



CORE

Simulating more realistic predation threat using attack playbacks

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ABSTRACT

Use of virtual proxies of live animals are rapidly gaining ground in studies of animal behaviour. Such proxies help to reduce the number of live experimental animals needed to stimulate the behaviour of experimental individuals and to increase standardisation. However, using too simplistic proxies may fail to induce a desired effect and/or lead to quick habituation. For instance, in a predation context, prey often employ multimodal cues to detect predators or use specific aspects of predator behaviour to assess threat. In a live interaction, predator and prey often show behaviours directed towards each other, which are absent in virtual proxies. Here we compared the effectiveness of chemical and visual predator cues in the cooperatively breeding cichlid Neolamprologus pulcher, a species in which predation pressure has been the evolutionary driver of its sociality. We created playbacks of predators simulating an attack and tested their effectiveness in comparison to a playback showing regular activity and to a live predator. We further compared the effectiveness of predator odour and conspecific skin extracts on behaviours directed towards a predator playback. Regular playbacks of calmly swimming predators were less effective than live predators in stimulating a focal individual's aggression and attention. However, playbacks mimicking an attacking predator induced responses much like a live predator. Chemical cues did not affect predator directed behaviour.

Subjects Animal Behavior

Keywords Virtual predator threat, Playbacks, Prey-directed behaviour, Threat simulation, Cichlid

INTRODUCTION

Predation risk is thought to play an important role in determining life-history trajectories, the social system and behaviours among animals (Holling, 1965; Tollrian, 1995; Stearns, 2000; Heg et al., 2004; Preisser & Bolnick, 2008; Walsh & Reznick, 2008; Groenewoud et al., 2016; Fischer et al., 2017a). While an obvious way of testing the impacts of predation risk is to manipulate the number of predators and/or to present predators to potential prey (e.g., Chiverton, 1986; Fontaine & Martin, 2006), such experiments are not always logistically or ethically feasible. An alternative approach is to use 'virtual proxies' on captive prey, which reduces the ethical implications of subjecting such prey to live predators and removes the logistical and ethical issues of manipulating predators. Many proxies such as decoys (Arroyo, Mougeot & Bretagnolle, 2001; Li et al., 2014; Catano et al., 2016; Griesser & Suzuki, 2017; Morales, Lucas & Velando, 2018), screen presentations (Johnson & Basolo, 2003;

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Fischer et al., 2014; Scherer, Godin & Schuett, 2017; Watve & Taborsky, 2019) or kairomones and alarm cues (reviewed in Ferrari, Wisenden & Chivers, 2010) have been used successfully so far. Nevertheless, the utility of virtual proxies rests with their ability to replicate the natural predator-prey encounter. Prey may use multiple cues to detect a predator (Gonzálvez & Rodríguez-Gironés, 2013) or multiple cues may have a different effect than a single cue (Hartman & Abrahams, 2000; Hickman, Stone & Mathis, 2004; Martin et al., 2010; Ward & Mehner, 2010). Further, prey may use specific features of the predator's behaviour or orientation to assess the immediate risk and fine-tune their behaviour (Kent et al., 2019). Most importantly, live predators and prey will naturally interact, an element inherently lacking in the use of proxies. Despite this, such interactions are rarely incorporated into experiments using virtual proxies (but see *Hartman & Abrahams*, 2000)—increasing the chances that prey will quickly habituate or even ignore the stimulus (Holomuzki & Hatchett, 1994; Raderschall, Magrath & Hemmi, 2011). To minimize these shortcomings, it is necessary to ensure that virtual proxies (i) stimulate the relevant sensory modalities and (ii) provide salient features of natural predators eliciting ecologically relevant prey responses.

In aquatic systems, the role of visual as well as chemical cues in predator detection and response is well studied. Chemical cues released from damaged tissues of injured conspecifics ('alarm cues') play an important role in detection of predators in many species and are used in experimental manipulation (e.g., Brown & Smith, 1998; Hartman & Abrahams, 2000; Brown, 2003; McCormick & Manassa, 2008; Ferrari, Wisenden & Chivers, 2010; Manassa & McCormick, 2012; Moscicki & Hurd, 2017; Lucon-Xiccato, 2019). For example, skin extract from conspecifics administered along with predator odour helps fishery-reared rainbow trouts to recognise predators (Brown & Smith, 1998). Similarly, the use of visual proxies in aquatic systems is also well-established (e.g., Johnson & Basolo, 2003; Gerlai, 2013; Fischer et al., 2014; O'Connor et al., 2015; Dieng et al., 2017). While most studies report a successful use of proxies in inducing predicted anti-predator response, none have compared their effectiveness to live predator presentations.

Neolamprologus pulcher is a cooperatively breeding cichlid from Lake Tanganyika that lives in groups of up to 25 individuals (*Taborsky*, 2016). The social structure of *N. pulcher* is driven primarily by predation pressure (*Groenewoud et al.*, 2016) and thus, manipulation of predation threat has been often employed to study their social dynamics (e.g., *Heg & Taborsky*, 2010; *Fischer et al.*, 2017a; *Watve & Taborsky*, 2019). Turbidity and water quality in the lake differs between populations (*Plisnier*, 2002) and between seasons (*Plisnier et al.*, 1999). Therefore, it is expected that *N. pulcher* would rely on both visual and chemical cues flexibly. *Lepidiolamprologus elongatus* is a common and ferocious predator of *N. pulcher* that is most often used in manipulation experiments in field as well as laboratory. It has been shown that *N. pulcher* can successfully distinguish images of *L. elongatus* from herbivorous cichlids of the same size and respond accordingly (*Fischer et al.*, 2014). Chemical cues from *L. elongatus* alone can also induce appropriate anti-predator responses in very young fry of *N. pulcher* (*Fischer et al.*, 2017a). Predator recognition in N. pulcher seems to be innate, as both these studies observed anti-predator response by naïve individuals. One study showed an unexpected reduction in response towards visual predator cues in the presence

of conspecific skin extract on the anti-predator response of *N. pulcher* (*O'Connor et al.*, 2015). It is, however, unknown if and how a combination of both predator odour and conspecific skin extract influences anti-predator response of *N. pulcher*.

Here we manipulated visual proxies to include prey-directed aggression by the predator *L. elongatus* to simulate a live interaction. We further tested the effect of predator odour and conspecific skin extract independently and in combination with each other on anti-predator responses of *N. pulcher*. We expected playbacks simulating directed attacks to induce responses more similar to a live predator but for lateral playbacks of normally swimming predators to induce lower responses than a live predator. We also predicted that *N. pulcher* will show a heightened response towards the predator visuals when predator odour was administered in conjunction with conspecific skin extract.

METHODS

Experimental animals and housing conditions

The experiments were conducted at the Ethologische Station Hasli, University of Bern, Switzerland, under license BE-74/15 of Veterinary office of Kanton Bern. All experimental animals were obtained from the University of Bern laboratory stock, maintained in a 13:11 h light-dark cycle at 26 °C \pm 1 °C mimicking the conditions of Lake Tanganyika. All focal N. pulcher were predator-naïve adults (size range 40-45 mm) reared in the laboratory for 5 generations. All predators were first generation offspring of wild caught fish and were in the size range of 80–140 mm. The experimental 20-litre tanks ($40 \text{ cm} \times 25 \text{ cm} \times 20 \text{ cm}$) were lined with 2 cm layer of sand and aerated by air stones. They were visually divided in three zones marked by pen on the front screen of each tank. The zone closest to the stimulus presentation was designated as 'zone of interaction', where the focal individual actively engaged in inspection of and defence against the stimulus. The farthest zone was considered as 'safe zone' and the time spent in this zone was taken as a measure of avoidance (Fig. 1). Each focal was tested in a separate experimental tank and was allowed to acclimatise overnight before testing. During acclimatisation, focal individuals were provided with ceramic pot halves as shelters, which were removed half an hour before each behavioural recording to avoid obstruction of observations. Visual playbacks were displayed on a 25 cm × 15 cm LED tablet screen ('HP ElitePad 900', resolution: 59 dot/cm) in both the experiments.

Experiment 1: Regular playbacks vs playbacks of attacks

Individual *L. elongatus* (N=2) were housed in individual 20 L tanks overnight and then filmed swimming calmly for 5 min (hereafter 'lateral playbacks'). The same individuals were then filmed through a one-way mirror, as they attacked their own reflection (hereafter 'frontal playbacks'). The mirror and camera were placed along the breadth of the filming tank, elevated to align with the level of sand (2 cm from bottom). The lens of the camera was higher by a further 3–4 cm. During presentations, the tablet was again slightly elevated, such that the apparent level of sand on screen matched the level of sand in the experimental tank, such that the visuals on screen appeared similar to a real adjacent tank of same dimensions. Frontal playbacks also lasted 5 min and were edited in Adobe Premier Pro such that the

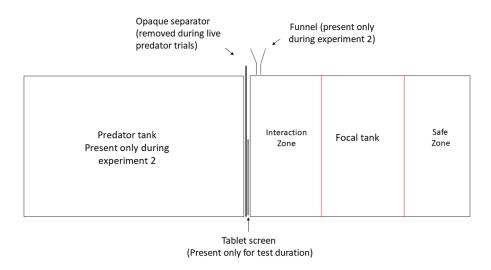


Figure 1 Experimental set up. Experimental manipulation of visual and olfactory cues. Predator tank was present only during Experiment 1 and was separated from the focal tank with an opaque partition at all times except during live-predator trials. Olfactory cues were administered close to the presentation through a funnel during Experiment 2.

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gap between any two attacks did not exceed 20 s. While it is theoretically possible that the responses towards a real prey and a conspecific mirror image differ in frequency and duration of aggressive displays (such as operculum spread, fin spread), frontal playback videos were edited to almost exclusively showed overt attacks (fast approach towards the target followed by biting), which at least for the human observer are indistinguishable from a targeted attack at prey. The filming tanks with predators were placed between two experimental tanks such that the same individual L. elongatus also acted as live stimulus. The predator and experimental tanks were separated by an opaque partition which was removed only during live presentation trials. A 5-minute long visual of an unoccupied filming tank was presented as blank control. To ensure that the focal fish were not simply responding to movement, an off-white square (approximately matching the colour of L. elongatus) of 5 cm × 5 cm was added on the background of the blank control and animated in PowerPoint to move randomly along the screen (hereafter 'moving square'). Each focal N. pulcher (N = 16) was exposed each to five presentations in a randomised order, with a gap of at least 3 h in between: (i) empty tank, (ii) moving square, (iii) calmly swimming predator, (iv) attacking predator, (v) live predator. The response of the focal towards the presentation was recorded for 5 min (see below).

Experiment 2: Chemical cues

In this experiment we tested the response of *N. pulcher* to chemical cues from *L. elongatus*, skin extract from a conspecific and a combination of the two, compared to a control cue of water collected from the focal's own tank. To obtain predator odour, individual *L. elongatus* (N = 3) were housed in individual 60 L tanks (60 cm \times 35 cm \times 30 cm) for 48 h, fed once a day with standard cichlid flake food. They were not fed for another 24 h after which 500 mL water was collected from the tank and stored in -20C. Conspecific skin extract

was prepared as per Brown & Smith (1998). Eleven N. pulcher in the size-range of 3.5–4.5 cm were sacrificed by stunning followed by severing the spinal cord. The skin was then separated, rinsed in distilled water to remove traces of other tissues and homogenised with equal volume distilled water (1 mL/1 cm²) to release any possible alarm cues in skin cells. The homogenised mixture was filtered to remove debris and diluted 16 times. Aliquots of 12 mL were stored at -20 °C and diluted 10 times with water from an unoccupied experimental tank before use. For combining the two cues, 12 mL of skin extract was diluted with water from predator holding tanks instead. The final volume of all cues was maintained at 120 mL (0.6% tank volume). Focal N. pulcher (N = 18) were exposed to all four cues in randomised order with at least 3 h in between. This time window is sufficient for the cues to dissipate and the focal individuals to return to normal behaviour (Watve & Taborsky, 2019). All chemical cues were combined with visuals of a predator (from 9 different L. elongatus individuals) swimming calmly in a 20-L tank to facilitate 'predator directed' behaviour by the focal N. pulcher. Chemical cues were administered through a funnel at the beginning of the playback (see Fig. 1 for set-up). Responses of the focal fish were recorded for 5 min (see 'Behavioural recordings').

Behavioural recordings

In both the experiments, the number of aggressive displays (head-down display, fin spread, operculum spread and fast approach towards the stimulus without attempting contact; see *Reyes-Contreras et al.*, 2019 for detailed ethogram) shown by the focal towards the stimulus display were counted as a measure of defence. If the focal individual was facing in straight line towards the screen while ceasing other activities (e.g., sand digging), it was considered as 'oriented towards the screen'. The amount of time a focal spent orienting towards the screen was used as a measure of attentiveness. Engagement with the stimulus was measured as the time spent in the zone closest to the predator presentation. Finally, avoidance or fear was measured as the amount of time spent in the safe zone (Fig. 1). Freezing, a common fear response of *N. pulcher*, was extremely rare and was therefore not included in the analysis. All observations were video recorded and later coded in Solomon Coder (András Petér, University of Budapest) with observer (SP) being blind to the type of cue.

Statistical analysis

All analyses were conducted in R 3.5 (*R Core Team, 2017*). The frequencies of aggressive displays were analysed using generalised mixed models (GLMMs) with negative binomial distribution in package 'glmmTMB' (*Brooks et al., 2017*). The times spent being attentive to the screen and in safe zone were analysed using linear mixed models (LMMs) in package 'lme4' (*Bates et al., 2015*). Normality assumptions were tested by Shapiro–Wilk normality tests and by visually examining Quantile-Quantile (Q-Q) plots as well as histograms of residuals and plots of residuals against fitted values. Log and root transformations were employed to achieve normality when the model residuals violated normality assumptions. All models included cue as the main effect (five levels for experiment 1, four levels for experiment 2) and focal identity as a random effect. As there were only two predator individuals in experiment 1, they were included as a factor in initial models but had no

effect in any of the models. Removing predator identity decreased the AIC by at least 2 in all models and therefore, the final models only included cue as the main factor. The identity of the predator in visual displays (N=9 different individuals) shown together with the chemical cues was included as a random effect in all models for experiment 2. In case of significant cue effects, pairwise comparisons between levels of the cue were carried out in package 'emmeans' (*Lenth*, 2018). *P*-values were adjusted for multiple testing using multivariate t distribution (mvt) and only adjusted values are reported here.

RESULTS

Experiment 1

The number of aggressive displays towards the stimulus was determined by the type of presentation (GLMM, $\chi^2 = 23.41$, p = 0.0001; Fig. 2A). Pairwise comparisons between presentation types revealed that there was no difference between the frontal playbacks and live stimulus (p = 0.13). However, lateral playbacks induced much lower aggression than live stimuli (p = 0.0008). Further, there was also a significant difference between the control stimulus and frontal playbacks (p = 0.02), which was not the case for lateral playbacks (p = 0.73 and the moving square (p = 0.98; Table 1, Fig. 2). Also the time spent being attentive to the stimulus was affected by the type of presentation (LMM, F = 4.14, p = 0.004; Fig. 2B). Focals were more attentive towards the live stimulus as compared to frontal playbacks (p = 0.02), the moving square (p = 0.004) and the control stimulus (p = 0.02), but not compared to lateral playbacks (p = 0.10; Table 2). No difference was found between lateral and frontal playbacks (p = 0.98; Table 2). Further, the time spent in the interaction zone was also influenced by presentation type (LMM power transformed, F = 5.34, p = 0.0009; Fig. 2C). Fish spent much longer in the zone of interaction when encountering a live predator compared to a blank control (p = 0.0007) or a moving square (p = 0.01) and marginally longer when compared to a lateral display (p = 0.09). Attention time did not differ between frontal playbacks and live predator (p = 0.46) or between lateral and frontal playbacks (p = 0.90; Table 3). Time in the safe zone was not affected by the type of display (LMM log-transformed, F = 1.29, p = 0.28; Fig. 2D).

Experiment 2

There was no effect of any chemical cue on the number of aggressive displays directed towards the screen (GLMM, $\chi^2 = 2.08$, p = 0.55; Fig. 3A), on the time spent attentive to the screen (LMM, F = 1.15, p = 0.33; Fig. 3B), or on the time spent in the safe zone (LMM, F = 1.44, p = 0.24; Fig. 3C) or in the interaction zone (LMM, F = 0.36, p = 0.77; Fig. 3D).

DISCUSSION

Here we tested the influence of different chemical and visual predator cues on the response of *N. pulcher*. The type of visual playbacks effected only those variables related to an interaction between the focal fish and screen, but it did not affect avoidance behaviour. Focals spent similar time in the interaction zone and showed similar aggression towards live stimuli and frontal playbacks, however, the response towards lateral playbacks was

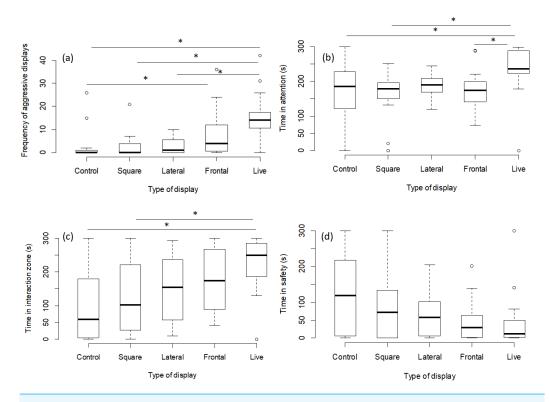


Figure 2 Behavioural observations for experiment 1. Effects of playback type on predator-directed behaviour (N=80 observations of 16 fish). The graphs show the effects of lateral vs. frontal (attacking) predator playbacks in comparison to a live predator, a blank control and a moving square. Significant differences are indicated by asterisks.

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Table 1 Aggressive displays towards different types of predator presentations. Pairwise comparisons between aggressive displays shown by focal N. pulcher (N=16) towards different types visual predator presentations. All p-values presented are corrected for multiple comparisons using 'mvt' method. Significant p-values are marked with asterisks.

Display	Estimate	SE	df	t	p
Control-Square	-0.26	0.55	73	-0.47	0.98
Control-Lateral	-0.70	0.57	73	-1.22	0.73
Control-Frontal	-1.77	0.57	73	-3.08	0.02*
Control-Live	-2.98	0.62	73	-4.76	0.0001*
Square-Lateral	-0.43	0.56	73	-0.77	0.93
Square-Frontal	-1.50	0.56	73	-2.67	0.06.
Square-Live	-2.71	0.59	73	-4.56	0.0002*
Lateral-Frontal	-1.06	0.52	73	-2.05	0.25
Lateral-Live	-2.28	0.54	73	-4.17	0.0008*
Frontal-Live	-1.21	0.51	73	-2.35	0.13

Table 2 Attention towards different types of predator presentations. Pairwise comparisons between time spent being attentive to different visual proxies by focal N. pulcher (N = 16). All p-values presented are corrected for multiple comparisons using 'mvt' method. Significant p-values are marked with asterisks.

Display	Estimate	SE	df	t	P
Control-Square	62.8	122	60	0.51	0.98
Control-Lateral	-80.9	122	60	-0.66	0.96
Control-Frontal	-13.9	122	60	-0.11	1.00
Control-Live	-384.4	122	60	-3.14	0.02*
Square-Lateral	-143.7	122	60	-1.17	0.76
Square-Frontal	-76.7	122	60	-0.62	0.96
Square-Live	-447.2	122	60	-3.66	0.004*
Lateral-Frontal	66.9	122	60	0.54	0.98
Lateral-Live	-303.5	122	60	-2.48	0.10
Frontal-Live	-370.5	122	60	-3.03	0.02*

Table 3 Time in interaction zone during different predator presentations. Pairwise comparisons between time spent in interaction zone by focal N. pulcher (N=16) during different visual proxies. All p-values presented are corrected for multiple comparisons using 'mvt' method. Significant p-values are marked with asterisks.

Display	Estimate	SE	df	t	P
Control-Square	-26.1	30.7	60	-0.85	0.91
Control-Lateral	-52.1	30.7	60	-1.70	0.44
Control-Frontal	-79.2	30.7	60	-2.58	0.08
Control-Live	-130.0	30.7	60	-4.24	0.0007*
Square-Lateral	-26.1	30.7	60	-0.85	0.91
Square-Frontal	-53.1	30.7	60	-1.73	0.42
Square-Live	-103.9	30.7	60	-3.38	0.01*
Lateral-Frontal	-27.1	30.7	60	-0.88	0.90
Lateral-Live	-77.9	30.7	60	-2.53	0.09
Frontal-Live	-50.8	30.7	60	-1.65	0.46

relatively weaker. Surprisingly, focals payed less attention towards the frontal playbacks than the live stimulus, whereas attention between the lateral playback and the live stimulus did not differ. The chemical cues did not affect the anti-predator behaviour of focals.

N. pulcher are sensitive to visual predator cues. For instance, they can distinguish heterospecifics, based simply on moving images (*Fischer et al.*, 2014). However, images can fail to simulate a more realistic threat. Here we show that a playback of predator attack can induce a response similar to that shown towards a live predator. While prey species may recognize a predator based on images, they may not necessarily perceive them as threatening (*Belin et al.*, 2018). Even for a moving visual, prey may only perceive the display as a threat if they include certain angles (*Kent et al.*, 2019), postures (*Etting & Isbell*, 2014) or behaviours (*Su & Lim*, 2017) of the predator. Video playbacks have been used as predator cues in the past (*Rowland*, 1999; *Carlile*, *Peters & Evans*, 2006; *Butler*, *Magrath & Peters*, 2017). However, they lacked the element of prey-directed behaviour of the predator.

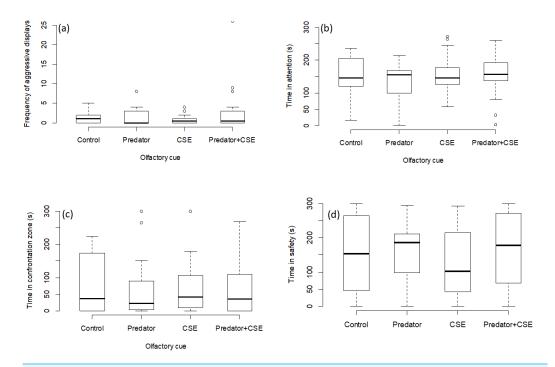


Figure 3 Results for experiment 2. Manipulation of olfactory cues (N = 72 observations of 18 focal fish). The graphs show the effects of predator odour and conspecific skin extract (CSE) on the aggression, time spent being attentive to predator display and the times spent engaging with or avoiding the presentation. Significant differences are marked with asterisks.

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We attempted to design visual predator cues that are as similar to an actual encounter as possible. While robotic models have been shown to hold potential (*Krause, Winfield & Deneubourg, 2011; Ladu et al., 2015; Landgraf et al., 2016; Porfiri et al., 2019*), they may not always be easily accessible. We propose a technically simple method of simulating a predator attack, which can be produced with ease and in a short time. Like other video playback, also frontal playbacks of attacking predators may eventually lead to habituations though. However, this can effectively be prevented by running them in an unpredictable temporal scheme (M Watve, 2019, unpublished data).

Even though the focal fish interacted with live stimuli and frontal playbacks, the apparent attacks from the predators in either case did not elicit a fear response in the fish. One possible explanation is that the predators never actually managed to harm the focals even during a live presentation, which may have emboldened the focals.

O'Connor et al. (2015) previously found conspecific skin extract to reduce the response towards a visual predator cue in *N. pulcher*, which is opposite to the prediction that skin extracts represent and alarm cue and should increase anti-predator responses (see *Chivers & Smith*, 1998 for comprehensive review). However, we did not find any such difference, neither when presenting skin extract alone or when paired with predator smell. Skin extract is assumed to contain alarm cues indicative of an injured conspecific, as shown in many other fish species including three-spine sticklebacks, *Gasterosteus aculeatus* (*Brown & Godin*, 1997) Rainbow trouts, *Onchoryhnchus mykiss* (*Brown & Smith*, 1997; *Brown*

& Smith, 1998), red swordtails Xiphophorus helleri (Mirza, Scott & Chivers, 2001) and rainbow darters, Etheostoma caeruleum (Commens & Mathis, 1999), although this is not always the case. A recent study showed that muscle tissue extracts induce antipredator responses more effectively than skin or other tissue extracts (Meuthen et al., 2018) in another African cichlid P. taeniatus. It is highly likely that skin extract does not contain alarm substances in N. pulcher either. Further investigation is required to determine whether and to what extent alarm cues are present in other tissues of N. pulcher. Fischer et al. (2017b) showed that both chemical and visual predator cues are used independently by N. pulcher in predator detection, but they did not administer them together. We administered chemical cues in combination with visual cues and our results suggest that N. pulcher ignore cues from predator odour or conspecific skin extract in the presence of visual cues. Similar results were seen in juvenile roach, Rutilus rutilus, which responded differently towards visual and chemical predator cues, but when presented with both cues together, the response was similar to visual cues alone (Martin et al., 2010).

Artificial proxies can help adhere to the 3Rs (Reduce, Refine, Replace) of ethical framework (*Russell & Burch*, 1959). While this is also an excellent way to increase standardization, it is important to make sure that the employed proxies indeed have the desired effect. For example, a species or population living under consistently high predation risk may ignore predator cues unless immediate threat is sensed (*Creel et al.*, 2008; *Bell et al.*, 2009). In such a case, it is important use a proxy that can simulate an immediate threat to induce measurable response in the prey.

CONCLUSIONS

Here we show that *N. pulcher* do not use chemical cues from conspecific skin tissue as a indication of predation threat when combined with visual predator cues. The failure to respond to chemical cues may have been caused by precedence of visual cues over chemical cues when both cues in both modalities are presented together. We further show that including directed attacks by predator in a visual proxy induces responses similar to a live predator encounter. Most of previous work has focused on non-interactive predator visuals. We show that it is important to include prey-directed behaviour to simulate a threat level similar to that posed by a live predator.

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ADDITIONAL INFORMATION AND DECLARATIONS

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Mukta Watve conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, approved the final draft.
- Sebastian Prati performed the experiments, authored or reviewed drafts of the paper, approved the final draft.
- Barbara Taborsky conceived and designed the experiments, analyzed the data, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper, approved the final draft.

Animal Ethics

The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):

This study was conducted at the Ethologische Center Hasli, University of Bern, Switzerland under license BE-74/15, approved by the Veterinary Office of Kanton Bern, Switzerland.

Data Availability

The following information was supplied regarding data availability: Raw data is available in the Supplemental File.

Supplemental Information

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.8149#supplemental-information.

REFERENCES

Arroyo B, Mougeot F, Bretagnolle V. 2001. Colonial breeding and nest defence in Montagu's harrier (*Circus pygargus*). *Behavioral Ecology and Sociobiology* **50**:109–115 DOI 10.1007/s002650100342.

- **Bates D, Machler M, Bolker BM, Walker SC. 2015.** Fitting linear mixed-effects models using lme4. *Journal of Statistical Software* **67**:1–48.
- Belin L, Formanek L, Heyraud C, Hausberger M, Henry L. 2018. Influence of early experience on processing 2D threatening pictures by European starlings (Sturnus vulgaris). *Animal Cognition* 21(6):749–758 DOI 10.1007/s10071-018-1207-x.
- Bell MBV, Radford AN, Rose R, Wade HM, Ridley AR. 2009. The value of constant surveillance in a risky environment. *Proceedings of the Royal Society B-Biological Sciences* 276:2997–3005 DOI 10.1098/rspb.2009.0276.
- Brooks ME, Kristensen K, Benthem KJvan, Magnusson A, Berg CW, Nielsen A, Skaug HJ, Maechler M, Bolker BM. 2017. glmmTMB balances speed and flexibility among packages for zero-inflated generalized linear mixed modeling. *The R Journal* 9:378–400 DOI 10.32614/RJ-2017-066.
- **Brown GE. 2003.** Learning about danger: chemical alarm cues and local risk assessment in prey fishes. *Fish and Fisheries* **4**:227–234 DOI 10.1046/j.1467-2979.2003.00132.
- **Brown GE, Godin J-GJ. 1997.** Anti-predator responses to conspecific and heterospecific skin extracts by threespine sticklebacks: alarm pheromones revisited. *Behaviour* **134**:1123–1134 DOI 10.1163/156853997X00098.
- **Brown GE, Smith RJF. 1997.** Conspecific skin extracts elicit anti-predator responses in juvenile rainbow trout (*Oncoryhnchus mykiss*). *Canadian Journal of Zoology* **75**:1916–1922 DOI 10.1139/z97-821.
- **Brown GE, Smith RJF. 1998.** Acquired predator recognition in juvenile rainbow trout (*Oncorhynchus mykiss*): conditioning hatchery-reared fish to recognize chemical cues of a predator. *Canadian Journal of Fisheries and Aquatic Sciences* **55**:611–617 DOI 10.1139/cjfas-55-3-611.
- **Butler NE, Magrath RD, Peters RA. 2017.** Lack of alarm calls in a gregarious bird: models and videos of predators prompt alarm responses but no alarm calls by zebra finches. *Behavioral Ecology and Sociobiology* **71**:113

 DOI 10.1007/s00265-017-2343-z.
- Carlile PA, Peters RA, Evans CS. 2006. Detection of a looming stimulus by the Jacky dragon: selective sensitivity to characteristics of an aerial predator. *Animal Behaviour* 72:553–562 DOI 10.1016/j.anbehav.2005.10.027.
- Catano LB, Rojas MC, Malossi RJ, Peters JR, Heithaus MR, Fourqurean JW, Burkepile DE. 2016. Reefscapes of fear: predation risk and reef hetero-geneity interact to shape herbivore foraging behaviour. *Journal of Animal Ecology* 85:146–156 DOI 10.1111/1365-2656.12440.
- **Chivers DP, Smith RJF. 1998.** Chemical alarm signalling in aquatic predator–prey systems: a review and prospectus. *Ecoscience* **5**:338–352 DOI 10.1080/11956860.1998.11682471.
- **Chiverton PA. 1986.** Predator density manipulation and its effects on populations of Rhopalosiphumpadi (Horn.: Aphididae) in spring barley. *Annals of Applied Biology* **109(1)**:49–60 DOI 10.1111/j.1744-7348.1986.tb03183.x.

- Commens AM, Mathis A. 1999. Alarm pheromones of rainbow darters: responses to skin extracts of conspecifics and congeners. *Journal of Fish Biology* 55:1359–1362 DOI 10.1111/j.1095-8649.1999.tb02081.x.
- Creel S, Winnie JA, Christianson D, Liley S. 2008. Time and space in general models of antipredator response: tests with wolves and elk. *Animal Behaviour* 76:1139–1146 DOI 10.1016/j.anbehav.2008.07.006.
- Dieng H, Satho T, Suradi NFB, Hakim H, Abang F, Aliasan NE, Miake F, Zuharah WF, Kassim NFA, Majid AHA, Fadzly N, Vargas REM, Morales NP, Noweg GT. 2017. Presence of a predator image in potential breeding sites and oviposition responses of a dengue vector. *Acta Tropica* 146:446–454 DOI 10.1016/j.actatropica.2017.08.033.
- **Etting SF, Isbell LA. 2014.** Rhesus Macaques (Macaca mulatta) Use Posture to Assess Level of Threat From Snakes. *Ethology* **120**(12):1177–1184 DOI 10.1111/eth.12293.
- **Ferrari MCO, Wisenden BD, Chivers DP. 2010.** Chemical ecology of predator–prey interactions in aquatic ecosystems: a review and prospectus. *Canadian Journal of Zoology* **88**:698–724 DOI 10.1139/Z10-029.
- **Fischer S, Bohn L, Oberhummer E, Nyman C, Taborsky B. 2017a.** Divergence of developmental trajectories is triggered interactively by early social and ecological experience in a cooperative breeder. *Proceedings of the National Academy of Sciences of the United States of America* **114**:E9300–E9307 DOI 10.1073/pnas.1705934114.
- **Fischer S, Oberhummer E, Cunha-Saraiva F, Gerber N, Taborsky B. 2017b.** Smell or vision? The use of different sensory modalities in predator discrimination. *Behavioral Ecology and Sociobiology* **71**:143 DOI 10.1007/s00265-017-2371-8.
- **Fischer S, Taborsky B, Burlaud R, Fernandez AA, Hess S, Oberhummer E, Frommen JG. 2014.** Animated images as a tool to study visual communication: a case study in a cooperatively breeding cichlid. *Behaviour* **151**:1921–1942 DOI 10.1163/1568539X-00003223.
- **Fontaine JJ, Martin TE. 2006.** Parent birds assess nest predation risk and adjust their reproductive strategies. *Ecology Letters* **9**:428–434 DOI 10.1111/j.1461-0248.2006.00892.x.
- **Gerlai R. 2013.** Antipredatory behavior of zebrafish: adaptive function and a tool for translational research. *Evolutionary Psychology* **11**:591–605.
- **Gonzálvez FG, Rodríguez-Gironés MA. 2013.** Seeing is believing: information content and behavioural response to visual and chemical cues. *Proceedings of the Royal Society B: Biological Sciences* **280** DOI 10.1098/rspb.2013.0886.
- **Griesser M, Suzuki TN. 2017.** Naive juveniles are more likely to become breeders after witnessing predator mobbing. *The American Naturalist* **189**:58–66 DOI 10.1086/689477.
- Groenewoud F, Frommen JG, Josi D, Tanaka H, Jungwirth A, Taborsky M. 2016.

 Predation risk drives social complexity in cooperative breeders. *Proceedings of the National Academy of Sciences of the United States of America* 113:4104–4109 DOI 10.1073/pnas.1524178113.

- Hartman EJ, Abrahams MV. 2000. Sensory compensation and the detection of predators: the interaction between chemical and visual information. *Proceedings of the Royal Society B: Biological Sciences* **267** DOI 10.1098/rspb.2000.1039.
- Heg D, Bachar Z, Brouwer L, Taborsky M. 2004. Predation risk is an ecological constraint for helper dispersal in a cooperatively breeding cichlid. *Proceedings of the Royal Society B: Biological Sciences* 271:2367–2374 DOI 10.1098/rspb.2004.2855.
- **Heg D, Taborsky M. 2010.** Helper response to experimentally manipulated predation risk in the cooperatively breeding cichlid *Neolamprologus pulcher*. *PLOS ONE* **5**:e10784 DOI 10.1371/journal.pone.0010784.
- **Hickman CR, Stone MD, Mathis A. 2004.** Priority use of chemical over visual cues for detection of predators by graybelly salamanders, *Eurycea multiplicata griseogaster*. *Herpetologica* **60(2)**:203–210 DOI 10.1655/03-26.
- **Holling CS. 1965.** The functional response of predators to prey density and its role in mimicry and population regulation. *Memoirs of the Entomological Society of Canada* **97**:5–60 DOI 10.4039/entm9745.
- **Holomuzki JR, Hatchett LA. 1994.** Predator avoidance costs and habituation to fish chemicals by a stream isopod. *Freshwater Biology* **32**:585–592 DOI 10.1111/j.1365-2427.1994.tb01149.
- **Johnson JB, Basolo AL. 2003.** Predator exposure alters female mate choice in the green swordtail. *Behavioral Ecology* **14**:690–625 DOI 10.1093/beheco/arg046.
- **Kent MIA, Herbert-read JE, Mcdonald G, Wood AJ, Ward AJW. 2019.** Fine-scale behavioural adjustments of prey on a continuum of risk. *Proceedings of the Royal Society B-Biological Sciences* **286**.
- **Krause J, Winfield AFT, Deneubourg JL. 2011.** Interactive robots in experimental biology. *Trends in Ecology and Evolution* **26**:369–375 DOI 10.1016/j.tree.2011.03.015.
- Ladu F, Bartolini T, Panitz SG, Chiarotti F, Butail S, Macrì S, Porfiri M. 2015. Live predators, robots, and computer-animated images elicit differential avoidance responses in zebrafish. *Zebrafish* 12:205–214 DOI 10.1089/zeb.2014.1041.
- Landgraf T, Bierbach D, Nguyen H, Muggelberg N, Romanczuk P, Krause J. 2016.

 RoboFish: increased acceptance of interactive robotic fish with realistic eyes and natural motion patterns by live Trinidadian guppies. *Bioinspiration and Biomimetics* 11:015001 DOI 10.1088/1748-3190/11/1/015001.
- **Lenth R. 2018.** emmeans: estimated marginal means, aka least-squares means. R package version 1.3.1. *Available at https://CRAN.R-project.org/package=emmeans.*
- **Li B, Belasen A, Pafilis P, Bednekoff P, Foufopoulos J. 2014.** Effects of feral cats on the evolution of anti-predator behaviours in Island reptiles: insights from an ancient introduction. *Proceedings of the Royal Society B: Biological Sciences* **281** DOI 10.1098/rspb.2014.0339.
- **Lucon-Xiccato T. 2019.** Chemical alarm cues allow prey to adjust their defensive behaviour to cover abundance. *Behavioural Processes* **162**:86–89 DOI 10.1016/j.beproc.2019.02.004.

- Manassa RP, McCormick MI. 2012. Social learning and acquired recognition of a predator by a marine fish. *Animal Cognition* 15:559–565 DOI 10.1007/s10071-012-0484.
- Martin CW, Fodrie FJ, Heck KL, Mattila J. 2010. Differential habitat use and antipredator response of juvenile roach (*Rutilus rutilus*) to olfactory and visual cues from multiple predators. *Oecologia* 162:893–902 DOI 10.1007/s00442-010-1564.
- McCormick MI, Manassa R. 2008. Predation risk assessment by olfactory and visual cues in a coral reef fish. *Coral Reefs* 27:105–113 DOI 10.1007/s00338-007-0296-9.
- **Meuthen D, Flege P, Brandt R, Thünken T. 2018.** The location of damage-released alarm cues in a cichlid fish. *Aegyptus* **19**(**5**):529–546.
- Mirza RS, Scott JJ, Chivers DP. 2001. Differential responses of male and female red swordtails to chemical alarm cues. *Journal of Fish Biology* **59**:716–728 DOI 10.1006/jfbi.2001.1683.
- **Morales J, Lucas A, Velando A. 2018.** Maternal programming of offspring antipredator behavior in a seabird. *Behavioral Ecology* **29**:489–485 DOI 10.1093/beheco/arx197.
- **Moscicki MK, Hurd PL. 2017.** Damage-induced alarm cues influence lateralized behaviour but not the relationship between behavioural and habenular asymmetry in convict cichlids (*Amatitlania nigrofasciata*). *Animal Cognition* **20**:537–551 DOI 10.1007/s10071-017-1081.
- O'Connor CM, Reddon AR, Odetunde A, Jindal S, Balshine S. 2015. Social cichlid fish change behaviour in response to a visual predator stimulus, but not the odour of damaged conspecifics. *Behavioural Processes* 121:21–29 DOI 10.1016/j.beproc.2015.10.002.
- **Plisnier PD. 2002.** Limnological profiles and their variability in Lake Tanganyika. In: Odada EO, Olago DO, eds. *The East African great lakes: limnology, palaeolimnology and biodiversity.* Dordrecht: Kluwer Academic Publishers, 349–366.
- Plisnier PD, Chitamwebwa D, Mwape L, Tshibangu K, Langenberg V, Coenen E. 1999. Limnological annual cycle inferred from physical—chemical fluctuations at three stations of Lake Tanganyika. *Hydrobiologia* **407**:45–58 DOI 10.1023/A:1003762119873.
- **Porfiri M, Spinello C, Yang Y, Macrì S. 2019.** Zebrafish adjust their behavior in response to an interactive robotic predator. *Frontiers in Robotics and AI* **6**:38 DOI 10.3389/FROBT.2019.00038.
- **Preisser EL, Bolnick DI. 2008.** The many faces of fear: comparing the pathways and impacts of nonconsumptive predator effects on prey populations. *PLOS ONE* **6**:e2465 DOI 10.1371/journal.pone.0002465.
- **R Core Team. 2017.** R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing. *Available at https://www.R-project.org/*.
- Raderschall CA, Magrath RD, Hemmi JM. 2011. Habituation under natural conditions: model predators are distinguished by approach direction. *Journal of Experimental Biology* 214:4209–4216 DOI 10.1242/jeb.061614.
- **Reyes-Contreras M, Glauser G, Rennison DJ, Taborsky B. 2019.** Early-life manipulation of cortisol and its receptor alters stress axis programming and social competence. *Philosophical Transactions of the Royal Society B: Biological Sciences* **374** DOI 10.1098/rstb.2018.0119.

- **Rowland WJ. 1999.** Studying visual cues in fish behavior: a review of ethological techniques. *Environmental Biology of Fishes* **56**:285–305 DOI 10.1023/A:1007517720723.
- **Russell WMS, Burch RL. 1959.** *The principles of humane experimental technique.* London: Methuen & Co. Special edition published by Universities Federation for Animal Welfare (UFAW), 1992.
- **Scherer U, Godin JGJ, Schuett W. 2017.** Validation of 2D-animated pictures as an investigative tool in the behavioural sciences: a case study with a West African cichlid fish, Pelvicachromis pulcher. *Ethology* **123**:560–570 DOI 10.1111/eth.12630.
- **Stearns SC. 2000.** Life history evolution: successes, limitations, and prospects. *Naturwissenschaften* **87**:476–486 DOI 10.1007/s001140050763.
- **Su TL, Lim SSL. 2017.** To flee or not to flee: characterising differentiated anti-predatory responses of two mangrove crabs. *Ethology Ecology & Evolution* **29(2)**:181–192 DOI 10.1080/03949370.2015.1129989.
- **Taborsky M. 2016.** Cichlid fishes: a model for the integrative study of social behavior. In: Koenig W, Dickinson J, eds. *Cooperative breeding in vertebrates: studies of ecology, evolution, and behavior.* Cambridge: Cambridge University Press, 272–293 DOI 10.1017/CBO9781107338357.017.
- **Tollrian R. 1995.** Predator-induced morphological defenses: costs, life history shifts, and maternal effects in *Daphnia pulex*. *Ecology* **76**:1691–1705 DOI 10.2307/1940703.
- **Walsh MR, Reznick DN. 2008.** Interactions between the direct and indirect effects of predators determine life history evolution in a killifish. *Proceedings of the National Academy of Sciences of the United States of America* **105**:594–599 DOI 10.1073/PNAS.0710051105.
- Ward AJW, Mehner T. 2010. Multimodal mixed messages: the use of multiple cues allows greater accuracy in social recognition and predator detection decisions in the mosquitofish, *Gambusia holbrooki*. *Behavioral Ecology* 21:1315–1320 DOI 10.1093/beheco/arq152.
- **Watve M, Taborsky B. 2019.** Presence of parents during early rearing affects offspring responses towards predators. *Animal Behaviour* **158**:239–247 DOI 10.1016/j.anbehav.2019.09.012.