod and seed yields; therefore, the interaction pattern of genotypes in all environments can be analyzed by visual inspection of the AMMI2 biplot (Figure 1). Oliveira & Godoy (2006) worked with 20 peanut genotypes distributed in ten environments and observed that the AMMI2 model captured almost 59% of variation.

The genotype BRS Pérola Branca (G1) was located closer to the origin, showing little contribution to GE interaction, and was also more stable (Figure 1). The best contribution to GE interaction was found in Colinas, MA (A5) and Barbalha, CE (A9), for pod and seed yield; whereas in Petrolina, PE (A1), Monteiro, PB (A2 and A8), and Abreu e Lima, PE (A7), the behavior of the genotype was very close, indicating

that, in future trials, only one location of those can be chosen to cut down expenses, without the risk of losing the representativeness of the trials. Silva Filho et al. (2008) used several methods, besides AMMI analysis, to compare the stability of cotton genotypes in 23 locations, in several states of Brazil, and also observed that it is possible to reduce the number of trials based on the clustering tendency in the same state.

The stability parameters, based on the nonlinear model of Toler (1990), for pod and seed yield are given in Table 5. The highest yields were obtained for extra-large seed genotypes – LViPE-06 (G7) and LGoPE-06 (G6) –, with 3.1 and 2.2 Mg ha⁻¹ for pod and seed yield, respectively. The early, large seed genotype BRS Pérola Branca also showed high yield, with 2.6 Mg ha⁻¹ in pods and 1.8 Mg ha⁻¹ in seeds.

Among the environments, the highest yields were obtained in Coruripe, AL (A11), Barbalha, CE (A4 and A9), and Colinas, MA (A5). In the semiarid region, which encompassed the municipalities of Petrolina, PE (A1), Monteiro, PB (A2), and São José do Piauí, PI (A6), runner genotypes showed lower performance, with pod and seed yields next to those obtained with upright genotypes in the rainy season (Santos et al., 2010).

Table 4. Joint analysis of variance of peanut genotypes evaluated in different environments as to pod and seed yield, and auto values obtained by principal components analysis.

Source of variation	Pod yield		Seed yield	
	DF	F calculated	DF	F calculated
Environment (E)	10	52.13**	10	42.84**
Block/E	44	0.69 ^{ns}	44	0.83 ^{ns}
Genotype (G)	6	25.43**	6	23.50**
E x G	60 (49) ⁽¹⁾	1.96**	60 (41) ⁽¹⁾	1.84**
Error	257 (209) ⁽¹⁾		257 (173) ⁽¹⁾	
R ²		0.76		0.73
CV (%)		24.3		26.58
Mean		2,543.6		1,773.4
Principal component	Auto values		Auto values	
	% explained	% accumulated	% explained	% accumulated
1	54.74	54.74	46.57	46.57
2	17.45	72.19	24.33	70.90
3	12.52	84.71	13.10	84.00
4	8.86	93.56	9.17	93.17
5	4.51	98.08	5.70	98.87
6	1.92	100.00	1.13	100.00

⁽¹⁾Corrected degree of freedom (DF), in parenthesis, for mean error and GE.

^{ns}Not significant. **Significant by the F test, at 1% probability.

Based on the pattern of the different groups found by the Toler & Burrows (1998) method, it was observed that LGoPE-06 (G6) showed convex and doubly desirable response (group A). However, LB308 (G2) had concave and doubly undesirable response, showing unsatisfactory yield and low stability in all trials.

According Toler (1990), genotypes clustered in group A were considered stable in favorable and unfavorable environments, and responsive to the improvement of management conditions. However,

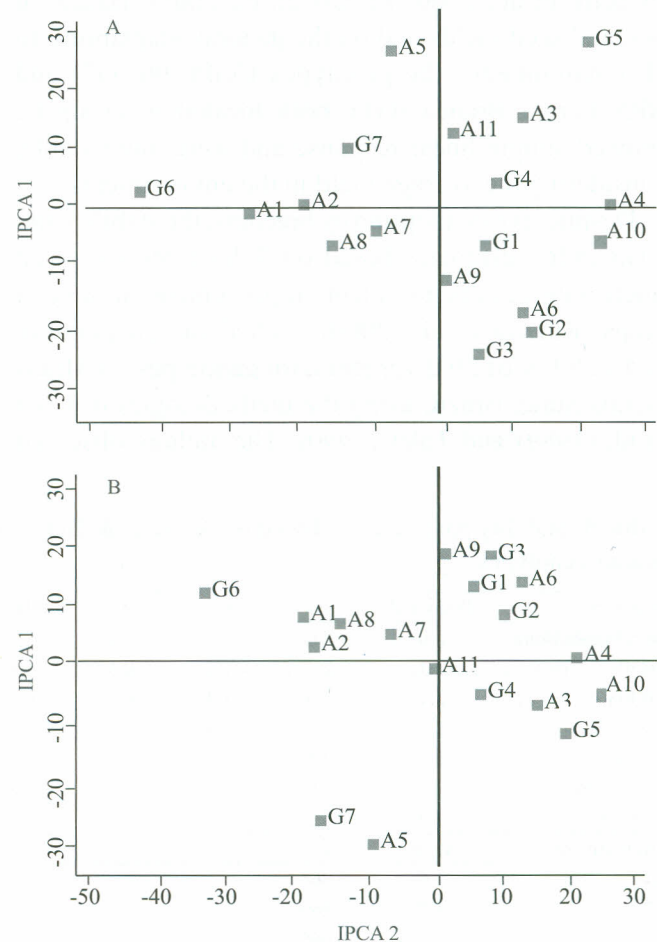


Figure 1. Genotype distribution in different environments regarding pod (A) and seed (B) yield, based on analysis by visual inspection of the AMMI2 biplot. G1, BRS Pérola Branca; G2, LB308; G3, LB408; G4, IAC Caiapó; G5, Florunner; G6, LGoPE-06; G7, LViPE-06. A1, Petrolina, PE, 2008; A2, Monteiro, PB, 2007; A3, Barbalha, CE, 2008; A4, Barbalha, CE, 2007; A5, Colinas, MA, 2009; A6, São João do Piauí, PI, 2009; A7, Abreu e Lima, PE, 2007; A8, Monteiro, PB, 2008; A9, Barbalha, CE, 2009 (trial 1); A10, Barbalha, 2009 (trial 2); A11, Coruripe, AL, 2009. IPCA, Interactive principal component axis

Rosse & Vencovsky (2000), when using the Toler model to estimate stability and adaptability in bean genotypes, found that genotypes with double favorable standard had low productivity and that those with double unfavorable standard had higher yields.

Most of the genotypes were situated in group C, with simple linear response, and did not deviate from the average response. This type of clustering often takes place when cultivars or advanced bred lines are evaluated. The yield of these materials is in accordance with the reported environmental average (Ribeiro et al., 2000; Morais et al., 2008). Based on pod and seed yields and on the general adaptability to all environments, the genotypes LViPE-06 (G7) and BRS Pérola Branca (G1), both located in group C, showed simple linear response and were more stable, considering the average yield in the environments.

Despite its low adoption by breeders, the stability and adaptability estimates based on Toler (1990) showed interesting results for plant improvement in several crops. Ribeiro et al. (2000) studied the adaptability and stability of 20 divergent corn genotypes, in Minas Gerais State, Brazil, using the methodologies of Cruz et al. (1989) and Toler (1990). The authors observed

that these models were very similar and efficient to predict the performance of genotypes. The correlation between both models was close to 97%.

Morais et al. (2008) evaluated 69 different soybean genotypes in several environments from the Central-West Region of Brazil and found different values for environmental quality. Considering group classifications based on yield and on response pattern, the authors were able to define genotypes with double desirable response pattern (group A) or with response pattern of mono-segmented genotypes (B, C, and D), with variable yield. This methodology was advantageous to estimate soybean adaptability and stability, since it discriminated genotypes with narrow genetics based on groups with different response pattern.

The results obtained in the present work using the Toler (1990) and AMMI methodologies were consistent with trimmings of genotypes during the improvement processes, especially for BRS Pérola Branca and LViPE-06.

BRS Pérola Branca, a recent early-runner peanut cultivar developed by Embrapa, was obtained via crossing between BR 1 (upright and short cycle,

Table 5. Stability parameters⁽¹⁾, based on the Toler & Burrows (1998) methodology, regarding pod and seed yields in runner peanut genotypes.

Genotype	Pod yield	Ai	Bli	B2i	Bi	Group	Seed yield	Ai	Bli	B2i	Bi	Group
BRS Pérola Branca	2.6	2.6	0.9	1.2	1.17	C	1.8	1.18	1.0	0.9	1.0	C
LB308	2.1	2.1	1.3	0.5*	0.8	E	1.4	1.6	1.3	0.5	0.9	C
LB408	2.1	2.3	1.4	0.7	0.9	C	1.5	1.6	1.4	0.6	1.0	C
IAC Caiapó	2.2	2.3	0.8	1.3	1.1	C	1.6	1.6	0.8	0.8	0.8	C
Florunner	2.5	2.5	1.2	1.2	1.2	C	1.7	1.7	1.2	1.3	1.2*	B
LGoPE 06	3.0	2.6	0.3*	1.3	0.8	A	2.0	1.8	0.4*	1.3	0.9	A
LViPE 06	3.2	3.2	1.1	0.9	1.0	C	2.3	2.2	0.9	1.5	1.2	C
Environment ⁽²⁾	Mean	Ui					Mean	Ui				
A1	1.6	-965					1.1	-682				
A2	1.7	-869					1.2	-608				
A3	1.8	-665					1.3	-461				
A4	2.9	345					2.0	231				
A5	3.3	784					2.4	712				
A6	1.7	-751					1.2	-513				
A7	2.5	-134					1.7	-92				
A8	2.7	202					1.9	177				
A9	3.0	467					2.0	253				
A10	2.7	123					1.9	96				
A11	4.0	1,463					2.7	887				

⁽¹⁾Ai, performance of the ith genotype (1, 2, ..., 7) in an environment of average yield; Bli and B2i, sensibility to the performance of the ith genotype in favorable and unfavorable environments, respectively; Ui, effect of the ith environment; Bi, the only parameter of the model when Bli and B2i are both not significant.

⁽²⁾A1, Petrolina, PE, 2008; A2, Monteiro, PB, 2007; A3, Barbalha, CE, 2008; A4, Barbalha, CE, 2007; A5, Colinas, MA, 2009; A6, São João do Piauí, PI, 2009; A7, Abreu e Lima, PE, 2007; A8, Monteiro, PB, 2008; A9, Barbalha, CE, 2009 (trial 1); A10, Barbalha, 2009 (trial 2); A11, Coruripe, AL, 2009.

Valência type) and the top line LViPE-06 (late cycle, Virginia type). The general breeding proceedings for the fitting of this cultivar were performed in Barbalha and Missão Velha, both located in the semiarid region of Ceará State, Brazil. In several field trials carried out in the rainy season, 'BRS Pérola Branca' (previously named Branco Rasteiro 1/08) showed moderate tolerance to drought (Santos et al., 2012). In a study carried out by Pereira (2010), involving osmotic adjustment, gas exchanges, and agronomic traits in peanut genotypes subjected to 20 days of water stress, BRS Pérola Branca (named LBR Branco) showed reasonable ability to prevent water losses. This trait, added to earliness, was genetically inherited from BR 1, an early drought tolerant cultivar developed by Embrapa and recommended for semiarid environments (Gomes et al., 2007; Luz et al., 2010; Graciano et al., 2011). From LViPE-06, 'BRS Pérola Branca' inherited its high tolerance to leaf diseases and high capacity for oil and pod production (Santos et al., 2010). The choice of LViPE-06 as a progenitor was due to its large genetic variability revealed by molecular markers in runner genotypes (Pereira et al., 2008). Both LViPE-06 and BRS Pérola Branca genotypes showed high oil content (51–52%), with oleic/linoleic ratio >1.6 (Santos et al., 2012), representing an excellent material for edible or fuel oil markets.

Conclusions

1. The peanut genotypes LViPE-06 and BRS Pérola Branca are more stable and adapted to the semiarid environment.

2. The peanut genotype LGoPE-06 is a promising material for pod production, but is highly dependent on favorable environments.

Acknowledgements

To Banco do Nordeste do Brasil, for financial support.

References

- CRUZ, C.D.; TORRES, R.A. de A.; VENCOVSKY, R. An alternative approach to the stability analysis proposed by Silva and Barreto. **Revista Brasileira de Genética**, v.12, p.567-580, 1989.
- DIAS, C.T. dos S. **Métodos para escolha de componentes em modelo de efeito principal aditivo e interação multiplicativa (AMMI)**. 2005. 73p. Tese (Livro Docência) – Escola Superior de Agricultura Luiz de Queiroz, Piracicaba.
- EBERHART, S.A.; RUSSELL, W.A. Stability parameters for comparing varieties. **Crop Science**, v.6, p.36-40, 1966.
- FERREIRA, D.F.; DEMÉTRIO, C.G.B.; MANLY, B.F.J.; MACHADO, A. de A.; VENKOVSKY, R. Statistical models in agriculture: biometrical methods for evaluating phenotypic stability in plant breeding. **Cerne**, v.12, p. 373-388, 2006.
- GOMES, L. de R.; SANTOS, R.C. dos; ANUNCIACÃO FILHO, C.J. da; MELO FILHO, P. de A. Adaptabilidade e estabilidade fenotípica de genótipos de amendoim de porte ereto. **Pesquisa Agropecuária Brasileira**, v.42, p.985-989, 2007.
- GRACIANO, E.S.A.; NOGUEIRA, R.J.M.C.; LIMA, D.R.M.; PACHECO, C.M.; SANTOS, R.C. Crescimento e capacidade fotossintética da cultivar de amendoim BR 1 sob condições de salinidade. **Revista Brasileira de Engenharia Agrícola e Ambiental**, v.15, p.794-800, 2011.
- LUZ, L.N. da; SANTOS, R.C. dos; SILVA FILHO, J.L. da; MELO FILHO, P. de A. Estimativas de parâmetros genéticos em linhagens de amendoim baseadas em descritores associados ao ginóforo. **Revista Ciência Agronômica**, v.41, p.132-138, 2010.
- MORAIS, L.K. de; MOURA, M.F.; VENCOVSKY, R.; PINHEIRO, J.B. Adaptabilidade e estabilidade fenotípica em soja avaliada pelo método de Toler. **Bragantia**, v.67, p.275-284, 2008.
- MURAKAMI, D.M.; CARDOSO, A.A.; CRUZ, C.D.; BIZÃO, N. Considerações sobre duas metodologias de análise de estabilidade e adaptabilidade. **Ciência Rural**, v.34, p.71-78, 2004.
- OLIVEIRA, E.J. de.; GODOY, I.J. de. Pod yield stability analysis of runner peanut lines using AMMI. **Crop Breeding and Applied Biotechnology**, v.6, p.311-317, 2006.
- PACHECO, R.M.; DUARTE, J.B.; VENCOVSKY, R.; PINHEIRO, J.B.; OLIVEIRA, A.B. Use of supplementary genotypes in AMMI analysis. **Theoretical and Applied Genetics**, v.110, p.812-818, 2005.
- PEREIRA, J.W. de L. **Respostas fisiológica e agrônômica de genótipos de amendoim sob condição de estresse hídrico**. 2010. 64p. Dissertação (Mestrado) – Universidade Federal Rural de Pernambuco, Recife.
- PEREIRA, J.W. de L.; MELO FILHO, P. de A.; SILVA, F.A.C.; SANTOS, R.C. dos. Variabilidade genética de acessos de amendoim do tipo *runner* com base em marcadores RAPD. **Revista Brasileira de Oleaginosas e Fibrosas**, v.12, p.35-40, 2008.
- PIMENTEL-GOMES, F. **Curso de estatística experimental**. 13.ed. Piracicaba: Nobel, 1990. 468p.
- RIBEIRO, P.H.E.; RAMALHO, M.A.P.; FERREIRA, D.F. Adaptabilidade e estabilidade de genótipos de milho em diferentes condições ambientais. **Pesquisa Agropecuária Brasileira**, v.35, p.2213-2222, 2000.
- ROSSE, L.N.; VENCOVSKY, R. Modelo de regressão não-linear aplicado ao estudo da estabilidade fenotípica de genótipos de feijão no Estado do Paraná. **Bragantia**, v.59, p.99-107, 2000.
- SANTOS, R.C. dos; FREIRE, R.M.M.; LIMA, L.M. de; ZAGONEL, G.F.; COSTA, B.J. Produtividade de grãos e óleo de genótipos de amendoim para o mercado oleoquímico. **Revista Ciência Agronômica**, v.43, p.72-77, 2012.

SANTOS, R.C. dos; REGO, G.M.; SANTOS, C.A.F.; MELO FILHO, P.A.; SILVA, A.P.G.; GONDIM, T.M.S.; SUASSUNA, T.F. **Recomendações técnicas para o cultivo do amendoim em pequenas propriedades agrícolas do Nordeste brasileiro**. Campina Grande: Embrapa Algodão, 2006. 7p. (Embrapa Algodão. Circular técnica, 102).

SANTOS, R.C. dos; REGO, G.M.; SILVA, A.P.G. da; VASCONCELOS, J.O.L.; COUTINHO, J.L.B.; MELO FILHO, P. de A. Produtividade de linhagens avançadas de amendoim em condições de sequeiro no Nordeste brasileiro. **Revista Brasileira de Engenharia Agrícola e Ambiental**, v.14, p.589-593, 2010.

SAS INSTITUTE. **SAS/STAT user's guide**. Version 8. Cary: SAS Institute, 1999. 3811p.

SILVA FILHO, J.L. da; MORELLO, C. de L.; FARIAS, F.J.C.; LAMAS, F.M.; PEDROSA, M.B.; RIBEIRO, J.L. Comparação de métodos para avaliar a adaptabilidade e estabilidade produtiva em algodoeiro. **Pesquisa Agropecuária Brasileira**, v.43, p.349-355, 2008.

TOLER J.E. **Patterns of genotypic performance over environmental arrays**. 1990. 154p. Thesis (Ph.D) – Clemson University, Clemson.

TOLER, J.E.; BURROWS, P.M. Genotypic performance over environmental arrays: a non-linear grouping protocol. **Journal of Applied Statistics**, v.25, p.131-143, 1998.

ZOBEL, R.W.; WRIGHT, M.J.; GAUCH, H.G. Statistical analysis of a yield trial. **Agronomy Journal**, v.80, p.388-393, 1988.

Received on July 29, 2011 and accepted on July 23, 2012